



IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of:

David BOTSTEIN, *et al.*

Application Serial No. 09/993,687

Filed: November 14, 2001

For: **SECRETED AND TRANSMEMBRANE
POLYPEPTIDES AND NUCLEIC
ACIDS ENCODING THE SAME**

) Examiner: Kemmerer, Elizabeth
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) Art Unit: 1646
)
) Confirmation No: 4943
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) Attorney's Docket No. 39780-2730 P1C11
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ON APPEAL TO THE BOARD OF PATENT APPEALS AND INTERFERENCES
APPELLANTS' BRIEF

MAIL STOP APPEAL BRIEF - PATENTS

Commissioner for Patents

P.O. Box 1450

Alexandria, Virginia 22304-1450

Dear Sir:

This Appeal Brief, filed in connection with the above captioned patent application, is responsive to the Final Office Actions mailed on September 27, 2006 and October 13, 2006. A Notice of Appeal was filed herein on March 13, 2007. A request for a one-month extension of time is requested herewith. Appellants hereby appeal to the Board of Patent Appeals and Interferences from the final rejection in this case.

The following constitutes the Appellants' Brief on Appeal.

06/18/2007 AADOF01 00000002 001641 09993687
01 FC:1251 120.00 DA

06/18/2007 AADOF01 00000001 001641 09993687
01 FC:1402 500.00 DA

I. REAL PARTY IN INTEREST

The real party in interest is Genentech, Inc., South San Francisco, California, by an assignment of the parent application, U.S. Patent Application Serial No. 09/941,992 recorded November 16, 2001, at Reel 012176 and Frame 0450.

II. RELATED APPEALS AND INTERFERENCES

The claims pending in the current application are directed to a polypeptide referred to herein as "PRO1009". There exist two related patent applications, (1) U.S. Patent Application Serial No. 09/991,181, now Patent No. 6, 913, 919, issued 07-05-2005 (containing claims directed to nucleic acids encoding PRO1009 polypeptides), and (2) U.S. Patent Application Serial No. 09/990,443, filed November 14, 2001 (containing claims directed to antibodies to PRO1009 polypeptides). Related U.S. Patent Application Serial No. 09/990,443 application is also under final rejection by the same Examiner and based upon the same outstanding rejections, an appeal is being pursued independently and concurrently herewith.

III. STATUS OF CLAIMS

Claims 119-126 and 129-131 are in this application.

Claims 1-118 and 127-128 have been canceled.

Claims 119-126 and 129-131 stand rejected and Appellants appeal the rejection of these claims.

A copy of the rejected claims in the present Appeal is provided as Claims Appendix (Section VIII).

IV. STATUS OF AMENDMENTS

A summary of the prosecution history for this case is as follows:

Previously, in response to a Final Office Action mailed on September 7, 2005, a Notice of Appeal was filed on December 6, 2005 and an Appeal Brief was filed on February 7, 2006. An Examiner's Answer was mailed on May 2, 2006, and an RCE Response with additional references and affidavits supporting Appellants' arguments was filed on June 29, 2006. Two Final Office Actions were mailed on September 27, 2006 and October 13, 2006, and a Notice of Appeal was filed March 13, 2007.

No claim amendments have been submitted after the last final rejection of October 13, 2006. A copy of the rejected claims in the present Appeal is provided as Appendix A.

V. SUMMARY OF CLAIMED SUBJECT MATTER

The invention claimed in the present application is related to an isolated polypeptide comprising the amino acid sequence of the polypeptide of SEQ ID NO:194, referred to in the present application as "PRO1009." The PRO1009 gene was shown for the first time in the present application to be significantly amplified in human colon cancers as compared to normal, non-cancerous human tissue controls (Example 170). This feature is specifically recited in Claim 124, and carried by all claims dependent from Claim 124. In addition, the invention also claims the amino acid sequence of the polypeptide of SEQ ID NO:194, lacking its associated signal-peptide; or the amino acid sequence of the polypeptide encoded by the full-length coding sequence of the cDNA deposited under ATCC accession number 209977 (Claims 124-126 and 129). The invention is further directed to polypeptides having at least 80% to 99% amino acid sequence identity to the amino acid sequence of the polypeptide of SEQ ID NO:194; the amino acid sequence of the polypeptide of SEQ ID NO:194, lacking its associated signal peptide; or the amino acid sequence of the polypeptide encoded by the full-length coding sequence of the cDNA deposited under ATCC accession number 209977, wherein the nucleic acid encoding said polypeptide is amplified in colon tumor (Claims 119-123). The invention is further directed to a chimeric polypeptide comprising one of the above polypeptides fused to a heterologous polypeptide (Claim 130), and to a chimeric polypeptide wherein the heterologous polypeptide is an epitope tag or an Fc region of an immunoglobulin (Claim 131). PRO polypeptide variants having at least about 80-99% amino acid sequence identity with a full length PRO polypeptide sequence, or a PRO polypeptide sequence lacking the signal peptide are generally described in the specification at, for example, page 305, line 23 onwards, and percent amino acid sequence identity determination is generally described at least at, for example, pages 306-308, line 14 onwards. The preparation of chimeric PRO polypeptides (Claims 130 and 131), including those wherein the heterologous polypeptide is an epitope tag or an Fc region of an immunoglobulin, is set forth in the specification at page 374, lines 24 to page 375, line 9. Examples 140-143 and page 376, line 12 onwards describe the expression of PRO polypeptides in various host cells, including *E. coli*, mammalian cells, yeast and Baculovirus-infected insect cells.

The amino acid sequence of the native "PRO1009" polypeptide and the nucleic acid sequence encoding this polypeptide (referred to in the present application as "DNA57129-1413") are shown in the present specification as SEQ ID NOs: 194 and 193, respectively, and in Figures 121 and 122, described on pages 293, lines 23-26. The full-length PRO1009 polypeptide having the amino acid sequence of SEQ ID NO:194 is described in the specification at, for example, on page 15 and pages 116-118 and the isolation of cDNA clones encoding PRO1009 of SEQ ID NO:194 is described in Example 51, page 444-445 of the specification.

Finally, Example 170, in the specification at page 539, line 19, to page 555, line 5, sets forth a 'Gene Amplification assay' which shows that the PRO1009 gene is amplified in the genome of certain human colon cancers (see Table 9B, page 552-553). The profiles of various primary colon tumors used for screening the PRO polypeptide compounds of the invention in the gene amplification assay are summarized on Table 8, page 546 of the specification.

VI. GROUND OF REJECTION TO BE REVIEWED ON APPEAL

1. Whether Claims 119-126 and 129-131 satisfy the utility/ enablement requirement under 35 U.S.C. §§101/112, first paragraph.
2. Whether Claims 119-123 and 130-131 satisfy the written description requirement under 35 U.S.C. §112, first paragraph.

VII. ARGUMENTS

Summary of the Arguments

Issue 1: Utility/ Enablement

Appellants rely upon the gene amplification data of the PRO1009 gene for patentable utility of the PRO1009 polypeptides. This data is clearly disclosed in the instant specification in Example 170 which discloses that the gene encoding PRO1009 showed significant amplification, ranging from **2.085 fold to 4.287-fold in twelve different colon primary tumors**. Appellants have submitted, in their Response filed August 4, 2005, a Declaration by Dr. Audrey Goddard, which explains that a gene identified as being amplified at least 2-fold by the disclosed gene amplification assay in a tumor sample relative to a normal sample is useful as a marker for the diagnosis of cancer, and for monitoring cancer development and/or for measuring the efficacy of cancer therapy. Therefore, such a gene is useful as a marker for the diagnosis of colon cancer, and for monitoring cancer development and/or for measuring the efficacy of cancer therapy.

Appellants have also submitted, in their Response filed June 18, 2004, ample evidence to show that, in general, if a gene is amplified in cancer, it is more likely than not that the encoded protein will be expressed at an elevated level. First, the articles by Orntoft *et al.*, Hyman *et al.*, and Pollack *et al.* collectively teach that in general, gene amplification increases mRNA expression. Second, the Declaration of Dr. Paul Polakis, principal investigator of the Tumor Antigen Project of Genentech, Inc., the assignee of the present application, shows that, in general, there is a correlation between mRNA levels and polypeptide levels. Third, Appellants further submit that even if there were no correlation between gene amplification and increased mRNA/protein expression, (which Appellants expressly do not concede), a polypeptide encoded by a gene that is amplified in cancer would still have a specific, substantial, and credible utility. Appellants submit that, as evidenced by the Ashkenazi Declaration and the teachings of Hanna and Mornin (both made of record in Appellants' Response filed June 18, 2004), simultaneous testing of gene amplification and gene product over-expression enables more accurate tumor classification, even if the gene-product, the protein, is not over-expressed. This leads to better determination of a suitable therapy for the tumor, as demonstrated by a real-world example of the breast cancer marker HER-2/neu. Appellants further note that the sale of gene expression chips to measure mRNA levels is a highly successful business, with a company such as Affymetrix recording 168.3 million dollars in sales of their GeneChip arrays in 2004. Clearly, the research community believes that the information obtained from these chips is useful (*i.e.*, that it is more likely than not informative of the protein level). Therefore, as a general rule, one skilled in the art would find it more likely than not that PRO1009 is useful as a diagnostic tool for detecting colon tumors.

The Examiner acknowledges on page 3 of the Final Office Action mailed September 7, 2005 that "the data in Table 9 may provide a basis for utility and enablement of PRO1009 nucleic acid," but contends that the data "does not provide a basis for utility or enablement of the claimed polypeptides. The art supports this position by establishing that there is no strong correlation between gene amplification and increased mRNA or protein levels." The Examiner maintains the rejection based on previously cited references Pennica *et al.*, Konopka *et al.*, Hu *et al.* and Haynes *et al.* and newly cited references Lian *et al.*, Fessler *et al.*, and Chen *et al.*

Appellants submit that the teachings of the Examiner's cited references do not conclusively establish a *prima facie* case for lack of utility (as will be discussed in detail below). In particular, contrary to the Examiner's interpretation, the data of Haynes *et al.* (see Figure 1) and Chen *et al.* (see Tables I and II of the paper) suggest that a positive correlation does exist between gene and protein expression. In addition, the teachings of Hu *et al.*, Lian *et al.* and Fessler *et al.* do not show a lack of correlation between mRNA and protein expression for genes in general. In fact, these cited references make clear references to various limitations in their studies and to their conclusions that were drawn by excluding certain data points. Appellants respectfully submit that such conclusions cannot be used to validate the Examiner's conclusions regarding the correlation between gene and mRNA/ protein expression in general. Since the Examiner has not cited evidentiary material that clearly addresses gene and mRNA/ protein expression in general, a *prima facie* case for lack of utility has not been made.

Taken together, although there are some examples in the scientific art that do not fit within the central dogma of molecular biology that there is generally a positive correlation between DNA, mRNA, and polypeptide levels, in general, in the majority of amplified genes, as exemplified by the teachings of Orntoft *et al.*, Hyman *et al.*, Pollack *et al.*, the Polakis Declaration, the art overwhelmingly show that gene amplification influences gene expression at the mRNA and protein levels. The widespread, art accepted use of information obtained from array chips for detecting diagnostic markers lend further support that in general, one of skill in the art would reasonably expect in this instance, based on the amplification data for the PRO1009 gene, that the PRO1009 polypeptide is concomitantly overexpressed and has utility in the diagnosis of colon cancer.

Accordingly, Appellants submit that when the proper legal standard is applied, one should reach the conclusion that the present application discloses at least one patentable utility for the claimed PRO1009 polypeptides. The specification also provides detailed guidance as to how to identify and make polypeptides having at least 80-99% amino acid sequence identity to PRO1009. Accordingly, one of ordinary skill in the art would understand how to make and use the recited polypeptide variants for the diagnosis of colon cancer without any undue experimentation.

Issue 2: Written Description

The factors to be considered in evidencing possession of a claimed genus include "disclosure of complete or partial structure, physical and/or chemical properties, functional characteristics, structure/function correlation, methods of making the claimed product, or any combination thereof." Current applicable case law holds that biological sequences are not adequately described solely by a description of their desired functional activities. It is, however, well established that a combination of functional and structural features suffices to describe a claimed genus, as discussed in the PTO's own Written Description Guidelines, and as set forth in *Enzo Biochem., Inc. v. Genprobe, Inc.* Appellants note that the claims recite structural features, namely, 80-99% sequence identity to the native sequence of SEQ ID NO:194, which are common to the genus. The genus of claimed polypeptides is further defined by having a specific functional activity for the encoding nucleic acids, namely, that the encoding nucleic acid is amplified in colon tumors. The specification provides detailed guidance as to how to identify the recited variants of SEQ ID NO:194, including methods for determining percent identity between two amino acid sequences, as well as listings of exemplary and preferred sequence substitutions, as well as detailed protocols for determining whether a gene encoding a variant PRO1009 protein is amplified in colon tumor. Thus, one of skill in the art could easily identify whether a variant PRO1009 sequence falls within the parameters of the claimed invention.

Accordingly, a description of the claimed genus has been achieved by the recitation of both structural and functional characteristics.

These arguments are all discussed in further detail below under the appropriate headings.

Response to Rejections

ISSUE 1. Claims 119-126 and 129-131 are Supported by a Credible, Specific and Substantial Asserted Utility, and Thus, Meet the Utility Requirement of 35 U.S.C. §§101/112, First Paragraph

The sole basis for the Examiner's rejection of Claims 119-126 and 129-131 under this section is that the data presented in Example 170 of the present specification is allegedly insufficient under the present legal standards to establish a patentable utility under 35 U.S.C. §101 for the presently claimed subject matter.

Claims 119-126 and 129-131 stand further rejected under 35 U.S.C. §112, first paragraph, allegedly "since the claimed invention is not supported by either a specific and substantial asserted

utility or a well established utility for the reasons set forth above, one skilled in the art clearly would not know how to use the claimed invention."

Appellants strongly disagree and, therefore, respectfully traverse the rejection.

A. The Legal Standard For Utility Under 35 U.S.C. §101

According to 35 U.S.C. §101:

Whoever invents or discovers any new and *useful* process, machine, manufacture, or composition of matter, or any new and *useful* improvement thereof, may obtain a patent therefor, subject to the conditions and requirements of this title.
(Emphasis added).

In interpreting the utility requirement, in *Brenner v. Manson*,¹ the Supreme Court held that the *quid pro quo* contemplated by the U.S. Constitution between the public interest and the interest of the inventors required that a patent Applicant disclose a "substantial utility" for his or her invention, *i.e.*, a utility "where specific benefit exists in currently available form."² The Court concluded that "a patent is not a hunting license. It is not a reward for the search, but compensation for its successful conclusion. A patent system must be related to the world of commerce rather than the realm of philosophy."³

Later, in *Nelson v. Bowler*,⁴ the C.C.P.A. acknowledged that tests evidencing pharmacological activity of a compound may establish practical utility, even though they may not establish a specific therapeutic use. The Court held that "since it is crucial to provide researchers with an incentive to disclose pharmaceutical activities in as many compounds as possible, we conclude adequate proof of any such activity constitutes a showing of practical utility."⁵

¹ *Brenner v. Manson*, 383 U.S. 519, 148 U.S.P.Q. (BNA) 689 (1966).

² *Id.* at 534, 148 U.S.P.Q. (BNA) at 695.

³ *Id.* at 536, 148 U.S.P.Q. (BNA) at 696.

⁴ *Nelson v. Bowler*, 626 F.2d 853, 206 U.S.P.Q. (BNA) 881 (C.C.P.A. 1980).

⁵ *Id.* at 856, 206 U.S.P.Q. (BNA) at 883.

In *Cross v. Iizuka*,⁶ the C.A.F.C. reaffirmed *Nelson*, and added that *in vitro* results might be sufficient to support practical utility, explaining that "*in vitro* testing, in general, is relatively less complex, less time consuming, and less expensive than *in vivo* testing. Moreover, *in vitro* results with the particular pharmacological activity are generally predictive of *in vivo* test results, i.e. there is a reasonable correlation there between."⁷ The Court perceived, "No insurmountable difficulty" in finding that, under appropriate circumstances, "*in vitro* testing, may establish a practical utility."⁸

The case law has also clearly established that Appellants' statements of utility are usually sufficient, unless such statement of utility is unbelievable on its face.⁹ The PTO has the initial burden to prove that Appellants' claims of usefulness are not believable on their face.¹⁰ In general, an Appellant's assertion of utility creates a presumption of utility that will be sufficient to satisfy the utility requirement of 35 U.S.C. §101, "unless there is a reason for one skilled in the art to question the objective truth of the statement of utility or its scope."^{11, 12}

Compliance with 35 U.S.C. §101 is a question of fact.¹³ The evidentiary standard to be used throughout *ex parte* examination in setting forth a rejection is a preponderance of the

⁶ *Cross v. Iizuka*, 753 F.2d 1047, 224 U.S.P.Q. (BNA) 739 (Fed. Cir. 1985).

⁷ *Id.* at 1050, 224 U.S.P.Q. (BNA) at 747.

⁸ *Id.*

⁹ *In re Gazave*, 379 F.2d 973, 154 U.S.P.Q. (BNA) 92 (C.C.P.A. 1967).

¹⁰ *Ibid.*

¹¹ *In re Langer*, 503 F.2d 1380, 1391, 183 U.S.P.Q. (BNA) 288, 297 (C.C.P.A. 1974).

¹² See also *In re Jolles*, 628 F.2d 1322, 206 U.S.P.Q. 885 (C.C.P.A. 1980); *In re Irons*, 340 F.2d 974, 144 U.S.P.Q. 351 (1965); *In re Sichert*, 566 F.2d 1154, 1159, 196 U.S.P.Q. 209, 212-13 (C.C.P.A. 1977).

¹³ *Raytheon v. Roper*, 724 F.2d 951, 956, 220 U.S.P.Q. (BNA) 592, 596 (Fed. Cir. 1983) *cert. denied*, 469 US 835 (1984).

totality of the evidence under consideration.¹⁴ Thus, to overcome the presumption of truth that an assertion of utility by the Appellant enjoys, the Examiner must establish that it is more likely than not that one of ordinary skill in the art would doubt the truth of the statement of utility. Only after the Examiner made a proper *prima facie* showing of lack of utility, does the burden of rebuttal shift to the Appellant. The issue will then be decided on the totality of evidence.

The well established case law is clearly reflected in the Utility Examination Guidelines (“Utility Guidelines”),¹⁵ which acknowledge that an invention complies with the utility requirement of 35 U.S.C. §101, if it has at least one asserted “specific, substantial, and credible utility” or a “well-established utility.” Under the Utility Guidelines, a utility is “specific” when it is particular to the subject matter claimed. For example, it is generally not enough to state that a nucleic acid is useful as a diagnostic without also identifying the conditions that are to be diagnosed.

In explaining the “substantial utility” standard, M.P.E.P. §2107.01 cautions, however, that Office personnel must be careful not to interpret the phrase “immediate benefit to the public” or similar formulations used in certain court decisions to mean that products or services based on the claimed invention must be “currently available” to the public in order to satisfy the utility requirement. “Rather, any reasonable use that an applicant has identified for the invention that can be viewed as providing a public benefit should be accepted as sufficient, at least with regard to defining a ‘substantial’ utility.”¹⁶ Indeed, the Guidelines for Examination of Applications for Compliance With the Utility Requirement,¹⁷ gives the following instruction to patent examiners: “If the Applicant has asserted that the claimed invention is useful for any particular practical purpose . . . and the assertion would be considered credible by a person of ordinary skill in the art, do not impose a rejection based on lack of utility.”

¹⁴ *In re Oetiker*, 977 F.2d 1443, 1445, 24 U.S.P.Q.2d (BNA) 1443, 1444 (Fed. Cir. 1992).

¹⁵ 66 Fed. Reg. 1092 (2001).

¹⁶ M.P.E.P. §2107.01.

¹⁷ M.P.E.P. §2107 II(B)(1).

B. Proper Application of the Legal Standard

Appellants submit that the evidentiary standard to be used throughout *ex parte* examination of a patent application is a preponderance of the totality of the evidence under consideration. Thus, to overcome the presumption of truth that an assertion of utility by the Appellant enjoys, the Examiner must establish that it is more likely than not that one of ordinary skill in the art would doubt the truth of the statement of utility. Only after the Examiner has made a proper *prima facie* showing of lack of utility, does the burden of rebuttal shift to the Appellant.

Appellants respectfully submit that the data presented in Example 170 starting on page 539 of the specification of the specification and the cumulative evidence of record, which underlies the current dispute, indeed support a "specific, substantial and credible" asserted utility for the presently claimed invention.

Patentable utility for the PRO1009 polypeptides is based upon the gene amplification data for the gene encoding the PRO1009 polypeptide. Example 170 describes the results obtained using a very well-known and routinely employed polymerase chain reaction (PCR)-based assay, the TaqManTM PCR assay, also referred to herein as the gene amplification assay. This assay allows one to quantitatively measure the level of gene amplification in a given sample, say, a tumor extract, or a cell line. It was well known in the art at the time the invention was made that gene amplification is an essential mechanism for oncogene activation. Appellants isolated genomic DNA from a variety of primary cancers and cancer cell lines that are listed in Table 9 (pages 539 onwards of the specification), including primary colon cancers of the type and stage indicated in Table 8 (page 546). The tumor samples were tested in triplicates with TaqmanTM primers and with internal controls, beta-actin and GADPH in order to quantitatively compare DNA levels between samples (page 548, lines 33-34). As a negative control, DNA was isolated from the cells of ten normal healthy individuals, which was pooled and used as a control (page 539, lines 27-29) and also, no-template controls (page 548, lines 33-34). The results of TaqManTM PCR are reported in ΔC_t units, as explained in the passage on page 539, lines 37-39. One unit corresponds to one PCR cycle or approximately a 2-fold amplification, relative to control, two units correspond to 4-fold, 3 units to 8-fold amplification and so on. Using this PCR-based assay, Appellants showed that the gene encoding for PRO1009 was amplified, that is,

it showed approximately 1.06-2.10 Δ Ct units which corresponds to $2^{1.06}$ - $2^{2.10}$ fold amplification or **2.085 fold to 4.287-fold** in twelve different colon primary tumors.

However, the Examiner states regarding the teachings of the Goddard Declaration that “the argument has been fully considered but is not deemed persuasive because it evidences that the instant specification provides a mere invitation to experiment, and not a readily available utility” (page 12, third paragraph of the Final Office Action mailed September 7, 2005. Appellants respectfully disagree.

Appellants point out that the Declaration by Dr. Audrey Goddard provides a statement by an expert in the relevant art that “fold amplification” values of at least 2-fold are considered significant in the TaqMan™ PCR gene amplification assay. Appellants particularly draw the Board's attention to page 3 of the Goddard Declaration which clearly states that:

It is further my considered scientific opinion that an at least **2-fold increase** in gene copy number in a tumor tissue sample relative to a normal (*i.e.*, non-tumor) sample is significant and useful in that the detected increase in gene copy number in the tumor sample relative to the normal sample serves as a basis for using relative gene copy number as quantitated by the TaqMan PCR technique as a diagnostic marker for the presence or absence of tumor in a tissue sample of unknown pathology. Accordingly, a gene identified as being amplified at least 2-fold by the quantitative TaqMan PCR assay in a tumor sample relative to a normal sample is **useful as a marker for the diagnosis of cancer**, for monitoring cancer development and/or for measuring the efficacy of cancer therapy. (Emphasis added).

Accordingly, the **2.085 fold to 4.287-fold** in twelve different colon primary tumors would be considered significant and credible by one skilled in the art, based upon the facts disclosed in the Goddard Declaration. As any skilled artisan in the field of oncology would easily appreciate that this gene is a good candidate marker for diagnosing colon tumors and would clearly find utility for the PRO1009 gene as a diagnostic for colon cancer or for diagnosing individuals at risk for developing colon cancer.

Further, as discussed below, Appellants had provided ample evidence in the form of articles from the art, like Orntoft *et al.*, Hyman *et al.*, Pollack *et al.*, and also the Polakis and Ashkenazi declarations, to show that, in general, if a gene is amplified in cancer, it is “more likely than not” that the encoded protein will also be expressed at an elevated level. The Examiner appears to disregard the ample evidence provided by the Appellants based on

misinterpretations of the teachings therein, as will be discussed below. Further, it is not a legal requirement to establish a necessary correlation between an increase in the copy number of the DNA and protein expression levels that would correlate to the disease state or that it is imperative to find evidence that DNA amplification is "necessarily" or "always" associated with overexpression of the gene product. Appellants respectfully submit that when the proper evidentiary standard is applied, a correlation must be acknowledged. The "more likely than not" standard is a much lower standard, and Appellants submit that, in fact, this standard is clearly met by the instant disclosure.

C. A prima facie case of lack of utility has not been established

The Examiner indicates based on Pennica *et al.*, Konopka *et al.* and Haynes *et al.* that "(w)hile the data in Table 9 may provide a basis for utility and enablement of PRO1009 nucleic acid, it does not provide a basis for utility or enablement of the claimed polypeptides" (Pages 3, last 5 lines of the Final Office Action mailed September 7, 2005).

Appellants respectfully submit that, contrary to the Examiner's assertion, none of the cited reference conclusively establish a *prima facie* case for lack of utility for the PRO1009 molecule. For instance, the teachings of Pennica *et al.* are specific to *WISP* genes, a specific class of closely related molecules. Pennica *et al.* showed that there was good correlation between DNA and mRNA expression levels for the *WISP-1* gene but not for *WISP-2* and *WISP-3* genes. But, the fact that in the case of closely related molecules, there seemed to be no correlation between gene amplification and the level of mRNA/protein expression does not establish that it is more likely than not, in general, that such correlation does not exist. As discussed above, the standard is not absolute certainty. Pennica *et al.* has no teaching whatsoever about the correlation of gene amplification and protein expression for genes in general. Similarly, in Konopka *et al.*, Appellants submit that the Examiner has generalized a very specific result disclosed by Konopka *et al.* to cover all genes. Konopka *et al.* actually state that "[p]rotein expression is not related to amplification of the *abl* gene but to variation in the level of *bcr-abl* mRNA produced from a single Ph¹ template." (See Konopka *et al.*, Abstract, emphasis added). The paper does not teach anything whatsoever about the correlation of protein expression and gene amplification in general, and provides no basis for the generalization that apparently underlies the present rejection. The statement of Konopka *et al.* that "[p]rotein

expression is not related to amplification of the *abl* gene . . . ” is not sufficient to establish a *prima facie* case of lack of utility. Therefore, the combined teachings of Pennica *et al.* and Konopka *et al.* are not directed towards genes in general but to a single gene or genes within a single family and thus, their teachings cannot support a general conclusion regarding correlation between gene amplification and mRNA or protein levels.

Actually, the cited reference Haynes *et al.*, showed that “there was a general trend, although no strong correlation between protein [expression] and transcript levels.” (see Figure 1 and page 1863, paragraph 2.1, last line). Therefore, when the proper legal standard is used, Haynes clearly supports the Appellants' position. This is all that's needed to meet the "more likely than not" evidentiary standard. Again, accurate prediction is not the standard. Therefore, a *prima facie* case of lack of utility has not been met based on the cited references Pennica *et al.*, Konopka *et al.* and Haynes *et al.*

Further, Appellants respectfully submit that, contrary to the Examiner's assertion, the cited Hu *et al.* reference, which was previously discussed in the response filed June 13, 2005, does not conclusively establish a *prima facie* case for lack of utility for the PRO1009 molecule. The Hu *et al.* reference is entitled “Analysis of Genomic and Proteomic Data using Advanced Literature Mining” (emphasis added). Therefore, as the title itself suggests, the conclusions in this reference are based upon statistical analysis of information obtained from published literature, and not from experimental data. Hu *et al.* performed statistical analysis to provide evidence for a relationship between mRNA expression and biological function of a given molecule (as in disease). The conclusions of Hu *et al.* however, only apply to a specific type of breast tumor (estrogen receptor (ER)-positive breast tumor) and cannot be generalized to breast cancer genes in general, let alone to cancer genes in general. Interestingly, the observed correlation was only found among ER-positive (breast) tumors not ER-negative tumors.” (See page 412, left column).

Moreover, the analytical methods utilized by Hu *et al.* have certain statistical drawbacks, as the authors themselves admit. For instance, according to Hu *et al.*, “*different statistical methods*” were applied to “*estimate the strength of gene-disease relationships and evaluated the results.*” (See page 406, left column, emphasis added). Using these different statistical methods, Hu *et al.* “[a]ssessed the relative strengths of gene-disease relationships based on the frequency of both co-citation and single citation.” (See page 411, left column). As is well known in the

art, different statistical methods allow different variables to be manipulated to affect the resulting outcome. In this regard, the authors disclose that, "Initial attempts to search the literature " using the list of genes, gene names, gene symbols, and frequently used synonyms generated by the authors "revealed several sources of false positives and false negatives." (See page 406, right column). The authors add that the false positives caused by "duplicative and unrelated meanings for the term" were "difficult to manage." Therefore, in order to minimize such false positives, Hu *et al.* disclose that these terms "had to be eliminated entirely, thereby reducing the false positive rate but unavoidably under-representing some genes." *Id.* (emphasis added). Hence, Hu *et al.* had to manipulate certain aspects of the input data, in order to generate, in their opinion, meaningful results. Further, because the frequency of citation for a given molecule and its relationship to disease only reflects the current research interest of a molecule, and not the true biological function of the molecule, as the authors themselves acknowledge, the "[r]elationship established by frequency of co-citation do not necessarily represent a true biological link." (See page 411, right column). Therefore, based on these findings, the authors add, "[t]his may reflect a bias in the literature to study the more prevalent type of tumor in the population. Furthermore, this emphasizes that caution must be taken when interpreting experiments that may contain subpopulations that behave very differently." *Id.* (Emphasis added). In other words, some molecules may have been underrepresented merely because they were less frequently cited or studied in literature compared to other more well-cited or studied genes. Therefore, Hu *et al.*'s conclusions are not based on genes/mRNA *in general*.

Therefore, Appellants submit that, based on the nature of the statistical analysis performed herein, and in particular, based on Hu's analysis of *one* class of genes, namely, the estrogen receptor (ER)-positive breast tumor genes, the conclusions drawn by the Examiner, namely that, "genes displaying a 5-fold change or less (mRNA expression) in tumors compared to normal showed no evidence of a correlation between altered gene expression and a known role in the disease (in general)" is not reliably supported.

The Examiner further cites new references by Lian *et al.*, Fessler *et al.* and Chen *et al.* in support of her interpretation that "protein levels cannot be accurately predicted from the level of the corresponding mRNA transcript." (Page 5 and 6 of the instant Final Office Action mailed September 7, 2005).

Appellants respectfully submit that Lian *et al.* only teach that protein expression may not correlate mRNA level in differentiating myeloid cells and does not teach anything of such a lack of correlation for genes in general. In fact, the authors themselves admit that there were a number of problems with their data. For instance, at page 520 of this article, the authors explicitly express their concerns regarding the methods they utilized and the interpretation of their data stating that "[t]hese data must be considered with several caveats: membrane and other hydrophobic proteins and very basic proteins are not well displayed by the standard 2DE approach, and proteins presented at low level will be missed. In addition, to simplify MS analysis, we used a Coomassie dye stain rather than silver to visualize proteins, and this decreased the sensitivity of detection of minor proteins." (Emphasis added). Appellants submit, as is well-known in the art, the Coomassie dye staining method is a very insensitive method of measuring protein. Therefore, the conclusions based on such measurements would hardly be considered accurate by the skilled artisan, or at least, would not be extrapolated to generally reflect the gene: mRNA/ protein relationships for proteins in general. Therefore, even if the teachings of Lian *et al.* reflects a lack of correlation between the genes and mRNA/ proteins in differentiating myeloid cells (which Appellants submit is not a representative sample of genes in general since only certain genes are expressed during differentiation), their conclusions are based on a widely accepted, insensitive method for protein staining, namely, the Coomassie dye staining, which cannot be applied to genes in general.

Similarly, in Fessler *et al.*, Appellants submit that the PTO has overlooked a number of limitations in their, which the authors themselves acknowledge. For instance, Fessler *et al.* only examined lipopolysaccharide-activated neutrophilins, so, as with Lian *et al.*, only examined the expression level of a few proteins/RNAs in response to LPS stimulation. Fessler *et al.* also concede that, since they used the Coomassie Blue dye staining method, which is known to have a limited protein binding range and a non-linear curve for protein detection, the resulting image analysis of the Coomassie Blue-stained proteins ought to be considered as semi-quantitative only (see page 31301, col. 1). Further, Fessler *et al.* submit that protein identification in their study was done using two-dimensional PAGE but admit that the analysis was limited only to well-resolved regions of the gel, which Fessler *et al.* explicitly concede, tends to select for more abundant protein species and therefore, may have performed less well with hydrophobic and high molecular weight proteins (see page 31301, col. 1). In addition, the Fessler *et al.* paper also

indicates that the harvesting of the LPS-incubated PMNs at 4 hours may have prevented the detection of early, transiently appearing proteins, and that the process of post-LPS incubation and pre-two-dimensional PAGE cell washes would expectedly remove secreted proteins from further analysis, perhaps contributing to the observed transcript-protein discordance. Therefore, the Fessler *et al.* reference explains the reasons for their transcript-protein discordance and like the Lian *et al.* reference, cannot be relied upon to make a general proposition that protein levels cannot be accurately predicted from mRNA levels.

In addition, as discussed above with Fessler *et al.*, Chen *et al.* also concede that there are problems with 2D gel protein detection and therefore, cannot accurately predict protein levels. For instance, Chen *et al.* says that, "(i)t is apparent that without prior enrichment only a relatively small and highly selected population of *long-lived, highly expressed proteins* is observed. There are many more proteins in a given cell which are not visualized by such methods. Frequently it is the low abundance proteins that execute key regulatory functions" (page 1870, col. 1). Thus, Chen *et al.*, concede that by selecting proteins visualized by 2D gels, they are likely to have excluded in their analysis many key regulatory proteins which could be candidate cancer markers.

However, the analysis provided by Chen *et al.* in fact support the Appellants general proposition that, even if protein levels cannot be accurately predicted (which is not required by the utility standard), in the majority of the proteins studied, it is most likely than not that an increase in gene amplification or mRNA levels generally correlates well with increased protein levels. A review of the correlation coefficient data presented in the Chen *et al.* paper indicates that, for instance, in Table 1, which lists 66 genes [the paper incorrectly states there are 69 genes listed] for which only one protein isoform is expressed, shows that 40 genes out of 66 had a positive correlation between mRNA expression and protein expression. This data clearly meets the standard for "more likely than not". Similarly, in Table II, 30 genes with multiple isoforms [again the paper incorrectly states there are 29] were presented. In this case, in 22 genes out of 30, at least one isoform showed a positive correlation between mRNA expression and protein expression. Furthermore, 12 genes out of 29 showed a strong positive correlation [as determined by the authors] for at least one isoform. No genes showed a significant negative correlation. Thus, Table II of Chen *et al.* also provides that it is more likely than not that protein levels will correlate with mRNA expression levels. In fact, the same authors in Chen *et al.*, published a

latter paper (Beer *et al.*, Nature Medicine 8(8) 816-824 (2002)- copy enclosed) which described the expression of genes in adenocarcinomas as compared to protein expression. They observed that "these results suggest that the oligonucleotide microarrays provided reliable measures of gene expression" (pg 317) and further stated that "these studies indicate that many of the genes identified using gene expression profiles are likely relevant to lung adenocarcinoma". Therefore, the authors of the Chen paper clearly agreed that microarrays provided a reliable measure of the expression levels of the gene and could be used to identify genes whose overexpression is associated with tumors.

In summary, Hu *et al.*, Lian *et al.* and Fessler *et al.* do not conclusively teach that, in general, protein levels cannot be accurately predicted from mRNA/ gene amplification levels. These authors concede that either due to insensitive protein detection methods or due their methodologies utilized in their protocols, some protein species may have been underrepresented over others. Therefore, the teachings of these references cannot be relied upon to establish a *prima facie* showing of lack of utility. On the other hand, as noted even in Haynes *et al.* and Chen *et al.*, most genes showed a positive correlation between increased gene amplification, mRNA and translated protein.

Appellants once again remind the Examiner that only after the Examiner has made a proper prima facie showing of lack of utility, does the burden of rebuttal shift to the applicant. Based on the above discussions, such a showing has not been made in this instance. Accordingly, the instant rejection should be withdrawn for the Examiner's lack of establishment of a *prima facie* showing.

D. The Gene Amplification Data Establishes Credible, Substantial and Specific Patentable Utility for the PRO1009 Polypeptide

In fact, as discussed throughout prosecution, Appellants submit that Example 170 of the specification further discloses that, "(a)mplification is associated with overexpression of the gene product, indicating that the polypeptides are useful targets for therapeutic intervention in certain cancers such as lung, colon, breast and other cancers and diagnostic determination of the presence of those cancers" (Emphasis added). Appellants have also submitted ample evidence to show that, in general, if a gene is amplified in cancer, it is "more likely than not" that the encoded protein will also be expressed at an elevated level.

For instance, Appellants presented the articles by Orntoft *et al.*, Hyman *et al.*, and Pollack *et al.* (made of record in Appellants' Response filed June 18, 2004), who collectively teach that in general, for most genes, DNA amplification increases mRNA expression. The results presented by Orntoft *et al.*, Hyman *et al.*, and Pollack *et al.* are based upon wide ranging analyses of a large number of tumor associated genes. Orntoft *et al.* studied transcript levels of 5600 genes in malignant bladder cancers, many of which were linked to the gain or loss of chromosomal material, and found that in general (18 of 23 cases) chromosomal areas with more than 2-fold gain of DNA showed a corresponding increase in mRNA transcripts. Hyman *et al.* compared DNA copy numbers and mRNA expression of over 12,000 genes in breast cancer tumors and cell lines, and found that there was evidence of a prominent global influence of copy number changes on gene expression levels. In Pollack *et al.*, the authors profiled DNA copy number alteration across 6,691 mapped human genes in 44 predominantly advanced primary breast tumors and 10 breast cancer cell lines, and found that on average, a 2-fold change in DNA copy number was associated with a corresponding 1.5-fold change in mRNA levels. In summary, the evidence supports the Appellants' position that gene amplification is more likely than not predictive of increased mRNA and polypeptide levels.

Second, the Declaration of Dr. Paul Polakis (made of record in Appellants' Response filed June 29, 2006), principal investigator of the Tumor Antigen Project of Genentech, Inc., the assignee of the present application, explains that in the course of Dr. Polakis' research using microarray analysis, he and his co-workers identified approximately 200 gene transcripts that are present in human tumor cells at significantly higher levels than in corresponding normal human cells. Appellants submit that Dr. Polakis' Declaration was presented to support the position that there is a correlation between mRNA levels and polypeptide levels, the correlation between gene amplification and mRNA levels having already been established by the data shown in the Orntoft *et al.*, Hyman *et al.*, and Pollack *et al.* articles. Appellants further emphasize that the opinions expressed in the Polakis Declaration, including in the above quoted statement, are all based on factual findings. For instance, antibodies binding to about 30 of these tumor antigens were prepared, and mRNA and protein levels were compared. In approximately 80% of the cases, the researchers found that increases in the level of a particular mRNA correlated with changes in the level of protein expressed from that mRNA when human tumor cells are compared with their corresponding normal cells. Therefore, Dr. Polakis' research, which is referenced in his

Declaration, shows that, in general, there is a correlation between increased mRNA and polypeptide levels. Hence, one of skill in the art would reasonably expect that, based on the gene amplification data of the PRO1009 gene, the PRO1009 polypeptide is concomitantly overexpressed in the colon tumors studied as well.

Appellants further note that the sale of gene expression chips to measure mRNA levels is a highly successful business, with a company such as Affymetrix recording 168.3 million dollars in sales of their GeneChip® arrays in 2004. Clearly, the research community believe that the information obtained from these chips is useful (*i.e.*, that it is more likely than not that the results are informative of protein levels).

The Examiner appears to disregard the ample evidence provided in the above referenced articles based on misinterpretations of their teachings. The “more likely than not” standard is a much lower standard than a “necessary” correlation or “accurate” prediction, and Appellants submit that in fact, this standard is clearly met by the instant disclosure, and furthermore, the Declarations and the articles by the Appellants lend significant support to the fact that for an amplified gene, it is more likely than not that the protein will also be overexpressed. Moreover, the Examiner’s cited references do not present a *prima facie* case of lack of utility, as discussed above.

Thus, based on the asserted utility for PRO1009 in the diagnosis of selected colon tumors, the reduction to practice of the instantly claimed protein sequence of SEQ ID NO:194 in the present application, the disclosure of the step-by-step protocols for making chimeric PRO polypeptides, including those wherein the heterologous polypeptide is an epitope tag or an Fc region of an immunoglobulin in the specification (at page 374, lines 24 to page 375, line 9), the disclosure of a step-by-step protocol for making and expressing PRO1009 in appropriate host cells (in Examples 140-143 and page 376, line 12), the step-by-step protocol for the preparation, isolation and detection of monoclonal, polyclonal and other types of antibodies against the PRO1009 protein in the specification (at pages 390-395) and the disclosure of the gene amplification assay in Example 170, the skilled artisan would know exactly how to make and use the claimed polypeptide for the diagnosis of colon cancers. Appellants submit that based on the detailed information presented in the specification and the advanced state of the art in oncology, the skilled artisan would have found such testing routine and not ‘undue’.

Therefore, Appellants respectfully request reconsideration and reversal of this outstanding rejections under 35 U.S.C. §101 and §112, First Paragraph to Claims 119-126 and 129-131.

ISSUE 2: Claims 119-123, 130 and 131 Satisfy the Written Description Requirement of 35 U.S.C. §112, First Paragraph

Claims 119-123, 130 and 131 stand rejected under 35 U.S.C. §112, first paragraph, as allegedly containing "subject matter which was not described in the specification in such a way as to reasonably convey to one skilled in the relevant art that the inventor(s), at the time the application was filed, had possession of the claimed invention." In particular, the Examiner has asserted that "the specification does not (disclose) any variants of SEQ ID NO: 194, naturally occurring or not, nor whether such sequences are amplified in colon tumors" (Page 14, Final Office Action mailed September 7, 2005). The Examiner also cites *Fiers v. Revel*, 25 U.S.P.Q.2d 1601 (Fed. Cir. 1993) and *Fiddes v. Baird*, 30 U.S.P.Q.2d 1481, 1483 (1993) to show that .

Appellants respectfully disagree.

A. The Legal Test for Written Description

The well-established test for sufficiency of support under the written description requirement of 35 U.S.C. §112, first paragraph is "whether the disclosure of the application as originally filed reasonably conveys to the artisan that the inventor had possession at that time of the later claimed subject matter, rather than the presence or absence of literal support in the specification for the claim language."^{18,19} The adequacy of written description support is a factual issue and is to be determined on a case-by-case basis.²⁰ The factual determination in a

¹⁸ *In re Kaslow*, 707 F.2d 1366, 1374, 212 U.S.P.Q. 1089, 1096 (Fed. Cir. 1983).

¹⁹ *See also Vas-Cath, Inc. v. Mahurkar*, 935 F.2d at 1563, 19 U.S.P.Q.2d at 1116 (Fed. Cir. 1991).

²⁰ *See e.g., Vas-Cath*, 935 F.2d at 1563; 19 U.S.P.Q.2d at 1116.

written description analysis depends on the nature of the invention and the amount of knowledge imparted to those skilled in the art by the disclosure.^{21, 22}

In *Environmental Designs, Ltd. v. Union Oil Co.*,²³ the Federal Circuit held, "Factors that may be considered in determining level of ordinary skill in the art include: (1) the educational level of the inventor; (2) type of problems encountered in the art; (3) prior art solutions to those problems; (4) rapidity with which innovations are made; (5) sophistication of the technology; and (6) educational level of active workers in the field."²⁴ Further, the "hypothetical 'person having ordinary skill in the art' to which the claimed subject matter pertains would, of necessity have the capability of understanding the scientific and engineering principles applicable to the pertinent art."^{25, 26}

B. The Disclosure Provides Sufficient Written Description for the Claimed Invention

Appellants respectfully submit that the instant specification evidences the actual reduction to practice of the native amino acid sequence of SEQ ID NO: 194. Support for "native sequences" can be found in the instant specification at least at page 304, line 26. Appellants also submit that the specification provides ample written support for determining percent sequence identity between two amino acid sequences (See pages 306-308, line 14 onwards). In fact, the specification teaches specific parameters to be associated with the term "percent identity" as applied to the present invention. The specification further provides detailed guidance as to changes that may be made to a PRO polypeptide without adversely affecting its activity

²¹ *Union Oil v. Atlantic Richfield Co.*, 208 F.2d 989, 996 (Fed. Cir. 2000).

²² *See also* M.P.E.P. §2163 II(A).

²³ 713 F.2d 693, 696, 218 U.S.P.Q. 865, 868 (Fed. Cir. 1983), *cert. denied*, 464 U.S. 1043 (1984).

²⁴ *See also* M.P.E.P. §2141.03.

²⁵ *Ex parte Hiyamizu*, 10 U.S.P.Q.2d 1393, 1394 (Bd. Pat. App. & Inter. 1988) (emphasis added).

²⁶ *See also* M.P.E.P. §2141.03.

(page 372, line 36 to page 373, line 17). This guidance includes a listing of exemplary and preferred substitutions for each of the twenty naturally occurring amino acids (Table 6, page 372). Accordingly, one of skill in the art could identify whether the variant PRO1009 sequence falls within the parameters of the claimed invention. Once such an amino acid sequence was identified, the specification sets forth methods for making the amino acid sequences (see page 376, line 9) and methods of preparing the PRO polypeptides (see Examples 140-143).

Currently pending Claims 119-123 and 130-131 recite the functional recitation that the nucleic acid encoding the claimed polypeptides are amplified in colon tumors. Appellants further submit that the specification provides ample written support for detecting and quantifying amplification of such nucleic acids in several tumors and/or cell lines as described in Example 170. Example 170 of the present application provides step-by-step guidelines and protocols for the gene amplification assay. By following this disclosure, one skilled in the art would know that it is easy to test whether a gene encoding a variant PRO1009 protein is amplified in colon tumors by the methods set forth in Example 170.

Appellants refer to the arguments and information presented above in response to the outstanding rejections under 35 U.S.C. §101 and 35 U.S.C. §112, first paragraph, for alleged lack of utility and enablement. These arguments are incorporated by reference herein. Appellants respectfully submit that as discussed above under Issue I, the teachings in the art, as exemplified by Orntoft *et al.*, Hyman *et al.*, Pollack *et al.*, and the Polakis Declaration, overwhelmingly show that gene amplification influences gene expression at the mRNA and protein levels. Thus, the amplification of the encoding polynucleotide in tumors does provide useful information regarding the functional property of the polypeptide in being overexpressed in tumor tissues.

Appellants further respectfully submit that whether or not the polypeptide is also overexpressed in tumor tissues is irrelevant to the consideration of adequate written description. The claims have characterized the recited polypeptides as having the property that their encoding polynucleotides are amplified in colon tumors. As discussed above, the specification describes methods for identifying genes which are amplified in colon tumors. Therefore, one of skill in the art could readily test a nucleic acid sequence which encodes a variant polypeptide to determine whether it is amplified by the methods set forth in Example 170. Thus, the recited property of

amplification of the encoding gene adds to the characterization of the claimed polypeptide sequences in a manner that one of skill in the art could readily assess and understand.

Applicants respectfully submit that the issue is whether the claimed sequences are adequately described. Appellants respectfully submit that the full-length sequence of PRO1009 is clearly provided in the specification. The Examiner has acknowledged that the specification discloses the full-length sequence of SEQ ID NO:194 and that polypeptides comprising the sequence set forth in SEQ ID NO:194 meet the written description provision of 35 U.S.C. §112, first paragraph.

Secondly, Appellants submit that the court in *Fiers v. Revel* held that “[i]f a conception of a DNA requires a precise definition, such as by structure, formula, chemical name, or physical properties, as we have held, then a description also requires that degree of specificity.” *Fiers*, 984 F.2d at 1171. Since the instant claims are directed to polypeptides, *Fiers* is distinguished on the facts and does not apply. Similarly, the *Fiddes* decision holds that naturally-occurring gene sequences cannot be patented unless the actual sequence is clearly disclosed in the patent application. Again, since the instant claims are directed to polypeptides, *Fiddes* is distinguished on the facts and does not apply.

In *Fiddes v. Baird*, the Board of Patent Appeals and Interferences held that party Fiddes’ claims to a human gene for basic fibroblast growth factor were separately patentable over party Baird’s issued and pending claims specifying a sequence encoding “mammalian” basic fibroblast growth factor. Party Baird’s issued patent disclosed the amino acid sequence that had been isolated from bovine pituitary and a theoretical DNA sequence encoding it. Party Baird’s pending claims were from a continuation-in-part application disclosing the naturally-occurring coding sequence for bovine fibroblast growth factor. Between the filing of the first application and the continuation-in-part application, DNA sequences anticipating claims to the naturally-occurring human fibroblast growth factor were published. Party Baird argued that the published sequence could not be used as prior art because it was entitled to the filing date of the first application.

The Board held that party Baird was not entitled to the filing date of the first application of its claims to the mammalian DNA sequence because it did not set out specific DNA sequences of naturally-occurring mammalian genes in the first application. Therefore, the first application did not meet the “written description” for the specific DNA sequences of naturally-occurring

mammalian genes. The Board added that party Baird was not in possession of the naturally occurring bovine gene at the time of filing the first application even though its encoded amino acid sequence was known.

Appellants respectfully submit that in the present application, Appellants have clearly disclosed the full-length sequence of SEQ ID NO:194 and its encoding nucleic acid sequence SEQ ID NO:193. In addition, one of skill in the art could identify whether the variant PRO1009 sequence falls within the parameters of the claimed invention. Furthermore, as mentioned above, since the instant claims are directed to polypeptides, *Fiddes* is distinguished on the facts and does not apply. Accordingly, the Examiner's assertion that Appellants provide no written description in the specification for any other species of PRO1009 molecules is based on the incorrect interpretation of the holding in the *Fiddes* case.

More recently, in *Enzo Biochem., Inc. v. Genprobe, Inc.* 296 F.3d 1316 (Fed. Cir. 2002), the court adopted the standard that "the written description requirement can be met by showing that the invention is complete by disclosure of sufficiently detailed, relevant identifying characteristics, . . . *i.e.*, complete or partial structure, other physical and/or chemical properties, functional characteristics when coupled with a known or disclosed correlation between function and structure, or some combination of such characteristics." *Id.* at 1324. While the invention in *Enzo* was still a DNA, the holding has been treated as being applicable to proteins as well. Indeed, the court adopted the standard from the USPTO's Written Description Examination Guidelines, which apply to both proteins and nucleic acids.

Accordingly, current applicable case law holds that biological sequences are not adequately described solely by a description of their desired functional activities. The instant claims meet the standard set by the *Enzo* court in that the claimed sequences are defined not only by functional properties, but also by structural limitations. It is well established that a combination of functional and structural features may suffice to describe a claimed genus. "An applicant may also show that an invention is complete by disclosure of sufficiently detailed, relevant identifying characteristics which provide evidence that applicant was in possession of the claimed invention, *i.e.*, complete or partial structure, other physical and/or chemical properties, functional characteristics when coupled with a known or disclosed correlation

between function and structure, or some combination of such characteristics."²⁷ Thus, the genus of polypeptides with at least 80-99% sequence identity to SEQ ID NO:194, which possess the functional property of having a nucleic acid which is amplified in colon tumor would meet the requirement of 35 U.S.C. §112, first paragraph, as providing adequate written description. Accordingly, one skilled in the art would have known that Appellants had knowledge and possessed the claimed polypeptides with 80-99% sequence identity to SEQ ID NO: 194 whose encoding nucleic acids were amplified in colon tumors. The recited property of amplification of the encoding gene adds to the characterization of the claimed polypeptide sequences in a manner that one of skill in the art could readily assess and understand. As discussed above, Appellants have recited structural features, namely, 80-99% sequence identity to SEQ ID NO: 194, which are common to the genus. Appellants have also provided guidance as to how to make the recited variants of SEQ ID NO:194, including listings of exemplary and preferred sequence substitutions. The genus of claimed polypeptides is further defined by having a specific functional activity for the encoding nucleic acids. Accordingly, a description of the claimed genus has been achieved.

Accordingly, Appellants respectfully request reconsideration and reversal of the written description rejection of Claims 119-123 and 130-131 under 35 U.S.C. §112, first paragraph.

²⁷ M.P.E.P. §2163 II(A)(3)(a).

CONCLUSION

For the reasons given above, Appellants submit that present specification clearly describes, details and provides a patentable utility for the claimed invention. Moreover, it is respectfully submitted that based upon this disclosed patentable utility, the present specification clearly teaches "how to use" the presently claimed polypeptide. As such, Appellants respectfully request reconsideration and reversal of the outstanding rejection of Claims 119-126 and 129-131.

The Commissioner is authorized to charge any fees which may be required, including extension fees, or credit any overpayment to Deposit Account No. **08-1641** (referencing Attorney's Docket No. **39780-2730 P1C11**).

Respectfully submitted,

Date: June 13, 2007

By: *Daphne Reddy*
Daphne Reddy (Reg. No. 53,507)

HELLER EHRMAN LLP
275 Middlefield Road
Menlo Park, California 94025-3506
Telephone: (650) 324-7000
Facsimile: (650) 324-0638

VIII. CLAIMS APPENDIX

Claims on Appeal

119. An isolated native sequence polypeptide having at least 80% amino acid sequence identity to:

- (a) the amino acid sequence of the polypeptide of SEQ ID NO: 194;
 - (b) the amino acid sequence of the polypeptide of SEQ ID NO: 194, lacking its associated signal peptide;
 - (c) the amino acid sequence of the polypeptide encoded by the full-length coding sequence of the cDNA deposited under ATCC accession number 209977,
- wherein, the nucleic acid encoding said polypeptide is amplified in colon tumors.

120. The isolated native sequence polypeptide of Claim 119 having at least 85% amino acid sequence identity to:

- (a) the amino acid sequence of the polypeptide of SEQ ID NO: 194;
 - (b) the amino acid sequence of the polypeptide of SEQ ID NO: 194, lacking its associated signal peptide;
 - (c) the amino acid sequence of the polypeptide encoded by the full-length coding sequence of the cDNA deposited under ATCC accession number 209977,
- wherein, the nucleic acid encoding said polypeptide is amplified in colon tumors.

121. The isolated native sequence polypeptide of Claim 119 having at least 90% amino acid sequence identity to:

- (a) the amino acid sequence of the polypeptide of SEQ ID NO: 194;
 - (b) the amino acid sequence of the polypeptide of SEQ ID NO: 194, lacking its associated signal peptide;
 - (c) the amino acid sequence of the polypeptide encoded by the full-length coding sequence of the cDNA deposited under ATCC accession number 209977,
- wherein, the nucleic acid encoding said polypeptide is amplified in colon tumors.

122. The isolated native sequence polypeptide of Claim 119 having at least 95% amino acid sequence identity to:

- (a) the amino acid sequence of the polypeptide of SEQ ID NO: 194;

(b) the amino acid sequence of the polypeptide of SEQ ID NO: 194, lacking its associated signal peptide;

(c) the amino acid sequence of the polypeptide encoded by the full-length coding sequence of the cDNA deposited under ATCC accession number 209977,

wherein, the nucleic acid encoding said polypeptide is amplified in colon tumors.

123. The isolated native sequence polypeptide of Claim 119 having at least 99% amino acid sequence identity to:

(a) the amino acid sequence of the polypeptide of SEQ ID NO: 194;

(b) the amino acid sequence of the polypeptide of SEQ ID NO: 194, lacking its associated signal peptide,

(c) the amino acid sequence of the polypeptide encoded by the full-length coding sequence of the cDNA deposited under ATCC accession number 209977,

wherein, the nucleic acid encoding said polypeptide is amplified in colon tumors.

124. An isolated polypeptide comprising:

(a) the amino acid sequence of the polypeptide of SEQ ID NO: 194;

(b) the amino acid sequence of the polypeptide of SEQ ID NO: 194, lacking its associated signal peptide,

(c) the amino acid sequence of the polypeptide encoded by the full-length coding sequence of the cDNA deposited under ATCC accession number 209977,

wherein, the nucleic acid encoding said polypeptide is amplified in colon tumors.

125. The isolated polypeptide of Claim 124 comprising the amino acid sequence of the polypeptide of SEQ ID NO: 194.

126. The isolated polypeptide of Claim 124 comprising the amino acid sequence of the polypeptide of SEQ ID NO: 194, lacking its associated signal peptide.

129. The isolated polypeptide of Claim 124 comprising the amino acid sequence of the polypeptide encoded by the full-length coding sequence of the cDNA deposited under ATCC accession number 209977.

130. A chimeric polypeptide comprising a polypeptide according to Claim 124 fused to a heterologous polypeptide.

131. The chimeric polypeptide of Claim 130, wherein said heterologous polypeptide is an epitope tag or an Fc region of an immunoglobulin.

IX. EVIDENCE APPENDIX

1. Declaration of Audrey Goddard, Ph.D. under 35 C.F.R. §1.132, with attached Exhibits A-G:
 - A. Curriculum Vitae of Audrey D. Goddard, Ph.D.
 - B. Higuchi, R. *et al.*, "Simultaneous amplification and detection of specific DNA sequences," *Biotechnology* 10:413-417 (1992).
 - C. Livak, K.J., *et al.*, "Oligonucleotides with fluorescent dyes at opposite ends provide a quenched probe system useful for detecting PCR product and nucleic acid hybridization," *PCR Methods Appl.* 4:357-362 (1995).
 - D. Heid, C.A. *et al.*, "Real time quantitative PCR," *Genome Res.* 6:986-994 (1996).
 - E. Pennica, D. *et al.*, "WISP genes are members of the connective tissue growth factor family that are up-regulated in Wnt-1-transformed cells and aberrantly expressed in human colon tumors," *Proc. Natl. Acad. Sci. USA* 95:14717-14722 (1998).
 - F. Pitti, R.M. *et al.*, "Genomic amplification of a decoy receptor for Fas ligand in lung and colon cancer," *Nature* 396:699-703 (1998).
 - G. Bieche, I. *et al.*, "Novel approach to quantitative polymerase chain reaction using real-time detection: Application to the detection of gene amplification in breast cancer," *Int. J. Cancer* 78:661-666 (1998).
2. Declaration of Avi Ashkenazi, Ph.D. under 35 C.F.R. §1.132, with attached Exhibit A (Curriculum Vitae).
3. Declaration of Paul Polakis, Ph.D. under 35 C.F.R. §1.132 (Polakis I).
4. Declaration of Paul Polakis, Ph.D. under 35 C.F.R. §1.132 (Polakis II).
5. Hyman, E., *et al.*, "Impact of DNA Amplification on Gene Expression Patterns in Breast Cancer," *Cancer Research* 62:6240-6245 (2002).
6. Pollack, J.R., *et al.*, "Microarray Analysis Reveals a Major Direct Role of DNA Copy Number Alteration in the Transcriptional Program of Human Breast Tumors," *Proc. Natl. Acad. Sci. USA* 99:12963-12968 (2002).
7. Hanna *et al.*, "HER-2/neu Breast Cancer Predictive Testing," Pathology Associates Medical Laboratories (1999).
8. Konopka *et al.*, "Variable Expression of the Translocated c-abl oncogene in Philadelphia-chromosome-positive B-lymphoid cell lines from chronic myelogenous leukemia patients" *Proc. Natl. Acad. Sci. USA* 83: 4049-52, (1986).
9. Haynes *et al.*, "Proteome analysis: Biological assay or data archive?" *Electrophoresis* 19:1862-1871 (1996).
10. Hu *et al.*, "Analysis of genomic and proteomic data using advanced literature mining," *J. Proteome Res.* 2: 405-412 (2003).
11. Lian *et al.*, "Genomic and proteomic analysis of the myeloid differentiation program," *Blood* 98: 513-524 (2001).

12. Fessler *et al.*, "A genomic and proteomic analysis of activation of the human neutrophil by lipopolysaccharide and its mediation by p38 mitogen-activated protein kinase," *J. Biol. Chem.* 277: 31291-31302 (2002).
13. Chen *et al.*, "Discordant Protein and mRNA Expression in Lung Adeno-carcinomas," *Mol. Cellular Proteomics*, 1: 304-313 (2002).
14. Abe, N., *et al.*, *Br J Cancer* - 89(11):2104-9 (2003).
15. Alberts, B., *et al.*, *Molecular Biology of the Cell* (3rd ed. 1994) Cell 3rd at 453 Figure 9-2 of Cell 3rd Cell 3rd at 403.
16. Alberts, B., *et al.*, *Molecular Biology of the Cell* (4rd ed.) In Cell 4th, Figure 6-3 on page 302 Figure 6-90 on page 364 of Cell 4th Cell 4th at 364 Cell 4th at 379.
17. Ando, M., *et al.*, *Br J Haematol.*, - 130(6):860-8 (2005).
18. Aust, G., *et al.*, *Thyroid* - 7(5):713-24 (1997).
19. Barnes, V.L., *et al.*, *J Histochem Cytochem.* - 47(6):787-98 (1999).
20. Bea, S., *et al.*, *Cancer Res.* - 61(6):2409-12 (2001).
21. Beer, *et al.*, *Nature Medicine* - 8(8):816-824 (2002).
22. Blaschke, V., *et al.*, *J Immunol Methods.* - 246(1-2):79-90 (2000).
23. Buckley, A.R., *et al.*, *Apoptosis.* - 2(6):518-28 (1997).
24. Caberlotto, L. *et al.*, *Neurosci Lett.* - 265(3):191-4 (1999).
25. Caberlotto, L., *et al.*, *Eur J Neurosci.* - 17(9):1736-46 (2003).
26. Choi, D., *et al.*, *J Soc Gynecol Investig.* - 9(1):41-6 (2002).
27. Couvelard, A., *et al.*, *Virchows Arch.* - 445(3):279-84 (2004).
28. Dagenais, A., *et al.*, *Am J. Physiol Lung Cell Mol Physiol.* - 286(2):L301-11 (2004).
29. De Boer, C.J., *et al.*, *Ann Oncol.* - 7(3):251-6 (1996).
30. Debieve, F., *et al.*, *Mol Hum Reprod.* - 6(8):743-9 (2000).
31. Dong, Z., *et al.*, *Invest Ophthalmol Vis Sci* - 42(13):3223- (2001).
32. Duchrow, M., *et al.*, *Cancer Invest.* - 19(6):588-96 (2001).
33. Dyer, J., *et al.*, *Equine Vet J.* - 34(4):349-58 (2002).

34. Egwuagu, C.E., *et al.*, J Immunol. - 168(7):3181-7 (2002).
35. El-Ghrably, I.A., *et al.*, Br J Ophthalmol. - 85(4):461-70 (2001).
36. Eleore, L., *et al.*, Neuroscience - 136(1):147-60 (2005).
37. Forsberg, H., *et al.*, Free Radic Res. - 24(6):451-9 (1996).
38. Freyschuss, B., *et al.*, Endocrinology - 133(4):1548-54 (1993).
39. Fu, K., *et al.*, Blood - 106(13):4315-21 (2005).
40. Fuchs, A.R., *et al.*, Biol Reprod. - 54(3):700-8 (1996).
41. Furuta, J., *et al.*, Melanoma Res. - 15(1):15-20 (2005).
42. Futcher, B., *et al.*, Mol Cell Biol., - 19(11):7357-68 (1999).
43. George, J., *et al.*, Biochem Pharmacol. - 49(7):873-81 (1995).
44. Giroux, M., *et al.*, J immunol. - 165(7):3985-91 (2000).
45. Gnatenko, D.V., *et al.*, Blood - 101(6):2285-93 (2003).
46. Godbout, R., *et al.*, J Biol Chem, - 273(33):21161-8 (1998).
47. Goldenberg, R.C., *et al.*, J Endocrinol. - 177(2):327-35 (2003).
48. Golebiowski, F., *et al.*, Acta Biochim Pol. - 50(4):1065-73 (2003).
49. Greenbaum, D., *et al.*, BioinformaticsI - 18(4):585-496 (2002).
50. Grem, J.L., *et al.*, Clin Cancer Res. - 7(4):999-1009 (2001).
51. Grenback, E., *et al.*, Regul Pept, - 117(2):127-39 (2004).
52. Gromova, I., *et al.*, Int J. Oncol. - 13(2):379-83 (1998).
53. Guo, Y., *et al.*, Zhinghua Jie He He Hu Xi Za Zhi - 25(6):337-40 (2002).
54. Habu, Y., *et al.*, Biochem Pharmacol - 69(6):993-9 (2005).
55. Hahn, M.E., *et al.*, Toxicol Appl Pharmacol. - 127(2):187-98 (1994).
56. Hähnel, E., *et al.*, Breast Cancer Research and Treatment - 24:71-74 (1992).
57. Hamilton, L.M., *et al.*, Clin Exp Allergy. - 33(2):233-40 (2003).

82. Lemstrom, K.B., *et al.*, Circulation - 105(21):2524-30 (2002).
83. Lewin, B., Genes VI Genes VI at 847-848 (1997).
84. Li, Y., *et al.*, Invest Ophthalmol Vis Sci. - 44(3):1299-304 (2003).
85. Li, Z.B., *et al.*, Biochem Biophys Res Commun. - 327(4):1163-9 (2005).
86. Lindberg, P., *et al.*, Arch Oral Biol. - 46(1):23-31 (2001).
87. Macabeo-Ong, M., *et al.*, 15(9):979-87 (2002).
88. Maruyama, H., *et al.*, Am J Pathol. - 155(3):815-22 (1999).
89. Meehan, T.P., *et al.*, J Mol Endocrinol. - 32(1):247-55 (2004).
90. Mendoza-Rodriguez, C.A., *et al.*, Mol Reprod Dev. 64(4):379-88 (2003).
91. Meoni, P., *et al.*, Brain Res Mol Res. - 54(1):13-23 (1998).
92. Meric, F., *et al.*, Molecular Cancer Therapeutics - 1:971-979 (2002).
93. Mezzano, S.A., *et al.*, Kiney Int. - 57(1):147-58 (2000).
94. Mingrone, G., *et al.*, Obes Res. - 11(5):632-40 (2003).
95. Miralles, C.P., *et al.*, Brain Res Mol Res. - 24(1-4):129-39 (1994).
96. Mizrachi, D., *et al.*, Biol Reprod. - 61(3):776-84 (1999).
97. Monaghan, P., *et al.*, J Gen Virol. - 86(Pt 10):2769-80 (2005).
98. Montuori, N., *et al.*, Int J Cancer - 105(3):353-60 (2003).
99. Munaut, C., *et al.*, Int J Cancer - 106(6):848-55 (2003).
100. Nie, Y., *et al.*, Carcinogenesis - 22(10):1615-23 (2001).
101. Nuciforo, P.G., *et al.*, Hum Pathol. - 34(7):639-45 (2003).
102. Oberringer, M., *et al.*, Biochem Biophys Res Commun. - 214(3):1009-14 (1995).
103. Orntoft, T.F., *et al.* Molecular & Cellular Proteomics – 1:37-45 (2002).
104. Pachmann, K., *et al.*, Br J. Haematol - 112(3):749-59 (2001).
105. Pairon, J.C., *et al.*, Am J Respir Cell Mol Biol. - 11(4):386-96 (1994).

58. Hassett, C., *et al.*, Arch Biochem Biophys. - 337(2):275-83 (1997).
59. Hirsch, F.R., *et al.*, Clinical Cancer Research - 7:5-22 (2001).
60. Holten-Andersen, M.N., *et al.*, Int J Cance, - 113(2):198-206 (2005).
61. Huang, Y.H., *et al.*, J Med Virol. - 70(4):610-6 (2003).
62. Huettner, P.C., *et al.*, Mod Pathol. - 5(3):250-6 (1992).
63. Hui, P., *et al.*, Leuk Lymphoma. 44(8):1385-94 (2003).
64. Husain, I., *et al.*, Cancer Res. - 54(2):539-46 (1994).
65. Ihmann, T., *et al.*, J Cancer Res Clin Oncol. - 130(12):749-56 (2004).
66. Ikegami, T., *et al.*, Am J Physiol Cell Physiol. - 281(4):C1396-402 (2001).
67. Jacquemin, E., *et al.*, Int J Biol. - 37(3):417-23 (1993).
68. Jaime, M., *et al.*, Hepatology - 35(5):1063-71 (2002).
69. Janssens, N., *et al.*, Tumour Biol. - 25(4):161-71 (2004).
70. Jungbluth, A.A., *et al.*, Int J. Cancer - 92(6):856-60 (2001).
71. Kalabis, G.M., *et al.*, Biol Reprod. - 73(4):591-7 (2005).
72. Kammaori, M., *et al.*, Int J Oncol. - 27(5):1257-63 (2005).
73. Khal, J., *et al.*, Br J Cancer - 93(7):774-80 (2005).
74. Khal, J., *et al.*, Int J. Biochem Cell Biol. - 37(10):2196-206 (2005).
75. Kogo, H., *et al.*, J Biol Chem. - 279(24):25574-81 (2004).
76. Kommoss, F., *et al.*, Acta Obstet Gynecol Scand Suppl. - 155:19-24 (1992).
77. Kumar, U., *et al.*, Breast Cancer Res. Treat. - 92(2):175-86 (2005).
78. Kuo, C.C., *et al.*, Proteomics - 5(4):894-906 (2005).
79. Landmark, B.F., *et al.*, J Reprod Fertil. - 99(2):323-34 (1993).
80. Lassmann, S., *et al.*, J Pathol. - 198(2):198-206 (2002).
81. Legrand, O., *et al.*, Br J, Haematol. - 94(1):23-33 (1996).

106. Papotti, M., *et al.*, Diagn Mol Pathol. - 9(1):47-57 (2000).
107. Papotti, M., *et al.*, Virchows Arch. - 440(5):461-75 (2002).
108. Paredes, J., *et al.*, Clin Cancer Res. - 11(16):5869-77 (2005).
109. Politis, I., *et al.*, J Dairy Sci. - 75(6):1423-9 (1992).
110. Preesman, A.H., *et al.*, J Invest Dermatol. - 99(5):587-93 (1992).
111. Pullig, O., *et al.*, Osteoarthritis Cartilage - 10(4):253-63 (2002).
112. Rey, C., *et al.*, Biochem Pharmacol. - 60(11):1636-46 (2000).
113. Rudlowski, C., *et al.*, Am J. Clin Pathol. - 120(5):691-8 (2003).
114. Sasaki, T., *et al.*, Exp Cell Res. - 275(2):189-99 (2002).
115. Sedelies, K.A., *et al.*, J Biol Chem. - 279(25):26581-7 (2004).
116. Shen, Y., *et al.*, Blood - 104(9):2936-9 (2004).
117. Shinohara, Y., *et al.*, Biochim Biophys Acta - 1368(1):129-36 (1998).
118. Silvers, A.L., *et al.*, Photochem Photobiol. - 75(3):302-10 (2002).
119. Song, L., *et al.*, Am J. Physiol. - 267(4 Pt 2):F546-57 (1994).
120. Spaziani, E.P., *et al.*, J Interferon Cytokine Res. - 18(12):1039-44 (1998).
121. Spika, I., *et al.*, Skin Pharmacol Appl Skin Physiol. - 16(3):143-50 (2003).
122. Splinter, P.L., *et al.*, J Biol Chem - 278(8):6268-74 (2003).
123. Stearns, M.E., *et al.*, Cancer Res. - 53(4):878-83 (1993).
124. Stein, R., *et al.*, J Urol. - 164(3Pt 2):1026-30 (2000).
125. Strickland, I., *et al.*, J Invest Dermatol. - 108(5):763-8 (1997).
126. Strutz, F., *et al.*, Kidney Int. - 57(4):1521-38 (2000).
127. Takahashi, K., *et al.*, J Biol. Chem. - 278(47):46654-60 (2003).
128. Takimoto, Y., *et al.*, Circulation - 105(4):490-6 (2002).
129. Telek, G., *et al.*, J Surg Res. - 96(1):56-67 (2001).

130. Timchenko, L., *et al.*, Semin Cell Biol. - 6(1):13-9 (1995).
131. Torronen, R., *et al.*, Chem Biol. Interact. - 83(2):107-19 (1992).
132. Ullmannova, V., *et al.*, Leuk Res. - 27(12):1115-23 (2003).
133. Van Beers, E.H., *et al.*, J Histochem Cytochem. - 46(2):231-40 (1998).
134. Van Der Wilt, C.L., *et al.*, Eur J Cancer - 39(5):691-7 (2003).
135. Waldherr, R., *et al.*, Pediatr Nephrol. - 7(4):471-8 (1993).
136. Walmer, D.K., *et al.*, Cancer Res. - 55(5):1168-75 (1995).
137. Wang, J., *et al.*, Cancer Res. - 54(2):560-4 (1994).
138. Wang, J., *et al.*, Urol Res. - 28(5):308-15 (2000).
139. Wang, L.G., *et al.*, Cancer Res. - 57(4):714-9 (1997).
140. Weterman, M.A., *et al.*, Cancer Res. - 53(24):6061-6 (1993).
141. Williams, E.T., *et al.*, J Pharmacol Exp Ther. 311(2):728-35 (2004).
142. Wojtaszek, P.A., *et al.*, Oncogene - 8(3):755-60 (1993).
143. Zhigang, Z., *et al.*, World Journal of Surgical Oncology - 2-13 (2004).
144. Zhong, W., *et al.*, Free Radic Biol Med. - 27(11-12):1334-45 (1999).
145. Xi, L., *et al.*, Zhonghua Fu Chan Ke Za Zhi - 40(6):407-10 (2005).
146. Nagaraja *et al.*, 2006, Oncogene 25: 2328-2338.
147. Waghray *et al.*, 2001, Proteomics 1: 1327-1338.
148. Sagynaliev *et al.*, 2005, Proteomics 5: 3066-3076.
149. Lilley *et al.*, "Proteomics" Mol. Biol. in Cellular Path. (2003) England: John Wiley & Sons, p 351-352.
150. Li *et al.*, 2006, Oncogene 25: 2628-2635.

Item 1 was submitted with Appellants' Response filed August 4, 2005, and was considered by the Examiner as indicated in the Final Office Action mailed September 7, 2005.

Items 2-3 and 5-7 were submitted with Appellants' Response filed June 18, 2004, and were considered by the Examiner as indicated in the Final Office Action mailed September 16, 2004.

Item 4 was submitted with Appellants' Response filed June 29, 2006, and was considered by the Examiner as indicated in the Final Office Action mailed September 27, 2006.

Items 8-9 were made of record by the Examiner in the Office Action mailed March 19, 2004.

Item 10 was made of record by the Examiner in the Final Office Action mailed September 16, 2004.

Items 11-13 were made of record by the Examiner in the Final Office Action mailed September 7, 2005.

Items 14-145 were submitted with Appellants' Response filed June 29, 2006, and were considered by the Examiner as indicated in the Final Office Action mailed September 27, 2006.

Items 146-149 were made of record by the Examiner in the Final Office Action mailed September 27, 2006.

Item 150 was made of record by the Examiner in the Final Office Action mailed October 13, 2006.

X. RELATED PROCEEDINGS APPENDIX

None.

SV 2281182 v1
6/13/07 1:47 PM (39780.2730)

PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

| | |
|---|---|
| In re Application of: Ashkenazi et al. | Group Art Unit: 1647 |
| Serial No.: 09/903,925 | Examiner: Fozia Hamid |
| Filed: July 11, 2001 | CERTIFICATE OF MAILING I hereby certify that this correspondence is being deposited with the United States Postal Service with sufficient postage as first class mail in an envelope addressed to: Assistant Commissioner of Patents, Washington, D.C. 20231 on |
| For: SECRETED AND TRANSMEMBRANE POLYPEPTIDES AND NUCLEIC ACIDS | Date |

DECLARATION OF AUDREY D. GODDARD, Ph.D UNDER 37 C.F.R. § 1.132

Assistant Commissioner of Patents
Washington, D.C. 20231

Sir:

I, Audrey D. Goddard, Ph.D. do hereby declare and say as follows:

1. I am a Senior Clinical Scientist at the Experimental Medicine/BioOncology, Medical Affairs Department of Genentech, Inc., South San Francisco, California 94080.
2. Between 1993 and 2001, I headed the DNA Sequencing Laboratory at the Molecular Biology Department of Genentech, Inc. During this time, my responsibilities included the identification and characterization of genes contributing to the oncogenic process, and determination of the chromosomal localization of novel genes.
3. My scientific Curriculum Vitae, including my list of publications, is attached to and forms part of this Declaration (Exhibit A).

Serial No.: *

Filed: *

4. I am familiar with a variety of techniques known in the art for detecting and quantifying the amplification of oncogenes in cancer, including the quantitative TaqMan PCR (i.e., "gene amplification") assay described in the above captioned patent application.

5. The TaqMan PCR assay is described, for example, in the following scientific publications: Higuchi *et al.*, Biotechnology 10:413-417 (1992) (Exhibit B); Livak *et al.*, PCR Methods Appl. 4:357-362 (1995) (Exhibit C) and Heid *et al.*, Genome Res. 6:986-994 (1996) (Exhibit D). Briefly, the assay is based on the principle that successful PCR yields a fluorescent signal due to Taq DNA polymerase-mediated exonuclease digestion of a fluorescently labeled oligonucleotide that is homologous to a sequence between two PCR primers. The extent of digestion depends directly on the amount of PCR, and can be quantified accurately by measuring the increment in fluorescence that results from decreased energy transfer. This is an extremely sensitive technique, which allows detection in the exponential phase of the PCR reaction and, as a result, leads to accurate determination of gene copy number.

6. The quantitative fluorescent TaqMan PCR assay has been extensively and successfully used to characterize genes involved in cancer development and progression. Amplification of protooncogenes has been studied in a variety of human tumors, and is widely considered as having etiological, diagnostic and prognostic significance. This use of the quantitative TaqMan PCR assay is exemplified by the following scientific publications: Pennica *et al.*, Proc. Natl. Acad. Sci. USA 95(25):14717-14722 (1998) (Exhibit E); Pitti *et al.*, Nature 396(6712):699-703 (1998) (Exhibit F) and Bieche *et al.*, Int. J. Cancer 78:661-666 (1998) (Exhibit G), the first two of which I am co-author. In particular, Pennica *et al.* have used the quantitative TaqMan PCR assay to study relative gene amplification of WISP and c-myc in various cell lines, colorectal tumors and normal mucosa. Pitti *et al.* studied the genomic amplification of a decoy receptor for Fas ligand in lung and colon cancer, using the quantitative TaqMan PCR assay. Bieche *et al.* used the assay to study gene amplification in breast cancer.

Serial No.: *

Filed: *

7. It is my personal experience that the quantitative TaqMan PCR technique is technically sensitive enough to detect at least a 2-fold increase in gene copy number relative to control. It is further my considered scientific opinion that an at least 2-fold increase in gene copy number in a tumor tissue sample relative to a normal (i.e., non-tumor) sample is significant and useful in that the detected increase in gene copy number in the tumor sample relative to the normal sample serves as a basis for using relative gene copy number as quantitated by the TaqMan PCR technique as a diagnostic marker for the presence or absence of tumor in a tissue sample of unknown pathology. Accordingly, a gene identified as being amplified at least 2-fold by the quantitative TaqMan PCR assay in a tumor sample relative to a normal sample is useful as a marker for the diagnosis of cancer, for monitoring cancer development and/or for measuring the efficacy of cancer therapy.

8. I declare further that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true. I declare that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Jan. 16, 2003

Date

Audrey D. Goddard

Audrey D. Goddard, Ph.D.

AUDREY D. GODDARD, Ph.D.

Genentech, Inc.
1 DNA Way
South San Francisco, CA, 94080
650.225.6429
goddarda@gene.com

110 Congo St.
San Francisco, CA, 94131
415.841.9154
415.819.2247 (mobile)
agoddard@pacbell.net

PROFESSIONAL EXPERIENCE

Genentech, Inc.
South San Francisco, CA

1993-present

2001 - present Senior Clinical Scientist
Experimental Medicine / BioOncology, Medical Affairs

Responsibilities:

- *Companion diagnostic oncology products*
- *Acquisition of clinical samples from Genentech's clinical trials for translational research*
- *Translational research using clinical specimen and data for drug development and diagnostics*
- *Member of Development Science Review Committee, Diagnostic Oversight Team, 21 CFR Part 11 Subteam*

Interests:

- *Ethical and legal implications of experiments with clinical specimens and data*
- *Application of pharmacogenomics in clinical trials*

1998 - 2001 Senior Scientist
Head of the DNA Sequencing Laboratory, Molecular Biology Department, Research

Responsibilities:

- *Management of a laboratory of up to nineteen –including postdoctoral fellow, associate scientist, senior research associate and research assistants/associate levels*
- *Management of a \$750K budget*
- *DNA sequencing core facility supporting a 350+ person research facility.*
- *DNA sequencing for high throughput gene discovery, - ESTs, cDNAs, and constructs*
- *Genomic sequence analysis and gene identification*
- *DNA sequence and primary protein analysis*

Research:

- *Chromosomal localization of novel genes*
- *Identification and characterization of genes contributing to the oncogenic process*
- *Identification and characterization of genes contributing to inflammatory diseases*
- *Design and development of schemes for high throughput genomic DNA sequence analysis*
- *Candidate gene prediction and evaluation*

1993 - 1998 Scientist

Head of the DNA Sequencing Laboratory, Molecular Biology Department, Research

Responsibilities

- *DNA sequencing core facility supporting a 350+ person research facility*
- *Assumed responsibility for a pre-existing team of five technicians and expanded the group into fifteen, introducing a level of middle management and additional areas of research*
- *Participated in the development of the basic plan for high throughput secreted protein discovery program – sequencing strategies, data analysis and tracking, database design*
- *High throughput EST and cDNA sequencing for new gene identification.*
- *Design and implementation of analysis tools required for high throughput gene identification.*
- *Chromosomal localization of genes encoding novel secreted proteins.*

Research:

- *Genomic sequence scanning for new gene discovery.*
- *Development of signal peptide selection methods.*
- *Evaluation of candidate disease genes.*
- *Growth hormone receptor gene SNPs in children with Idiopathic short stature*

**Imperial Cancer Research Fund
London, UK with Dr. Ellen Solomon**

1989-1992

6/89 – 12/92 Postdoctoral Fellow

- *Cloning and characterization of the genes fused at the acute promyelocytic leukemia translocation breakpoints on chromosomes 17 and 15.*
- *Prepared a successfully funded European Union multi-center grant application*

**McMaster University
Hamilton, Ontario, Canada with Dr. G. D. Sweeney**

1983

5/83 – 8/83: NSERC Summer Student

- *In vitro* metabolism of β -naphthoflavone in C57Bl/6J and DBA mice

EDUCATION

Ph.D.

"Phenotypic and genotypic effects of mutations in the human retinoblastoma gene."

Supervisor: Dr. R. A. Phillips

University of Toronto
Toronto, Ontario, Canada.
Department of Medical
Biophysics.

1989

Honours B.Sc

"The *in vitro* metabolism of the cytochrome P-448 inducer β -naphthoflavone in C57BL/6J mice."

Supervisor: Dr. G. D. Sweeney

McMaster University,
Hamilton, Ontario, Canada.
Department of Biochemistry

1983

ACADEMIC AWARDS

| | |
|---|-----------|
| Imperial Cancer Research Fund Postdoctoral Fellowship | 1989-1992 |
| Medical Research Council Studentship | 1983-1988 |
| NSERC Undergraduate Summer Research Award | 1983 |
| Society of Chemical Industry Merit Award (Hons. Biochem.) | 1983 |
| Dr. Harry Lyman Hooker Scholarship | 1981-1983 |
| J.L.W. Gill Scholarship | 1981-1982 |
| Business and Professional Women's Club Scholarship | 1980-1981 |
| Wyerhauser Foundation Scholarship | 1979-1980 |

INVITED PRESENTATIONS

Genentech's gene discovery pipeline: High throughput identification, cloning and characterization of novel genes. Functional Genomics: From Genome to Function, Litchfield Park, AZ, USA. October 2000

High throughput identification, cloning and characterization of novel genes. G2K:Back to Science, Advances in Genome Biology and Technology I. Marco Island, FL, USA. February 2000

Quality control in DNA Sequencing: The use of Phred and Phrap. Bay Area Sequencing Users Meeting, Berkeley, CA, USA. April 1999

High throughput secreted protein identification and cloning. Tenth International Genome Sequencing and Analysis Conference, Miami, FL, USA. September 1998

The evolution of DNA sequencing: The Genentech perspective. Bay Area Sequencing Users Meeting, Berkeley, CA, USA. May 1998

Partial Growth Hormone Insensitivity: The role of GH-receptor mutations in Idiopathic Short Stature. Tenth Annual National Cooperative Growth Study Investigators Meeting, San Francisco, CA, USA. October, 1996

Growth hormone (GH) receptor defects are present in selected children with non-GH-deficient short stature: A molecular basis for partial GH-insensitivity. 76th Annual Meeting of The Endocrine Society, Anaheim, CA, USA. June 1994

A previously uncharacterized gene, myl, is fused to the retinoic acid receptor alpha gene in acute promyelocytic leukemia. XV International Association for Comparative Research on Leukemia and Related Disease, Padua, Italy. October 1991

PATENTS

Goddard A, Godowski PJ, Gurney AL. NL2 Tie ligand homologue polypeptide. Patent Number: 6,455,496. Date of Patent: Sept. 24, 2002.

Goddard A, Godowski PJ and Gurney AL. NL3 Tie ligand homologue nucleic acids. Patent Number: 6,426,218. Date of Patent: July 30, 2002.

Godowski P, Gurney A, Hillan KJ, Botstein D, **Goddard A**, Roy M, Ferrara N, Tumas D, Schwall R. NL4 Tie ligand homologue nucleic acid. Patent Number: 6,4137,770. Date of Patent: July 2, 2002.

Ashkenazi A, Fong S, **Goddard A**, Gurney AL, Napier MA, Tumas D, Wood WI. Nucleic acid encoding A-33 related antigen poly peptides. Patent Number: 6,410,708. Date of Patent: Jun. 25, 2002.

Botstein DA, Cohen RL, **Goddard AD**, Gurney AL, Hillan KJ, Lawrence DA, Levine AJ, Pennica D, Roy MA and Wood WI. WISP polypeptides and nucleic acids encoding same. Patent Number: 6,387,657. Date of Patent: May 14, 2002.

Goddard A, Godowski PJ and Gurney AL. Tie ligands. Patent Number: 6,372,491. Date of Patent: April 16, 2002.

Godowski PJ, Gurney AL, **Goddard A** and Hillan K. TIE ligand homologue antibody. Patent Number: 6,350,450. Date of Patent: Feb. 26, 2002.

Fong S, Ferrara N, **Goddard A**, Godowski PJ, Gurney AL, Hillan K and Williams PM. Tie receptor tyrosine kinase ligand homologues. Patent Number: 6,348,351. Date of Patent: Feb. 19, 2002.

Goddard A, Godowski PJ and Gurney AL. Ligand homologues. Patent Number: 6,348,350. Date of Patent: Feb. 19, 2002.

Attie KM, Carlsson LMS, Gesundheit N and **Goddard A**. Treatment of partial growth hormone insensitivity syndrome. Patent Number: 6,207,640. Date of Patent: March 27, 2001.

Fong S, Ferrara N, **Goddard A**, Godowski PJ, Gurney AL, Hillan K and Williams PM. Nucleic acids encoding NL-3. Patent Number: 6,074,873. Date of Patent: June 13, 2000

Attie K, Carlsson LMS, Gesundheit N and **Goddard A**. Treatment of partial growth hormone insensitivity syndrome. Patent Number: 5,824,642. Date of Patent: October 20, 1998

Attie K, Carlsson LMS, Gesundheit N and **Goddard A**. Treatment of partial growth hormone insensitivity syndrome. Patent Number: 5,646,113. Date of Patent: July 8, 1997

Multiple additional provisional applications filed

PUBLICATIONS

Seshasayee D, Dowd P, Gu Q, Erickson S, **Goddard AD** Comparative sequence analysis of the *HER2* locus in mouse and man. Manuscript in preparation.

Abuzzahab MJ, **Goddard A**, Grigorescu F, Lautier C, Smith RJ and Chernausk SD. Human IGF-1 receptor mutations resulting in pre- and post-natal growth retardation. Manuscript in preparation.

Aggarwal S, Xie, M-H, Foster J, Frantz G, Stinson J, Corpuz RT, Simmons L, Hillan K, Yansura DG, Vandlen RL, **Goddard AD** and Gurney AL. FHFR, a novel receptor for the fibroblast growth factors. Manuscript submitted.

Adams SH, Chui C, Schilbach SL, Yu XX, **Goddard AD**, Grimaldi JC, Lee J, Dowd P, Colman S., Lewin DA. (2001) BFIT, a unique acyl-CoA thioesterase induced in thermogenic brown adipose tissue: Cloning, organization of the human gene, and assessment of a potential link to obesity. *Biochemical Journal* **360**: 135-142.

Lee J, Ho WH, Maruoka M, Corpuz RT, Baldwin DT, Foster JS, **Goddard AD**, Yansura DG, Vandlen RL, Wood WI, Gurney AL. (2001) IL-17E, a novel proinflammatory ligand for the IL-17 receptor homolog IL-17Rh1. *Journal of Biological Chemistry* **276**(2): 1660-1664.

Xie M-H, Aggarwal S, Ho W-H, Foster J, Zhang Z, Stinson J, Wood WI, **Goddard AD** and Gurney AL. (2000) Interleukin (IL)-22, a novel human cytokine that signals through the interferon-receptor related proteins CRF2-4 and IL-22R. *Journal of Biological Chemistry* **275**: 31335-31339.

Weiss GA, Watanabe CK, Zhong A, **Goddard A** and Sidhu SS. (2000) Rapid mapping of protein functional epitopes by combinatorial alanine scanning. *Proc. Natl. Acad. Sci. USA* **97**: 8950-8954.

Guo S, Yamaguchi Y, Schilbach S, Wada T., Lee J, **Goddard A**, French D, Handa H, Rosenthal A. (2000) A regulator of transcriptional elongation controls vertebrate neuronal development. *Nature* **408**: 366-369.

Yan M, Wang L-C, Hymowitz SG, Schilbach S, Lee J, **Goddard A**, de Vos AM, Gao WQ, Dixit VM. (2000) Two-amino acid molecular switch in an epithelial morphogen that regulates binding to two distinct receptors. *Science* **290**: 523-527.

Sehl PD, Tai JTN, Hillan KJ, Brown LA, **Goddard A**, Yang R, Jin H and Lowe DG. (2000) Application of cDNA microarrays in determining molecular phenotype in cardiac growth, development, and response to injury. *Circulation* **101**: 1990-1999.

Guo S, Brush J, Teraoka H, **Goddard A**, Wilson SW, Mullins MC and Rosenthal A. (1999) Development of noradrenergic neurons in the zebrafish hindbrain requires BMP, FGF8, and the homeodomain protein soulless/Phox2A. *Neuron* **24**: 555-566.

Stone D, Murone, M, Luoh, S, Ye W, Armanini P, Gurney A, Phillips HS, Brush, J, **Goddard A**, de Sauvage FJ and Rosenthal A. (1999) Characterization of the human suppressor of fused; a negative regulator of the zinc-finger transcription factor Gli. *J. Cell Sci.* **112**: 4437-4448.

Xie M-H, Holcomb I, Deuel B, Dowd P, Huang A, Vagts A, Foster J, Liang J, Brush J, Gu Q, Hillan K, **Goddard A** and Gurney, A.L. (1999) FGF-19, a novel fibroblast growth factor with unique specificity for FGFR4. *Cytokine* **11**: 729-735.

Yan M, Lee J, Schilbach S, **Goddard A** and Dixit V. (1999) mE10, a novel caspase recruitment domain-containing proapoptotic molecule. *J. Biol. Chem.* **274**(15): 10287-10292.

Gurney AL, Marsters SA, Huang RM, Pitti RM, Mark DT, Baldwin DT, Gray AM, Dowd P, Brush J, Heldens S, Schow P, **Goddard AD**, Wood WI, Baker KP, Godowski PJ and Ashkenazi A. (1999) Identification of a new member of the tumor necrosis factor family and its receptor, a human ortholog of mouse GITR. *Current Biology* **9**(4): 215-218.

Ridgway JBB, Ng E, Kern JA, Lee J, Brush J, **Goddard A** and Carter P. (1999) Identification of a human anti-CD55 single-chain Fv by subtractive panning of a phage library using tumor and nontumor cell lines. *Cancer Research* **59**: 2718-2723.

Pitti RM, Marsters SA, Lawrence DA, Roy M, Kischkel FC, Dowd P, Huang A, Donahue CJ, Sherwood SW, Baldwin DT, Godowski PJ, Wood WI, Gurney AL, Hillan KJ, Cohen RL, **Goddard AD**, Botstein D and Ashkenazi A. (1998) Genomic amplification of a decoy receptor for Fas ligand in lung and colon cancer. *Nature* **396**(6712): 699-703.

Pennica D, Swanson TA, Welsh JW, Roy MA, Lawrence DA, Lee J, Brush J, Taneyhill LA, Deuel B, Lew M, Watanabe C, Cohen RL, Melhem MF, Finley GG, Quirke P, **Goddard AD**, Hillan KJ, Gurney AL, Botstein D and Levine AJ. (1998) WISP genes are members of the connective tissue growth factor family that are up-regulated in wnt-1-transformed cells and aberrantly expressed in human colon tumors. *Proc. Natl. Acad. Sci. USA.* **95**(25): 14717-14722.

Yang RB, Mark MR, Gray A, Huang A, Xie MH, Zhang M, **Goddard A**, Wood WI, Gurney AL and Godowski PJ. (1998) Toll-like receptor-2 mediates lipopolysaccharide-induced cellular signalling. *Nature* **395**(6699): 284-288.

Merchant AM, Zhu Z, Yuan JQ, **Goddard A**, Adams CW, Presta LG and Carter P. (1998) An efficient route to human bispecific IgG. *Nature Biotechnology* **16**(7): 677-681.

Marsters SA, Sheridan JP, Pitti RM, Brush J, **Goddard A** and Ashkenazi A. (1998) Identification of a ligand for the death-domain-containing receptor Apo3. *Current Biology* **8**(9): 525-528.

Xie J, Murone M, Luoh SM, Ryan A, Gu Q, Zhang C, Bonifas JM, Lam CW, Hynes M, **Goddard A**, Rosenthal A, Epstein EH Jr. and de Sauvage FJ. (1998) Activating Smoothed mutations in sporadic basal-cell carcinoma. *Nature.* **391**(6662): 90-92.

Marsters SA, Sheridan JP, Pitti RM, Huang A, Skubatch M, Baldwin D, Yuan J, Gurney A, **Goddard AD**, Godowski P and Ashkenazi A. (1997) A novel receptor for Apo2L/TRAIL contains a truncated death domain. *Current Biology.* **7**(12): 1003-1006.

Hynes M, Stone DM, Dowd M, Pitts-Meek S, **Goddard A**, Gurney A and Rosenthal A. (1997) Control of cell pattern in the neural tube by the zinc finger transcription factor *Gli-1*. *Neuron* **19**: 15-26.

Sheridan JP, Marsters SA, Pitti RM, Gurney A., Skubatch M, Baldwin D, Ramakrishnan L, Gray CL, Baker K, Wood WI, **Goddard AD**, Godowski P, and Ashkenazi A. (1997) Control of TRAIL-Induced Apoptosis by a Family of Signaling and Decoy Receptors. *Science* **277** (5327): 818-821.

Goddard AD, Dowd P, Chernauek S, Geffner M, Gertner J, Hintz R, Hopwood N, Kaplan S, Plotnick L, Rogol A, Rosenfield R, Saenger P, Mauras N, Hershkopf R, Angulo M and Attie, K. (1997) Partial growth hormone insensitivity: The role of growth hormone receptor mutations in idiopathic short stature. *J. Pediatr.* **131**: S51-55.

Klein RD, Sherman D, Ho WH, Stone D, Bennett GL, Moffat B, Vandlen R, Simmons L, Gu Q, Hongo JA, Devaux B, Poulsen K, Armanini M, Nozaki C, Asai N, **Goddard A**, Phillips H, Henderson CE, Takahashi M and Rosenthal A. (1997) A GPI-linked protein that interacts with Ret to form a candidate neurturin receptor. *Nature*. **387**(6634): 717-21.

Stone DM, Hynes M, Armanini M, Swanson TA, Gu Q, Johnson RL, Scott MP, Pennica D, **Goddard A**, Phillips H, Noll M, Hooper JE, de Sauvage F and Rosenthal A. (1996) The tumour-suppressor gene patched encodes a candidate receptor for Sonic hedgehog. *Nature* **384**(6605): 129-34.

Marsters SA, Sheridan JP, Donahue CJ, Pitti RM, Gray CL, **Goddard AD**, Bauer KD and Ashkenazi A. (1996) Apo-3, a new member of the tumor necrosis factor receptor family, contains a death domain and activates apoptosis and NF-kappa β . *Current Biology* **6**(12): 1669-76.

Rothe M, Xiong J, Shu HB, Williamson K, **Goddard A** and Goeddel DV. (1996) I-TRAF is a novel TRAF-interacting protein that regulates TRAF-mediated signal transduction. *Proc. Natl. Acad. Sci. USA* **93**: 8241-8246.

Yang M, Luoh SM, **Goddard A**, Reilly D, Henzel W and Bass S. (1996) The bglX gene located at 47.8 min on the Escherichia coli chromosome encodes a periplasmic beta-glucosidase. *Microbiology* **142**: 1659-65.

Goddard AD and Black DM. (1996) Familial Cancer in Molecular Endocrinology of Cancer. Waxman, J. Ed. Cambridge University Press, Cambridge UK, pp.187-215.

Treanor JJS, Goodman L, de Sauvage F, Stone DM, Poulson KT, Beck CD, Gray C, Armanini MP, Pollocks RA, Hefti F, Phillips HS, **Goddard A**, Moore MW, Buj-Bello A, Davis AM, Asai N, Takahashi M, Vandlen R, Henderson CE and Rosenthal A. (1996) Characterization of a receptor for GDNF. *Nature* **382**: 80-83.

Klein RD, Gu Q, **Goddard A** and Rosenthal A. (1996) Selection for genes encoding secreted proteins and receptors. *Proc. Natl. Acad. Sci. USA* **93**: 7108-7113.

Winslow JW, Moran P, Valverde J, Shih A, Yuan JQ, Wong SC, Tsai SP, **Goddard A**, Henzel WJ, Hefti F and Caras I. (1995) Cloning of AL-1, a ligand for an Eph-related tyrosine kinase receptor involved in axon bundle formation. *Neuron* **14**: 973-981.

Bennett BD, Zeigler FC, Gu Q, Fendly B, **Goddard AD**, Gillett N and Matthews W. (1995) Molecular cloning of a ligand for the EPH-related receptor protein-tyrosine kinase Htk. *Proc. Natl. Acad. Sci. USA* **92**: 1866-1870.

Huang X, Yuang J, **Goddard A**, Foulis A, James RF, Lernmark A, Pujol-Borrell R, Rabinovitch A, Somoza N and Stewart TA. (1995) Interferon expression in the pancreases of patients with type I diabetes. *Diabetes* **44**: 658-664.

Goddard AD, Yuan JQ, Fairbairn L, Dexter M, Borrow J, Kozak C and Solomon E. (1995) Cloning of the murine homolog of the leukemia-associated PML gene. *Mammalian Genome* **6**: 732-737.

Goddard AD, Covello R, Luoh SM, Clackson T, Attie KM, Gesundheit N, Rundle AC, Wells JA, Carlsson LMTI and The Growth Hormone Insensitivity Study Group. (1995) Mutations of the growth hormone receptor in children with idiopathic short stature. *N. Engl. J. Med.* **333**: 1093-1098.

Kuo SS, Moran P, Gripp J, Armanini M, Phillips HS, **Goddard A** and Caras IW. (1994) Identification and characterization of Batk, a predominantly brain-specific non-receptor protein tyrosine kinase related to Csk. *J. Neurosci. Res.* **38**: 705-715.

Mark MR, Scadden DT, Wang Z, Gu Q, **Goddard A** and Godowski PJ. (1994) Rse, a novel receptor-type tyrosine kinase with homology to Axl/Ufo, is expressed at high levels in the brain. *Journal of Biological Chemistry* **269**: 10720-10728.

Borrow J, Shipley J, Howe K, Kiely F, **Goddard A**, Sheer D, Srivastava A, Antony AC, Fioretos T, Mitelman F and Solomon E. (1994) Molecular analysis of simple variant translocations in acute promyelocytic leukemia. *Genes Chromosomes Cancer* **9**: 234-243.

Goddard AD and Solomon E. (1993) Genetics of Cancer. *Adv. Hum. Genet.* **21**: 321-376.

Borrow J, **Goddard AD**, Gibbons B, Katz F, Swirsky D, Fioretos T, Dube I, Winfield DA, Kingston J, Hagemeijer A, Rees JKH, Lister AT and Solomon E. (1992) Diagnosis of acute promyelocytic leukemia by RT-PCR: Detection of *PML-RARA* and *RARA-PML* fusion transcripts. *Br. J. Haematol.* **82**: 529-540.

Goddard AD, Borrow J and Solomon E. (1992) A previously uncharacterized gene, PML, is fused to the retinoic acid receptor alpha gene in acute promyelocytic leukemia. *Leukemia* **6 Suppl 3**: 117S-119S.

Zhu X, Dunn JM, **Goddard AD**, Squire JA, Becker A, Phillips RA and Gallie BL. (1992) Mechanisms of loss of heterozygosity in retinoblastoma. *Cytogenet. Cell. Genet.* **59**: 248-252.

Foulkes W, **Goddard A.** and Patel K. (1991) Retinoblastoma linked with Seascale [letter]. *British Med. J.* **302**: 409.

Goddard AD, Borrow J, Freemont PS and Solomon E. (1991) Characterization of a novel zinc finger gene disrupted by the t(15;17) in acute promyelocytic leukemia. *Science* **254**: 1371-1374.

Solomon E, Borrow J and **Goddard AD**. (1991) Chromosomal aberrations in cancer. *Science* **254**: 1153-1160.

Pajunen L, Jones TA, **Goddard A**, Sheer D, Solomon E, Pihlajaniemi T and Kivirikko KI. (1991) Regional assignment of the human gene coding for a multifunctional peptide (P4HB) acting as the β -subunit of prolyl-4-hydroxylase and the enzyme protein disulfide isomerase to 17q25. *Cytogenet. Cell. Genet.* **56**: 165-168.

Borrow J, Black DM, **Goddard AD**, Yagle MK, Frischauf A.-M and Solomon E. (1991) Construction and regional localization of a *NotI* linking library from human chromosome 17q. *Genomics* **10**: 477-480.

Borrow J, **Goddard AD**, Sheer D and Solomon E. (1990) Molecular analysis of acute promyelocytic leukemia breakpoint cluster region on chromosome 17. *Science* **249**: 1577-1580.

Myers JC, Jones TA, Pohjolainen E-R, Kadri AS, **Goddard AD**, Sheer D, Solomon E and Pihlajaniemi T. (1990) Molecular cloning of 5(IV) collagen and assignment of the gene to the region of the region of the X-chromosome containing the Alport Syndrome locus. *Am. J. Hum. Genet.* **46**: 1024-1033.

Gallie BL, Squire JA, **Goddard A**, Dunn JM, Canton M, Hinton D, Zhu X and Phillips RA. (1990) Mechanisms of oncogenesis in retinoblastoma. *Lab. Invest.* **62**: 394-408.

Goddard AD, Phillips RA, Greger V, Passarge E, Hopping W, Gallie BL and Horsthemke B. (1990) Use of the RB1 cDNA as a diagnostic probe in retinoblastoma families. *Clinical Genetics* **37**: 117-126.

Zhu XP, Dunn JM, Phillips RA, **Goddard AD**, Paton KE, Becker A and Gallie BL. (1989) Germline, but not somatic, mutations of the RB1 gene preferentially involve the paternal allele. *Nature* **340**: 312-314.

Gallie BL, Dunn JM, **Goddard A**, Becker A and Phillips RA. (1988) Identification of mutations in the putative retinoblastoma gene. In Molecular Biology of The Eye: Genes, Vision and Ocular Disease. UCLA Symposia on Molecular and Cellular Biology, New Series, Volume 88. J. Piatigorsky, T. Shinohara and P.S. Zelenka, Eds. Alan R. Liss, Inc., New York, 1988, pp. 427-436.

Goddard AD, Balakier H, Canton M, Dunn J, Squire J, Reyes E, Becker A, Phillips RA and Gallie BL. (1988) Infrequent genomic rearrangement and normal expression of the putative RB1 gene in retinoblastoma tumors. *Mol. Cell. Biol.* **8**: 2082-2088.

Squire J, Dunn J, **Goddard A**, Hoffman T, Musarella M, Willard HF, Becker AJ, Gallie BL and Phillips RA. (1986) Cloning of the esterase D gene: A polymorphic gene probe closely linked to the retinoblastoma locus on chromosome 13. *Proc. Natl. Acad. Sci. USA* **83**: 6573-6577.

Squire J, **Goddard AD**, Canton M, Becker A, Phillips RA and Gallie BL (1986) Tumour induction by the retinoblastoma mutation is independent of N-myc expression. *Nature* **322**: 555-557.

Goddard AD, Heddle JA, Gallie BL and Phillips RA. (1985) Radiation sensitivity of fibroblasts of bilateral retinoblastoma patients as determined by micronucleus induction *in vitro*. *Mutation Research* **152**: 31-38.

RESEARCH

SIMULTANEOUS AMPLIFICATION AND DETECTION OF SPECIFIC DNA SEQUENCES

Russell Higuchi*, Gavin Dollinger¹, P. Sean Walsh and Robert GriffithRoche Molecular Systems, Inc., 1400 53rd St., Emeryville, CA 94608. ¹Chiron Corporation, 1400 53rd St., Emeryville, CA 94608. *Corresponding author.

We have enhanced the polymerase chain reaction (PCR) such that specific DNA sequences can be detected without opening the reaction tube. This enhancement requires the addition of ethidium bromide (EtBr) to a PCR. Since the fluorescence of EtBr increases in the presence of double-stranded (ds) DNA an increase in fluorescence in such a PCR indicates a positive amplification, which can be easily monitored externally. In fact, amplification can be continuously monitored in order to follow its progress. The ability to simultaneously amplify specific DNA sequences and detect the product of the amplification both simplifies and improves PCR and may facilitate its automation and more widespread use in the clinic or in other situations requiring high sample throughput.

Although the potential benefits of PCR¹ to clinical diagnostics are well known^{2,3}, it is still not widely used in this setting, even though it is four years since thermostable DNA polymerases⁴ made PCR practical. Some of the reasons for its slow acceptance are high cost, lack of automation of pre- and post-PCR processing steps, and false positive results from carryover-contamination. The first two points are related in that labor is the largest contributor to cost at the present stage of PCR development. Most current assays require some form of "downstream" processing once thermocycling is done in order to determine whether the target DNA sequence was present and has amplified. These include DNA hybridization^{5,6}, gel electrophoresis with or without use of restriction digestion^{7,8}, HPLC⁹, or capillary electrophoresis¹⁰. These methods are labor-intensive, have low throughput, and are difficult to automate. The third point is also closely related to downstream processing. The handling of the PCR product in these downstream processes increases the chances that amplified DNA will spread through the typing lab, resulting in a risk of

"carryover" false positives in subsequent testing¹¹.

These downstream processing steps would be eliminated if specific amplification and detection of amplified DNA took place simultaneously within an unopened reaction vessel. Assays in which such different processes take place without the need to separate reaction components have been termed "homogeneous". No truly homogeneous PCR assay has been demonstrated to date, although progress towards this end has been reported. Chehab, et al.¹², developed a PCR product detection scheme using fluorescent primers that resulted in a fluorescent PCR product. Allele-specific primers, each with different fluorescent tags, were used to indicate the genotype of the DNA. However, the unincorporated primers must still be removed in a downstream process in order to visualize the result. Recently, Holland, et al.¹³, developed an assay in which the endogenous 5' exonuclease assay of *Taq* DNA polymerase was exploited to cleave a labeled oligonucleotide probe. The probe would only cleave if PCR amplification had produced its complementary sequence. In order to detect the cleavage products, however, a subsequent process is again needed.

We have developed a truly homogeneous assay for PCR and PCR product detection based upon the greatly increased fluorescence that ethidium bromide and other DNA binding dyes exhibit when they are bound to dsDNA¹⁴⁻¹⁶. As outlined in Figure 1, a prototypic PCR

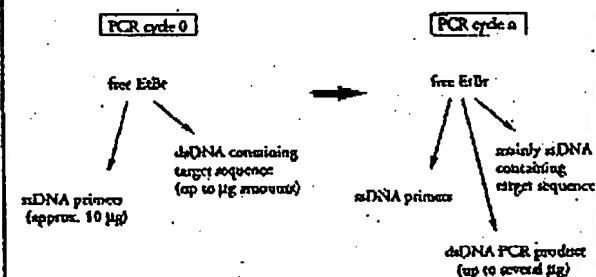


FIGURE 1 Principle of simultaneous amplification and detection of PCR product. The components of a PCR containing EtBr that are fluorescent are listed—EtBr itself, EtBr bound to either ssDNA or dsDNA. There is a large fluorescence enhancement when EtBr is bound to DNA and binding is greatly enhanced when DNA is double-stranded. After sufficient (n) cycles of PCR, the net increase in dsDNA results in additional EtBr binding, and a net increase in total fluorescence.

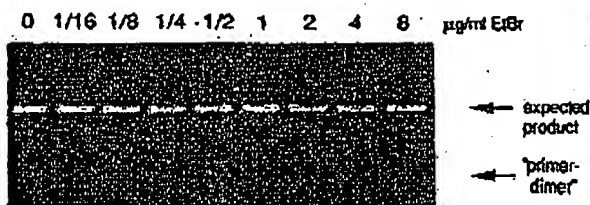


FIGURE 2 Gel electrophoresis of PCR amplification products of the human nuclear gene, HLA DQα, made in the presence of increasing amounts of EtBr (up to 8 μg/ml). The presence of EtBr has no obvious effect on the yield or specificity of amplification.

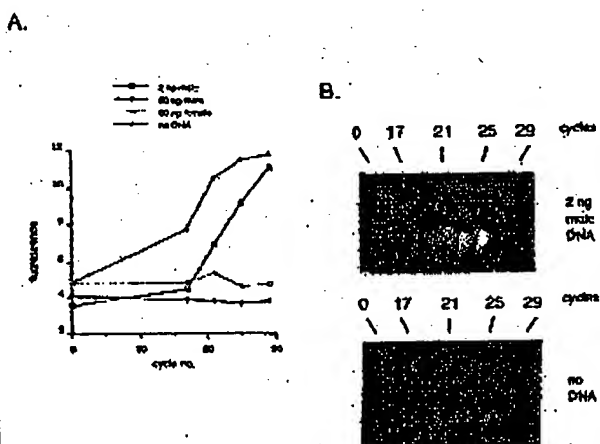


FIGURE 3 (A) Fluorescence measurements from PCR tubes that contain 0.5 μg/ml EtBr and that are specific for Y-chromosome repeat sequences. Five replicate PCRs were begun containing each of the DNAs specified. At each indicated cycle, one of the five replicate PCRs for each DNA was removed from thermocycling and its fluorescence measured. Units of fluorescence are arbitrary. (B) UV photograph of PCR tubes (0.5 ml Eppendorf-style, polypropylene micro-centrifuge tubes) containing reactions, those starting from 2 ng male DNA and control reactions without any DNA, from (A).

begins with primers that are single-stranded DNA (ssDNA), dNTPs, and DNA polymerase. An amount of dsDNA containing the target sequence (target DNA) is also typically present. This amount can vary, depending on the application, from single-cell amounts of DNA¹⁷ to micrograms per PCR¹⁸. If EtBr is present, the reagents that will fluoresce, in order of increasing fluorescence, are free EtBr itself, and EtBr bound to the single-stranded DNA primers and to the double-stranded target DNA (by its intercalation between the stacked bases of the DNA double-helix). After the first denaturation cycle, target DNA will be largely single-stranded. After a PCR is completed, the most significant change is the increase in the amount of dsDNA (the PCR product itself) of up to several micrograms. Formerly free EtBr is bound to the additional dsDNA, resulting in an increase in fluorescence. There is also some decrease in the amount of ssDNA primer, but because the binding of EtBr to ssDNA is much less than to dsDNA, the effect of this change on the total fluorescence of the sample is small. The fluorescence increase can be measured by directing excitation illumination through the walls of the amplification vessel

before and after, or even continuously during, thermocycling.

RESULTS

PCR in the presence of EtBr. In order to assess the effect of EtBr in PCR, amplifications of the human HLA DQα gene¹⁹ were performed with the dye present at concentrations from 0.06 to 8.0 μg/ml (a typical concentration of EtBr used in staining of nucleic acids following gel electrophoresis is 0.5 μg/ml). As shown in Figure 2, gel electrophoresis revealed little or no difference in the yield or quality of the amplification product whether EtBr was absent or present at any of these concentrations, indicating that EtBr does not inhibit PCR.

Detection of human Y-chromosome specific sequences. Sequence-specific, fluorescence enhancement of EtBr as a result of PCR was demonstrated in a series of amplifications containing 0.5 μg/ml EtBr and primers specific to repeat DNA sequences found on the human Y-chromosome²⁰. These PCRs initially contained either 60 ng male, 60 ng female, 2 ng male human or no DNA. Five replicate PCRs were begun for each DNA. After 0, 17, 21, 24 and 29 cycles of thermocycling, a PCR for each DNA was removed from the thermocycler, and its fluorescence measured in a spectrofluorometer and plotted vs. amplification cycle number (Fig. 3A). The shape of this curve reflects the fact that by the time an increase in fluorescence can be detected, the increase in DNA is becoming linear and not exponential with cycle number. As shown, the fluorescence increased about three-fold over the background fluorescence for the PCRs containing human male DNA, but did not significantly increase for negative control PCRs, which contained either no DNA or human female DNA. The more male DNA present to begin with—60 ng versus 2 ng—the fewer cycles were needed to give a detectable increase in fluorescence. Gel electrophoresis on the products of these amplifications showed that DNA fragments of the expected size were made in the male DNA containing reactions and that little DNA synthesis took place in the control samples.

In addition, the increase in fluorescence was visualized by simply laying the completed, unopened PCR tubes on a UV transilluminator and photographing them through a red filter. This is shown in figure 3B for the reactions that began with 2 ng male DNA and those with no DNA.

Detection of specific alleles of the human β-globin gene. In order to demonstrate that this approach has adequate specificity to allow genetic screening, a detection of the sickle-cell anemia mutation was performed. Figure 4 shows the fluorescence from completed amplifications containing EtBr (0.5 μg/ml) as detected by photography of the reaction tubes on a UV transilluminator. These reactions were performed using primers specific for either the wild-type or sickle-cell mutation of the human β-globin gene²¹. The specificity for each allele is imparted by placing the sickle-mutation site at the terminal 3' nucleotide of one primer. By using an appropriate primer annealing temperature, primer extension—and thus amplification—can take place only if the 3' nucleotide of the primer is complementary to the β-globin allele present^{21,22}.

Each pair of amplifications shown in Figure 4 consists of a reaction with either the wild-type allele specific (left tube) or sickle-allele specific (right tube) primers. Three different DNAs were typed: DNA from a homozygous, wild-type β-globin individual (AA); from a heterozygous sickle β-globin individual (AS); and from a homozygous sickle β-globin individual (SS). Each DNA (50 ng genomic DNA to start each PCR) was analyzed in triplicate (3 pairs

of reactions each). The DNA type was reflected in the relative fluorescence intensities in each pair of completed amplifications. There was a significant increase in fluorescence only where a β -globin allele DNA matched the primer set. When measured on a spectrofluorometer (data not shown), this fluorescence was about three times that present in a PCR where both β -globin alleles were mismatched to the primer set. Gel electrophoresis (not shown) established that this increase in fluorescence was due to the synthesis of nearly a microgram of a DNA fragment of the expected size for β -globin. There was little synthesis of dsDNA in reactions in which the allele-specific primer was mismatched to both alleles.

Continuous monitoring of a PCR. Using a fiber optic device it is possible to direct excitation illumination from a spectrofluorometer to a PCR undergoing thermocycling and to return its fluorescence to the spectrofluorometer. The fluorescence readout of such an arrangement, directed at an EtBr-containing amplification of Y-chromosome specific sequences from 25 ng of human male DNA, is shown in Figure 5. The readout from a control PCR with no target DNA is also shown. Thirty cycles of PCR were monitored for each.

The fluorescence trace as a function of time clearly shows the effect of the thermocycling. Fluorescence intensity rises and falls inversely with temperature. The fluorescence intensity is minimum at the denaturation temperature (94°C) and maximum at the annealing/extension temperature (50°C). In the negative-control PCR, these fluorescence maxima and minima do not change significantly over the thirty thermocycles, indicating that there is little dsDNA synthesis without the appropriate target DNA, and there is little if any bleaching of EtBr during the continuous illumination of the sample.

In the PCR containing male DNA, the fluorescence maxima at the annealing/extension temperature begin to increase at about 4000 seconds of thermocycling, and continue to increase with time, indicating that dsDNA is being produced at a detectable level. Note that the fluorescence minima at the denaturation temperature do not significantly increase, presumably because at this temperature there is no dsDNA for EtBr to bind. Thus the course of the amplification is followed by tracking the fluorescence increase at the annealing temperature. Analysis of the products of these two amplifications by gel electrophoresis showed a DNA fragment of the expected size for the male DNA containing sample and no detectable DNA synthesis for the control sample.

DISCUSSION

Downstream processes such as hybridization to a sequence-specific probe can enhance the specificity of DNA detection by PCR. The elimination of these processes means that the specificity of this homogeneous assay depends solely on that of PCR. In the case of sickle-cell disease, we have shown that PCR alone has sufficient DNA sequence specificity to permit genetic screening. Using appropriate amplification conditions, there is little non-specific production of dsDNA in the absence of the appropriate target allele.

The specificity required to detect pathogens can be more or less than that required to do genetic screening, depending on the number of pathogens in the sample and the amount of other DNA that must be taken with the sample. A difficult target is HIV, which requires detection of a viral genome that can be at the level of a few copies per thousands of host cells⁶. Compared with genetic screening, which is performed on cells containing at least one copy of the target sequence, HIV detection requires both more specificity and the input of more total

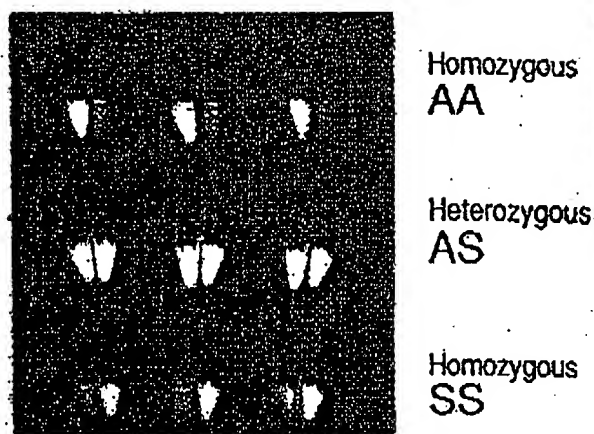


FIGURE 4 UV photograph of PCR tubes containing amplifications using EtBr that are specific to wild-type (A) or sickle (S) alleles of the human β -globin gene. The left of each pair of tubes contains allele-specific primers to the wild-type alleles, the right tube primers to the sickle allele. The photograph was taken after 30 cycles of PCR, and the input DNAs and the alleles they contain are indicated. Fifty ng of DNA was used to begin PCR. Typing was done in triplicate (3 pairs of PCR's) for each input DNA.

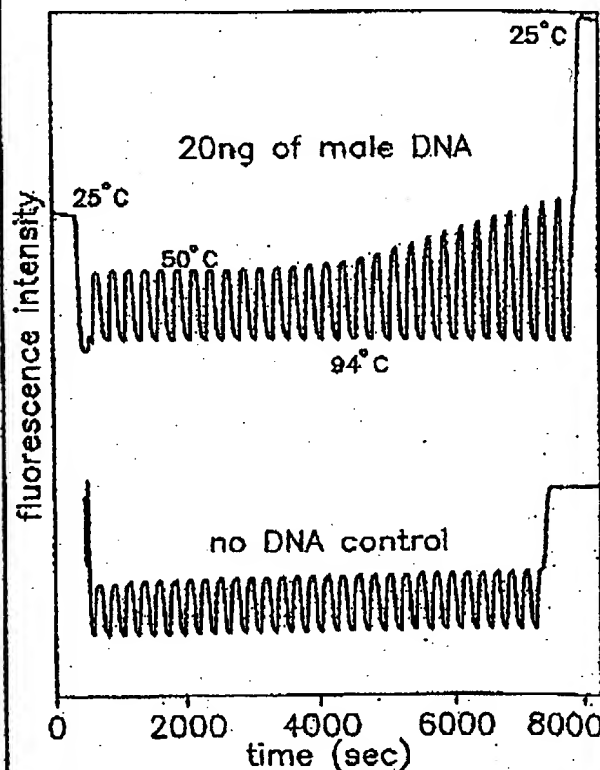


FIGURE 5 Continuous, real-time monitoring of a PCR. A fiber optic was used to carry excitation light to a PCR in progress and also emitted light back to a fluorometer (see Experimental Protocol). Amplification using human male-DNA specific primers in a PCR starting with 20 ng of human male DNA (top), or in a control PCR without DNA (bottom), were monitored. Thirty cycles of PCR were followed for each. The temperature cycled between 94°C (denaturation) and 50°C (annealing and extension). Note in the male DNA PCR, the cycle (time) dependent increase in fluorescence at the annealing/extension temperature.

DNA—up to microgram amounts—in order to have sufficient numbers of target sequences. This large amount of starting DNA in an amplification significantly increases the background fluorescence over which any additional fluorescence produced by PCR must be detected. An additional complication that occurs with targets in low copy-number is the formation of the "primer-dimer" artifact. This is the result of the extension of one primer using the other primer as a template. Although this occurs infrequently, once it occurs the extension product is a substrate for PCR amplification, and can compete with true PCR targets if those targets are rare. The primer-dimer product is of course dsDNA and thus is a potential source of false signal in this homogeneous assay.

To increase PCR specificity and reduce the effect of primer-dimer amplification, we are investigating a number of approaches, including the use of nested-primer amplifications that take place in a single tube³, and the "hot-start", in which nonspecific amplification is reduced by raising the temperature of the reaction before DNA synthesis begins²³. Preliminary results using these approaches suggest that primer-dimer is effectively reduced and it is possible to detect the increase in EtBr fluorescence in a PCR instigated by a single HIV genome in a background of 10^5 cells. With larger numbers of cells, the background fluorescence contributed by genomic DNA becomes problematic. To reduce this background, it may be possible to use sequence-specific DNA-binding dyes that can be made to preferentially bind PCR product over genomic DNA by incorporating the dye-binding DNA sequence into the PCR product through a 5' "add-on" to the oligonucleotide primer²⁴.

We have shown that the detection of fluorescence generated by an EtBr-containing PCR is straightforward, both once PCR is completed and continuously during thermocycling. The ease with which automation of specific DNA detection can be accomplished is the most promising aspect of this assay. The fluorescence analysis of completed PCRs is already possible with existing instrumentation in 96-well format²⁵. In this format, the fluorescence in each PCR can be quantitated before, after, and even at selected points during thermocycling by moving the rack of PCRs to a 96-microwell plate fluorescence reader²⁶.

The instrumentation necessary to continuously monitor multiple PCRs simultaneously is also simple in principle. A direct extension of the apparatus used here is to have multiple fiberoptics transmit the excitation light and fluorescent emissions to and from multiple PCRs. The ability to monitor multiple PCRs continuously may allow quantitation of target DNA copy number. Figure 3 shows that the larger the amount of starting target DNA, the sooner during PCR a fluorescence increase is detected. Preliminary experiments (Higuchi and Dollinger, manuscript in preparation) with continuous monitoring have shown a sensitivity to two-fold differences in initial target DNA concentration.

Conversely, if the number of target molecules is known—as it can be in genetic screening—continuous monitoring may provide a means of detecting false positive and false negative results. With a known number of target molecules, a true positive would exhibit detectable fluorescence by a predictable number of cycles of PCR. Increases in fluorescence detected before or after that cycle would indicate potential artifacts. False negative results due to, for example, inhibition of DNA polymerase, may be detected by including within each PCR an inefficiently amplifying marker. This marker results in a fluorescence increase only after a large number of cycles—many more than are necessary to detect a true

positive. If a sample fails to have a fluorescence increase after this many cycles, inhibition may be suspected. Since, in this assay, conclusions are drawn based on the presence or absence of fluorescence signal alone, such controls may be important. In any event, before any test based on this principle is ready for the clinic, an assessment of its false positive/false negative rates will need to be obtained using a large number of known samples.

In summary, the inclusion in PCR of dyes whose fluorescence is enhanced upon binding dsDNA makes it possible to detect specific DNA amplification from outside the PCR tube. In the future, instruments based upon this principle may facilitate the more widespread use of PCR in applications that demand the high throughput of samples.

EXPERIMENTAL PROTOCOL

Human HLA-DQ α gene amplifications containing EtBr. PCRs were set up in 100 μ l volumes containing 10 mM Tris-HCl, pH 8.3; 50 mM KCl; 4 mM MgCl₂; 2.5 units of *Taq* DNA polymerase (Perkin-Elmer Cetus, Norwalk, CT); 20 pmole each of human HLA-DQ α gene specific oligonucleotide primers (CH26 and CH27¹⁹ and approximately 10^5 copies of DQ α PCR product diluted from a previous reaction. Ethidium bromide (EtBr; Sigma) was used at the concentrations indicated in Figure 2. Thermocycling proceeded for 20 cycles in a model 480 thermocycler (Perkin-Elmer Cetus, Norwalk, CT) using a "step-cycle" program of 94°C for 1 min, denaturation and 60°C for 30 sec, annealing and 72°C for 30 sec, extension.

Y-chromosome specific PCR. PCRs (100 μ l total reaction volume) containing 0.5 μ g/ml EtBr were prepared as described for HLA-DQ α , except with different primers and target DNAs. These PCRs contained 15 pmole each male DNA-specific primers Y1.1 and Y1.2²⁰, and either 60 ng male, 60 ng female, 2 ng male, or no human DNA. Thermocycling was 94°C for 1 min, and 60°C for 1 min using a "step-cycle" program. The number of cycles for a sample were as indicated in Figure 3. Fluorescence measurement is described below.

Allele-specific, human β -globin gene PCR. Amplifications of 100 μ l volume using 0.5 μ g/ml of EtBr were prepared as described for HLA-DQ α above except with different primers and target DNAs. These PCRs contained either primer pair HGP2/Hp14A (wild-type globin specific primers) or HGP2/Hp14S (sickle-globin specific primers) at 10 pmole each primer per PCR. These primers were developed by Wu et al.²¹. Three different target DNAs were used in separate amplifications—50 ng each of human DNA that was homozygous for the sickle trait (SS), DNA that was heterozygous for the sickle trait (AS), or DNA that was homozygous for the w.t. globin (AA). Thermocycling was for 30 cycles at 94°C for 1 min, and 55°C for 1 min, using a "step-cycle" program. An annealing temperature of 55°C had been shown by Wu et al.²¹ to provide allele-specific amplification. Completed PCRs were photographed through a red filter (Wratten 23A) after placing the reaction tubes atop a model TM-36 transilluminator (UV-products San-Gabriel, CA).

Fluorescence measurement. Fluorescence measurements were made on PCRs containing EtBr in a Fluorolog-2 fluorometer (SPEX, Edison, NJ). Excitation was at the 500 nm band with about 2 nm bandwidth with a GG 435 nm cut-off filter (Melles Griest, Inc., Irvine, CA) to exclude second-order light. Emitted light was detected at 570 nm with a band width of about 7 nm. An OG 530 nm cut-off filter was used to remove the excitation light.

Continuous fluorescence monitoring of PCR. Continuous monitoring of a PCR in progress was accomplished using the spectrofluorometer and settings described above as well as a fiberoptic accessory (SPEX cat. no. 1950) to both send excitation light to, and receive emitted light from, a PCR placed in a well of a model 480 thermocycler (Perkin-Elmer Cetus). The probe end of the fiberoptic cable was attached with "5 minute-epoxy" to the open top of a PCR tube (a 0.5 ml polypropylene centrifuge tube with its cap removed) effectively sealing it. The exposed top of the PCR tube and the end of the fiberoptic cable were shielded from room light and the room lights were kept dimmed during each run. The monitored PCR was an amplification of Y-chromosome-specific repeat sequences as described above, except using an annealing/extension temperature of 50°C. The reaction was covered with mineral oil (2 drops) to prevent evaporation. Thermocycling and fluorescence measurement were started simultaneously. A time-base scan with a 10 second integration time

was used and the emission signal was ratioed to the excitation signal to control for changes in light-source intensity. Data were collected using the dm3000f, version 2.5 (SPEX) data system.

Acknowledgments

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References

- Mullis, K., Faloona, F., Scharf, S., Saiki, R., Horn, G. and Erlich, H. 1986. Specific enzymatic amplification of DNA *in vitro*: The polymerase chain reaction. *CSHSQ* 51:263-273.
- White, T. J., Arnheim, N. and Erlich, H. A. 1989. The polymerase chain reaction. *Trends Genet.* 5:185-189.
- Erlich, H. A., Gelfand, D. and Sninsky, J. J. 1991. Recent advances in the polymerase chain reaction. *Science* 252:1643-1651.
- Saiki, R. K., Gelfand, D. H., Stoffel, S., Scharf, S. J., Higuchi, R., Horn, G. T., Mullis, K. B. and Erlich, H. A. 1988. Primer-directed enzymatic amplification of DNA with a thermostable DNA polymerase. *Science* 239:487-491.
- Saiki, R. K., Walsh, P. S., Levenson, C. H. and Erlich, H. A. 1989. Genetic analysis of amplified DNA with immobilized sequence-specific oligonucleotide probes. *Proc. Natl. Acad. Sci. USA* 86:6230-6234.
- Kwok, S. Y., Mack, D. H., Mullis, K. B., Poiesz, B. J., Ehrlich, G. D., Blair, D. and Friedman-Kien, A. S. 1987. Identification of human immunodeficiency virus sequences by using *in vitro* enzymatic amplification and oligomer cleavage detection. *J. Virol.* 61:1690-1694.
- Chhab, F. F., Doherty, M., Cai, S. P., Kan, Y. W., Cooper, S. and Rubin, E. M. 1987. Detection of sickle cell anemia and thalassemia. *Nature* 329:203-204.
- Horn, G. T., Richards, B. and Klunger, K. W. 1989. Amplification of a highly polymorphic VNTR segment by the polymerase chain reaction. *Nuc. Acids Res.* 16:2140.
- Katz, E. D. and Dong, M. W. 1990. Rapid analysis and purification of polymerase chain reaction products by high-performance liquid chromatography. *Biotechniques* 8:546-555.
- Helger, D. N., Cohen, A. S. and Karger, B. L. 1990. Separation of DNA restriction fragments by high performance capillary electrophoresis with low and zero crosslinked polyacrylamide using continuous and pulsed electric fields. *J. Chromatogr.* 516:33-48.
- Kwok, S. Y. and Higuchi, R. G. 1989. Avoiding false positives with PCR. *Nature* 339:237-238.
- Chhab, F. F. and Kan, Y. W. 1989. Detection of specific DNA sequences by fluorescence amplification: a color complementation assay. *Proc. Natl. Acad. Sci. USA* 86:9178-9182.
- Holland, P. M., Abramson, R. D., Watson, R. and Gelfand, D. H. 1991. Detection of specific polymerase chain reaction products by utilizing the 5' to 3' exonuclease activity of *Thermus aquaticus* DNA polymerase. *Proc. Natl. Acad. Sci. USA* 88:7276-7280.
- Mariotti, J., Roques, B. P. and Le Peq, J. B. 1979. Ethidium dimer: a new reagent for the fluorimetric determination of nucleic acids. *Anal. Biochem.* 94:259-264.
- Kapuscinski, J. and Sacz, W. 1979. Interactions of 4',6'-diamidino-2-phenylindole with synthetic polynucleotides. *Nuc. Acids Res.* 6:5519-5534.
- Searle, M. S. and Embrey, K. J. 1990. Sequence-specific interaction of Hoechst 33258 with the minor groove of an adenine-tract DNA duplex studied in solution by ¹H NMR spectroscopy. *Nuc. Acids Res.* 18:3752-3762.
- Li, H. H., Gyllenstein, U. B., Cui, X. F., Saiki, R. K., Erlich, H. A. and Arnheim, N. 1988. Amplification and analysis of DNA sequences in single human sperm and diploid cells. *Nature* 336:414-417.
- Abbott, M. A., Poiesz, B. J., Byrne, B. C., Kwok, S. Y., Sninsky, J. J. and Erlich, H. A. 1988. Enzymatic gene amplification: qualitative and quantitative methods for detecting proviral DNA amplified *in vitro*. *J. Infect. Dis.* 158:1158.
- Saiki, R. K., Bugawan, T. L., Horn, G. T., Mullis, K. B. and Erlich, H. A. 1986. Analysis of enzymatically amplified β -globin and HLA-DQ α DNA with allele-specific oligonucleotide probes. *Nature* 324:163-166.
- Kogan, S. C., Doherty, M. and Gitschier, J. 1987. An improved method for prenatal diagnosis of genetic diseases by analysis of amplified DNA sequences. *N. Engl. J. Med.* 317:985-990.
- Wu, D. Y., Ugocz, L., Pol, B. K. and Wallace, R. B. 1989. Allele-specific enzymatic amplification of β -globin genomic DNA for diagnosis of sickle cell anemia. *Proc. Natl. Acad. Sci. USA* 86:2757-2760.
- Kwok, S., Kellogg, D. E., McKinney, N., Spasic, D., Guda, L., Levenson, C. and Sninsky, J. J. 1990. Effects of primer-template mismatches on the polymerase chain reaction: Human immunodeficiency virus type 1 model studies. *Nuc. Acids Res.* 18:999-1005.
- Chou, Q., Russell, M., Birck, D., Raymond, J. and Bloch, W. 1992. Prevention of pre-PCR mis-priming and primer dimerization improves low-copy-number amplifications. *Submitted.*
- Higuchi, R. 1989. Using PCR to engineer DNA, p. 61-70. In: *PCR Technology*. H. A. Erlich (Ed.). Stockton Press, New York, N.Y.
- Hall, L., Atwood, J. G., DiCesare, J., Katz, E., Picot, E., Williams, J. F. and Woudenberg, T. 1991. A high-performance system for automation of the polymerase chain reaction. *Biotechniques* 10:102-103, 106-112.
- Tumosa, N. and Kalwa, L. 1989. Fluorescent EIA screening of monoclonal antibodies to cell surface antigens. *J. Immun. Meth.* 116:59-63.

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Oligonucleotides with Fluorescent Dyes at Opposite Ends Provide a Quenched Probe System Useful for Detecting PCR Product and Nucleic Acid Hybridization

Kenneth J. Livak, Susan J.A. Flood, Jeffrey Marmaro, William Giusti, and Karin Deetz

Perkin-Elmer, Applied Biosystems Division, Foster City, California 94404

The 5' nuclease PCR assay detects the accumulation of specific PCR product by hybridization and cleavage of a double-labeled fluorogenic probe during the amplification reaction. The probe is an oligonucleotide with both a reporter fluorescent dye and a quencher dye attached. An increase in reporter fluorescence intensity indicates that the probe has hybridized to the target PCR product and has been cleaved by the 5' → 3' nucleolytic activity of *Taq* DNA polymerase. In this study, probes with the quencher dye attached to an internal nucleotide were compared with probes with the quencher dye attached to the 3'-end nucleotide. In all cases, the reporter dye was attached to the 5' end. All intact probes showed quenching of the reporter fluorescence. In general, probes with the quencher dye attached to the 3'-end nucleotide exhibited a larger signal in the 5' nuclease PCR assay than the internally labeled probes. It is proposed that the larger signal is caused by increased likelihood of cleavage by *Taq* DNA polymerase when the probe is hybridized to a template strand during PCR. Probes with the quencher dye attached to the 3'-end nucleotide also exhibited an increase in reporter fluorescence intensity when hybridized to a complementary strand. Thus, oligonucleotides with reporter and quencher dyes attached at opposite ends can be used as homogeneous hybridization probes.

A homogeneous assay for detecting the accumulation of specific PCR product that uses a double-labeled fluorogenic probe was described by Lee et al.⁽¹⁾ The assay exploits the 5' → 3' nucleolytic activity of *Taq* DNA polymerase^(2,3) and is diagramed in Figure 1. The fluorogenic probe consists of an oligonucleotide with a reporter fluorescent dye, such as a fluorescein, attached to the 5' end; and a quencher dye, such as a rhodamine, attached internally. When the fluorescein is excited by irradiation, its fluorescent emission will be quenched if the rhodamine is close enough to be excited through the process of fluorescence energy transfer (FET).^(4,5) During PCR, if the probe is hybridized to a template strand, *Taq* DNA polymerase will cleave the probe because of its inherent 5' → 3' nucleolytic activity. If the cleavage occurs between the fluorescein and rhodamine dyes, it causes an increase in fluorescein fluorescence intensity because the fluorescein is no longer quenched. The increase in fluorescein fluorescence intensity indicates that the probe-specific PCR product has been generated. Thus, FET between a reporter dye and a quencher dye is critical to the performance of the probe in the 5' nuclease PCR assay.

Quenching is completely dependent on the physical proximity of the two dyes.⁽⁶⁾ Because of this, it has been assumed that the quencher dye must be attached near the 5' end. Surprisingly, we have found that attaching a rhodamine dye at the 3' end of a probe still provides adequate quenching for the probe to perform in the 5' nuclease

PCR assay. Furthermore, cleavage of this type of probe is not required to achieve some reduction in quenching. Oligonucleotides with a reporter dye on the 5' end and a quencher dye on the 3' end exhibit a much higher reporter fluorescence when double-stranded as compared with single-stranded. This should make it possible to use this type of double-labeled probe for homogeneous detection of nucleic acid hybridization.

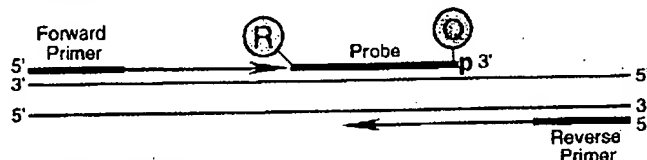
MATERIALS AND METHODS

Oligonucleotides

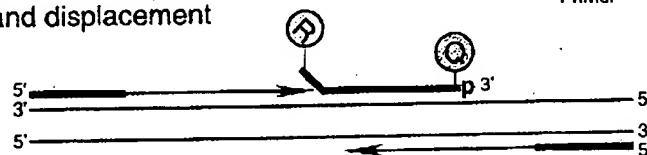
Table 1 shows the nucleotide sequence of the oligonucleotides used in this study. Linker arm nucleotide (LAN) phosphoramidite was obtained from Glen Research. The standard DNA phosphoramidites, 6-carboxyfluorescein (6-FAM) phosphoramidite, 6-carboxytetramethylrhodamine succinimidyl ester (TAMRA NHS ester), and Phosphalink for attaching a 3'-blocking phosphate, were obtained from Perkin-Elmer, Applied Biosystems Division. Oligonucleotide synthesis was performed using an ABI model 394 DNA synthesizer (Applied Biosystems). Primer and complement oligonucleotides were purified using Oligo Purification Cartridges (Applied Biosystems). Double-labeled probes were synthesized with 6-FAM-labeled phosphoramidite at the 5' end, LAN replacing one of the T's in the sequence, and Phosphalink at the 3' end. Following deprotection and ethanol precipitation, TAMRA NHS ester was coupled to the LAN-containing oligonucleotide in 250

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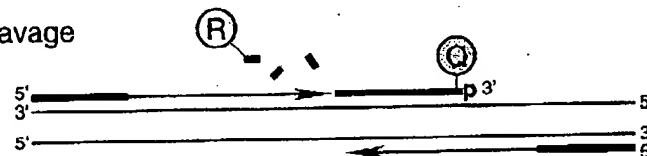
Polymerization



Strand displacement



Cleavage



Polymerization completed

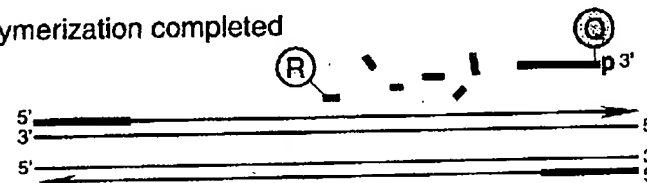


FIGURE 1 Diagram of 5' nuclease assay. Stepwise representation of the 5' → 3' nucleolytic activity of *Taq* DNA polymerase acting on a fluorogenic probe during one extension phase of PCR.

mm Na-bicarbonate buffer (pH 9.0) at room temperature. Unreacted dye was removed by passage over a PD-10 Sephadex column. Finally, the double-labeled probe was purified by preparative high-performance liquid chromatography (HPLC) using an Aquapore C₈ 220×4.6-mm column with 7-μm particle size. The column was developed with a 24-min linear gradient of 8–20% acetonitrile in 0.1 M TEAA (triethylamine acetate). Probes are named by designating the sequence from Table 1 and the position of the LAN-TAMRA moiety. For example, probe A1-7 has sequence A1 with LAN-TAMRA at nucleotide position 7 from the 5' end.

PCR Systems

All PCR amplifications were performed in the Perkin-Elmer GeneAmp PCR System 9600 using 50-μl reactions that contained 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 200 μM dATP, 200 μM dCTP, 200 μM dGTP, 400 μM dUTP, 0.5 unit of AmpErase uracil N-glycosylase (Perkin-Elmer), and 1.25 unit of AmpliTaq DNA polymerase (Perkin-Elmer). A 295-bp segment from exon 3 of the human β-actin

gene (nucleotides 2141–2435 in the sequence of Nakajima-Iijima et al.⁽⁷⁾) was amplified using primers AFP and ARP (Table 1), which are modified slightly from those of du Breuil et al.⁽⁸⁾ Actin amplification reactions contained 4 mM MgCl₂, 20 ng of human genomic DNA, 50 nM A1 or A3 probe, and 300 nM each

primer. The thermal regimen was 50°C (2 min), 95°C (10 min), 40 cycles of 95°C (20 sec), 60°C (1 min), and hold at 72°C. A 515-bp segment was amplified from a plasmid that consists of a segment of λ DNA (nucleotides 32,220–32,747) inserted in the *Sma*I site of vector pUC119. These reactions contained 3.5 mM MgCl₂, 1 ng of plasmid DNA, 50 nM P2 or P5 probe, 200 nM primer F119, and 200 nM primer R119. The thermal regimen was 50°C (2 min), 95°C (10 min), 25 cycles of 95°C (20 sec), 57°C (1 min), and hold at 72°C.

Fluorescence Detection

For each amplification reaction, a 40-μl aliquot of a sample was transferred to an individual well of a white, 96-well microtiter plate (Perkin-Elmer). Fluorescence was measured on the Perkin-Elmer TaqMan LS-50B System, which consists of a luminescence spectrometer with plate reader assembly, a 485-nm excitation filter, and a 515-nm emission filter. Excitation was at 488 nm using a 5-nm slit width. Emission was measured at 518 nm for 6-FAM (the reporter or R value) and 582 nm for TAMRA (the quencher or Q value) using a 10-nm slit width. To determine the increase in reporter emission that is caused by cleavage of the probe during PCR, three normalizations are applied to the raw emission data. First, emission intensity of a buffer blank is subtracted for each wavelength. Second, emission intensity of the reporter is

TABLE 1 Sequences of Oligonucleotides

| Name | Type | Sequence |
|------|------------|---|
| F119 | primer | ACCCACAGGAAGTATCACCACCTC |
| R119 | primer | ATGTCGCGTTCGGCTGACGTTCTGC |
| P2 | probe | TCGCATTACTGATCGTTCACCAACAGT _p |
| P2C | complement | GTACTGGTTGGCAACGATCAGTAATGCGATG |
| P5 | probe | CGGATTTGCTGGTATCTATGACAAGGAT _p |
| P5C | complement | TTCATCCTTGTATAGATACCAAGAAATCCG |
| AFP | primer | TCACCCACACTGTGCCATCTACGA |
| ARP | primer | CAGCGGAACCGTCTATTGCCAATGG |
| A1 | probe | ATGCCCTCCCCATGCCATCTGCGT _p |
| A1C | complement | AGACGCAGGATGGCATGGGGGAGGGCATAC |
| A3 | probe | CGCCCCTGGACTTCGAGCAAGAGAT _p |
| A3C | complement | CCATCTCTTGCTCGAAGTCCAGGGCGAC |

For each oligonucleotide used in this study, the nucleic acid sequence is given, written in the 5' → 3' direction. There are three types of oligonucleotides: PCR primer, fluorogenic probe used in the 5' nuclease assay, and complement used to hybridize to the corresponding probe. For the probes, the underlined base indicates a position where LAN with TAMRA attached was substituted for a T. (p) The presence of a 3' phosphate on each probe.

A1-2 RAQGCCCTCCCCCATGCCATCCTGCGTp
 A1-7 RATGCCCQCCCCCATGCCATCCTGCGTp
 A1-14 RATGCCCTCCCCCAQGCCATCCTGCGTp
 A1-19 RATGCCCTCCCCCATGCCAQCCTGCGTp
 A1-22 RATGCCCTCCCCCATGCCATCCQCGTp
 A1-26 RATGCCCTCCCCCATGCCATCCTGCGQp

| Probe | 518 nm | | 582 nm | | RQ ⁻ | RQ ⁺ | Δ RQ |
|-------|--------------|--------------|-------------|-------------|-----------------|-----------------|-------------|
| | no temp. | + temp. | no temp. | + temp. | | | |
| A1-2 | 25.5 ± 2.1 | 32.7 ± 1.9 | 38.2 ± 3.0 | 38.2 ± 2.0 | 0.67 ± 0.01 | 0.86 ± 0.06 | 0.19 ± 0.06 |
| A1-7 | 53.5 ± 4.3 | 395.1 ± 21.4 | 108.5 ± 6.3 | 110.3 ± 5.3 | 0.49 ± 0.03 | 3.58 ± 0.17 | 3.09 ± 0.18 |
| A1-14 | 127.0 ± 4.9 | 403.5 ± 19.1 | 109.7 ± 5.3 | 93.1 ± 6.3 | 1.16 ± 0.02 | 4.34 ± 0.15 | 3.18 ± 0.15 |
| A1-19 | 187.5 ± 17.9 | 422.7 ± 7.7 | 70.3 ± 7.4 | 73.0 ± 2.8 | 2.67 ± 0.05 | 5.80 ± 0.15 | 3.13 ± 0.16 |
| A1-22 | 224.6 ± 9.4 | 482.2 ± 43.6 | 100.0 ± 4.0 | 96.2 ± 9.6 | 2.25 ± 0.03 | 5.02 ± 0.11 | 2.77 ± 0.12 |
| A1-26 | 160.2 ± 8.9 | 454.1 ± 18.4 | 93.1 ± 5.4 | 90.7 ± 3.2 | 1.72 ± 0.02 | 5.01 ± 0.08 | 3.29 ± 0.08 |

FIGURE 2 Results of 5' nuclease assay comparing β -actin probes with TAMRA at different nucleotide positions. As described in Materials and Methods, PCR amplifications containing the indicated probes were performed, and the fluorescence emission was measured at 518 and 582 nm. Reported values are the average \pm 1 s.d. for six reactions run without added template (no temp.) and six reactions run with template (+ temp.). The RQ ratio was calculated for each individual reaction and averaged to give the reported RQ⁻ and RQ⁺ values.

divided by the emission intensity of the quencher to give an RQ ratio for each reaction tube. This normalizes for well-to-well variations in probe concentration and fluorescence measurement. Finally, Δ RQ is calculated by subtracting the RQ value of the no-template control (RQ⁻) from the RQ value for the complete reaction including template (RQ⁺).

RESULTS

A series of probes with increasing distances between the fluorescein reporter and rhodamine quencher were tested to investigate the minimum and maximum spacing that would give an acceptable performance in the 5' nuclease PCR assay. These probes hybridize to a target

sequence in the human β -actin gene. Figure 2 shows the results of an experiment in which these probes were included in PCR that amplified a segment of the β -actin gene containing the target sequence. Performance in the 5' nuclease PCR assay is monitored by the magnitude of Δ RQ, which is a measure of the increase in reporter fluorescence caused by PCR amplification of the probe target. Probe A1-2 has a Δ RQ value that is close to zero, indicating that the probe was not cleaved appreciably during the amplification reaction. This suggests that with the quencher dye on the second nucleotide from the 5' end, there is insufficient room for *Taq* polymerase to cleave efficiently between the reporter and quencher. The other five probes exhibited comparable Δ RQ values that are

clearly different from zero. Thus, all five probes are being cleaved during PCR amplification resulting in a similar increase in reporter fluorescence. It should be noted that complete digestion of a probe produces a much larger increase in reporter fluorescence than that observed in Figure 2 (data not shown). Thus, even in reactions where amplification occurs, the majority of probe molecules remain uncleaved. It is mainly for this reason that the fluorescence intensity of the quencher dye TAMRA changes little with amplification of the target. This is what allows us to use the 582-nm fluorescence reading as a normalization factor.

The magnitude of RQ⁻ depends mainly on the quenching efficiency inherent in the specific structure of the probe and the purity of the oligonucleotide. Thus, the larger RQ⁻ values indicate that probes A1-14, A1-19, A1-22, and A1-26 probably have reduced quenching as compared with A1-7. Still, the degree of quenching is sufficient to detect a highly significant increase in reporter fluorescence when each of these probes is cleaved during PCR.

To further investigate the ability of TAMRA on the 3' end to quench 6-FAM on the 5' end, three additional pairs of probes were tested in the 5' nuclease PCR assay. For each pair, one probe has TAMRA attached to an internal nucleotide and the other has TAMRA attached to the 3' end nucleotide. The results are shown in Table 2. For all three sets, the probe with the 3' quencher exhibits a Δ RQ value that is considerably higher than for the probe with the internal quencher. The RQ⁻ values suggest that differences in quenching are not as great as those observed with some of the A1 probes. These results demonstrate that a quencher dye on the 3' end of an oligonucleotide can quench efficiently the

TABLE 2 Results of 5' Nuclease Assay Comparing Probes with TAMRA Attached to an Internal or 3'-terminal Nucleotide

| Probe | 518 nm | | 582 nm | | RQ ⁻ | RQ ⁺ | Δ RQ |
|-------|-------------|--------------|-------------|--------------|-----------------|-----------------|-------------|
| | no temp. | + temp. | no temp. | + temp. | | | |
| A3-6 | 54.6 ± 3.2 | 84.8 ± 3.7 | 116.2 ± 6.4 | 115.6 ± 2.5 | 0.47 ± 0.02 | 0.73 ± 0.03 | 0.26 ± 0.04 |
| A3-24 | 72.1 ± 2.9 | 236.5 ± 11.1 | 84.2 ± 4.0 | 90.2 ± 3.8 | 0.86 ± 0.02 | 2.62 ± 0.05 | 1.76 ± 0.05 |
| P2-7 | 82.8 ± 4.4 | 384.0 ± 34.1 | 105.1 ± 6.4 | 120.4 ± 10.2 | 0.79 ± 0.02 | 3.19 ± 0.16 | 2.40 ± 0.16 |
| P2-27 | 113.4 ± 6.6 | 555.4 ± 14.1 | 140.7 ± 8.5 | 118.7 ± 4.8 | 0.81 ± 0.01 | 4.68 ± 0.10 | 3.88 ± 0.10 |
| P3-10 | 77.5 ± 6.5 | 244.4 ± 15.9 | 86.7 ± 4.3 | 95.8 ± 6.7 | 0.89 ± 0.05 | 2.55 ± 0.06 | 1.66 ± 0.08 |
| P5-28 | 64.0 ± 5.2 | 333.6 ± 12.1 | 100.6 ± 6.1 | 94.7 ± 6.3 | 0.63 ± 0.02 | 3.53 ± 0.12 | 2.89 ± 0.13 |

Reactions containing the indicated probes and calculations were performed as described in Material and Methods and in the legend to Fig. 2.

Research

fluorescence of a reporter dye on the 5' end. The degree of quenching is sufficient for this type of oligonucleotide to be used as a probe in the 5' nuclease PCR assay.

To test the hypothesis that quenching by a 3' TAMRA depends on the flexibility of the oligonucleotide, fluorescence was measured for probes in the single-stranded and double-stranded states. Table 3 reports the fluorescence observed at 518 and 582 nm. The relative degree of quenching is assessed by calculating the RQ ratio. For probes with TAMRA 6–10 nucleotides from the 5' end, there is little difference in the RQ values when comparing single-stranded with double-stranded oligonucleotides. The results for probes with TAMRA at the 3' end are much different. For these probes, hybridization to a complementary strand causes a dramatic increase in RQ. We propose that this loss of quenching is caused by the rigid structure of double-stranded DNA, which prevents the 5' and 3' ends from being in proximity.

When TAMRA is placed toward the 3' end, there is a marked Mg^{2+} effect on quenching. Figure 3 shows a plot of observed RQ values for the A1 series of probes as a function of Mg^{2+} concentration. With TAMRA attached near the 5' end (probe A1-2 or A1-7), the RQ value at 0 mM Mg^{2+} is only slightly higher than RQ at 10 mM Mg^{2+} . For probes A1-19, A1-22, and A1-26, the RQ values at 0 mM Mg^{2+} are very high, indicating a much

reduced quenching efficiency. For each of these probes, there is a marked decrease in RQ at 1 mM Mg^{2+} followed by a gradual decline as the Mg^{2+} concentration increases to 10 mM. Probe A1-14 shows an intermediate RQ value at 0 mM Mg^{2+} with a gradual decline at higher Mg^{2+} concentrations. In a low-salt environment with no Mg^{2+} present, a single-stranded oligonucleotide would be expected to adopt an extended conformation because of electrostatic repulsion. The binding of Mg^{2+} ions acts to shield the negative charge of the phosphate backbone so that the oligonucleotide can adopt conformations where the 3' end is close to the 5' end. Therefore, the observed Mg^{2+} effects support the notion that quenching of a 5' reporter dye by TAMRA at or near the 3' end depends on the flexibility of the oligonucleotide.

DISCUSSION

The striking finding of this study is that it seems the rhodamine dye TAMRA, placed at any position in an oligonucleotide, can quench the fluorescent emission of a fluorescein (6-FAM) placed at the 5' end. This implies that a single-stranded, double-labeled oligonucleotide must be able to adopt conformations where the TAMRA is close to the 5' end. It should be noted that the decay of 6-FAM in the excited state requires a certain amount of time. Therefore, what

matters for quenching is not the average distance between 6-FAM and TAMRA but, rather, how close TAMRA can get to 6-FAM during the lifetime of the 6-FAM excited state. As long as the decay time of the excited state is relatively long compared with the molecular motions of the oligonucleotide, quenching can occur. Thus, we propose that TAMRA at the 3' end, or any other position, can quench 6-FAM at the 5' end because TAMRA is in proximity to 6-FAM often enough to be able to accept energy transfer from an excited 6-FAM.

Details of the fluorescence measurements remain puzzling. For example, Table 3 shows that hybridization of probes A1-26, A3-24, and P5-28 to their complementary strands not only causes a large increase in 6-FAM fluorescence at 518 nm but also causes a modest increase in TAMRA fluorescence at 582 nm. If TAMRA is being excited by energy transfer from quenched 6-FAM, then loss of quenching attributable to hybridization should cause a decrease in the fluorescence emission of TAMRA. The fact that the fluorescence emission of TAMRA increases indicates that the situation is more complex. For example, we have anecdotal evidence that the bases of the oligonucleotide, especially G, quench the fluorescence of both 6-FAM and TAMRA to some degree. When double-stranded, base-pairing may reduce the ability of the bases to quench. The primary factor causing the quenching of 6-FAM in an intact probe is the TAMRA dye. Evidence for the importance of TAMRA is that 6-FAM fluorescence remains relatively unchanged when probes labeled only with 6-FAM are used in the 5' nuclease PCR assay (data not shown). Secondary effectors of fluorescence, both before and after cleavage of the probe, need to be explored further.

Regardless of the physical mechanism, the relative independence of position and quenching greatly simplifies the design of probes for the 5' nuclease PCR assay. There are three main factors that determine the performance of a double-labeled fluorescent probe in the 5' nuclease PCR assay. The first factor is the degree of quenching observed in the intact probe. This is characterized by the value of RQ^{-} , which is the ratio of reporter to quencher fluorescent emissions for a no template control PCR. Influences on the value of RQ^{-} include the particular reporter and quencher

TABLE 3 Comparison of Fluorescence Emissions of Single-stranded and Double-stranded Fluorogenic Probes

| Probe | 518 nm | | 582 nm | | RQ | |
|-------|--------|--------|--------|--------|------|------|
| | ss | ds | ss | ds | ss | ds |
| A1-7 | 27.75 | 68.53 | 61.08 | 138.18 | 0.45 | 0.50 |
| A1-26 | 43.31 | 509.38 | 53.50 | 93.86 | 0.81 | 5.43 |
| A3-6 | 16.75 | 62.88 | 39.33 | 165.57 | 0.43 | 0.38 |
| A3-24 | 30.05 | 578.64 | 67.72 | 140.25 | 0.45 | 3.21 |
| P2-7 | 35.02 | 70.13 | 54.63 | 121.09 | 0.64 | 0.58 |
| P2-27 | 39.89 | 320.47 | 65.10 | 61.13 | 0.61 | 5.25 |
| P5-10 | 27.34 | 144.85 | 61.95 | 165.54 | 0.44 | 0.87 |
| P5-28 | 33.65 | 462.29 | 72.39 | 104.61 | 0.46 | 4.43 |

(ss) Single-stranded. The fluorescence emissions at 518 or 582 nm for solutions containing a final concentration of 50 nM indicated probe, 10 mM Tris-HCl (pH 8.3), 50 mM KCl, and 10 mM $MgCl_2$. (ds) Double-stranded. The solutions contained, in addition, 100 nM A1C for probes A1-7 and A1-26, 100 nM A3C for probes A3-6 and A3-24, 100 nM P2C for probes P2-7 and P2-27, or 100 nM P5C for probes P5-10 and P5-28. Before the addition of $MgCl_2$, 120 μ l of each sample was heated at 95°C for 5 min. Following the addition of 80 μ l of 25 mM $MgCl_2$, each sample was allowed to cool to room temperature and the fluorescence emissions were measured. Reported values are the average of three determinations.

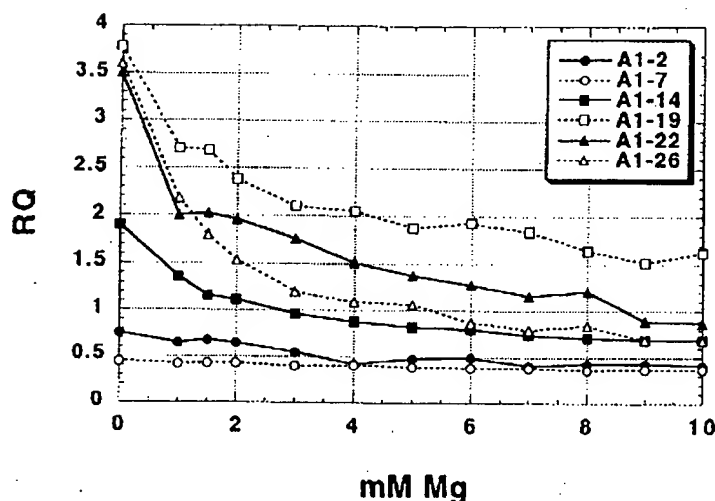


FIGURE 3 Effect of Mg^{2+} concentration on RQ ratio for the A1 series of probes. The fluorescence emission intensity at 518 and 582 nm was measured for solutions containing 50 nM probe, 10 mM Tris-HCl (pH 8.3), 50 mM KCl, and varying amounts (0–10 mM) of $MgCl_2$. The calculated RQ ratios (518 nm intensity divided by 582 nm intensity) are plotted vs. $MgCl_2$ concentration (mM Mg). The key (upper right) shows the probes examined.

dyes used, spacing between reporter and quencher dyes, nucleotide sequence context effects, presence of structure or other factors that reduce flexibility of the oligonucleotide, and purity of the probe. The second factor is the efficiency of hybridization, which depends on probe T_m , presence of secondary structure in probe or template, annealing temperature, and other reaction conditions. The third factor is the efficiency at which *Taq* DNA polymerase cleaves the bound probe between the reporter and quencher dyes. This cleavage is dependent on sequence complementarity between probe and template as shown by the observation that mismatches in the segment between reporter and quencher dyes drastically reduce the cleavage of probe.⁽¹⁾

The rise in RQ^- values for the A1 series of probes seems to indicate that the degree of quenching is reduced somewhat as the quencher is placed toward the 3' end. The lowest apparent quenching is observed for probe A1-19 (see Fig. 3) rather than for the probe where the TAMRA is at the 3' end (A1-26). This is understandable, as the conformation of the 3' end position would be expected to be less restricted than the conformation of an internal position. In effect, a quencher at the 3' end is freer to adopt conformations close to the 5' reporter dye than is an internally placed quencher. For the other three sets of

probes, the interpretation of RQ^- values is less clear-cut. The A3 probes show the same trend as A1, with the 3' TAMRA probe having a larger RQ^- than the internal TAMRA probe. For the P2 pair, both probes have about the same RQ^- value. For the P5 probes, the RQ^- for the 3' probe is less than for the internally labeled probe. Another factor that may explain some of the observed variation is that purity affects the RQ^- value. Although all probes are HPLC purified, a small amount of contamination with unquenched reporter can have a large effect on RQ^- .

Although there may be a modest effect on degree of quenching, the position of the quencher apparently can have a large effect on the efficiency of probe cleavage. The most drastic effect is observed with probe A1-2, where placement of the TAMRA on the second nucleotide reduces the efficiency of cleavage to almost zero. For the A3, P2, and P5 probes, ΔRQ is much greater for the 3' TAMRA probes as compared with the internal TAMRA probes. This is explained most easily by assuming that probes with TAMRA at the 3' end are more likely to be cleaved between reporter and quencher than are probes with TAMRA attached internally. For the A1 probes, the cleavage efficiency of probe A1-7 must already be quite high, as ΔRQ does not increase when the quencher is placed closer to the 3' end. This illus-

trates the importance of being able to use probes with a quencher on the 3' end in the 5' nuclease PCR assay. In this assay, an increase in the intensity of reporter fluorescence is observed only when the probe is cleaved between the reporter and quencher dyes. By placing the reporter and quencher dyes on the opposite ends of an oligonucleotide probe, any cleavage that occurs will be detected. When the quencher is attached to an internal nucleotide, sometimes the probe works well (A1-7) and other times not so well (A3-6). The relatively poor performance of probe A3-6 presumably means the probe is being cleaved 3' to the quencher rather than between the reporter and quencher. Therefore, the best chance of having a probe that reliably detects accumulation of PCR product in the 5' nuclease PCR assay is to use a probe with the reporter and quencher dyes on opposite ends.

Placing the quencher dye on the 3' end may also provide a slight benefit in terms of hybridization efficiency. The presence of a quencher attached to an internal nucleotide might be expected to disrupt base-pairing and reduce the T_m of a probe. In fact, a 2°C–3°C reduction in T_m has been observed for two probes with internally attached TAMRAs.⁽⁹⁾ This disruptive effect would be minimized by placing the quencher at the 3' end. Thus, probes with 3' quenchers might exhibit slightly higher hybridization efficiencies than probes with internal quenchers.

The combination of increased cleavage and hybridization efficiencies means that probes with 3' quenchers probably will be more tolerant of mismatches between probe and target as compared with internally labeled probes. This tolerance of mismatches can be advantageous, as when trying to use a single probe to detect PCR-amplified products from samples of different species. Also, it means that cleavage of probe during PCR is less sensitive to alterations in annealing temperature or other reaction conditions. The one application where tolerance of mismatches may be a disadvantage is for allelic discrimination. Lee et al.⁽¹⁾ demonstrated that allele-specific probes were cleaved between reporter and quencher only when hybridized to a perfectly complementary target. This allowed them to distinguish the normal human cystic fibrosis allele from the $\Delta F508$ mutant. Their probes had TAMRA attached to the seventh nucleotide from

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the 5' end and were designed so that any mismatches were between the reporter and quencher. Increasing the distance between reporter and quencher would lessen the disruptive effect of mismatches and allow cleavage of the probe on the incorrect target. Thus, probes with a quencher attached to an internal nucleotide may still be useful for allelic discrimination.

In this study loss of quenching upon hybridization was used to show that quenching by a 3' TAMRA is dependent on the flexibility of a single-stranded oligonucleotide. The increase in reporter fluorescence intensity, though, could also be used to determine whether hybridization has occurred or not. Thus, oligonucleotides with reporter and quencher dyes attached at opposite ends should also be useful as hybridization probes. The ability to detect hybridization in real time means that these probes could be used to measure hybridization kinetics. Also, this type of probe could be used to develop homogeneous hybridization assays for diagnostics or other applications. Bagwell et al.⁽¹⁰⁾ describe just this type of homogeneous assay where hybridization of a probe causes an increase in fluorescence caused by a loss of quenching. However, they utilized a complex probe design that requires adding nucleotides to both ends of the probe sequence to form two imperfect hairpins. The results presented here demonstrate that the simple addition of a reporter dye to one end of an oligonucleotide and a quencher dye to the other end generates a fluorogenic probe that can detect hybridization or PCR amplification.

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REFERENCES

1. Lee, L.G., C.R. Connell, and W. Bloch. 1993. Allelic discrimination by nick-translation PCR with fluorogenic probes. *Nucleic Acids Res.* **21**: 3761-3766.
2. Holland, P.M., R.D. Abramson, R. Watson, and D.H. Gelfand. 1991. Detection of specific polymerase chain reaction prod-

- uct by utilizing the 5' to 3' exonuclease activity of *Thermus aquaticus* DNA polymerase. *Proc. Natl. Acad. Sci.* **88**: 7276-7280.
3. Lyamichev, V., M.A.D. Brow, and J.E. Dahlberg. 1993. Structure-specific endonucleolytic cleavage of nucleic acids by eubacterial DNA polymerases. *Science* **260**: 778-783.
4. Förster, V.Th. 1948. Zwischenmolekulare Energiewanderung und Fluoreszenz. *Ann. Phys. (Leipzig)* **2**: 55-75.
5. Lakowicz, J.R. 1983. Energy transfer. In *Principles of fluorescent spectroscopy*, pp. 303-339. Plenum Press, New York, NY.
6. Stryer, L. and R.P. Haugland. 1967. Energy transfer: A spectroscopic ruler. *Proc. Natl. Acad. Sci.* **58**: 719-726.
7. Nakajima-Iijima, S., H. Hamada, P. Reddy, and T. Kakunaga. 1985. Molecular structure of the human cytoplasmic beta-actin gene: Inter-species homology of sequences in the introns. *Proc. Natl. Acad. Sci.* **82**: 6133-6137.
8. du Breuil, R.M., J.M. Patel, and B.V. Mendelow. 1993. Quantitation of β -actin-specific mRNA transcripts using xeno-competitive PCR. *PCR Methods Applic.* **3**: 57-59.
9. Livak, K.J. (unpubl.).
10. Bagwell, C.B., M.E. Munson, R.L. Christensen, and E.J. Lovett. 1994. A new homogeneous assay system for specific nucleic acid sequences: Poly-dA and poly-A detection. *Nucleic Acids Res.* **22**: 2424-2425.

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GENOMI METHODS

Real Time Quantitative PCR

Christian A. Heid,¹ Junko Stevens,² Kenneth J. Livak,² and
P. Mickey Williams^{1,3}

¹BioAnalytical Technology Department, Genentech, Inc., South San Francisco, California 94080;

²Applied BioSystems Division of Perkin Elmer Corp., Foster City, California 94404

We have developed a novel "real time" quantitative PCR method. The method measures PCR product accumulation through a dual-labeled fluorogenic probe (i.e., TaqMan Probe). This method provides very accurate and reproducible quantitation of gene copies. Unlike other quantitative PCR methods, real-time PCR does not require post-PCR sample handling, preventing potential PCR product carry-over contamination and resulting in much faster and higher throughput assays. The real-time PCR method has a very large dynamic range of starting target molecule determination (at least five orders of magnitude). Real-time quantitative PCR is extremely accurate and less-labor-intensive than current quantitative PCR methods.

Quantitative nucleic acid sequence analysis has had an important role in many fields of biological research. Measurement of gene expression (RNA) has been used extensively in monitoring biological responses to various stimuli (Tan et al. 1994; Huang et al. 1995a,b; Prud'homme et al. 1995). Quantitative gene analysis (DNA) has been used to determine the genomic quantity of a particular gene, as in the case of the human *HER2* gene, which is amplified in ~30% of breast tumors (Slamon et al. 1987). Gene and genome quantitation (DNA and RNA) also have been used for analysis of human immunodeficiency virus (HIV) burden demonstrating changes in the levels of virus throughout the different phases of the disease (Connor et al. 1993; Platak et al. 1993b; Furtado et al. 1995).

Many methods have been described for the quantitative analysis of nucleic acid sequences (both for RNA and DNA; Southern 1975; Sharp et al. 1980; Thomas 1980). Recently, PCR has proven to be a powerful tool for quantitative nucleic acid analysis. PCR and reverse transcriptase (RT)-PCR have permitted the analysis of minimal starting quantities of nucleic acid (as little as one cell equivalent). This has made possible many experiments that could not have been performed with traditional methods. Although PCR has provided a powerful tool, it is imperative

that it be used properly for quantitation (Rasmussen 1995). Many early reports of quantitative PCR and RT-PCR described quantitation of the PCR product but did not measure the initial target sequence quantity. It is essential to design proper controls for the quantitation of the initial target sequences (Perre 1992; Clementi et al. 1993).

Researchers have developed several methods of quantitative PCR and RT-PCR. One approach measures PCR product quantity in the log phase of the reaction before the plateau (Kellogg et al. 1990; Pang et al. 1990). This method requires that each sample has equal input amounts of nucleic acid and that each sample under analysis amplifies with identical efficiency up to the point of quantitative analysis. A gene sequence (contained in all samples at relatively constant quantities, such as β -actin) can be used for sample amplification efficiency normalization. Using conventional methods of PCR detection and quantitation (gel electrophoresis or plate capture hybridization), it is extremely laborious to assure that all samples are analyzed during the log phase of the reaction (for both the target gene and the normalization gene). Another method, quantitative competitive (QC)-PCR, has been developed and is used widely for PCR quantitation. QC-PCR relies on the inclusion of an internal control competitor in each reaction (Becker-Andre 1991; Platak et al. 1993a,b). The efficiency of each reaction is normalized to the internal competitor. A known amount of internal competitor can be

³Corresponding author.

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added to each sample. To obtain relative quantitation, the unknown target PCR product is compared with the known competitor PCR product. Success of a quantitative competitive PCR assay relies on developing an internal control that amplifies with the same efficiency as the target molecule. The design of the competitor and the validation of amplification efficiencies require a dedicated effort. However, because QC-PCR does not require that PCR products be analyzed during the log phase of the amplification, it is the easier of the two methods to use.

Several detection systems are used for quantitative PCR and RT-PCR analysis: (1) agarose gels, (2) fluorescent labeling of PCR products and detection with laser-induced fluorescence using capillary electrophoresis (Fusco et al. 1995; Williams et al. 1996) or acrylamide gels, and (3) plate capture and sandwich probe hybridization (Mulder et al. 1994). Although these methods proved successful, each method requires post-PCR manipulations that add time to the analysis and may lead to laboratory contamination. The sample throughput of these methods is limited (with the exception of the plate capture approach), and, therefore, these methods are not well suited for uses demanding high sample throughput (i.e., screening of large numbers of biomolecules or analyzing samples for diagnostics or clinical trials).

Here we report the development of a novel assay for quantitative DNA analysis. The assay is based on the use of the 5' nuclease assay first described by Holland et al. (1991). The method uses the 5' nuclease activity of *Taq* polymerase to cleave a nonextendible hybridization probe during the extension phase of PCR. The approach uses dual-labeled fluorogenic hybridization probes (Lee et al. 1993; Bassler et al. 1995; Livak et al. 1995a,b). One fluorescent dye serves as a reporter [FAM (i.e., 6-carboxyfluorescein)] and its emission spectra is quenched by the second fluorescent dye, TAMRA (i.e., 6-carboxy-tetramethylrhodamine). The nuclease degradation of the hybridization probe releases the quenching of the FAM fluorescent emission, resulting in an increase in peak fluorescent emission at 518 nm. The use of a sequence detector (ABI Prism) allows measurement of fluorescent spectra of all 96 wells of the thermal cycler continuously during the PCR amplification. Therefore, the reactions are monitored in real time. The output data is described and quantitative analysis of input target DNA sequences is discussed below.

RESULTS

PCR Product Detection in Real Time

The goal was to develop a high-throughput, sensitive, and accurate gene quantitation assay for use in monitoring lipid mediated therapeutic gene delivery. A plasmid encoding human factor VIII gene sequence, pl8TM (see Methods), was used as a model therapeutic gene. The assay uses fluorescent Taqman methodology and an instrument capable of measuring fluorescence in real time (ABI Prism 7700 Sequence Detector). The Taqman reaction requires a hybridization probe labeled with two different fluorescent dyes. One dye is a reporter dye (FAM), the other is a quenching dye (TAMRA). When the probe is intact, fluorescent energy transfer occurs and the reporter dye fluorescent emission is absorbed by the quenching dye (TAMRA). During the extension phase of the PCR cycle, the fluorescent hybridization probe is cleaved by the 5'-3' nucleolytic activity of the DNA polymerase. On cleavage of the probe, the reporter dye emission is no longer transferred efficiently to the quenching dye, resulting in an increase of the reporter dye fluorescent emission spectra. PCR primers and probes were designed for the human factor VIII sequence and human β -actin gene (as described in Methods). Optimization reactions were performed to choose the appropriate probe and magnesium concentrations yielding the highest intensity of reporter fluorescent signal without sacrificing specificity. The instrument uses a charge-coupled device (i.e., CCD camera) for measuring the fluorescent emission spectra from 500 to 650 nm. Each PCR tube was monitored sequentially for 25 msec with continuous monitoring throughout the amplification. Each tube was re-examined every 8.5 sec. Computer software was designed to examine the fluorescent intensity of both the reporter dye (FAM) and the quenching dye (TAMRA). The fluorescent intensity of the quenching dye, TAMRA, changes very little over the course of the PCR amplification (data not shown). Therefore, the intensity of TAMRA dye emission serves as an internal standard with which to normalize the reporter dye (FAM) emission variations. The software calculates a value termed ΔRn (or $\Delta R(2)$) using the following equation: $\Delta Rn = (Rn^t) / (Rn^i)$, where Rn^t = emission intensity of reporter/emission intensity of quencher at any given time in a reaction tube, and Rn^i = emission intensity of re-

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porter/emission intensity of quencher measured prior to PCR amplification in that same reaction tube. For the purpose of quantitation, the last three data points (ΔRn s) collected during the extension step for each PCR cycle were analyzed. The nucleolytic degradation of the hybridization probe occurs during the extension phase of PCR, and, therefore, reporter fluorescent emission increases during this time. The three data points were averaged for each PCR cycle and the mean value for each was plotted in an "amplification plot" shown in Figure 1A. The ΔRn mean value is plotted on the y-axis, and time, represented by cycle number, is plotted on the x-axis. During the early cycles of the PCR amplification, the ΔRn

value remains at base line. When sufficient hybridization probe has been cleaved by the *Taq* polymerase nuclease activity, the intensity of reporter fluorescent emission increases. Most PCR amplifications reach a plateau phase of reporter fluorescent emission if the reaction is carried out to high cycle numbers. The amplification plot is examined early in the reaction, at a point that represents the log phase of product accumulation. This is done by assigning an arbitrary threshold that is based on the variability of the base-line data. In Figure 1A, the threshold was set at 10 standard deviations above the mean of base line emission calculated from cycles 1 to 15. Once the threshold is chosen, the point at which

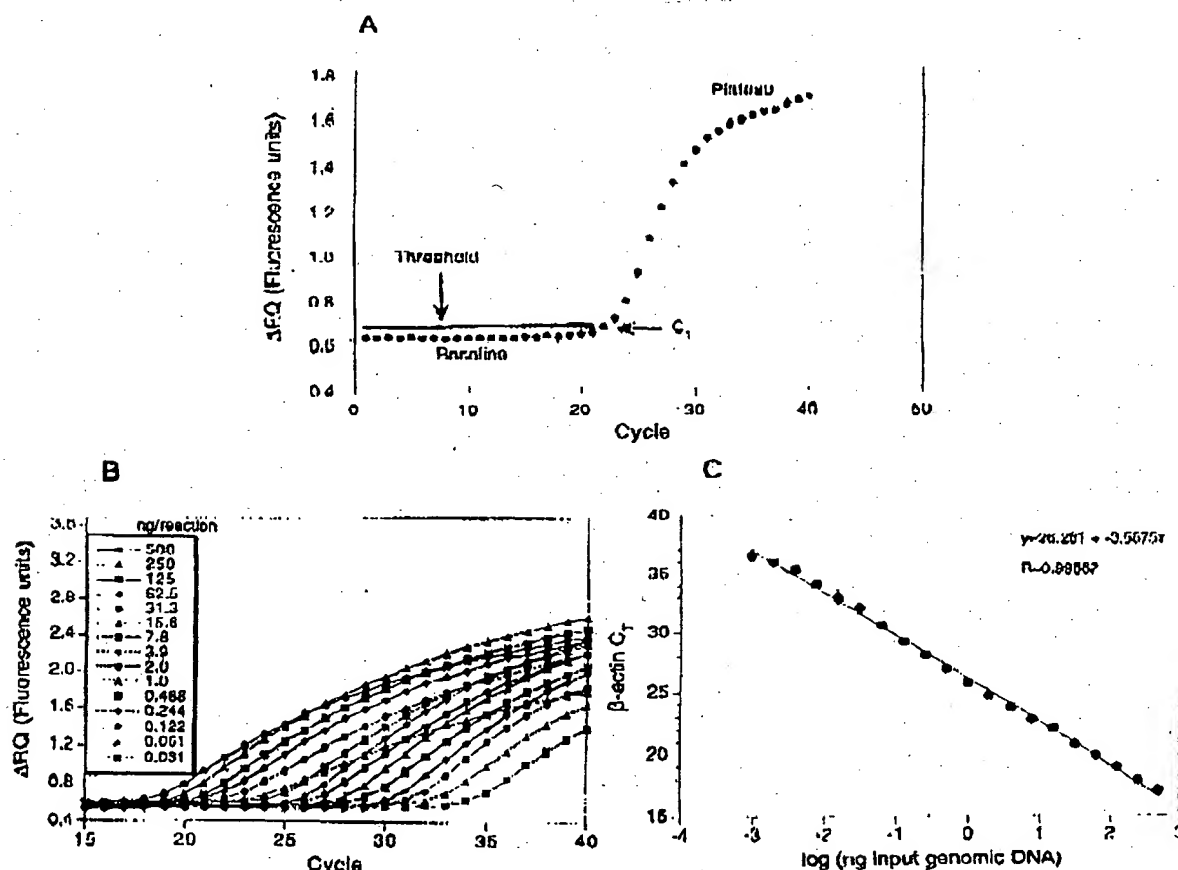


Figure 1 PCR product detection in real time. (A) The Model 7700 software will construct amplification plots from the extension phase fluorescent emission data collected during the PCR amplification. The standard deviation is determined from the data points collected from the base line of the amplification plot. C_T values are calculated by determining the point at which the fluorescence exceeds a threshold limit (usually 10 times the standard deviation of the base line). (B) Overlay of amplification plots of serially (1:2) diluted human genomic DNA samples amplified with β -actin primers. (C) Input DNA concentration of the samples plotted versus C_T . All

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the amplification plot crosses the threshold is defined as C_T . C_T is reported as the cycle number at this point. As will be demonstrated, the C_T value is predictive of the quantity of input target.

C_T Values Provide a Quantitative Measurement of Input Target Sequences

Figure 1B shows amplification plots of 15 different PCR amplifications overlaid. The amplifications were performed on a 1:2 serial dilution of human genomic DNA. The amplified target was human β actin. The amplification plots shift to the right (to higher threshold cycles) as the input target quantity is reduced. This is expected because reactions with fewer starting copies of the target molecule require greater amplification to degrade enough probe to attain the threshold fluorescence. An arbitrary threshold of 10 standard deviations above the base line was used to determine the C_T values. Figure 1C represents the C_T values plotted versus the sample dilution value. Each dilution was amplified in triplicate PCR amplifications and plotted as mean values with error bars representing one standard deviation. The C_T values decrease linearly with increasing target quantity. Thus, C_T values can be used as a quantitative measurement of the input target number. It should be noted that the amplification plot for the 15.6-ng sample shown in Figure 1B does not reflect the same fluorescent rate of increase exhibited by most of the other samples. The 15.6-ng sample also achieves endpoint plateau at a lower fluorescent value than would be expected based on the input DNA. This phenomenon has been observed occasionally with other samples (data not shown) and may be attributable to late cycle inhibition; this hypothesis is still under investigation. It is important to note that the flattened slope and early plateau do not impact significantly the calculated C_T value as demonstrated by the fit on the line shown in Figure 1C. All triplicate amplifications resulted in very similar C_T values—the standard deviation did not exceed 0.5 for any dilution. This experiment contains a >100,000-fold range of input target molecules. Using C_T values for quantitation permits a much larger assay range than directly using total fluorescent emission intensity for quantitation. The linear range of fluorescent intensity measurement of the ABI Prism 7700 Se-

ments over a very large range of relative starting target quantities.

Sample Preparation Validation

Several parameters influence the efficiency of PCR amplification: magnesium and salt concentrations, reaction conditions (i.e., time and temperature), PCR target size and composition, primer sequences, and sample purity. All of the above factors are common to a single PCR assay, except sample to sample purity. In an effort to validate the method of sample preparation for the factor VIII assay, PCR amplification reproducibility and efficiency of 10 replicate sample preparations were examined. After genomic DNA was prepared from the 10 replicate samples, the DNA was quantitated by ultraviolet spectroscopy. Amplifications were performed analyzing β -actin gene content in 100 and 25 ng of total genomic DNA. Each PCR amplification was performed in triplicate. Comparison of C_T values for each triplicate sample show minimal variation based on standard deviation and coefficient of variance (Table 1). Therefore, each of the triplicate PCR amplifications was highly reproducible, demonstrating that real time PCR using this instrumentation introduces minimal variation into the quantitative PCR analysis. Comparison of the mean C_T values of the 10 replicate sample preparations also showed minimal variability, indicating that each sample preparation yielded similar results for β -actin gene quantity. The highest C_T difference between any of the samples was 0.85 and 0.71 for the 100 and 25 ng samples, respectively. Additionally, the amplification of each sample exhibited an equivalent rate of fluorescent emission intensity change per amount of DNA target analyzed as indicated by similar slopes derived from the sample dilutions (Fig. 2). Any sample containing an excess of a PCR inhibitor would exhibit a greater measured β -actin C_T value for a given quantity of DNA. In addition, the inhibitor would be diluted along with the sample in the dilution analysis (Fig. 2), altering the expected C_T value change. Each sample amplification yielded a similar result in the analysis, demonstrating that this method of sample preparation is highly reproducible with regard to sample purity.

Quantitative Analysis of a Plasmid After

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Table 1. Reproducibility of Sample Preparation Method

| Sample no. | 100 ng | | | | 25 ng | | | |
|------------|----------------|-------|--------------------|------|----------------|-------|--------------------|------|
| | C _T | mean | standard deviation | CV | C _T | mean | standard deviation | CV |
| 1 | 18.24 | 18.27 | 0.06 | 0.32 | 20.48 | 20.51 | 0.03 | 0.17 |
| | 18.23 | | | | 20.55 | | | |
| | 18.33 | | | | 20.5 | | | |
| 2 | 18.33 | 18.37 | 0.06 | 0.32 | 20.61 | 20.54 | 0.11 | 0.54 |
| | 18.35 | | | | 20.59 | | | |
| | 18.44 | | | | 20.41 | | | |
| 3 | 18.3 | 18.34 | 0.07 | 0.36 | 20.54 | 20.54 | 0.06 | 0.28 |
| | 18.3 | | | | 20.6 | | | |
| | 18.42 | | | | 20.49 | | | |
| 4 | 18.15 | 18.23 | 0.08 | 0.46 | 20.48 | 20.43 | 0.05 | 0.26 |
| | 18.23 | | | | 20.44 | | | |
| | 18.32 | | | | 20.38 | | | |
| 5 | 18.4 | 18.42 | 0.04 | 0.23 | 20.68 | 20.73 | 0.13 | 0.61 |
| | 18.38 | | | | 20.87 | | | |
| | 18.46 | | | | 20.63 | | | |
| 6 | 18.54 | 18.74 | 0.24 | 1.26 | 21.09 | 21.06 | 0.03 | 0.15 |
| | 18.67 | | | | 21.04 | | | |
| | 19 | | | | 21.04 | | | |
| 7 | 18.28 | 18.39 | 0.12 | 0.66 | 20.67 | 20.68 | 0.04 | 0.2 |
| | 18.36 | | | | 20.73 | | | |
| | 18.52 | | | | 20.65 | | | |
| 8 | 18.45 | 18.63 | 0.16 | 0.83 | 20.98 | 20.86 | 0.12 | 0.57 |
| | 18.7 | | | | 20.84 | | | |
| | 18.73 | | | | 20.75 | | | |
| 9 | 18.18 | 18.29 | 0.1 | 0.55 | 20.46 | 20.51 | 0.07 | 0.32 |
| | 18.34 | | | | 20.54 | | | |
| | 18.26 | | | | 20.48 | | | |
| 10 | 18.42 | 18.55 | 0.12 | 0.65 | 20.79 | 20.73 | 0.1 | 0.16 |
| | 18.57 | | | | 20.78 | | | |
| | 18.66 | | | | 20.62 | | | |
| Mean | (1 10) | 18.42 | 0.17 | 0.90 | | 20.66 | 0.19 | 0.94 |

for containing a partial cDNA for human factor VIII, pF8TM. A series of transfections was set up using a decreasing amount of the plasmid (40, 4, 0.5, and 0.1 µg). Twenty-four hours post-transfection, total DNA was purified from each flask of cells. β-Actin gene quantity was chosen as a value for normalization of genomic DNA concentration from each sample. In this experiment, β-actin gene content should remain constant relative to total genomic DNA. Figure 3 shows the result of the β-actin DNA measurement (100 ng total DNA determined by ultraviolet spectroscopy) of each sample. Each sample was analyzed in triplicate and the mean β-actin C_T values of the triplicates were plotted (error bars represent one standard deviation). The highest difference

between any two sample means was 0.95 C_T. Ten nanograms of total DNA of each sample were also examined for β-actin. The results again showed that very similar amounts of genomic DNA were present; the maximum mean β-actin C_T value difference was 1.0. As Figure 3 shows, the rate of β-actin C_T change between the 100 and 10-ng samples was similar (slope values range between 3.56 and -3.45). This verifies again that the method of sample preparation yields samples of identical PCR integrity (i.e., no sample contained an excessive amount of a PCR inhibitor). However, these results indicate that each sample contained slight differences in the actual amount of genomic DNA analyzed. Determination of actual genomic DNA concentration was accomplished

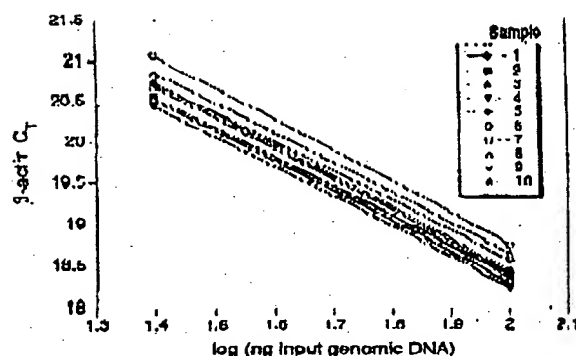


Figure 2 Sample preparation purity. The replicate samples shown in Table 1 were also amplified in triplicate using 25 ng of each DNA sample. The figure shows the input DNA concentration (100 and 25 ng) vs. C_t . In the figure, the 100 and 25 ng points for each sample are connected by a line.

by plotting the mean β -actin C_t value obtained for each 100-ng sample on a β -actin standard curve (shown in Fig. 4C). The actual genomic DNA concentration of each sample, a , was obtained by extrapolation to the x-axis.

Figure 4A shows the measured (i.e., non-normalized) quantities of factor VIII plasmid DNA (pF8TM) from each of the four transient cell transfections. Each reaction contained 100 ng of total sample DNA (as determined by UV spectroscopy). Each sample was analyzed in triplicate

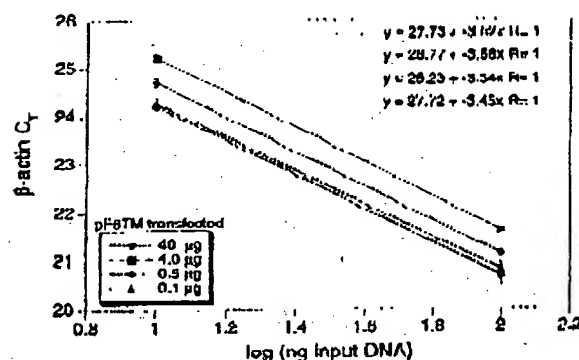


Figure 3 Analysis of transfected cell DNA quantity and purity. The DNA preparations of the four 293 cell transfections (40, 4, 0.5, and 0.1 µg of pF8TM) were analyzed for the β -actin gene. 100 and 10 ng (determined by ultraviolet spectroscopy) of each sample were amplified in triplicate. For each amount of pF8TM that was transfected, the β -actin C_t values are plotted versus the total input DNA concentration.

REAL TIME QUANTITATIVE PCR

PCR amplifications. As shown, pF8TM purified from the 293 cells decreases (mean C_t values increase) with decreasing amounts of plasmid (transfected). The mean C_t values obtained for pF8TM in Figure 4A were plotted on a standard curve comprised of serially diluted pF8TM, shown in Figure 4B. The quantity of pF8TM, b , found in each of the four transfections was determined by extrapolation to the x-axis of the standard curve in Figure 4B. These uncorrected values, b , for pF8TM were normalized to determine the actual amount of pF8TM found per 100 ng of genomic DNA by using the equation:

$$\frac{b \times 100 \text{ ng}}{a} = \text{actual pF8TM copies per 100 ng of genomic DNA}$$

where a = actual genomic DNA in a sample and b = pF8TM copies from the standard curve. The normalized quantity of pF8TM per 100 ng of genomic DNA for each of the four transfections is shown in Figure 4D. These results show that the quantity of factor VIII plasmid associated with the 293 cells, 24 hr after transfection, decreases with decreasing plasmid concentration used in the transfection. The quantity of pF8TM associated with 293 cells, after transfection with 40 µg of plasmid, was 35 pg per 100 ng genomic DNA. This results in ~520 plasmid copies per cell.

DISCUSSION

We have described a new method for quantitating gene copy numbers using real-time analysis of PCR amplifications. Real-time PCR is compatible with either of the two PCR (RT-PCR) approaches: (1) quantitative competitive where an internal competitor for each target sequence is used for normalization (data not shown) or (2) quantitative comparative PCR using a normalization gene contained within the sample (i.e., β -actin) or a "housekeeping" gene for RT-PCR. If equal amounts of nucleic acid are analyzed for each sample and if the amplification efficiency before quantitative analysis is identical for each sample, the internal control (normalization gene or competitor) should give equal signals for all samples.

The real-time PCR method offers several advantages over the other two methods currently employed (see the Introduction). First, the real-time PCR method is performed in a closed-tube system and requires no post-PCR manipulation

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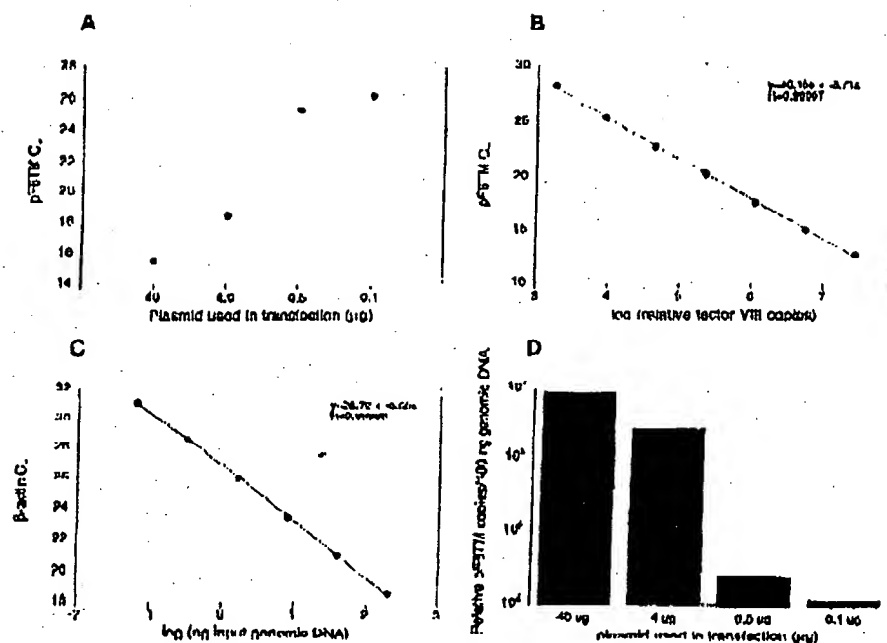


Figure 4. Quantitative analysis of pF8TM in transfected cells. (A) Amount of plasmid DNA used for the transfection plotted against the mean C_T value determined for pF8TM remaining 24 hr after transfection. (B,C) Standard curves of pF8TM and β-actin, respectively. pF8TM DNA (B) and genomic DNA (C) were diluted serially 1:5 before amplification with the appropriate primers. The β-actin standard curve was used to normalize the results of A to 100 ng of genomic DNA. (D) The amount of pF8TM present per 100 ng of genomic DNA.

of sample. Therefore, the potential for PCR contamination in the laboratory is reduced because amplified products can be analyzed and disposed of without opening the reaction tubes. Second, this method supports the use of a normalization gene (i.e., β-actin) for quantitative PCR or housekeeping genes for quantitative RT-PCR controls. Analysis is performed in real time during the log phase of product accumulation. Analysis during log phase permits many different genes (over a wide input target range) to be analyzed simultaneously, without concern of reaching reaction plateau at different cycles. This will make multi-gene analysis assays much easier to develop, because individual internal competitors will not be needed for each gene under analysis. Third, sample throughput will increase dramatically with the new method because there is no post-PCR processing time. Additionally, working in a 96-well format is highly compatible with automation technology.

The real-time PCR method is highly reproducible. Replicate amplifications can be analyzed

for each sample minimizing potential error. The system allows for a very large assay dynamic range (approaching 1,000,000-fold starting target). Using a standard curve for the target of interest, relative copy number values can be determined for any unknown sample. Fluorescent threshold values, C_T, correlate linearly with relative DNA copy numbers. Real time quantitative RT-PCR methodology (Gibson et al., this issue) has also been developed. Finally, real time quantitative PCR methodology can be used to develop high-throughput screening assays for a variety of applications [quantitative gene expression (RT-PCR), gene copy assays (Her2, HIV, etc.), genotyping (knockout mouse analysis), and Immuno-PCR].

Real-time PCR may also be performed using intercalating dyes (Higuchi et al. 1992) such as ethidium bromide. The fluorogenic probe method offers a major advantage over intercalating dyes—greater specificity (i.e., primer dimers and nonspecific PCR products are not detected).

REAL TIME QUANTITATIVE PCR

METHODS

Generation of a Plasmid Containing a Partial cDNA for Human Factor VIII

Total RNA was harvested (RNAzol B from Tel Test, Inc., Friendswood, TX) from cells transfected with a factor VIII expression vector, pCIS2.8(251) (Eaton et al. 1986; Gorman et al. 1990). A factor VIII partial cDNA sequence was generated by RT-PCR (Clontech Amp 12.1 (1st RNA PCR Kit) (part N808-0179, PE Applied Biosystems, Foster City, CA)) using the PCR primers F8for and F8rev (primer sequences are shown below). The amplicon was reamplified using modified F8for and F8rev primers (appended with *HindIII* and *HindIII* restriction site sequences at the 5' end) and cloned into pCIS2.32 (Promega Corp., Madison, WI). The resulting clone, pF8TM, was used for transient transfection of 293 cells.

Amplification of Target DNA and Detection of Amplicon Factor VIII Plasmid DNA

(pF8TM) was amplified with the primers F8for 5'-CCCCCTTCCCAAGAGTGAACCTGTC-3' and F8rev 5'-AAACCTTCAACCTTGGATGCTTACGTC-3'. The reaction produced a 422-bp PCR product. The forward primer was designed to recognize a unique sequence found in the 5' untranslated region of the parent pCIS2.8(251) plasmid and therefore does not recognize and amplify the human factor VIII gene. Primers were chosen with the assistance of the computer program Oligo 4.0 (National Biosciences, Inc., Plymouth, MN). The human β -actin gene was amplified with the primers β -actin forward primer 5'-TCACCCACACTGTTGCCATCTTACCA-3' and β -actin reverse primer 5'-CAGCGGAACCCGCTCATTCGCAATGG-3'. The reaction produced a 295-bp PCR product.

Amplification reactions (50 μ l) contained a DNA sample, 10 \times PCR Buffer II (5 μ l), 200 μ M dATP, dCTP, dGTP, and 400 μ M dTTP, 4 mM MgCl₂, 1.25 Units AmpliTaq DNA polymerase, 0.5 unit AmpErase uracil N-glycosylase (UNG), 50 pmole of each factor VIII primer, and 15 pmole of each β -actin primer. The reactions also contained one of the following detection probes (100 nM each): F8probe 5'-(FAM)AGCTTCTTCCACCTTCTTCTTCTGTCGCCCTT(TAMRA)p 3' and β -actin probe 5'-(FAM)ATGCCCTT(X(TAMRA)CCCCCATGCCATC)p-3' where p indicates phosphorylation and X indicates a linker arm nucleotide. Reaction tubes were MicroAmp Optical Tubes (part number N801 0933, Perkin Elmer) that were frosted (at Perkin Elmer) to prevent light from reflecting. Tube caps were similar to MicroAmp Caps but specially designed to prevent light scattering. All of the PCR consumables were supplied by PE Applied Biosystems (Foster City, CA) except the factor VIII primers, which were synthesized at Genentech, Inc. (South San Francisco, CA). Probes were designed using the Oligo 4.0 software, following guidelines suggested in the Model 7700 Sequence Detector Instrument manual. Briefly, probe T_m should be at least 5°C higher than the annealing temperature used during thermal cycling; primers should not form stable duplexes with the probe.

The thermal cycling conditions included 2 min at 50°C and 10 min at 95°C. Thermal cycling proceeded with

reactions were performed in the Model 7700 Sequence Detector (PE Applied Biosystems), which contains a GeneAmp PCR System 9600. Reaction conditions were programmed on a Power Macintosh 7100 (Apple Computer, Santa Clara, CA) linked directly to the Model 7700 Sequence Detector. Analysis of data was also performed on the Macintosh computer. Collection and analysis software was developed at PE Applied Biosystems.

Transfection of Cells with Factor VIII Construct

Four T175 flasks of 293 cells (ATCC CRL 1573), a human fetal kidney suspension cell line, were grown to 80% confluency and transfected pF8TM. Cells were grown in the following media: 50% HAM'S F12 without GHT, 50% low glucose Dulbecco's modified Eagle medium (DMEM) without glycine with sodium bicarbonate, 10% fetal bovine serum, 2 mM L-glutamine, and 1% penicillin-streptomycin. The media was changed 30 min before the transfection. pF8TM DNA amounts of 40, 4, 0.5, and 0.1 μ g were added to 1.5 ml of a solution containing 0.125 M CaCl₂ and 1 \times HEPES. The four mixtures were left at room temperature for 10 min and then added dropwise to the cells. The flasks were incubated at 37°C and 5% CO₂ for 24 hr, washed with PBS, and resuspended in PBS. The resuspended cells were divided into aliquots and DNA was extracted immediately using the QIAamp Blood Kit (Qiagen, Chatsworth, CA). DNA was eluted into 200 μ l of 20 mM Tris-HCl at pH 8.0.

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REFERENCES

- Bassler, H.A., S.J. Flood, K.J. Livak, J. Marmaro, R. Koon, and C.A. Batt. 1995. Use of a fluorogenic probe in a PCR-based assay for the detection of *Listeria monocytogenes*. *App. Environ. Microbiol.* 61: 3724-3728.
- Becker-Andre, M. 1991. Quantitative evaluation of mRNA levels. *Meth. Mol. Cell. Biol.* 2: 189-201.
- Clement, M., S. Menzo, P. Bagnarelli, A. Manzini, A. Valenza, and P.E. Varaldo. 1993. Quantitative PCR and RT-PCR in virology. [Review]. *PCR Methods Applic.* 2: 191-196.
- Connor, R.I., H. Mohri, Y. Cao, and D.D. Ho. 1993. Increased viral burden and cytopathicity correlate temporally with CD4⁺ T-lymphocyte decline and clinical progression in human immunodeficiency virus type 1-infected individuals. *J. Virol.* 67: 1772-1777.
- Eaton, D.L., W.J. Wood, D. Eaton, P.E. Hass, P.

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- Venar, and C. Gorman. 1986. Construction and characterization of an active factor VIII variant lacking the central one third of the molecule. *Biochemistry* 25: 8343-8347.
- Fasco, M.J., C.P. Treanor, S. Spivack, H.L. Wigge, and L.S. Kaminsky. 1995. Quantitative RNA-polymerase chain reaction-DNA analysis by capillary electrophoresis and laser-induced fluorescence. *Anal. Biochem.* 224: 140-147.
- Forre, J. 1992. Quantitative or semi-quantitative PCR: Reality versus myth. *PCR Methods Applic.* 2: 1-9.
- Furtado, M.R., L.A. Kingsley, and S.M. Wollinsky. 1995. Changes in the viral mRNA expression pattern correlate with a rapid rate of CD4+ T-cell number decline in human immunodeficiency virus type 1-infected individuals. *J. Virol.* 69: 2092-2100.
- Gibson, U.E.M., C.A. Heid, and P.M. Williams. 1996. A novel method for real time quantitative competitive RT-PCR. *Genome Res.* (this issue).
- Gorman, C.M., D.R. Gies, and G. McCray. 1990. Transient production of proteins using an adenovirus transformed cell line. *DNA Prot. Engin. Tech.* 2: 3-10.
- Higuchi, R., G. Dollinger, P.S. Walsh, and R. Griffith. 1992. Simultaneous amplification and detection of specific DNA sequences. *Biochemistry* 10: 413-417.
- Holland, P.M., R.D. Abramson, R. Watson, and D.J. Gelfand. 1991. Detection of specific polymerase chain reaction product by utilizing the 5'-3' exonuclease activity of *Thermus aquaticus* DNA polymerase. *Proc. Natl. Acad. Sci.* 88: 7276-7280.
- Huang, S.K., H.Q. Xiao, T.J. Klein, G. Paciotti, D.G. Marsh, L.M. Lichtenstein, and M.C. Liu. 1995a. IL-13 expression at the sites of allergen challenge in patients with asthma. *J. Immun.* 155: 2688-2694.
- Huang, S.K., M. Yi, E. Palmer, and D.G. Marsh. 1995b. A dominant T cell receptor beta-chain in response to a short ragweed allergen. *Am J. Immun.* 164: 6157-6162.
- Kellogg, D.E., J.J. Slinisky, and S. Kowk. 1990. Quantitation of HIV-1 proviral DNA relative to cellular DNA by the polymerase chain reaction. *Anal. Biochem.* 189: 202-208.
- Lee, J.-G., C.R. Connell, and W. Bloch. 1993. Allelic discrimination by nick-translation PCR with fluorogenic probes. *Nucleic Acids Res.* 21: 3761-3766.
- Livak, K.J., S.J. Flood, J. Marmaro, W. Giusti, and K. Deetz. 1995a. Oligonucleotides with fluorescent dyes at opposite ends provide a quenched probe system useful for detecting PCR product and nucleic acid hybridization. *PCR Methods Applic.* 4: 357-362.
- Livak, K.J., J. Marmaro, and J.A. Todd. 1995b. Towards fully automated genome-wide polymorphism screening [letter]. *Nature Genet.* 9: 341-342.
- Mulder, J., N. McKinney, C. Christopherson, J. Slinisky, L. Greenfield, and S. Kwak. 1994. Rapid and simple PCR assay for quantitation of human immunodeficiency virus type 1 RNA in plasma: Application to acute retroviral infection. *J. Clin. Microbiol.* 32: 292-300.
- Pang, S., Y. Koyanagi, S. Miller, C. Wiloy, H.V. Vinters, and L.S. Chen. 1990. High levels of unintegrated HIV-1 DNA in brain tissue of AIDS dementia patients. *Nature* 343: 85-89.
- Platak, M.J., K.C. Luk, B. Williams, and J.D. Lifson. 1995a. Quantitative competitive polymerase chain reaction for accurate quantitation of HIV DNA and RNA species. *Aid/Techniques* 14: 70-81.
- Platak, M.J., M.S. Saag, L.C. Yang, S.J. Clark, J.C. Kappes, K.C. Luk, B.H. Hann, G.M. Shaw, and J.D. Lifson. 1995b. High levels of HIV-1 in plasma during all stages of infection determined by competitive PCR [see Comments]. *Science* 269: 1749-1754.
- Prod'homme, G.J., D.H. Kono, and A.N. Theofilopoulos. 1995. Quantitative polymerase chain reaction analysis reveals marked overexpression of interleukin-1 beta, interleukin-1 and interferon-gamma mRNA in the lymph nodes of lupus-prone mice. *Mol. Immunol.* 32: 495-503.
- Racymackers, J. 1995. A commentary on the practical applications of competitive PCR. *Genome Res.* 5: 91-94.
- Sharp, P.A., A.J. Berk, and S.M. Berget. 1980. Transcription maps of adenovirus. *Methods Enzymol.* 65: 750-768.
- Slamon, D.J., G.M. Clark, S.C. Wong, W.J. Levin, A. Ullrich, and W.L. McGuire. 1987. Human breast cancer: Correlation of relapse and survival with amplification of the HER-2/neu oncogene. *Science* 235: 177-182.
- Southern, E.M. 1975. Detection of specific sequences among DNA fragments separated by gel electrophoresis. *J. Mol. Biol.* 98: 503-517.
- Tan, X., X. Sun, C.F. Gonzalez, and W. Hsueh. 1994. PAI and TNF increase the precursor of B-lymphocyte mRNA in mouse intestine: Quantitative analysis by competitive PCR. *Biochim. Biophys. Acta* 1215: 157-162.
- Thomas, P.S. 1980. Hybridization of denatured RNA and small DNA fragments transferred to nitrocellulose. *Proc. Natl. Acad. Sci.* 77: 5201-5205.
- Williams, S., C. Scher, A. Krishnamo, C. Heid, B. Karger, and P.M. Williams. 1996. Quantitative competitive PCR: Analysis of amplified products of the HIV-1 gag gene by capillary electrophoresis with laser induced fluorescence detection. *Anal. Biochem.* (in press).

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WISP genes are members of the connective tissue growth factor family that are up-regulated in Wnt-1-transformed cells and aberrantly expressed in human colon tumors

DIANE PENNICA*†, TODD A. SWANSON*, JAMES W. WELSH*, MARGARET A. ROY‡, DAVID A. LAWRENCE*, JAMES LEE‡, JENNIFER BRUSH‡, LISA A. TANEYHILL§, BETHANNE DEUEL‡, MICHAEL LEW¶, COLIN WATANABE¶, ROBERT L. COHEN*, MONA F. MELHEM**, GENE G. FINLEY**, PHIL QUIRKE††, AUDREY D. GODDARD‡, KENNETH J. HILLAN¶, AUSTIN L. GURNEY‡, DAVID BOTSTEIN‡,††, AND ARNOLD J. LEVINE§

Departments of *Molecular Oncology, †Molecular Biology, ‡Scientific Computing, and §Pathology, Genentech Inc., 1 DNA Way, South San Francisco, CA 94080; **University of Pittsburgh School of Medicine, Veterans Administration Medical Center, Pittsburgh, PA 15240; ††University of Leeds, Leeds, LS29JT United Kingdom; ‡Department of Genetics, Stanford University, Palo Alto, CA 94305; and §Department of Molecular Biology, Princeton University, Princeton, NJ 08544

Contributed by David Botstein and Arnold J. Levine, October 21, 1998

ABSTRACT Wnt family members are critical to many developmental processes, and components of the Wnt signaling pathway have been linked to tumorigenesis in familial and sporadic colon carcinomas. Here we report the identification of two genes, *WISP-1* and *WISP-2*, that are up-regulated in the mouse mammary epithelial cell line C57MG transformed by Wnt-1, but not by Wnt-4. Together with a third related gene, *WISP-3*, these proteins define a subfamily of the connective tissue growth factor family. Two distinct systems demonstrated *WISP* induction to be associated with the expression of Wnt-1. These included (i) C57MG cells infected with a Wnt-1 retroviral vector or expressing Wnt-1 under the control of a tetracycline repressible promoter, and (ii) Wnt-1 transgenic mice. The *WISP-1* gene was localized to human chromosome 8q24.1-8q24.3. *WISP-1* genomic DNA was amplified in colon cancer cell lines and in human colon tumors and its RNA overexpressed (2- to >30-fold) in 84% of the tumors examined compared with patient-matched normal mucosa. *WISP-3* mapped to chromosome 6q22-6q23 and also was overexpressed (4- to >40-fold) in 63% of the colon tumors analyzed. In contrast, *WISP-2* mapped to human chromosome 20q12-20q13 and its DNA was amplified, but RNA expression was reduced (2- to >30-fold) in 79% of the tumors. These results suggest that the *WISP* genes may be downstream of Wnt-1 signaling and that aberrant levels of *WISP* expression in colon cancer may play a role in colon tumorigenesis.

Wnt-1 is a member of an expanding family of cysteine-rich, glycosylated signaling proteins that mediate diverse developmental processes such as the control of cell proliferation, adhesion, cell polarity, and the establishment of cell fates (1, 2). Wnt-1 originally was identified as an oncogene activated by the insertion of mouse mammary tumor virus in virus-induced mammary adenocarcinomas (3, 4). Although Wnt-1 is not expressed in the normal mammary gland, expression of Wnt-1 in transgenic mice causes mammary tumors (5).

In mammalian cells, Wnt family members initiate signaling by binding to the seven-transmembrane spanning Frizzled receptors and recruiting the cytoplasmic protein Dishevelled (Dsh) to the cell membrane (1, 2, 6). Dsh then inhibits the kinase activity of the normally constitutively active glycogen synthase kinase-3 β (GSK-3 β) resulting in an increase in β -catenin levels. Stabilized β -catenin interacts with the transcription factor TCF/Lef1, forming a complex that appears in

the nucleus and binds TCF/Lef1 target DNA elements to activate transcription (7, 8). Other experiments suggest that the adenomatous polyposis coli (APC) tumor suppressor gene also plays an important role in Wnt signaling by regulating β -catenin levels (9). APC is phosphorylated by GSK-3 β , binds to β -catenin, and facilitates its degradation. Mutations in either APC or β -catenin have been associated with colon carcinomas and melanomas, suggesting these mutations contribute to the development of these types of cancer, implicating the Wnt pathway in tumorigenesis (1).

Although much has been learned about the Wnt signaling pathway over the past several years, only a few of the transcriptionally activated downstream components activated by Wnt have been characterized. Those that have been described cannot account for all of the diverse functions attributed to Wnt signaling. Among the candidate Wnt target genes are those encoding the nodal-related 3 gene, *Xnr3*, a member of the transforming growth factor (TGF)- β superfamily, and the homeobox genes, *engrailed*, *goosecoid*, *twin* (*Xtwn*), and *siamois* (2). A recent report also identifies *c-myc* as a target gene of the Wnt signaling pathway (10).

To identify additional downstream genes in the Wnt signaling pathway that are relevant to the transformed cell phenotype, we used a PCR-based cDNA subtraction strategy, suppression subtractive hybridization (SSH) (11), using RNA isolated from C57MG mouse mammary epithelial cells and C57MG cells stably transformed by a Wnt-1 retrovirus. Overexpression of Wnt-1 in this cell line is sufficient to induce a partially transformed phenotype, characterized by elongated and refractile cells that lose contact inhibition and form a multilayered array (12, 13). We reasoned that genes differentially expressed between these two cell lines might contribute to the transformed phenotype.

In this paper, we describe the cloning and characterization of two genes up-regulated in Wnt-1 transformed cells, *WISP-1* and *WISP-2*, and a third related gene, *WISP-3*. The *WISP* genes are members of the CCN family of growth factors, which includes connective tissue growth factor (CTGF), Cyr61, and *nov*, a family not previously linked to Wnt signaling.

MATERIALS AND METHODS

SSH. SSH was performed by using the PCR-Select cDNA Subtraction Kit (CLONTECH). Tester double-stranded

Abbreviations: TGF, transforming growth factor; CTGF, connective tissue growth factor; SSH, suppression subtractive hybridization; WVC, von Willebrand factor type C module.

Data deposition: The sequences reported in this paper have been deposited in the Genbank database (accession nos. AF100777, AF100778, AF100779, AF100780, and AF100781).

†To whom reprint requests should be addressed. e-mail: diane@gene.com.

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cDNA was synthesized from 2 μ g of poly(A)⁺ RNA isolated from the C57MG/Wnt-1 cell line and driver cDNA from 2 μ g of poly(A)⁺ RNA from the parent C57MG cells. The subtracted cDNA library was subcloned into a pGEM-T vector for further analysis.

cDNA Library Screening. Clones encoding full-length mouse *WISP-1* were isolated by screening a λ gt10 mouse embryo cDNA library (CLONTECH) with a 70-bp probe from the original partial clone 568 sequence corresponding to amino acids 128–169. Clones encoding full-length human *WISP-1* were isolated by screening λ gt10 lung and fetal kidney cDNA libraries with the same probe at low stringency. Clones encoding full-length mouse and human *WISP-2* were isolated by screening a C57MG/Wnt-1 or human fetal lung cDNA library with a probe corresponding to nucleotides 1463–1512. Full-length cDNAs encoding *WISP-3* were cloned from human bone marrow and fetal kidney libraries.

Expression of Human *WISP* RNA. PCR amplification of first-strand cDNA was performed with human Multiple Tissue cDNA panels (CLONTECH) and 300 μ M of each dNTP at 94°C for 1 sec, 62°C for 30 sec, 72°C for 1 min, for 22–32 cycles. *WISP* and glyceraldehyde-3-phosphate dehydrogenase primer sequences are available on request.

In Situ Hybridization. ³²P-labeled sense and antisense riboprobes were transcribed from an 897-bp PCR product corresponding to nucleotides 601–1440 of mouse *WISP-1* or a 294-bp PCR product corresponding to nucleotides 82–375 of mouse *WISP-2*. All tissues were processed as described (40).

Radiation Hybrid Mapping. Genomic DNA from each hybrid in the Stanford G3 and Genebridge4 Radiation Hybrid Panels (Research Genetics, Huntsville, AL) and human and hamster control DNAs were PCR-amplified, and the results were submitted to the Stanford or Massachusetts Institute of Technology web servers.

Cell Lines, Tumors, and Mucosa Specimens. Tissue specimens were obtained from the Department of Pathology (University of Pittsburgh) for patients undergoing colon resection and from the University of Leeds, United Kingdom. Genomic DNA was isolated (Qiagen) from the pooled blood of 10 normal human donors, surgical specimens, and the following ATCC human cell lines: SW480, COLO 320DM, HT-29, WiDr, and SW403 (colon adenocarcinomas), SW620 (lymph node metastasis, colon adenocarcinoma), HCT 116 (colon carcinoma), SK-CO-1 (colon adenocarcinoma, ascites), and HM7 (a variant of ATCC colon adenocarcinoma cell line LS 174T). DNA concentration was determined by using Hoechst dye 33258 intercalation fluorimetry. Total RNA was prepared by homogenization in 7 M GuSCN followed by centrifugation over CsCl cushions or prepared by using RNeasy.

Gene Amplification and RNA Expression Analysis. Relative gene amplification and RNA expression of *WISPs* and *c-myc* in the cell lines, colorectal tumors, and normal mucosa were determined by quantitative PCR. Gene-specific primers and fluorogenic probes (sequences available on request) were designed and used to amplify and quantitate the genes. The relative gene copy number was derived by using the formula $2^{(\Delta Ct)}$ where ΔCt represents the difference in amplification cycles required to detect the *WISP* genes in peripheral blood lymphocyte DNA compared with colon tumor DNA or colon tumor RNA compared with normal mucosal RNA. The δ -method was used for calculation of the SE of the gene copy number or RNA expression level. The *WISP*-specific signal was normalized to that of the glyceraldehyde-3-phosphate dehydrogenase housekeeping gene. All TaqMan assay reagents were obtained from Perkin-Elmer Applied Biosystems.

RESULTS

Isolation of *WISP-1* and *WISP-2* by SSH. To identify Wnt-1-inducible genes, we used the technique of SSH using the

mouse mammary epithelial cell line C57MG and C57MG cells that stably express Wnt-1 (11). Candidate differentially expressed cDNAs (1,384 total) were sequenced. Thirty-nine percent of the sequences matched known genes or homologues, 32% matched expressed sequence tags, and 29% had no match. To confirm that the transcript was differentially expressed, semiquantitative reverse transcription-PCR and Northern analysis were performed by using mRNA from the C57MG and C57MG/Wnt-1 cells.

Two of the cDNAs, *WISP-1* and *WISP-2*, were differentially expressed, being induced in the C57MG/Wnt-1 cell line, but not in the parent C57MG cells or C57MG cells overexpressing Wnt-4 (Fig. 1 A and B). Wnt-4, unlike Wnt-1, does not induce the morphological transformation of C57MG cells and has no effect on β -catenin levels (13, 14). Expression of *WISP-1* was up-regulated approximately 3-fold in the C57MG/Wnt-1 cell line and *WISP-2* by approximately 5-fold by both Northern analysis and reverse transcription-PCR.

An independent, but similar, system was used to examine *WISP* expression after Wnt-1 induction. C57MG cells expressing the *Wnt-1* gene under the control of a tetracycline-repressible promoter produce low amounts of Wnt-1 in the repressed state but show a strong induction of *Wnt-1* mRNA and protein within 24 hr after tetracycline removal (8). The levels of Wnt-1 and *WISP* RNA isolated from these cells at various times after tetracycline removal were assessed by quantitative PCR. Strong induction of Wnt-1 mRNA was seen as early as 10 hr after tetracycline removal. Induction of *WISP* mRNA (2- to 6-fold) was seen at 48 and 72 hr (data not shown). These data support our previous observations that show that *WISP* induction is correlated with Wnt-1 expression. Because the induction is slow, occurring after approximately 48 hr, the induction of *WISPs* may be an indirect response to Wnt-1 signaling.

cDNA clones of human *WISP-1* were isolated and the sequence compared with mouse *WISP-1*. The cDNA sequences of mouse and human *WISP-1* were 1,766 and 2,830 bp in length, respectively, and encode proteins of 367 aa, with predicted relative molecular masses of $\approx 40,000$ (M_r 40 K). Both have hydrophobic N-terminal signal sequences, 38 conserved cysteine residues, and four potential N-linked glycosylation sites and are 84% identical (Fig. 2A).

Full-length cDNA clones of mouse and human *WISP-2* were 1,734 and 1,293 bp in length, respectively, and encode proteins of 251 and 250 aa, respectively, with predicted relative molecular masses of $\approx 27,000$ (M_r 27 K) (Fig. 2B). Mouse and human *WISP-2* are 73% identical. Human *WISP-2* has no potential N-linked glycosylation sites, and mouse *WISP-2* has one at

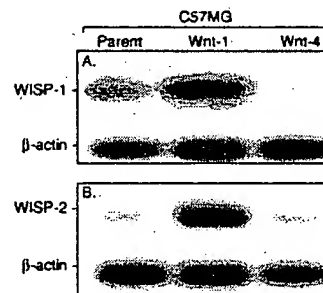


Fig. 1. *WISP-1* and *WISP-2* are induced by Wnt-1, but not Wnt-4, expression in C57MG cells. Northern analysis of *WISP-1* (A) and *WISP-2* (B) expression in C57MG, C57MG/Wnt-1, and C57MG/Wnt-4 cells. Poly(A)⁺ RNA (2 μ g) was subjected to Northern blot analysis and hybridized with a 70-bp mouse *WISP-1*-specific probe (amino acids 278–300) or a 190-bp *WISP-2*-specific probe (nucleotides 1438–1627) in the 3' untranslated region. Blots were rehybridized with human β -actin probe.

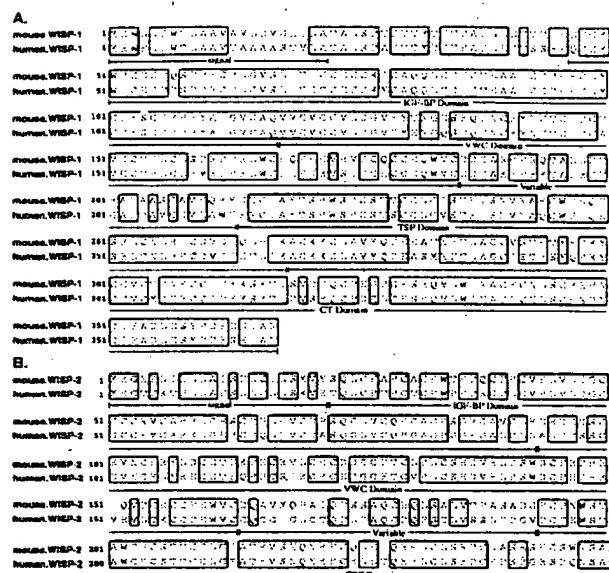


FIG. 2. Encoded amino acid sequence alignment of mouse and human *WISP-1* (A) and mouse and human *WISP-2* (B). The potential signal sequence, insulin-like growth factor-binding protein (IGF-BP), VWC, thrombospondin (TSP), and C-terminal (CT) domains are underlined.

position 197. *WISP-2* has 28 cysteine residues that are conserved among the 38 cysteines found in *WISP-1*.

Identification of *WISP-3*. To search for related proteins, we screened expressed sequence tag (EST) databases with the *WISP-1* protein sequence and identified several ESTs as potentially related sequences. We identified a homologous protein that we have called *WISP-3*. A full-length human *WISP-3* cDNA of 1,371 bp was isolated corresponding to those ESTs that encode a 354-aa protein with a predicted molecular mass of 39,293. *WISP-3* has two potential N-linked glycosylation sites and 36 cysteine residues. An alignment of the three human *WISP* proteins shows that *WISP-1* and *WISP-3* are the most similar (42% identity), whereas *WISP-2* has 37% identity with *WISP-1* and 32% identity with *WISP-3* (Fig. 3A).

***WISPs* Are Homologous to the CTGF Family of Proteins.** Human *WISP-1*, *WISP-2*, and *WISP-3* are novel sequences; however, mouse *WISP-1* is the same as the recently identified *Elm1* gene. *Elm1* is expressed in low, but not high, metastatic mouse melanoma cells, and suppresses the *in vivo* growth and metastatic potential of K-1735 mouse melanoma cells (15). Human and mouse *WISP-2* are homologous to the recently described rat gene, *rCop-1* (16). Significant homology (36–44%) was seen to the CCN family of growth factors. This family includes three members, CTGF, Cyr61, and the protooncogene *nov*. CTGF is a chemotactic and mitogenic factor for fibroblasts that is implicated in wound healing and fibrotic disorders and is induced by TGF- β (17). Cyr61 is an extracellular matrix signaling molecule that promotes cell adhesion, proliferation, migration, angiogenesis, and tumor growth (18, 19). *nov* (nephroblastoma overexpressed) is an immediate early gene associated with quiescence and found altered in Wilms tumors (20). The proteins of the CCN family share functional, but not sequence, similarity to Wnt-1. All are secreted, cysteine-rich heparin binding glycoproteins that associate with the cell surface and extracellular matrix.

WISP proteins exhibit the modular architecture of the CCN family, characterized by four conserved cysteine-rich domains (Fig. 3B) (21). The N-terminal domain, which includes the first 12 cysteine residues, contains a consensus sequence (GCGC-CXXC) conserved in most insulin-like growth factor (IGF)-

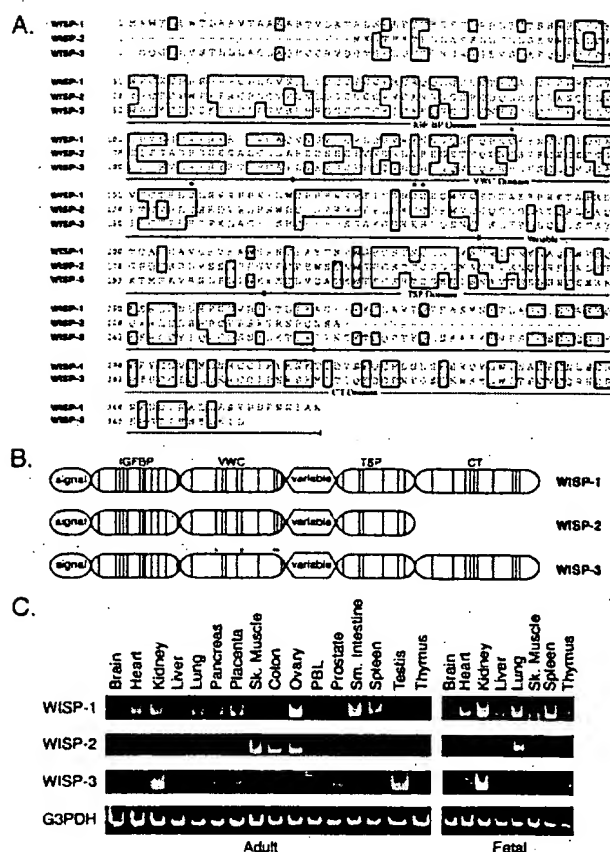


FIG. 3. (A) Encoded amino acid sequence alignment of human *WISPs*. The cysteine residues of *WISP-1* and *WISP-2* that are not present in *WISP-3* are indicated with a dot. (B) Schematic representation of the *WISP* proteins showing the domain structure and cysteine residues (vertical lines). The four cysteine residues in the VWC domain that are absent in *WISP-3* are indicated with a dot. (C) Expression of *WISP* mRNA in human tissues. PCR was performed on human multiple-tissue cDNA panels (CLONTECH) from the indicated adult and fetal tissues.

binding proteins (BP). This sequence is conserved in *WISP-2* and *WISP-3*, whereas *WISP-1* has a glutamine in the third position instead of a glycine. CTGF recently has been shown to specifically bind IGF (22) and a truncated *nov* protein lacking the IGF-BP domain is oncogenic (23). The von Willebrand factor type C module (VWC), also found in certain collagens and mucins, covers the next 10 cysteine residues, and is thought to participate in protein complex formation and oligomerization (24). The VWC domain of *WISP-3* differs from all CCN family members described previously, in that it contains only six of the 10 cysteine residues (Fig. 3A and B). A short variable region follows the VWC domain. The third module, the thrombospondin (TSP) domain is involved in binding to sulfated glycoconjugates and contains six cysteine residues and a conserved WSXCSXCG motif first identified in thrombospondin (25). The C-terminal (CT) module containing the remaining 10 cysteines is thought to be involved in dimerization and receptor binding (26). The CT domain is present in all CCN family members described to date but is absent in *WISP-2* (Fig. 3A and B). The existence of a putative signal sequence and the absence of a transmembrane domain suggest that *WISPs* are secreted proteins, an observation supported by an analysis of their expression and secretion from mammalian cell and baculovirus cultures (data not shown).

Expression of *WISP* mRNA in Human Tissues. Tissue-specific expression of human *WISPs* was characterized by PCR

analysis on adult and fetal multiple tissue cDNA panels. *WISP-1* expression was seen in the adult heart, kidney, lung, pancreas, placenta, ovary, small intestine, and spleen (Fig. 3C). Little or no expression was detected in the brain, liver, skeletal muscle, colon, peripheral blood leukocytes, prostate, testis, or thymus. *WISP-2* had a more restricted tissue expression and was detected in adult skeletal muscle, colon, ovary, and fetal lung. Predominant expression of *WISP-3* was seen in adult kidney and testis and fetal kidney. Lower levels of *WISP-3* expression were detected in placenta, ovary, prostate, and small intestine.

In Situ Localization of *WISP-1* and *WISP-2*. Expression of *WISP-1* and *WISP-2* was assessed by *in situ* hybridization in mammary tumors from Wnt-1 transgenic mice. Strong expression of *WISP-1* was observed in stromal fibroblasts lying within the fibrovascular tumor stroma (Fig. 4 A–D). However, low-level *WISP-1* expression also was observed focally within tumor cells (data not shown). No expression was observed in normal breast. Like *WISP-1*, *WISP-2* expression also was seen in the tumor stroma in breast tumors from Wnt-1 transgenic animals (Fig. 4 E–H). However, *WISP-2* expression in the stroma was in spindle-shaped cells adjacent to capillary vessels, whereas

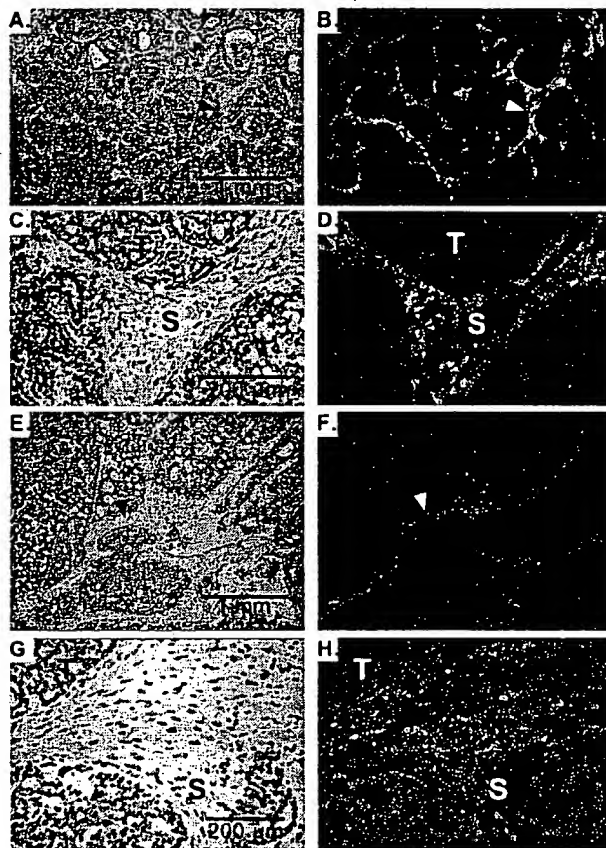


FIG. 4. (A, C, E, and G) Representative hematoxylin/eosin-stained images from breast tumors in Wnt-1 transgenic mice. The corresponding dark-field images showing *WISP-1* expression are shown in B and D. The tumor is a moderately well-differentiated adenocarcinoma showing evidence of adenoid cystic change. At low power (A and B), expression of *WISP-1* is seen in the delicate branching fibrovascular tumor stroma (arrowhead). At higher magnification, expression is seen in the stromal(s) fibroblasts (C and D), and tumor cells are negative. Focal expression of *WISP-1*, however, was observed in tumor cells in some areas. Images of *WISP-2* expression are shown in E–H. At low power (E and F), expression of *WISP-2* is seen in cells lying within the fibrovascular tumor stroma. At higher magnification, these cells appeared to be adjacent to capillary vessels whereas tumor cells are negative (G and H).

the predominant cell type expressing *WISP-1* was the stromal fibroblasts.

Chromosome Localization of the *WISP* Genes. The chromosomal location of the human *WISP* genes was determined by radiation hybrid mapping panels. *WISP-1* is approximately 3.48 cR from the meiotic marker AFM259xc5 [logarithm of odds (lod) score 16.31] on chromosome 8q24.1 to 8q24.3, in the same region as the human locus of the *novH* family member (27) and roughly 4 Mbs distal to *c-myc* (28). Preliminary fine mapping indicates that *WISP-1* is located near D8S1712 STS. *WISP-2* is linked to the marker SHGC-33922 (lod = 1,000) on chromosome 20q12–20q13.1. Human *WISP-3* mapped to chromosome 6q22–6q23 and is linked to the marker AFM211ze5 (lod = 1,000). *WISP-3* is approximately 18 Mbs proximal to CTGF and 23 Mbs proximal to the human cellular oncogene *MYB* (27, 29).

Amplification and Aberrant Expression of *WISPs* in Human Colon Tumors. Amplification of protooncogenes is seen in many human tumors and has etiological and prognostic significance. For example, in a variety of tumor types, *c-myc* amplification has been associated with malignant progression and poor prognosis (30). Because *WISP-1* resides in the same general chromosomal location (8q24) as *c-myc*, we asked whether it was a target of gene amplification, and, if so, whether this amplification was independent of the *c-myc* locus. Genomic DNA from human colon cancer cell lines was assessed by quantitative PCR and Southern blot analysis. (Fig. 5 A and B). Both methods detected similar degrees of *WISP-1* amplification. Most cell lines showed significant (2- to 4-fold) amplification, with the HT-29 and WiDr cell lines demonstrating an 8-fold increase. Significantly, the pattern of amplification observed did not correlate with that observed for *c-myc*, indicating that the *c-myc* gene is not part of the amplicon that involves the *WISP-1* locus.

We next examined whether the *WISP* genes were amplified in a panel of 25 primary human colon adenocarcinomas. The relative *WISP* gene copy number in each colon tumor DNA was compared with pooled normal DNA from 10 donors by quantitative PCR (Fig. 6). The copy number of *WISP-1* and *WISP-2* was significantly greater than one, approximately 2-fold for *WISP-1* in about 60% of the tumors and 2- to 4-fold for *WISP-2* in 92% of the tumors ($P < 0.001$ for each). The copy number for *WISP-3* was indistinguishable from one ($P = 0.166$). In addition, the copy number of *WISP-2* was significantly higher than that of *WISP-1* ($P < 0.001$).

The levels of *WISP* transcripts in RNA isolated from 19 adenocarcinomas and their matched normal mucosa were

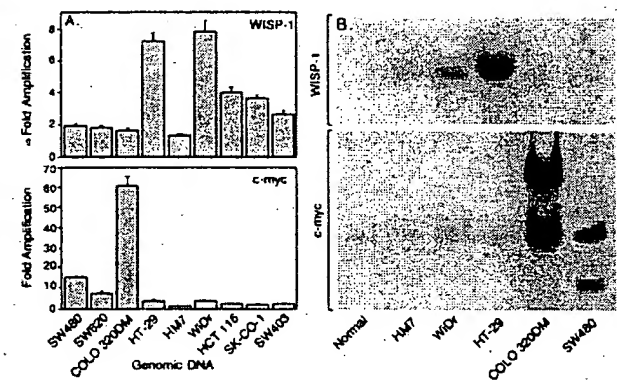


FIG. 5. Amplification of *WISP-1* genomic DNA in colon cancer cell lines. (A) Amplification in cell line DNA was determined by quantitative PCR. (B) Southern blots containing genomic DNA (10 μ g) digested with *Eco*RI (*WISP-1*) or *Xba*I (*c-myc*) were hybridized with a 100-bp human *WISP-1* probe (amino acids 186–219) or a human *c-myc* probe (located at bp 1901–2000). The *WISP* and *myc* genes are detected in normal human genomic DNA after a longer film exposure.

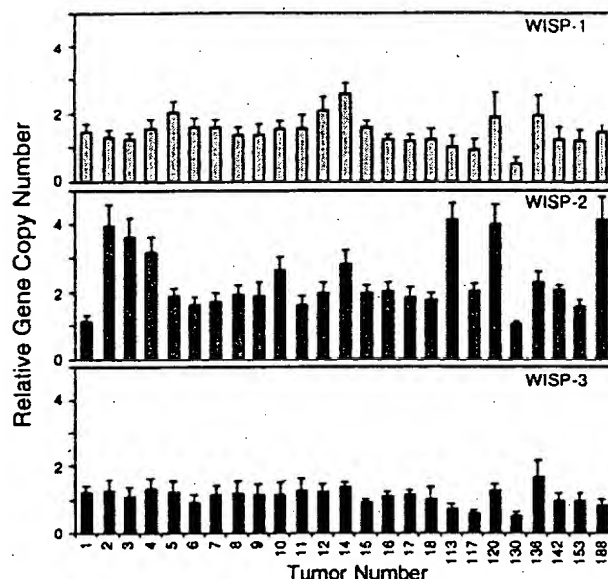


FIG. 6. Genomic amplification of *WISP* genes in human colon tumors. The relative gene copy number of the *WISP* genes in 25 adenocarcinomas was assayed by quantitative PCR, by comparing DNA from primary human tumors with pooled DNA from 10 healthy donors. The data are means \pm SEM from one experiment done in triplicate. The experiment was repeated at least three times.

assessed by quantitative PCR (Fig. 7). The level of *WISP-1* RNA present in tumor tissue varied but was significantly increased (2- to >25-fold) in 84% (16/19) of the human colon tumors examined compared with normal adjacent mucosa. Four of 19 tumors showed greater than 10-fold overexpression. In contrast, in 79% (15/19) of the tumors examined, *WISP-2* RNA expression was significantly lower in the tumor than the mucosa. Similar to *WISP-1*, *WISP-3* RNA was overexpressed in 63% (12/19) of the colon tumors compared with the normal

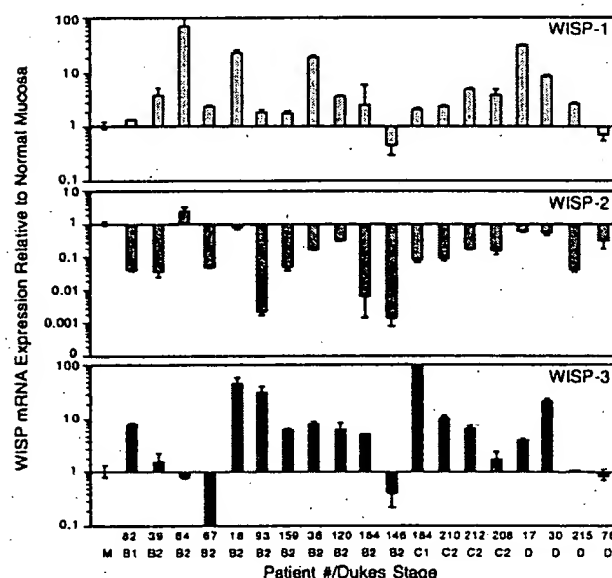


FIG. 7. *WISP* RNA expression in primary human colon tumors relative to expression in normal mucosa from the same patient. Expression of *WISP* mRNA in 19 adenocarcinomas was assayed by quantitative PCR. The Dukes stage of the tumor is listed under the sample number. The data are means \pm SEM from one experiment done in triplicate. The experiment was repeated at least twice.

mucosa. The amount of overexpression of *WISP-3* ranged from 4- to >40-fold.

DISCUSSION

One approach to understanding the molecular basis of cancer is to identify differences in gene expression between cancer cells and normal cells. Strategies based on assumptions that steady-state mRNA levels will differ between normal and malignant cells have been used to clone differentially expressed genes (31). We have used a PCR-based selection strategy, SSH, to identify genes selectively expressed in C57MG mouse mammary epithelial cells transformed by Wnt-1.

Three of the genes isolated, *WISP-1*, *WISP-2*, and *WISP-3*, are members of the CCN family of growth factors, which includes CTGF, Cyr61, and *nov*, a family not previously linked to Wnt signaling.

Two independent experimental systems demonstrated that *WISP* induction was associated with the expression of Wnt-1. The first was C57MG cells infected with a Wnt-1 retroviral vector or C57MG cells expressing Wnt-1 under the control of a tetracycline-repressible promoter, and the second was in Wnt-1 transgenic mice, where breast tissue expresses Wnt-1, whereas normal breast tissue does not. No *WISP* RNA expression was detected in mammary tumors induced by polyoma virus middle T antigen (data not shown). These data suggest a link between Wnt-1 and *WISPs* in that in these two situations, *WISP* induction was correlated with Wnt-1 expression.

It is not clear whether the *WISPs* are directly or indirectly induced by the downstream components of the Wnt-1 signaling pathway (i.e., β -catenin-TCF-1/Lef1). The increased levels of *WISP* RNA were measured in Wnt-1-transformed cells, hours or days after Wnt-1 transformation. Thus, *WISP* expression could result from Wnt-1 signaling directly through β -catenin transcription factor regulation or alternatively through Wnt-1 signaling turning on a transcription factor, which in turn regulates *WISPs*.

The *WISPs* define an additional subfamily of the CCN family of growth factors. One striking difference observed in the protein sequence of *WISP-2* is the absence of a CT domain, which is present in CTGF, Cyr61, *nov*, *WISP-1*, and *WISP-3*. This domain is thought to be involved in receptor binding and dimerization. Growth factors, such as TGF- β , platelet-derived growth factor, and nerve growth factor, which contain a cystine knot motif exist as dimers (32). It is tempting to speculate that *WISP-1* and *WISP-3* may exist as dimers, whereas *WISP-2* exists as a monomer. If the CT domain is also important for receptor binding, *WISP-2* may bind its receptor through a different region of the molecule than the other CCN family members. No specific receptors have been identified for CTGF or *nov*. A recent report has shown that integrin $\alpha_v\beta_3$ serves as an adhesion receptor for Cyr61 (33).

The strong expression of *WISP-1* and *WISP-2* in cells lying within the fibrovascular tumor stroma in breast tumors from Wnt-1 transgenic animals is consistent with previous observations that transcripts for the related CTGF gene are primarily expressed in the fibrous stroma of mammary tumors (34). Epithelial cells are thought to control the proliferation of connective tissue stroma in mammary tumors by a cascade of growth factor signals similar to that controlling connective tissue formation during wound repair. It has been proposed that mammary tumor cells or inflammatory cells at the tumor interstitial interface secrete TGF- β 1, which is the stimulus for stromal proliferation (34). TGF- β 1 is secreted by a large percentage of malignant breast tumors and may be one of the growth factors that stimulates the production of CTGF and *WISPs* in the stroma.

It was of interest that *WISP-1* and *WISP-2* expression was observed in the stromal cells that surrounded the tumor cells

(epithelial cells) in the Wnt-1 transgenic mouse sections of breast tissue. This finding suggests that paracrine signaling could occur in which the stromal cells could supply WISP-1 and WISP-2 to regulate tumor cell growth on the WISP extracellular matrix. Stromal cell-derived factors in the extracellular matrix have been postulated to play a role in tumor cell migration and proliferation (35). The localization of WISP-1 and WISP-2 in the stromal cells of breast tumors supports this paracrine model.

An analysis of WISP-1 gene amplification and expression in human colon tumors showed a correlation between DNA amplification and overexpression, whereas overexpression of WISP-3 RNA was seen in the absence of DNA amplification. In contrast, WISP-2 DNA was amplified in the colon tumors, but its mRNA expression was significantly reduced in the majority of tumors compared with the expression in normal colonic mucosa from the same patient. The gene for human WISP-2 was localized to chromosome 20q12-20q13, at a region frequently amplified and associated with poor prognosis in node negative breast cancer and many colon cancers, suggesting the existence of one or more oncogenes at this locus (36-38). Because the center of the 20q13 amplicon has not yet been identified, it is possible that the apparent amplification observed for WISP-2 may be caused by another gene in this amplicon.

A recent manuscript on *rCop-1*, the rat orthologue of WISP-2, describes the loss of expression of this gene after cell transformation, suggesting it may be a negative regulator of growth in cell lines (16). Although the mechanism by which WISP-2 RNA expression is down-regulated during malignant transformation is unknown, the reduced expression of WISP-2 in colon tumors and cell lines suggests that it may function as a tumor suppressor. These results show that the WISP genes are aberrantly expressed in colon cancer and suggest that their altered expression may confer selective growth advantage to the tumor.

Members of the Wnt signaling pathway have been implicated in the pathogenesis of colon cancer, breast cancer, and melanoma, including the tumor suppressor gene adenomatous polyposis coli and β -catenin (39). Mutations in specific regions of either gene can cause the stabilization and accumulation of cytoplasmic β -catenin, which presumably contributes to human carcinogenesis through the activation of target genes such as the WISPs. Although the mechanism by which Wnt-1 transforms cells and induces tumorigenesis is unknown, the identification of WISPs as genes that may be regulated downstream of Wnt-1 in C57MG cells suggests they could be important mediators of Wnt-1 transformation. The amplification and altered expression patterns of the WISPs in human colon tumors may indicate an important role for these genes in tumor development.

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- Cadigan, K. M. & Nusse, R. (1997) *Genes Dev.* 11, 3286-3305.
- Dale, T. C. (1998) *Biochem. J.* 329, 209-223.
- Nusse, R. & Varmus, H. E. (1982) *Cell* 31, 99-109.
- van Ooyen, A. & Nusse, R. (1984) *Cell* 39, 233-240.
- Tsukamoto, A. S., Grosschedl, R., Guzman, R. C., Parslow, T. & Varmus, H. E. (1988) *Cell* 55, 619-625.
- Brown, J. D. & Moon, R. T. (1998) *Curr. Opin. Cell Biol.* 10, 182-187.
- Molenaar, M., van de Wetering, M., Oosterwégel, M., Peterson-Maduro, J., Godsave, S., Korinek, V., Roose, J., Destree, O. & Clevers, H. (1996) *Cell* 86, 391-399.
- Korinek, V., Barker, N., Willert, K., Molenaar, M., Roose, J., Wagenaar, G., Markman, M., Lamers, W., Destree, O. & Clevers, H. (1998) *Mol. Cell Biol.* 18, 1248-1256.
- Munemitsu, S., Albert, I., Souza, B., Rubinfeld, B. & Polakis, P. (1995) *Proc. Natl. Acad. Sci. USA* 92, 3046-3050.
- He, T. C., Sparks, A. B., Rago, C., Hermeking, H., Zawel, L., da Costa, L. T., Morin, P. J., Vogelstein, B. & Kinzler, K. W. (1998) *Science* 281, 1509-1512.
- Diatchenko, L., Lau, Y. F., Campbell, A. P., Chenchik, A., Moqadam, F., Huang, B., Lukyanov, S., Lukyanov, K., Gurskaya, N., Sverdlov, E. D. & Siebert, P. D. (1996) *Proc. Natl. Acad. Sci. USA* 93, 6025-6030.
- Brown, A. M., Wildin, R. S., Prendergast, T. J. & Varmus, H. E. (1986) *Cell* 46, 1001-1009.
- Wong, G. T., Gavin, B. J. & McMahon, A. P. (1994) *Mol. Cell Biol.* 14, 6278-6286.
- Shimizu, H., Julius, M. A., Giarre, M., Zheng, Z., Brown, A. M. & Kitajewski, J. (1997) *Cell Growth Differ.* 8, 1349-1358.
- Hashimoto, Y., Shindo-Okada, N., Tani, M., Nagamachi, Y., Takeuchi, K., Shiroishi, T., Toma, H. & Yokota, J. (1998) *J. Exp. Med.* 187, 289-296.
- Zhang, R., Averboukh, L., Zhu, W., Zhang, H., Jo, H., Dempsey, P. J., Coffey, R. J., Pardee, A. B. & Liang, P. (1998) *Mol. Cell Biol.* 18, 6131-6141.
- Grotendorst, G. R. (1997) *Cytokine Growth Factor Rev.* 8, 171-179.
- Kireeva, M. L., Mo, F. E., Yang, G. P. & Lau, L. F. (1996) *Mol. Cell Biol.* 16, 1326-1334.
- Babic, A. M., Kireeva, M. L., Kolesnikova, T. V. & Lau, L. F. (1998) *Proc. Natl. Acad. Sci. USA* 95, 6355-6360.
- Martinerie, C., Huff, V., Joubert, I., Badzioch, M., Saunders, G., Strong, L. & Perbal, B. (1994) *Oncogene* 9, 2729-2732.
- Bork, P. (1993) *FEBS Lett.* 327, 125-130.
- Kim, H. S., Nagalla, S. R., Oh, Y., Wilson, E., Roberts, C. T., Jr. & Rosenfeld, R. G. (1997) *Proc. Natl. Acad. Sci. USA* 94, 12981-12986.
- Joliet, V., Martinerie, C., Dambrine, G., Plassiart, G., Brisac, M., Crochet, J. & Perbal, B. (1992) *Mol. Cell Biol.* 12, 10-21.
- Mancuso, D. J., Tuley, E. A., Westfield, L. A., Worrall, N. K., Shelton-Inloes, B. B., Sorace, J. M., Alevy, Y. G. & Sadler, J. E. (1989) *J. Biol. Chem.* 264, 19514-19527.
- Holt, G. D., Pangburn, M. K. & Ginsburg, V. (1990) *J. Biol. Chem.* 265, 2852-2855.
- Voorberg, J., Fontijn, R., Calafat, J., Janssen, H., van Mourik, J. A. & Pannekoek, H. (1991) *J. Cell Biol.* 113, 195-205.
- Martinerie, C., Viegas-Pequignot, E., Guenard, I., Dutrillaux, B., Nguyen, V. C., Bernheim, A. & Perbal, B. (1992) *Oncogene* 7, 2529-2534.
- Takahashi, E., Hori, T., O'Connell, P., Leppert, M. & White, R. (1991) *Cytogenet. Cell. Genet.* 57, 109-111.
- Meesse, E., Meltzer, P. S., Witkowski, C. M. & Trent, J. M. (1989) *Genes Chromosomes Cancer* 1, 88-94.
- Garte, S. J. (1993) *Crit. Rev. Oncog.* 4, 435-449.
- Zhang, L., Zhou, W., Velculescu, V. E., Kern, S. E., Hruban, R. H., Hamilton, S. R., Vogelstein, B. & Kinzler, K. W. (1997) *Science* 276, 1268-1272.
- Sun, P. D. & Davies, D. R. (1995) *Annu. Rev. Biophys. Biomol. Struct.* 24, 269-291.
- Kireeva, M. L., Lam, S. C. T. & Lau, L. F. (1998) *J. Biol. Chem.* 273, 3090-3096.
- Frazier, K. S. & Grotendorst, G. R. (1997) *Int. J. Biochem. Cell Biol.* 29, 153-161.
- Wernert, N. (1997) *Virchows Arch.* 430, 433-443.
- Tanner, M. M., Tirkkonen, M., Kallioniemi, A., Collins, C., Stokke, T., Karhu, R., Kowbel, D., Shadravan, F., Hintz, M., Kuo, W. L., et al. (1994) *Cancer Res.* 54, 4257-4260.
- Brinkmann, U., Gallo, M., Polymeropoulos, M. H. & Pastan, I. (1996) *Genome Res.* 6, 187-194.
- Bischoff, J. R., Anderson, L., Zhu, Y., Mossie, K., Ng, L., Souza, B., Schryver, B., Flanagan, P., Clairvoyant, F., Ginther, C., et al. (1998) *EMBO J.* 17, 3052-3065.
- Morin, P. J., Sparks, A. B., Korinek, V., Barker, N., Clevers, H., Vogelstein, B. & Kinzler, K. W. (1997) *Science* 275, 1787-1790.
- Lu, L. H. & Gillett, N. (1994) *Cell Vision* 1, 169-176.

Genomic amplification of a decoy receptor for Fas ligand in lung and colon cancer

Robert M. Pitti[†], Scot A. Marsters^{††}, David A. Lawrence^{††}, Margaret Roy^{*}, Frank C. Kischkel^{*}, Patrick Dowd^{*}, Arthur Huang^{*}, Christopher J. Donahue^{*}, Steven W. Sherwood^{*}, Daryl T. Baldwin^{*}, Paul J. Godowski^{*}, William I. Wood^{*}, Austin L. Gurney^{*}, Kenneth J. Hillan^{*}, Robert L. Cohen^{*}, Audrey D. Goddard^{*}, David Botstein[‡] & Avi Ashkenazi^{*}

^{*} Departments of Molecular Oncology, Molecular Biology, and Immunology, Genentech Inc., 1 DNA Way, South San Francisco, California 94080, USA

[‡] Department of Genetics, Stanford University, Stanford, California 94305, USA

[†] These authors contributed equally to this work

Fas ligand (FasL) is produced by activated T cells and natural killer cells and it induces apoptosis (programmed cell death) in target cells through the death receptor Fas/Apo1/CD95 (ref. 1). One important role of FasL and Fas is to mediate immune-cytotoxic killing of cells that are potentially harmful to the organism, such as virus-infected or tumour cells¹. Here we report the discovery of a soluble decoy receptor, termed decoy receptor 3 (DcR3), that binds to FasL and inhibits FasL-induced apoptosis. The DcR3 gene was amplified in about half of 35 primary lung and colon tumours studied, and DcR3 messenger RNA was expressed in malignant tissue. Thus, certain tumours may escape FasL-dependent immune-cytotoxic attack by expressing a decoy receptor that blocks FasL.

By searching expressed sequence tag (EST) databases, we identified a set of related ESTs that showed homology to the tumour necrosis factor (TNF) receptor (TNFR) gene superfamily². Using the overlapping sequence, we isolated a previously unknown full-length complementary DNA from human fetal lung. We named the protein encoded by this cDNA decoy receptor 3 (DcR3). The cDNA encodes a 300-amino-acid polypeptide that resembles members of the TNFR family (Fig. 1a): the amino terminus contains a leader sequence, which is followed by four tandem cysteine-rich domains (CRDs). Like one other TNFR homologue, osteoprotegerin (OPG)³, DcR3 lacks an apparent transmembrane sequence, which indicates that it may be a secreted, rather than a membrane-associated, molecule. We expressed a recombinant, histidine-tagged form of DcR3 in mammalian cells; DcR3 was secreted into the cell culture medium, and migrated on polyacrylamide gels as a protein of relative molecular mass 35,000 (data not shown). DcR3 shares sequence identity in particular with OPG (31%) and TNFR2 (29%), and has relatively less homology with Fas (17%). All of the cysteines in the four CRDs of DcR3 and OPG are conserved; however, the carboxy-terminal portion of DcR3 is 101 residues shorter.

We analysed expression of DcR3 mRNA in human tissues by northern blotting (Fig. 1b). We detected a predominant 1.2-kilobase transcript in fetal lung, brain, and liver, and in adult spleen, colon and lung. In addition, we observed relatively high DcR3 mRNA expression in the human colon carcinoma cell line SW480.

To investigate potential ligand interactions of DcR3, we generated a recombinant, Fc-tagged DcR3 protein. We tested binding of DcR3-Fc to human 293 cells transfected with individual TNF-family ligands, which are expressed as type 2 transmembrane proteins (these transmembrane proteins have their N termini in the cytosol). DcR3-Fc showed a significant increase in binding to cells transfected with FasL⁴ (Fig. 2a), but not to cells transfected with TNF⁵, Apo2L/TRAIL^{6,7}, Apo3L/TWEAK^{8,9}, or OPG/TRANCE/

methods. Peptides AENK or AEQK were dissolved in water, made isotonic with NaCl and diluted into RPMI growth medium. T-cell-proliferation assays were done essentially as described^{20,21}. Briefly, after antigen pulsing (30 µg ml⁻¹ TTCF) with tetrapeptides (1–2 mg ml⁻¹), PBMCs or EBV-B cells were washed in PBS and fixed for 45 s in 0.05% glutaraldehyde. Glycine was added to a final concentration of 0.1M and the cells were washed five times in RPMI 1640 medium containing 1% FCS before co-culture with T-cell clones in round-bottom 96-well microtitre plates. After 48 h, the cultures were pulsed with 1 µCi of ³H-thymidine and harvested for scintillation counting 16 h later. Predigestion of native TTCF was done by incubating 200 µg TTCF with 0.25 µg pig kidney legumain in 500 µl 50 mM citrate buffer, pH 5.5, for 1 h at 37°C. **Glycopeptide digestions.** The peptides HIDNEED1, HIDN(N-glucosamine) EED1 and HIDNESD1, which are based on the TTCF sequence, and QQQLHFGSNVTDCSGNFCLFR(KKK), which is based on human transferrin, were obtained by custom synthesis. The three C-terminal lysine residues were added to the natural sequence to aid solubility. The transferrin glycopeptide QQQLHFGSNVTDCSGNFCLFR was prepared by tryptic (Promega) digestion of 5 mg reduced, carboxy-methylated human transferrin followed by concanavalin A chromatography¹¹. Glycopeptides corresponding to residues 622–642 and 421–452 were isolated by reverse-phase HPLC and identified by mass spectrometry and N-terminal sequencing. The lyophilized transferrin-derived peptides were redissolved in 50 mM sodium acetate, pH 5.5, 10 mM dithiothreitol, 20% methanol. Digestions were performed for 3 h at 30°C with 5–50 mU ml⁻¹ pig kidney legumain or B-cell AEP. Products were analysed by HPLC or MALDI-TOF mass spectrometry using a matrix of 10 mg ml⁻¹ α-cyanocinnamic acid in 50% acetonitrile/0.1% TFA and a PerSeptive Biosystems Elite STR mass spectrometer set to linear or reflector mode. Internal standardization was obtained with a matrix ion of 568.13 mass units.

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- Chen, J. M. *et al.* Cloning, isolation, and characterisation of mammalian legumain, an asparaginyl endopeptidase. *J. Biol. Chem.* 272, 8090–8098 (1997).
- Kembhavi, A. A., Buttle, D. J., Knight, C. G. & Barrett, A. J. The two cysteine endopeptidases of legume seeds: purification and characterization by use of specific fluorometric assays. *Arch. Biochem. Biophys.* 303, 208–213 (1993).
- Dalton, J. P., Hla Jmriska, L. & Bridley, P. J. Asparaginyl endopeptidase activity in adult *Schistosoma mansoni*. *Parasitology* 111, 575–580 (1995).
- Bennett, K. *et al.* Antigen processing for presentation by class II major histocompatibility complex requires cleavage by cathepsin E. *Eur. J. Immunol.* 22, 1519–1524 (1992).
- Riese, R. J. *et al.* Essential role for cathepsin S in MHC class II-associated invariant chain processing and peptide loading. *Immunity* 4, 357–366 (1996).
- Rodriguez, G. M. & Diment, S. Role of cathepsin D in antigen presentation of ovalbumin. *J. Immunol.* 149, 2894–2898 (1992).
- Hewitt, E. W. *et al.* Natural processing sites for human cathepsin E and cathepsin D in tetanus toxin: implications for T cell epitope generation. *J. Immunol.* 159, 4693–4699 (1997).
- Watts, C. Capture and processing of exogenous antigens for presentation on MHC molecules. *Annu. Rev. Immunol.* 15, 821–850 (1997).
- Chapman, H. A. Endosomal proteases and MHC class II function. *Curr. Opin. Immunol.* 10, 93–102 (1998).
- Fineschi, B. & Miller, J. Endosomal proteases and antigen processing. *Trends Biochem. Sci.* 22, 377–382 (1997).
- Lu, J. & van Halbeek, H. Complete ¹H and ¹³C resonance assignments of a 21-amino acid glycopeptide prepared from human serum transferrin. *Carbohydr. Res.* 296, 1–21 (1996).
- Fearon, D. T. & Locksley, R. M. The instructive role of innate immunity in the acquired immune response. *Science* 272, 50–54 (1996).
- Medzhitov, R. & Janeway, C. A. J. Innate immunity: the virtues of a nonclonal system of recognition. *Cell* 91, 295–298 (1997).
- Wyatt, R. *et al.* The antigenic structure of the HIV gp120 envelope glycoprotein. *Nature* 393, 705–711 (1998).
- Botarelli, P. *et al.* N-glycosylation of HIV gp120 may constrain recognition by T lymphocytes. *J. Immunol.* 147, 3128–3132 (1991).
- Davidson, H. W., West, M. A. & Watts, C. Endocytosis, intracellular trafficking, and processing of membrane IgG and monovalent antigen/membrane IgG complexes in B lymphocytes. *J. Immunol.* 144, 4101–4109 (1990).
- Barrett, A. J. & Kirschke, H. Cathepsin B, cathepsin H and cathepsin L. *Methods Enzymol.* 80, 535–559 (1981).
- Makoff, A. J., Ballantine, S. P., Smallwood, A. E. & Fairweather, N. F. Expression of tetanus toxin fragment C in *E. coli*: its purification and potential use as a vaccine. *Biotechnology* 7, 1043–1046 (1989).
- Lane, D. P. & Harlow, E. *Antibodies: A Laboratory Manual* (Cold Spring Harbor Laboratory Press, 1988).
- Lanzavecchia, A. Antigen-specific interaction between T and B cells. *Nature* 314, 537–539 (1985).
- Pond, L. & Watts, C. Characterization of transport of newly assembled, T cell-stimulatory MHC class II-peptide complexes from MHC class II compartments to the cell surface. *J. Immunol.* 159, 543–553 (1997).

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Correspondence and requests for materials should be addressed to C.W. (e-mail: c.watts@dundee.ac.uk).

RANKL¹⁰⁻¹² (data not shown). DcR3-Fc immunoprecipitated shed FasL from FasL-transfected 293 cells (Fig. 2b) and purified soluble FasL (Fig. 2c), as did the Fc-tagged ectodomain of Fas but not TNFR1. Gel-filtration chromatography showed that DcR3-Fc and soluble FasL formed a stable complex (Fig. 2d). Equilibrium analysis indicated that DcR3-Fc and Fas-Fc bound to soluble FasL with a comparable affinity ($K_d = 0.8 \pm 0.2$ and 1.1 ± 0.1 nM, respectively; Fig. 2e), and that DcR3-Fc could block nearly all of the binding of soluble FasL to Fas-Fc (Fig. 2e, inset). Thus, DcR3 competes with Fas for binding to FasL.

To determine whether binding of DcR3 inhibits FasL activity, we tested the effect of DcR3-Fc on apoptosis induction by soluble FasL in Jurkat T leukaemia cells, which express Fas (Fig. 3a). DcR3-Fc and Fas-Fc blocked soluble-FasL-induced apoptosis in a similar dose-dependent manner, with half-maximal inhibition at $\sim 0.1 \mu\text{g ml}^{-1}$. Time-course analysis showed that the inhibition did not merely delay cell death, but rather persisted for at least 24 hours (Fig. 3b). We also tested the effect of DcR3-Fc on activation-induced cell death (AICD) of mature T lymphocytes, a FasL-dependent process¹. Consistent with previous results¹³, activation of interleukin-2-stimulated CD4-positive T cells with anti-CD3 antibody increased the level of apoptosis twofold, and Fas-Fc blocked this effect substantially (Fig. 3c); DcR3-Fc blocked the

induction of apoptosis to a similar extent. Thus, DcR3 binding blocks apoptosis induction by FasL.

FasL-induced apoptosis is important in elimination of virus-infected cells and cancer cells by natural killer cells and cytotoxic T lymphocytes; an alternative mechanism involves perforin and granzymes^{1,14-16}. Peripheral blood natural killer cells triggered marked cell death in Jurkat T leukaemia cells (Fig. 3d); DcR3-Fc and Fas-Fc each reduced killing of target cells from $\sim 65\%$ to $\sim 30\%$, with half-maximal inhibition at $\sim 1 \mu\text{g ml}^{-1}$; the residual killing was probably mediated by the perforin/granzyme pathway. Thus, DcR3 binding blocks FasL-dependent natural killer cell activity. Higher DcR3-Fc and Fas-Fc concentrations were required to block soluble FasL activity, which is consistent with the greater potency of membrane-associated FasL compared with soluble FasL¹⁷.

Given the role of immune-cytotoxic cells in elimination of tumour cells and the fact that DcR3 can act as an inhibitor of FasL, we proposed that DcR3 expression might contribute to the ability of some tumours to escape immune-cytotoxic attack. As genomic amplification frequently contributes to tumorigenesis, we investigated whether the DcR3 gene is amplified in cancer. We analysed DcR3 gene-copy number by quantitative polymerase chain

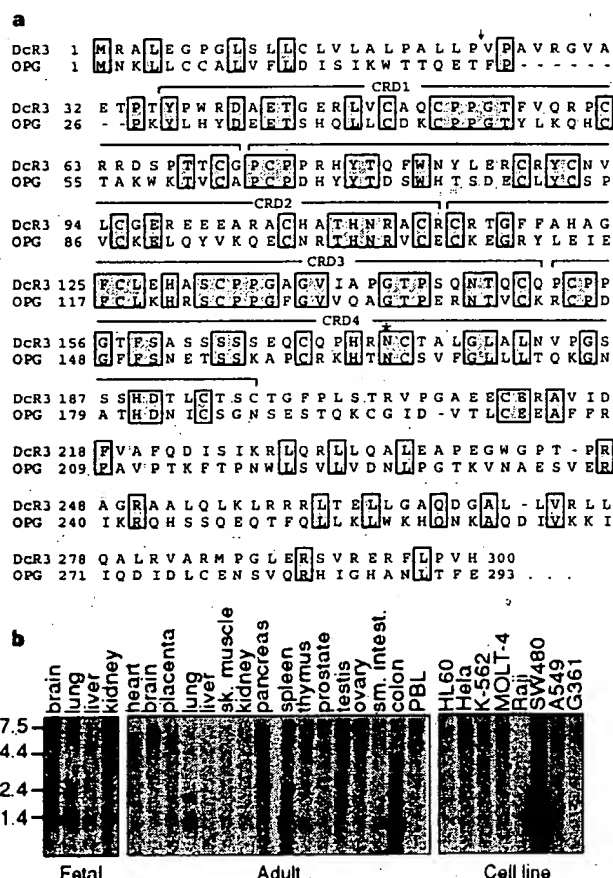


Figure 1 Primary structure and expression of human DcR3. **a**, Alignment of the amino-acid sequences of DcR3 and of osteoprotegerin (OPG); the C-terminal 101 residues of OPG are not shown. The putative signal cleavage site (arrow), the cysteine-rich domains (CRD 1-4), and the N-linked glycosylation site (asterisk) are shown. **b**, Expression of DcR3 mRNA. Northern hybridization analysis was done using the DcR3 cDNA as a probe and blots of poly(A)⁺ RNA (Clontech) from human fetal and adult tissues or cancer cell lines. PBL, peripheral blood lymphocyte.

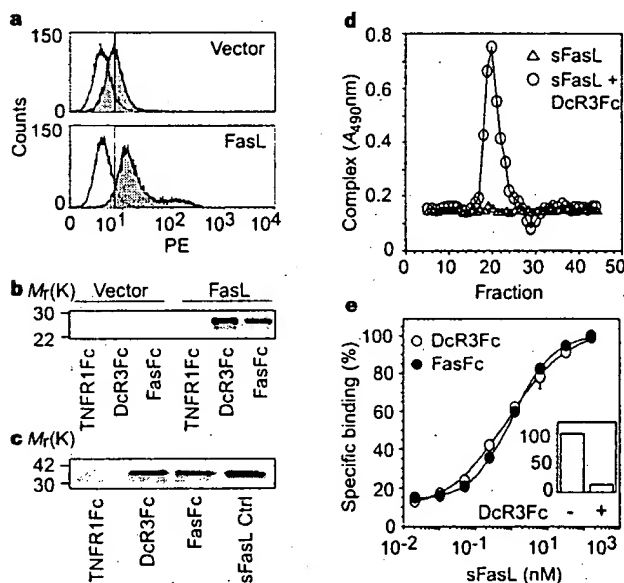


Figure 2 Interaction of DcR3 with FasL. **a**, 293 cells were transfected with pRK5 vector (top) or with pRK5 encoding full-length FasL (bottom), incubated with DcR3-Fc (solid line, shaded area), TNFR1-Fc (dotted line) or buffer control (dashed line) (the dashed and dotted lines overlap), and analysed for binding by FACS. Statistical analysis showed a significant difference ($P < 0.001$) between the binding of DcR3-Fc to cells transfected with FasL or pRK5. PE, phycoerythrin-labelled cells. **b**, 293 cells were transfected as in **a** and metabolically labelled, and cell supernatants were immunoprecipitated with Fc-tagged TNFR1, DcR3 or Fas. **c**, Purified soluble FasL (sFasL) was immunoprecipitated with TNFR1-Fc, DcR3-Fc or Fas-Fc and visualized by immunoblot with anti-FasL antibody. sFasL was loaded directly for comparison in the right-hand lane. **d**, Flag-tagged sFasL was incubated with DcR3-Fc or with buffer and resolved by gel filtration; column fractions were analysed in an assay that detects complexes containing DcR3-Fc and sFasL-Flag. **e**, Equilibrium binding of DcR3-Fc or Fas-Fc to sFasL-Flag. Inset, competition of DcR3-Fc with Fas-Fc for binding to sFasL-Flag.

reaction (PCR)¹⁸ in genomic DNA from 35 primary lung and colon tumours, relative to pooled genomic DNA from peripheral blood leukocytes (PBLs) of 10 healthy donors. Eight of 18 lung tumours and 9 of 17 colon tumours showed DcR3 gene amplification, ranging from 2- to 18-fold (Fig. 4a, b). To confirm this result, we analysed the colon tumour DNAs with three more, independent sets of DcR3-based PCR primers and probes; we observed nearly the same amplification (data not shown).

We then analysed DcR3 mRNA expression in primary tumour tissue sections by *in situ* hybridization. We detected DcR3 expression in 6 out of 15 lung tumours, 2 out of 2 colon tumours, 2 out of 5 breast tumours, and 1 out of 1 gastric tumour (data not shown). A section through a squamous-cell carcinoma of the lung is shown in Fig. 4c. DcR3 mRNA was localized to infiltrating malignant epithelium, but was essentially absent from adjacent stroma, indicating tumour-specific expression. Although the individual tumour specimens that we analysed for mRNA expression and gene amplification were different, the *in situ* hybridization results are consistent with the finding that the DcR3 gene is amplified frequently in tumours. SW480 colon carcinoma cells, which showed abundant DcR3 mRNA expression (Fig. 1b), also had marked DcR3 gene amplification, as shown by quantitative PCR (fourfold) and by Southern blot hybridization (fivefold) (data not shown).

If DcR3 amplification in cancer is functionally relevant, then DcR3 should be amplified more than neighbouring genomic regions that are not important for tumour survival. To test this,

we mapped the human DcR3 gene by radiation-hybrid analysis; DcR3 showed linkage to marker AFM218xe7 (T160), which maps to chromosome position 20q13. Next, we isolated from a bacterial artificial chromosome (BAC) library a human genomic clone that carries DcR3, and sequenced the ends of the clone's insert. We then determined, from the nine colon tumours that showed twofold or greater amplification of DcR3, the copy number of the DcR3-flanking sequences (reverse and forward) from the BAC, and of seven genomic markers that span chromosome 20 (Fig. 4d). The DcR3-linked reverse marker showed an average amplification of roughly threefold, slightly less than the approximately fourfold amplification of DcR3; the other markers showed little or no amplification. These data indicate that DcR3 may be at the 'epicentre' of a distal chromosome 20 region that is amplified in colon cancer, consistent with the possibility that DcR3 amplification promotes tumour survival.

Our results show that DcR3 binds specifically to FasL and inhibits FasL activity. We did not detect DcR3 binding to several other TNF-ligand-family members; however, this does not rule out the possibility that DcR3 interacts with other ligands, as do some other TNFR family members, including OPG^{2,19}.

FasL is important in regulating the immune response; however, little is known about how FasL function is controlled. One mechanism involves the molecule cFLIP, which modulates apoptosis signalling downstream of Fas²⁰. A second mechanism involves proteolytic shedding of FasL from the cell surface¹⁷. DcR3 competes with Fas for

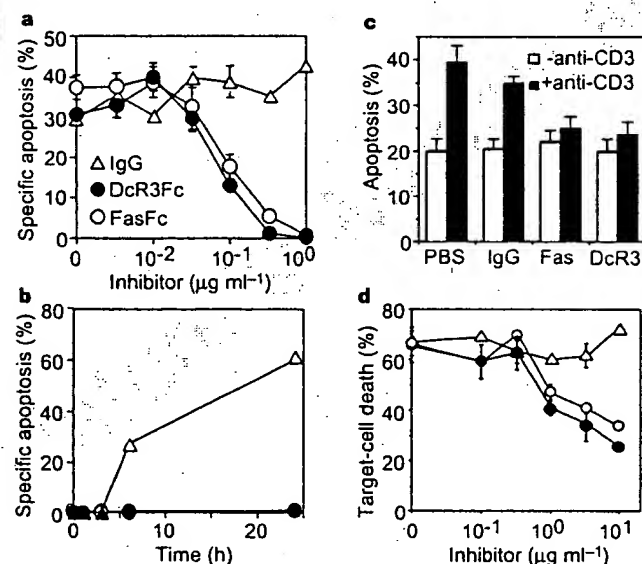


Figure 3 Inhibition of FasL activity by DcR3. **a**, Human Jurkat T leukaemia cells were incubated with Flag-tagged soluble FasL (sFasL; 5 ng ml⁻¹) oligomerized with anti-Flag antibody (0.1 μg ml⁻¹) in the presence of the proposed inhibitors DcR3-Fc, Fas-Fc or human IgG1 and assayed for apoptosis (mean ± s.e.m. of triplicates). **b**, Jurkat cells were incubated with sFasL-Flag plus anti-Flag antibody as in **a**, in presence of 1 μg ml⁻¹ DcR3-Fc (filled circles), Fas-Fc (open circles) or human IgG1 (triangles), and apoptosis was determined at the indicated time points. **c**, Peripheral blood T cells were stimulated with PHA and interleukin-2, followed by control (white bars) or anti-CD3 antibody (filled bars), together with phosphate-buffered saline (PBS), human IgG1, Fas-Fc, or DcR3-Fc (10 μg ml⁻¹). After 16 h, apoptosis of CD4⁺ cells was determined (mean ± s.e.m. of results from five donors). **d**, Peripheral blood natural killer cells were incubated with ⁵¹Cr-labelled Jurkat cells in the presence of DcR3-Fc (filled circles), Fas-Fc (open circles) or human IgG1 (triangles), and target-cell death was determined by release of ⁵¹Cr (mean ± s.d. for two donors, each in triplicate).

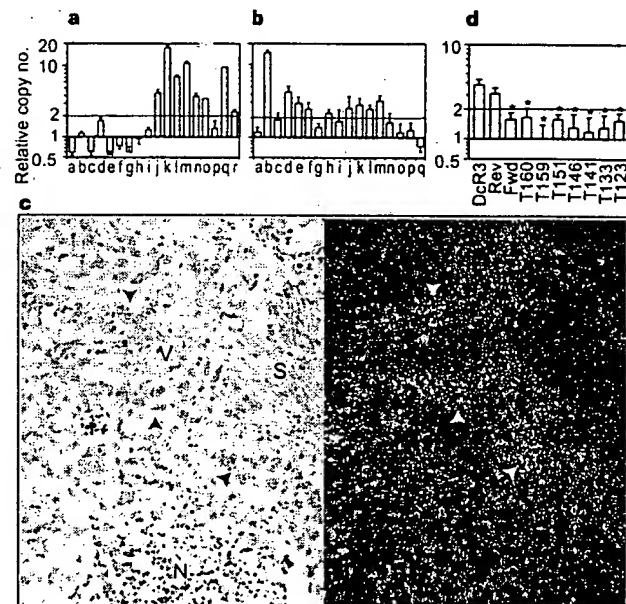


Figure 4 Genomic amplification of DcR3 in tumours. **a**, Lung cancers, comprising eight adenocarcinomas (c, d, f, g, h, j, k, r), seven squamous-cell carcinomas (a, e, m, n, o, p, q), one non-small-cell carcinoma (b), one small-cell carcinoma (i), and one bronchial adenocarcinoma (l). The data are means ± s.d. of 2 experiments done in duplicate. **b**, Colon tumours, comprising 17 adenocarcinomas. Data are means ± s.e.m. of five experiments done in duplicate. **c**, *In situ* hybridization analysis of DcR3 mRNA expression in a squamous-cell carcinoma of the lung. A representative bright-field image (left) and the corresponding dark-field image (right) show DcR3 mRNA over infiltrating malignant epithelium (arrowheads). Adjacent non-malignant stroma (S), blood vessel (V) and necrotic tumour tissue (N) are also shown. **d**, Average amplification of DcR3 compared with amplification of neighbouring genomic regions (reverse and forward, Rev and Fwd), the DcR3-linked marker T160, and other chromosome-20 markers, in the nine colon tumours showing DcR3 amplification of twofold or more (b). Data are from two experiments done in duplicate. Asterisk indicates $P < 0.01$ for a Student's *t*-test comparing each marker with DcR3.

FasL binding; hence, it may represent a third mechanism of extracellular regulation of FasL activity. A decoy receptor that modulates the function of the cytokine interleukin-1 has been described²¹. In addition, two decoy receptors that belong to the TNFR family, DcR1 and DcR2, regulate the FasL-related apoptosis-inducing molecule Apo2L²². Unlike DcR1 and DcR2, which are membrane-associated proteins, DcR3 is directly secreted into the extracellular space. One other secreted TNFR-family member is OPG³, which shares greater sequence homology with DcR3 (31%) than do DcR1 (17%) or DcR2 (19%); OPG functions as a third decoy for Apo2L¹⁹. Thus, DcR3 and OPG define a new subset of TNFR-family members that function as secreted decoys to modulate ligands that induce apoptosis. Pox viruses produce soluble TNFR homologues that neutralize specific TNF-family ligands, thereby modulating the antiviral immune response². Our results indicate that a similar mechanism, namely, production of a soluble decoy receptor for FasL, may contribute to immune evasion by certain tumours. □

Methods

Isolation of DcR3 cDNA. Several overlapping ESTs in GenBank (accession numbers AA025672, AA025673 and W67560) and in LifeseqTM (Incyte Pharmaceuticals; accession numbers 1339238, 1533571, 1533650, 1542861, 1789372 and 2207027) showed similarity to members of the TNFR family. We screened human cDNA libraries by PCR with primers based on the region of EST consensus; fetal lung was positive for a product of the expected size. By hybridization to a PCR-generated probe based on the ESTs, one positive clone (DNA30942) was identified. When searching for potential alternatively spliced forms of DcR3 that might encode a transmembrane protein, we isolated 50 more clones; the coding regions of these clones were identical in size to that of the initial clone (data not shown).

Fc-fusion proteins (immunoadhesins). The entire DcR3 sequence, or the ectodomain of Fas or TNFR1, was fused to the hinge and Fc region of human IgG1, expressed in insect SF9 cells or in human 293 cells, and purified as described²³.

Fluorescence-activated cell sorting (FACS) analysis. We transfected 293 cells using calcium phosphate or Effectene (Qiagen) with pRK5 vector or pRK5 encoding full-length human FasL⁴ (2 µg), together with pRK5 encoding CrmA (2 µg) to prevent cell death. After 16 h, the cells were incubated with biotinylated DcR3-Fc or TNFR1-Fc and then with phycoerythrin-conjugated streptavidin (GibcoBRL), and were assayed by FACS. The data were analysed by Kolmogorov-Smirnov statistical analysis. There was some detectable staining of vector-transfected cells by DcR3-Fc; as these cells express little FasL (data not shown), it is possible that DcR3 recognized some other factor that is expressed constitutively on 293 cells.

Immunoprecipitation. Human 293 cells were transfected as above, and metabolically labelled with [³⁵S]cysteine and [³⁵S]methionine (0.5 mCi; Amersham). After 16 h of culture in the presence of z-VAD-fmk (10 µM), the medium was immunoprecipitated with DcR3-Fc, Fas-Fc or TNFR1-Fc (5 µg), followed by protein A-Sepharose (Repligen). The precipitates were resolved by SDS-PAGE and visualized on a phosphorimager (Fuji BAS2000). Alternatively, purified, Flag-tagged soluble FasL (1 µg) (Alexis) was incubated with each Fc-fusion protein (1 µg), precipitated with protein A-Sepharose, resolved by SDS-PAGE and visualized by immunoblotting with rabbit anti-FasL antibody (Oncogene Research).

Analysis of complex formation. Flag-tagged soluble FasL (25 µg) was incubated with buffer or with DcR3-Fc (40 µg) for 1.5 h at 24 °C. The reaction was loaded onto a Superdex 200 HR 10/30 column (Pharmacia) and developed with PBS; 0.6-ml fractions were collected. The presence of DcR3-Fc-FasL complex in each fraction was analysed by placing 100 µl aliquots into microtitre wells pre-coated with anti-human IgG (Boehringer) to capture DcR3-Fc, followed by detection with biotinylated anti-Flag antibody Bio M2 (Kodak) and streptavidin-horseradish peroxidase (Amersham). Calibration of the column indicated an apparent relative molecular mass of the complex of 420K (data not shown), which is consistent with a stoichiometry of two DcR3-Fc homodimers to two soluble FasL homotrimers.

Equilibrium binding analysis. Microtitre wells were coated with anti-human

IgG, blocked with 2% BSA in PBS. DcR3-Fc or Fas-Fc was added, followed by serially diluted Flag-tagged soluble FasL. Bound ligand was detected with anti-Flag antibody as above. In the competition assay, Fas-Fc was immobilized as above, and the wells were blocked with excess IgG1 before addition of Flag-tagged soluble FasL plus DcR3-Fc.

T-cell AICD. CD3⁺ lymphocytes were isolated from peripheral blood of individual donors using anti-CD3 magnetic beads (Miltenyi Biotech), stimulated with phytohaemagglutinin (PHA; 2 µg ml⁻¹) for 24 h, and cultured in the presence of interleukin-2 (100 U ml⁻¹) for 5 days. The cells were plated in wells coated with anti-CD3 antibody (Pharmingen) and analysed for apoptosis 16 h later by FACS analysis of annexin-V-binding of CD4⁺ cells²⁴.

Natural killer cell activity. Natural killer cells were isolated from peripheral blood of individual donors using anti-CD56 magnetic beads (Miltenyi Biotech), and incubated for 16 h with ⁵¹Cr-loaded Jurkat cells at an effector-to-target ratio of 1:1 in the presence of DcR3-Fc, Fas-Fc or human IgG1. Target-cell death was determined by release of ⁵¹Cr in effector-target co-cultures relative to release of ⁵¹Cr by detergent lysis of equal numbers of Jurkat cells.

Gene-amplification analysis. Surgical specimens were provided by J. Kern (lung tumours) and P. Quirke (colon tumours). Genomic DNA was extracted (Qiagen) and the concentration was determined using Hoechst dye 33258 intercalation fluorometry. Amplification was determined by quantitative PCR¹⁸ using a TaqMan instrument (ABI). The method was validated by comparison of PCR and Southern hybridization data for the Myc and HER-2 oncogenes (data not shown). Gene-specific primers and fluorogenic probes were designed on the basis of the sequence of DcR3 or of nearby regions identified on a BAC carrying the human DcR3 gene; alternatively, primers and probes were based on Stanford Human Genome Center marker AFM218xe7 (T160), which is linked to DcR3 (likelihood score = 5.4), SHGC-36268 (T159), the nearest available marker which maps to ~500 kilobases from T160, and five extra markers that span chromosome 20. The DcR3-specific primer sequences were 5'-CTTCTTCGCGCAGCTG-3' and 5'-ATCAGCCCGCACCAG-3' and the fluorogenic probe sequence was 5'-(FAM-ACACGATGCGTGCTCCAAGCAG Aap-(TAMARA), where FAM is 5'-fluorescein phosphoramidite. Relative gene-copy numbers were derived using the formula 2^(ΔCT), where ΔCT is the difference in amplification cycles required to detect DcR3 in peripheral blood lymphocyte DNA compared to test DNA.

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- Nagata, S. Apoptosis by death factor. *Cell* 88, 355-365 (1997).
- Smith, C. A., Farrah, T. & Goodwin, R. G. The TNF receptor superfamily of cellular and viral proteins: activation, costimulation, and death. *Cell* 76, 959-962 (1994).
- Simonet, W. S. et al. Osteoprotegerin: a novel secreted protein involved in the regulation of bone density. *Cell* 89, 309-319 (1997).
- Suda, T., Takahashi, T., Golstein, P. & Nagata, S. Molecular cloning and expression of Fas ligand, a novel member of the TNF family. *Cell* 75, 1169-1178 (1993).
- Pennica, D. et al. Human tumour necrosis factor: precursor structure, expression and homology to lymphotoxin. *Nature* 312, 724-729 (1984).
- Pitti, R. M. et al. Induction of apoptosis by Apo-2 ligand, a new member of the tumor necrosis factor receptor family. *J. Biol. Chem.* 271, 12687-12690 (1996).
- Wiley, S. R. et al. Identification and characterization of a new member of the TNF family that induces apoptosis. *Immunity* 3, 673-682 (1995).
- Marsters, S. A. et al. Identification of a ligand for the death-domain-containing receptor Apo3. *Curr. Biol.* 8, 525-528 (1998).
- Chicheportiche, Y. et al. TWEAK, a new secreted ligand in the TNF family that weakly induces apoptosis. *J. Biol. Chem.* 272, 32401-32410 (1997).
- Wong, B. R. et al. TRANCE is a novel ligand of the TNFR family that activates c-Jun-N-terminal kinase in T cells. *J. Biol. Chem.* 272, 25190-25194 (1997).
- Anderson, D. M. et al. A homolog of the TNF receptor and its ligand enhance T-cell growth and dendritic-cell function. *Nature* 390, 175-179 (1997).
- Lacey, D. L. et al. Osteoprotegerin ligand is a cytokine that regulates osteoclast differentiation and activation. *Cell* 93, 165-176 (1998).
- Dhein, J., Walczak, H., Baumler, C., Debatin, K. M. & Krammer, P. H. Autocrine T-cell suicide mediated by Apo1/Fas/CD95. *Nature* 373, 438-441 (1995).
- Arase, H., Arase, N. & Saito, T. Fas-mediated cytotoxicity by freshly isolated natural killer cells. *J. Exp. Med.* 181, 1235-1238 (1995).
- Medvedev, A. E. et al. Regulation of Fas and Fas ligand expression in NK cells by cytokines and the involvement of Fas ligand in NK/LAK cell-mediated cytotoxicity. *Cytokine* 9, 394-404 (1997).
- Moretta, A. Mechanisms in cell-mediated cytotoxicity. *Cell* 90, 13-18 (1997).
- Tanaka, M., Imai, T., Adachi, M. & Nagata, S. Downregulation of Fas ligand by shedding. *Nature Med.* 4, 31-36 (1998).
- Gelmini, S. et al. Quantitative PCR-based homogeneous assay with fluorogenic probes to measure c-erbB-2 oncogene amplification. *Clin. Chem.* 43, 752-758 (1997).
- Emery, J. G. et al. Osteoprotegerin is a receptor for the cytotoxic ligand TRAIL. *J. Biol. Chem.* 273, 14363-14367 (1998).
- Wallach, D. Placing death under control. *Nature* 388, 123-125 (1997).
- Colotta, F. et al. Interleukin-1 type II receptor: a decoy target for IL-1 that is regulated by IL-4. *Science* 261, 472-475 (1993).

22. Ashkenazi, A. & Dixit, V. M. Death receptors: signaling and modulation. *Science* **281**, 1305–1308 (1998).
23. Ashkenazi, A. & Chomow, S. M. Immunoadhesins as research tools and therapeutic agents. *Curr. Opin. Immunol.* **9**, 195–200 (1997).
24. Marsters, S. *et al.* Activation of apoptosis by Apo-2 ligand is independent of FADD but blocked by CrmA. *Curr. Biol.* **6**, 750–752 (1996).

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Correspondence and requests for materials should be addressed to A.A. (e-mail: aa@gene.com). The GenBank accession number for the DcR3 cDNA sequence is AF104419.

Crystal structure of the ATP-binding subunit of an ABC transporter

Li-Wei Hung*, Iris Xiaoyan Wang†, Kishiko Nikaido†, Pei-Qi Liu†, Giovanna Ferro-Luzzi Ames† & Sung-Hou Kim*‡

* E. O. Lawrence Berkeley National Laboratory, † Department of Molecular and Cell Biology, and ‡ Department of Chemistry, University of California at Berkeley, Berkeley, California 94720, USA

ABC transporters (also known as traffic ATPases) form a large family of proteins responsible for the translocation of a variety of compounds across membranes of both prokaryotes and eukaryotes¹. The recently completed *Escherichia coli* genome sequence revealed that the largest family of paralogous *E. coli* proteins is composed of ABC transporters². Many eukaryotic proteins of medical significance belong to this family, such as the cystic fibrosis transmembrane conductance regulator (CFTR), the P-glycoprotein (or multidrug-resistance protein) and the heterodimeric transporter associated with antigen processing (Tap1–Tap2). Here we report the crystal structure at 1.5 Å resolution of HisP, the ATP-binding subunit of the histidine permease, which is an ABC transporter from *Salmonella typhimurium*. We correlate the details of this structure with the biochemical, genetic and biophysical properties of the wild-type and several mutant HisP proteins. The structure provides a basis for understanding properties of ABC transporters and of defective CFTR proteins.

ABC transporters contain four structural domains: two nucleotide-binding domains (NBDs), which are highly conserved throughout the family, and two transmembrane domains¹. In prokaryotes these domains are often separate subunits which are assembled into a membrane-bound complex; in eukaryotes the domains are generally fused into a single polypeptide chain. The periplasmic histidine permease of *S. typhimurium* and *E. coli*^{1,3–8} is a well-characterized ABC transporter that is a good model for this superfamily. It consists of a membrane-bound complex, HisQMP₂, which comprises integral membrane subunits, HisQ and HisM, and two copies of HisP, the ATP-binding subunit. HisP, which has properties intermediate between those of integral and peripheral membrane proteins⁹, is accessible from both sides of the membrane, presumably by its interaction with HisQ and HisM⁶. The two HisP subunits form a dimer, as shown by their cooperativity in ATP hydrolysis⁵, the requirement for both subunits to be present for activity⁴, and the formation of a HisP dimer upon chemical cross-linking. Soluble HisP also forms a dimer³. HisP has been purified and characterized in an active soluble form³ which can be reconstituted into a fully active membrane-bound complex⁴.

The overall shape of the crystal structure of the HisP monomer is that of an 'L' with two thick arms (arm I and arm II); the ATP-binding pocket is near the end of arm I (Fig. 1). A six-stranded β -sheet ($\beta 3$ and $\beta 8$ – $\beta 12$) spans both arms of the L, with a domain of α - plus β -type structure ($\beta 1$, $\beta 2$, $\beta 4$ – $\beta 7$, $\alpha 1$ and $\alpha 2$) on one side (within arm I) and a domain of mostly α -helices ($\alpha 3$ – $\alpha 9$) on the

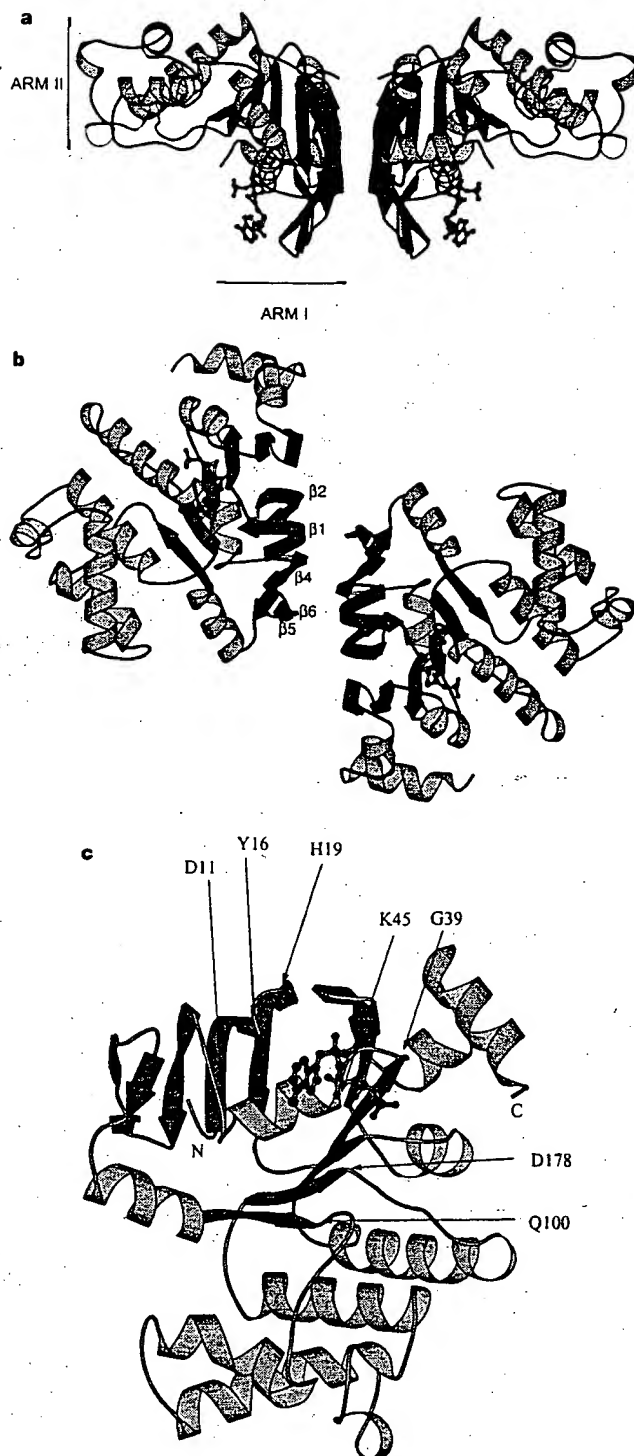


Figure 1. Crystal structure of HisP. **a**, View of the dimer along an axis perpendicular to its two-fold axis. The top and bottom of the dimer are suggested to face towards the periplasmic and cytoplasmic sides, respectively (see text). The thickness of arm II is about 25 Å, comparable to that of membrane. α -Helices are shown in orange and β -sheets in green. **b**, View along the two-fold axis of the HisP dimer, showing the relative displacement of the monomers not apparent in **a**. The β -strands at the dimer interface are labelled. **c**, View of one monomer from the bottom of arm I, as shown in **a**, towards arm II, showing the ATP-binding pocket. **a**–**c**, The protein and the bound ATP are in 'ribbon' and 'ball-and-stick' representations, respectively. Key residues discussed in the text are indicated in **c**. These figures were prepared with MOLSCRIPT²⁰. N, amino terminus; C, C terminus.

NOVEL APPROACH TO QUANTITATIVE POLYMERASE CHAIN REACTION USING REAL-TIME DETECTION: APPLICATION TO THE DETECTION OF GENE AMPLIFICATION IN BREAST CANCER

Ivan BIÈCHE^{1,2}, Martine OLIVI¹, Marie-Hélène CHAMPÈME², Dominique VIDAUD¹, Rosette LIDEREAU² and Michel VIDAUD^{1*}

¹Laboratoire de Génétique Moléculaire, Faculté des Sciences Pharmaceutiques et Biologiques de Paris, Paris, France

²Laboratoire d'Oncogénétique, Centre René Huguenin, St-Cloud, France

Gene amplification is a common event in the progression of human cancers, and amplified oncogenes have been shown to have diagnostic, prognostic and therapeutic relevance. A kinetic quantitative polymerase-chain-reaction (PCR) method, based on fluorescent TaqMan methodology and a new instrument (ABI Prism 7700 Sequence Detection System) capable of measuring fluorescence in real-time, was used to quantify gene amplification in tumor DNA. Reactions are characterized by the point during cycling when PCR amplification is still in the exponential phase, rather than the amount of PCR product accumulated after a fixed number of cycles. None of the reaction components is limited during the exponential phase, meaning that values are highly reproducible in reactions starting with the same copy number. This greatly improves the precision of DNA quantification. Moreover, real-time PCR does not require post-PCR sample handling, thereby preventing potential PCR-product carry-over contamination; it possesses a wide dynamic range of quantification and results in much faster and higher sample throughput. The real-time PCR method, was used to develop and validate a simple and rapid assay for the detection and quantification of the 3 most frequently amplified genes (*myc*, *ccnd1* and *erbB2*) in breast tumors. Extra copies of *myc*, *ccnd1* and *erbB2* were observed in 10, 23 and 15%, respectively, of 108 breast-tumor DNA; the largest observed numbers of gene copies were 4.6, 18.6 and 15.1, respectively. These results correlated well with those of Southern blotting. The use of this new semi-automated technique will make molecular analysis of human cancers simpler and more reliable, and should find broad applications in clinical and research settings. *Int. J. Cancer* 78:661–666, 1998.

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Gene amplification plays an important role in the pathogenesis of various solid tumors, including breast cancer, probably because over-expression of the amplified target genes confers a selective advantage. The first technique used to detect genomic amplification was cytogenetic analysis. Amplification of several chromosome regions, visualized either as extrachromosomal double minutes (dmns) or as integrated homogeneously staining regions (HSRs), are among the main visible cytogenetic abnormalities in breast tumors. Other techniques such as comparative genomic hybridization (CGH) (Kallioniemi *et al.*, 1994) have also been used in broad searches for regions of increased DNA copy numbers in tumor cells, and have revealed some 20 amplified chromosome regions in breast tumors. Positional cloning efforts are underway to identify the critical gene(s) in each amplified region. To date, genes known to be amplified frequently in breast cancers include *myc* (8q24), *ccnd1* (11q13), and *erbB2* (17q12-q21) (for review, see Bièche and Lidereau, 1995).

Amplification of the *myc*, *ccnd1*, and *erbB2* proto-oncogenes should have clinical relevance in breast cancer, since independent studies have shown that these alterations can be used to identify sub-populations with a worse prognosis (Berns *et al.*, 1992; Schuurin *et al.*, 1992; Slamon *et al.*, 1987). Muss *et al.* (1994) suggested that these gene alterations may also be useful for the prediction and assessment of the efficacy of adjuvant chemotherapy and hormone therapy.

However, published results diverge both in terms of the frequency of these alterations and their clinical value. For instance, over 500 studies in 10 years have failed to resolve the controversy

surrounding the link suggested by Slamon *et al.* (1987) between *erbB2* amplification and disease progression. These discrepancies are partly due to the clinical, histological and ethnic heterogeneity of breast cancer, but technical considerations are also probably involved.

Specific genes (DNA) were initially quantified in tumor cells by means of blotting procedures such as Southern and slot blotting. These batch techniques require large amounts of DNA (5–10 µg/reaction) to yield reliable quantitative results. Furthermore, meticulous care is required at all stages of the procedures to generate blots of sufficient quality for reliable dosage analysis. Recently, PCR has proven to be a powerful tool for quantitative DNA analysis, especially with minimal starting quantities of tumor samples (small, early-stage tumors and formalin-fixed, paraffin-embedded tissues).

Quantitative PCR can be performed by evaluating the amount of product either after a given number of cycles (end-point quantitative PCR) or after a varying number of cycles during the exponential phase (kinetic quantitative PCR). In the first case, an internal standard distinct from the target molecule is required to ascertain PCR efficiency. The method is relatively easy but implies generating, quantifying and storing an internal standard for each gene studied. Nevertheless, it is the most frequently applied method to date.

One of the major advantages of the kinetic method is its rapidity in quantifying a new gene, since no internal standard is required (an external standard curve is sufficient). Moreover, the kinetic method has a wide dynamic range (at least 5 orders of magnitude), giving an accurate value for samples differing in their copy number. Unfortunately, the method is cumbersome and has therefore been rarely used. It involves aliquot sampling of each assay mix at regular intervals and quantifying, for each aliquot, the amplification product. Interest in the kinetic method has been stimulated by a novel approach using fluorescent TaqMan methodology and a new instrument (ABI Prism 7700 Sequence Detection System) capable of measuring fluorescence in real time (Gibson *et al.*, 1996; Heid *et al.*, 1996). The TaqMan reaction is based on the 5' nuclease assay first described by Holland *et al.* (1991). The latter uses the 5' nuclease activity of Taq polymerase to cleave a specific fluorogenic oligonucleotide probe during the extension phase of PCR. The approach uses dual-labeled fluorogenic hybridization probes (Lee *et al.*, 1993). One fluorescent dye, co-valently linked to the 5' end of the oligonucleotide, serves as a reporter [FAM (*i.e.*, 6-carboxy-fluorescein)] and its emission spectrum is quenched by a second fluorescent dye, TAMRA (*i.e.*, 6-carboxy-tetramethyl-rhodamine) attached to the 3' end. During the extension phase of the PCR

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*Correspondence to: Laboratoire de Génétique Moléculaire, Faculté des Sciences Pharmaceutiques et Biologiques de Paris, 4 Avenue de l'Observatoire, F-75006 Paris, France. Fax: (33)1-4407-1754. E-mail: mvidaud@teaser.fr

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cycle, the fluorescent hybridization probe is hydrolyzed by the 5'-3' nucleolytic activity of DNA polymerase. Nuclease degradation of the probe releases the quenching of FAM fluorescence emission, resulting in an increase in peak fluorescence emission. The fluorescence signal is normalized by dividing the emission intensity of the reporter dye (FAM) by the emission intensity of a reference dye (i.e., ROX, 6-carboxy-X-rhodamine) included in TaqMan buffer, to obtain a ratio defined as the Rn (normalized reporter) for a given reaction tube. The use of a sequence detector enables the fluorescence spectra of all 96 wells of the thermal cycler to be measured continuously during PCR amplification.

The real-time PCR method offers several advantages over other current quantitative PCR methods (Celi *et al.*, 1994): (i) the probe-based homogeneous assay provides a real-time method for detecting only specific amplification products, since specific hybridization of both the primers and the probe is necessary to generate a signal; (ii) the C_t (threshold cycle) value used for quantification is measured when PCR amplification is still in the log phase of PCR product accumulation. This is the main reason why C_t is a more reliable measure of the starting copy number than are end-point measurements, in which a slight difference in a limiting component can have a drastic effect on the amount of product; (iii) use of C_t values gives a wider dynamic range (at least 5 orders of magnitude), reducing the need for serial dilution; (iv) The real-time PCR method is run in a closed-tube system and requires no post-PCR sample handling, thus avoiding potential contamination; (v) the system is highly automated, since the instrument continuously measures fluorescence in all 96 wells of the thermal cycler during PCR amplification and the corresponding software processes, and analyzes the fluorescence data; (vi) the assay is rapid, as results are available just one minute after thermal cycling is complete; (vii) the sample throughput of the method is high, since 96 reactions can be analyzed in 2 hr.

Here, we applied this semi-automated procedure to determine the copy numbers of the 3 most frequently amplified genes in breast tumors (*myc*, *ccnd1* and *erbB2*), as well as 2 genes (*alb* and *app*) located in a chromosome region in which no genetic changes have been observed in breast tumors. The results for 108 breast tumors were compared with previous Southern-blot data for the same samples.

MATERIAL AND METHODS

Tumor and blood samples

Samples were obtained from 108 primary breast tumors removed surgically from patients at the Centre René Huguenin; none of the patients had undergone radiotherapy or chemotherapy. Immediately after surgery, the tumor samples were placed in liquid nitrogen until extraction of high-molecular-weight DNA. Patients were included in this study if the tumor sample used for DNA preparation contained more than 60% of tumor cells (histological analysis). A blood sample was also taken from 18 of the same patients.

DNA was extracted from tumor tissue and blood leukocytes according to standard methods.

Real-time PCR

Theoretical basis. Reactions are characterized by the point during cycling when amplification of the PCR product is first detected, rather than by the amount of PCR product accumulated after a fixed number of cycles. The higher the starting copy number of the genomic DNA target, the earlier a significant increase in fluorescence is observed. The parameter C_t (threshold cycle) is defined as the fractional cycle number at which the fluorescence generated by cleavage of the probe passes a fixed threshold above baseline. The target gene copy number in unknown samples is quantified by measuring C_t and by using a standard curve to determine the starting copy number. The precise amount of genomic DNA (based on optical density) and its quality (i.e., lack

of extensive degradation) are both difficult to assess. We therefore also quantified a control gene (*alb*) mapping to chromosome region 4q11-q13, in which no genetic alterations have been found in breast-tumor DNA by means of CGH (Kallioniemi *et al.*, 1994).

Thus, the ratio of the copy number of the target gene to the copy number of the *alb* gene normalizes the amount and quality of genomic DNA. The ratio defining the level of amplification is termed "N", and is determined as follows:

$$N = \frac{\text{copy number of target gene (app, myc, ccnd1, erbB2)}}{\text{copy number of reference gene (alb)}}$$

Primers, probes, reference human genomic DNA and PCR consumables. Primers and probes were chosen with the assistance of the computer programs Oligo 4.0 (National Biosciences, Plymouth, MN), EuGene (Daniben Systems, Cincinnati, OH) and Primer Express (Perkin-Elmer Applied Biosystems, Foster City, CA).

Primers were purchased from DNAgency (Malvern, PA) and probes from Perkin-Elmer Applied Biosystems.

Nucleotide sequences for the oligonucleotide hybridization probes and primers are available on request.

The TaqMan PCR Core reagent kit, MicroAmp optical tubes, and MicroAmp caps were from Perkin-Elmer Applied Biosystems.

Standard-curve construction. The kinetic method requires a standard curve. The latter was constructed with serial dilutions of specific PCR products, according to Piatak *et al.* (1993). In practice, each specific PCR product was obtained by amplifying 20 ng of a standard human genomic DNA (Boehringer, Mannheim, Germany) with the same primer pairs as those used later for real-time quantitative PCR. The 5 PCR products were purified using MicroSpin S-400 HR columns (Pharmacia, Uppsala, Sweden) electrophoresed through an acrylamide gel and stained with ethidium bromide to check their quality. The PCR products were then quantified spectrophotometrically and pooled, and serially diluted 10-fold in mouse genomic DNA (Clontech, Palo Alto, CA) at a constant concentration of 2 ng/ μ l. The standard curve used for real-time quantitative PCR was based on serial dilutions of the pool of PCR products ranging from 10^{-7} (10^5 copies of each gene) to 10^{-10} (10^2 copies). This series of diluted PCR products was aliquoted and stored at -80°C until use.

The standard curve was validated by analyzing 2 known quantities of calibrator human genomic DNA (20 ng and 50 ng).

PCR amplification. Amplification mixes (50 μ l) contained the sample DNA (around 20 ng, around 6600 copies of disomic genes), $10\times$ TaqMan buffer (5 μ l), 200 μ M dATP, dCTP, dGTP, and 400 μ M dUTP, 5 mM MgCl_2 , 1.25 units of AmpliTaq Gold, 0.5 units of AmpErase uracil N-glycosylase (UNG), 200 nM each primer and 100 nM probe. The thermal cycling conditions comprised 2 min at 50°C and 10 min at 95°C . Thermal cycling consisted of 40 cycles at 95°C for 15 s and 65°C for 1 min. Each assay included: a standard curve (from 10^5 to 10^2 copies) in duplicate, a no-template control, 20 ng and 50 ng of calibrator human genomic DNA (Boehringer) in triplicate, and about 20 ng of unknown genomic DNA in triplicate (26 samples can thus be analyzed on a 96-well microplate). All samples with a coefficient of variation (CV) higher than 10% were retested.

All reactions were performed in the ABI Prism 7700 Sequence Detection System (Perkin-Elmer Applied Biosystems), which detects the signal from the fluorogenic probe during PCR.

Equipment for real-time detection. The 7700 system has a built-in thermal cycler and a laser directed via fiber optical cables to each of the 96 sample wells. A charge-coupled-device (CDD) camera collects the emission from each sample and the data are analyzed automatically. The software accompanying the 7700 system calculates C_t and determines the starting copy number in the samples.

Determination of gene amplification. Gene amplification was calculated as described above. Only samples with an N value higher than 2 were considered to be amplified.

RESULTS

To validate the method, real-time PCR was performed on genomic DNA extracted from 108 primary breast tumors, and 18 normal leukocyte DNA samples from some of the same patients. The target genes were the *myc*, *ccnd1* and *erbB2* proto-oncogenes, and the β -amyloid precursor protein gene (*app*), which maps to a chromosome region (21q21.2) in which no genetic alterations have been found in breast tumors (Kallioniemi *et al.*, 1994). The reference disomic gene was the albumin gene (*alb*, chromosome 4q11-q13).

Validation of the standard curve and dynamic range of real-time PCR

The standard curve was constructed from PCR products serially diluted in genomic mouse DNA at a constant concentration of 2 ng/ μ l. It should be noted that the 5 primer pairs chosen to analyze the 5 target genes do not amplify genomic mouse DNA (data not shown). Figure 1 shows the real-time PCR standard curve for the *alb* gene. The dynamic range was wide (at least 4 orders of magnitude), with samples containing as few as 10^2 copies or as many as 10^5 copies.

Copy-number ratio of the 2 reference genes (*app* and *alb*)

The *app* to *alb* copy-number ratio was determined in 18 normal leukocyte DNA samples and all 108 primary breast-tumor DNA

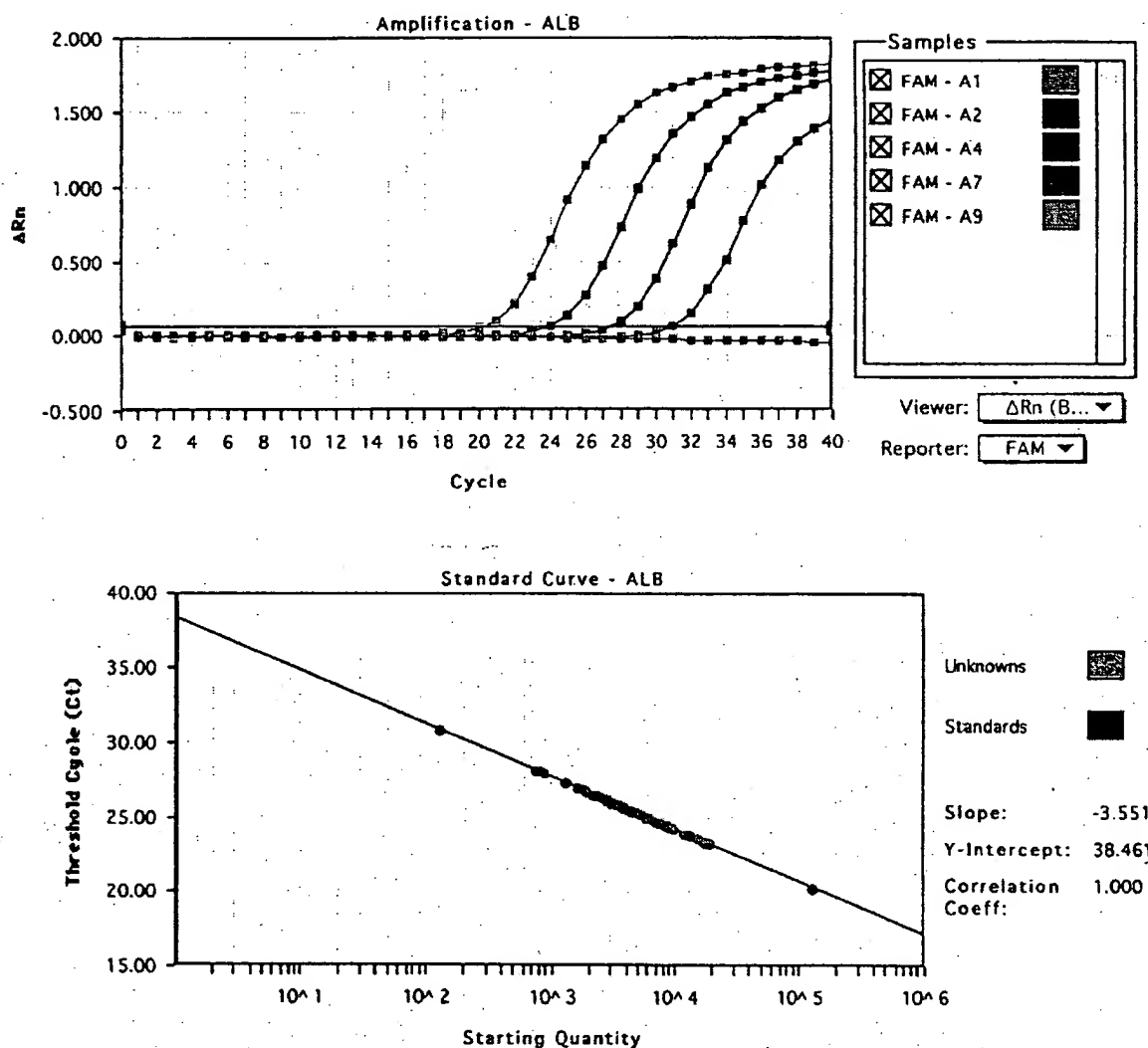


FIGURE 1 – Albumin (*alb*) gene dosage by real-time PCR. Top: Amplification plots for reactions with starting *alb* gene copy number ranging from 10^5 (A9), 10^4 (A7), 10^3 (A4) to 10^2 (A2) and a no-template control (A1). Cycle number is plotted vs. change in normalized reporter signal (ΔRn). For each reaction tube, the fluorescence signal of the reporter dye (FAM) is divided by the fluorescence signal of the passive reference dye (ROX), to obtain a ratio defined as the normalized reporter signal (Rn). ΔRn represents the normalized reporter signal (Rn) minus the baseline signal established in the first 15 PCR cycles. ΔRn increases during PCR as *alb* PCR product copy number increases until the reaction reaches a plateau. C_t (threshold cycle) represents the fractional cycle number at which a significant increase in Rn above a baseline signal (horizontal black line) can first be detected. Two replicate plots were performed for each standard sample, but the data for only one are shown here. Bottom: Standard curve plotting log starting copy number vs. C_t (threshold cycle). The black dots represent the data for standard samples plotted in duplicate and the red dots the data for unknown genomic DNA samples plotted in triplicate. The standard curve shows 4 orders of linear dynamic range.

samples. We selected these 2 genes because they are located in 2 chromosome regions (*app*, 21q21.2; *alb*, 4q11-q13) in which no obvious genetic changes (including gains or losses) have been observed in breast cancers (Kallioniemi *et al.*, 1994). The ratio for the 18 normal leukocyte DNA samples fell between 0.7 and 1.3 (mean 1.02 ± 0.21), and was similar for the 108 primary breast-tumor DNA samples (0.6 to 1.6, mean 1.06 ± 0.25), confirming that *alb* and *app* are appropriate reference disomic genes for breast-tumor DNA. The low range of the ratios also confirmed that the nucleotide sequences chosen for the primers and probes were not polymorphic, as mismatches of their primers or probes with the subject's DNA would have resulted in differential amplification.

myc, *ccnd1* and *erbB2* gene dose in normal leukocyte DNA

To determine the cut-off point for gene amplification in breast-cancer tissue, 18 normal leukocyte DNA samples were tested for the gene dose (N), calculated as described in "Material and Methods". The N value of these samples ranged from 0.5 to 1.3 (mean 0.84 ± 0.22) for *myc*; 0.7 to 1.6 (mean 1.06 ± 0.23) for *ccnd1* and 0.6 to 1.3 (mean 0.91 ± 0.19) for *erbB2*. Since N values for *myc*, *ccnd1* and *erbB2* in normal leukocyte DNA consistently fell between 0.5 and 1.6, values of 2 or more were considered to represent gene amplification in tumor DNA.

myc, *ccnd1* and *erbB2* gene dose in breast-tumor DNA

myc, *ccnd1* and *erbB2* gene copy numbers in the 108 primary breast tumors are reported in Table I. Extra copies of *ccnd1* were more frequent (23%, 25/108) than extra copies of *erbB2* (15%, 16/108) and *myc* (10%, 11/108), and ranged from 2 to 18.6 for *ccnd1*, 2 to 15.1 for *erbB2*, and only 2 to 4.6 for the *myc* gene. Figure 2 and Table II represent tumors in which the *ccnd1* gene was amplified 16-fold (T145), 6-fold (T133) and non-amplified (T118). The 3 genes were never found to be co-amplified in the same tumor. *erbB2* and *ccnd1* were co-amplified in only 3 cases, *myc* and *ccnd1* in 2 cases and *myc* and *erbB2* in 1 case. This favors the hypothesis that gene amplifications are independent events in breast cancer. Interestingly, 5 tumors showed a decrease of at least 50% in the *erbB2* copy number ($N < 0.5$), suggesting that they bore deletions of the 17q21 region (the site of *erbB2*). No such decrease in copy number was observed with the other 2 proto-oncogenes.

Comparison of gene dose determined by real-time quantitative PCR and Southern-blot analysis

Southern-blot analysis of *myc*, *ccnd1* and *erbB2* amplifications had previously been done on the same 108 primary breast tumors. A perfect correlation between the results of real-time PCR and Southern blot was obtained for tumors with high copy numbers ($N \geq 5$). However, there were cases (1 *myc*, 6 *ccnd1* and 4 *erbB2*) in which real-time PCR showed gene amplification whereas Southern-blot did not, but these were mainly cases with low extra copy numbers (N from 2 to 2.9).

DISCUSSION

The clinical applications of gene amplification assays are currently limited, but would certainly increase if a simple, standardized and rapid method were perfected. Gene amplification status has been studied mainly by means of Southern blotting, but this method is not sensitive enough to detect low-level gene amplification nor accurate enough to quantify the full range of amplification values. Southern blotting is also time-consuming, uses radioactive

reagents and requires relatively large amounts of high-quality genomic DNA, which means it cannot be used routinely in many laboratories. An amplification step is therefore required to determine the copy number of a given target gene from minimal quantities of tumor DNA (small early-stage tumors, cytopuncture specimens or formalin-fixed, paraffin-embedded tissues).

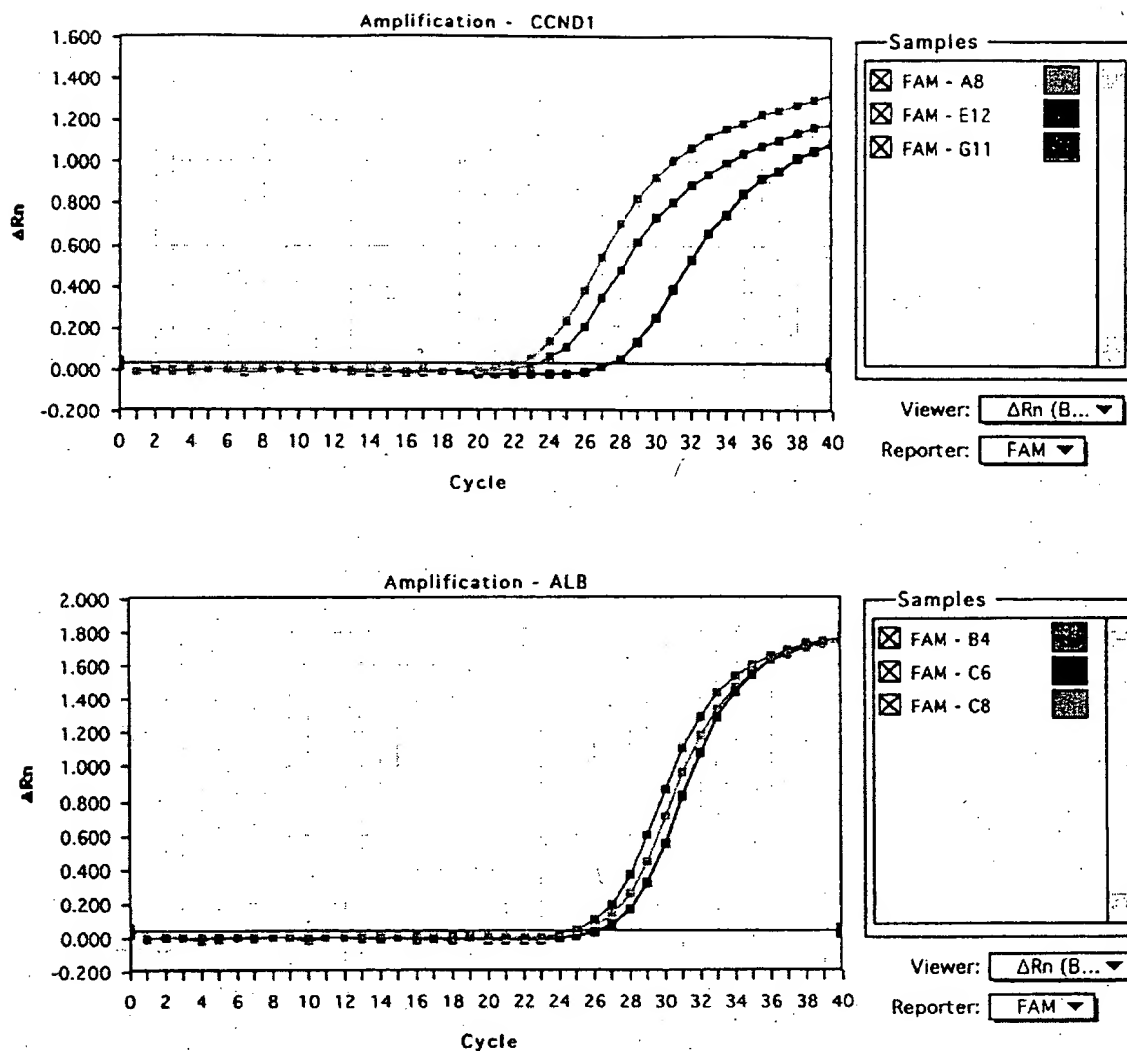
In this study, we validated a PCR method developed for the quantification of gene over-representation in tumors. The method, based on real-time analysis of PCR amplification, has several advantages over other PCR-based quantitative assays such as competitive quantitative PCR (Celi *et al.*, 1994). First, the real-time PCR method is performed in a closed-tube system, avoiding the risk of contamination by amplified products. Re-amplification of carryover PCR products in subsequent experiments can also be prevented by using the enzyme uracil N-glycosylase (UNG) (Longo *et al.*, 1990). The second advantage is the simplicity and rapidity of sample analysis, since no post-PCR manipulations are required. Our results show that the automated method is reliable. We found it possible to determine, in triplicate, the number of copies of a target gene in more than 100 tumors per day. Third, the system has a linear dynamic range of at least 4 orders of magnitude, meaning that samples do not have to contain equal starting amounts of DNA. This technique should therefore be suitable for analyzing formalin-fixed, paraffin-embedded tissues. Fourth, and above all, real-time PCR makes DNA quantification much more precise and reproducible, since it is based on C_t values rather than end-point measurement of the amount of accumulated PCR product. Indeed, the ABI Prism 7700 Sequence Detection System enables C_t to be calculated when PCR amplification is still in the exponential phase and when none of the reaction components is rate-limiting. The within-run CV of the C_t value for calibrator human DNA (5 replicates) was always below 5%, and the between-assay precision in 5 different runs was always below 10% (data not shown). In addition, the use of a standard curve is not absolutely necessary, since the copy number can be determined simply by comparing the C_t ratio of the target gene with that of reference genes. The results obtained by the 2 methods (with and without a standard curve) are similar in our experiments (data not shown). Moreover, unlike competitive quantitative PCR, real-time PCR does not require an internal control (the design and storage of internal controls and the validation of their amplification efficiency is laborious).

The only potential disadvantage of real-time PCR, like all other PCR-based methods and solid-matrix blotting techniques (Southern blots and dot blots) is that it cannot avoid dilution artifacts inherent in the extraction of DNA from tumor cells contained in heterogeneous tissue specimens. Only FISH and immunohistochemistry can measure alterations on a cell-by-cell basis (Pauletti *et al.*, 1996; Slamon *et al.*, 1989). However, FISH requires expensive equipment and trained personnel and is also time-consuming. Moreover, FISH does not assess gene expression and therefore cannot detect cases in which the gene product is over-expressed in the absence of gene amplification, which will be possible in the future by real-time quantitative RT-PCR. Immunohistochemistry is subject to considerable variations in the hands of different teams, owing to alterations of target proteins during the procedure, the different primary antibodies and fixation methods used and the criteria used to define positive staining.

The results of this study are in agreement with those reported in the literature. (i) Chromosome regions 4q11-q13 and 21q21.2 (which bear *alb* and *app*, respectively) showed no genetic alterations in the breast-cancer samples studied here, in keeping with the results of CGH (Kallioniemi *et al.*, 1994). (ii) We found that amplifications of these 3 oncogenes were independent events, as reported by other teams (Berns *et al.*, 1992; Borg *et al.*, 1992). (iii) The frequency and degree of *myc* amplification in our breast tumor DNA series were lower than those of *ccnd1* and *erbB2* amplification, confirming the findings of Borg *et al.* (1992) and Courjal *et al.* (1997). (iv) The maxima of *ccnd1* and *erbB2* over-representation were 18-fold and 15-fold, also in keeping with earlier results (about

TABLE I - DISTRIBUTION OF AMPLIFICATION LEVEL (N) FOR *myc*, *ccnd1* AND *erbB2* GENES IN 108 HUMAN BREAST TUMORS

| Gene | Amplification level (N) | | | |
|--------------|-------------------------|------------|------------|----------|
| | <0.5 | 0.5-1.9 | 2-4.9 | ≥ 5 |
| <i>myc</i> | 0 | 97 (89.8%) | 11 (10.2%) | 0 |
| <i>ccnd1</i> | 0 | 83 (76.9%) | 17 (15.7%) | 8 (7.4%) |
| <i>erbB2</i> | 5 (4.6%) | 87 (80.6%) | 8 (7.4%) | 8 (7.4%) |



| Tumor | CCND1 | | ALB | |
|--------|----------------|-------------|----------------|-------------|
| | C _t | Copy number | C _t | Copy number |
| ■ T118 | 27.3 | 4605 | 26.5 | 4365 |
| ■ T133 | 23.2 | 61659 | 25.2 | 10092 |
| ■ T145 | 22.1 | 125892 | 25.6 | 7762 |

FIGURE 2 - *ccnd1* and *alb* gene dosage by real-time PCR in 3 breast tumor samples: T118 (E12, C6, black squares), T133 (G11, B4, red squares) and T145 (A8, C8, blue squares). Given the C_t of each sample, the initial copy number is inferred from the standard curve obtained during the same experiment. Triplicate plots were performed for each tumor sample, but the data for only one are shown here. The results are shown in Table II.

30-fold maximum) (Berns *et al.*, 1992; Borg *et al.*, 1992; Courjal *et al.*, 1997). (v) The *erbB2* copy numbers obtained with real-time PCR were in good agreement with data obtained with other quantitative PCR-based assays in terms of the frequency and degree of amplification (An *et al.*, 1995; Deng *et al.*, 1996; Valeron

et al., 1996). Our results also correlate well with those recently published by Gelmini *et al.* (1997), who used the TaqMan system to measure *erbB2* amplification in a small series of breast tumors (n = 25), but with an instrument (LS-50B luminescence spectrometer, Perkin-Elmer Applied Biosystems) which only allows end-

TABLE II - EXAMPLES OF *ccnd1* GENE DOSAGE RESULTS FROM 3 BREAST TUMORS¹

| Tumor | <i>ccnd1</i> | | | <i>alb</i> | | | <i>Nccnd1/alb</i> |
|-------|--------------|--------|------|-------------|-------|-----|-------------------|
| | Copy number | Mean | SD | Copy number | Mean | SD | |
| T118 | 4525 | | | 4223 | | | |
| | 4605 | 4603 | 77 | 4365 | 4325 | 89 | 1.06 |
| | 4678 | | | 4387 | | | |
| T133 | 59821 | | | 9787 | | | |
| | 61659 | 61100 | 1111 | 10092 | 10137 | 375 | 6.03 |
| | 61821 | | | 10533 | | | |
| T145 | 128563 | | | 7321 | | | |
| | 125892 | 125392 | 3448 | 7762 | 7672 | 316 | 16.34 |
| | 121722 | | | 7933 | | | |

¹For each sample, 3 replicate experiments were performed and the mean and the standard deviation (SD) was determined. The level of *ccnd1* gene amplification (*Nccnd1/alb*) is determined by dividing the average *ccnd1* copy number value by the average *alb* copy number value.

point measurement of fluorescence intensity. Here we report *myc* and *ccnd1* gene dosage in breast cancer by means of quantitative PCR. (vi) We found a high degree of concordance between real-time quantitative PCR and Southern blot analysis in terms of gene amplification, especially for samples with high copy numbers (≥ 5 -fold). The slightly higher frequency of gene amplification (especially *ccnd1* and *erbB2*) observed by means of real-time quantitative PCR as compared with Southern-blot analysis may be explained by the higher sensitivity of the former method. However, we cannot rule out the possibility that some tumors with a few extra

gene copies observed in real-time PCR had additional copies of an arm or a whole chromosome (trisomy, tetrasomy or polysomy) rather than true gene amplification. These 2 types of genetic alteration (polysomy and gene amplification) could be easily distinguished in the future by using an additional probe located on the same chromosome arm, but some distance from the target gene. It is noteworthy that high gene copy numbers have the greatest prognostic significance in breast carcinoma (Borg *et al.*, 1992; Slamon *et al.*, 1987).

Finally, this technique can be applied to the detection of gene deletion as well as gene amplification. Indeed, we found a decreased copy number of *erbB2* (but not of the other 2 proto-oncogenes) in several tumors; *erbB2* is located in a chromosome region (17q21) reported to contain both deletions and amplifications in breast cancer (Bièche and Lidereau, 1995).

In conclusion, gene amplification in various cancers can be used as a marker of pre-neoplasia, also for early diagnosis of cancer, staging, prognostication and choice of treatment. Southern blotting is not sufficiently sensitive, and FISH is lengthy and complex. Real-time quantitative PCR overcomes both these limitations, and is a sensitive and accurate method of analyzing large numbers of samples in a short time. It should find a place in routine clinical gene dosage.

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REFERENCES

- AN, H.X., NIEDERACHER, D., BECKMANN, M.W., GÖHRING, U.J., SCHARL, A., PICARD, F., VAN ROEYEN, C., SCHNÜRCH, H.G. and BENDER, H.G., *erbB2* gene amplification detected by fluorescent differential polymerase chain reaction in paraffin-embedded breast carcinoma tissues. *Int. J. Cancer (Pred. Oncol.)*, **64**, 291-297 (1995).
- BERNS, E.M.J.J., KLIJN, J.G.M., VAN PUTTEN, W.L.J., VAN STAVEREN, I.L., PORTINGEN, H. and FOEKENS, J.A., *c-myc* amplification is a better prognostic factor than HER2/*neu* amplification in primary breast cancer. *Cancer Res.*, **52**, 1107-1113 (1992).
- BIÈCHE, I. and LIDEREAU, R., Genetic alterations in breast cancer. *Genes Chrom. Cancer*, **14**, 227-251 (1995).
- BORG, A., BALDETORP, B., FERNO, M., OLSSON, H. and SIGURDSSON, H., *c-myc* amplification is an independent prognostic factor in post-menopausal breast cancer. *Int. J. Cancer*, **51**, 687-691 (1992).
- CÉLI, F.S., COHEN, M.M., ANTONARAKIS, S.E., WERTHEIMER, E., ROTH, J. and SHULDINER, A.R., Determination of gene dosage by a quantitative adaptation of the polymerase chain reaction (qd-PCR): rapid detection of deletions and duplications of gene sequences. *Genomics*, **21**, 304-310 (1994).
- COURJAL, F., CUNY, M., SIMONY-LAFONTAINE, J., LOUASSON, G., SPEISER, P., ZEILLINGER, R., RODRIGUEZ, C. and THEILLET, C., Mapping of DNA amplifications at 15 chromosomal localizations in 1875 breast tumors: definition of phenotypic groups. *Cancer Res.*, **57**, 4360-4367 (1997).
- DENG, G., YU, M., CHEN, L.C., MOORE, D., KURISU, W., KALLIONIEMI, A., WALDMAN, F.M., COLLINS, C. and SMITH, H.S., Amplifications of oncogene *erbB-2* and chromosome 20q in breast cancer determined by differentially competitive polymerase chain reaction. *Breast Cancer Res. Treat.*, **40**, 271-281 (1996).
- GELMINI, S., ORLANDO, C., SESTINI, R., VONA, G., PINZANI, P., RUOCO, L. and PAZZAGLI, M., Quantitative polymerase chain reaction-based homogeneous assay with fluorogenic probes to measure *c-erbB-2* oncogene amplification. *Clin. Chem.*, **43**, 752-758 (1997).
- GIBSON, U.E.M., HEID, C.A. and WILLIAMS, P.M., A novel method for real-time quantitative RT-PCR. *Genome Res.*, **6**, 995-1001 (1996).
- HEID, C.A., STEVENS, J., LIVAK, K.J. and WILLIAMS, P.M., Real-time quantitative PCR. *Genome Res.*, **6**, 986-994 (1996).
- HOLLAND, P.M., ABRAMSON, R.D., WATSON, R. and GELFAND, D.H., Detection of specific polymerase chain reaction product by utilizing the 5' to 3' exonuclease activity of *Thermus aquaticus* DNA polymerase. *Proc. nat. Acad. Sci. (Wash.)*, **88**, 7276-7280 (1991).
- KALLIONIEMI, A., KALLIONIEMI, O.P., PIPER, J., TANNER, M., STOKKES, T., CHEN, L., SMITH, H.S., PINKEL, D., GRAY, J.W. and WALDMAN, F.M., Detection and mapping of amplified DNA sequences in breast cancer by comparative genomic hybridization. *Proc. nat. Acad. Sci. (Wash.)*, **91**, 2156-2160 (1994).
- LEE, L.G., CONNELL, C.R. and BIOCH, W., Allelic discrimination by nick-translation PCR with fluorogenic probe. *Nucleic Acids Res.*, **21**, 3761-3766 (1993).
- LONGO, N., BERNINGER, N.S. and HARTLEY, J.L., Use of uracil DNA glycosylase to control carry-over contamination in polymerase chain reactions. *Gene*, **93**, 125-128 (1990).
- MUSS, H.B., THOR, A.D., BERRY, D.A., KUTE, T., LIU, E.T., KOERNER, F., CIRINCIONE, C.T., BUDMAN, D.R., WOOD, W.C., BARCOS, M. and HENDERSON, I.C., *c-erbB-2* expression and response to adjuvant therapy in women with node-positive early breast cancer. *New Engl. J. Med.*, **330**, 1260-1266 (1994).
- PAULETTI, G., GODOLPHIN, W., PRESS, M.F. and SALMON, D.J., Detection and quantification of HER-2/*neu* gene amplification in human breast cancer archival material using fluorescence *in situ* hybridization. *Oncogene*, **13**, 63-72 (1996).
- PIATAK, M., LUK, K.C., WILLIAMS, B. and LIFSON, J.D., Quantitative competitive polymerase chain reaction for accurate quantitation of HIV DNA and RNA species. *Biotechniques*, **14**, 70-80 (1993).
- SCHUURING, E., VERHOEVEN, E., VAN TINTEREN, H., PETERSE, J.L., NUNNIK, B., THUNNISSEN, F.B.J.M., DEVILEE, P., CORNELISSE, C.J., VAN DE VIJVER, M.J., MOOI, W.J. and MICHALIDES, R.J.A.M., Amplification of genes within the chromosome 11q13 region is indicative of poor prognosis in patients with operable breast cancer. *Cancer Res.*, **52**, 5229-5234 (1992).
- SLAMON, D.J., CLARK, G.M., WONG, S.G., LEVIN, W.S., ULLRICH, A. and MCGUIRE, W.L., Human breast cancer: correlation of relapse and survival with amplification of the HER-2/*neu* oncogene. *Science*, **235**, 177-182 (1987).
- SLAMON, D.J., GODOLPHIN, W., JONES, L.A., HOLT, J.A., WONG, S.G., KEITH, D.E., LEVIN, W.J., STUART, S.G., UDOWE, J., ULLRICH, A. and PRESS, M.F., Studies of the HER-2/*neu* proto-oncogene in human breast and ovarian cancer. *Science*, **244**, 707-712 (1989).
- VALERON, P.F., CHIRINO, R., FERNANDEZ, L., TORRES, S., NAVARRO, D., AGUIAR, J., CABRERA, J.J., DIAZ-CHICO, B.N. and DIAZ-CHICO, J.C., Validation of a differential PCR and an ELISA procedure in studying HER-2/*neu* status in breast cancer. *Int. J. Cancer*, **65**, 129-133 (1996).

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant : Ashkenazi et al.
App. No. : 09/903,925
Filed : July 11, 2001
For : SECRETED AND
TRANSMEMBRANE
POLYPEPTIDES AND NUCLEIC
ACIDS ENCODING THE SAME
Examiner : Hamud, Fozia M

Group Art Unit 1647

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DECLARATION OF AVI ASHKENAZI, Ph.D UNDER 37 C.F.R. § 1.132

I, Avi Ashkenazi, Ph.D. declare and say as follows: -

1. I am Director and Staff Scientist at the Molecular Oncology Department of Genentech, Inc., South San Francisco, CA 94080.
2. I joined Genentech in 1988 as a postdoctoral fellow. Since then, I have investigated a variety of cellular signal transduction mechanisms, including apoptosis, and have developed technologies to modulate such mechanisms as a means of therapeutic intervention in cancer and autoimmune disease. I am currently involved in the investigation of a series of secreted proteins over-expressed in tumors, with the aim to identify useful targets for the development of therapeutic antibodies for cancer treatment.
3. My scientific Curriculum Vitae, including my list of publications, is attached to and forms part of this Declaration (Exhibit A).
4. Gene amplification is a process in which chromosomes undergo changes to contain multiple copies of certain genes that normally exist as a single copy, and is an important factor in the pathophysiology of cancer. Amplification of certain genes (e.g., Myc or Her2/Neu)

gives cancer cells a growth or survival advantage relative to normal cells, and might also provide a mechanism of tumor cell resistance to chemotherapy or radiotherapy.

5. If gene amplification results in over-expression of the mRNA and the corresponding gene product, then it identifies that gene product as a promising target for cancer therapy, for example by the therapeutic antibody approach. Even in the absence of over-expression of the gene product, amplification of a cancer marker gene - as detected, for example, by the reverse transcriptase TaqMan[®] PCR or the fluorescence *in situ* hybridization (FISH) assays - is useful in the diagnosis or classification of cancer, or in predicting or monitoring the efficacy of cancer therapy. An increase in gene copy number can result not only from intrachromosomal changes but also from chromosomal aneuploidy. It is important to understand that detection of gene amplification can be used for cancer diagnosis even if the determination includes measurement of chromosomal aneuploidy. Indeed, as long as a significant difference relative to normal tissue is detected, it is irrelevant if the signal originates from an increase in the number of gene copies per chromosome and/or an abnormal number of chromosomes.

6. I understand that according to the Patent Office, absent data demonstrating that the increased copy number of a gene in certain types of cancer leads to increased expression of its product, gene amplification data are insufficient to provide substantial utility or well established utility for the gene product (the encoded polypeptide), or an antibody specifically binding the encoded polypeptide. However, even when amplification of a cancer marker gene does not result in significant over-expression of the corresponding gene product, this very absence of gene product over-expression still provides significant information for cancer diagnosis and treatment. Thus, if over-expression of the gene product does not parallel gene amplification in certain tumor types but does so in others, then parallel monitoring of gene amplification and gene product over-expression enables more accurate tumor classification and hence better determination of suitable therapy. In addition, absence of over-expression is crucial information for the practicing clinician. If a gene is amplified but the corresponding gene product is not over-expressed, the clinician accordingly will decide not to treat a patient with agents that target that gene product.

7. I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information or belief are believed to be true, and further that these statements were made with the knowledge that willful false statements and the like so

made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful statements may jeopardize the validity of the application or any patent issued thereon.

By: Avi Ashkenazi
Avi Ashkenazi, Ph.D.

Date: 9/15/03

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9/12/03 3:06 PM (39780.7000)

CURRICULUM VITAE

Avi Ashkenazi

July 2003

Personal:

Date of birth: 29 November, 1956
Address: 1456 Tarrytown Street, San Mateo, CA 94402
Phone: (650) 578-9199 (home); (650) 225-1853 (office)
Fax: (650) 225-6443 (office)
Email: aa@gene.com

Education:

1983: B.S. in Biochemistry, with honors, Hebrew University, Israel
1986: Ph.D. in Biochemistry, Hebrew University, Israel

Employment:

1983-1986: Teaching assistant, undergraduate level course in Biochemistry
1985-1986: Teaching assistant, graduate level course on Signal Transduction
1986 - 1988: Postdoctoral fellow, Hormone Research Dept., UCSF, and
Developmental Biology Dept., Genentech, Inc., with J. Ramachandran
1988 - 1989: Postdoctoral fellow, Molecular Biology Dept., Genentech, Inc.,
with D. Capon
1989 - 1993: Scientist, Molecular Biology Dept., Genentech, Inc.
1994 -1996: Senior Scientist, Molecular Oncology Dept., Genentech, Inc.
1996-1997: Senior Scientist and Interim director, Molecular Oncology Dept.,
Genentech, Inc.
1997-1990: Senior Scientist and preclinical project team leader, Genentech, Inc.
1999 -2002: Staff Scientist in Molecular Oncology, Genentech, Inc.
2002-present: Staff Scientist and Director in Molecular Oncology, Genentech, Inc.

Awards:

1988: First prize, The Boehringer Ingelheim Award

Editorial:

Editorial Board Member: Current Biology

Associate Editor, Clinical Cancer Research.

Associate Editor, Cancer Biology and Therapy.

Refereed papers:

1. Gertler, A., Ashkenazi, A., and Madar, Z. Binding sites for human growth hormone and ovine and bovine prolactins in the mammary gland and liver of the lactating cow. *Mol. Cell. Endocrinol.* 34, 51-57 (1984).
2. Gertler, A., Shamay, A., Cohen, N., Ashkenazi, A., Friesen, H., Levanon, A., Gorecki, M., Aviv, H., Hadari, D., and Vogel, T. Inhibition of lactogenic activities of ovine prolactin and human growth hormone (hGH) by a novel form of a modified recombinant hGH. *Endocrinology* 118, 720-726 (1986).
3. Ashkenazi, A., Madar, Z., and Gertler, A. Partial purification and characterization of bovine mammary gland prolactin receptor. *Mol. Cell. Endocrinol.* 50, 79-87 (1987).
4. Ashkenazi, A., Pines, M., and Gertler, A. Down-regulation of lactogenic hormone receptors in Nb2 lymphoma cells by cholera toxin. *Biochemistry Internatl.* 14, 1065-1072 (1987).
5. Ashkenazi, A., Cohen, R., and Gertler, A. Characterization of lactogen receptors in lactogenic hormone-dependent and independent Nb2 lymphoma cell lines. *FEBS Lett.* 210, 51-55 (1987).
6. Ashkenazi, A., Vogel, T., Barash, I., Hadari, D., Levanon, A., Gorecki, M., and Gertler, A. Comparative study on in vitro and in vivo modulation of lactogenic and somatotropic receptors by native human growth hormone and its modified recombinant analog. *Endocrinology* 121, 414-419 (1987).
7. Peralta, E., Winslow, J., Peterson, G., Smith, D., Ashkenazi, A., Ramachandran, J., Schimerlik, M., and Capon, D. Primary structure and biochemical properties of an M2 muscarinic receptor. *Science* 236, 600-605 (1987).
8. Peralta, E., Ashkenazi, A., Winslow, J., Smith, D., Ramachandran, J., and Capon, D. J. Distinct primary structures, ligand-binding properties and tissue-specific expression of four human muscarinic acetylcholine receptors. *EMBO J.* 6, 3923-3929 (1987).
9. Ashkenazi, A., Winslow, J., Peralta, E., Peterson, G., Schimerlik, M., Capon, D., and Ramachandran, J. An M2 muscarinic receptor subtype coupled to both adenylyl cyclase and phosphoinositide turnover. *Science* 238, 672-675 (1987).

10. Pines, M., Ashkenazi, A., Cohen-Chapnik, N., Binder, L., and Gertler, A. Inhibition of the proliferation of Nb2 lymphoma cells by femtomolar concentrations of cholera toxin and partial reversal of the effect by 12-o-tetradecanoyl-phorbol-13-acetate. *J. Cell. Biochem.* 37, 119-129 (1988).
11. Peralta, E., Ashkenazi, A., Winslow, J., Ramachandran, J., and Capon, D. Differential regulation of PI hydrolysis and adenylyl cyclase by muscarinic receptor subtypes. *Nature* 334, 434-437 (1988).
12. Ashkenazi, A., Peralta, E., Winslow, J., Ramachandran, J., and Capon, D. Functionally distinct G proteins couple different receptors to PI hydrolysis in the same cell. *Cell* 56, 487-493 (1989).
13. Ashkenazi, A., Ramachandran, J., and Capon, D. Acetylcholine analogue stimulates DNA synthesis in brain-derived cells via specific muscarinic acetylcholine receptor subtypes. *Nature* 340, 146-150 (1989).
14. Lammare, D., Ashkenazi, A., Fleury, S., Smith, D., Sekaly, R., and Capon, D. The MHC-binding and gp120-binding domains of CD4 are distinct and separable. *Science* 245, 743-745 (1989).
15. Ashkenazi, A., Presta, L., Marsters, S., Camerato, T., Rosenthal, K., Fendly, B., and Capon, D. Mapping the CD4 binding site for human immunodeficiency virus type 1 by alanine-scanning mutagenesis. *Proc. Natl. Acad. Sci. USA.* 87, 7150-7154 (1990).
16. Chamow, S., Peers, D., Byrn, R., Mulkerrin, M., Harris, R., Wang, W., Bjorkman, P., Capon, D., and Ashkenazi, A. Enzymatic cleavage of a CD4 immunoadhesin generates crystallizable, biologically active Fd-like fragments. *Biochemistry* 29, 9885-9891 (1990).
17. Ashkenazi, A., Smith, D., Marsters, S., Riddle, L., Gregory, T., Ho, D., and Capon, D. Resistance of primary isolates of human immunodeficiency virus type 1 to soluble CD4 is independent of CD4-gp120 binding affinity. *Proc. Natl. Acad. Sci. USA.* 88, 7056-7060 (1991).
18. Ashkenazi, A., Marsters, S., Capon, D., Chamow, S., Figari, I., Pennica, D., Goeddel, D., Palladino, M., and Smith, D. Protection against endotoxic shock by a tumor necrosis factor receptor immunoadhesin. *Proc. Natl. Acad. Sci. USA.* 88, 10535-10539 (1991).
19. Moore, J., McKeating, J., Huang, Y., Ashkenazi, A., and Ho, D. Virions of primary HIV-1 isolates resistant to sCD4 neutralization differ in sCD4 affinity and glycoprotein gp120 retention from sCD4-sensitive isolates. *J. Virol.* 66, 235-243 (1992).

20. Jin, H., Oksenberg, D., Ashkenazi, A., Peroutka, S., Duncan, A., Rozmahel, R., Yang, Y., Mengod, G., Palacios, J., and O'Dowd, B. Characterization of the human 5-hydroxytryptamine_{1B} receptor. *J. Biol. Chem.* **267**, 5735-5738 (1992).
21. Marsters, A., Frutkin, A., Simpson, N., Fendly, B. and Ashkenazi, A. Identification of cysteine-rich domains of the type 1 tumor necrosis receptor involved in ligand binding. *J. Biol. Chem.* **267**, 5747-5750 (1992).
22. Chamow, S., Kogan, T., Peers, D., Hastings, R., Byrn, R., and Ashkenazi, A. Conjugation of sCD4 without loss of biological activity via a novel carbohydrate-directed cross-linking reagent. *J. Biol. Chem.* **267**, 15916-15922 (1992).
23. Oksenberg, D., Marsters, A., O'Dowd, B., Jin, H., Havlik, S., Peroutka, S., and Ashkenazi, A. A single amino-acid difference confers major pharmacologic variation between human and rodent 5-HT_{1B} receptors. *Nature* **360**, 161-163 (1992).
24. Haak-Frendscho, M., Marsters, S., Chamow, S., Peers, D., Simpson, N., and Ashkenazi, A. Inhibition of interferon γ by an interferon γ receptor immunoadhesin. *Immunology* **79**, 594-599 (1993).
25. Penica, D., Lam, V., Weber, R., Kohr, W., Basa, L., Spellman, M., Ashkenazi, A., Shire, S., and Goeddel, D. Biochemical characterization of the extracellular domain of the 75-kd tumor necrosis factor receptor. *Biochemistry* **32**, 3131-3138. (1993).
26. Barfod, L., Zheng, Y., Kuang, W., Hart, M., Evans, T., Cerione, R., and Ashkenazi, A. Cloning and expression of a human CDC42 GTPase Activating Protein reveals a functional SH3-binding domain. *J. Biol. Chem.* **268**, 26059-26062 (1993).
27. Chamow, S., Zhang, D., Tan, X., Mhtre, S., Marsters, S., Peers, D., Byrn, R., Ashkenazi, A., and Yunghans, R. A humanized bispecific immunoadhesin-antibody that retargets CD3⁺ effectors to kill HIV-1-infected cells. *J. Immunol.* **153**, 4268-4280 (1994).
28. Means, R., Krantz, S., Luna, J., Marsters, S., and Ashkenazi, A. Inhibition of murine erythroid colony formation in vitro by interferon γ and correction by interferon γ receptor immunoadhesin. *Blood* **83**, 911-915 (1994).
29. Haak-Frendscho, M., Marsters, S., Mordenti, J., Gillet, N., Chen, S., and Ashkenazi, A. Inhibition of TNF by a TNF receptor immunoadhesin: comparison with an anti-TNF mAb. *J. Immunol.* **152**, 1347-1353 (1994).

30. Chamow, S., Kogan, T., Venuti, M., Gadek, T., Peers, D., Mordenti, J., Shak, S., and Ashkenazi, A. Modification of CD4 immunoadhesin with monomethoxy-PEG aldehyde via reductive alkylation. *Bioconj. Chem.* 5, 133-140 (1994).
31. Jin, H., Yang, R., Marsters, S., Bunting, S., Wurm, F., Chamow, S., and Ashkenazi, A. Protection against rat endotoxic shock by p55 tumor necrosis factor (TNF) receptor immunoadhesin: comparison to anti-TNF monoclonal antibody. *J. Infect. Diseases* 170, 1323-1326 (1994).
32. Beck, J., Marsters, S., Harris, R., Ashkenazi, A., and Chamow, S. Generation of soluble interleukin-1 receptor from an immunoadhesin by specific cleavage. *Mol. Immunol.* 31, 1335-1344 (1994).
33. Pitti, B., Marsters, M., Haak-Frendscho, M., Osaka, G., Mordenti, J., Chamow, S., and Ashkenazi, A. Molecular and biological properties of an interleukin-1 receptor immunoadhesin. *Mol. Immunol.* 31, 1345-1351 (1994).
34. Oksenberg, D., Havlik, S., Peroutka, S., and Ashkenazi, A. The third intracellular loop of the 5-HT₂ receptor specifies effector coupling. *J. Neurochem.* 64, 1440-1447 (1995).
35. Bach, E., Szabo, S., Dighe, A., Ashkenazi, A., Aguet, M., Murphy, K., and Schreiber, R. Ligand-induced autoregulation of IFN- γ receptor β chain expression in T helper cell subsets. *Science* 270, 1215-1218 (1995).
36. Jin, H., Yang, R., Marsters, S., Ashkenazi, A., Bunting, S., Marra, M., Scott, R., and Baker, J. Protection against endotoxic shock by bactericidal/permeability-increasing protein in rats. *J. Clin. Invest.* 95, 1947-1952 (1995).
37. Marsters, S., Penica, D., Bach, E., Schreiber, R., and Ashkenazi, A. Interferon γ signals via a high-affinity multisubunit receptor complex that contains two types of polypeptide chain. *Proc. Natl. Acad. Sci. USA.* 92, 5401-5405 (1995).
38. Van Zee, K., Moldawer, L., Oldenburg, H., Thompson, W., Stackpole, S., Montegut, W., Rogy, M., Meschter, C., Gallati, H., Schiller, C., Richter, W., Loetcher, H., Ashkenazi, A., Chamow, S., Wurm, F., Calvano, S., Lowry, S., and Lesslauer, W. Protection against lethal *E. coli* bacteremia in baboons by pretreatment with a 55-kDa TNF receptor-Ig fusion protein, Ro45-2081. *J. Immunol.* 156, 2221-2230 (1996).
39. Pitti, R., Marsters, S., Ruppert, S., Donahue, C., Moore, A., and Ashkenazi, A. Induction of apoptosis by Apo-2 Ligand, a new member of the tumor necrosis factor cytokine family. *J. Biol. Chem.* 271, 12687-12690 (1996).

40. Marsters, S., Pitti, R., Donahue, C., Rupert, S., Bauer, K., and Ashkenazi, A. Activation of apoptosis by Apo-2 ligand is independent of FADD but blocked by CrmA. *Curr. Biol.* 6, 1669-1676 (1996).
41. Marsters, S., Skubatch, M., Gray, C., and Ashkenazi, A. Herpesvirus entry mediator, a novel member of the tumor necrosis factor receptor family, activates the NF- κ B and AP-1 transcription factors. *J. Biol. Chem.* 272, 14029-14032 (1997).
42. Sheridan, J., Marsters, S., Pitti, R., Gurney, A., Skubatch, M., Baldwin, D., Ramakrishnan, L., Gray, C., Baker, K., Wood, W.I., Goddard, A., Godowski, P., and Ashkenazi, A. Control of TRAIL-induced apoptosis by a family of signaling and decoy receptors. *Science* 277, 818-821 (1997).
43. Marsters, S., Sheridan, J., Pitti, R., Gurney, A., Skubatch, M., Baldwin, D., Huang, A., Yuan, J., Goddard, A., Godowski, P., and Ashkenazi, A. A novel receptor for Apo2L/TRAIL contains a truncated death domain. *Curr. Biol.* 7, 1003-1006 (1997).
44. Marsters, A., Sheridan, J., Pitti, R., Brush, J., Goddard, A., and Ashkenazi, A. Identification of a ligand for the death-domain-containing receptor Apo3. *Curr. Biol.* 8, 525-528 (1998).
45. Rieger, J., Naumann, U., Glaser, T., Ashkenazi, A., and Weller, M. Apo2 ligand: a novel weapon against malignant glioma? *FEBS Lett.* 427, 124-128 (1998).
46. Pender, S., Fell, J., Chamow, S., Ashkenazi, A., and MacDonald, T. A p55 TNF receptor immunoadhesin prevents T cell mediated intestinal injury by inhibiting matrix metalloproteinase production. *J. Immunol.* 160, 4098-4103 (1998).
47. Pitti, R., Marsters, S., Lawrence, D., Roy, Kischkel, F., M., Dowd, P., Huang, A., Donahue, C., Sherwood, S., Baldwin, D., Godowski, P., Wood, W., Gurney, A., Hillan, K., Cohen, R., Goddard, A., Botstein, D., and Ashkenazi, A. Genomic amplification of a decoy receptor for Fas ligand in lung and colon cancer. *Nature* 396, 699-703 (1998).
48. Mori, S., Marakami-Mori, K., Nakamura, S., Ashkenazi, A., and Bonavida, B. Sensitization of AIDS Kaposi's sarcoma cells to Apo-2 ligand-induced apoptosis by actinomycin D. *J. Immunol.* 162, 5616-5623 (1999).
49. Gurney, A. Marsters, S., Huang, A., Pitti, R., Mark, M., Baldwin, D., Gray, A., Dowd, P., Brush, J., Heldens, S., Schow, P., Goddard, A., Wood, W., Baker, K., Godowski, P., and Ashkenazi, A. Identification of a new member of the tumor necrosis factor family and its receptor, a human ortholog of mouse GITR. *Curr. Biol.* 9, 215-218 (1999).

50. Ashkenazi, A., Pai, R., Fong, s., Leung, S., Lawrence, D., Marsters, S., Blackie, C., Chang, L., McMurtrey, A., Hebert, A., DeForge, L., Khoumenis, I., Lewis, D., Harris, L., Bussiere, J., Koeppen, H., Shahrokh, Z., and Schwall, R. Safety and anti-tumor activity of recombinant soluble Apo2 ligand. *J. Clin. Invest.* **104**, 155-162 (1999).
51. Chuntharapai, A., Gibbs, V., Lu, J., Ow, A., Marsters, S., Ashkenazi, A., De Vos, A., Kim, K.J. Determination of residues involved in ligand binding and signal transmissiion in the human IFN- α receptor 2. *J. Immunol.* **163**, 766-773 (1999).
52. Johnsen, A.-C., Haux, J., Steinkjer, B., Nonstad, U., Egeberg, K., Sundan, A., Ashkenazi, A., and Espevik, T. Regulation of Apo2L/TRAIL expression in NK cells – involvement in NK cell-mediated cytotoxicity. *Cytokine* **11**, 664-672 (1999).
53. Roth, W., Isenmann, S., Naumann, U., Kugler, S., Bahr, M., Dichgans, J., Ashkenazi, A., and Weller, M. Eradication of intracranial human malignant glioma xenografts by Apo2L/TRAIL. *Biochem. Biophys. Res. Commun.* **265**, 479-483 (1999).
54. Hymowitz, S.G., Christinger, H.W., Fuh, G., Ultsch, M., O'Connell, M., Kelley, R.F., Ashkenazi, A. and de Vos, A.M. Triggering Cell Death: The Crystal Structure of Apo2L/TRAIL in a Complex with Death Receptor 5. *Molec. Cell* **4**, 563–571 (1999).
55. Hymowitz, S.G., O'Connel, M.P., Utsch, M.H., Hurst, A., Totpal, K., Ashkenazi, A., de Vos, A.M., Kelley, R.F. A unique zinc-binding site revealed by a high-resolution X-ray structure of homotrimeric Apo2L/TRAIL. *Biochemistry* **39**, 633-640 (2000).
56. Zhou, Q., Fukushima, P., DeGraff, W., Mitchell, J.B., Stetler-Stevenson, M., Ashkenazi, A., and Steeg, P.S. Radiation and the Apo2L/TRAIL apoptotic pathway preferentially inhibit the colonization of premalignant human breast cancer cells overexpressing cyclin D1. *Cancer Res.* **60**, 2611-2615 (2000).
57. Kischkel, F.C., Lawrence, D. A., Chuntharapai, A., Schow, P., Kim, J., and Ashkenazi, A. Apo2L/TRAIL-dependent recruitment of endogenous FADD and Caspase-8 to death receptors 4 and 5. *Immunity* **12**, 611-620 (2000).
58. Yan, M., Marsters, S.A., Grewal, I.S., Wang, H., *Ashkenazi, A., and *Dixit, V.M. Identification of a receptor for BlyS demonstrates a crucial role in humoral immunity. *Nature Immunol.* **1**, 37-41 (2000).

59. Marsters, S.A., Yan, M., Pitti, R.M., Haas, P.E., Dixit, V.M., and Ashkenazi, A. Interaction of the TNF homologues BLyS and APRIL with the TNF receptor homologues BCMA and TACI. *Curr. Biol.* 10, 785-788 (2000).
60. Kischkel, F.C., and Ashkenazi, A. Combining enhanced metabolic labeling with immunoblotting to detect interactions of endogenous cellular proteins. *Biotechniques* 29, 506-512 (2000).
61. Lawrence, D., Shahrokh, Z., Marsters, S., Achilles, K., Shih, D. Mounho, B., Hillan, K., Totpal, K. DeForge, L., Schow, P., Hooley, J., Sherwood, S., Pai, R., Leung, S., Khan, L., Gliniak, B., Bussiere, J., Smith, C., Strom, S., Kelley, S., Fox, J., Thomas, D., and Ashkenazi, A. Differential hepatocyte toxicity of recombinant Apo2L/TRAIL versions. *Nature Med.* 7, 383-385 (2001).
62. Chuntharapai, A., Dodge, K., Grimmer, K., Schroeder, K., Marsters, S.A., Koeppen, H., Ashkenazi, A., and Kim, K.J. Isotype-dependent inhibition of tumor growth in vivo by monoclonal antibodies to death receptor 4. *J. Immunol.* 166, 4891-4898 (2001).
63. Pollack, I.F., Erff, M., and Ashkenazi, A. Direct stimulation of apoptotic signaling by soluble Apo2L/tumor necrosis factor-related apoptosis-inducing ligand leads to selective killing of glioma cells. *Clin. Cancer Res.* 7, 1362-1369 (2001).
64. Wang, H., Marsters, S.A., Baker, T., Chan, B., Lee, W.P., Fu, L., Tumas, D., Yan, M., Dixit, V.M., *Ashkenazi, A., and *Grewal, I.S. TACI-ligand interactions are required for T cell activation and collagen-induced arthritis in mice. *Nature Immunol.* 2, 632-637 (2001).
65. Kischkel, F.C., Lawrence, D. A., Tinel, A., Virmani, A., Schow, P., Gazdar, A., Blenis, J., Arnott, D., and Ashkenazi, A. Death receptor recruitment of endogenous caspase-10 and apoptosis initiation in the absence of caspase-8. *J. Biol. Chem.* 276, 46639-46646 (2001).
66. LeBlanc, H., Lawrence, D.A., Varfolomeev, E., Totpal, K., Morlan, J., Schow, P., Fong, S., Schwall, R., Sinicropi, D., and Ashkenazi, A. Tumor cell resistance to death receptor induced apoptosis through mutational inactivation of the proapoptotic Bcl-2 homolog Bax. *Nature Med.* 8, 274-281 (2002).
67. Miller, K., Meng, G., Liu, J., Hurst, A., Hsei, V., Wong, W-L., Ekert, R., Lawrence, D., Sherwood, S., DeForge, L., Gaudreault, Keller, G., Sliwkowski, M., Ashkenazi, A., and Presta, L. Design, Construction, and analyses of multivalent antibodies. *J. Immunol.* 170, 4854-4861 (2003).

68. Varfolomeev, E., Kischkel, F., Martin, F., Wanh, H., Lawrence, D., Olsson, C., Tom, L., Erickson, S., French, D., Schow, P., Grewal, I. and Ashkenazi, A. Immune system development in APRIL knockout mice. Submitted.

Review articles:

1. Ashkenazi, A., Peralta, E., Winslow, J., Ramachandran, J., and Capon, D., J. Functional role of muscarinic acetylcholine receptor subtype diversity. *Cold Spring Harbor Symposium on Quantitative Biology*. **LIII**, 263-272 (1988).
2. Ashkenazi, A., Peralta, E., Winslow, J., Ramachandran, J., and Capon, D. Functional diversity of muscarinic receptor subtypes in cellular signal transduction and growth. *Trends Pharmacol. Sci.* Dec Supplement, 12-21 (1989).
3. Chamow, S., Duliege, A., Ammann, A., Kahn, J., Allen, D., Eichberg, J., Byrn, R., Capon, D., Ward, R., and Ashkenazi, A. CD4 immunoadhesins in anti-HIV therapy: new developments. *Int. J. Cancer* Supplement 7, 69-72 (1992).
4. Ashkenazi, A., Capon, and D. Ward, R. Immunoadhesins. *Int. Rev. Immunol.* **10**, 217-225 (1993).
5. Ashkenazi, A., and Peralta, E. Muscarinic Receptors. In *Handbook of Receptors and Channels*. (S. Peroutka, ed.), CRC Press, Boca Raton, Vol. I, p. 1-27, (1994).
6. Krantz, S. B., Means, R. T., Jr., Lina, J., Marsters, S. A., and Ashkenazi, A. Inhibition of erythroid colony formation in vitro by gamma interferon. In *Molecular Biology of Hematopoiesis* (N. Abraham, R. Shadduck, A. Levine F. Takaku, eds.) Intercept Ltd. Paris, Vol. 3, p. 135-147 (1994).
7. Ashkenazi, A. Cytokine neutralization as a potential therapeutic approach for SIRS and shock. *J. Biotechnology in Healthcare* **1**, 197-206 (1994).
8. Ashkenazi, A., and Chamow, S. M. Immunoadhesins: an alternative to human monoclonal antibodies. *Immunomethods: A companion to Methods in Enzymology* **8**, 104-115 (1995).
9. Chamow, S., and Ashkenazi, A. Immunoadhesins: Principles and Applications. *Trends Biotech.* **14**, 52-60 (1996).
10. Ashkenazi, A., and Chamow, S. M. Immunoadhesins as research tools and therapeutic agents. *Curr. Opin. Immunol.* **9**, 195-200 (1997).
11. Ashkenazi, A., and Dixit, V. Death receptors: signaling and modulation. *Science* **281**, 1305-1308 (1998).
12. Ashkenazi, A., and Dixit, V. Apoptosis control by death and decoy receptors. *Curr. Opin. Cell. Biol.* **11**, 255-260 (1999).

13. Ashkenazi, A. Chapters on Apo2L/TRAIL; DR4, DR5, DcR1, DcR2; and DcR3. Online Cytokine Handbook (www.apnet.com/cytokinereference/).
14. Ashkenazi, A. Targeting death and decoy receptors of the tumor necrosis factor superfamily. *Nature Rev. Cancer* 2, 420-430 (2002).
15. LeBlanc, H. and Ashkenazi, A. Apoptosis signaling by Apo2L/TRAIL. *Cell Death and Differentiation* 10, 66-75 (2003).
16. Almasan, A. and Ashkenazi, A. Apo2L/TRAIL: apoptosis signaling, biology, and potential for cancer therapy. *Cytokine and Growth Factor Reviews* 14, 337-348 (2003).

Book:

Antibody Fusion Proteins (Chamow, S., and Ashkenazi, A., eds., John Wiley and Sons Inc.) (1999).

Talks:

1. Resistance of primary HIV isolates to CD4 is independent of CD4-gp120 binding affinity. UCSF Symposium, HIV Disease: Pathogenesis and Therapy. Greenelefe, FL, March 1991.
2. Use of immuno-hybrids to extend the half-life of receptors. IBC conference on Biopharmaceutical Half-life Extension. New Orleans, LA, June 1992.
3. Results with TNF receptor Immunoconjugates for the Treatment of Sepsis. IBC conference on Endotoxemia and Sepsis. Philadelphia, PA, June 1992.
4. Immunoconjugates: an alternative to human antibodies. IBC conference on Antibody Engineering. San Diego, CA, December 1993.
5. Tumor necrosis factor receptor: a potential therapeutic for human septic shock. American Society for Microbiology Meeting, Atlanta, GA, May 1993.
6. Protective efficacy of TNF receptor immunoconjugate vs anti-TNF monoclonal antibody in a rat model for endotoxic shock. 5th International Congress on TNF. Asilomar, CA, May 1994.
7. Interferon- γ signals via a multisubunit receptor complex that contains two types of polypeptide chain. American Association of Immunologists Conference. San Francisco, CA, July 1995.
8. Immunoconjugates: Principles and Applications. Gordon Research Conference on Drug Delivery in Biology and Medicine. Ventura, CA, February 1996.

9. Apo-2 Ligand, a new member of the TNF family that induces apoptosis in tumor cells. Cambridge Symposium on TNF and Related Cytokines in Treatment of Cancer. Hilton-Head, NC, March 1996.
10. Induction of apoptosis by Apo2 Ligand. American Society for Biochemistry and Molecular Biology, Symposium on Growth Factors and Cytokine Receptors. New Orleans, LA, June, 1996.
11. Apo2 ligand, an extracellular trigger of apoptosis. 2nd Clontech Symposium, Palo Alto, CA, October 1996.
12. Regulation of apoptosis by members of the TNF ligand and receptor families. Stanford University School of Medicine, Palo Alto, CA, December 1996.
13. Apo-3: a novel receptor that regulates cell death and inflammation. 4th International Congress on Immune Consequences of Trauma, Shock, and Sepsis. Munich, Germany, March 1997.
14. New members of the TNF ligand and receptor families that regulate apoptosis, inflammation, and immunity. UCLA School of Medicine, LA, CA, March 1997.
15. Immunoadhesins: an alternative to monoclonal antibodies. 5th World Conference on Bispecific Antibodies. Volendam, Holland, June 1997.
16. Control of Apo2L signaling. Cold Spring Harbor Laboratory Symposium on Programmed Cell Death. Cold Spring Harbor, New York. September, 1997.
17. Chairman and speaker, Apoptosis Signaling session. IBC's 4th Annual Conference on Apoptosis. San Diego, CA., October 1997.
18. Control of Apo2L signaling by death and decoy receptors. American Association for the Advancement of Science. Philadelphia, PA, February 1998.
19. Apo2 ligand and its receptors. American Society of Immunologists. San Francisco, CA, April 1998.
20. Death receptors and ligands. 7th International TNF Congress. Cape Cod, MA, May 1998.
21. Apo2L as a potential therapeutic for cancer. UCLA School of Medicine. LA, CA, June 1998.
22. Apo2L as a potential therapeutic for cancer. Gordon Research Conference on Cancer Chemotherapy. New London, NH, July 1998.
23. Control of apoptosis by Apo2L. Endocrine Society Conference, Stevenson, WA, August 1998.
24. Control of apoptosis by Apo2L. International Cytokine Society Conference, Jerusalem, Israel, October 1998.

25. Apoptosis control by death and decoy receptors. American Association for Cancer Research Conference, Whistler, BC, Canada, March 1999.
26. Apoptosis control by death and decoy receptors. American Society for Biochemistry and Molecular Biology Conference, San Francisco, CA, May 1999.
27. Apoptosis control by death and decoy receptors. Gordon Research Conference on Apoptosis, New London, NH, June 1999.
28. Apoptosis control by death and decoy receptors. Arthritis Foundation Research Conference, Alexandria GA, Aug 1999.
29. Safety and anti-tumor activity of recombinant soluble Apo2L/TRAIL. Cold Spring Harbor Laboratory Symposium on Programmed Cell Death. . Cold Spring Harbor, NY, September 1999.
30. The Apo2L/TRAIL system: therapeutic potential. American Association for Cancer Research, Lake Tahoe, NV, Feb 2000.
31. Apoptosis and cancer therapy. Stanford University School of Medicine, Stanford, CA, Mar 2000.
32. Apoptosis and cancer therapy. University of Pennsylvania School of Medicine, Philadelphia, PA, Apr 2000.
33. Apoptosis signaling by Apo2L/TRAIL. International Congress on TNF. Trondheim, Norway, May 2000.
34. The Apo2L/TRAIL system: therapeutic potential. Cap-CURE summit meeting. Santa Monica, CA, June 2000.
35. The Apo2L/TRAIL system: therapeutic potential. MD Anderson Cancer Center. Houston, TX, June 2000.
36. Apoptosis signaling by Apo2L/TRAIL. The Protein Society, 14th Symposium. San Diego, CA, August 2000.
37. Anti-tumor activity of Apo2L/TRAIL. AAPS annual meeting. Indianapolis, IN Aug 2000.
38. Apoptosis signaling and anti-cancer potential of Apo2L/TRAIL. Cancer Research Institute, UC San Francisco, CA, September 2000.
39. Apoptosis signaling by Apo2L/TRAIL. Kenote address, TNF family Minisymposium, NIH. Bethesda, MD, September 2000.
40. Death receptors: signaling and modulation. Keystone symposium on the Molecular basis of cancer. Taos, NM, Jan 2001.
41. Preclinical studies of Apo2L/TRAIL in cancer. Symposium on Targeted therapies in the treatment of lung cancer. Aspen, CO, Jan 2001.

42. Apoptosis signaling by Apo2L/TRAIL. Weizmann Institute of Science, Rehovot, Israel, March 2001.
43. Apo2L/TRAIL: Apoptosis signaling and potential for cancer therapy. Weizmann Institute of Science, Rehovot, Israel, March 2001.
44. Targeting death receptors in cancer with Apo2L/TRAIL. Cell Death and Disease conference, North Falmouth, MA, Jun 2001.
45. Targeting death receptors in cancer with Apo2L/TRAIL. Biotechnology Organization conference, San Diego, CA, Jun 2001.
46. Apo2L/TRAIL signaling and apoptosis resistance mechanisms. Gordon Research Conference on Apoptosis, Oxford, UK, July 2001.
47. Apo2L/TRAIL signaling and apoptosis resistance mechanisms. Cleveland Clinic Foundation, Cleveland, OH, Oct 2001.
48. Apoptosis signaling by death receptors: overview. International Society for Interferon and Cytokine Research conference, Cleveland, OH, Oct 2001.
49. Apoptosis signaling by death receptors. American Society of Nephrology Conference. San Francisco, CA, Oct 2001.
50. Targeting death receptors in cancer. Apoptosis: commercial opportunities. San Diego, CA, Apr 2002.
51. Apo2L/TRAIL signaling and apoptosis resistance mechanisms. Kimmel Cancer Research Center, Johns Hopkins University, Baltimore MD. May 2002.
52. Apoptosis control by Apo2L/TRAIL. (Keynote Address) University of Alabama Cancer Center Retreat, Birmingham, Ab. October 2002.
53. Apoptosis signaling by Apo2L/TRAIL. (Session co-chair) TNF international conference. San Diego, CA. October 2002.
54. Apoptosis signaling by Apo2L/TRAIL. Swiss Institute for Cancer Research (ISREC). Lausanne, Swizerland. Jan 2003.
55. Apoptosis induction with Apo2L/TRAIL. Conference on New Targets and Innovative Strategies in Cancer Treatment. Monte Carlo. February 2003.
56. Apoptosis signaling by Apo2L/TRAIL. Hermelin Brain Tumor Center Symposium on Apoptosis. Detroit, MI. April 2003.
57. Targeting apoptosis through death receptors. Sixth Annual Conference on Targeted Therapies in the Treatment of Breast Cancer. Kona, Hawaii. July 2003.
58. Targeting apoptosis through death receptors. Second International Conference on Targeted Cancer Therapy. Washington, DC. Aug 2003.

Issued Patents:

1. Ashkenazi, A., Chamow, S. and Kogan, T. Carbohydrate-directed crosslinking reagents. US patent 5,329,028 (Jul 12, 1994).
2. Ashkenazi, A., Chamow, S. and Kogan, T. Carbohydrate-directed crosslinking reagents. US patent 5,605,791 (Feb 25, 1997).
3. Ashkenazi, A., Chamow, S. and Kogan, T. Carbohydrate-directed crosslinking reagents. US patent 5,889,155 (Jul 27, 1999).
4. Ashkenazi, A., APO-2 Ligand. US patent 6,030,945 (Feb 29, 2000).
5. Ashkenazi, A., Chuntharapai, A., Kim, J., APO-2 ligand antibodies. US patent 6,046,048 (Apr 4, 2000).
6. Ashkenazi, A., Chamow, S. and Kogan, T. Carbohydrate-directed crosslinking reagents. US patent 6,124,435 (Sep 26, 2000).
7. Ashkenazi, A., Chuntharapai, A., Kim, J., Method for making monoclonal and cross-reactive antibodies. US patent 6,252,050 (Jun 26, 2001).
8. Ashkenazi, A. APO-2 Receptor. US patent 6,342,369 (Jan 29, 2002).
9. Ashkenazi, A. Fong, S., Goddard, A., Gurney, A., Napier, M., Tumas, D., Wood, W. A-33 polypeptides. US patent 6,410,708 (Jun 25, 2002).
10. Ashkenazi, A. APO-3 Receptor. US patent 6,462,176 B1 (Oct 8, 2002).
11. Ashkenazi, A. APO-2LI and APO-3 polypeptide antibodies. US patent 6,469,144 B1 (Oct 22, 2002).
12. Ashkenazi, A., Chamow, S. and Kogan, T. Carbohydrate-directed crosslinking reagents. US patent 6,582,928B1 (Jun 24, 2003).

DECLARATION OF PAUL POLAKIS, Ph.D.

I, Paul Polakis, Ph.D., declare and say as follows:

1. I was awarded a Ph.D. by the Department of Biochemistry of the Michigan State University in 1984. My scientific Curriculum Vitae is attached to and forms part of this Declaration (Exhibit A).

2. I am currently employed by Genentech, Inc. where my job title is Staff Scientist. Since joining Genentech in 1999, one of my primary responsibilities has been leading Genentech's Tumor Antigen Project, which is a large research project with a primary focus on identifying tumor cell markers that find use as targets for both the diagnosis and treatment of cancer in humans.

3. As part of the Tumor Antigen Project, my laboratory has been analyzing differential expression of various genes in tumor cells relative to normal cells. The purpose of this research is to identify proteins that are abundantly expressed on certain tumor cells and that are either (i) not expressed, or (ii) expressed at lower levels, on corresponding normal cells. We call such differentially expressed proteins "tumor antigen proteins". When such a tumor antigen protein is identified, one can produce an antibody that recognizes and binds to that protein. Such an antibody finds use in the diagnosis of human cancer and may ultimately serve as an effective therapeutic in the treatment of human cancer.

4. In the course of the research conducted by Genentech's Tumor Antigen Project, we have employed a variety of scientific techniques for detecting and studying differential gene expression in human tumor cells relative to normal cells, at genomic DNA, mRNA and protein levels. An important example of one such technique is the well known and widely used technique of microarray analysis which has proven to be extremely useful for the identification of mRNA molecules that are differentially expressed in one tissue or cell type relative to another. In the course of our research using microarray analysis, we have identified approximately 200 gene transcripts that are present in human tumor cells at significantly higher levels than in corresponding normal human cells. To date, we have generated antibodies that bind to about 30 of the tumor antigen proteins expressed from these differentially expressed gene transcripts and have used these antibodies to quantitatively determine the level of production of these tumor antigen proteins in both human cancer cells and corresponding normal cells. We have then compared the levels of mRNA and protein in both the tumor and normal cells analyzed.

5. From the mRNA and protein expression analyses described in paragraph 4 above, we have observed that there is a strong correlation between changes in the level of mRNA present in any particular cell type and the level of protein

expressed from that mRNA in that cell type. In approximately 80% of our observations we have found that increases in the level of a particular mRNA correlates with changes in the level of protein expressed from that mRNA when human tumor cells are compared with their corresponding normal cells.

6. Based upon my own experience accumulated in more than 20 years of research, including the data discussed in paragraphs 4 and 5 above and my knowledge of the relevant scientific literature, it is my considered scientific opinion that for human genes, an increased level of mRNA in a tumor cell relative to a normal cell typically correlates to a similar increase in abundance of the encoded protein in the tumor cell relative to the normal cell. In fact, it remains a central dogma in molecular biology that increased mRNA levels are predictive of corresponding increased levels of the encoded protein. While there have been published reports of genes for which such a correlation does not exist, it is my opinion that such reports are exceptions to the commonly understood general rule that increased mRNA levels are predictive of corresponding increased levels of the encoded protein.

7. I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information or belief are believed to be true, and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful statements may jeopardize the validity of the application or any patent issued thereon.

Dated: 5/07/04

By: Paul Polakis

Paul Polakis, Ph.D.

CURRICULUM VITAE

PAUL G. POLAKIS
Staff Scientist
Genentech, Inc
1 DNA Way, MS#40
S. San Francisco, CA 94080

EDUCATION:

Ph.D., Biochemistry, Department of Biochemistry,
Michigan State University (1984)

B.S., Biology. College of Natural Science, Michigan State University (1977)

PROFESSIONAL EXPERIENCE:

| | |
|--------------|---|
| 2002-present | Staff Scientist, Genentech, Inc S. San Francisco, CA |
| 1999- 2002 | Senior Scientist, Genentech, Inc., S. San Francisco, CA |
| 1997 -1999 | Research Director Onyx Pharmaceuticals, Richmond, CA |
| 1992- 1996 | Senior Scientist, Project Leader, Onyx Pharmaceuticals, Richmond, CA |
| <hr/> | |
| 1991-1992 | Senior Scientist, Chiron Corporation, Emeryville, CA. |
| 1989-1991 | Scientist, Cetus Corporation, Emeryville CA. |
| 1987-1989 | Postdoctoral Research Associate, Genentech, Inc., South San Francisco, CA. |
| 1985-1987 | Postdoctoral Research Associate, Department of Medicine, Duke University Medical Center, Durham, NC |

1984-1985

Assistant Professor, Department of Chemistry,
Oberlin College, Oberlin, Ohio

1980-1984

Graduate Research Assistant, Department of
Biochemistry, Michigan State University
East Lansing, Michigan

PUBLICATIONS:

1. Polakis, P. G. and Wilson, J. E. 1982 Purification of a Highly Bindable Rat Brain Hexokinase by High Performance Liquid Chromatography. **Biochem. Biophys. Res. Commun.** 107, 937-943.
2. Polakis, P.G. and Wilson, J. E. 1984 Proteolytic Dissection of Rat Brain Hexokinase: Determination of the Cleavage Pattern during Limited Digestion with Trypsin. **Arch. Biochem. Biophys.** 234, 341-352.
3. Polakis, P. G. and Wilson, J. E. 1985 An Intact Hydrophobic N-Terminal Sequence is Required for the Binding Rat Brain Hexokinase to Mitochondria. **Arch. Biochem. Biophys.** 236, 328-337.
4. Uhing, R.J., Polakis, P.G. and Snyderman, R. 1987 Isolation of GTP-binding Proteins from Myeloid HL60 Cells. **J. Biol. Chem.** 262, 15575-15579.
5. Polakis, P.G., Uhing, R.J. and Snyderman, R. 1988 The Formylpeptide Chemoattractant Receptor Copurifies with a GTP-binding Protein Containing a Distinct 40 kDa Pertussis Toxin Substrate. **J. Biol. Chem.** 263, 4969-4979.
6. Uhing, R. J., Dillon, S., Polakis, P. G., Truett, A. P. and Snyderman, R. 1988 Chemoattractant Receptors and Signal Transduction Processes in Cellular and Molecular Aspects of Inflammation (Poste, G. and Crooke, S. T. eds.) pp 335-379.
7. Polakis, P.G., Evans, T. and Snyderman 1989 Multiple Chromatographic Forms of the Formylpeptide Chemoattractant Receptor and their Relationship to GTP-binding Proteins. **Biochem. Biophys. Res. Commun.** 161, 276-283.
8. Polakis, P. G., Snyderman, R. and Evans, T. 1989 Characterization of G25K, a GTP-binding Protein Containing a Novel Putative Nucleotide Binding Domain. **Biochem. Biophys. Res. Commun.** 160, 25-32.
9. Polakis, P., Weber, R.F., Nevins, B., Didsbury, J. Evans, T. and Snyderman, R. 1989 Identification of the *ral* and *rac1* Gene Products, Low Molecular Mass GTP-binding Proteins from Human Platelets. **J. Biol. Chem.** 264, 16383-16389.
10. Snyderman, R., Perianin, A., Evans, T., Polakis, P. and Didsbury, J. 1989 G Proteins and Neutrophil Function. In ADP-Ribosylating Toxins and G Proteins: Insights into Signal Transduction. (J. Moss and M. Vaughn, eds.) Amer. Soc. Microbiol. pp. 295-323.

11. Hart, M.J., Polakis, P.G., Evans, T. and Cerrione, R.A. 1990 The identification and Characterization of an Epidermal Growth Factor-Stimulated Phosphorylation of a Specific Low Molecular Mass GTP-binding Protein in a Reconstituted Phospholipid Vesicle System. *J. Biol. Chem.* 265, 5990-6001.

12. Yatani, A., Okabe, K., Polakis, P., Halenbeck, R., McCormick, F. and Brown, A. M. 1990 ras p21 and GAP Inhibit Coupling of Muscarinic Receptors to Atrial K⁺ Channels. *Cell*. 61, 769-776.

13. Munemitsu, S., Innis, M.A., Clark, R., McCormick, F., Ullrich, A. and Polakis, P.G. 1990 Molecular Cloning and Expression of a G25K cDNA, the Human Homolog of the Yeast Cell Cycle Gene CDC42. *Mol. Cell. Biol.* 10, 5977-5982.

14. Polakis, P.G., Rubinfeld, B., Evans, T. and McCormick, F. 1991 Purification of Plasma Membrane-Associated GTPase Activating Protein (GAP) Specific for rap-1/krev-1 from HL60 Cells. *Proc. Natl. Acad. Sci. USA* 88, 239-243.

15. Moran, M. F., Polakis, P., McCormick, F., Pawson, T. and Ellis, C. 1991 Protein Tyrosine Kinases Regulate the Phosphorylation, Protein Interactions, Subcellular Distribution, and Activity of p21ras GTPase Activating Protein. *Mol. Cell. Biol.* 11, 1804-1812

16. Rubinfeld, B., Wong, G., Bekesi, E., Wood, A., McCormick, F. and Polakis, P. G. 1991 A Synthetic Peptide Corresponding to a Sequence in the GTPase Activating Protein Inhibits p21^{ras} Stimulation and Promotes Guanine Nucleotide Exchange. *Internatl. J. Peptide and Prot. Res.* 38, 47-53.

17. Rubinfeld, B., Munemitsu, S., Clark, R., Conroy, L., Watt, K., Crosier, W., McCormick, F., and Polakis, P. 1991 Molecular Cloning of a GTPase Activating Protein Specific for the Krev-1 Protein p21^{rap1}. *Cell* 65, 1033-1042.

18. Zhang, K., Papageorge, A., G., Martin, P., Vass, W. C., Olah, Z., Polakis, P., McCormick, F. and Lowy, D. R. 1991 Heterogenous Amino Acids in RAS and Rap1A Specifying Sensitivity to GAP Proteins. *Science* 254, 1630-1634.

19. Martin, G., Yatani, A., Clark, R., Polakis, P., Brown, A. M. and McCormick, F. 1992 GAP Domains Responsible for p21^{ras}-dependent Inhibition of Muscarinic Atrial K⁺ Channel Currents. *Science* 255, 192-194.

20. McCormick, F., Martin, G. A., Clark, R., Bollag, G. and Polakis, P. 1992 Regulation of p21ras by GTPase Activating Proteins. *Cold Spring Harbor Symposia on Quantitative Biology*. Vol. 56, 237-241.

21. Pronk, G. B., Polakis, P., Wong, G., deVries-Smits, A. M., Bos J. L. and McCormick, F. 1992 p60^{v-src} Can Associate with and Phosphorylate the p21^{ras} GTPase Activating Protein. *Oncogene* 7, 389-394.

22. Polakis P. and McCormick, F. 1992 Interactions Between p21^{ras} Proteins and Their GTPase Activating Proteins. In *Cancer Surveys* (Franks, L. M., ed.) 12, 25-42.

23. Wong, G., Muller, O., Clark, R., Conroy, L., Moran, M., Polakis, P. and McCormick, F. 1992 Molecular cloning and nucleic acid binding properties of the GAP-associated tyrosine phosphoprotein p62. *Cell* 69, 551-558.
24. Polakis, P., Rubinfeld, B. and McCormick, F. 1992 Phosphorylation of rap1GAP in vivo and by cAMP-dependent Kinase and the Cell Cycle p34^{cdc2} Kinase in vitro. *J. Biol. Chem.* 267, 10780-10785.
25. McCabe, P.C., Haubrauck, H., Polakis, P., McCormick, F., and Innis, M. A. 1992 Functional Interactions Between p21^{rap1A} and Components of the Budding pathway of *Saccharomyces cerevisiae*. *Mol. Cell. Biol.* 12, 4084-4092.
26. Rubinfeld, B., Crosier, W.J., Albert, I., Conroy, L., Clark, R., McCormick, F. and Polakis, P. 1992 Localization of the rap1GAP Catalytic Domain and Sites of Phosphorylation by Mutational Analysis. *Mol. Cell. Biol.* 12, 4634-4642.
27. Ando, S., Kaibuchi, K., Sasaki, K., Hiraoka, T., Nishiyama, T., Mizuno, T., Asada, M., Nunoi, H., Matsuda, I., Matsuura, Y., Polakis, P., McCormick, F. and Takai, Y. 1992 Post-translational processing of rac p21s is important both for their interaction with the GDP/GTP exchange proteins and for their activation of NADPH oxidase. *J. Biol. Chem.* 267, 25709-25713.
28. Janoueix-Lerosey, I., Polakis, P., Tavitian, A. and deGunzberg, J. 1992 Regulation of the GTPase activity of the ras-related rap2 protein. *Biochem. Biophys. Res. Commun.* 189, 455-464.
29. Polakis, P. 1993 GAPs Specific for the rap1/Krev-1 Protein. in GTP-binding Proteins: the ras-superfamily. (J.C. LaCale and F. McCormick, eds.) 445-452.
30. Polakis, P. and McCormick, F. 1993 Structural requirements for the interaction of p21^{ras} with GAP, exchange factors, and its biological effector target. *J. Biol. Chem.* 268, 9157-9160.
31. Rubinfeld, B., Souza, B., Albert, I., Muller, O., Chamberlain, S., Masiarz, F., Munemitsu, S. and Polakis, P. 1993 Association of the APC gene product with beta- catenin. *Science* 262, 1731-1734.
32. Weiss, J., Rubinfeld, B., Polakis, P., McCormick, F., Cavenee, W. A. and Arden, K. 1993 The gene for human rap1-GTPase activating protein (rap1GAP) maps to chromosome 1p35-1p36.1. *Cytogenet. Cell Genet.* 66, 18-21.
33. Sato, K. Y., Polakis, P., Haubruck, H., Fasching, C. L., McCormick, F. and Stanbridge, E. J. 1994 Analysis of the tumor suppressor activity of the K-rev gene in human tumor cell lines. *Cancer Res.* 54, 552-559.
34. Janoueix-Lerosey, I., Fontenay, M., Tobelem, G., Tavitian, A., Polakis, P. and DeGunzburg, J. 1994 Phosphorylation of rap1GAP during the cell cycle. *Biochem. Biophys. Res. Commun.* 202, 967-975
35. Munemitsu, S., Souza, B., Mueller, O., Albert, I., Rubinfeld, B., and Polakis, P. 1994 The APC gene product associates with microtubules in vivo and affects their assembly in vitro. *Cancer Res.* 54, 3676-3681.

36. Rubinfeld, B. and Polakis, P. 1995 Purification of baculovirus produced rap1GAP. **Methods Enz.** 255,31
37. Polakis, P. 1995 Mutations in the APC gene and their implications for protein structure and function. **Current Opinions in Genetics and Development** 5, 66-71
38. Rubinfeld, B., Souza, B., Albert, I., Munemitsu, S. and Polakis P. 1995 The APC protein and E-cadherin form similar but independent complexes with α -catenin, β -catenin and Plakoglobin. **J. Biol. Chem.** 270, 5549-5555
39. Munemitsu, S., Albert, I., Souza, B., Rubinfeld, B., and Polakis, P. 1995 Regulation of intracellular β -catenin levels by the APC tumor suppressor gene. **Proc. Natl. Acad. Sci.** 92, 3046-3050.
40. Lock, P., Fumagalli, S., Polakis, P. McCormick, F. and Courtneidge, S. A. 1996 The human p62 cDNA encodes Sam68 and not the rasGAP-associated p62 protein. **Cell** 84, 23-24.
41. Papkoff, J., Rubinfeld, B., Schryver, B. and Polakis, P. 1996 Wnt-1 regulates free pools of catenins and stabilizes APC-catenin complexes. **Mol. Cell. Biol.** 16, 2128-2134.
42. Rubinfeld, B., Albert, I., Porfiri, E., Fiol, C., Munemitsu, S. and Polakis, P. 1996 Binding of GSK3 β to the APC- β -catenin complex and regulation of complex assembly. **Science** 272, 1023-1026.
43. Munemitsu, S., Albert, I., Rubinfeld, B. and Polakis, P. 1996 Deletion of amino-terminal structure stabilizes β -catenin in vivo and promotes the hyperphosphorylation of the APC tumor suppressor protein. **Mol. Cell. Biol.** 16, 4088-4094.
44. Hart, M. J., Callow, M. G., Sousa, B. and Polakis P. 1996 IQGAP1, a calmodulin binding protein with a rasGAP related domain, is a potential effector for cdc42Hs. **EMBO J.** 15, 2997-3005.
45. Nathke, I. S., Adams, C. L., Polakis, P., Sellin, J. and Nelson, W. J. 1996 The adenomatous polyposis coli (APC) tumor suppressor protein is localized to plasma membrane sites involved in active epithelial cell migration. **J. Cell. Biol.** 134, 165-180.
46. Hart, M. J., Sharma, S., elMasry, N., Qui, R-G., McCabe, P., Polakis, P. and Bollag, G. 1996 Identification of a novel guanine nucleotide exchange factor for the rho GTPase. **J. Biol. Chem.** 271, 25452.
47. Thomas JE, Smith M, Rubinfeld B, Gutowski M, Beckmann RP, and Polakis P. 1996 Subcellular localization and analysis of apparent 180-kDa and 220-kDa proteins of the breast cancer susceptibility gene, BRCA1. **J. Biol. Chem.** 1996 271, 28630-28635
48. Hayashi, S., Rubinfeld, B., Souza, B., Polakis, P., Wieschaus, E., and Levine, A. 1997 A Drosophila homolog of the tumor suppressor adenomatous polyposis coli

down-regulates β -catenin but its zygotic expression is not essential for the regulation of armadillo. **Proc. Natl. Acad. Sci.** 94, 242-247.

49. Vleminckx, K., Rubinfeld, B., Polakis, P. and Gumbiner, B. 1997 The APC tumor suppressor protein induces a new axis in *Xenopus* embryos. **J. Cell. Biol.** 136, 411-420.

50. Rubinfeld, B., Robbins, P., El-Gamil, M., Albert, I., Porfiri, P. and Polakis, P. 1997 Stabilization of β -catenin by genetic defects in melanoma cell lines. **Science** 275, 1790-1792.

51. Polakis, P. The adenomatous polyposis coli (APC) tumor suppressor. 1997 **Biochem. Biophys. Acta**, 1332, F127-F147.

52. Rubinfeld, B., Albert, I., Porfiri, E., Munemitsu, S., and Polakis, P. 1997 Loss of β -catenin regulation by the APC tumor suppressor protein correlates with loss of structure due to common somatic mutations of the gene. **Cancer Res.** 57, 4624-4630.

53. Porfiri, E., Rubinfeld, B., Albert, I., Hovanes, K., Waterman, M., and Polakis, P. 1997 Induction of a β -catenin-LEF-1 complex by wnt-1 and transforming mutants of β -catenin. **Oncogene** 15, 2833-2839.

54. Thomas JE, Smith M, Tonkinson JL, Rubinfeld B, and Polakis P., 1997 Induction of phosphorylation on BRCA1 during the cell cycle and after DNA damage. **Cell Growth Differ.** 8, 801-809.

55. Hart, M., de los Santos, R., Albert, I., Rubinfeld, B., and Polakis P., 1998 Down regulation of β -catenin by human Axin and its association with the adenomatous polyposis coli (APC) tumor suppressor, β -catenin and glycogen synthase kinase 3 β . **Current Biology** 8, 573-581.

56. Polakis, P. 1998 The oncogenic activation of β -catenin. **Current Opinions in Genetics and Development** 9, 15-21

57. Matt Hart, Jean-Paul Concordet, Irina Lassot, Iris Albert, Rico del los Santos, Herve Durand, Christine Perret, Bonnee Rubinfeld, Florence Margottin, Richard Benarous and Paul Polakis. 1999 The F-box protein β -TrCP associates with phosphorylated β -catenin and regulates its activity in the cell. **Current Biology** 9, 207-10.

58. Howard C. Crawford, Barbara M. Fingleton, Bonnee Rubinfeld, Paul Polakis and Lynn M. Matrisian 1999 The metalloproteinase matrilysin is a target of β -catenin transactivation in intestinal tumours. **Oncogene** 18, 2883-91.

59. Meng J, Glick JL, Polakis P, Casey PJ. 1999 Functional interaction between G α (z) and Rap1GAP suggests a novel form of cellular cross-talk. **J Biol Chem.** 17, 36663-9

60. Vijayasurian Easwaran, Virginia Song, **Paul Polakis** and Steve Byers 1999 The ubiquitin-proteasome pathway and serine kinase activity modulate APC mediated regulation of β -catenin-LEF signaling. **J. Biol. Chem.** 274(23):16641-5.

61 **Polakis P**, Hart M and Rubinfeld B. 1999. Defects in the regulation of beta-catenin in colorectal cancer. **Adv Exp Med Biol.** 470, 23-32

62 Shen Z, Batzer A, Koehler JA, **Polakis P**, Schlessinger J, Lydon NB, Moran MF. 1999 Evidence for SH3 domain directed binding and phosphorylation of Sam68 by Src. **Oncogene.** 18, 4647-53

64. Thomas GM, Frame S, Goedert M, Nathke I, **Polakis P**, Cohen P. 1999 A GSK3- binding peptide from FRAT1 selectively inhibits the GSK3-catalysed phosphorylation of axin and beta-catenin. **FEBS Lett.** 458, 247-51.

65. Peifer M, **Polakis P**. 2000 Wnt signaling in oncogenesis and embryogenesis—a look outside the nucleus. **Science** 287,1606-9.

66. **Polakis P**. 2000 Wnt signaling and cancer. **Genes Dev**;14, 1837-1851.

67. Spink KE, **Polakis P**, Weis WI 2000 Structural basis of the Axin-adenomatous polyposis coli interaction. **EMBO J** 19, 2270-2279.

68. Szeto, W., Jiang, W., Tice, D.A., Rubinfeld, B., Hollingshead, P.G., Fong, S.E., Dugger, D.L., Pham, T., Yansura, D.E., Wong, T.A., Grimaldi, J.C., Corpuz, R.T., Singh J.S., Frantz, G.D., Devaux, B., Crowley, C.W., Schwall, R.H., Eberhard, D.A., Rastelli, L., **Polakis, P.** and Pennica, D. 2001 Overexpression of the Retinoic Acid-Responsive Gene Stra6 in Human Cancers and its Synergistic Induction by Wnt-1 and Retinoic Acid. **Cancer Res** 61, 4197-4204.

69. Rubinfeld B, Tice DA, **Polakis P**. 2001 Axin dependent phosphorylation of the adenomatous polyposis coli protein mediated by casein kinase 1 epsilon. **J Biol Chem** 276, 39037-39045.

70. **Polakis P**. 2001 More than one way to skin a catenin. **Cell** 2001 105, 563-566.

71. Tice DA, Soloviev I, **Polakis P**. 2002 Activation of the Wnt Pathway Interferes with Serum Response Element-driven Transcription of Immediate Early Genes. **J Biol. Chem.** 277, 6118-6123.

72. Tice DA, Szeto W, Soloviev I, Rubinfeld B, Fong SE, Dugger DL, Winer J,

Williams PM, Wieand D, Smith V, Schwall RH, Pennica D, Polakis P. 2002 Synergistic activation of tumor antigens by wnt-1 signaling and retinoic acid revealed by gene expression profiling. **J Biol Chem.** 277,14329-14335.

73. Polakis, P. 2002 Casein kinase I: A wnt'er of disconnect. **Curr. Biol.** 12, R499.

74. Mao, W., Luis, E., Ross, S., Silva, J., Tan, C., Crowley, C., Chui, C., Franz, G., Senter, P., Koeppen, H., Polakis, P. 2004 EphB2 as a therapeutic antibody drug target for the treatment of colorectal cancer. **Cancer Res.** 64, 781-788.

75. Shibamoto, S., Winer, J., Williams, M., Polakis, P. 2003 A Blockade in Wnt signaling is activated following the differentiation of F9 teratocarcinoma cells. **Exp. Cell Res.** 29211-20.

76. Zhang Y, Eberhard DA, Frantz GD, Dowd P, Wu TD, Zhou Y, Watanabe C, Luoh SM, Polakis P, Hillan KJ, Wood WI, Zhang Z. 2004 GEPIS--quantitative gene expression profiling in normal and cancer tissues. **Bioinformatics**, April 8

DECLARATION OF PAUL POLAKIS, Ph.D.

I, Paul Polakis, Ph.D., declare and say as follows:

1. I was awarded a Ph.D. by the Department of Biochemistry of the Michigan State University in 1984. My scientific Curriculum Vitae is attached to and forms part of this Declaration (Exhibit A).
2. I am currently employed by Genentech, Inc. where my job title is Staff Scientist. Since joining Genentech in 1999, one of my primary responsibilities has been leading Genentech's Tumor Antigen Project, which is a large research project with a primary focus on identifying tumor cell markers that find use as targets for both the diagnosis and treatment of cancer in humans.
3. As part of the Tumor Antigen Project, my laboratory has been analyzing differential expression of various genes in tumor cells relative to normal cells. The purpose of this research is to identify proteins that are abundantly expressed on certain tumor cells and that are either (i) not expressed, or (ii) expressed at lower levels, on corresponding normal cells. We call such differentially expressed proteins "tumor antigen proteins". When such a tumor antigen protein is identified, one can produce an antibody that recognizes and binds to that protein. Such an antibody finds use in the diagnosis of human cancer and may ultimately serve as an effective therapeutic in the treatment of human cancer.
4. In the course of the research conducted by Genentech's Tumor Antigen Project, we have employed a variety of scientific techniques for detecting and studying differential gene expression in human tumor cells relative to normal cells, at genomic DNA, mRNA and protein levels. An important example of one such technique is the well known and widely used technique of microarray analysis which has proven to be extremely useful for the identification of mRNA molecules that are differentially expressed in one tissue or cell type relative to another. In the course of our research using microarray analysis, we have identified approximately 200 gene transcripts that are present in human tumor cells at significantly higher levels than in corresponding normal human cells. To date, we have generated antibodies that bind to about 30 of the tumor antigen proteins expressed from these differentially expressed gene transcripts and have used these antibodies to quantitatively determine the level of production of these tumor antigen proteins in both human cancer cells and corresponding normal cells. We have then compared the levels of mRNA and protein in both the tumor and normal cells analyzed.
5. From the mRNA and protein expression analyses described in paragraph 4 above, we have observed that there is a strong correlation between changes in the level of mRNA present in any particular cell type and the level of protein

expressed from that mRNA in that cell type. In approximately 80% of our observations we have found that increases in the level of a particular mRNA correlates with changes in the level of protein expressed from that mRNA when human tumor cells are compared with their corresponding normal cells.

6. Based upon my own experience accumulated in more than 20 years of research, including the data discussed in paragraphs 4 and 5 above and my knowledge of the relevant scientific literature, it is my considered scientific opinion that for human genes, an increased level of mRNA in a tumor cell relative to a normal cell typically correlates to a similar increase in abundance of the encoded protein in the tumor cell relative to the normal cell. In fact, it remains a central dogma in molecular biology that increased mRNA levels are predictive of corresponding increased levels of the encoded protein. While there have been published reports of genes for which such a correlation does not exist, it is my opinion that such reports are exceptions to the commonly understood general rule that increased mRNA levels are predictive of corresponding increased levels of the encoded protein.

7. I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information or belief are believed to be true, and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful statements may jeopardize the validity of the application or any patent issued thereon.

Dated: 5/07/04

By: Paul Polakis

Paul Polakis, Ph.D.

CURRICULUM VITAE

PAUL G. POLAKIS
Staff Scientist
Genentech, Inc
1 DNA Way, MS#40
S. San Francisco, CA 94080

EDUCATION:

Ph.D., Biochemistry, Department of Biochemistry,
Michigan State University (1984)

B.S., Biology. College of Natural Science, Michigan State University (1977)

PROFESSIONAL EXPERIENCE:

2002-present

Staff Scientist, Genentech, Inc
S. San Francisco, CA

1999- 2002

Senior Scientist, Genentech, Inc.,
S. San Francisco, CA

1997 -1999

Research Director
Onyx Pharmaceuticals, Richmond, CA

1992- 1996

Senior Scientist, Project Leader, Onyx
Pharmaceuticals, Richmond, CA

1991-1992

Senior Scientist, Chiron Corporation,
Emeryville, CA.

1989-1991

Scientist, Cetus Corporation, Emeryville CA.

1987-1989

Postdoctoral Research Associate, Genentech,
Inc., South San Francisco, CA.

1985-1987

Postdoctoral Research Associate, Department
of Medicine, Duke University Medical Center,
Durham, NC

1984-1985

Assistant Professor, Department of Chemistry,
Oberlin College, Oberlin, Ohio

1980-1984

Graduate Research Assistant, Department of
Biochemistry, Michigan State University
East Lansing, Michigan

PUBLICATIONS:

1. Polakis, P. G. and Wilson, J. E. 1982 Purification of a Highly Bindable Rat Brain Hexokinase by High Performance Liquid Chromatography. **Biochem. Biophys. Res. Commun.** 107, 937-943.
2. Polakis, P.G. and Wilson, J. E. 1984 Proteolytic Dissection of Rat Brain Hexokinase: Determination of the Cleavage Pattern during Limited Digestion with Trypsin. **Arch. Biochem. Biophys.** 234, 341-352.
3. Polakis, P. G. and Wilson, J. E. 1985 An Intact Hydrophobic N-Terminal Sequence is Required for the Binding Rat Brain Hexokinase to Mitochondria. **Arch. Biochem. Biophys.** 236, 328-337.
4. Uhing, R.J., Polakis, P.G. and Snyderman, R. 1987 Isolation of GTP-binding Proteins from Myeloid HL60 Cells. **J. Biol. Chem.** 262, 15575-15579.
5. Polakis, P.G., Uhing, R.J. and Snyderman, R. 1988 The Formylpeptide Chemoattractant Receptor Copurifies with a GTP-binding Protein Containing a Distinct 40 kDa Pertussis Toxin Substrate. **J. Biol. Chem.** 263, 4969-4979.
6. Uhing, R. J., Dillon, S., Polakis, P. G., Truett, A. P. and Snyderman, R. 1988 Chemoattractant Receptors and Signal Transduction Processes in Cellular and Molecular Aspects of Inflammation (Poste, G. and Crooke, S. T. eds.) pp 335-379.
7. Polakis, P.G., Evans, T. and Snyderman 1989 Multiple Chromatographic Forms of the Formylpeptide Chemoattractant Receptor and their Relationship to GTP-binding Proteins. **Biochem. Biophys. Res. Commun.** 161, 276-283.
8. Polakis, P. G., Snyderman, R. and Evans, T. 1989 Characterization of G25K, a GTP-binding Protein Containing a Novel Putative Nucleotide Binding Domain. **Biochem. Biophys. Res. Commun.** 160, 25-32.
9. Polakis, P., Weber, R.F., Nevins, B., Didsbury, J. Evans, T. and Snyderman, R. 1989 Identification of the ral and rac1 Gene Products, Low Molecular Mass GTP-binding Proteins from Human Platelets. **J. Biol. Chem.** 264, 16383-16389.
10. Snyderman, R., Perianin, A., Evans, T., Polakis, P. and Didsbury, J. 1989 G Proteins and Neutrophil Function. In ADP-Ribosylating Toxins and G Proteins: Insights into Signal Transduction. (J. Moss and M. Vaughn, eds.) Amer. Soc. Microbiol. pp. 295-323.

11. Hart, M.J., Polakis, P.G., Evans, T. and Cerrione, R.A. 1990 The Identification and Characterization of an Epidermal Growth Factor-Stimulated Phosphorylation of a Specific Low Molecular Mass GTP-binding Protein in a Reconstituted Phospholipid Vesicle System. *J. Biol. Chem.* 265, 5990-6001.
12. Yatani, A., Okabe, K., Polakis, P., Halenbeck, R., McCormick, F. and Brown, A. M. 1990 ras p21 and GAP Inhibit Coupling of Muscarinic Receptors to Atrial K⁺ Channels. *Cell*. 61, 769-776.
13. Munemitsu, S., Innis, M.A., Clark, R., McCormick, F., Ullrich, A. and Polakis, P.G. 1990 Molecular Cloning and Expression of a G25K cDNA, the Human Homolog of the Yeast Cell Cycle Gene CDC42. *Mol. Cell. Biol.* 10, 5977-5982.
14. Polakis, P.G., Rubinfeld, B., Evans, T. and McCormick, F. 1991 Purification of Plasma Membrane-Associated GTPase Activating Protein (GAP) Specific for rap-1/krev-1 from HL60 Cells. *Proc. Natl. Acad. Sci. USA* 88, 239-243.
15. Moran, M. F., Polakis, P., McCormick, F., Pawson, T. and Ellis, C. 1991 Protein Tyrosine Kinases Regulate the Phosphorylation, Protein Interactions, Subcellular Distribution, and Activity of p21ras GTPase Activating Protein. *Mol. Cell. Biol.* 11, 1804-1812.
16. Rubinfeld, B., Wong, G., Bekesi, E., Wood, A., McCormick, F. and Polakis, P. G. 1991 A Synthetic Peptide Corresponding to a Sequence in the GTPase Activating Protein Inhibits p21^{ras} Stimulation and Promotes Guanine Nucleotide Exchange. *Internatl. J. Peptide and Prot. Res.* 38, 47-53.
17. Rubinfeld, B., Munemitsu, S., Clark, R., Conroy, L., Watt, K., Crosier, W., McCormick, F., and Polakis, P. 1991 Molecular Cloning of a GTPase Activating Protein Specific for the Krev-1 Protein p21^{rap1}. *Cell* 65, 1033-1042.
18. Zhang, K., Papageorge, A., G., Martin, P., Vass, W. C., Olah, Z., Polakis, P., McCormick, F. and Lowy, D. R. 1991 Heterogenous Amino Acids in RAS and Rap1A Specifying Sensitivity to GAP Proteins. *Science* 254, 1630-1634.
19. Martin, G., Yatani, A., Clark, R., Polakis, P., Brown, A. M. and McCormick, F. 1992 GAP Domains Responsible for p21^{ras}-dependent Inhibition of Muscarinic Atrial K⁺ Channel Currents. *Science* 255, 192-194.
20. McCormick, F., Martin, G. A., Clark, R., Bollag, G. and Polakis, P. 1992 Regulation of p21^{ras} by GTPase Activating Proteins. *Cold Spring Harbor Symposia on Quantitative Biology*. Vol. 56, 237-241.
21. Pronk, G. B., Polakis, P., Wong, G., deVries-Smits, A. M., Bos J. L. and McCormick, F. 1992 p60^{v-src} Can Associate with and Phosphorylate the p21^{ras} GTPase Activating Protein. *Oncogene* 7, 389-394.
22. Polakis P. and McCormick, F. 1992 Interactions Between p21^{ras} Proteins and Their GTPase Activating Proteins. In Cancer Surveys (Franks, L. M., ed.) 12, 25-42.

23. Wong, G., Muller, O., Clark, R., Conroy, L., Moran, M., Polakis, P. and McCormick, F. 1992 Molecular cloning and nucleic acid binding properties of the GAP-associated tyrosine phosphoprotein p62. **Cell** 69, 551-558.
24. Polakis, P., Rubinfeld, B. and McCormick, F. 1992 Phosphorylation of rap1GAP in vivo and by cAMP-dependent Kinase and the Cell Cycle p34^{cdc2} Kinase in vitro. **J. Biol. Chem.** 267, 10780-10785.
25. McCabe, P.C., Haubrauck, H., Polakis, P., McCormick, F., and Innis, M. A. 1992 Functional Interactions Between p21^{rap1A} and Components of the Budding pathway of *Saccharomyces cerevisiae*. **Mol. Cell. Biol.** 12, 4084-4092.
26. Rubinfeld, B., Crosier, W.J., Albert, I., Conroy, L., Clark, R., McCormick, F. and Polakis, P. 1992 Localization of the rap1GAP Catalytic Domain and Sites of Phosphorylation by Mutational Analysis. **Mol. Cell. Biol.** 12, 4634-4642.
27. Ando, S., Kaibuchi, K., Sasaki, K., Hiraoka, T., Nishiyama, T., Mizuno, T., Asada, M., Nuno, H., Matsuda, I., Matsuura, Y., Polakis, P., McCormick, F. and Takai, Y. 1992 Post-translational processing of rac p21s is important both for their interaction with the GDP/GTP exchange proteins and for their activation of NADPH oxidase. **J. Biol. Chem.** 267, 25709-25713.
28. Janoueix-Lerosey, I., Polakis, P., Tavittian, A. and deGunzburg, J. 1992 Regulation of the GTPase activity of the ras-related rap2 protein. **Biochem. Biophys. Res. Commun.** 189, 455-464.
29. Polakis, P. 1993 GAPs Specific for the rap1/Krev-1 Protein. in GTP-binding Proteins: the ras-superfamily. (J.C. LaCale and F. McCormick, eds.) 445-452.
30. Polakis, P. and McCormick, F. 1993 Structural requirements for the interaction of p21^{ras} with GAP, exchange factors, and its biological effector target. **J. Biol. Chem.** 268, 9157-9160.
31. Rubinfeld, B., Souza, B., Albert, I., Muller, O., Chamberlain, S., Masiarz, F., Munemitsu, S. and Polakis, P. 1993 Association of the APC gene product with beta- catenin. **Science** 262, 1731-1734.
32. Weiss, J., Rubinfeld, B., Polakis, P., McCormick, F., Cavenee, W. A. and Arden, K. 1993 The gene for human rap1-GTPase activating protein (rap1GAP) maps to chromosome 1p35-1p36.1. **Cytogenet. Cell Genet.** 66, 18-21.
33. Sato, K. Y., Polakis, P., Haubruck, H., Fasching, C. L., McCormick, F. and Stanbridge, E. J. 1994 Analysis of the tumor suppressor activity of the K-rev gene in human tumor cell lines. **Cancer Res.** 54, 552-559.
34. Janoueix-Lerosey, I., Fontenay, M., Tobelem, G., Tavittian, A., Polakis, P. and DeGunzburg, J. 1994 Phosphorylation of rap1GAP during the cell cycle. **Biochem. Biophys. Res. Commun.** 202, 967-975
35. Munemitsu, S., Souza, B., Mueller, O., Albert, I., Rubinfeld, B., and Polakis, P. 1994 The APC gene product associates with microtubules in vivo and affects their assembly in vitro. **Cancer Res.** 54, 3676-3681.

36. Rubinfeld, B. and Polakis, P. 1995 Purification of baculovirus produced rap1GAP. **Methods Enz.** 255,31
37. Polakis, P. 1995 Mutations in the APC gene and their implications for protein structure and function. **Current Opinions in Genetics and Development** 5, 66-71
38. Rubinfeld, B., Souza, B., Albert, I., Munemitsu, S. and Polakis P. 1995 The APC protein and E-cadherin form similar but independent complexes with α -catenin, β -catenin and Plakoglobin. **J. Biol. Chem.** 270, 5549-5555
39. Munemitsu, S., Albert, I., Souza, B., Rubinfeld, B., and Polakis, P. 1995 Regulation of intracellular β -catenin levels by the APC tumor suppressor gene. **Proc. Natl. Acad. Sci.** 92, 3046-3050.
40. Lock, P., Fumagalli, S., Polakis, P. McCormick, F. and Courtneidge, S. A. 1996 The human p62 cDNA encodes Sam68 and not the rasGAP-associated p62 protein. **Cell** 84, 23-24.
41. Papkoff, J., Rubinfeld, B., Schryver, B. and Polakis, P. 1996 Wnt-1 regulates free pools of catenins and stabilizes APC-catenin complexes. **Mol. Cell. Biol.** 16, 2128-2134.
42. Rubinfeld, B., Albert, I., Porfiri, E., Fiol, C., Munemitsu, S. and Polakis, P. 1996 Binding of GSK3 β to the APC- β -catenin complex and regulation of complex assembly. **Science** 272, 1023-1026.
43. Munemitsu, S., Albert, I., Rubinfeld, B. and Polakis, P. 1996 Deletion of amino-terminal structure stabilizes β -catenin in vivo and promotes the hyperphosphorylation of the APC tumor suppressor protein. **Mol. Cell. Biol.** 16, 4088-4094.
44. Hart, M. J., Callow, M. G., Sousa, B. and Polakis P. 1996 IQGAP1, a calmodulin binding protein with a rasGAP related domain, is a potential effector for cdc42Hs. **EMBO J.** 15, 2997-3005.
45. Nathke, I. S., Adams, C. L., Polakis, P., Sellin, J. and Nelson, W. J. 1996 The adenomatous polyposis coli (APC) tumor suppressor protein is localized to plasma membrane sites involved in active epithelial cell migration. **J. Cell. Biol.** 134, 165-180.
46. Hart, M. J., Sharma, S., elMasry, N., Qui, R-G., McCabe, P., Polakis, P. and Bollag, G. 1996 Identification of a novel guanine nucleotide exchange factor for the rho GTPase. **J. Biol. Chem.** 271, 25452.
47. Thomas JE, Smith M, Rubinfeld B, Gutowski M, Beckmann RP, and Polakis P. 1996 Subcellular localization and analysis of apparent 180-kDa and 220-kDa proteins of the breast cancer susceptibility gene, BRCA1. **J. Biol. Chem.** 1996 271, 28630-28635
48. Hayashi, S., Rubinfeld, B., Souza, B., Polakis, P., Wieschaus, E., and Levine, A. 1997 A Drosophila homolog of the tumor suppressor adenomatous polyposis coli

down-regulates β -catenin but its zygotically expression is not essential for the regulation of armadillo. *Proc. Natl. Acad. Sci.* 94, 242-247.

49. Vleminckx, K., Rubinfeld, B., Polakis, P. and Gumbiner, B. 1997 The APC tumor suppressor protein induces a new axis in *Xenopus* embryos. *J. Cell. Biol.* 136, 411-420.
50. Rubinfeld, B., Robbins, P., El-Gamil, M., Albert, I., Porfiri, P. and Polakis, P. 1997 Stabilization of β -catenin by genetic defects in melanoma cell lines. *Science* 275, 1790-1792.
51. Polakis, P. The adenomatous polyposis coli (APC) tumor suppressor. 1997 *Biochem. Biophys. Acta*, 1332, F127-F147.
52. Rubinfeld, B., Albert, I., Porfiri, E., Munemitsu, S., and Polakis, P. 1997 Loss of β -catenin regulation by the APC tumor suppressor protein correlates with loss of structure due to common somatic mutations of the gene. *Cancer Res.* 57, 4624-4630.
53. Porfiri, E., Rubinfeld, B., Albert, I., Hovanes, K., Waterman, M., and Polakis, P. 1997 Induction of a β -catenin-LEF-1 complex by wnt-1 and transforming mutants of β -catenin. *Oncogene* 15, 2833-2839.
54. Thomas JE, Smith M, Tonkinson JL, Rubinfeld B, and Polakis P., 1997 Induction of phosphorylation on BRCA1 during the cell cycle and after DNA damage. *Cell Growth Differ.* 8, 801-809.
55. Hart, M., de los Santos, R., Albert, I., Rubinfeld, B., and Polakis P., 1998 Down regulation of β -catenin by human Axin and its association with the adenomatous polyposis coli (APC) tumor suppressor, β -catenin and glycogen synthase kinase 3 β . *Current Biology* 8, 573-581.
56. Polakis, P. 1998 The oncogenic activation of β -catenin. *Current Opinions in Genetics and Development* 9, 15-21
57. Matt Hart, Jean-Paul Concordet, Irina Lassot, Iris Albert, Rico del los Santos, Herve Durand, Christine Perret, Bonnee Rubinfeld, Florence Margottin, Richard Benarous and Paul Polakis. 1999 The F-box protein β -TrCP associates with phosphorylated β -catenin and regulates its activity in the cell. *Current Biology* 9, 207-10.
58. Howard C. Crawford, Barbara M. Fingleton, Bonnee Rubinfeld, Paul Polakis and Lynn M. Matrisian 1999 The metalloproteinase matrilysin is a target of β -catenin transactivation in intestinal tumours. *Oncogene* 18, 2883-91.
59. Meng J, Glick JL, Polakis P, Casey PJ. 1999 Functional interaction between Galpha(z) and Rap1GAP suggests a novel form of cellular cross-talk. *J Biol Chem.* 17, 36663-9

60. Vijayasurian Easwaran, Virginia Song, **Paul Polakis** and Steve Byers 1999 The ubiquitin-proteasome pathway and serine kinase activity modulate APC mediated regulation of β -catenin-LEF signaling. **J. Biol. Chem.** 274(23):16641-5.
61. **Polakis P**, Hart M and Rubinfeld B. 1999 Defects in the regulation of beta-catenin in colorectal cancer. **Adv Exp Med Biol.** 470, 23-32
62. Shen Z, Batzer A, Koehler JA, **Polakis P**, Schlessinger J, Lydon NB, Moran MF. 1999 Evidence for SH3 domain directed binding and phosphorylation of Sam68 by Src. **Oncogene.** 18, 4647-53
64. Thomas GM, Frame S, Goedert M, Nathke I, **Polakis P**, Cohen P. 1999 A GSK3- binding peptide from FRAT1 selectively inhibits the GSK3-catalysed phosphorylation of axin and beta-catenin. **FEBS Lett.** 458, 247-51.
65. Peifer M, **Polakis P**. 2000 Wnt signaling in oncogenesis and embryogenesis—a look outside the nucleus. **Science** 287,1606-9.
66. **Polakis P**. 2000 Wnt signaling and cancer. **Genes Dev**;14, 1837-1851.
67. Spink KE, **Polakis P**, Weis WI 2000 Structural basis of the Axin-adenomatous polyposis coli interaction. **EMBO J** 19, 2270-2279.
68. Szeto, W., Jiang, W., Tice, D.A., Rubinfeld, B., Hollingshead, P.G., Fong, S.E., Dugger, D.L., Pham, T., Yansura, D.E., Wong, T.A., Grimaldi, J.C., Corpuz, R.T., Singh J.S., Frantz, G.D., Devaux, B., Crowley, C.W., Schwall, R.H., Eberhard, D.A., Rastelli, L., **Polakis, P.** and Pennica, D. 2001 Overexpression of the Retinoic Acid-Responsive Gene Stra6 in Human Cancers and its Synergistic Induction by Wnt-1 and Retinoic Acid. **Cancer Res** 61, 4197-4204.
69. Rubinfeld B, Tice DA, **Polakis P**. 2001 Axin dependent phosphorylation of the adenomatous polyposis coli protein mediated by casein kinase 1 epsilon. **J Biol Chem** 276, 39037-39045.
70. **Polakis P**. 2001 More than one way to skin a catenin. **Cell** 2001 105, 563-566.
71. Tice DA, Soloviev I, **Polakis P**. 2002 Activation of the Wnt Pathway Interferes with Serum Response Element-driven Transcription of Immediate Early Genes. **J Biol. Chem.** 277, 6118-6123.
72. Tice DA, Szeto W, Soloviev I, Rubinfeld B, Fong SE, Dugger DL, Winer J,

Williams PM, Wieand D, Smith V, Schwall RH, Pennica D, Polakis P. 2002 Synergistic activation of tumor antigens by wnt-1 signaling and retinoic acid revealed by gene expression profiling. **J Biol Chem.** 277,14329-14335.

3. Polakis, P. 2002 Casein kinase I: A wnt'er of disconnect. **Curr. Biol.** 12, R499.

74. Mao, W., Luis, E., Ross, S., Silva, J., Tan, C., Crowley, C., Chui, C., Franz, G., Senter, P., Koeppen, H., Polakis, P. 2004 EphB2 as a therapeutic antibody drug target for the treatment of colorectal cancer. **Cancer Res.** 64, 781-788.

75. Shibamoto, S., Winer, J., Williams, M., Polakis, P. 2003 A Blockade in Wnt signaling is activated following the differentiation of F9 teratocarcinoma cells. **Exp. Cell Res.** 29211-20.

76. Zhang Y, Eberhard DA, Frantz GD, Dowd P, Wu TD, Zhou Y, Watanabe C, Luoh SM, Polakis P, Hillan KJ, Wood WI, Zhang Z. 2004 GEPIS—quantitative gene expression profiling in normal and cancer tissues. **Bioinformatics**, April 8

SECOND DECLARATION OF PAUL POLAKIS, Ph.D.

I, Paul Polakis, Ph.D., declare and say as follows:

1. I am currently employed by Genentech, Inc. where my job title is Staff Scientist.
2. Since joining Genentech in 1999, one of my primary responsibilities has been leading Genentech's Tumor Antigen Project, which is a large research project with a primary focus on identifying tumor cell markers that find use as targets for both the diagnosis and treatment of cancer in humans.
3. As I stated in my previous Declaration dated May 7, 2004 (attached as Exhibit A), my laboratory has been employing a variety of techniques, including microarray analysis, to identify genes which are differentially expressed in human tumor tissue relative to normal human tissue. The primary purpose of this research is to identify proteins that are abundantly expressed on certain human tumor tissue(s) and that are either (i) not expressed, or (ii) expressed at detectably lower levels, on normal tissue(s).
4. In the course of our research using microarray analysis, we have identified approximately 200 gene transcripts that are present in human tumor tissue at significantly higher levels than in normal human tissue. To date, we have successfully generated antibodies that bind to 31 of the tumor antigen proteins expressed from these differentially expressed gene transcripts and have used these antibodies to quantitatively determine the level of production of these tumor antigen proteins in both human tumor tissue and normal tissue. We have then quantitatively compared the levels of mRNA and protein in both the tumor and normal tissues analyzed. The results of these analyses are attached herewith as Exhibit B. In Exhibit B, "+" means that the mRNA or protein was detectably overexpressed in the tumor tissue relative to normal tissue and "-" means that no detectable overexpression was observed in the tumor tissue relative to normal tissue.
5. As shown in Exhibit B, of the 31 genes identified as being detectably overexpressed in human tumor tissue as compared to normal human tissue at the mRNA level, 28 of them (i.e., greater than 90%) are also detectably overexpressed in human tumor tissue as compared to normal human tissue at the protein level. As such, in the cases where we have been able to quantitatively measure both (i) mRNA and (ii) protein levels in both (i) tumor tissue and (ii) normal tissue, we have observed that in the vast majority of cases, there is a very strong correlation between increases in mRNA expression and increases in the level of protein encoded by that mRNA.

6. Based upon my own experience accumulated in more than 20 years of research, including the data discussed in paragraphs 4-5 above and my knowledge of the relevant scientific literature, it is my considered scientific opinion that for human genes, an increased level of mRNA in a tumor tissue relative to a normal tissue more often than not correlates to a similar increase in abundance of the encoded protein in the tumor tissue relative to the normal tissue. In fact, it remains a generally accepted working assumption in molecular biology that increased mRNA levels are more often than not predictive of elevated levels of the encoded protein. In fact, an entire industry focusing on the research and development of therapeutic antibodies to treat a variety of human diseases, such as cancer, operates on this working assumption.
7. I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information or belief are believed to be true, and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful statements may jeopardize the validity of the application or any patent issued thereon.

Dated: 3-29-00

By: Paul Polakis

Paul Polakis, Ph.D.

EXHIBIT A

DECLARATION OF PAUL POLAKIS, Ph.D.

I, Paul Polakis, Ph.D., declare and say as follows:

1. I was awarded a Ph.D. by the Department of Biochemistry of the Michigan State University in 1984. My scientific Curriculum Vitae is attached to and forms part of this Declaration (Exhibit A).
2. I am currently employed by Genentech, Inc. where my job title is Staff Scientist. Since joining Genentech in 1999, one of my primary responsibilities has been leading Genentech's Tumor Antigen Project, which is a large research project with a primary focus on identifying tumor cell markers that find use as targets for both the diagnosis and treatment of cancer in humans.
3. As part of the Tumor Antigen Project, my laboratory has been analyzing differential expression of various genes in tumor cells relative to normal cells. The purpose of this research is to identify proteins that are abundantly expressed on certain tumor cells and that are either (i) not expressed, or (ii) expressed at lower levels, on corresponding normal cells. We call such differentially expressed proteins "tumor antigen proteins". When such a tumor antigen protein is identified, one can produce an antibody that recognizes and binds to that protein. Such an antibody finds use in the diagnosis of human cancer and may ultimately serve as an effective therapeutic in the treatment of human cancer.
4. In the course of the research conducted by Genentech's Tumor Antigen Project, we have employed a variety of scientific techniques for detecting and studying differential gene expression in human tumor cells relative to normal cells, at genomic DNA, mRNA and protein levels. An important example of one such technique is the well known and widely used technique of microarray analysis which has proven to be extremely useful for the identification of mRNA molecules that are differentially expressed in one tissue or cell type relative to another. In the course of our research using microarray analysis, we have identified approximately 200 gene transcripts that are present in human tumor cells at significantly higher levels than in corresponding normal human cells. To date, we have generated antibodies that bind to about 30 of the tumor antigen proteins expressed from these differentially expressed gene transcripts and have used these antibodies to quantitatively determine the level of production of these tumor antigen proteins in both human cancer cells and corresponding normal cells. We have then compared the levels of mRNA and protein in both the tumor and normal cells analyzed.
5. From the mRNA and protein expression analyses described in paragraph 4 above, we have observed that there is a strong correlation between changes in the level of mRNA present in any particular cell type and the level of protein

expressed from that mRNA in that cell type. In approximately 80% of our observations we have found that increases in the level of a particular mRNA correlates with changes in the level of protein expressed from that mRNA when human tumor cells are compared with their corresponding normal cells.

6. Based upon my own experience accumulated in more than 20 years of research, including the data discussed in paragraphs 4 and 5 above and my knowledge of the relevant scientific literature, it is my considered scientific opinion that for human genes, an increased level of mRNA in a tumor cell relative to a normal cell typically correlates to a similar increase in abundance of the encoded protein in the tumor cell relative to the normal cell. In fact, it remains a central dogma in molecular biology that increased mRNA levels are predictive of corresponding increased levels of the encoded protein. While there have been published reports of genes for which such a correlation does not exist, it is my opinion that such reports are exceptions to the commonly understood general rule that increased mRNA levels are predictive of corresponding increased levels of the encoded protein.

7. I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information or belief are believed to be true, and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful statements may jeopardize the validity of the application or any patent issued thereon.

Dated: 5/07/04

By: Paul Polakis

Paul Polakis, Ph.D.

CURRICULUM VITAE

PAUL G. POLAKIS
Staff Scientist
Genentech, Inc
1 DNA Way, MS#40
S. San Francisco, CA 94080

EDUCATION:

Ph.D., Biochemistry, Department of Biochemistry,
Michigan State University (1984)

B.S., Biology. College of Natural Science, Michigan State University (1977)

PROFESSIONAL EXPERIENCE:

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|--------------|---|
| 2002-present | Staff Scientist, Genentech, Inc S. San Francisco, CA |
| 1999- 2002 | Senior Scientist, Genentech, Inc., S. San Francisco, CA |
| 1997 -1999 | Research Director Onyx Pharmaceuticals, Richmond, CA |
| 1992- 1996 | Senior Scientist, Project Leader, Onyx Pharmaceuticals, Richmond, CA |
| 1991-1992 | Senior Scientist, Chiron Corporation, Emeryville, CA. |
| 1989-1991 | Scientist, Cetus Corporation, Emeryville CA. |
| 1987-1989 | Postdoctoral Research Associate, Genentech, Inc., South San Francisco, CA. |
| 1985-1987 | Postdoctoral Research Associate, Department of Medicine, Duke University Medical Center, Durham, NC |

1984-1985

Assistant Professor, Department of Chemistry,
Oberlin College, Oberlin, Ohio

1980-1984

Graduate Research Assistant, Department of
Biochemistry, Michigan State University
East Lansing, Michigan

PUBLICATIONS:

1. Polakis, P. G. and Wilson, J. E. 1982 Purification of a Highly Bindable Rat Brain Hexokinase by High Performance Liquid Chromatography. *Biochem. Biophys. Res. Commun.* 107, 937-943.
2. Polakis, P.G. and Wilson, J. E. 1984 Proteolytic Dissection of Rat Brain Hexokinase: Determination of the Cleavage Pattern during Limited Digestion with Trypsin. *Arch. Biochem. Biophys.* 234, 341-352.
3. Polakis, P. G. and Wilson, J. E. 1985 An Intact Hydrophobic N-Terminal Sequence is Required for the Binding Rat Brain Hexokinase to Mitochondria. *Arch. Biochem. Biophys.* 236, 328-337.
4. Uhing, R.J., Polakis, P.G. and Snyderman, R. 1987 Isolation of GTP-binding Proteins from Myeloid HL60 Cells. *J. Biol. Chem.* 262, 15575-15579.
5. Polakis, P.G., Uhing, R.J. and Snyderman, R. 1988 The Formylpeptide Chemoattractant Receptor Copurifies with a GTP-binding Protein Containing a Distinct 40 kDa Pertussis Toxin Substrate. *J. Biol. Chem.* 263, 4969-4979.
6. Uhing, R. J., Dillon, S., Polakis, P. G., Truett, A. P. and Snyderman, R. 1988 Chemoattractant Receptors and Signal Transduction Processes in Cellular and Molecular Aspects of Inflammation (Poste, G. and Crooke, S. T. eds.) pp 335-379.
7. Polakis, P.G., Evans, T. and Snyderman 1989 Multiple Chromatographic Forms of the Formylpeptide Chemoattractant Receptor and their Relationship to GTP-binding Proteins. *Biochem. Biophys. Res. Commun.* 161, 276-283.
8. Polakis, P. G., Snyderman, R. and Evans, T. 1989 Characterization of G25K, a GTP-binding Protein Containing a Novel Putative Nucleotide Binding Domain. *Biochem. Biophys. Res. Commun.* 160, 25-32.
9. Polakis, P., Weber, R.F., Nevins, B., Didsbury, J. Evans, T. and Snyderman, R. 1989 Identification of the *ral* and *rac1* Gene Products, Low Molecular Mass GTP-binding Proteins from Human Platelets. *J. Biol. Chem.* 264, 16383-16389.
10. Snyderman, R., Perianin, A., Evans, T., Polakis, P. and Didsbury, J. 1989 G Proteins and Neutrophil Function. In *ADP-Ribosylating Toxins and G Proteins: Insights into Signal Transduction.* (J. Moss and M. Vaughn, eds.) Amer. Soc. Microbiol. pp. 295-323.

11. Hart, M.J., Polakis, P.G., Evans, T. and Cerrione, R.A. 1990 The Identification and Characterization of an Epidermal Growth Factor-Stimulated Phosphorylation of a Specific Low Molecular Mass GTP-binding Protein in a Reconstituted Phospholipid Vesicle System. *J. Biol. Chem.* 265, 5990-6001.
12. Yatani, A., Okabe, K., Polakis, P., Halenbeck, R., McCormick, F. and Brown, A. M. 1990 ras p21 and GAP Inhibit Coupling of Muscarinic Receptors to Atrial K⁺ Channels. *Cell*. 61, 769-776.
13. Munemitsu, S., Innis, M.A., Clark, R., McCormick, F., Ullrich, A. and Polakis, P.G. 1990 Molecular Cloning and Expression of a G25K cDNA, the Human Homolog of the Yeast Cell Cycle Gene CDC42. *Mol. Cell. Biol.* 10, 5977-5982.
14. Polakis, P.G., Rubinfeld, B., Evans, T. and McCormick, F. 1991 Purification of Plasma Membrane-Associated GTPase Activating Protein (GAP) Specific for rap-1/krev-1 from HL60 Cells. *Proc. Natl. Acad. Sci. USA* 88, 239-243.
15. Moran, M. F., Polakis, P., McCormick, F., Pawson, T. and Ellis, C. 1991 Protein Tyrosine Kinases Regulate the Phosphorylation, Protein Interactions, Subcellular Distribution, and Activity of p21ras GTPase Activating Protein. *Mol. Cell. Biol.* 11, 1804-1812.
16. Rubinfeld, B., Wong, G., Bekesi, E. Wood, A. McCormick, F. and Polakis, P. G. 1991 A Synthetic Peptide Corresponding to a Sequence in the GTPase Activating Protein Inhibits p21^{ras} Stimulation and Promotes Guanine Nucleotide Exchange. *Internatl. J. Peptide and Prot. Res.* 38, 47-53.
17. Rubinfeld, B., Munemitsu, S., Clark, R., Conroy, L., Watt, K., Crosier, W., McCormick, F., and Polakis, P. 1991 Molecular Cloning of a GTPase Activating Protein Specific for the Krev-1 Protein p21^{rap1}. *Cell* 65, 1033-1042.
18. Zhang, K., Papageorge, A., G., Martin, P., Vass, W. C., Olah, Z., Polakis, P., McCormick, F. and Lowy, D. R. 1991 Heterogenous Amino Acids in RAS and Rap1A Specifying Sensitivity to GAP Proteins. *Science* 254, 1630-1634.
19. Martin, G., Yatani, A., Clark, R., Polakis, P., Brown, A. M. and McCormick, F. 1992 GAP Domains Responsible for p21^{ras}-dependent Inhibition of Muscarinic Atrial K⁺ Channel Currents. *Science* 255, 192-194.
20. McCormick, F., Martin, G. A., Clark, R., Bollag, G. and Polakis, P. 1992 Regulation of p21ras by GTPase Activating Proteins. *Cold Spring Harbor Symposia on Quantitative Biology*. Vol. 56, 237-241.
21. Pronk, G. B., Polakis, P., Wong, G., deVries-Smits, A. M., Bos J. L. and McCormick, F. 1992 p60^{v-src} Can Associate with and Phosphorylate the p21^{ras} GTPase Activating Protein. *Oncogene* 7,389-394.
22. Polakis P. and McCormick, F. 1992 Interactions Between p21^{ras} Proteins and Their GTPase Activating Proteins. In Cancer Surveys (Franks, L. M., ed.) 12, 25-42.

23. Wong, G., Muller, O., Clark, R., Conroy, L., Moran, M., Polakis, P. and McCormick, F. 1992 Molecular cloning and nucleic acid binding properties of the GAP-associated tyrosine phosphoprotein p62. *Cell* 69, 551-558.
24. Polakis, P., Rubinfeld, B. and McCormick, F. 1992 Phosphorylation of rap1GAP in vivo and by cAMP-dependent Kinase and the Cell Cycle p34^{cdc2} Kinase in vitro. *J. Biol. Chem.* 267, 10780-10785.
25. McCabe, P.C., Haubrauck, H., Polakis, P., McCormick, F., and Innis, M. A. 1992 Functional Interactions Between p21^{rap1A} and Components of the Budding pathway of *Saccharomyces cerevisiae*. *Mol. Cell. Biol.* 12, 4084-4092.
26. Rubinfeld, B., Crosier, W.J., Albert, I., Conroy, L., Clark, R., McCormick, F. and Polakis, P. 1992 Localization of the rap1GAP Catalytic Domain and Sites of Phosphorylation by Mutational Analysis. *Mol. Cell. Biol.* 12, 4634-4642.
27. Ando, S., Kaibuchi, K., Sasaki, K., Hiraoka, T., Nishiyama, T., Mizuno, T., Asada, M., Nuno, H., Matsuda, I., Matsuura, Y., Polakis, P., McCormick, F. and Takai, Y. 1992 Post-translational processing of rac p21s is important both for their interaction with the GDP/GTP exchange proteins and for their activation of NADPH oxidase. *J. Biol. Chem.* 267, 25709-25713.
28. Janoueix-Lerosey, I., Polakis, P., Tavitian, A. and deGunzburg, J. 1992 Regulation of the GTPase activity of the ras-related rap2 protein. *Biochem. Biophys. Res. Commun.* 189, 455-464.
29. Polakis, P. 1993 GAPs Specific for the rap1/Krev-1 Protein. in GTP-binding Proteins: the ras-superfamily. (J.C. LaCale and F. McCormick, eds.) 445-452.
30. Polakis, P. and McCormick, F. 1993 Structural requirements for the interaction of p21^{ras} with GAP, exchange factors, and its biological effector target. *J. Biol. Chem.* 268, 9157-9160.
31. Rubinfeld, B., Souza, B., Albert, I., Muller, O., Chamberlain, S., Masiarz, F., Munemitsu, S. and Polakis, P. 1993 Association of the APC gene product with beta-catenin. *Science* 262, 1731-1734.
32. Weiss, J., Rubinfeld, B., Polakis, P., McCormick, F., Cavenee, W. A. and Arden, K. 1993 The gene for human rap1-GTPase activating protein (rap1GAP) maps to chromosome 1p35-1p36.1. *Cytogenet. Cell Genet.* 66, 18-21.

33. Sato, K. Y., Polakis, P., Haubruck, H., Fasching, C. L., McCormick, F. and Stanbridge, E. J. 1994 Analysis of the tumor suppressor activity of the K-rev gene in human tumor cell lines. *Cancer Res.* 54, 552-559.
34. Janoueix-Lerosey, I., Fontenay, M., Tobelem, G., Tavitian, A., Polakis, P. and DeGunzburg, J. 1994 Phosphorylation of rap1GAP during the cell cycle. *Biochem. Biophys. Res. Commun.* 202, 967-975.
35. Munemitsu, S., Souza, B., Mueller, O., Albert, I., Rubinfeld, B., and Polakis, P. 1994 The APC gene product associates with microtubules in vivo and affects their assembly in vitro. *Cancer Res.* 54, 3676-3681.

36. Rubinfeld, B. and Polakis, P. 1995 Purification of baculovirus produced rap1GAP. *Methods Enz.* 255,31
37. Polakis, P. 1995 Mutations in the APC gene and their implications for protein structure and function: *Current Opinions in Genetics and Development* 5, 66-71
38. Rubinfeld, B., Souza, B., Albert, I., Munemitsu, S. and Polakis P. 1995 The APC protein and E-cadherin form similar but independent complexes with α -catenin, β -catenin and Plakoglobin. *J. Biol. Chem.* 270, 5549-5555
39. Munemitsu, S., Albert, I., Souza, B., Rubinfeld, B., and Polakis, P. 1995 Regulation of intracellular β -catenin levels by the APC tumor suppressor gene. *Proc. Natl. Acad. Sci.* 92, 3046-3050.
40. Lock, P., Fumagalli, S., Polakis, P. McCormick, F. and Courtneidge, S. A. 1996 The human p62 cDNA encodes Sam68 and not the rasGAP-associated p62 protein. *Cell* 84, 23-24.
41. Papkoff, J., Rubinfeld, B., Schryver, B. and Polakis, P. 1996 Wnt-1 regulates free pools of catenins and stabilizes APC-catenin complexes. *Mol. Cell. Biol.* 16, 2128-2134.
42. Rubinfeld, B., Albert, I., Porfiri, E., Fiol, C., Munemitsu, S. and Polakis, P. 1996 Binding of GSK3 β to the APC- β -catenin complex and regulation of complex assembly. *Science* 272, 1023-1026.
43. Munemitsu, S., Albert, I., Rubinfeld, B. and Polakis, P. 1996 Deletion of amino-terminal structure stabilizes β -catenin in vivo and promotes the hyperphosphorylation of the APC tumor suppressor protein. *Mol. Cell. Biol.* 16, 4088-4094.
44. Hart, M. J., Callow, M. G., Sousa, B. and Polakis P. 1996 IQGAP1, a calmodulin binding protein with a rasGAP related domain, is a potential effector for cdc42Hs. *EMBO J.* 15, 2997-3005.
45. Nathke, I. S., Adams, C. L., Polakis, P., Sellin, J. and Nelson, W. J. 1996 The adenomatous polyposis coli (APC) tumor suppressor protein is localized to plasma membrane sites involved in active epithelial cell migration. *J. Cell. Biol.* 134, 165-180.
-
46. Hart, M. J., Sharma, S., elMasry, N., Qui, R-G., McCabe, P., Polakis, P. and Bollag, G. 1996 Identification of a novel guanine nucleotide exchange factor for the rho GTPase. *J. Biol. Chem.* 271, 25452.
47. Thomas JE, Smith M, Rubinfeld B, Gutowski M, Beckmann RP, and Polakis P. 1996 Subcellular localization and analysis of apparent 180-kDa and 220-kDa proteins of the breast cancer susceptibility gene, BRCA1. *J. Biol. Chem.* 1996 271, 28630-28635
48. Hayashi, S., Rubinfeld, B., Souza, B., Polakis, P., Wieschaus, E., and Levine, A. 1997 A Drosophila homolog of the tumor suppressor adenomatous polyposis coli

down-regulates β -catenin but its zygotic expression is not essential for the regulation of armadillo. *Proc. Natl. Acad. Sci.* 94, 242-247.

49. Vleminckx, K., Rubinfeld, B., Polakis, P. and Gumbiner, B. 1997 The APC tumor suppressor protein induces a new axis in *Xenopus* embryos. *J. Cell. Biol.* 136, 411-420.

50. Rubinfeld, B., Robbins, P., El-Gamil, M., Albert, I., Porfiri, P. and Polakis, P. 1997 Stabilization of β -catenin by genetic defects in melanoma cell lines. *Science* 275, 1790-1792.

51. Polakis, P. The adenomatous polyposis coli (APC) tumor suppressor. 1997 *Biochem. Biophys. Acta*, 1332, F127-F147.

52. Rubinfeld, B., Albert, I., Porfiri, E., Munemitsu, S., and Polakis, P. 1997 Loss of β -catenin regulation by the APC tumor suppressor protein correlates with loss of structure due to common somatic mutations of the gene. *Cancer Res.* 57, 4624-4630.

53. Porfiri, E., Rubinfeld, B., Albert, I., Hovanes, K., Waterman, M., and Polakis, P. 1997 Induction of a β -catenin-LEF-1 complex by wnt-1 and transforming mutants of β -catenin. *Oncogene* 15, 2833-2839.

54. Thomas JE, Smith M, Tonkinson JL, Rubinfeld B, and Polakis P., 1997 Induction of phosphorylation on BRCA1 during the cell cycle and after DNA damage. *Cell Growth Differ.* 8, 801-809.

55. Hart, M., de los Santos, R., Albert, I., Rubinfeld, B., and Polakis P., 1998 Down regulation of β -catenin by human Axin and its association with the adenomatous polyposis coli (APC) tumor suppressor, β -catenin and glycogen synthase kinase 3 β . *Current Biology* 8, 573-581.

56. Polakis, P. 1998 The oncogenic activation of β -catenin. *Current Opinions in Genetics and Development* 9, 15-21

57. Matt Hart, Jean-Paul Concordet, Irina Lassot, Iris Albert, Rico del los Santos, Herve Durand, Christine Perret, Bonnee Rubinfeld, Florence Margottin, Richard Benarous and Paul Polakis. 1999 The F-box protein β -TrCP associates with phosphorylated β -catenin and regulates its activity in the cell. *Current Biology* 9, 207-10.

58. Howard C. Crawford, Barbara M. Fingleton, Bonnee Rubinfeld, Paul Polakis and Lynn M. Matrisian 1999 The metalloproteinase matrilysin is a target of β -catenin transactivation in intestinal tumours. *Oncogene* 18, 2883-91.

59. Meng J, Glick JL, Polakis P, Casey PJ. 1999 Functional interaction between Galpha(z) and Rap1GAP suggests a novel form of cellular cross-talk. *J Biol Chem.* 17, 36663-9

60. Vijayasurian Easwaran, Virginia Song, Paul Polakis and Steve Byers 1999 The ubiquitin-proteosome pathway and serine kinase activity modulate APC mediated regulation of β -catenin-LEF signaling. *J. Biol. Chem.* 274(23):16641-5.
 61. Polakis P, Hart M and Rubinfeld B. 1999 Defects in the regulation of beta-catenin in colorectal cancer. *Adv Exp Med Biol.* 470, 23-32
 62. Shen Z, Batzer A, Koehler JA, Polakis P, Schlessinger J, Lydon NB, Moran MF. 1999 Evidence for SH3 domain directed binding and phosphorylation of Sam68 by Src. *Oncogene.* 18, 4647-53
 64. Thomas GM, Frame S, Goedert M, Nathke I, Polakis P, Cohen P. 1999 A GSK3- binding peptide from FRAT1 selectively inhibits the GSK3-catalysed phosphorylation of axin and beta-catenin. *FEBS Lett.* 458, 247-51.
 65. Peifer M, Polakis P. 2000 Wnt signaling in oncogenesis and embryogenesis—a look outside the nucleus. *Science* 287,1606-9.
 66. Polakis P. 2000 Wnt signaling and cancer. *Genes Dev*;14, 1837-1851.
 67. Spink KE, Polakis P, Weis WI 2000 Structural basis of the Axin-adenomatous polyposis coli interaction. *EMBO J* 19, 2270-2279.
 68. Szeto, W., Jiang, W., Tice, D.A., Rubinfeld, B., Hollingshead, P.G., Fong, S.E., Dugger, D.L., Pham, T., Yansura, D.E., Wong, T.A., Grimaldi, J.C., Corpuz, R.T., Singh J.S., Frantz, G.D., Devaux, B., Crowley, C.W., Schwall, R.H., Eberhard, D.A., Rastelli, L., Polakis, P. and Pennica, D. 2001 Overexpression of the Retinoic Acid-Responsive Gene Stra6 in Human Cancers and its Synergistic Induction by Wnt-1 and Retinoic Acid. *Cancer Res* 61, 4197-4204.
 69. Rubinfeld B, Tice DA, Polakis P. 2001 Axin dependent phosphorylation of the adenomatous polyposis coli protein mediated by casein kinase 1 epsilon. *J Biol Chem* 276, 39037-39045.
-
70. Polakis P. 2001 More than one way to skin a catenin. *Cell* 2001 105, 563-566.
 71. Tice DA, Soloviev I, Polakis P. 2002 Activation of the Wnt Pathway Interferes with Serum Response Element-driven Transcription of Immediate Early Genes. *J Biol. Chem.* 277, 6118-6123.
 72. Tice DA, Szeto W, Soloviev I, Rubinfeld B, Fong SE, Dugger DL, Winer J,

- Williams PM, Wieand D, Smith V, Schwall RH, Pennica D, Polakis P. 2002 Synergistic activation of tumor antigens by wnt-1 signaling and retinoic acid revealed by gene expression profiling. *J Biol Chem.* 277,14329-14335.
73. Polakis, P. 2002 Casein kinase I: A wnt'er of disconnect. *Curr. Biol.* 12, R499.
74. Mao, W., Luis, E., Ross, S., Silva, J., Tan, C., Crowley, C., Chui, C., Franz, G., Senter, P., Koeppen, H., Polakis, P. 2004 EphB2 as a therapeutic antibody drug target for the treatment of colorectal cancer. *Cancer Res.* 64, 781-788.
75. Shibamoto, S., Winer, J., Williams, M., Polakis, P. 2003 A Blockade in Wnt signaling is activated following the differentiation of F9 teratocarcinoma cells. *Exp. Cell Res.* 29211-20.
76. Zhang Y, Eberhard DA, Frantz GD, Dowd P, Wu TD, Zhou Y, Watanabe C, Luoh SM, Polakis P, Hillan KJ, Wood WI, Zhang Z. 2004 GEPIS--quantitative gene expression profiling in normal and cancer tissues. *Bioinformatics*, April 8.
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EXHIBIT B

| | tumor mRNA | tumor IHC |
|----------|------------|-----------|
| UNQ2525 | + | + |
| UNQ2378 | + | + |
| UNQ972 | + | - |
| UNQ97671 | + | + |
| UNQ2964 | + | + |
| UNQ323 | + | + |
| UNQ1655 | + | + |
| UNQ2333 | + | + |
| UNQ9638 | + | + |
| UNQ8209 | + | + |
| UNQ6507 | + | + |
| UNQ8196 | + | + |
| UNQ9109 | + | + |
| UNQ100 | + | + |
| UNQ178 | + | + |
| UNQ1477 | + | + |
| UNQ1839 | + | + |
| UNQ2079 | + | + |
| UNQ8782 | + | + |
| UNQ9646 | + | - |
| UNQ111 | + | + |
| UNQ3079 | + | + |
| UNQ8175 | + | + |
| UNQ9509 | + | + |
| UNQ10978 | + | - |
| UNQ2103 | + | + |
| UNQ1563 | + | + |
| UNQ16188 | + | + |
| UNQ13589 | + | + |
| UNQ1078 | + | + |
| UNQ879 | + | + |

Impact of DNA Amplification on Gene Expression Patterns in Breast Cancer^{1,2}Elizabeth Hyman,³ Päävikki Kauraniemi,³ Sampsa Hautaniemi, Maija Wolf, Spyro Mousses, Ester Rozenblum, Markus Ringnér, Guido Sauter, Outi Monni, Abdel Elkhouloun, Olli-P. Kallioniemi, and Anne Kallioniemi⁴

Howard Hughes Medical Institute-NIH Research Scholar, Bethesda, Maryland 20892 [E.H.]; Cancer Genetics Branch, National Human Genome Research Institute, NIH, Bethesda, Maryland 20892 [E.H., P.K., S.H., M.W., S.M., E.R., M.R., A.E., O.K., A.K.]; Laboratory of Cancer Genetics, Institute of Medical Technology, University of Tampere and Tampere University Hospital, FIN-33520 Tampere, Finland [P.K., A.K.]; Signal Processing Laboratory, Tampere University of Technology, FIN-33101 Tampere, Finland [S.H.]; Institute of Pathology, University of Basel, CH-4003 Basel, Switzerland [G.S.]; and Biomedicum Biochip Center, Helsinki University Hospital, Biomedicum Helsinki, FIN-00014 Helsinki, Finland [O.M.]

ABSTRACT

Genetic changes underlie tumor progression and may lead to cancer-specific expression of critical genes. Over 1100 publications have described the use of comparative genomic hybridization (CGH) to analyze the pattern of copy number alterations in cancer, but very few of the genes affected are known. Here, we performed high-resolution CGH analysis on cDNA microarrays in breast cancer and directly compared copy number and mRNA expression levels of 13,824 genes to quantitate the impact of genomic changes on gene expression. We identified and mapped the boundaries of 24 independent amplicons, ranging in size from 0.2 to 12 Mb. Throughout the genome, both high- and low-level copy number changes had a substantial impact on gene expression, with 44% of the highly amplified genes showing overexpression and 10.5% of the highly overexpressed genes being amplified. Statistical analysis with random permutation tests identified 270 genes whose expression levels across 14 samples were systematically attributable to gene amplification. These included most previously described amplified genes in breast cancer and many novel targets for genomic alterations, including the *HOXB7* gene, the presence of which in a novel amplicon at 17q21.3 was validated in 10.2% of primary breast cancers and associated with poor patient prognosis. In conclusion, CGH on cDNA microarrays revealed hundreds of novel genes whose overexpression is attributable to gene amplification. These genes may provide insights to the clonal evolution and progression of breast cancer and highlight promising therapeutic targets.

INTRODUCTION

Gene expression patterns revealed by cDNA microarrays have facilitated classification of cancers into biologically distinct categories, some of which may explain the clinical behavior of the tumors (1-6). Despite this progress in diagnostic classification, the molecular mechanisms underlying gene expression patterns in cancer have remained elusive, and the utility of gene expression profiling in the identification of specific therapeutic targets remains limited.

Accumulation of genetic defects is thought to underlie the clonal evolution of cancer. Identification of the genes that mediate the effects of genetic changes may be important by highlighting transcripts that are actively involved in tumor progression. Such transcripts and their encoded proteins would be ideal targets for anticancer therapies, as demonstrated by the clinical success of new therapies against amplified oncogenes, such as *ERBB2* and *EGFR* (7, 8), in breast cancer and other solid tumors. Besides amplifications of known oncogenes, over

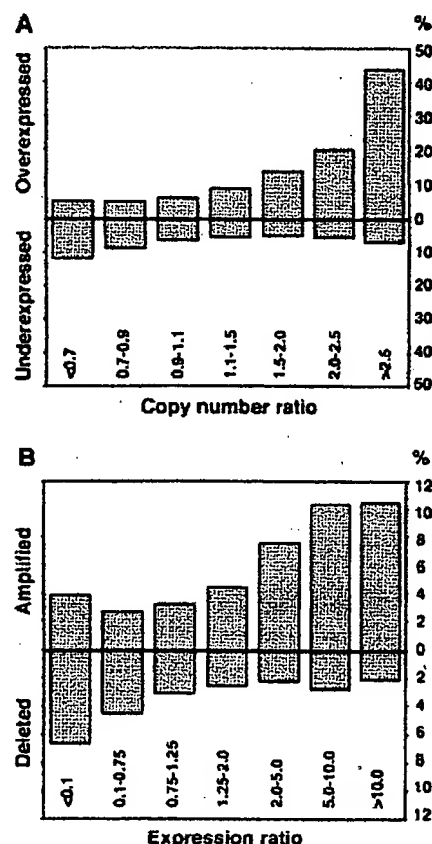


Fig. 1. Impact of gene copy number on global gene expression levels. A, percentage of over- and underexpressed genes (Y axis) according to copy number ratios (X axis). Threshold values used for over- and underexpression were >2.184 (global upper 7% of the cDNA ratios) and <0.4826 (global lower 7% of the expression ratios). B, percentage of amplified and deleted genes according to expression ratios. Threshold values for amplification and deletion were >1.5 and <0.7 .

20 recurrent regions of DNA amplification have been mapped in breast cancer by CGH⁵ (9, 10). However, these amplicons are often large and poorly defined, and their impact on gene expression remains unknown.

We hypothesized that genome-wide identification of those gene expression changes that are attributable to underlying gene copy number alterations would highlight transcripts that are actively involved in the causation or maintenance of the malignant phenotype. To identify such transcripts, we applied a combination of cDNA and CGH microarrays to: (a) determine the global impact that gene copy number variation plays in breast cancer development and progression; and (b) identify and characterize those genes whose mRNA expres-

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² Supplementary data for this article are available at Cancer Research Online (<http://cancerres.aacrjournals.org>).

³ Contributed equally to this work.

⁴ To whom requests for reprints should be addressed, at Laboratory of Cancer Genetics, Institute of Medical Technology, Lenckellijankatu 6, FIN-33520 Tampere, Finland. Phone: 358-3247-4125; Fax: 358-3247-4168; E-mail: anne.kallioniemi@uta.fi.

⁵ The abbreviations used are: CGH, comparative genomic hybridization; FISH, fluorescence *in situ* hybridization; RT-PCR, reverse transcription-PCR.

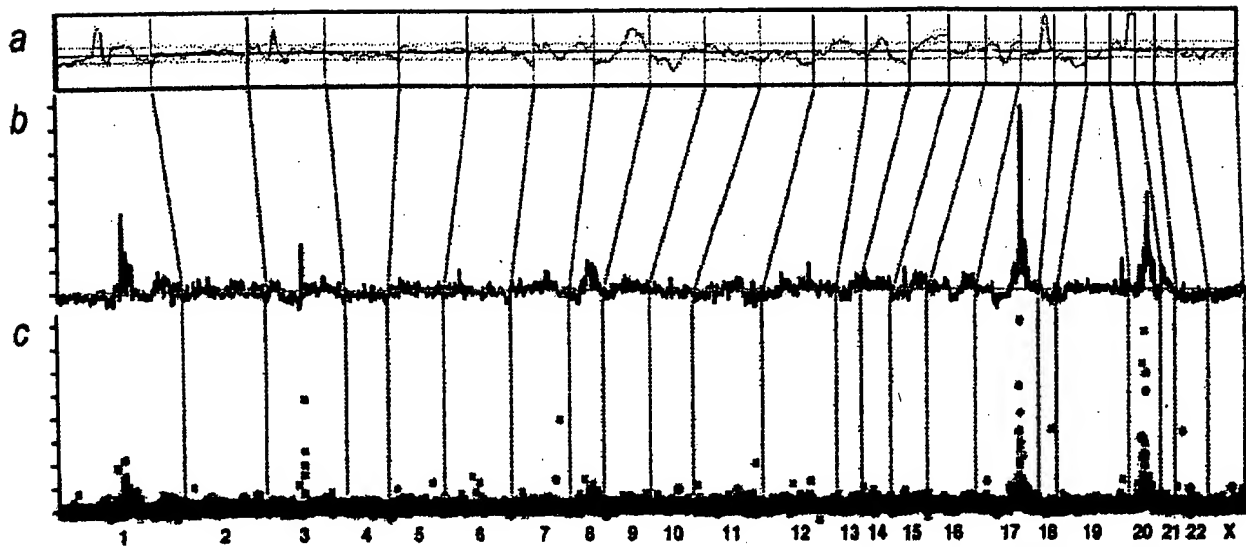


Fig. 2. Genome-wide copy number and expression analysis in the MCF-7 breast cancer cell line. *A*, chromosomal CGH analysis of MCF-7. The copy number ratio profile (blue line) across the entire genome from 1p telomere to Xq telomere is shown along with ± 1 SD (orange lines). The black horizontal line indicates a ratio of 1.0; red line, a ratio of 0.8; and green line, a ratio of 1.2. *B–C*, genome-wide copy number analysis in MCF-7 by CGH on cDNA microarray. The copy number ratios were plotted as a function of the position of the cDNA clones along the human genome. In *B*, individual data points are connected with a line, and a moving median of 10 adjacent clones is shown. Red horizontal line, the copy number ratio of 1.0. In *C*, individual data points are labeled by color coding according to cDNA expression ratios. The bright green dots indicate the upper 2%, and dark red dots, the next 5% of the expression ratios in MCF-7 cells (overexpressed genes); bright green dots indicate the lowest 2%, and dark green dots, the next 5% of the expression ratios (underexpressed genes); the rest of the observations are shown with black crosses. The chromosome numbers are shown at the bottom of the figure, and chromosome boundaries are indicated with a dashed line.

sion is most significantly associated with amplification of the corresponding genomic template.

MATERIALS AND METHODS

Breast Cancer Cell Lines. Fourteen breast cancer cell lines (BT-20, BT-474, HCC1428, Hs578t, MCF7, MDA-361, MDA-436, MDA-453, MDA-468, SKBR-3, T-47D, UACC812, ZR-75-1, and ZR-75-30) were obtained from the American Type Culture Collection (Manassas, VA). Cells were grown under recommended culture conditions. Genomic DNA and mRNA were isolated using standard protocols.

Copy Number and Expression Analyses by cDNA Microarrays. The preparation and printing of the 13,824 cDNA clones on glass slides were performed as described (11–13). Of these clones, 244 represented uncharacterized expressed sequence tags, and the remainder corresponded to known genes. CGH experiments on cDNA microarrays were done as described (14, 15). Briefly, 20 μ g of genomic DNA from breast cancer cell lines and normal human WBCs were digested for 14–18 h with *AluI* and *RsaI* (Life Technologies, Inc., Rockville, MD) and purified by phenol/chloroform extraction. Six μ g of digested cell line DNAs were labeled with Cy3-dUTP (Amersham Pharmacia) and normal DNA with Cy5-dUTP (Amersham Pharmacia) using the Bioprime Labeling kit (Life Technologies, Inc.). Hybridization (14, 15) and posthybridization washes (13) were done as described. For the expression analyses, a standard reference (Universal Human Reference RNA; Stratagene, La Jolla, CA) was used in all experiments. Forty μ g of reference RNA were labeled with Cy3-dUTP and 3.5 μ g of test mRNA with Cy5-dUTP, and the labeled cDNAs were hybridized on microarrays as described (13, 15). For both microarray analyses, a laser confocal scanner (Agilent Technologies, Palo Alto, CA) was used to measure the fluorescence intensities at the target locations using the DEARRAY software (16). After background subtraction, average intensities at each clone in the test hybridization were divided by the average intensity of the corresponding clone in the control hybridization. For the copy number analysis, the ratios were normalized on the basis of the distribution of ratios of all targets on the array and for the expression analysis on the basis of 88 housekeeping genes, which were spotted four times onto the array. Low quality measurements (*i.e.*, copy number data with mean reference intensity <100 fluorescent units, and expression data with both test and reference intensity <100 fluorescent units and/or with spot size <50 units)

were excluded from the analysis and were treated as missing values. The distributions of fluorescence ratios were used to define cutpoints for increased/decreased copy number. Genes with CGH ratio >1.43 (representing the upper 5% of the CGH ratios across all experiments) were considered to be amplified, and genes with ratio <0.73 (representing the lower 5%) were considered to be deleted.

Statistical Analysis of CGH and cDNA Microarray Data. To evaluate the influence of copy number alterations on gene expression, we applied the following statistical approach. CGH and cDNA calibrated intensity ratios were log-transformed and normalized using median centering of the values in each cell line. Furthermore, cDNA ratios for each gene across all 14 cell lines were median centered. For each gene, the CGH data were represented by a vector that was labeled 1 for amplification (ratio, >1.43) and 0 for no amplification. Amplification was correlated with gene expression using the signal-to-noise statistics (1). We calculated a weight, w_g , for each gene as follows:

$$w_g = \frac{m_{g1} - m_{g0}}{\sigma_{g1} + \sigma_{g0}}$$

where m_{g1} , σ_{g1} and m_{g0} , σ_{g0} denote the means and SDs for the expression levels for amplified and nonamplified cell lines, respectively. To assess the statistical significance of each weight, we performed 10,000 random permutations of the label vector. The probability that a gene had a larger or equal weight by random permutation than the original weight was denoted by α . A low α (<0.05) indicates a strong association between gene expression and amplification.

Genomic Localization of cDNA Clones and Amplicon Mapping. Each cDNA clone on the microarray was assigned to a Unigene cluster using the Unigene Build 141.⁶ A database of genomic sequence alignment information for mRNA sequences was created from the August 2001 freeze of the University of California Santa Cruz's GoldenPath database.⁷ The chromosome and bp positions for each cDNA clone were then retrieved by relating these data sets. Amplicons were defined as a CGH copy number ratio >2.0 in at least two adjacent clones in two or more cell lines or a CGH ratio >2.0 in at least three adjacent clones in a single cell line. The amplicon start and end positions were

⁶ Internet address: http://research.nhgri.nih.gov/microarray/downloadable_cdna.html.

⁷ Internet address: www.genome.ucsc.edu.

Table 1. Summary of independent amplicons in 14 breast cancer cell lines by CGH microarray

| Location | Start (Mb) | End (Mb) | Size (Mb) |
|-----------------|------------|----------|-----------|
| 1p13 | 132.79 | 132.94 | 0.2 |
| 1q21 | 173.92 | 177.25 | 3.3 |
| 1q22 | 179.28 | 179.57 | 0.3 |
| 3p14 | 71.94 | 74.66 | 2.7 |
| 7p12.1-7p11.2 | 55.62 | 60.95 | 5.3 |
| 7q31 | 125.73 | 130.96 | 5.2 |
| 7q32 | 140.01 | 140.68 | 0.7 |
| 8q21.11-8q21.13 | 86.45 | 92.46 | 6.0 |
| 8q21.3 | 98.45 | 103.05 | 4.6 |
| 8q23.3-8q24.14 | 129.88 | 142.15 | 12.3 |
| 8q24.22 | 151.21 | 152.16 | 1.0 |
| 9p13 | 38.63 | 39.25 | 0.6 |
| 13q22-q31 | 77.15 | 81.38 | 4.2 |
| 16q22 | 86.70 | 87.62 | 0.9 |
| 17q11 | 29.30 | 30.85 | 1.6 |
| 17q12-q21.2 | 39.79 | 42.80 | 3.0 |
| 17q21.32-q21.33 | 52.47 | 55.80 | 3.3 |
| 17q22-q23.3 | 63.81 | 69.70 | 5.9 |
| 17q23.3-q24.3 | 69.93 | 74.99 | 5.1 |
| 19q13 | 40.63 | 41.40 | 0.8 |
| 20q11.22 | 34.59 | 35.85 | 1.3 |
| 20q13.12 | 44.00 | 45.62 | 1.6 |
| 20q13.12-q13.13 | 46.45 | 49.43 | 3.0 |
| 20q13.2-q13.32 | 51.32 | 59.12 | 7.8 |

extended to include neighboring nonamplified clones (ratio, <1.5). The amplicon size determination was partially dependent on local clone density.

FISH. Dual-color interphase FISH to breast cancer cell lines was done as described (17). Bacterial artificial chromosome clone RP11-361K8 was labeled with SpectrumOrange (Vysis, Downers Grove, IL), and SpectrumOrange-labeled probe for *EGFR* was obtained from Vysis. SpectrumGreen-labeled chromosome 7 and 17 centromere probes (Vysis) were used as a reference. A tissue microarray containing 612 formalin-fixed, paraffin-embedded primary breast cancers (17) was applied in FISH analyses as described (18). The use of these specimens was approved by the Ethics Committee of the University of Basel and by the NIH. Specimens containing a 2-fold or higher increase in the number of test probe signals, as compared with corresponding centromere signals, in at least 10% of the tumor cells were considered to be amplified. Survival analysis was performed using the Kaplan-Meier method and the log-rank test.

RT-PCR. The *HOXB7* expression level was determined relative to *GAPDH*. Reverse transcription and PCR amplification were performed using Access RT-PCR System (Promega Corp., Madison, WI) with 10 ng of mRNA as a template. *HOXB7* primers were 5'-GAGCAGAGGGACTCGGACTT-3' and 5'-GCCTCAGGTAGCGATTGTAG-3'.

RESULTS

Global Effect of Copy Number on Gene Expression. 13,824 arrayed cDNA clones were applied for analysis of gene expression and gene copy number (CGH microarrays) in 14 breast cancer cell lines. The results illustrate a considerable influence of copy number on gene expression patterns. Up to 44% of the highly amplified transcripts (CGH ratio, >2.5) were overexpressed (i.e., belonged to the global upper 7% of expression ratios), compared with only 6% for genes with normal copy number levels (Fig. 1A). Conversely, 10.5% of the transcripts with high-level expression (cDNA ratio, >10) showed increased copy number (Fig. 1B). Low-level copy number increases and decreases were also associated with similar, although less dramatic, outcomes on gene expression (Fig. 1).

Identification of Distinct Breast Cancer Amplicons. Base-pair locations obtained for 11,994 cDNAs (86.8%) were used to plot copy number changes as a function of genomic position (Fig. 2, Supplement Fig. A). The average spacing of clones throughout the genome was 267 kb. This high-resolution mapping identified 24 independent breast cancer amplicons, spanning from 0.2 to 12 Mb of DNA (Table 1). Several amplification sites detected previously by chromosomal

CGH were validated, with 1q21, 17q12-q21.2, 17q22-q23, 20q13.1, and 20q13.2 regions being most commonly amplified. Furthermore, the boundaries of these amplicons were precisely delineated. In addition, novel amplicons were identified at 9p13 (38.65-39.25 Mb), and 17q21.3 (52.47-55.80 Mb).

Direct Identification of Putative Amplification Target Genes. The cDNA/CGH microarray technique enables the direct correlation of copy number and expression data on a gene-by-gene basis throughout the genome. We directly annotated high-resolution CGH plots with gene expression data using color coding. Fig. 2C shows that most of the amplified genes in the MCF-7 breast cancer cell line at 1p13, 17q22-q23, and 20q13 were highly overexpressed. A view of chromosome 7 in the MDA-468 cell line implicates *EGFR* as the most highly overexpressed and amplified gene at 7p11-p12 (Fig. 3A). In BT-474, the two known amplicons at 17q12 and 17q22-q23 contained numerous highly overexpressed genes (Fig. 3B). In addition, several genes, including the homeobox genes *HOXB2* and *HOXB7*, were highly amplified in a previously undescribed independent amplicon at 17q21.3. *HOXB7* was systematically amplified (as validated by FISH, Fig. 3B, inset) as well as overexpressed (as verified by RT-PCR, data not shown) in BT-474, UACC812, and ZR-75-30 cells. Furthermore, this novel

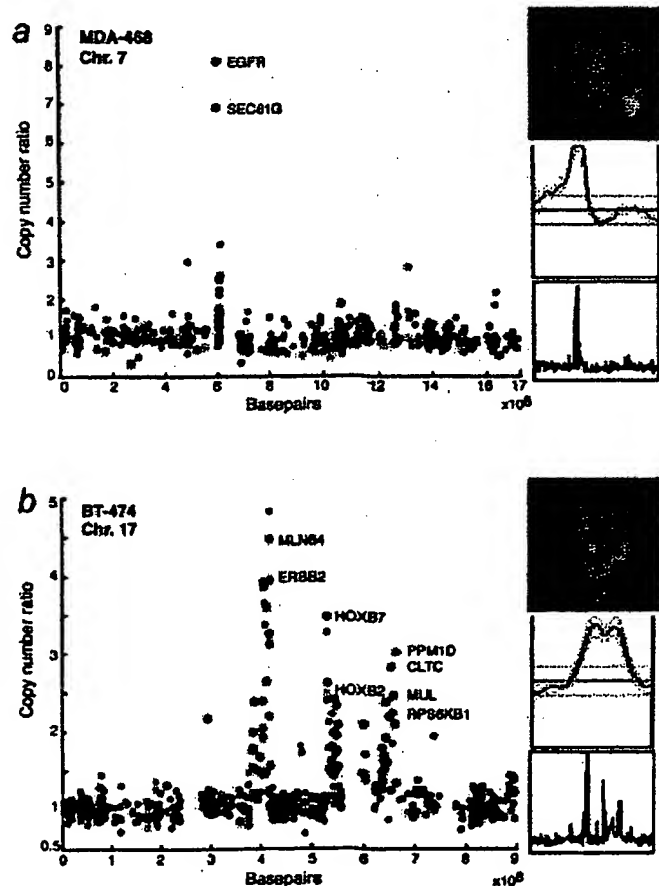
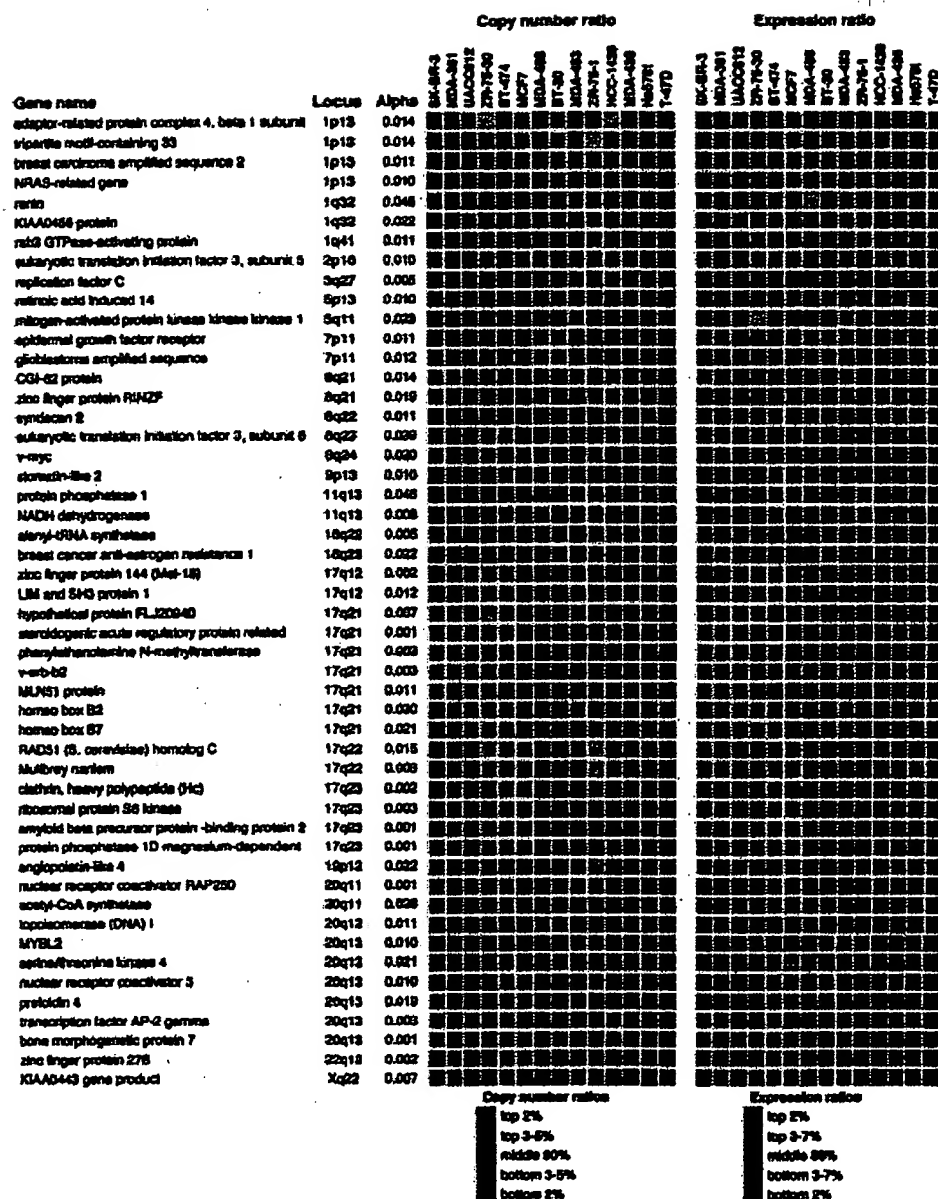


Fig. 3. Annotation of gene expression data on CGH microarray profiles. A, genes in the 7p11-p12 amplicon in the MDA-468 cell line are highly expressed (red dots) and include the *EGFR* oncogene. B, several genes in the 17q12, 17q21.3, and 17q23 amplicons in the BT-474 breast cancer cell line are highly overexpressed (red) and include the *HOXB7* gene. The data labels and color coding are as indicated for Fig. 2C. Insets show chromosomal CGH profiles for the corresponding chromosomes and validation of the increased copy number by interphase FISH using *EGFR* (red) and chromosome 7 centromere probe (green) to MDA-468 (A) and *HOXB7*-specific probe (red) and chromosome 17 centromere (green) to BT-474 cells (B).

Fig. 4. List of 50 genes with a statistically significant correlation (α value <0.05) between gene copy number and gene expression. Name, chromosomal location, and the α value for each gene are indicated. The genes have been ordered according to their position in the genome. The color maps on the right illustrate the copy number and expression ratio patterns in the 14 cell lines. The key to the color code is shown at the bottom of the graph. Gray squares, missing values. The complete list of 270 genes is shown in supplemental Fig. B.



amplification was validated to be present in 10.2% of 363 primary breast cancers by FISH to a tissue microarray and was associated with poor prognosis of the patients ($P = 0.001$).

Statistical Identification and Characterization of 270 Highly Expressed Genes in Amplicons. Statistical comparison of expression levels of all genes as a function of gene amplification identified 270 genes whose expression was significantly influenced by copy number across all 14 cell lines (Fig. 4, Supplemental Fig. B). According to the gene ontology data,⁸ 91 of the 270 genes represented hypothetical proteins or genes with no functional annotation, whereas 179 had associated functional information available. Of these, 151 (84%) are implicated in apoptosis, cell proliferation, signal transduction, and transcription, whereas 28 (16%) had functional annotations that could not be directly linked with cancer.

DISCUSSION

The importance of recurrent gene and chromosome copy number changes in the development and progression of solid tumors has been characterized in >1000 publications applying CGH⁹ (9, 10), as well as in a large number of other molecular cytogenetic, cytogenetic, and molecular genetic studies. The effects of these somatic genetic changes on gene expression levels have remained largely unknown, although a few studies have explored gene expression changes occurring in specific amplicons (15, 19–21). Here, we applied genome-wide cDNA microarrays to identify transcripts whose expression changes were attributable to underlying gene copy number alterations in breast cancer.

The overall impact of copy number on gene expression patterns was substantial with the most dramatic effects seen in the case of high-

* Internet address: <http://www.geneontology.org/>.

⁹ Internet address: <http://www.ncbi.nlm.nih.gov/entrez>.

level copy number increase. Low-level copy number gains and losses also had a significant influence on expression levels of genes in the regions affected, but these effects were more subtle on a gene-by-gene basis than those of high-level amplifications. However, the impact of low-level gains on the dysregulation of gene expression patterns in cancer may be equally important if not more important than that of high-level amplifications. Aneuploidy and low-level gains and losses of chromosomal arms represent the most common types of genetic alterations in breast and other cancers and, therefore, have an influence on many genes. Our results in breast cancer extend the recent studies on the impact of aneuploidy on global gene expression patterns in yeast cells, acute myeloid leukemia, and a prostate cancer model system (22–24).

The CGH microarray analysis identified 24 independent breast cancer amplicons. We defined the precise boundaries for many amplicons detected previously by chromosomal CGH (9, 10, 25, 26) and also discovered novel amplicons that had not been detected previously, presumably because of their small size (only 1–2 Mb) or close proximity to other larger amplicons. One of these novel amplicons involved the homeobox gene region at 17q21.3 and led to the overexpression of the *HOXB7* and *HOXB2* genes. The homeodomain transcription factors are known to be key regulators of embryonic development and have been occasionally reported to undergo aberrant expression in cancer (27, 28). *HOXB7* transfection induced cell proliferation in melanoma, breast, and ovarian cancer cells and increased tumorigenicity and angiogenesis in breast cancer (29–32). The present results imply that gene amplification may be a prominent mechanism for overexpressing *HOXB7* in breast cancer and suggest that *HOXB7* contributes to tumor progression and confers an aggressive disease phenotype in breast cancer. This view is supported by our finding of amplification of *HOXB7* in 10% of 363 primary breast cancers, as well as an association of amplification with poor prognosis of the patients.

We carried out a systematic search to identify genes whose expression levels across all 14 cell lines were attributable to amplification status. Statistical analysis revealed 270 such genes (representing ~2% of all genes on the array), including not only previously described amplified genes, such as *HER-2*, *MYC*, *EGFR*, ribosomal protein S6 kinase, and *AIB3*, but also numerous novel genes such as *NRAS-related gene* (1p13), *syndecan-2* (8q22), and *bone morphogenic protein* (20q13.1), whose activation by amplification may similarly promote breast cancer progression. Most of the 270 genes have not been implicated previously in breast cancer development and suggest novel pathogenetic mechanisms. Although we would not expect all of them to be causally involved, it is intriguing that 84% of the genes with associated functional information were implicated in apoptosis, cell proliferation, signal transduction, transcription, or other cellular processes that could directly imply a possible role in cancer progression. Therefore, a detailed characterization of these genes may provide biological insights to breast cancer progression and might lead to the development of novel therapeutic strategies.

In summary, we demonstrate application of cDNA microarrays to the analysis of both copy number and expression levels of over 12,000 transcripts throughout the breast cancer genome, roughly once every 267 kb. This analysis provided: (a) evidence of a prominent global influence of copy number changes on gene expression levels; (b) a high-resolution map of 24 independent amplicons in breast cancer; and (c) identification of a set of 270 genes, the overexpression of which was statistically attributable to gene amplification. Characterization of a novel amplicon at 17q21.3 implicated amplification and overexpression of the *HOXB7* gene in breast cancer, including a clinical association

between *HOXB7* amplification and poor patient prognosis. Overall, our results illustrate how the identification of genes activated by gene amplification provides a powerful approach to highlight genes with an important role in cancer as well as to prioritize and validate putative targets for therapy development.

REFERENCES

- Golub, T. R., Slonim, D. K., Tamayo, P., Huard, C., Gaasenbeek, M., Mesirov, J. P., Coller, H., Loh, M. L., Downing, J. R., Caligiuri, M. A., Bloomfield, C. D., and Lander, E. S. Molecular classification of cancer: class discovery and class prediction by gene expression monitoring. *Science* (Wash. DC), 286: 531–537, 1999.
- Alizadeh, A. A., Eisen, M. B., Davis, R. E., Ma, C., Lossos, I. S., Rosenwald, A., Boldrick, J. C., Sabet, H., Tran, T., Yu, X., et al. Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling. *Nature* (Lond.), 403: 503–511, 2000.
- Bittner, M., Meltzer, P., Chen, Y., Jiang, Y., Seftor, E., Hendrix, M., Radmacher, M., Simon, R., Yakhini, Z., Ben-Dor, A., et al. Molecular classification of cutaneous malignant melanoma by gene expression profiling. *Nature* (Lond.), 406: 536–540, 2000.
- Perou, C. M., Sorlie, T., Eisen, M. B., van de Rijn, M., Jeffrey, S. S., Rees, C. A., Pollack, J. R., Ross, D. T., Johnson, H., Akslen, L. A., et al. Molecular portraits of human breast tumours. *Nature* (Lond.), 406: 747–752, 2000.
- Dhanasekaran, S. M., Barrette, T. R., Ghossein, D., Shah, R., Varambally, S., Kurachi, K., Pienta, K. J., Rubin, M. A., and Chinraiyani, A. M. Delineation of prognostic biomarkers in prostate cancer. *Nature* (Lond.), 412: 822–826, 2001.
- Sorlie, T., Perou, C. M., Tibshirani, R., Aas, T., Geisler, S., Johnson, H., Hastie, T., Eisen, M. B., van de Rijn, M., Jeffrey, S. S., et al. Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc. Natl. Acad. Sci. USA*, 98: 10869–10874, 2001.
- Ross, J. S., and Fletcher, J. A. The *HER-2/neu* oncogene: prognostic factor, predictive factor and target for therapy. *Semin. Cancer Biol.*, 9: 125–138, 1999.
- Arteaga, C. L. The epidermal growth factor receptor: from mutant oncogene in nonhuman cancers to therapeutic target in human neoplasia. *J. Clin. Oncol.*, 19: 32–40, 2001.
- Knuutila, S., Bjorkqvist, A. M., Autio, K., Tarkkanen, M., Wolf, M., Monni, O., Szymanska, J., Larramendi, M. L., Tapper, J., Pere, H., El-Rifai, W., et al. DNA copy number amplifications in human neoplasms: review of comparative genomic hybridization studies. *Am. J. Pathol.*, 152: 1107–1123, 1998.
- Knuutila, S., Autio, K., and Aalto, Y. Online access to CGH data of DNA sequence copy number changes. *Am. J. Pathol.*, 157: 689, 2000.
- DeRisi, J., Penland, L., Brown, P. O., Bittner, M. L., Meltzer, P. S., Ray, M., Chen, Y., Su, Y. A., and Trent, J. M. Use of a cDNA microarray to analyse gene expression patterns in human cancer. *Nat. Genet.*, 14: 457–460, 1996.
- Shalon, D., Smith, S. J., and Brown, P. O. A DNA microarray system for analyzing complex DNA samples using two-color fluorescent probe hybridization. *Genome Res.*, 6: 639–645, 1996.
- Mousses, S., Bittner, M. L., Chen, Y., Dougherty, E. R., Baxevanis, A., Meltzer, P. S., and Trent, J. M. Gene expression analysis by cDNA microarrays. In: F. J. Livesey and S. P. Hunt (eds.), *Functional Genomics*, pp. 113–137. Oxford: Oxford University Press, 2000.
- Pollack, J. R., Perou, C. M., Alizadeh, A. A., Eisen, M. B., Pergamenschikov, A., Williams, C. F., Jeffrey, S. S., Botstein, D., and Brown, P. O. Genome-wide analysis of DNA copy-number changes using cDNA microarrays. *Nat. Genet.*, 23: 41–46, 1999.
- Monni, O., Bärilund, M., Mousses, S., Kononen, J., Sauter, G., Heiskanen, M., Paavola, P., Avela, K., Chen, Y., Bittner, M. L., and Kallioniemi, A. Comprehensive copy number and gene expression profiling of the 17q23 amplicon in human breast cancer. *Proc. Natl. Acad. Sci. USA*, 98: 5711–5716, 2001.
- Chen, Y., Dougherty, E. R., and Bittner, M. L. Ratio-based decisions and the quantitative analysis of cDNA microarray images. *J. Biomed. Optics*, 2: 364–374, 1997.
- Bärilund, M., Forozan, F., Kononen, J., Eubendorf, L., Chen, Y., Bittner, M. L., Thorst, J., Haas, P., Bucher, C., Sauter, G., et al. Detecting activation of ribosomal protein S6 kinase by complementary DNA and tissue microarray analysis. *J. Natl. Cancer Inst.*, 92: 1252–1259, 2000.
- Andersen, C. L., Hostetter, G., Grigoryan, A., Sauter, G., and Kallioniemi, A. Improved procedure for fluorescence *in situ* hybridization on tissue microarrays. *Cytometry*, 43: 83–86, 2001.
- Kauraniemi, P., Bärilund, M., Monni, O., and Kallioniemi, A. New amplified and highly expressed genes discovered in the ERBB2 amplicon in breast cancer by cDNA microarrays. *Cancer Res.*, 61: 8235–8240, 2001.
- Clark, J., Edwards, S., John, M., Flohr, P., Gordon, T., Maillard, K., Giddings, I., Brown, C., Bagherzadeh, A., Campbell, C., Shipley, J., Wooster, R., and Cooper, C. S. Identification of amplified and expressed genes in breast cancer by comparative hybridization onto microarrays of randomly selected cDNA clones. *Genes Chromosomes Cancer*, 34: 104–114, 2002.
- Varis, A., Wolf, M., Monni, O., Vakkari, M. L., Kokkola, A., Moskaluk, C., Frierson, H., Powell, S. M., Knuutila, S., Kallioniemi, A., and El-Rifai, W. Targets of gene amplification and overexpression at 17q in gastric cancer. *Cancer Res.*, 62: 2625–2629, 2002.
- Hughes, T. R., Roberts, C. J., Dai, H., Jones, A. R., Meyer, M. R., Slade, D., Burchard, J., Dow, S., Ward, T. R., Kidd, M. J., Friend, S. H., and Marton, M. J.

- Widespread aneuploidy revealed by DNA microarray expression profiling. *Nat. Genet.*, 25: 333-337, 2000.
23. Virtaneva, K., Wright, F. A., Tanner, S. M., Yuan, B., Lemon, W. J., Caligiuri, M. A., Bloomfield, C. D., de La Chapelle, A., and Krahe, R. Expression profiling reveals fundamental biological differences in acute myeloid leukemia with isolated trisomy 8 and normal cytogenetics. *Proc. Natl. Acad. Sci. USA*, 98: 1124-1129, 2001.
24. Phillips, J. L., Hayward, S. W., Wang, Y., Vasselli, J., Pavlovich, C., Padilla-Nash, H., Pezullo, J. R., Ghadimi, B. M., Grossfeld, G. D., Rivera, A., Linchan, W. M., Cunha, G. R., and Ried, T. The consequences of chromosomal aneuploidy on gene expression profiles in a cell line model for prostate carcinogenesis. *Cancer Res.*, 61: 8143-8149, 2001.
25. Bärlund, M., Tirkkonen, M., Forozan, F., Tanner, M. M., Kallioniemi, O. P., and Kallioniemi, A. Increased copy number at 17q22-q24 by CGH in breast cancer is due to high-level amplification of two separate regions. *Genes Chromosomes Cancer*, 20: 372-376, 1997.
26. Tanner, M. M., Tirkkonen, M., Kallioniemi, A., Isola, J., Kuukasjärvi, T., Collins, C., Kowbel, D., Guan, X. Y., Trent, J., Gray, J. W., Meltzer, P., and Kallioniemi, O. P. Independent amplification and frequent co-amplification of three nonsyntenic regions on the long arm of chromosome 20 in human breast cancer. *Cancer Res.*, 56: 3441-3445, 1996.
27. Cillo, C., Faiella, A., Cantile, M., and Boncinelli, E. Homeobox genes and cancer. *Exp. Cell Res.*, 248: 1-9, 1999.
28. Cillo, C., Cantile, M., Faiella, A., and Boncinelli, E. Homeobox genes in normal and malignant cells. *J. Cell. Physiol.*, 188: 161-169, 2001.
29. Care, A., Silvani, A., Meccia, E., Mantia, G., Stoppacciaro, A., Parmiani, G., Peschle, C., and Colombo, M. P. HOXB7 constitutively activates basic fibroblast growth factor in melanomas. *Mol. Cell. Biol.*, 16: 4842-4851, 1996.
30. Care, A., Silvani, A., Meccia, E., Mantia, G., Peschle, C., and Colombo, M. P. Transduction of the SkBr3 breast carcinoma cell line with the HOXB7 gene induces bFGF expression, increases cell proliferation and reduces growth factor dependence. *Oncogene*, 16: 3285-3289, 1998.
31. Care, A., Felicetti, F., Meccia, E., Bottero, L., Parenza, M., Stoppacciaro, A., Peschle, C., and Colombo, M. P. HOXB7: a key factor for tumor-associated angiogenic switch. *Cancer Res.*, 61: 6532-6539, 2001.
32. Naora, H., Yang, Y. Q., Montz, F. J., Seidman, J. D., Kurman, R. J., and Roden, R. B. A serologically identified tumor antigen encoded by a homeobox gene promotes growth of ovarian epithelial cells. *Proc. Natl. Acad. Sci. USA*, 98: 4060-4065, 2001.

Microarray analysis reveals a major direct role of DNA copy number alteration in the transcriptional program of human breast tumors

Jonathan R. Pollack^{1,2*}, Therese Sørlie³, Charles M. Perou⁴, Christian A. Rees^{1,2*}, Stefanie S. Jeffrey^{2*}, Per E. Lønning^{2*}, Robert Tibshirani⁵, David Botstein¹, Anne-Lise Børresen-Dale³, and Patrick O. Brown^{1,2*}

Departments of ¹Pathology, ²Genetics, ³Surgery, ⁴Health Research and Policy, and ⁵Biochemistry, and ⁶Howard Hughes Medical Institute, Stanford University School of Medicine, Stanford, CA 94305; ¹Department of Genetics, Norwegian Radium Hospital, Montebello, N-0310 Oslo, Norway; ²Department of Medicine (Oncology), Haukeland University Hospital, N-5021 Bergen, Norway; and ³Department of Genetics and Lineberger Comprehensive Cancer Center, University of North Carolina, Chapel Hill, NC 27599

Contributed by Patrick O. Brown, August 6, 2002

Genomic DNA copy number alterations are key genetic events in the development and progression of human cancers. Here we report a genome-wide microarray comparative genomic hybridization (array CGH) analysis of DNA copy number variation in a series of primary human breast tumors. We have profiled DNA copy number alteration across 6,691 mapped human genes, in 44 predominantly advanced, primary breast tumors and 10 breast cancer cell lines. While the overall patterns of DNA amplification and deletion corroborate previous cytogenetic studies, the high-resolution (gene-by-gene) mapping of amplicon boundaries and the quantitative analysis of amplicon shape provide significant improvement in the localization of candidate oncogenes. Parallel microarray measurements of mRNA levels reveal the remarkable degree to which variation in gene copy number contributes to variation in gene expression in tumor cells. Specifically, we find that 62% of highly amplified genes show moderately or highly elevated expression, that DNA copy number influences gene expression across a wide range of DNA copy number alterations (deletion, low-, mid- and high-level amplification), that on average, a 2-fold change in DNA copy number is associated with a corresponding 1.5-fold change in mRNA levels, and that overall, at least 12% of all the variation in gene expression among the breast tumors is directly attributable to underlying variation in gene copy number. These findings provide evidence that widespread DNA copy number alteration can lead directly to global deregulation of gene expression, which may contribute to the development or progression of cancer.

Conventional cytogenetic techniques, including comparative genomic hybridization (CGH) (1), have led to the identification of a number of recurrent regions of DNA copy number alteration in breast cancer cell lines and tumors (2–4). While some of these regions contain known or candidate oncogenes [e.g., *FGFR1* (8p11), *MYC* (8q24), *CCND1* (11q13), *ERBB2* (17q12), and *ZNF217* (20q13)] and tumor suppressor genes [*RB1* (13q14) and *TP53* (17p13)], the relevant gene(s) within other regions (e.g., gain of 1q, 8q22, and 17q22–24, and loss of 8p) remain to be identified. A high-resolution genome-wide map, delineating the boundaries of DNA copy number alterations in tumors, should facilitate the localization and identification of oncogenes and tumor suppressor genes in breast cancer. In this study, we have created such a map, using array-based CGH (5–7) to profile DNA copy number alteration in a series of breast cancer cell lines and primary tumors.

An unresolved question is the extent to which the widespread DNA copy number changes that we and others have identified in breast tumors alter expression of genes within involved regions. Because we had measured mRNA levels in parallel in the same samples (8), using the same DNA microarrays, we had an opportunity to explore on a genomic scale the relationship between DNA copy number changes and gene expression. From

this analysis, we have identified a significant impact of widespread DNA copy number alteration on the transcriptional programs of breast tumors.

Materials and Methods

Tumors and Cell Lines. Primary breast tumors were predominantly large (>3 cm), intermediate-grade, infiltrating ductal carcinomas, with more than 50% being lymph node positive. The fraction of tumor cells within specimens averaged at least 50%. Details of individual tumors have been published (8, 9), and are summarized in Table 1, which is published as supporting information on the PNAS web site, www.pnas.org. Breast cancer cell lines were obtained from the American Type Culture Collection. Genomic DNA was isolated either using Qiagen genomic DNA columns, or by phenol/chloroform extraction followed by ethanol precipitation.

DNA Labeling and Microarray Hybridizations. Genomic DNA labeling and hybridizations were performed essentially as described in Pollack *et al.* (7), with slight modifications. Two micrograms of DNA was labeled in a total volume of 50 microliters and the volumes of all reagents were adjusted accordingly. “Test” DNA (from tumors and cell lines) was fluorescently labeled (Cy5) and hybridized to a human cDNA microarray containing 6,691 different mapped human genes (i.e., UniGene clusters). The “reference” (labeled with Cy3) for each hybridization was normal female leukocyte DNA from a single donor. The fabrication of cDNA microarrays and the labeling and hybridization of mRNA samples have been described (8).

Data Analysis and Map Positions. Hybridized arrays were scanned on a GenePix scanner (Axon Instruments, Foster City, CA), and fluorescence ratios (test/reference) calculated using SCANALYZE software (available at <http://rana.lbl.gov>). Fluorescence ratios were normalized for each array by setting the average log fluorescence ratio for all array elements equal to 0. Measurements with fluorescence intensities more than 20% above background were considered reliable. DNA copy number profiles that deviated significantly from background ratios measured in normal genomic DNA control hybridizations were interpreted as evidence of real DNA copy number alteration (see *Estimating Significance of Altered Fluorescence Ratios* in the supporting information). When indicated, DNA copy number profiles are displayed as a moving average (symmetric 5-nearest neighbors). Map positions for arrayed human cDNAs were assigned by

Abbreviation: CGH, comparative genomic hybridization.

*To whom reprint requests should be addressed at: Department of Pathology, Stanford University School of Medicine, CCSR Building, Room 3245A, 269 Campus Drive, Stanford, CA 94305-5176. E-mail: pollack1@stanford.edu.

**Present address: Zymyx Inc., Hayward, CA 94545.

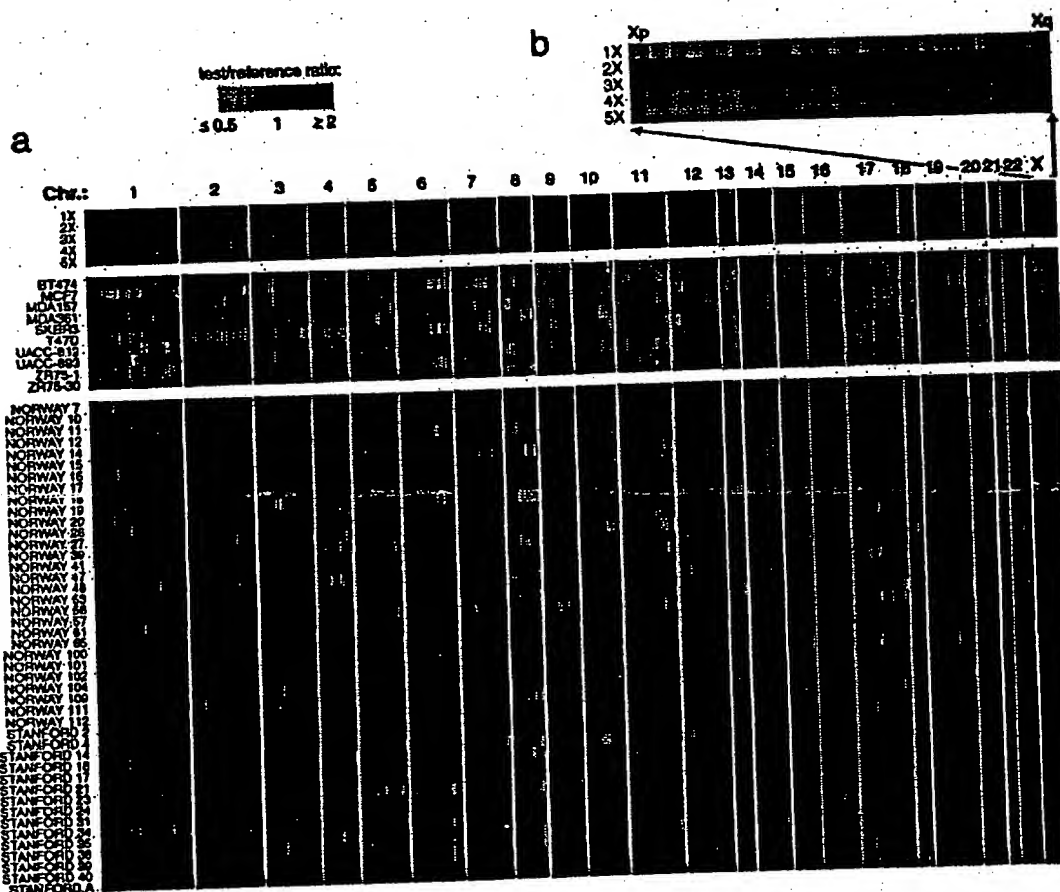


Fig. 1. Genome-wide measurement of DNA copy number alteration by array CGH. (a) DNA copy number profiles are illustrated for cell lines containing different numbers of X chromosomes, for breast cancer cell lines, and for breast tumors. Each row represents a different cell line or tumor, and each column represents one of 6,691 different mapped human genes present on the microarray, ordered by genome map position from 1pter through Xqter. Moving average (symmetric 5-nearest neighbors) fluorescence ratios (test/reference) are depicted using a log₂-based pseudocolor scale (indicated), such that red luminescence reflects fold-amplification, green luminescence reflects fold-deletion, and black indicates no change (gray indicates poorly measured data). (b) Enlarged view of DNA copy number profiles across the X chromosome, shown for cell lines containing different numbers of X chromosomes.

identifying the starting position of the best and longest match of any DNA sequence represented in the corresponding UniGene cluster (10) against the "Golden Path" genome assembly (<http://genome.ucsc.edu/>; Oct 7, 2000 Freeze). For UniGene clusters represented by multiple arrayed elements, mean fluorescence ratios (for all elements representing the same UniGene cluster) are reported. For mRNA measurements, fluorescence ratios are "mean-centered" (i.e., reported relative to the mean ratio across the 44 tumor samples). The data set described here can be accessed in its entirety in the supporting information.

Results

We performed CGH on 44 predominantly locally advanced, primary breast tumors and 10 breast cancer cell lines, using cDNA microarrays containing 6,691 different mapped human genes (Fig. 1a; also see *Materials and Methods* for details of microarray hybridizations). To take full advantage of the improved spatial resolution of array CGH, we ordered (fluorescence ratios for) the 6,691 cDNAs according to the "Golden Path" (<http://genome.ucsc.edu/>) genome assembly of the draft human genome sequences (11). In so doing, arrayed cDNAs not only themselves represent genes of potential interest (e.g., candidate oncogenes within amplicons), but also provide precise genetic landmarks for chromosomal regions of amplification and

deletion. Parallel analysis of DNA from cell lines containing different numbers of X chromosomes (Fig. 1b), as we did before (7), demonstrated the sensitivity of our method to detect single-copy loss (45, XO), and 1.5- (47,XXX), 2- (48,XXXX), or 2.5-fold (49,XXXXX) gains (also see Fig. 5, which is published as supporting information on the PNAS web site). Fluorescence ratios were linearly proportional to copy number ratios, which were slightly underestimated, in agreement with previous observations (7). Numerous DNA copy number alterations were evident in both the breast cancer cell lines and primary tumors (Fig. 1a), detected in the tumors despite the presence of euploid non-tumor cell types; the magnitudes of the observed changes were generally lower in the tumor samples. DNA copy-number alterations were found in every cancer cell line and tumor, and on every human chromosome in at least one sample. Recurrent regions of DNA copy number gain and loss were readily identifiable. For example, gains within 1q, 8q, 17q, and 20q were observed in a high proportion of breast cancer cell lines/tumors (90%/69%, 100%/47%, 100%/60%, and 90%/44%, respectively), as were losses within 1p, 3p, 8p, and 13q (80%/24%, 80%/22%, 80%/22%, and 70%/18%, respectively), consistent with published cytogenetic studies (refs. 2-4; a complete listing of gains/losses is provided in Tables 2 and 3, which are published as supporting information on the PNAS web site). The total

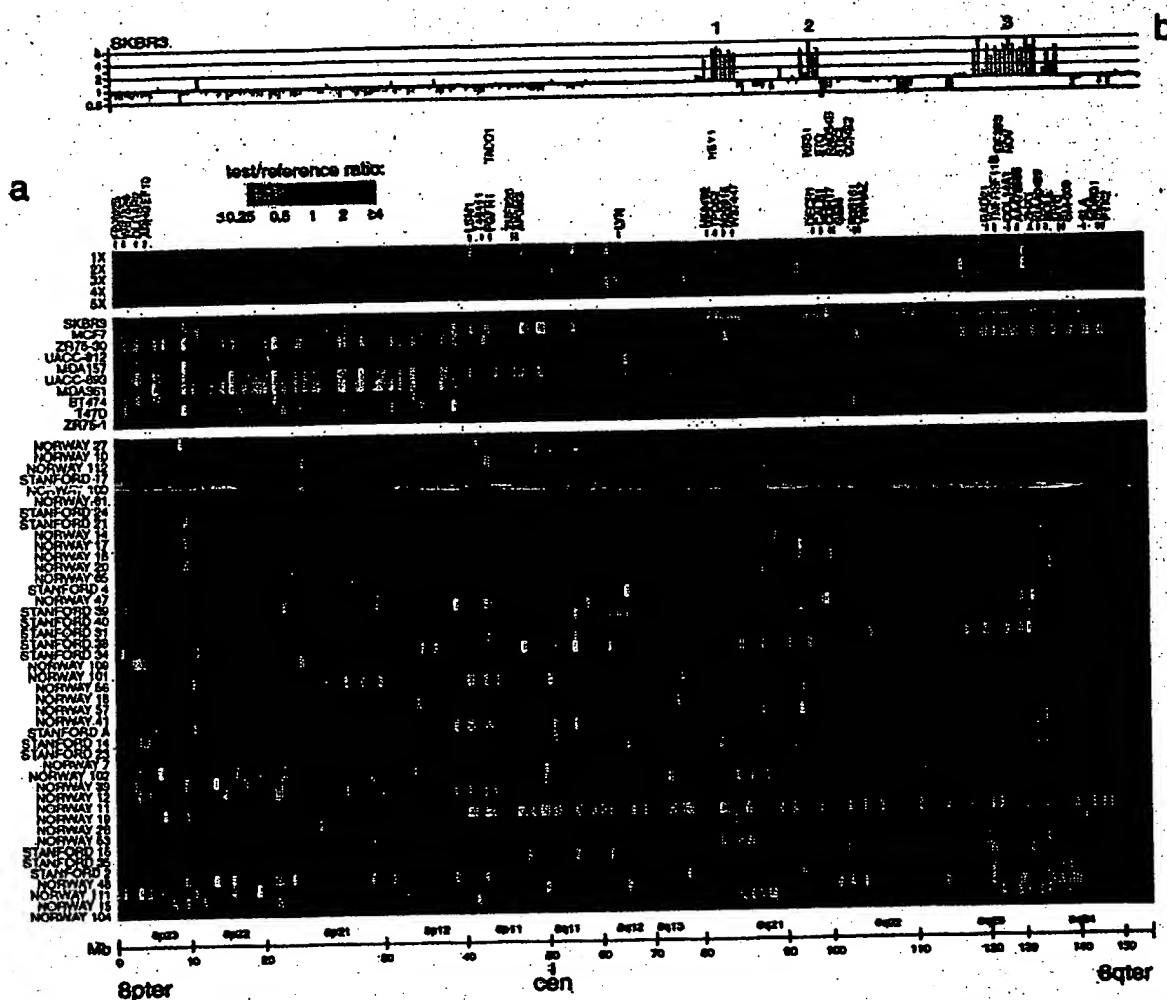


Fig. 2. DNA copy number alteration across chromosome 8 by array CGH. (a) DNA copy number profiles are illustrated for cell lines containing different numbers of X chromosomes, for breast cancer cell lines, and for breast tumors. Breast cancer cell lines and tumors are separately ordered by hierarchical clustering to highlight recurrent copy number changes. The 241 genes present on the microarrays and mapping to chromosome 8 are ordered by position along the chromosome. Fluorescence ratios (test/reference) are depicted by a \log_2 pseudocolor scale (indicated). Selected genes are indicated with color-coded text (red, increased; green, decreased; black, no change; gray, not well measured) to reflect correspondingly altered mRNA levels (observed in the majority of the subset of samples displaying the DNA copy number change). The map positions for genes of interest that are not represented on the microarray are indicated in the row above those genes represented on the array. (b) Graphical display of DNA copy number profile for breast cancer cell line SKBR3. Fluorescence ratios (tumor/normal) are plotted on a \log_2 scale for chromosome 8 genes, ordered along the chromosome.

number of genomic alterations (gains and losses) was found to be significantly higher in breast tumors that were high grade ($P = 0.008$), consistent with published CGH data (3), estrogen receptor negative ($P = 0.04$), and harboring TP53 mutations ($P = 0.0006$) (see Table 4, which is published as supporting information on the PNAS web site).

The improved spatial resolution of our array CGH analysis is illustrated for chromosome 8, which displayed extensive DNA copy number alteration in our series. A detailed view of the variation in the copy number of 241 genes mapping to chromosome 8 revealed multiple regions of recurrent amplification; each of these potentially harbors a different known or previously uncharacterized oncogene (Fig. 2a). The complexity of amplicon structure is most easily appreciated in the breast cancer cell line SKBR3. Although a conventional CGH analysis of 8q in SKBR3 identified only two distinct regions of amplification (12), we observed three distinct regions of high-level amplification (labeled 1–3 in Fig. 2b). For each of these regions we can define the

boundaries of the interval recurrently amplified in the tumors we examined; in each case, known or plausible candidate oncogenes can be identified (a description of these regions, as well as the recurrently amplified regions on chromosomes 17 and 20, can be found in Figs. 6 and 7, which are published as supporting information on the PNAS web site).

For a subset of breast cancer cell lines and tumors (4 and 37, respectively), and a subset of arrayed genes (6,095), mRNA levels were quantitatively measured in parallel by using cDNA microarrays (8). The parallel assessment of mRNA levels is useful in the interpretation of DNA copy number changes. For example, the highly amplified genes that are also highly expressed are the strongest candidate oncogenes within an amplicon. Perhaps more significantly, our parallel analysis of DNA copy number changes and mRNA levels provides us the opportunity to assess the global impact of widespread DNA copy number alteration on gene expression in tumor cells.

A strong influence of DNA copy number on gene expression is evident in an examination of the pseudocolor representations

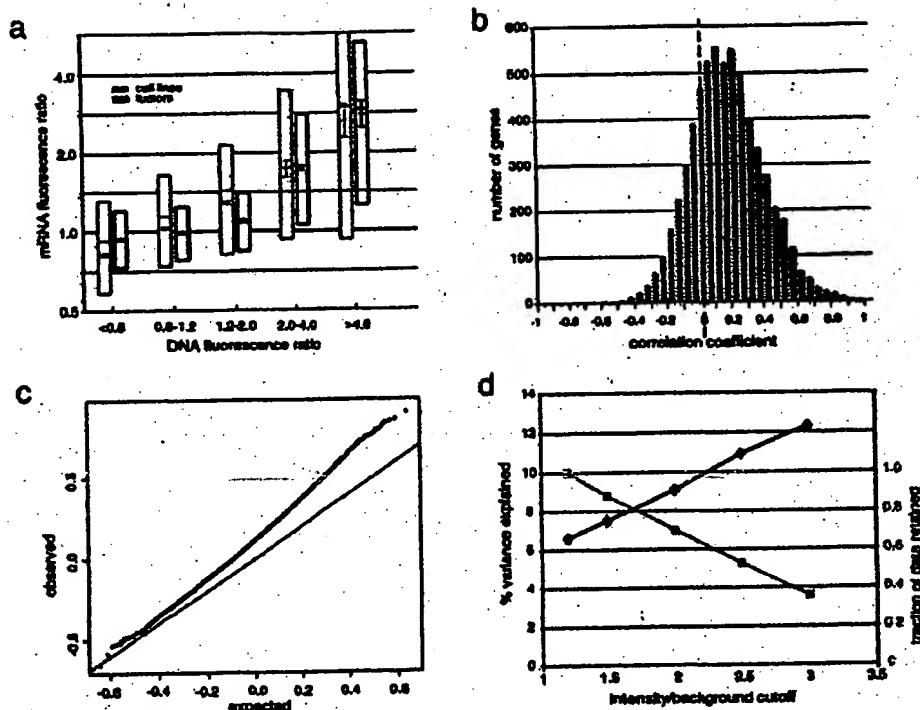


Fig. 4. Genome-wide influence of DNA copy number alterations on mRNA levels. (a) For breast cancer cell lines (gray) and tumor samples (black), both mean-centered mRNA fluorescence ratio (log₂ scale) quartiles (box plots indicate 25th, 50th, and 75th percentile) and averages (diamonds; Y-value error bars indicate standard errors of the mean) are plotted for each of five classes of genes, representing DNA deletion (tumor/normal ratio < 0.8), no change (0.8–1.2), low- (1.2–2), medium- (2–4), and high-level (>4) amplification. *P* values for pair-wise Student's *t* tests, comparing averages between adjacent classes (moving left to right), are 4×10^{-40} , 1×10^{-40} , 5×10^{-35} , 1×10^{-23} (cell lines), and 1×10^{-40} , 1×10^{-214} , 5×10^{-41} , 1×10^{-4} (tumors). (b) Distribution of correlations between DNA copy number and mRNA levels, for 6,095 different human genes across 37 breast tumor samples. (c) Plot of observed versus expected correlation coefficients. The expected values were obtained by randomization of the sample labels in the DNA copy number data set. The line of unity is indicated. (d) Percent variance in gene expression (among tumors) directly explained by variation in gene copy number. Percent variance explained (black line) and fraction of data retained (gray line) are plotted for different fluorescence intensity/background (a rough surrogate for signal/noise) cutoff values. Fraction of data retained is relative to the 1.2 intensity/background cutoff. Details of the linear regression model used to estimate the fraction of variation in gene expression attributable to underlying DNA copy number alteration can be found in the supporting information (see *Estimating the Fraction of Variation in Gene Expression Attributable to Underlying DNA Copy Number Alteration*).

tumors that could be attributed to underlying variation in DNA copy number. From this analysis, we estimate that, overall, about 7% of all of the observed variation in mRNA levels can be explained directly by variation in copy number of the altered genes (Fig. 4d). We can reduce the effects of experimental measurement error on this estimate by using only that fraction of the data most reliably measured (fluorescence intensity/background > 3); using that data, our estimate of the percent variation in mRNA levels directly attributed to variation in gene copy number increases to 12% (Fig. 4d). This still undoubtedly represents a significant underestimate, as the observed variation in global gene expression is affected not only by true variation in the expression programs of the tumor cells themselves, but also by the variable presence of non-tumor cell types within clinical samples.

Discussion

This genome-wide, array CGH analysis of DNA copy number alteration in a series of human breast tumors demonstrates the usefulness of defining amplicon boundaries at high resolution (gene-by-gene), and quantitatively measuring amplicon shape, to assist in locating and identifying candidate oncogenes. By analyzing mRNA levels in parallel, we have also discovered that changes in DNA copy number have a large, pervasive, direct effect on global gene expression patterns in both breast cancer

cell lines and tumors. Although the DNA microarrays used in our analysis may display a bias toward characterized and/or highly expressed genes, because we are examining such a large fraction of the genome (approximately 20% of all human genes), and because, as detailed above, we are likely underestimating the contribution of DNA copy number changes to altered gene expression, we believe our findings are likely to be generalizable (but would nevertheless still be remarkable if only applicable to this set of ~6,100 genes).

In budding yeast, aneuploidy has been shown to result in chromosome-wide gene expression biases (13). Two recent studies have begun to examine the global relationship between DNA copy number and gene expression in cancer cells. In agreement with our findings, Phillips *et al.* (14) have shown that with the acquisition of tumorigenicity in an immortalized prostate epithelial cell line, new chromosomal gains and losses resulted in a statistically significant respective increase and decrease in the average expression level of involved genes. In contrast, Platzer *et al.* (15) recently reported that in metastatic colon tumors only ~4% of genes within amplified regions were found more highly (>2-fold) expressed, when compared with normal colonic epithelium. This report differs substantially from our finding that 62% of highly amplified genes in breast cancer exhibit at least 2-fold increased expression. These contrasting findings may reflect methodological differences between the

studies. For example, the study of Platzer *et al.* (15) may have systematically under-measured gene expression changes. In this regard it is remarkable that only 14 transcripts of many thousand residing within unamplified chromosomal regions were found to exhibit at least 4-fold altered expression in metastatic colon cancer. Additionally, their reliance on lower-resolution chromosomal CGH may have resulted in poorly delimiting the boundaries of high-complexity amplicons, effectively overcalling regions with amplification. Alternatively, the contrasting findings for amplified genes may represent real biological differences between breast and metastatic colon tumors; resolution of this issue will require further studies.

Our finding that widespread DNA copy number alteration has a large, pervasive and direct effect on global gene expression patterns in breast cancer has several important implications. First, this finding supports a high degree of copy number-dependent gene expression in tumors. Second, it suggests that most genes are not subject to specific autoregulation or dosage compensation. Third, this finding cautions that elevated expression of an amplified gene cannot alone be considered strong independent evidence of a candidate oncogene's role in tumorigenesis. In our study, fully 62% of highly amplified genes demonstrated moderately or highly elevated expression. This highlights the importance of high-resolution mapping of amplicon boundaries and shape [to identify the "driving" gene(s) within amplicons (16)], on a large number of samples, in addition to functional studies. Fourth, this finding suggests that analyzing

the genomic distribution of expressed genes, even within existing microarray gene expression data sets, may permit the inference of DNA copy number aberration, particularly aneuploidy (where gene expression can be averaged across large chromosomal regions; see Fig. 3 and supporting information). Fifth, this finding implies that a substantial portion of the phenotypic uniqueness (and by extension, the heterogeneity in clinical behavior) among patients' tumors may be traceable to underlying variation in DNA copy number. Sixth, this finding supports a possible role for widespread DNA copy number alteration in tumorigenesis (17, 18), beyond the amplification of specific oncogenes and deletion of specific tumor suppressor genes. Widespread DNA copy number alteration, and the concomitant widespread imbalance in gene expression, might disrupt critical stoichiometric relationships in cell metabolism and physiology (e.g., proteasome, mitotic spindle), possibly promoting further chromosomal instability and directly contributing to tumor development or progression. Finally, our findings suggest the possibility of cancer therapies that exploit specific or global imbalances in gene expression in cancer.

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1. Kallioniemi, A., Kallioniemi, O. P., Sudar, D., Rutovitz, D., Gray, J. W., Waldman, F. & Pinkel, D. (1992) *Science* 258, 818–821.
2. Kallioniemi, A., Kallioniemi, O. P., Piper, J., Tanner, M., Stokke, T., Chen, L., Smith, H. S., Pinkel, D., Gray, J. W. & Waldman, F. M. (1994) *Proc. Natl. Acad. Sci. USA* 91, 2156–2160.
3. Tirkkoonen, M., Tanner, M., Karhu, R., Kallioniemi, A., Isola, J. & Kallioniemi, O. P. (1998) *Genes Chromosomes Cancer* 21, 177–184.
4. Forozan, F., Mahlamaki, E. H., Monni, O., Chen, Y., Veldman, R., Jiang, Y., Gooden, G. C., Ethier, S. P., Kallioniemi, A. & Kallioniemi, O. P. (2000) *Cancer Res.* 60, 4519–4525.
5. Solinas-Toldo, S., Lampel, S., Stilgenbauer, S., Nickolenko, J., Benner, A., Dohner, H., Cremer, T. & Lichter, P. (1997) *Genes Chromosomes Cancer* 20, 399–407.
6. Pinkel, D., Segreves, R., Sudar, D., Clark, S., Poole, L., Kowbel, D., Collins, C., Kuo, W. L., Chen, C., Zhai, Y., *et al.* (1998) *Nat. Genet.* 20, 207–211.
7. Pollack, J. R., Perou, C. M., Alizadeh, A. A., Eisen, M. B., Pergamenschikov, A., Williams, C. F., Jeffrey, S. S., Botstein, D. & Brown, P. O. (1999) *Nat. Genet.* 23, 41–46.
8. Perou, C. M., Sorlie, T., Eisen, M. B., van de Rijn, M., Jeffrey, S. S., Rees, C. A., Pollack, J. R., Ross, D. T., Johnsen, H., Akslen, L. A., *et al.* (2000) *Nature (London)* 406, 747–752.
9. Sorlie, T., Perou, C. M., Tibshirani, R., Aas, T., Geister, S., Johnsen, H., Hastie, T., Eisen, M. B., van de Rijn, M., Jeffrey, S. S., *et al.* (2001) *Proc. Natl. Acad. Sci. USA* 98, 10869–10874.
10. Schuler, G. D. (1997) *J. Mol. Med.* 75, 694–698.
11. Lander, E. S., Linton, L. M., Birren, B., Nusbaum, C., Zody, M. C., Baldwin, J., Devon, K., Dewar, K., Doyle, M., FitzHugh, W., *et al.* (2001) *Nature (London)* 409, 860–921.
12. Fejzo, M. S., Godfrey, T., Chen, C., Waldman, F. & Gray, J. W. (1998) *Genes Chromosomes Cancer* 22, 105–113.
13. Hughes, T. R., Roberts, C. J., Dai, H., Jones, A. R., Meyer, M. R., Slade, D., Burchard, J., Dow, S., Ward, T. R., Kidd, M. J., *et al.* (2000) *Nat. Genet.* 25, 333–337.
14. Phillips, J. L., Hayward, S. W., Wang, Y., Vasselli, J., Pavlovich, C., Padilla-Nash, H., Pezullo, J. R., Ghadimi, B. M., Grossfeld, G. D., Rivera, A., *et al.* (2001) *Cancer Res.* 61, 8143–8149.
15. Platzer, P., Upender, M. B., Wilson, K., Willis, J., Lutterbaugh, J., Nosrati, A., Wilson, J. K., Mack, D., Ried, T. & Markowitz, S. (2002) *Cancer Res.* 62, 1134–1138.
16. Albertson, D. G., Ylstra, B., Segraves, R., Collins, C., Dairkee, S. H., Kowbel, D., Kuo, W. L., Gray, J. W. & Pinkel, D. (2000) *Nat. Genet.* 25, 144–146.
17. Li, R., Yerganian, G., Duesberg, P., Kraemer, A., Willer, A., Rausch, C. & Hehlmann, R. (1997) *Proc. Natl. Acad. Sci. USA* 94, 14506–14511.
18. Rasnick, D. & Duesberg, P. H. (1999) *Biochem. J.* 348, 621–630.



TECHNICAL UPDATE

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HER-2/neu Breast Cancer Predictive Testing

Julie Sanford Hanna, Ph.D. and Dan Mornin, M.D.

EACH YEAR, OVER 182,000 WOMEN in the United States are diagnosed with breast cancer, and approximately 45,000 die of the disease.¹ Incidence appears to be increasing in the United States at a rate of roughly 2% per year. The reasons for the increase are unclear, but non-genetic risk factors appear to play a large role.²

Five-year survival rates range from approximately 65%-85%, depending on demographic group, with a significant percentage of women experiencing recurrence of their cancer within 10 years of diagnosis. One of the factors most predictive for recurrence once a diagnosis of breast cancer has been made is the number of axillary lymph nodes to which tumor has metastasized. Most node-positive women are given adjuvant therapy, which increases their survival. However, 20%-30% of patients without axillary node involvement also develop recurrent disease, and the difficulty lies in how to identify this high-risk subset of patients. These patients could benefit from increased surveillance, early intervention, and treatment.

Prognostic markers currently used in breast cancer recurrence prediction include tumor size, histological grade, steroid hormone receptor status, DNA ploidy, proliferative index, and cathepsin D status. Expression of growth factor receptors and over-expression of the HER-2/neu oncogene have also been identified as having value regarding treatment regimen and prognosis.

HER-2/neu (also known as c-erbB2) is an oncogene that encodes a transmembrane glycoprotein that is homologous to, but distinct from, the epidermal growth factor receptor. Numerous studies have indicated that high levels of expression of this protein are associated with rapid tumor growth, certain forms of therapy resistance, and shorter disease-free survival. The gene has been shown to be amplified and/or overexpressed in 10%-30% of invasive breast cancers and in 40%-60% of intraductal breast carcinoma.³

There are two distinct FDA-approved methods by which HER-2/neu status can be evaluated: immunohistochemistry (IHC, HercepTest™) and FISH (fluorescent in situ hybridization, PathVysion™ Kit). Both methods can be performed on archived and current specimens. The first method allows visual assessment of the amount of HER-2/neu protein present on the cell membrane. The latter method allows direct quantification of the level of gene amplification present in the tumor, enabling differentiation between low- versus high-amplification. At least one study has demonstrated a difference in

recurrence risk in women younger than 40 years of age for low- versus high-amplified tumors (54.5% compared to 85.7%); this is compared to a recurrence rate of 16.7% for patients with no HER-2/neu gene amplification.⁴ HER-2/neu status may be particularly important to establish in women with small (≤ 1 cm) tumor size.

The choice of methodology for determination of HER-2/neu status depends in part on the clinical setting. FDA approval for the Vysis FISH test was granted based on clinical trials involving 1549 node-positive patients. Patients received one of three different treatments consisting of different doses of cyclophosphamide, Adriamycin, and 5-fluorouracil (CAF). The study showed that patients with amplified HER-2/neu benefited from treatment with higher doses of adriamycin-based therapy, while those with normal HER-2/neu levels did not. The study therefore identified a sub-set of women, who because they did not benefit from more aggressive treatment, did not need to be exposed to the associated side effects. In addition, other evidence indicates that HER-2/neu amplification in node-negative patients can be used as an independent prognostic indicator for early recurrence, recurrent disease at any time and disease-related death.⁵ Demonstration of HER-2/neu gene amplification by FISH has also been shown to be of value in predicting response to chemotherapy in stage-2 breast cancer patients.

Selection of patients for Herceptin® (Trastuzumab) monoclonal antibody therapy, however, is based upon demonstration of HER-2/neu protein overexpression using HercepTest™. Studies using Herceptin® in patients with metastatic breast cancer show an increase in time to disease progression, increased response rate to chemotherapeutic agents and a small increase in overall survival rate. The FISH assays have not yet been approved for this purpose, and studies looking at response to Herceptin® in patients with or without gene amplification status determined by FISH are in progress.

In general, FISH and IHC results correlate well. However, subsets of tumors are found which show discordant results; i.e., protein overexpression without gene amplification or lack of protein overexpression with gene amplification. The clinical significance of such results is unclear. Based on the above considerations, HER-2/neu testing at SHMC/PAML will utilize immunohistochemistry (HercepTest®) as a screen, followed by FISH in IHC-negative cases. Alternatively, either method may be ordered individually depending on the clinical setting or clinician preference.

CPT code information

HER-2/neu via IHC

88342 (including interpretive report)

HER-2/neu via FISH

88271×2 Molecular cytogenetics, DNA probe, each

88274 Molecular cytogenetics, interphase in situ hybridization, analyze 25-99 cells

88291 Cytogenetics and molecular cytogenetics, interpretation and report

Procedural Information

Immunohistochemistry is performed using the FDA-approved DAKO antibody kit, Herceptest®. The DAKO kit contains reagents required to complete a two-step immunohistochemical staining procedure for routinely processed, paraffin-embedded specimens. Following incubation with the primary rabbit antibody to human HER-2/neu protein, the kit employs a ready-to-use dextran-based visualization reagent. This reagent consists of both secondary goat anti-rabbit antibody molecules with horseradish peroxidase molecules linked to a common dextran polymer backbone, thus eliminating the need for sequential application of link antibody and peroxidase conjugated antibody. Enzymatic conversion of the subsequently added chromogen results in formation of visible reaction product at the antigen site. The specimen is then counterstained; a pathologist using light-microscopy interprets results.

FISH analysis at SHMC/PAML is performed using the FDA-approved PathVysion™ HER-2/neu DNA probe kit, produced by Vysis, Inc. Formalin fixed, paraffin-embedded breast tissue is processed using routine histological methods, and then slides are treated to allow hybridization of DNA probes to the nuclei present in the tissue section. The PathVysion™ kit contains two direct-labeled DNA probes, one specific for the alphoid repetitive DNA (CEP 17, spectrum orange) present at the chromosome 17 centromere and the second for the HER-2/neu oncogene located at 17q11.2-12 (spectrum green). Enumeration of the probes allows a ratio of the number of copies of chromosome 17 to the number of copies of HER-2/neu to be obtained; this enables quantification of low versus high amplification levels, and allows an estimate of the percentage of cells with HER-2/neu gene amplification. The clinically relevant distinction is whether the gene amplification is due to increased gene copy number on the two chromosome 17 homologues normally present or an increase in the number of chromosome 17s in the cells. In the majority of cases, ratio equivalents less than 2.0 are indicative of a normal/negative result, ratios of 2.1 and over indicate that amplification is present and to what degree. Interpretation of this data will be performed and reported from the Vysis-certified Cytogenetics laboratory at SHMC.

References

1. Wingo, P.A., Tong, T., Bolden, S., "Cancer Statistics", 1995;45:1:8-31.
2. "Cancer Rates and Risks", 4th ed., National Institutes of Health, National Cancer Institute, 1996, p. 120.
3. Slamon, D.J., Clark, G.M., Song, S.G., Levin, W.J., Ullrich, A., McGuire, W.L. "Human breast Cancer: Correlation of relapse and survival with amplification of the her-2/neu oncogene". Science, 235:177-182, 1987.
4. Xing, W.R., Gilchrist, K.W., Harris, C.P., Samson, W., Meisner, L.F. "FISH detection of HER-2/neu oncogene amplification in early onset breast cancer". Breast Cancer Res. And Treatment 39(2):203-212, 1996.
5. Press, M.F., Bernstein, L., Thomas, P.A., Meisner, L.F., Zhou, J.Y., Ma, Y., Hung, G., Robinson, R.A., Harris, C., El-Naggar, A., Slamon, D.J., Phillips, R.N., Ross, J.S., Wolman, S.R., Flom, K.J., "Her-2/neu gene amplification characterized by fluorescence in situ hybridization: poor prognosis in node-negative breast carcinomas", J. Clinical Oncology 15(8):2894-2904, 1997.

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Variable expression of the translocated *c-abl* oncogene in Philadelphia-chromosome-positive B-lymphoid cell lines from chronic myelogenous leukemia patients

JAMES B. KONOPKA^{*,†}, STEVEN CLARK^{*}, JAMI McLAUGHLIN^{*}, MASAKUZU NITTA[†], YOSHIRO KATO[†], ANNABEL STRIFE[†], BAYARD CLARKSON[†], AND OWEN N. WITTE^{*,§}

^{*}Department of Microbiology and Molecular Biology Institute, University of California, Los Angeles, 405 Hilgard Avenue, Los Angeles, CA 90024; and [†]The Laboratory of Hematopoietic Cell Kinetics and The Laboratory of Cancer Genetics and Cytogenetics, Memorial Sloan-Kettering Cancer Center, 1275 York Avenue, New York, NY 10021

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ABSTRACT The consistent cytogenetic translocation of chronic myelogenous leukemia (the Philadelphia chromosome, Ph¹) has been observed in cells of multiple hematopoietic lineages. This translocation creates a chimeric gene composed of breakpoint-cluster-region (*bcr*) sequences from chromosome 22 fused to a portion of the *abl* oncogene on chromosome 9. The resulting gene product (P210^{c-abl}) resembles the transforming protein of the Abelson murine leukemia virus in its structure and tyrosine kinase activity. P210^{c-abl} is expressed in Ph¹-positive cell lines of myeloid lineage and in clinical specimens with myeloid predominance. We show here that Epstein-Barr virus-transformed B-lymphocyte lines that retain Ph¹ can express P210^{c-abl}. The level of expression in these B-cell lines is generally lower and more variable than that observed for myeloid lines. Protein expression is not related to amplification of the *abl* gene but to variation in the level of *bcr-abl* mRNA produced from a single Ph¹ template.

Chronic myelogenous leukemia (CML) is a disease of the pluripotent stem cell (1). In greater than 95% of patients, the leukemic cells contain the cytogenetic marker known as the Philadelphia chromosome, or Ph¹ (2). This reciprocal translocation event between the long arms of chromosomes 9 and 22 has been used as a disease-specific marker for diagnosis and evaluation of therapy. Multiple hematopoietic lineages, including myeloid and B-lymphoid, contain Ph¹ in early or chronic phase, as well as in the more acute accelerated and blast crisis phases of the disease.

One molecular consequence of Ph¹ is the translocation of the chromosomal arm containing the *c-abl* gene on chromosome 9 into the middle of the breakpoint-cluster region (*bcr*) gene on chromosome 22 (3-6). Although the precise translocation breakpoints are variable, an RNA-splicing mechanism generates a very similar 8-kilobase (kb) mRNA in each case (5-9). The hybrid *bcr-abl* message encodes a structurally altered form of the *abl* oncogene product, called P210^{c-abl} (10-13), with an amino-terminal segment derived from a portion of the exons of *bcr* on chromosome 22 and a carboxyl-terminal segment derived from a major portion of the exons of the *c-abl* gene on chromosome 9. The chimeric structure of *bcr-abl* and the resulting P210^{c-abl} is similar to the structure of the Abelson murine leukemia virus *gag-abl* genome and resulting P160^{v-abl} transforming gene product. Both proteins have very similar tyrosine kinase activities (10, 11, 14) which can be distinguished by their relative stability to denaturing detergents and by their ATP requirements from the recently described tyrosine kinase activity of the *c-abl* gene product (15).

In concert with structural modification of the amino-terminal portion of the *abl* gene, increased level of expression has been implicated in activation of *c-abl* oncogenic potential. Myeloid and erythroid cell lines and clinical samples derived from acute-phase CML patients contain about 10-fold higher levels of the 8-kb *bcr-abl* mRNA and P210^{c-abl} than the *c-abl* mRNA forms (6 and 7 kb) and P145^{c-abl} gene product (5, 8, 9, 11). The higher level of expression of the chimeric *bcr-abl* message in acute-phase cells is not likely to be solely due to the presence of the *bcr* promoter sequences at the 5' end of the gene, since the normal 4.5-kb and 6.7-kb *bcr*-encoded mRNA species are expressed at an even lower level than the normal *c-abl* messages (5, 6).

We have analyzed a series of Epstein-Barr virus-immortalized B-lymphoid cell lines derived from CML patients (16). With such *in vitro* clonal cell lines, we can evaluate whether the presence of Ph¹ always results in synthesis of the chimeric *bcr-abl* message and protein, and whether the quantitative expression varies for cells of B-lymphoid lineage as compared to previously examined myeloid cell lines. Our results show that cell lines that retain Ph¹ do express *bcr-abl* message and protein, but that the level is generally lower and more variable than previously seen for myeloid cell lines. The demonstration that the Ph¹ chromosomal template can vary in its level of expression of P210^{c-abl} suggests that secondary mechanisms, beyond the translocation itself, contribute to the regulation of the *bcr-abl* gene in different cell types or subclones that derive from the affected stem cell.

MATERIALS AND METHODS

Cells and Cell Labelings. Epstein-Barr virus-transformed B-lymphoid cell lines were established from peripheral blood samples of chronic- and acute-phase CML patients as reported (16). The cell lines are designated according to patient number, karyotype, and lineage. For example, SK-CML7Bt(9,22)-33 refers to CML patient 7, B-lymphoid cell line, 9;22 translocation (Ph¹), cell line 33; and SK-CML7BN-2 refers to B-cell line 2 with a normal karyotype derived from the same patient. Repeat karyotype analysis was performed to verify the retention of Ph¹ just prior to analysis for *abl* protein and RNA. Cells were maintained in RPMI 1640 medium with 20% fetal bovine serum. We have not observed any consistent pattern of *in vitro* growth rate that correlates to the stage of disease at the time of transformation with Epstein-Barr virus. Cells (1.5×10^7) were washed twice with Dulbecco's modified Eagle's medium lacking phosphate and

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Abbreviations: *bcr*, breakpoint-cluster region; CML, chronic myelogenous leukemia; kb, kilobase(s).

[†]Present address: Department of Genetics, University of Washington, Seattle, WA 98195.

[§]To whom correspondence should be addressed.

supplemented with 5% dialyzed fetal bovine serum. Cells were then resuspended in 2 ml of the minimal medium. Labeling was started with the addition of [32 P]orthophosphate (1 mCi/ml; ICN; 1 Ci = 37 GBq) and continued at 37°C for 3–4 hr.

Immunoprecipitation and Immunoblotting. Immunoprecipitations were carried out as described (10). Cells (1.5×10^7) were washed with phosphate-buffered saline and extracted with 3–5 ml of phosphate lysis buffer (1% Triton X-100/0.1 NaDodSO₄/0.5% deoxycholate/10 mM Na₂HPO₄, pH 7.5/100 mM NaCl) with 5 mM EDTA and 5 mM phenylmethylsulfonyl fluoride. Extracts were clarified by centrifugation and precipitated with normal or rabbit anti-*abl* sera (anti-pEX-2 or anti-pEX-5) (17). The precipitated proteins were electrophoresed in a NaDodSO₄/8% polyacrylamide gel. 32 P-labeled proteins were detected by autoradiography. Alternatively, *abl* proteins were detected by immunoblotting. Extracts from unlabeled cells were clarified, and proteins were concentrated by immunoprecipitation with rabbit antisera against *abl*-encoded proteins [anti-pEX-2 and anti-pEX-5 combined (17)] and then fractionated in 8% acrylamide gels. The proteins were transferred from the gel to nitrocellulose filters, using protease-facilitated transfer (18). The *abl*-encoded proteins were detected using murine monoclonal antibodies as a probe and peroxidase-conjugated goat anti-mouse second stage antibody (Bio-Rad) for development. Rabbit antisera and mouse monoclonal antibodies to *abl* proteins were prepared using bacterially expressed regions of the *v-abl* protein as immunogens (17, 19). Anti-pEX-2 antibodies react with the internal tyrosine kinase domain and anti-pEX-5 antibodies react with the carboxyl-terminal segment of the *abl* proteins.

RNA Analysis. RNA was extracted from 10^8 cells by the NaDodSO₄/urea/phenol method (20). Polyadenylated RNA was purified by oligo(dT) affinity chromatography. Samples were electrophoresed in a 1% agarose/formaldehyde gel and transferred to nitrocellulose. *abl* RNA species were detected by hybridization with a nick-translated *v-abl* fragment probe (21).

DNA Analysis. DNA was prepared from 5×10^7 cells of each cell line and processed for Southern blots with a *v-abl* probe as described (21).

RESULTS

Variable Levels of P210^{c-abl} Are Detected in Ph⁺-Positive Cell Lines. Ph⁺-positive and Ph⁺-negative, Epstein-Barr virus-transformed B-lymphocyte cell lines derived from the same patient were examined for P210^{c-abl} synthesis by immunoprecipitation of [32 P]orthophosphate-labeled cell extracts with anti-*abl* sera (Fig. 1). The normal *c-abl* protein P145^{c-abl} was detected at a similar level in multiple Ph⁺-positive and Ph⁺-negative cell lines. P210^{c-abl} was only detected in the Ph⁺-positive cell lines because the *bcr-abl* chimeric gene which encodes P210^{c-abl} resides on the Ph⁺ (4, 5, 11, 13). The level of P210^{c-abl} was about 4- to 5-fold higher than the level of P145^{c-abl} in the SK-CML7Bt-33 cell line (Fig. 1A, +). The Ph⁺-positive erythroid-progenitor cell line K562 (C) showed a level of P210^{c-abl} about 10-fold higher than P145^{c-abl}. However, the level of P210^{c-abl} was about one-fifth that of P145^{c-abl} in the Ph⁺-positive SK-CML16Bt-1 cell line (Fig. 1B, +). Comparison of different autoradiographic exposures roughly indicated that the level of P210^{c-abl} varies over a 20-fold range between these Ph⁺-positive B-cell lines. Analysis of four additional Ph⁺-positive B-cell lines demonstrated that the level of P210^{c-abl} fell into two general classes; some cell lines had a level of P210^{c-abl} similar to SK-CML7Bt-33 and others had the low level similar to SK-CML16Bt-1 (Table 1). This differs from previous studies with Ph⁺-positive myeloid cell lines and patient samples derived from acute-

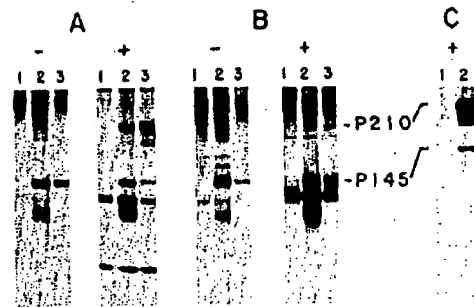


FIG. 1. Detection of variable levels of P210^{c-abl} in Ph⁺-positive B-cell lines. Production of P145^{c-abl} and P210^{c-abl} in Epstein-Barr virus-transformed B-cell lines derived from a blast-crisis (A) and a chronic-phase (B) CML patient was examined by metabolic labeling with [32 P]orthophosphate and immunoprecipitation. Ph⁺-negative (-) and Ph⁺-positive (+) cell lines derived from each patient were analyzed. The Ph⁺-negative cell line in A, - is SK-CML7Bt-2 and in B, - is SK-CML16Bt-1. The Ph⁺-positive cell line in A, + is SK-CML7Bt-33 and in B, + is SK-CML16Bt-1. The K562 cell line, a Ph⁺-positive erythroid progenitor cell line spontaneously derived from a blast-crisis patient (33), is represented in C. Cells (1.5×10^7) were metabolically labeled with 2 mCi of [32 P]orthophosphate for 3–4 hr and then were extracted and clarified by centrifugation. Samples were immunoprecipitated with control normal serum (lanes 1), anti-pEX-2 (lanes 2), or anti-pEX-5 (lanes 3) and analyzed by NaDodSO₄/8% PAGE followed by autoradiography with an intensifying screen (3 days for A and C, 10 days for B).

phase CML patients, in which P210^{c-abl} was detected at a 10-fold higher level than P145^{c-abl} (refs. 10 and 11; Table 1). There was no large difference in level of chimeric mRNA and P210^{c-abl} expressed in four myeloid/erythroid-lineage Ph⁺-positive cell lines (K562, EM2, EM3, CML22, and BV173; refs. 9 and 11), despite a 4- to 5-fold amplification of *abl*-related sequences in the K562 cell line.

Detection of different levels of P210^{c-abl} in Fig. 1 could be due to decreased phosphorylation of P210^{c-abl}, a lower level of P210^{c-abl} synthesis, or altered stability of the protein. To help distinguish among these possibilities, the steady-state level of P210^{c-abl} in the cell lines was assayed by immunoblotting. The results show that SK-CML7Bt-33 (Fig. 2A, +) had a higher level of P210^{c-abl} than P145, similar to the results with metabolic labeling (Fig. 1). We did not detect P210^{c-abl} by immunoblotting with 2×10^7 cells of line SK-CML8Bt-3 (Fig. 2B, +). Reconstruction experiments using dilutions of cell extracts showed that we could detect about 5–10% the level of P210^{c-abl} expressed in the K562 cell line (data not shown). We infer that the steady-state level of P210^{c-abl} in SK-CML8Bt-3 is lower than the level in SK-CML7Bt-33 by a factor of at least 10. The level of P210^{c-abl} detected in these assays correlated with the amount of P210^{c-abl} tyrosine kinase activity that could be detected *in vitro* (data not shown).

Different Levels of P210^{c-abl} Are Reflected in the Amount of Stable *bcr-abl* mRNA. To identify the basis for detection of variable levels of P210^{c-abl}, we examined the production of the *abl* RNA. RNA blot hybridization analysis using a *v-abl* probe (Fig. 3) showed that the normal 6- and 7-kb *c-abl* mRNAs were present at a similar level in Ph⁺-positive and -negative cell lines derived from different patients. However, the 8-kb mRNA that encodes P210^{c-abl} was detected at a 10-fold higher level in SK-CML7Bt-33 (Fig. 3A, +) than in SK-CML16Bt-1 (B, +), which correlated with the relative level of P210^{c-abl} detected in each cell line. Analysis of additional cell lines demonstrated that the level of 8-kb RNA directly correlated with the level of P210^{c-abl} (Table 1). The variation in level of 8-kb RNA detected in these cell lines was not due to loss or gain of Ph⁺, because cytogenetic analysis confirmed the presence of Ph⁺ in these cell lines (ref. 16 and

Table 1. Relative levels of *bcr-abl* expression in Epstein-Barr virus-immortalized B-cell lines and myeloid CML lines

| Cell line* | CML phase† | Ph ⁺ ‡ | P210§ | 8-kb mRNA¶ |
|--------------|------------|-------------------|-------|------------|
| SK-CML7BN-2 | BC | - | - | - |
| SK-CML8BN-10 | Chronic | - | - | - |
| SK-CML8BN-12 | Chronic | - | - | - |
| SK-CML16BN-1 | Chronic | - | - | - |
| SK-CML35BN-1 | Chronic | - | - | - |
| SK-CML7B5-33 | BC | + | +++ | +++ |
| SK-CML21Bt-1 | Acc | + | +++ | +++ |
| SK-CML21Bt-6 | Acc | + | +++ | +++ |
| SK-CML8Bt-3 | Chronic | + | + | ± |
| SK-CML16Bt-1 | Chronic | + | + | + |
| SK-CML35Bt-2 | Chronic | + | + | + |
| K562 | BC | + | +++++ | +++++ |
| BV173 | BC | + | +++++ | +++++ |
| EM2 | BC | + | +++++ | +++++ |

*Cell lines derived from CML patients by transformation with Epstein-Barr virus as described (16). Names of cell lines indicate patient number and Ph⁺ status: SK-CML7Bt indicates a cell line derived from patient 7 that carries the 9;22 Ph⁺ translocation; N indicates a normal karyotype. Myeloid-erythroid cell lines (K562, EM2, and BV173) are described in previous publications (9, 11, 22, 33).

†Status of patient at the time cell line was derived. BC, blast crisis; Acc, accelerated phase.

‡Presence (+) or absence (-) of Ph⁺ as demonstrated by karyotypic or Southern blot analysis.

§P210^{ab} detected as described in legend to Fig. 1. B-cell lines derived from blast-crisis and accelerated-phase patients had levels of P210 3- to 5-fold higher (++++) than levels of P145. Chronic-phase-derived cell lines had P210 levels lower than or just equivalent (+) to the level of P145. Myeloid and erythroid lines had levels of P210 5- to 10-fold higher than P145 (+++++).

¶Eight-kilobase *bcr-abl* mRNA detected as described in legend to Fig. 2. Symbols: ±, borderline detectable; +++++, level of 8-kb mRNA 5- to 10-fold higher than that of the 6- and 7-kb *c-abl* mRNA species; +++, level of 8-kb mRNA 3- to 5-fold higher than that of the 6- and 7-kb species; +, a level approximately equivalent to that of the 6- and 7-kb messages.

data not shown). There was no difference in the copy number of *abl*-related sequences as judged by Southern blot analysis (Fig. 4). Only the K562 cell line control showed an amplification of *abl* sequences, as previously reported (22, 23). These combined data suggest that differential *bcr-abl* mRNA expression from a single gene template is responsible for the variable levels of P210^{ab} detected. This could be mediated

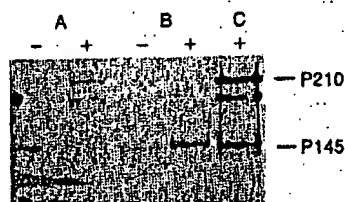


FIG. 2. Analysis of steady-state *abl* protein levels by immunoblotting. Cell extracts prepared from 2×10^7 cells of lines SK-CML7BN-2 (A, -), SK-CML7Bt-33 (A, +), SK-CML8BN-10 (B, -), and SK-CML8Bt-3 (B, +) were concentrated by immunoprecipitation with anti-pEX-2 plus anti-pEX-5. Samples were then electrophoresed in a NaDodSO₄/8% polyacrylamide gel and transferred to nitrocellulose, using protease-facilitated transfer (18). *abl* proteins were detected using a mixture of two monoclonal antibodies directed against the pEX-2 and pEX-5 *abl*-protein fragments produced in bacteria (19) as a probe and a peroxidase-conjugated goat anti-mouse second-stage antibody (Bio-Rad) for development.

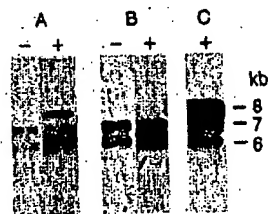


FIG. 3. Comparison of *abl* RNA levels in Ph⁺-positive and -negative B-cell lines. The levels of the normal 6- and 7-kb *c-abl* RNAs and the 8-kb *bcr-abl* RNA were analyzed by blot hybridization using a *v-abl* probe. RNA was extracted from Ph⁺-negative lines SK-CML7BN-2 (A, -) and SK-CML16BN-1 (B, -), from Ph⁺-positive lines SK-CML6Bt-33 (A, +) and SK-CML16Bt-3 (B, +), and from line K562 (C, +) by the NaDodSO₄/urea/phenol method (20). Polyadenylated RNA was purified by oligo(dT) affinity chromatography, and 15 µg of each sample was electrophoresed in a 1% agarose/formaldehyde gel and then transferred to nitrocellulose. The blotted RNAs were hybridized with a nick-translated *v-abl* fragment probe (21) and then autoradiographed for 4 days.

by factors influencing the transcription rate of the *bcr-abl* gene or the stability of the mRNA.

DISCUSSION

Several lines of evidence suggest that formation of Ph⁺ is not the primary event that affects the stem cell in CML. Patients have been identified that present with the clinical picture of CML but only later develop Ph⁺ (1). This observation, coupled with studies of *G6PD* (glucose-6-phosphate dehydrogenase)-heterozygous females with CML that demonstrate stem-cell clonality by isozyme analysis among cell

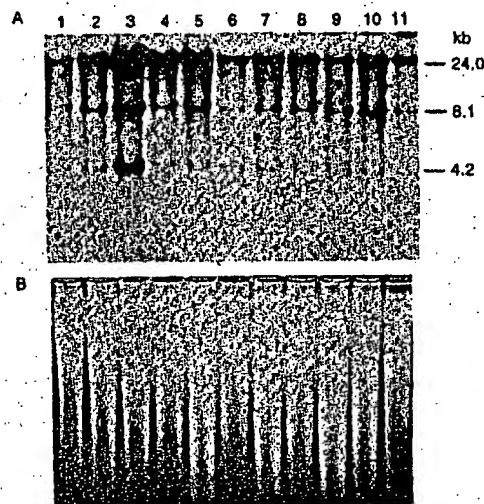


FIG. 4. Southern blot analysis of *abl* sequences in Ph⁺-positive and -negative B-cell lines. High molecular weight DNA (15 µg) was digested with restriction endonuclease *Bam*HI, separated in a 0.8% agarose gel, and then transferred to nitrocellulose. The blotted DNA fragments were hybridized with a nick-translated, 2.4-kb *Bgl* II *v-abl* fragment (1.5×10^8 cpm/µg; ref. 21) and exposed for 4 days. (A) Autoradiogram of *abl*-specific fragments in cell lines HL-60 (lane 1), EM2 (lane 2), K562 (lane 3), SK-CML7Bt-33 (lane 4), SK-CML8Bt-3 (lane 5), SK-CML16Bt-1 (lane 6), SK-CML21Bt-6 (lane 7), SK-CML35Bt-2 (lane 8), SK-CML7BN-2 (lane 9), SK-CML8BN-2 (lane 10), and SK-CML35BN-1 (lane 11). (B) Ethidium bromide staining of agarose gel prior to transfer to nitrocellulose, showing the level of variation in amount of DNA loaded per lane.

populations that lack the Ph¹ marker, supports a secondary or complementary role for Ph¹ in the progression of the disease (24, 25). This chromosome marker is found in chronic, accelerated, and blast-crisis phases of the disease. It is likely that Ph¹ confers some growth advantage, since cells with the marker chromosome eventually predominate the marrow and peripheral blood even in chronic phase. During the phase of blast crisis, many patients develop additional chromosome abnormalities, including duplication of Ph¹, a variety of trisomies, and complex translocations (26). This is suggestive evidence for Ph¹ being a necessary but not sufficient genetic change for the full evolution of the disease.

The realization that one molecular result of Ph¹ is the generation of a chimeric *bcr-abl* protein with functional characteristics and structure analogous to the *gag-abl* transforming protein of the Abelson murine leukemia virus strengthens the argument for an important role of Ph¹ in the pathogenesis of CML. Although the Abelson virus is generally considered a rapidly transforming retrovirus, its effects can range from overcoming growth factor requirements, to cellular lethality, to induction of highly oncogenic tumors in a number of hematopoietic cell lineages (27, 28). Even in the transformation of murine cell targets, there are several lines of evidence that suggest that the growth-promoting activity of the *v-abl* gene product is complemented by further cellular changes in the production of the malignant-cell phenotype (29–31).

The regulation of *bcr-abl* gene expression is complex because the 5' end of the gene is derived from the non-*abl* sequences, *bcr*, normally found on chromosome 22 (6). The level of stable message for the normal *bcr* gene and the normal *abl* gene are both much lower than the level of the *bcr-abl* message and protein from cell lines and clinical specimens derived from myeloid blast-crisis patients (5, 6, 11). Therefore, the high level of *bcr-abl* expression cannot simply be attributed to the regulatory sequences associated with *bcr*. Possibly, creation of the chimeric gene disrupts the normal regulatory sequences and results in a higher level of expression. Variation in *bcr-abl* expression may result from secondary changes in the structure of the chimeric gene or function of *trans*-acting factors that occur during evolution of the disease. Our analysis of P210^{c-abl} and the 8-kb mRNA in Epstein-Barr virus-transformed Ph¹-positive B-cell lines demonstrates that stable message and protein levels from the *bcr-abl* gene can vary over a wide range. This variation does not result from a change in the number of *bcr-abl* templates secondary to gene amplification but more likely from changes in either transcription rate or mRNA stability. We suspect this range of *bcr-abl* expression is not limited to lymphoid cells. Analysis of peripheral blood leukocytes derived from an unusual CML patient who has been in chronic phase with myeloid predominance for 16 years showed a level of P210^{c-abl} one-fifth that of P145^{c-abl}, as detected by metabolic labeling with [³²P]orthophosphate and immunoprecipitation (S.C., O.N.W., and P. Greenberg, unpublished observations). Lower levels of expression of the chimeric mRNA have been demonstrated in clinical samples from chronic-phase CML patients compared to acute-phase CML patients (9). Others have reported chronic-phase patients with variable but, in some cases, relatively high levels of the *bcr-abl* mRNA (32). The sampling variation and the heterogeneous mixture of cell types in clinical samples complicate such analyses. Further work is needed to evaluate whether there is a defined change in P210^{c-abl} expression during the progression of CML. It is interesting to note that among the limited sample of Ph¹-positive B-cell lines we have examined (Table 1), we have seen higher levels of P210^{c-abl} in those derived from patients at more advanced stages of the disease.

It will be important to search for cell-type-specific mechanisms that might regulate expression of *bcr-abl* from Ph¹.

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1. Champlin, R. E. & Golde, D. W. (1985) *Blood* 65, 1039–1047.
2. Rowley, J. D. (1973) *Nature (London)* 243, 290–291.
3. Heisterkamp, N., Stephenson, J. R., Groffen, J., Hansen, P. F., de Klein, A., Bartram, C. R. & Grosfeld, G. (1983) *Nature (London)* 306, 239–242.
4. Bartram, C. R., de Klein, A., Hagemeijer, A., van Agthoven, T., van Kessel, A. G., Bootsma, D., Grosfeld, G., Ferguson-Smith, M. A., Davies, T., Stone, M., Heisterkamp, N., Stephenson, J. R. & Groffen, J. (1983) *Nature (London)* 306, 277–280.
5. Shtivelman, E., Lifshitz, B., Gale, R. P. & Canaani, D. (1985) *Nature (London)* 315, 550–554.
6. Heisterkamp, N., Stam, K. & Groffen, J. (1985) *Nature (London)* 315, 758–761.
7. Groffen, J., Stephenson, J. R., Heisterkamp, N., de Klein, A., Bartram, C. R. & Grosfeld, G. (1984) *Cell* 36, 93–99.
8. Gale, R. P. & Canaani, E. (1984) *Proc. Natl. Acad. Sci. USA* 81, 5648–5652.
9. Collins, S., Kubonishi, L., Miyoshi, I. & Groudine, M. T. (1984) *Science* 225, 72–74.
10. Konopka, J. B., Watanabe, S. M. & Witte, O. N. (1984) *Cell* 7, 1035–1042.
11. Konopka, J. B., Watanabe, S. M., Singer, J., Collins, S. & Witte, O. N. (1985) *Proc. Natl. Acad. Sci. USA* 82, 1810–1814.
12. Klotzer, W., Kurzrock, R., Smith, L., Talpaz, M., Spiller, M., Gutterman, J. & Arlinghaus, R. (1985) *Virology* 140, 230–238.
13. Kozbor, D., Giallongo, A., Sierze, M. E., Konopka, J. B., Witte, O. N., Showe, L. C. & Croce, C. M. (1985) *Nature (London)*, in press.
14. Davis, R. L., Konopka, J. B. & Witte, O. N. (1985) *Mol. Cell Biol.* 5, 204–213.
15. Konopka, J. B. & Witte, O. N. (1985) *Mol. Cell Biol.* 5, 3116–3123.
16. Nitta, M., Kato, Y., Strife, A., Wachter, M., Fried, J., Perez, A., Jhanwar, S., Duigou, R., Chaganti, R. S. K. & Clarkson, B. (1985) *Blood* 66, 1053–1061.
17. Konopka, J. B., Davis, J. L., Watanabe, S. M., Ponticelli, A. S., Schiff-Maker, L., Rosenberg, N. & Witte, O. N. (1984) *Virology* 51, 223–232.
18. Gibson, W. (1981) *Anal. Biochem.* 118, 1–3.
19. Schiff-Maker, L., Konopka, J. B., Clark, S., Witte, O. N. & Rosenberg, N. (1986) *J. Virol.* 57, 1182–1186.
20. Schwartz, R. C., Sonenshein, G. E., Bothwell, A. & Gelfand, M. L. (1981) *J. Immunol.* 126, 2104–2108.
21. Goff, S. P., Gilboa, E., Witte, O. N. & Baltimore, D. (1980) *Cell* 22, 777–785.
22. Collins, S. J. & Groudine, M. T. (1983) *Proc. Natl. Acad. Sci. USA* 80, 4813–4817.
23. Selden, J. R., Emanuel, B. S., Wang, E., Cannizzaro, L., Palumbo, A., Erikson, J., Nowell, P. C., Rovera, G. & Croce, C. M. (1983) *Proc. Natl. Acad. Sci. USA* 80, 7289–7292.
24. Fialkow, P. J., Martin, P. J., Najfeld, V., Penfold, G. K., Jacobson, R. J. & Hansen, J. A. (1981) *Blood* 58, 158–163.
25. Martin, P. J., Najfeld, V. & Fialkow, P. J. (1982) *Can. Gen. Cytogenet.* 6, 359–368.
26. Rowley, J. D. (1980) *Annu. Rev. Genet.* 14, 17–40.
27. Whitlock, C. A. & Witte, O. N. (1984) *Adv. Immunol.* 37, 74–98.
28. Pierce, J. H., Di Fiore, P. P., Aaronson, S. A., Potter, M., Humphrey, J., Scott, A. & Ihle, J. N. (1985) *Cell* 41, 685–693.
29. Whitlock, C. A., Ziegler, S. & Witte, O. N. (1983) *Mol. Cell Biol.* 3, 596–604.
30. Wolf, D., Harris, N. & Rotter, V. (1984) *Cell* 38, 119–126.
31. Klein, G. & Klein, E. (1985) *Nature (London)* 315, 190–195.
32. Stam, K. Jr., Heisterkamp, N., Grosfeld, G., de Klein, A., Verma, R., Coleman, M., Dosik, H. & Groffen, J. (1985) *N. Engl. J. Med.* 313, 1429–1433.
33. Lozzio, C. B. & Lozzio, B. B. (1975) *Blood* 45, 321–334.

Review

Paul A. Haynes
Steven P. Cyp
Daniel Figeys
Ruedi Aebersold

Department of Molecular
Biotechnology, University of
Washington, Seattle, WA, USA

Proteome analysis: Biological assay or data archive?

In this review we examine the current state of proteome analysis. There are three main issues discussed: why it is necessary to study proteomes; how proteomes can be analyzed with current technology; and how proteome analysis can be used to enhance biological research. We conclude that proteome analysis is an essential tool in the understanding of regulated biological systems. Current technology, while still mostly limited to the more abundant proteins, enables the use of proteome analysis both to establish databases of proteins present, and to perform biological assays involving measurement of multiple variables. We believe that the utility of proteome analysis in future biological research will continue to be enhanced by further improvements in analytical technology.

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1 Introduction

A proteome has been defined as the protein complement expressed by the genome of an organism, or, in multicellular organisms, as the protein complement expressed by a tissue or differentiated cell [1]. In the most common implementation of proteome analysis the proteins extracted from the cell or tissue analyzed are separated by high

resolution two-dimensional gel electrophoresis (2-DE), detected in the gel and identified by their amino acid sequence. The ease, sensitivity and speed with which gel-separated proteins can be identified by the use of recently developed mass spectrometric techniques have dramatically increased the interest in proteome technology. One of the most attractive features of such analyses is that complex biological systems can potentially be studied in their entirety, rather than as a multitude of individual components. This makes it far easier to uncover the many complex, and often obscure, relationships between mature gene products in cells. Large-scale proteome characterization projects have been undertaken for a number of different organisms and cell types. Microbial proteome projects currently in progress include, for example: *Saccharomyces cerevisiae* [2], *Salmonella enterica* [3], *Spiroplasma melliferum* [4], *Mycobacterium tuberculosis* [5], *Ochrobactrum anthropi* [6], *Haemophilus influenzae* [7], *Synechocystis* spp. [8], *Escherichia coli* [9], *Rhizobium leguminosarum* [10], and *Dicystidium discoidum* [11]. Proteome projects underway for tissues of more complex organisms include those for: human bladder squamous cell carcinomas [12], human liver [13], human plasma [13], human keratinocytes [12], human fibroblasts [12], mouse kidney [12], and rat serum [14]. In this manuscript we critically assess the concept of proteome analysis and the technical feasibility of establishing complete proteome maps, and discuss ways in which proteome analysis and biological research intersect.

2 Rationale for proteome analysis

The dramatic growth in both the number of genome projects and the speed with which genome sequences are being determined has generated huge amounts of sequence information, for some species even complete genomic sequences ([15-17]). The description of the state of a biological system by the quantitative measurement of system components has long been a primary objective in molecular biology. With recent technical advances including the development of differential display-PCR [18], cDNA microarray and DNA chip technology [19, 20] and serial analysis of gene expression (SAGE) [21, 22], it is now feasible to establish global and quantitative mRNA expression maps of cells and tissues, in which the sequence of all the genes is known, at a speed and sensitivity which is not matched by current

Correspondence: Professor Ruedi Aebersold, Department of Molecular Biotechnology, University of Washington, Box 357730, Seattle, WA, 98195, USA (Tel: +206-616-4235; Fax: +206-616-6192; E-mail: ruedi@u.washington.edu)

Abbreviations: CID, collision-induced dissociation; MS/MS, tandem mass spectrometry; SAGE, serial analysis of gene expression

Keywords: Proteome / Two-dimensional polyacrylamide gel electrophoresis / Tandem mass spectrometry

protein analysis technology. Given the long-standing paradigm in biology that DNA synthesizes RNA which synthesizes protein, and the ability to rapidly establish global, quantitative mRNA expression maps, the questions which arise are why technically complex proteome projects should be undertaken and what specific types of information could be expected from proteome projects which cannot be obtained from genomic and transcript profiling projects. We see three main reasons for proteome analysis to become an essential component in the comprehensive analysis of biological systems. (i) Protein expression levels are not predictable from the mRNA expression levels, (ii) proteins are dynamically modified and processed in ways which are not necessarily apparent from the gene sequence, and (iii) proteomes are dynamic and reflect the state of a biological system.

2.1 Correlation between mRNA and protein expression levels

Interpretations of quantitative mRNA expression profiles frequently implicitly or explicitly assume that for specific genes the transcript levels are indicative of the levels of protein expression. As part of an ongoing study in our laboratory, we have determined the correlation of expression at the mRNA and protein levels for a population of selected genes in the yeast *Saccharomyces cerevisiae* growing at mid-log phase (S. P. Gygi *et al.*, submitted for publication): mRNA expression levels were calculated from published SAGE frequency tables [22]. Protein expression levels were quantified by metabolic radiolabeling of the yeast proteins, liquid scintillation counting of the protein spots separated by high resolution 2-DE and mass spectrometric identification of the protein(s) migrating to each spot. The selected 80 samples constitute a relatively homogeneous group with respect to predicted half-life and expression level of the protein products. Thus far, we have found a general trend but no strong correlation between protein and transcript levels (Fig. 1). For some genes studied equivalent mRNA transcript levels translated into protein abundances which varied by more than 50-fold. Similarly, equivalent steady-state protein expression levels were maintained by transcript levels varying by as much as 40-fold (S. P. Gygi *et al.*, submitted). These results suggest that even for a population of genes predicted to be relatively homogeneous with respect to protein half-life and gene expression, the protein levels cannot be accurately predicted from the level of the corresponding mRNA transcript.

2.2 Proteins are dynamically modified and processed

In the mature, biologically active form many proteins are post-translationally modified by glycosylation, phosphorylation, prenylation, acylation, ubiquitination or one or more of many other modifications [23] and many proteins are only functional if specifically associated or complexed with other molecules, including DNA, RNA, proteins and organic and inorganic cofactors. Frequently, modifications are dynamic and reversible and may alter the precise three-dimensional structure and the state of activity of a protein. Collectively, the state of modification of the proteins which constitute a biological system

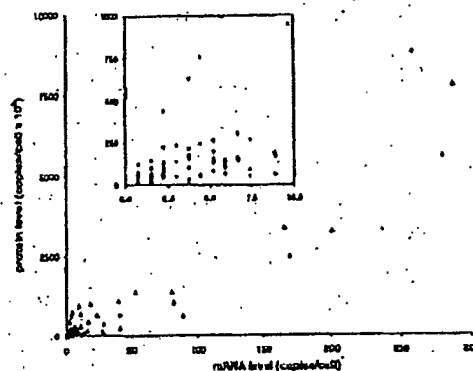


Figure 1. Correlation between mRNA and protein levels in yeast cells. For a selected population of 80 genes, protein levels were measured by ^{35}S -radiolabeling and mRNA levels were calculated from published SAGE tables. Inset: expanded view of the low abundance region. For more experimental details, also see Figs. 5 and 6, (S. P. Gygi *et al.*, submitted).

are important indicators for the state of the system. The type of protein modification and the sites modified at a specific cellular state can usually not be determined from the gene sequence alone.

2.3 Proteomes are dynamic and reflect the state of a biological system

A single genome can give rise to many qualitatively and quantitatively different proteomes. Specific stages of the cell cycle and states of differentiation, responses to growth and nutrient conditions, temperature and stress, and pathological conditions represent cellular states which are characterized by significantly different proteomes. The proteome, in principle, also reflects events that are under translational and post-translational control. It is therefore expected that proteomics will be able to provide the most precise and detailed molecular description of the state of a cell or tissue, provided that the external conditions defining the state are carefully determined. In answer to the question of whether the study of proteomes is necessary for the analysis of biomolecular systems, it is evident that the analysis of mature protein products in cells is essential as there are numerous levels of control of protein synthesis, degradation, processing and modification, which are only apparent by direct protein analysis.

3 Description and assessment of current proteome analysis technology

3.1 Technical requirements of proteome technology

In biological systems the level of expression as well as the states of modification, processing and macro-molecular association of proteins are controlled and modulated depending on the state of the system. Comprehensive analysis of the identity, quantity and state of modification of proteins therefore requires the detection and

quantitation of the proteins which constitute the system, and analysis of differentially processed forms. There are a number of inherent difficulties in protein analysis which complicate these tasks. First, proteins cannot be amplified. It is possible to produce large amounts of a particular protein by over-expression in specific cell systems. However, since many proteins are dynamically post-translationally modified, they cannot be easily amplified in the form in which they finally function in the biological system. It is frequently difficult to purify from the native source sufficient amounts of a protein for analysis. From a technological point of view this translates into the need for high sensitivity analytical techniques. Second, many proteins are modified and processed post-translationally. Therefore, in addition to the protein identity, the structural basis for differentially modified isoforms also needs to be determined. The distribution of a constant amount of protein over several differentially modified isoforms further reduces the amount of each species available for analysis. The complexity and dynamics of post-translational protein editing thus significantly complicates proteomic studies. Third, proteins vary dramatically with respect to their solubility in commonly used solvents. There are few, if any, solvent conditions in which all proteins are soluble and which are also compatible with protein analysis. This makes the development of protein purification methods particularly difficult since both protein purification and solubility have to be achieved under the same conditions. Detergents, in particular sodium dodecyl sulfate (SDS), are frequently added to aqueous solvents to maintain protein solubility. The compatibility with SDS is a big advantage of SDS polyacrylamide gel electrophoresis (SDS-PAGE) over other protein separation techniques. Thus, SDS-PAGE and two-dimensional gel electrophoresis, which also uses SDS and other detergents, are the most general and preferred methods for the purification of small amounts of proteins, provided that activity does not necessarily need to be maintained. Lastly, the number of proteins in a given cell system is typically in the thousands. Any attempt to identify and categorize all of these must use methods which are as rapid as possible to allow completion of the project within a reasonable time frame. Therefore, a successful, general proteomics technology requires high sensitivity, high throughput, the ability to differentiate differentially modified proteins, and the ability to quantitatively display and analyze all the proteins present in a sample.

3.2 2-D electrophoresis - mass spectrometry: a common implementation of proteomic analysis

The most common currently used implementation of proteomic analysis technology is based on the separation of proteins by two-dimensional (2-DE) gel electrophoresis and their subsequent identification and analysis by mass spectrometry (MS) or tandem mass spectrometry (MS/MS). In 2-DE, proteins are first separated by isoelectric focusing (IEF) and then by SDS-PAGE, in the second, perpendicular dimension. Separated proteins are visualized at high sensitivity by staining or autoradiography, producing two-dimensional arrays of proteins. 2-DE gels are, at present, the most commonly used means of global display of proteins in complex

samples. The separation of thousands of proteins has been achieved in a single gel [24, 25] and differentially modified proteins are frequently separated. Due to the compatibility of 2-DE with high concentrations of detergents, protein denaturants and other additives promoting protein solubility, the technique is widely used.

The second step of this type of proteomic analysis is the identification and analysis of separated proteins. Individual proteins from polyacrylamide gels have traditionally been identified using *N*-terminal sequencing [26, 27], internal peptide sequencing [28, 29], immunoblotting or comigration with known proteins [30]. The recent dramatic growth of large-scale genomic and expressed sequence tag (EST) sequence databases has resulted in a fundamental change in the way proteins are identified by their amino acid sequence. Rather than by the traditional methods described above, protein sequences are now frequently determined by correlating mass spectral or tandem mass spectral data of peptides derived from proteins, with the information contained in sequence databases [31-33].

There are a number of alternative approaches to proteomic analysis currently under development. There is considerable interest in developing a proteomic analysis strategy which bypasses 2-DE altogether, because it is considered a relatively slow and tedious process, and because of perceived difficulties in extracting proteins from the gel matrix for analysis. However, 2-DE as a starting point for proteomic analysis has many advantages compared to other techniques available today. The most significant strengths of the 2-DE-MS approach include the relatively uniform behavior of proteins in gels, the ability to quantify spots and the high resolution and simultaneous display of hundreds to thousands of proteins within a reasonable time frame.

A schematic diagram of a typical procedure of the identification of gel-separated proteins is shown in Fig. 2. Protein spots detected in the gel are enzymatically or chemically fragmented and the peptide fragments are isolated for analysis, as already indicated, most frequently by MS or MS/MS. There are numerous protocols for the generation of peptide fragments from gel-separated proteins. They can be grouped into two categories, digestion in the gel slice [28, 34] or digestion after electrotransfer out of the gel onto a suitable membrane [29, 35-37] and reviewed in [38]. In most instances either technique is applicable and yields good results. The analysis of MS or MS/MS data is an important step in the whole process because MS instruments can generate an enormous amount of information which cannot easily be managed manually. Recently, a number of groups have developed software systems dedicated to the use of peptide MS and MS/MS spectra for the identification of proteins. Proteins are identified by correlating the information contained in the MS spectra of protein digests or MS/MS spectra of individual peptides with data contained in DNA or protein sequence databases.

The systems we are currently using in our laboratory are based on the separation of the peptides contained in protein digests by narrow bore or capillary liquid chromatog-

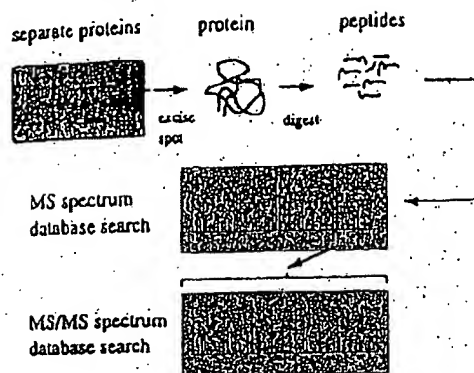


Figure 2. Schematic diagram of a procedure for identification of gel-separated proteins. Peptides can either be separated by a technique such as LC or CE, or infused as a mixture and sorted in the MS. Database searching can either be performed on peptide masses from an MS spectrum, peptide fragment masses from CID spectra of peptides, or a combination of both.

raphy [39, 40] or capillary electrophoresis [41], the analysis of the separated peptides by electrospray ionization (ESI) MS/MS, and the correlation of the generated peptide spectra with sequence databases using the SEQUEST program developed at the University of Washington [32, 33]. The system automatically performs the following operations: a particular peptide ion characterized by its mass-to-charge ratio is selected in the MS out of all the peptide ions present in the system at a particular time; the selected peptide ion is collided in a collision cell with argon (collision-induced dissociation, CID) and the masses of the resulting fragment ions are determined in the second sector of the tandem MS; this experimentally determined CID spectrum is then correlated with the CID spectra predicted from all the peptides in a sequence database which have essentially the same mass as the peptide selected for CID; this correlation matches the isolated peptide with a sequence segment in a database and thus identifies the protein from which the peptide was derived. There are a number of alternative programs which use peptide CID spectra for protein identification, but we use the SEQUEST system because it is currently the most highly automated program and has proven to be successful, versatile and robust.

3.3 Protein identification by LC-MS/MS, capillary LC-MS/MS and CE-MS/MS

It has been demonstrated repeatedly that MS has a very high intrinsic sensitivity. For the routine analysis of gel-separated proteins at high sensitivity, the most significant challenge is the handling of small amounts of sample. The crux of the problem is the extraction and transfer of peptide mixtures generated by the digestion of low nanogram amounts of protein, from gels into the MS/MS system without significant loss of sample or introduction of unwanted contaminants. We employ three different systems for introducing gel-purified samples into an MS, depending on the level of sensitivity

required. As an approximate guideline, for samples containing tens of picomoles of peptides, LC-MS/MS is most appropriate; for samples containing low picomole amounts to high femtomole amounts we use capillary LC-MS/MS; and for samples containing femtomoles or less, CE-MS/MS is the method of choice.

3.3.1 LC-MS/MS

The coupling of an MS to an HPLC system using a 0.5 mm diameter or bigger reverse phase (RP) column has been described in detail [42]. This system has several advantages if a large number of samples are to be analyzed and all are available in sufficient quantity. The LC-MS and database searching program can be run in a fully automated mode using an autosampler, thus maximizing sample throughput and minimizing the need for operator interference. The relatively large column is tolerant of high levels of impurities from either gel preparation or sample matrix. Lastly, if configured with a flow-splitter and micro-sprayer [40], analyses can be performed on a small fraction of the sample (less than 5%) while the remainder of the sample is recovered in very pure solvents. This latter feature is particularly useful when an orthogonal technique is also used to analyze peptide fractions, such as scintillation of an introduced radiolabel, and this data can be correlated with peptides identified by CID spectra.

3.3.2 Capillary LC-MS

An increase of sensitivity of approximately tenfold can be achieved by using a capillary LC system with a 100 μ m ID column rather than a 0.5 mm ID column as referred to above. Since very low flow rates are required for such columns, most reports have used a precolumn flow splitting system for producing solvent gradients. We have recently described the design and construction of a novel gradient mixing system which enables the formation of reproducible gradients at very low flow rates (low nL/min) without the need for flow splitting (A. Ducet *et al.*, submitted for publication). Using this capillary LC-MS/MS system we were able to identify gel-separated proteins if low picomole to high femtomole amounts were loaded onto the gel [40]. This system is as yet not automated and, like all capillary LC systems, is prone to blockage of the columns by microparticulates when analyzing gel-separated proteins.

3.3.3 CE-MS/MS

The highest level of sensitivity for analyzing gel-separated proteins can be achieved by using capillary electrophoresis – mass spectrometry (CE-MS). We have described in the past a solid-phase extraction capillary electrophoresis (SPE-CE) system which was used with triple quadrupole and ion trap ESI-MS/MS systems for the identification of proteins at the low femtomole to sub-femtomole sensitivity level [43, 44]. While this system is highly sensitive, its operation is labor-intensive and its operation has not been automated. In order to devise an analytical system with both the sensitivity of a CE and the level of automation of LC, we have constructed

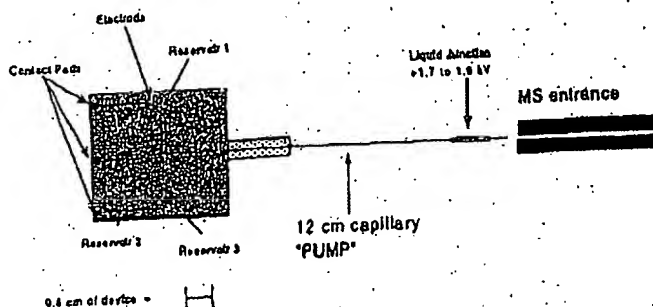


Figure 3. Schematic illustration of a microfabricated analytical system for CE, consisting of a micromachined device, coated capillary electroosmotic pump, and microelectrospray interface. The dimensions of the channels and reservoirs are as indicated in the text. The channels on the device were graphically enhanced to make them more visible. Reproduced from [45], with permission.

microfabricated devices for the introduction of samples into ESI-MS for high-sensitivity peptide analysis.

The basic device is a piece of glass, into which channels of 10–30 μm in depth and 50–70 μm in diameter are etched by using photolithography/etching techniques similar to the ones used in the semiconductor industry. (A simple device is shown in Fig. 3). The channels are connected to an external high voltage power supply [45]. Samples are manipulated on the device and off the device to the MS by applying different potentials to the reservoirs. This creates a solvent flow by electroosmotic pumping which can be redirected by changing the position of the electrode. Therefore, without the need for valves or gates and without any external pumping, the flow can be redirected by simply switching the position of the electrodes on the device. The direction and rate of the flow can be modulated by the size and the polarity of the electric field applied and also by the charge state of the surface.

The type of data generated by the system is illustrated in Fig. 4, which shows the mass spectrum of a peptide sample representing the tryptic digest of carbonic anhydrase at 290 fmol/ μL . Each numbered peak indicates a peptide successfully identified as being derived from carbonic an-

hydrase. Some of the unassigned signals may be chemical or peptide contaminants. The MS is programmed to automatically select each peak and subject the peptide to CID. The resulting CID spectra are then used to identify the protein by correlation with sequence databases. Therefore, this system allows us to concurrently apply a number of protein digests onto the device, to sequentially mobilize the samples, to automatically generate CID spectra of selected peptide ions and to search sequence databases for protein identification. These steps are performed automatically without the need for user input and proteins can be identified at very low femtomole level sensitivity at a rate of approximately one protein per 15 min.

3.4 Assessment of 2-DE-MS proteome technology

Using a combination of the analytical techniques described above we have identified the 80 protein spots indicated in Fig. 5. The protein pattern was generated by separating a total of 40 microgram of protein contained in a total cell lysate of the yeast strain YPH499 by high resolution 2-DE and silver staining of the separated proteins. To estimate how far this type of proteome analysis can penetrate towards the identification of low abundance proteins, we have calculated the codon bias of the genes encoding the respective proteins. Codon bias is a

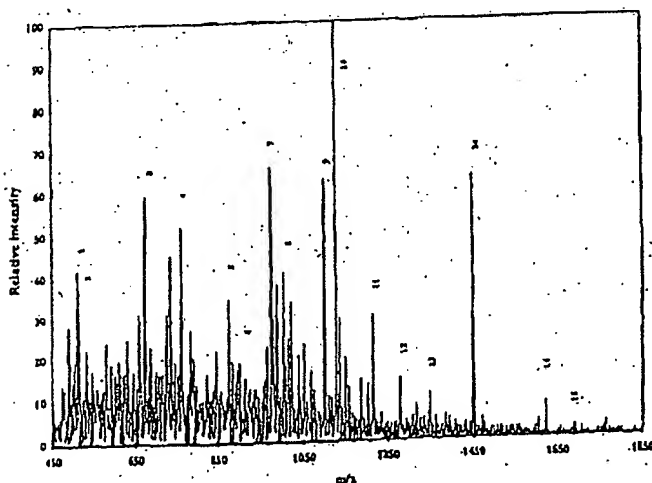


Figure 4. MS spectrum of a tryptic digest of carbonic anhydrase using the microfabricated system shown in Fig. 3. 290 fmol/ μL of carbonic anhydrase tryptic digest was infused into a Finnigan LCQ ion trap MS. Each peak was selected for CID, and those which were identified as containing peptides derived from carbonic anhydrase are numbered. Reproduced from [45], with permission.

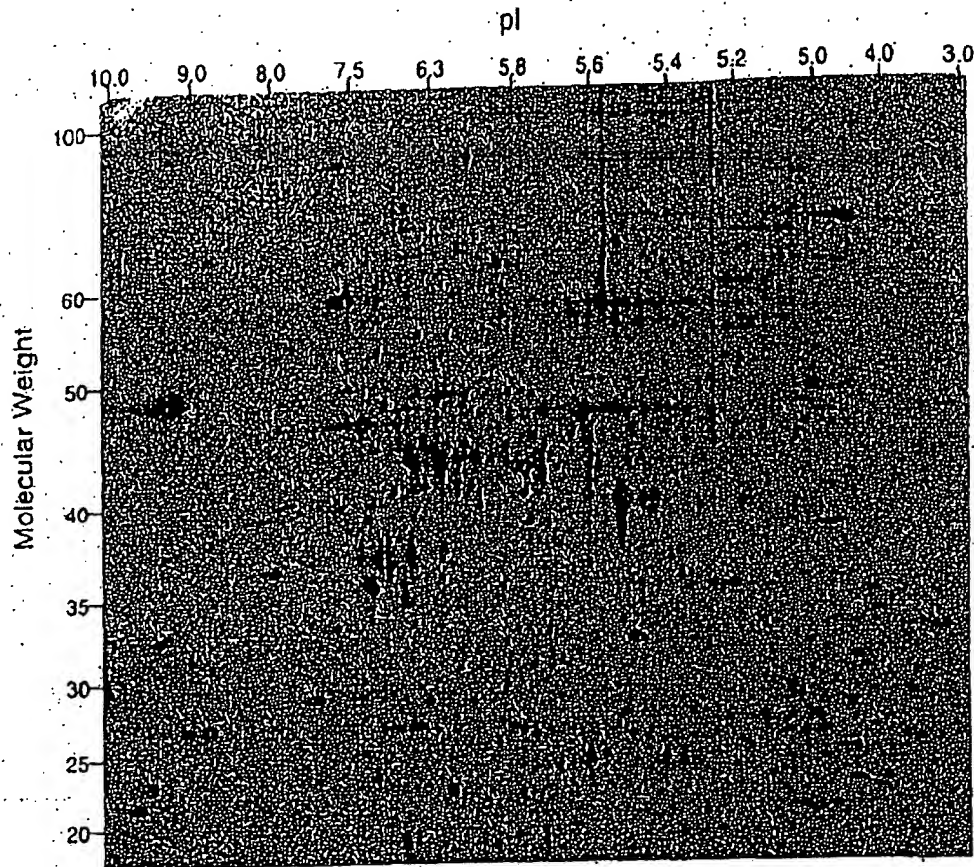


Figure 5. 2-DE separation of a lysate of yeast cells, with identified proteins highlighted. The first dimension of separation was an IFO (from pH 3-10), and the second dimension was a 10%T-SDS-PAGE gel. Proteins were visualized by silver staining. Further details of experimental procedures are included in S. P. Ouyi *et al.* (submitted).

calculated measure of the degree of redundancy of triplet DNA codons used to produce each amino acid in a particular gene sequence. It has been shown to be a useful indicator of the level of the protein product of a particular gene sequence present in a cell [46]. The general rule which applies is that the higher the value of the codon bias calculated for a gene, the more abundant the protein product of that gene becomes. The calculated codon bias values corresponding to the proteins identified in Fig. 5 are shown in Fig. 6b. Nearly all of the proteins identified (> 95%) have codon bias values of > 0.2, indicating they are highly abundant in cells. In contrast, codon bias values calculated for the entire yeast genome (Fig. 6a) show that the majority of proteins present in the proteome have a codon bias of < 0.2 and are thus of low abundance.

This finding is of considerable importance in our assessment of the current status of proteome analysis technology. It is clear that even using highly sensitive analytical techniques, we are only able to visualize and identify the

more abundant proteins. Since many important regulatory proteins are present only at low abundance, these would not be amenable to analysis using such techniques. This situation would be exacerbated in the analysis of proteomes containing many more proteins than the approximately 6000 gene products present in yeast cells [16]. In the analysis of, for example, the proteome of any human cells, there are potentially 50 000-100 000 gene products [47]. Inherent limitations on the amount of protein that can be loaded on 2-DE, and the number of components that can be resolved, indicate that only the most highly abundant fraction of the many gene products could be successfully analyzed. One approach that has been employed to circumvent these limitations is the use of very narrow range immobilized pH gradient strips for the first-dimension separation of 2-DE [48]. Since only those proteins which focus within the narrow range will enter the second dimension of separation, a much higher sample loading within the desired range is possible. This, in turn, can lead to the visualization and identification of less abundant proteins.

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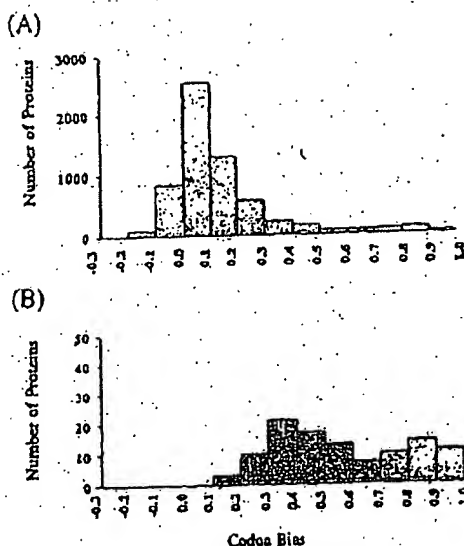


Figure 6. Calculated codon bias values for yeast proteins. (A) Distribution of calculated values for the entire yeast proteome. (B) Distribution of calculated values for the subset of 50 identified proteins also shown in Figs. 1 and 5. Further details of experimental procedures are included in S. P. Orgz et al. (submitted).

4 Utility of proteome analysis for biological research

For the success of proteomics as a mainstream approach to the analysis of biological systems it is essential to define how proteome analysis and biological research projects intersect. Without a clear plan for the implementation of proteome-type approaches into biological research projects the full impact of the technology can not be realized. The literature indicates that proteome analysis is used both as a database/data archive, and as a biological assay or biological research tool.

4.1 The proteome as a database

The use of proteomics as a database or data archive essentially entails an attempt to identify all the proteins in a cell or species and to annotate each protein with the known biological information that is relevant for each protein. The level of annotation can, of course, be extensive. The most common implementation of this idea is the separation of proteins by high resolution 2-DE, the identification of each detected protein spot and the annotation of the protein spots in a 2-DE gel database format. This approach is complicated by the fact that it is difficult to precisely define a proteome and to decide which proteome should be represented in the database. In contrast to the genome of a species, which is essentially static, the proteome is highly dynamic. Processes such as differentiation, cell activation and disease can all significantly change the proteome of a species. This is illustrated in Fig. 7. The figure shows two high-resolu-

tion 2-DE maps of proteins isolated from rat serum. Fig. 7A is from the serum of normal rats, while Fig. 7B is from the serum of rats in acute-phase serum after prior treatment with an inflammation-causing agent [49]. It is obvious that the protein patterns are significantly different in several areas, raising the question of exactly which proteome is being described.

Therefore, a comprehensive proteome database of a species or cell type needs to contain all of the parameters which describe the state and the type of the cells from which the proteins were extracted as well as the software tools to search the database with queries which reflect the dynamics of biological systems. A comprehensive proteome database should be capable of quantitatively describing the fate of each protein if specific systems and pathways are activated in the cell. Specifically, the quantity, the degree of modification, the subcellular location and the nature of molecules specifically interacting with a protein as well as the rate of change of these variables should be described. Using these admittedly stringent criteria, there is currently no complete proteome database. A number of such databases are, however, in the process of being constructed. The most advanced among them, in our opinion, are the yeast protein database YPD [50] (accessible at <http://www.ypd.com>) and the human 2D-PAGE databases of the Danish Centre for Human Genome Research [12] (accessible at <http://biobase.dk/cgi-bin/celis>). While neither can be considered complete as not all of the potential gene products are identified, both contain extensive annotation of supplemental information for many of the spots which are positively identified in reference samples.

4.2 The proteome as a biological assay

The use of proteome analysis as a biological assay or research tool represents an alternative approach to integrating biology with proteomics. To investigate the state of a system, samples are subjected to a specific process that allows the quantitative or qualitative measurement of some of the variables which describe the system. In typical biochemical assays one variable (e.g., enzyme activity) of a single component (e.g., a particular enzyme) is measured. Using proteomics as an assay, multiple variables (e.g., expression level, rate of synthesis, phosphorylation state, etc.) are measured concurrently on many (ideally all) of the proteins in a sample. The use of proteomics as an assay is a less far-reaching proposition than the construction of a comprehensive proteome database. It does, however, represent a pragmatic approach which can be adapted to investigate specific systems and pathways, as long as the interpretation of the results takes into account that with current technology not all of the variables which describe the system can be observed (see Section 3.4).

A common implementation of proteome analysis as a biological assay is when a 2-DE protein pattern generated from the analysis of an experimental sample is compared to an array of reference patterns representing different states of the system under investigation. The state of the experimental system at the time the sample was generated is therefore determined by the quantita-

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tive comparative analysis of hundreds to a few thousand proteins. Comparative analysis of the 2-DE patterns further highlights quantitative and qualitative differences in the protein profiles which correlate with the state of the system. For this type of analysis it is not essential that all the proteins are identified or even visu-

alized, although the results become more informative as more proteins are compared. It is obvious, however, that the possibility to identify any protein deemed characteristic for a particular state dramatically enhances this approach by opening up new avenues for experimentation.

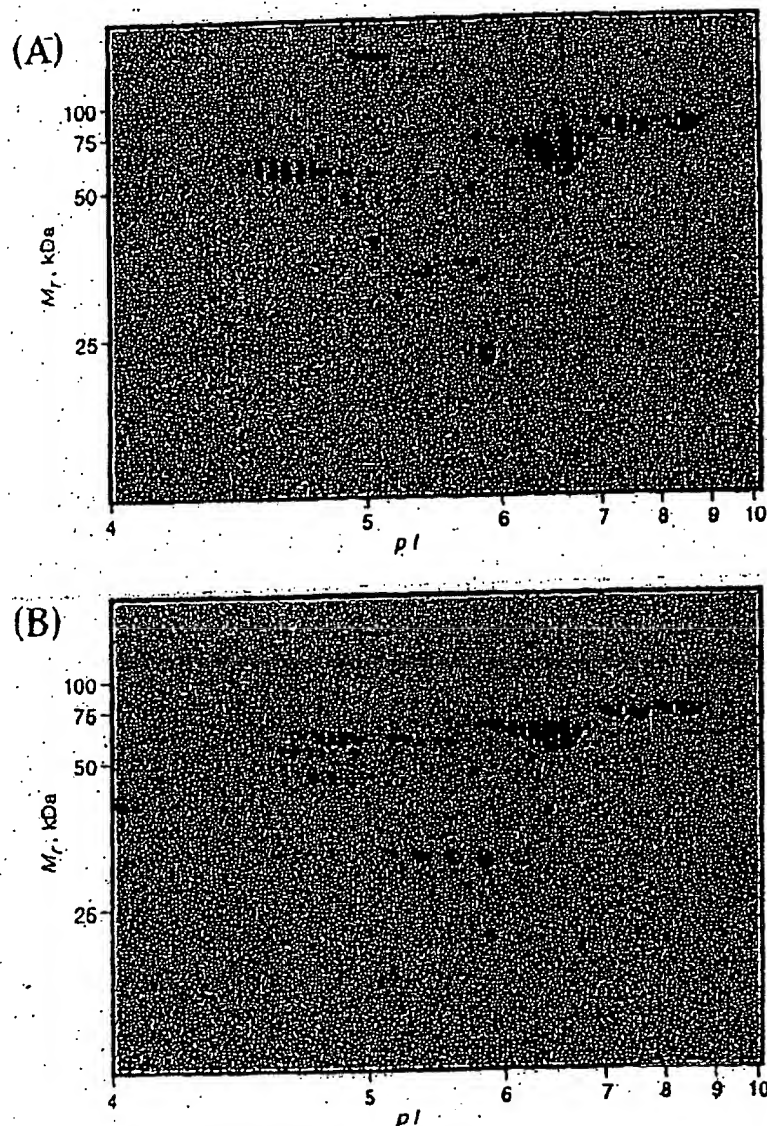


Figure 2. High resolution 2-DE map of proteins isolated from rat serum with or without prior exposure to an inflammation-causing agent. (A) normal rat serum, (B) acute-phase serum from rats which had previously been exposed to an inflammation-causing agent. The first dimension of separation is an IPG from pH 4-10, and the second dimension is a 7.5-17.5%T gradient SDS-PAGE gel. Proteins were visualized by staining with amido black. Further details of experimental procedures are included in [14, 49].

Proteome analysis as a biological assay has been successfully used in the field of toxicology, to characterize disease states or to study differential activation of cells. The approach is limited, of course, by the fact that only the visible protein spots are included in the assay; and it is well known that a substantial but far from complete fraction of cellular proteins are detected if a total cell lysate is separated by 2-DE. Proteins may not be detected in 2-DE gels because they are not abundant enough to be visualized by the detection method used, because they do not migrate within the boundaries (size, pI) resolved by the gel, because they are not soluble under the conditions used, or for other reasons.

A different way to use proteome analysis as a biological assay to define the state of a biological system is to take advantage of the wealth of information contained in 2-DE protein patterns. 2-DE is referred to as two-dimensional because of the electrophoretic mobility and the isoelectric points which define the position of each protein in a 2-DE pattern. In addition to the two dimensions used to generate the protein patterns, a number of additional data dimensions are contained in the protein patterns. Some of these dimensions such as protein expression level, phosphorylation state, subcellular location, association with other proteins, rate of synthesis or degradation indicate the activity state of a protein or a biological system. Comparative analysis of 2-DE protein patterns representing different states is therefore ideally suited for the detection, identification and analysis of suitable markers. Once again it must be emphasized that in this type of experiment only a fraction of the cellular proteins is analyzed. Since many regulatory proteins are of low abundance, this limitation is a concern, particularly in cases in which regulatory pathways are being investigated.

5 Concluding remarks

In this report we have addressed three main issues related to proteome analysis. First, we have discussed the rationale for studying proteomes. Second, we have assessed the technical feasibility of analyzing proteomes and described current proteome technology, and third, we have analyzed the utility of proteome analysis for biological research. It is apparent that proteome analysis is an essential tool in the analysis of biological systems. The multi-level control of protein synthesis and degradation in cells means that only the direct analysis of mature protein products can reveal their correct identities, their relevant state of modification and/or association and their amounts. Recently developed methods have enabled the identification of proteins at ever-increasing sensitivity levels and at a high level of automation of the analytical processes. A number of technical challenges, however, remain. While it is currently possible to identify essentially any protein spots that can be visualized by common staining methods, it is apparent that without prior enrichment only a relatively small and highly selected population of long-lived, highly expressed proteins is observed. There are many more proteins in a given cell which are not visualized by such methods. Frequently it is the low abundance proteins that execute key regulatory functions.

We have outlined the two principal ways proteome analysis is currently being used to intersect with biological research projects: the proteome as a database or data archive and proteome analysis as a biological assay. Both approaches have in common that at present they are conceptually and technically limited. Current proteome databases typically are limited to one cell type and one state of a cell and therefore do not account for the dynamics of biological systems. The use of proteome analysis as a biological assay can provide a wealth of information, but it is limited to the proteins detected and is therefore not truly proteome-wide. These limitations in proteomics are to a large extent a reflection of the fact that proteins in their fully processed form cannot easily be amplified and are therefore difficult to isolate in amounts sufficient for analysis or experimentation. The fact that to date no complete proteome has been described further attests to these difficulties. With continued rapid progress in protein analysis technology, however, we anticipate that the goal of complete proteome analysis will eventually become attainable.

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6 References

- [1] Wilkins, M. R., Pasquelli, C., Appel, R. D., Ou, K., O'Neil, O., Sanchez, J.-C., Yan, J. X., Goolley, A. A., Hughes, O., Humphrey-Smith, I., Williams, K. L., Hochstrasser, D. P., *BioTechnology* 1996, 14, 61-65.
- [2] Hodges, P. E., Payne, W. E., Garrels, J. I., *Nucleic Acids Res.* 1991, 19, 68-72.
- [3] O'Connor, C. D., Farris, M., Fyfe, R., Qi, S. Y., *Electrophoresis* 1997, 18, 1483-1490.
- [4] Cordwell, S. J., Bancel, D. J., Humphrey-Smith, I., *Electrophoresis* 1997, 18, 1335-1346.
- [5] Urquhart, B. L., Alsalas, T. R., Roach, D., Bancel, D. J., Bjellqvist, B., Britton, W. L., Humphrey-Smith, I., *Electrophoresis* 1997, 18, 1384-1392.
- [6] Wajsbom, V. C., Bjellqvist, B., Humphrey-Smith, I., *Electrophoresis* 1997, 18, 1333-1343.
- [7] Link, A. J., Hays, L. G., Carmack, E. B., Yates III, J. R., *Electrophoresis* 1997, 18, 1314-1324.
- [8] Suzuki, T., Ohara, O., *Electrophoresis* 1997, 18, 1252-1258.
- [9] VanBogelen, R. A., Abshire, K. Z., Molkover, B., Olson, B. R., Neidhardt, F. C., *Electrophoresis* 1997, 18, 1243-1251.
- [10] Overholo, M., Redmond, J. W., Rolfe, B. O., Djordjevic, M. A., *Mol. Plant Microbe Interact.* 1997, 10, 506-516.
- [11] Yan, J. X., Tonello, L., Sanchez, J.-C., Wilkins, M. R., Packer, N. H., Goolley, A. A., Hochstrasser, D. P., Williams, K. L., *Electrophoresis* 1997, 18, 491-497.
- [12] Celis, J., Oromor, P., Ostergaard, M., Madson, P., Honore, B., DeGaard, K., Olsen, E., Varum, H., Kristensen, D. B., Gromova, I., Haunsbo, A., Van Damme, J., Puyse, M., Vandekerckhove, J., Rasmussen, H. H., *FEBS Lett.* 1996, 398, 129-134.
- [13] Appel, R. D., Sanchez, J.-C., Baiocchi, A., O'Neil, O., Miu, M., Vargas, J. R., Hochstrasser, D. P., *Electrophoresis* 1993, 14, 1223-1231.
- [14] Haynes, P., Miller, I., Aebbersold, R., Gemelner, M., Eberlin, I., Lovat, R. M., Manzoni, C., Vignati, M., Gianazza, E., *Electrophoresis* 1998, 19, 1484-1492.

- [15] Fleischmann, R. D., Adams, M. D., White, O., Clayton, R. A., Kirkness, E. F., Kerlavage, A. R., Bult, C. J., Tomb, J.-P., Dougherty, B. A., Merrick, J. M., McKenney, K., Sutton, G., Fink, H., W. Fields, C., Gocayne, J. D., Scott, J., Shiley, R., Liu, L.-L., Glodet, A., Kelley, J. M., Weidman, J. P., Phillips, C. A., Spriggs, T., Hedblom, E., Cotton, M. D., Utterback, T. R., Hannu, N. C., Nguyen, D. T., Sauter, D. M., Brandon, R. C., Fink, L. D., Filichkin, J. L., Fuhrmann, J. L., Geoghegan, N. S. M., Osocher, C. L., McDonald, L. A., Small, K. V., Fraser, C. M., Smith, C. O., Venter, J. C., *Science* 1993, 269, 496-512.
- [16] Ouellet, A., Barrell, B. G., Boushey, H., Davis, R. W., Dujon, B., Feldman, M., Galibert, F., Hohlisch, J. D., Jacq, C., Johnston, M., Louis, B. J., McEwen, H. W., Murakami, Y., Philippon, P., Telford, H., Ollivier, S. O., *Science* 1996, 274, 546.
- [17] Fraser, C. M., Casjens, S., Huang, W. M., Sutton, O. O., Clayton, R., Lathigis, R., White, O., Kerlavage, A. R., Dodson, R., Hickey, E. K., O'Connell, M., Dougherty, B., Tomb, J. P., Fleischmann, R. D., Richardson, D., Peterson, J., Kerlavage, A. R., Quackenbush, J., Salzberg, S., Hanson, M., van Vugt, R., Palmer, M., Adams, M. D., Gocayne, J., Weidman, J., Utterback, T., Wathley, T., McDonald, L., Artlich, P., Bowman, C., Oatland, S., Fujii, C., Cotton, M. D., Horst, K., Roberts, K., Hatch, B., Smith, H. O., Venter, J. C., *Nature* 1997, 390, 580-586.
- [18] Liang, P., Pardee, A. B., *Science* 1993, 257, 967-971.
- [19] Lashkari, D. A., DeRisi, J. L., McCusker, J. H., Namath, A. P., Gentile, C., Huang, S. Y., Brown, P. O., Davis, R. W., *Proc. Natl. Acad. Sci. USA* 1997, 94, 13057-13062.
- [20] Shaloo, D., Smith, S. J., Brown, P. O., *Genome Res.* 1996, 6, 639-645.
- [21] Velculescu, V. E., Zhang, L., Vogelstein, B., Kinzler, K. W., *Science* 1995, 270, 484-487.
- [22] Velculescu, V. E., Zhang, L., Zhou, W., Vogelstein, B., Basrai, M. A., Bassett, D. B., Miter, P., Vogelstein, B., Kinzler, K. W., *Cell* 1997, 84, 243-251.
- [23] Krabb, R. O., Wolf, P., *Adv. Enzymol.* 1993, 67, 265-298.
- [24] Gorg, A., Postel, W., Gunther, S., *Electrophoresis* 1988, 9, 531-546.
- [25] Klose, J., Kobalz, U., *Electrophoresis* 1995, 16, 1034-1059.
- [26] Matuszewska, P., *J. Biol. Chem.* 1987, 262, 10035-10038.
- [27] Aebersold, R. H., Toplow, D. B., Hood, L. E., Kent, S. B., *J. Biol. Chem.* 1986, 261, 4229-4231.
- [28] Rosenfeld, J., Capdeville, J., Guilleminot, J. C., Ferran, P., *Anal. Biochem.* 1992, 203, 173-179.
- [29] Aebersold, R. H., Leavitt, J., Saavedra, R. A., Hood, L. E., Kent, S. B., *Proc. Natl. Acad. Sci. USA* 1987, 84, 6970-6974.
- [30] Honori, B., Löffler, H., Madson, P., Celis, J. E., *Eur. J. Biochem.* 1993, 218, 421-430.
- [31] Mann, M., Wilm, M., *Anal. Chem.* 1994, 66, 4390-4399.
- [32] Eng, J., McCormack, A. L., Yates III, J. R., *J. Amer. Mass Spectrom.* 1994, 5, 976-989.
- [33] Yates III, J. R., Eng, J. K., McCormack, A. L., Schlicht, D., *Anal. Chem.* 1995, 67, 1426-1436.
- [34] Shevchenko, A., Wilm, M., Vorm, O., Mann, M., *Anal. Chem.* 1996, 68, 810-818.
- [35] Hertz, D., Covey, T. C., Wink, R., Brownsey, R. W., Aebersold, R., *Protein Sci.* 1993, 2, 1342-1351.
- [36] van Oostveen, L., Ducret, A., Aebersold, R., *Anal. Biochem.* 1997, 247, 310-318.
- [37] Lul, M., Tempst, P., Erdjument-Bromage, H., *Anal. Biochem.* 1996, 241, 156-166.
- [38] Patterson, S. D., Aebersold, R. A., *Electrophoresis* 1995, 16, 1791-1814.
- [39] Ducret, A., Foy, Bruno, C., Bures, B. J., Marhaug, G., Husby, O. R. A., *Electrophoresis* 1996, 17, 866-876.
- [40] Haynes, P. A., Phipps, N., Aebersold, R., *Electrophoresis* 1998, 19, 939-945.
- [41] Figge, D., Van Oostveen, L., Ducret, A., Aebersold, R., *Anal. Chem.* 1996, 68, 1822-1828.
- [42] Ducret, A., Van Oostveen, L., Eng, J. K., Yates III, J. R., Aebersold, R., *Protein Sci.* 1997, 7, 796-799.
- [43] Figge, D., Ducret, A., Yates III, J. R., Aebersold, R., *Nature Biotech.* 1996, 14, 1579-1583.
- [44] Figge, D., Aebersold, R., *Electrophoresis* 1997, 18, 360-364.
- [45] Figge, D., Ning, Y., Aebersold, R., *Anal. Chem.* 1997, 69, 3150-3160.
- [46] Garrels, J. I., McLaughlin, C. S., Warner, J. R., Fletcher, B., Laiter, O. I., Kobayashi, R., Schwender, B., Volpe, T., Anderson, D. S., Mesquita-Puentes, R., Payne, W. B., *Electrophoresis* 1997, 18, 1347-1360.
- [47] Schuler, G. D., Boguski, M. S., Stewart, E. A., Stein, L. D., Gyapay, G., Rice, K., White, R. E., Rodriguez-Tome, P., Aggarwal, A., Bajorek, B., Beato, S., Birge, B. B., Butler, A., Castle, A. B., Chishti-Khushf, N., Chu, A., Clea, C., Cowles, S., Day, P. J., Dibling, T., Drouot, N., Dunham, I., Duprat, S., Edwards, C., Fan, J.-B., Fang, N., Frazee, C., Garrett, C., Green, L., Hadley, D., Harris, M., Harrison, P., Brady, S., Hicks, A., Holloway, E., Hul, L., Hussala, S., Louis-Dit-Sully, C., Ma, J., MacOlivier, A., Mader, C., Murakami, A., Mathis, T. C., McKusick, K. B., Morissette, J., Mungall, A., Muscat, D., Musbaum, H. C., Page, D. C., Peck, A., Perkins, S., Piercy, M., Qiu, F., Quackenbush, J., Ranby, S., Reif, T., Rosen, S., Sanders, X., She, X., Silva, J., Slosser, D. K., Soderlund, C., Sun, W.-L., Tabak, P., Thangarajah, T., Vega-Czarny, N., Vollrath, D., Voyticky, S., Wilmer, T., Wu, X., Adams, M. D., Ailiff, C., Walter, N. A., R., Brändén, R., Doherty, A., Goodfellow, P. N., Houlgate, R., Hudson, J. R., Jr., Ito, S. B., Iorio, K. R., Lee, W. Y., Saki, N., Nagata, T., Ishikawa, K., Momma, N., Phillips, C., Polymenopoulos, M. H., Sandusky, M., Schmitt, K., Berry, R., Swanson, K., Torres, R., Venter, J. C., Sikela, J. M., Beckmann, J. S., Weisenbach, J., Myers, R. M., Cox, D. R., James, M. R., Bentley, D., *et al. Science* 1996, 274, 540-546.
- [48] Sánchez, J.-C., Rouge, V., Fritsch, M., Ravet, P., Tonello, L., Moormayer, M., Wilkins, M. R., Hochstrasser, D. P., *Electrophoresis* 1997, 18, 324-327.
- [49] Miller, L., Haynes, P., Gmeliner, M., Aebersold, R., Manzoni, C., Loyall, M. R., Vignati, M., Eberlin, I., Gianazza, E., *Electrophoresis* 1998, 19, 1493-1500.
- [50] Garrels, J. I., *Nucleic Acids Res.* 1996, 24, 46-49.

Analysis of Genomic and Proteomic Data Using Advanced Literature Mining

Yanhui Hu, Lisa M. Hines, Haifeng Weng, Dongmei Zuo, Miguel Rivera,
Andrea Richardson, and Joshua LaBaer*

Institute of Proteomics, Harvard Medical School—BCMP, 240 Longwood Avenue, Boston, Massachusetts 02115

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High-throughput technologies, such as proteomic screening and DNA micro-arrays, produce vast amounts of data requiring comprehensive analytical methods to decipher the biologically relevant results. One approach would be to manually search the biomedical literature; however, this would be an arduous task. We developed an automated literature-mining tool, termed MedGene, which comprehensively summarizes and estimates the relative strengths of all human gene–disease relationships in Medline. Using MedGene, we analyzed a novel micro-array expression dataset comparing breast cancer and normal breast tissue in the context of existing knowledge. We found no correlation between the strength of the literature association and the magnitude of the difference in expression level when considering changes as high as 5-fold; however, a significant correlation was observed ($r = 0.41$; $p = 0.05$) among genes showing an expression difference of 10-fold or more. Interestingly, this only held true for estrogen receptor (ER) positive tumors, not ER negative. MedGene identified a set of relatively understudied, yet highly expressed genes in ER negative tumors worthy of further examination.

Keywords: bioinformatics • micro-array • text mining • gene-disease association • breast cancer

Introduction

At its current pace, the accumulation of biomedical literature outpaces the ability of most researchers and clinicians to stay abreast of their own immediate fields, let alone cover a broader range of topics. For example, to follow a single disease, e.g., breast cancer, a researcher would have had to scan 130 different journals and read 27 papers per day in 1999.¹ This problem is accentuated with high-throughput technologies such as DNA micro-arrays and proteomics, which require the analysis of large datasets involving thousands of genes, many of which are unfamiliar to a particular researcher. In any microarray experiment, thousands of genes may demonstrate statistically significant expression changes, but only a fraction of these may be relevant to the study. The ability to interpret these datasets would be enhanced if they could be compared to a comprehensive summary of what is known about all genes. Thus, there is a need to summarize existing knowledge in a format that allows for the rapid analysis of associations between genes and diseases or other specific biological concepts.

One solution to this problem is to compile structured digital resources, such as the Breast Cancer Gene Database¹ and the Tumor Gene Database.² However, as these resources are hand-curated, the labor-intensive review process becomes a rate-limiting step in the growth of the database. As a result, these

databases have a limited scale and the genes are not selected in a systematic fashion.

An alternative approach is automated text mining; a method which involves automated information extraction by searching documents for text strings and analyzing their frequency and context. This approach has been used successfully in several instances for biological applications. In most cases, it has been applied to extract information about the relationships or interactions that proteins or genes have with one another, in the literature or by functional annotation.^{3–7} Thus far, few publications have applied text-mining to examine the global relationships between genes and diseases. Perez-Iratxeta et al. automatically examined the GO (Gene Ontology) annotation of genes and their predicted chromosomal locations in order to identify genes linked to inherited disorders.⁸

To obtain a more global understanding of disease development, it would be valuable to incorporate information regarding all possible gene-disease relationships, including biochemical, physiological, pharmacological, epidemiological, as well as genetic. This information would enable comprehensive comparisons between large experimental datasets and existing knowledge in the literature. This would accomplish two things. First, it would serve to validate experiments by demonstrating that known responses occur as predicted. Second, it would rapidly highlight which genes are corroborated by the literature and which genes are novel in a given context. We have utilized a computational approach to literature mining to produce a

* To whom correspondence should be addressed: jlaBaer@hms.harvard.edu.

comprehensive set of gene-disease relationships. In addition, we have developed a novel approach to assess the strength of each association based on the frequency of citation and co-citation. We applied this tool to help interpret the data from a large micro-array gene expression experiment comparing normal and cancerous breast tissue.

Methods

MedGene Database. MedGene is a relational database, storing disease and gene information from NCBI, text mining results, statistical scores, and hyperlinks to the primary literature. MedGene has a web-based user interface for users to query the database (<http://hipseq.med.harvard.edu/MedGene/>).

Text Mining Algorithms. MeSH files were downloaded from the MeSH web site at NLM (National Library of Medicine) (<http://www.nlm.nih.gov/mesh/meshhome.html>) and human disease categories were selected. LocusLink files were downloaded from the LocusLink web site at NCBI (<http://www.ncbi.nih.gov/LocusLink/>). Official/preferred gene symbol, official/preferred gene name, and gene alternative symbols and names, all relevant annotations and URLs for each LocusLink record, were collected. Gene search terms were used for literature searching and included all qualified gene names, gene symbols, and gene family terms. Primary gene keys, predominantly qualified gene family terms and gene official/preferred symbols, were used to index Medline records. If the official/preferred gene symbols did not meet the standards to be an index, then qualified gene official/preferred names were used. A local copy of Medline records (up to July, 2002) was pre-selected.

A JAVA module examined the MeSH terms and then indexed each Medline record with the appropriate disease terms. A separate JAVA module was used to examine the titles and abstracts for gene search terms and then to index the gene-related Medline records with the relevant primary gene key(s).

Statistical Methods. For every gene and disease pair, we counted records that were indexed for both gene and disease (double positive hits), for disease only (disease single hits), for gene only (gene single hits), and for neither gene nor disease (double negative hits) to generate a 2×2 contingency table. On the basis of the contingency table-framework, we applied different statistical methods to estimate the strength of gene-disease relationships and evaluated the results. These methods included chi-square analysis, Fisher's exact probabilities, relative risk of gene, and relative risk of disease¹⁶ (<http://hipseq.med.harvard.edu/MedGene/>). In addition, we computed the "product of frequency", which is the product of the proportion of disease/gene double hits to disease single hits and the proportion of disease/gene double hits to gene single hits. To obtain a normal distribution, we transformed all the statistical scores using the natural logarithm. We selected the log of the product of frequency (LPF) to validate MedGene and to use for the analysis with the micro-array data. Spearman rank-correlation coefficients were used to assess the linear relationship between LPF and micro-array fold change in expression level.

Global Analysis. Diseases with at least 50 related genes were selected for clustering analysis, and the LPF scores were normalized with total score for each disease. Hierarchical clustering was done with the "Cluster" software and the clustering result was visualized using "TreeView" (<http://rana.lbl.gov/EisenSoftware.htm>).

Breast Tissue Micro-Arrays. Eighty-nine breast cancer samples (79% ER-positive) and 7 normal breast tissue samples were selected from the Harvard Breast SPORE frozen tissue repository and were representative of the spectrum of histological types, grades, and hormone receptor immuno-phenotypes of breast cancer. Biotinylated cRNA, generated from the total RNA extracted from the bulk tumor, was hybridized to Affymetrix U95A oligo-nucleotide micro-arrays. These micro-arrays consist of 12 400 probes, which represent approximately 9000 genes. Raw expression values were obtained using GENE-CHIP software from Affymetrix, and then further analyzed using the DNA-Chip Analyzer (dChip) custom software.

Results

Automated Indexing of Medline Records by Disease and Gene. To study the gene-disease associations in the literature, we first compiled complete lists for human diseases and human genes. To index all Medline records that were relevant to human diseases, the Medical Subject Heading (MeSH) index of Medline records was utilized. MeSH is a controlled medical vocabulary from the National Library of Medicine and consists of a set of terms or subject headings that are arranged in both an alphabetic and an hierarchical structure. Medline records are reviewed manually and MeSH terms are added to each with software assistance.^{9,10} Twenty-three human disease category headings along with all of their child terms (see the Supporting Information, Supplemental Table 1, or visit http://hipseq.med.harvard.edu/MedGene/publication/s_Table1.html) were selected from the 2002 MeSH index creating a list of 4033 human diseases.

No index comparable to the MeSH index exists for genes, and thus, it was necessary to apply a string search algorithm for gene names or symbols found in Medline text. A complete list of genes, gene names, gene symbols, and frequently used synonyms were collected from the LocusLink database at NCBI,^{11,12} which contains 53 259 independent records keyed by an official gene symbol or name (June 18th, 2002). For the purposes of this study, no distinction was made between genes and their gene products. Authors often use the same name for both, differentiating the two only by the use of italics, if at all. For the intended use of this study, this lack of distinction is unlikely to have a large effect and may in fact be beneficial.

Initial attempts to search the literature using these lists revealed several sources of false positives and false negatives (Table 1). False positives primarily arose when the searched term had other meanings, whereas false negatives arose from syntax discrepancies necessitating the development of filters to reduce these errors. The syntax issues were readily handled by including alternate syntax forms in the search terms. The false positive cases, caused by duplicative and unrelated meanings for the terms, were more difficult to manage. Where possible, case sensitive string mapping reduced inappropriate citations. In many cases, however, this was not sufficient and the terms had to be eliminated entirely, thereby reducing the false positive rate but unavoidably under-representing some genes.

For the purposes of data tracking, a primary gene key was selected to represent all synonyms that correspond to each gene. Medline records were indexed with a primary gene key when any synonym for that key was found in the title or abstract. Case-insensitive string mapping was used for all searches except as noted above. No additional weight was

Table 1. Systematic Sources of False Positives and False Negatives in Unfiltered Data^a

| source of error | error type | example | filter solution |
|---------------------------------------|----------------|--|------------------------------|
| gene symbol/name is not unique | false positive | MAG—myelin associated glycoprotein MAG—malignancy-associated protein | eliminate this term |
| gene symbol is unrelated abbreviation | false positive | PA—pallid homologue (mouse), pallidin (also abbrev. for Pennsylvania) | eliminate this term |
| gene symbol/name has language meaning | false positive | WAS—Wiskott–Aldrich Syndrome (also the word “was”) | case-sensitive string search |
| nonstandard syntax | false negative | BAG-1 instead of BAG1 | add dash term |
| unofficial gene name/symbol | false negative | P53 instead of TP53 | add all gene nicknames |
| nonspecified gene name | false negative | estrogen receptor instead of Estrogen receptor 1 | add family stem term |

^a In preliminary studies, Medline was searched for co-occurrence of genes and diseases and the resulting output was evaluated to identify error sources that were amenable to global filters. Each error source is categorized by the type of error it causes: false positives are suggested relationships that are not real and false negatives are real relationships that are underrepresented. The filter solutions used are indicated. Note that in some cases, the filter solution itself introduces error. In general, error rates maximized sensitivity, even at the expense of specificity if needed.

added for multiple occurrences of a term or the co-occurrence of multiple synonyms for the same gene key.

Medline records were searched with all qualified gene identifiers, such as the official/preferred gene symbol, the official/preferred gene name, all gene nicknames and all syntax variants. In situations where there are several members of a gene family or splice variants, some authors prefer to use a shortened gene family name, e.g., estrogen receptor instead of estrogen receptor 1 (*ESR1*), creating a source of false negatives. For this reason, gene family stem terms were created for all genes that have an alpha or numerical suffix (e.g., *IL2RA*, *TGFβ*, *ESR1*, etc.) and then used to search the literature. The family stem terms were handled separately from the specific gene names so that it would be clear when linkages were made to the gene family versus a specific member in that family.

To improve performance and accuracy, some pre-selection was applied to the records that were scanned. First, review articles were eliminated to avoid redundant treatment of citations. Second, non-English journals were removed because the natural language filters were only relevant to English publications. Finally, journals unlikely to contain primary data about gene-disease relationships were also removed (e.g., *Int. J. Health Educ.*, *Bedside Nurse*, and *J. Health Econ.*). Together, these filters reduced the 12 198 221 Medline publications (July 2002) by 37%.

Ranking the Relative Strengths of Gene-Disease Associations. In total, there were 618 708 gene-disease co-citations, in which 16% (8297) of all studied diseases had been associated to a disease and 96% (3875) of all diseases had been associated to at least one gene. To rank the relative strengths of gene disease relationships, we tested several different statistical methods and examined the results. With the exception of the relative risk estimates, the methods provided similar results with respect to the rank order of the gene-disease association strengths. However, after comparing the results to other databases and after consulting disease experts, the log of the product of frequency (LPF) was selected for further analysis because it gave the best results overall.

Validation of MedGene. In developing this tool, it was important to minimize the number of missed genes (false negatives) and miscalled genes (false positives). However, in situations when these goals were in conflict, inclusiveness was prioritized. To determine the false negative rate in MedGene, breast cancer was used as a test case because it was associated with more genes than any other human disease and because

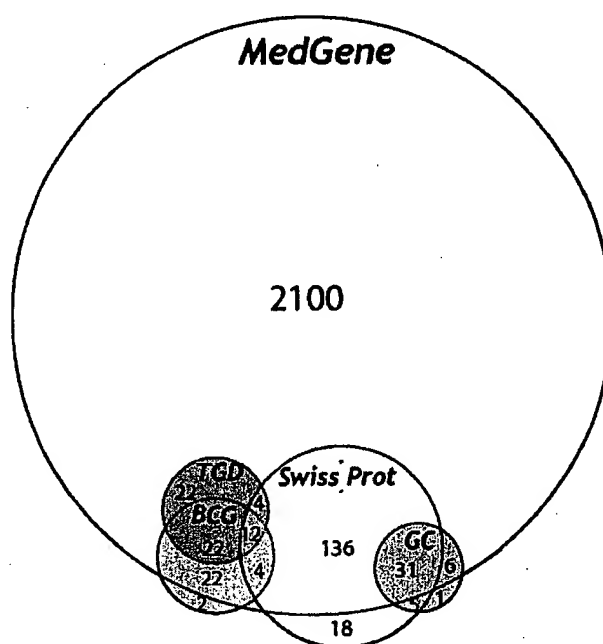


Figure 1. Estimation of the false negative rate by comparison with hand-curated databases. The breast cancer-related genes identified by MedGene were compared with those listed in several other databases including the Tumor Gene Database (TGD),² the Breast Cancer Gene Database (BCG),¹ GeneCards (GC)¹⁷ and Swissprot.¹⁸ Genes were considered false negatives if they were represented in at least one of these other databases and not in MedGene and their link to breast cancer was supported by at least one literature reference. All literature references were verified by manual review to confirm their validity. The number of genes in each database or shared by more than one database is indicated. The false negative rate was calculated by genes missed at MedGene (26)/total number of nonoverlapping genes in other databases (285).

there were several public databases that link genes to breast cancer. We compared the list of breast cancer-related genes from MedGene to these databases, illustrated in Figure 1. Among the 285 distinct breast cancer-related genes that were supported by at least one literature citation in these hand-curated databases, 26 were absent from MedGene, suggesting a false negative rate of approximately 9%. To determine why these were missed, all literature references for these genes (80

papers) were reviewed manually (see the Supporting Information, Supplemental Table 2, or visit http://hipseq.med.harvard.edu/MedGene/publication/s_Table 2.html). Among these papers, most false negatives were caused by nonstandard gene terms or gene terms eliminated by our specificity filters. Few genes were missed because they were only mentioned in review papers (0.4%) or they appeared only in the body of the manuscript but not the abstract or title (1.1%). Of note, MedGene identified approximately 2000 additional breast cancer-related genes not listed in any other database.

To assess the false positive error rate, two complementary approaches were used: a detailed analysis of one disease and a global examination of 1000 diseases. The detailed approach examined the false positive error rate and its sources, whereas the global approach tested whether the overall results made biomedical sense.

Using the LPF, 1467 genes related to prostate cancer were assembled in rank order. We then retrieved approximately 300 Medline records each for the highest ranked 100 and the lowest ranked 200 genes and manually reviewed the titles and abstracts to determine the verity of the association. Nearly 80% of the highest ranked 100 genes fell into one of the five categories that reflect meaningful gene-disease relationships (see the Supporting Information, Supplemental Table 3, or visit http://hipseq.med.harvard.edu/MedGene/publication/s_Table 3.html). Among the lowest ranked 200 genes, approximately 70% reflected true relationships. Of the 600 records reviewed, there were only two in which the association between the gene and the disease was described as negative. Both were genes with very low scores. In both cases, the authors did not argue the absence of any relationship, but rather that a particular feature of the gene or protein was not shown to be related to human prostate cancer.^{13,14}

The coincidence of some gene symbols with medical abbreviations, chemical abbreviations and biological abbreviations resulted in most of the false positives (see the Supporting Information, Supplemental Table 4, or visit http://hipseq.med.harvard.edu/MedGene/publication/s_Table 4.html), emphasizing the importance of the filters that were added in the search algorithm (Table 1). Without the filters, the false positive rate more than doubled, and the false negative rate rose dramatically (data not shown). For example, among the papers about breast cancer, there were only 12 Medline records that referred to *ESR1* and 10 to *ESR2*, whereas almost 2000 papers mentioned estrogen receptor without specifying *ESR1* or *ESR2*; this latter group was detected by the family stem term filter.

To further validate these results, a global analysis of the gene-disease relationships described by MedGene was performed. For this experiment, it was reasoned that the more closely related the diseases are to one another, the more they will be related to the same gene sets. Thus, if the relationships defined by MedGene accurately reflected the literature, then an unsupervised hierarchical clustering of the gene data should group diseases in a manner consistent with common medical thinking. Conversely, if the clustered diseases do not make sense biologically or medically, it may reflect excessive false positives, false negatives, or inappropriate scoring of the data.

To execute this experiment, the gene sets and the corresponding LPF values for 1000 randomly selected diseases (each with at least 50 gene relationships) were used as a dataset for clustering the diseases. A review of the results showed that the resulting disease clusters were indeed logical based upon common medical knowledge (see the Supporting Information,

Supplemental Figure 1, or visit http://hipseq.med.harvard.edu/MedGene/publication/s_Figure 1.html). For example, in one such cluster shown in Figure 2, diabetes and its complications grouped together and were also closely linked to diseases associated with starvation states.

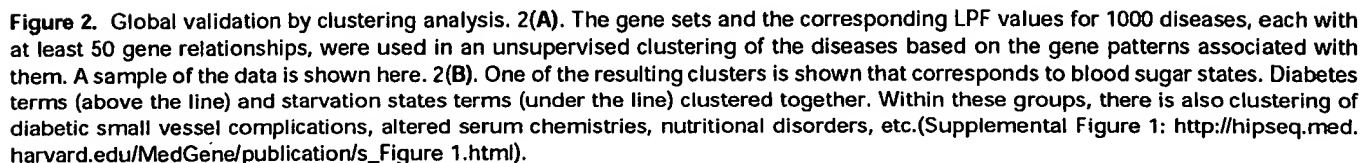
The number of genes associated with a given disease can be estimated by adjusting the MedGene number up by the false negative rate (~9%) and down by the false positive rate (~26% on average). Using this, the average disease has 103.7 ± 45.3 (mean \pm s.d.) genes associated with it, although the range is quite broad with 2359 genes related to breast cancer, 2122 genes related to lung cancer and no genes related to a number of diseases.

Applying MedGene to the Analysis of Large Datasets. Access to a comprehensive summary of the genes linked to human diseases provided an opportunity to analyze data obtained from a high-throughput experiment. We compared the MedGene breast cancer gene list to a gene expression data set generated from a micro-array analysis comparing breast cancer and normal breast tissue samples. Micro-array analysis identified 2286 genes that had greater than a 1-fold difference in mean expression level between breast cancer samples and normal breast samples. Using MedGene, we sorted the 2286 genes into four classes: 555 genes directly linked to breast cancer in the literature by gene term search (first-degree association by gene name); 328 genes directly linked by family term search (first-degree association by family term); 1021 genes linked to breast cancer only through other breast cancer genes (second-degree association); and 505 genes not previously associated with breast cancer. (See the Supporting Information, Supplemental Figure 2, or visit http://hipseq.med.harvard.edu/MedGene/publication/s_Figure 2.html.) Among the 505 previously unrelated genes, 467 were either newly identified genes or genes that had not previously been associated with any disease. Among the remaining 38 genes, 9 had been related to other cancers, specifically esophageal, colon, uterine, skin, and cervix.

To determine whether the genes highlighted by the micro-array analysis were more likely to have been previously linked to breast cancer in the literature, we created a two-dimensional plot of the fold change of expression level between breast cancer and normal tissue versus the literature score (LPF) (Figure 3A). There was a broad spread of expression changes among the genes directly linked to breast cancer ranging from less than 1-fold change (68%) to over 40-fold (0.3%). Notably, the majority of genes with greater than 10-fold expression changes were linked to breast cancer by first-degree association.

Among all 754 genes directly linked to breast cancer in the literature, there was no correlation between LPF and micro-array fold change ($r = 0.018$, p -value = 0.62). However, when we stratified the analysis based on the magnitude of the fold change, we observed an increasing trend in correlation (Figure 3B) suggesting that genes with a more substantial change in expression level were more likely to have a stronger association in the literature. For genes that had 10-fold change or more in expression level, the correlation increased to 0.41 (p -value = 0.05).

When we evaluated the micro-array data separately for ER positive and ER negative tumors, the trend in correlation between fold change and literature score was highly dependent on estrogen receptor status. Interestingly, there was a similar trend in correlation for ER positive tumors, but no trend in correlation for ER negative tumors.



disease unrelated to breast cancer. As expected, we did not observe an increasing trend in correlation for hypertension.

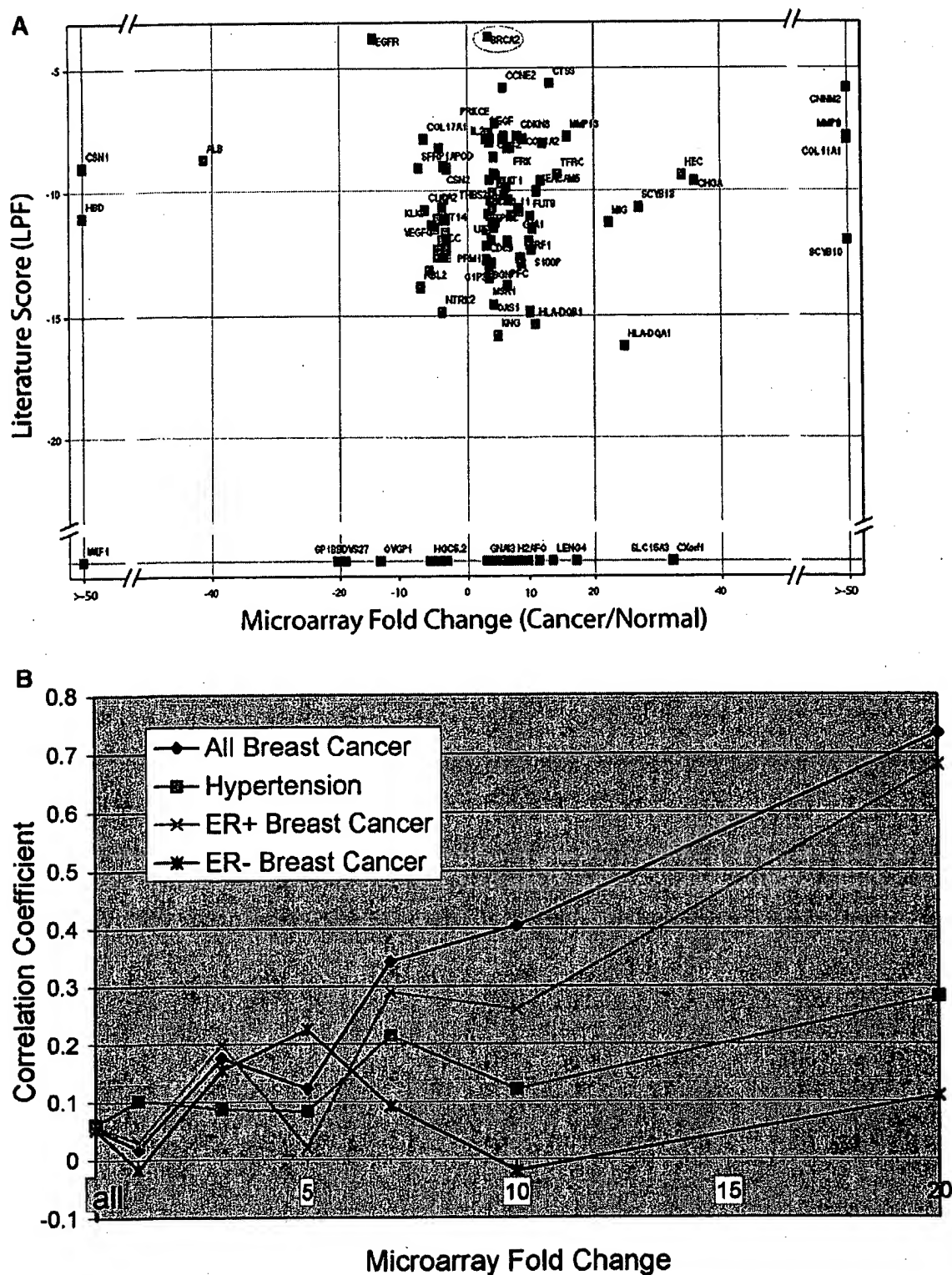


Figure 3. Relationship between literature score and functional data for breast cancer. **3A.** The data from an expression analysis of samples for breast tumors and normal breast tissue were analyzed to indicate the fold difference of expression level between breast tumor and normal sample (cutoff ≥ 3 -fold change). The fold changes were plotted against the literature score for the same gene set. Green dots represent first-degree association by gene search, blue dots represent first-degree association by family search and red dots represent no-association. Some well-studied genes, such as BRCA2 (pink circle), are not reflected by a substantial difference in expression level. Furthermore, the majority of genes that have no association with breast cancer in the literature had less than 10-fold expression changes (shaded area). **3B.** The Spearman rank-correlation coefficients between literature score (LPF) and the fold change of expression level between tumor and normal breast samples (y-axis) in relation to the amount of fold change of expression level (x-axis). Gene rank lists were generated for breast cancer (blue) and hypertension (pink). Correlations were also computed between the breast cancer gene LPF scores and fold change expression data among estrogen receptor positive tumors only (light blue) and estrogen receptor negative tumors only (purple).

Table 2. Top 25 Genes Related to Selected Human Diseases*

| breast neoplasms | hypertension | rheumatoid arthritis | bipolar disorder | atherosclerosis |
|-------------------|-------------------------|----------------------|------------------|---------------------|
| estrogen receptor | <i>REN</i> | <i>RA</i> | <i>ERDA1</i> | apolipoprotein |
| <i>PGR</i> | <i>DBP</i> | <i>TNFRSF10A</i> | <i>SNAP29</i> | <i>APOE</i> |
| <i>ERBB2</i> | <i>LEP</i> | <i>CRP</i> | <i>PFKL</i> | <i>LDLR</i> |
| <i>BRCA1</i> | <i>AGT</i> | <i>AS</i> | <i>DRD2</i> | <i>ELN</i> |
| <i>BRCA2</i> | <i>INS</i> | <i>ESR1</i> | <i>TRH</i> | <i>ARG1</i> |
| <i>EGFR</i> | kallikrein | <i>HLA-DRB1</i> | <i>IMPA2</i> | <i>APOB</i> |
| <i>CYP19</i> | <i>ACE</i> | <i>DR1</i> | <i>HTR3A</i> | <i>APOA1</i> |
| <i>TFF1</i> | endothelin | interleukin | <i>DRD3</i> | <i>MSR1</i> |
| <i>PSEN2</i> | <i>S100A6</i> | <i>TNF</i> | <i>REM</i> | <i>LPL</i> |
| <i>TP53</i> | <i>BDK</i> | <i>IL6</i> | <i>KCNN3</i> | <i>PON1</i> |
| <i>CES3</i> | <i>DIANPH</i> | collagen | <i>DRD4</i> | plasminogen |
| <i>CEACAM5</i> | <i>SAR1</i> | <i>IL1A</i> | <i>HTR2C</i> | activator inhibitor |
| <i>ERBB3</i> | <i>PIH</i> | <i>ACR</i> | <i>RELN</i> | <i>PLG</i> |
| cyclin | <i>CD59</i> | <i>TNFRSF12</i> | <i>DBH</i> | vascular cell |
| <i>COX5A</i> | <i>ALB</i> | <i>IL2</i> | <i>MAOA</i> | adhesion molecule |
| cathepsin | <i>CYP11B2</i> | <i>CHI3L1</i> | <i>COMT</i> | <i>ATOH1</i> |
| <i>ERBB4</i> | <i>MAT2B</i> | <i>IL8</i> | <i>HTR2A</i> | <i>VWF</i> |
| <i>TRAM</i> | angiotensin receptor | interleukin 1 | <i>SYNJ1</i> | <i>INS</i> |
| <i>CCND1</i> | <i>AGTR2</i> | matrix | <i>INPP1</i> | <i>ARG2</i> |
| <i>EGF</i> | <i>NPPA</i> | metalloproteinase | <i>NEDD4L</i> | <i>ABCA1</i> |
| <i>MUC1</i> | <i>LVM</i> | interferon | <i>FRA13C</i> | <i>OLR1</i> |
| insulin-like | <i>DBH</i> | <i>IL4</i> | transducer of | collagen |
| <i>BCL2</i> | <i>NPY</i> | <i>IL17</i> | <i>ERBB2</i> | <i>MCP</i> |
| mucin | <i>POMC</i> | <i>MMP3</i> | <i>BAIAP3</i> | lipoprotein |
| <i>FGF3</i> | neuropeptide | <i>SIL</i> | <i>ATP1B3</i> | <i>APOA2</i> |
| | | | <i>DRD5</i> | intercellular |
| | | | | adhesion molecule |
| | | | | <i>RAB27A</i> |

* MedGene results for the top 25 genes associated with breast neoplasms, hypertension, rheumatoid arthritis, bipolar disorder, and atherosclerosis, respectively, ranked by LPF scores. The hyperlink to all the papers co-citing the gene and the disease is available at MedGene website (<http://hipseq.med.harvard.edu/MedGene/>).

Discussion

The Human Genome Project heralded a new era in biological research where the emphasis on understanding specific pathways has expanded to global studies of genomic organization and biological systems. High-throughput technologies can provide novel insight into comprehensive biological function but also introduces new challenges. The utility of these technologies is limited to the ability to generate, analyze, and interpret large gene lists. MedGene, a relational database derived by mining the information in Medline, was created to address this need. MedGene users can query for a rank-ordered list of human gene-disease relationships (Table 2) for one or more diseases. Each entry is hyperlinked to the original papers supporting each association and to other relevant databases.

MedGene is an innovative extension of previous text mining approaches. Perez-Iratxeta et al. used the GO annotation and their chromosomal locations to predict genes that may contribute to inherited disorders.⁸ MedGene takes a broader view and includes all diseases and all possible gene-disease relationships. Furthermore, MedGene utilizes co-citation to indicate a relationship rather than GO annotation, which is limited to the subset of genes that have GO annotation. Our approach is complementary to that taken by Chaussabel and Sher, who used the frequency of co-cited terms to cluster genes into a hierarchy of gene-gene relationships.⁶

A unique aspect of this tool is the ability to assess the relative strengths of gene-disease relationships based on the frequency of both co-citation and single citation. This presupposes that most co-citations describe a positive association, often referred to as publication bias¹⁵ and is supported by our observations

that negative associations are rare (Supplemental Table 3: http://hipseq.med.harvard.edu/MedGene/publication/s_Table3.html). Of course, relationships established by frequency of co-citation do not necessarily represent a true biological link; however, it is strong evidence to support a true relationship.

Another important feature of MedGene is the implementation of software filters that substantially reduced the error rate. We estimate that less than 10% of all associations were missed and at least 70% of even the weakest associations were real. For this study, all of the filters that we applied were general ones, e.g., expanding the list of all gene names to address the different syntax forms used by different journals, eliminating gene names that correspond to common English words, etc. The majority of the remaining search term ambiguities were idiosyncratic and difficult to identify systematically without causing a significant rise in false negatives. Alternative approaches, such as the examination of the nearest neighbor terms, need to be considered to further reduce the false positive rate.

It is not uncommon to see expression changes in micro-array experiments as small as 2-fold reported in the literature. Even when these expression changes are statistically significant, it is not always clear if they are biologically meaningful. When comparing expression levels of disease to normal tissue, one expects an enrichment of known disease-related genes to appear in the altered expression group. MedGene provided a unique opportunity to test this notion in the context of existing knowledge on a novel breast cancer micro-array dataset. For genes displaying a 5-fold change or less in tumors compared to normal, there was no evidence of a correlation between altered gene expression and a known role in the disease. This

Table 3. Genes with Large Expression Changes in ER- but Not in ER+ Breast Tumors

| gene symbol | fold change (ER+) | fold change (ER-) |
|-----------------|-------------------|-------------------|
| <i>KRTHB1</i> | 1.0 | 610.8 |
| <i>BRS3</i> | 1.2 | 89.4 |
| <i>DKK1</i> | 1.2 | 69.8 |
| <i>ZIC1</i> | 1.9 | 59.6 |
| <i>TLR1</i> | 1.0 | 38.5 |
| <i>KIAA0680</i> | 2.6 | 33.2 |
| <i>CDKN3</i> | 1.0 | 30.6 |
| <i>EBI2</i> | 4.0 | 27.9 |
| <i>GZMB</i> | 3.8 | 21.9 |
| <i>STK18</i> | 4.7 | 18.6 |
| <i>GPR49</i> | 1.0 | 14.6 |
| <i>MYO10</i> | 1.6 | 14.4 |
| <i>LAD1</i> | -1.0 | 13.5 |
| <i>POLE2</i> | 4.2 | 13.0 |
| <i>HMG4</i> | 4.4 | 12.9 |
| <i>BCL2L11</i> | -1.2 | 12.3 |
| <i>LRP8</i> | 2.9 | 12.2 |
| <i>CCNB2</i> | 1.0 | 11.8 |
| <i>CCNE2</i> | 4.0 | 11.6 |
| <i>FGB</i> | -4.3 | 11.1 |
| <i>KNSL6</i> | 2.9 | 10.9 |
| <i>HIF5</i> | 3.0 | 10.2 |
| <i>SERPINH2</i> | 4.6 | 10.2 |
| <i>YAP1</i> | 1.0 | 10.0 |
| <i>LPHB</i> | -1.3 | -10.4 |
| <i>TCEA2</i> | -1.1 | -10.8 |
| <i>TFF1</i> | 1.3 | -11.4 |
| <i>COL17A1</i> | -4.1 | -15.7 |
| <i>POP5</i> | 1.1 | -16.2 |
| <i>BPAG1</i> | -4.6 | -22.3 |
| <i>PDZK1</i> | -1.1 | -36.8 |
| <i>VEGFC</i> | -2.8 | -51.5 |
| <i>MUC6</i> | -1.4 | -64.9 |
| <i>SERPINA5</i> | -1.0 | -83.1 |
| <i>MEIS1</i> | -1.6 | -85.9 |
| <i>CA12</i> | 2.4 | -150.3 |

Table 3. MedGene identified a set of relatively understudied, yet highly expressed genes in ER negative, but not ER positive breast tumors. All of these genes have either never been co-cited with breast cancer or have a weak association except those marked with an *.

reflects the many genes whose role in breast cancer may not involve large changes in expression in sporadic tumors (e.g., *BRCA1* and *BRCA2*) and genes whose modest changes in expression may be unrelated to the disease. Strikingly, among genes with a 10-fold change or more in expression level, there was a strong and significant correlation between expression level and a published role in the disease, providing the first global validation of the micro-array approach to identifying disease-specific genes.

The results derived from MedGene have two implications. First, a careful hunt for corroborating evidence of a role in breast cancer should precede any further study of genes with less than 5-fold expression level changes. Second, any genes with 10-fold changes or more are likely to be related to breast cancer and warrant attention. It is likely that this threshold will change depending on the disease as well as the experiment.

Interestingly, the observed correlation was only found among ER-positive tumors, not ER-negative. This may reflect a bias in the literature to study the more prevalent type of tumor in the population. Furthermore, this emphasizes that caution must be taken when interpreting experiments that may contain subpopulations that behave very differently. The MedGene approach identified a set of relatively understudied, yet highly expressed genes in ER-negative tumors that are worthy of further examination (Table 3).

In conclusion, we have developed an automated method of summarizing and organizing the vast biomedical literature. To our knowledge, the resulting database is the most comprehensive and accurate of its kind. By generating a score that reflects the strength of the association, it provides an important tool for the rapid and flexible analysis of large datasets from various high-throughput screening experiments. Furthermore, it can be used for selecting subsets of genes for functional studies, for building disease-specific arrays, for looking at genes common to multiple diseases and various other high-throughput applications. In the future, it will be possible to enhance the utility of the MedGene database by building links between genes and other MeSH terms as well as other biological processes and concepts, such as cell division and responses to small molecules.

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Supporting Information Available: Twenty-three human disease category headings along with all of their child terms selected from the 2002 MeSH index (Supplemental Table 1); analysis of the causes of false negatives in MedGene (Supplemental Table 2); meaningful gene-disease relationships found in MedGene (Supplemental Table 3); causes for incorrect assignment of gene indexes (Supplemental Table 4); a review of the results, showing that the resulting disease clusters were indeed logical (Supplemental Figure 1); and a review of the results showing that among the 505 previously unrelated genes, 467 were either newly identified genes or genes that had not previously been associated with any disease (Supplemental Figure 2). This material is available free of charge via the Internet at <http://pubs.acs.org> and at the web sites mentioned in the text.

References

- (1) Baasiri, R. A.; Glasser, S. R.; Steffen, D. L.; Wheeler, D. A. *Oncogene* 1999, 18, 7958-7965.
- (2) Steffen, D. L.; Levine, A. E.; Yarus, S.; Baasiri, R. A.; Wheeler, D. A. *Bioinformatics* 2000, 16, 639-649.
- (3) Marcotte, E. M.; Xenarios, I.; Eisenberg, D. *Bioinformatics* 2001, 17, 359-363.
- (4) Ono, T.; Hishigaki, H.; Tanigami, A.; Takagi, T. *Bioinformatics* 2001, 17, 155-161.
- (5) Jenssen, T. K.; Laegreid, A.; Komorowski, J.; Hovig, E. *Nat. Genet.* 2001, 28, 21-28.
- (6) Chaussabel, D.; Sher, A. *Genome Biol.* 2002, 3, RESEARCH0055.
- (7) Gibbons, F. D.; Roth, F. P. *Genome Res.* 2002, 12, 1574-1581.
- (8) Perez-Iratxeta, C.; Bork, P.; Andrade, M. A. *Nat. Genet.* 2002, 31, 316-319.
- (9) Funk, M. E.; Reid, C. A. *Bull. Med. Libr. Assoc.* 1983, 71, 176-183.
- (10) Humphrey, S. M.; Miller, N. E. *J. Am. Soc. Inf. Sci.* 1987, 38, 184-196.
- (11) Maglott, D. R.; Katz, K. S.; Sicotte, H.; Pruitt, K. D. *Nucleic Acids Res.* 2000, 28, 126-128.
- (12) Pruitt, K. D.; Maglott, D. R. *Nucleic Acids Res.* 2001, 29, 137-140.
- (13) Wadelius, M.; Andersson, A. O.; Johansson, J. E.; Wadelius, C.; Rane, E. *Pharmacogenetics* 1999, 9, 333-340.
- (14) Adam, R. M.; Borer, J. G.; Williams, J.; Eastham, J. A.; Loughlin, K. R.; Freeman, M. R. *Endocrinology* 1999, 140, 5866-5875.
- (15) Montori, V. M.; Smieja, M.; Guyatt, G. H. *Mayo Clin. Proc.* 2000, 75, 1284-1288.
- (16) Denenberg, V. H. *Statistics Experimental Design for Behavioral and Biological Researchers*; Wiley-Liss: New York, 1976.
- (17) Rebhan, M.; Chalifa-Caspi, V.; Prilusky, J.; Lancet, D. *Trends Genet.* 1997, 13, 163.
- (18) Bairoch, A.; Apweiler, R. *Nucleic Acids Res.* 2000, 28, 45-48. PR0340227

Genomic and proteomic analysis of the myeloid differentiation program

Zheng Lian, Le Wang, Shigeru Yamaga, Wesley Bonds, Y. Beazer-Barclay, Yuval Kluger, Mark Gerstein, Peter E. Newburger, Nancy Berliner, and Sherman M. Weissman

Although the mature neutrophil is one of the better characterized mammalian cell types, the mechanisms of myeloid differentiation are incompletely understood at the molecular level. A mouse promyelocytic cell line (MPRO), derived from murine bone marrow cells and arrested developmentally by a dominant-negative retinoic acid receptor, morphologically differentiates to mature neutrophils in the presence of 10 μ M retinoic acid. An exten-

sive catalog was prepared of the gene expression changes that occur during morphologic maturation. To do this, 3'-end differential display, oligonucleotide chip array hybridization, and 2-dimensional protein electrophoresis were used. A large number of genes whose mRNA levels are modulated during differentiation of MPRO cells were identified. The results suggest the involvement of several transcription regulatory factors not

previously implicated in this process, but they also emphasize the importance of events other than the production of new transcription factors. Furthermore, gene expression patterns were compared at the level of mRNA and protein, and the correlation between 2 parameters was studied. (Blood. 2001;98:513-524)

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Introduction

Studies of normal myeloid maturation from many laboratories have identified genes that may play critical roles in myeloid differentiation.¹⁻⁴ Current studies suggest that these events are dependent on a cascade of molecular changes that involve complex modulation of mRNA transcription. Furthermore, studies of acute leukemia have suggested that the disease arises from the accumulation of myeloid precursors arrested at early stages of differentiation and associated, in many cases, with chromosomal rearrangements that alter the structure of specific transcription factors.⁵ Nevertheless, the molecular events underlying the production of mature myeloid cells are not well understood and appear to use interacting pathways and networks, the elucidation of which requires an extensive description of the molecular components available to the myeloid cell.

An extensive body of information is accumulating with respect to gene expression profiles of mammalian cells. However, much of the information available in public databases has been accumulated by the use of techniques such as single oligonucleotide chips or cDNA arrays that measure fewer than 6000 of potentially 30 000 to 120 000 transcripts. The more limited range of analyses reported by the serial analysis of gene expression (SAGE)^{6,7} technique accurately estimates changes in levels of the more abundant mRNAs but requires extensive redundant analyses to measure changes in the patterns of expression of scarce mRNAs. We have used a modified polymerase chain reaction (PCR)-based cDNA differential display (DD) method in which single restriction fragments derived from the 3' end of cDNAs are separated on a sequencing gel.^{8,9} Bands from the gel can be identified initially by sequencing, but then

comparison of patterns from different samples can be made without further sequencing. This sensitive and reproducible method detects, in principle, most cDNAs regardless of whether they are represented in existing databases.

Systematic analysis of the function of genes can also be performed at the protein level. This approach has the advantage of being closest to function, because proteins perform most of the reactions necessary for the cell. The most common method of proteome analysis is the combination of 2-dimensional gel electrophoresis (2DE) to separate and visualize protein and mass spectrometry (MS) for protein identification.¹⁰ Several such analyses of yeast and of normal or malignant mammalian cells have been performed. To date, however, there have been few studies in which both mRNA and protein have been compared by applying analyses to the same samples. The studies of Anderson¹¹ and Gygi¹² showed that there is not a good correlation between mRNA and protein levels, in yeast or human liver cells. However, other analyses disagree with this conclusion (Greenbaum et al, manuscript submitted, and Fletcher et al¹⁴). Furthermore, global correlations between changes in mRNA and protein levels have not been examined during the execution of any developmental program.

The MPRO cell line was derived by transduction of a dominant-negative retinoic acid receptor construct into normal mouse bone marrow cells. It is a granulocyte-macrophage colony-stimulating factor (GM-CSF)-dependent line arrested at a promyelocytic stage of development.^{15,16} After treatment with all-trans retinoic acid (ATRA) most of the cells acquire the morphology of mature

From the Department of Genetics, Boyer Center for Molecular Medicine, the Section of Hematology, Department of Internal Medicine, and the Department of Molecular Biophysics and Biochemistry, Yale University School of Medicine, New Haven, CT; the Department of Pediatrics, University of Massachusetts Medical School, Worcester, MA; and Gene Logic, Gaithersburg, MD.

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L.W. and S.Y. contributed equally to this research.

Reprints: Sherman M. Weissman, Department of Genetics, Boyer Center for Molecular Medicine, Yale University School of Medicine, Rm 336, 295 Congress Ave, New Haven, CT 06536-0812; e-mail: sherman.weissman@yale.edu.

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neutrophils and begin to produce neutrophil lactoferrin and gelatinase, 2 proteins characteristic of neutrophil secondary granules.¹⁷ As such, it offers a valuable model for studying neutrophil differentiation *in vitro*.

We now report the analysis of mRNA expression changes during the process of MPRO cell maturation to neutrophils and compare the results with a limited analysis of cellular protein composition. mRNA expression changes were studied by combining the use of oligonucleotide arrays and DD. A database (dbMC) with comprehensive genomic information for myeloid differentiation program was constructed (accessible at <http://www.bioinfo.mbb.yale.edu/expression/neutrophil>). We have grouped the changes in mRNA levels of a large number of genes into 6 patterns, with implications for the genetic program of myeloid differentiation.

We also compared 2-dimensional high-resolution gel electrophoretograms from control cells and cells differentiated for 72 hours in the presence of ATRA. Fifty protein spots whose relative intensity changed prominently during differentiation were examined by mass spectrometry. The results suggest a poor correlation between mRNA expression and protein abundance, indicating that it may be difficult to extrapolate directly from individual mRNA changes to corresponding ones in protein levels (as estimated from 2DE).

Materials and methods

Cell lines

MPRO cells and HM-5 cells provided by Dr Schickwahn Tsai (Fred Hutchinson Cancer Research Center, Seattle, WA)¹⁵ were used throughout the study. The cells proliferated continuously as a GM-CSF-dependent cell line at 37°C in Iscoves modified Dulbecco medium (Gibco BRL, Grand Island, NY) supplemented with 5% to 10% fetal calf serum (Gibco BRL) and 10% HM-5-conditioned medium as a source of GM-CSF. Morphologic differentiation of the blocked MPRO promyelocytes was induced by treatment with 10 μ M ATRA (Sigma, St Louis, MO). Controls were cultured in the absence of ATRA but with the same volume of vehicle (ethanol).

RNA isolation and differential display

After exposure to 10 μ M ATRA for 0, 24, 48, or 72 hours, total cellular RNA was isolated from MPRO cells using TRIzol reagent (Life Technologies, Gaithersburg, MD). cDNA was then synthesized using a T-7 Sal-Oligo d(T) 32 primer as described previously.^{8,18} The double-stranded cDNA was digested with 1 of 9 different restriction enzymes (*Apa*I, *Bgl*II, *Bam*HI, *Eco*RI, *Hind*III, *Xba*I, *Kpn*I, and *Sph*I) and ligated to Y-shaped adaptors with a complementary overhang. DNA fragments were then amplified by PCR as described previously.^{8,18} PCR products were separated on a sequencing gel of 6% polyacrylamide with 7 M urea. The gel was dried and exposed to x-ray film. Genes from differential display gels, whose maximum intensity changes equaled 2+ on a scale of 1+ to 8+, were recorded as significantly changed.¹⁹ Individual DNA bands were recovered from the gels, amplified by PCR, and sequenced.

Oligonucleotide chip analysis of RNA samples

Ten micrograms total RNA from each sample (0, 24, 48, or 72 hours) was used to prepare cDNA. This cDNA was transcribed with T7 RNA polymerase to prepare a fluorescently labeled probe.^{20,21} Each sample was hybridized to mouse array chip (Multik Array; Affymetrix, Santa Clara, CA) containing oligonucleotide probe sets corresponding to approximately 7000 known genes or ESTs represented by UniGene clusters.²² cDNAs were considered present if their probe set results were rated as such by the GeneChip software (Affymetrix) and if the average difference (AD) between perfect match and mismatch probe pairs was not less 100 U. If a

gene was represented by more than one array probe set, the average of all probe sets for the gene was taken. Genes with AD values between 100 and 200 were considered unchanged because of their low expression levels. Those genes with AD values equal to or more than 200 U at one time point were further studied by rescaling, threshold, and normalization methods described in the MIT Center for Genome Research Web site.¹³ A value of 20 was assigned to any gene with an AD below 20 at some time point.

Bioinformatics and database development

All the sequences or gene fragments were searched using Blast against GenBank and TIGR gene indices. A database of genes or ESTs whose expression levels changed during myeloid differentiation was constructed containing information for each band or gene. This included GenBank matches, Locus Link or Unigene clusters, expression patterns, tissue distribution, synonym(s) protein name, gene name(s), notations of possible functions, poly A signal and sequence quality, and hyperlinks to the database searches, sequence trace files, and related references. All gene data were then gathered into a cluster file. Supplementary information is available at <http://bioinfo.mbb.yale.edu/expression/neutrophil>.

Classification and analysis of DNA fragments

Sequences from differential display analyses were classified as representing known genes, ESTs, genomic sequences, or novel genes as described.^{19,23} Known genes from both differential display and arrays were clustered into 27 functional categories and searched against SWISS-PROT (<http://www.expasy.ch/cn/cgibin/sprot-search-full>) or PIR (<http://www.pir.georgetown.edu/>). Information such as function, subcellular location, family and superfamily classification, map position, similarity, synonym(s) protein name, gene name(s), and so on was recorded in a variety of databases.

Northern blot analysis

Thirty micrograms total cellular RNA per lane from time-course MPRO cells were loaded onto 1.2% formaldehyde-agarose gels, then transferred to Hybond-N+ membranes (Amersham Pharmacia Biotech, Uppsala, Sweden). After standard prehybridization, membranes were hybridized overnight at 65°C with radiolabeled cDNA probes (ordered from Research Genetics according to their dbEST Image ID). Membranes were washed at a final stringency of 60°C in 0.1 \times SSC.

Immobilized pH gradient 2-dimensional gel electrophoresis and mass spectrometry

Induced MPRO cells collected at 0 and 72 hours were lysed with lysis buffer (540 mg urea, 20 mg dithiothreitol, 20 μ L Pharmalyte [3-10], 1.4 mg phenylmethylsulfonyl fluoride, 1 μ g each aprotinin, leupeptin, pepstatin A, and antipain 50 μ g TLCK, and 100 μ g TPCK/1 mL). We applied 100 μ L each MPRO cell lysate (2.5 \times 10⁶ cells/100 μ L) to immobilized pH gradient (IPG) strips (pH 3-10 L; Amersham Pharmacia Biotech), and IPG electrophoresis was conducted for 16 hours (20 100 Vh) using an Immobiline Drystrip Kit (Amersham Pharmacia Biotech). Electrophoresis in the second dimension was carried out in a 12% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) gel with the Laemmli-SDS continuous system in a Protean II xi 2-D cell (Bio-Rad) run at 40 mA constant current for 4.5 hours. Proteins were detected by Brilliant Blue G-cooloidal staining.²⁴ Protein spots were excised from the gel and digested with trypsin. ACTH clip (average [M+H] 2466.70) and bradykinin (average [M+H] 1061.23) were used for calibration of peptide masses. One microliter sample digest was mixed with 1.0 μ L α -cyano-4-hydroxy cinnamic acid (4.5 mg/mL in 50% CH₃CN, 0.05% TFA) matrix solution and 1 μ L calibrants (100 fmol) each. The spectra of the peptides were acquired in reflector/delayed extraction mode on a Voyager-DE STR mass spectrometer (Perceptive Biosystems, Foster City, CA). Peptides were identified using the ProFound search engine.²⁵

Results

Differentiation of MPRO cells

Figure 1 illustrates the morphologic changes in an MPRO cell population representative of those used for RNA expression analysis. Undifferentiated MPRO cells resembled promyelocytes under the light microscope (Figure 1A). After induction with ATRA for 24 hours, the cells morphologically differentiated into metamyelocytes (Figure 1B). At 48 hours, the cells further developed into metamyelocytes and band neutrophils (Figure 1C). At 72 hours, nearly 100% of MPRO cells became mature neutrophils (Figure 1D).

Identification of mRNAs by differential display assay

MPRO cellular mRNA was analyzed at 0, 24, 48, and 72 hours after ATRA treatment. Nine restriction enzymes were used in a 3'-end DD approach. During MPRO differentiation, 1109 fragments corresponding to 837 transcripts were found to change substantially in expression levels (Figure 2). These represented approximately 279 known genes, 112 ESTs, and 59 putative new genes, each with a perfect or fair polyadenylation signal at an appropriate distance from the oligo-dT priming site. The gene information detected by DD was collected in database dbMCd.

Identification of mRNAs by oligonucleotide chip assay

We used an oligonucleotide chip containing 13 179 probe sets corresponding to approximately 7000 murine genes to analyze patterns of mRNA expression in the same RNA samples used for DD. The information obtained by oligonucleotide arrays was collected in the database dbMCa.

We clustered the genes by their similarity to idealized expression patterns. For instance, the expression pattern of an ideal gene that is overexpressed (high) at time 0 and underexpressed (low) at 24, 48, and 72 hours, would be high-low-low-low (HLLL). Overall we have ($2^4 - 2$) idealized patterns excluding HHHH and LLLL. Pearson correlation was used as the

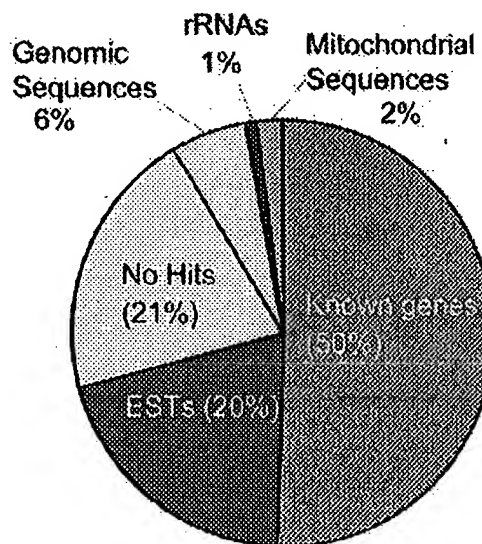


Figure 2. Distribution of genes obtained by DD assay. MPRO cell mRNA was analyzed at 0, 24, 48, and 72 hours after ATRA treatment; 1109 fragments corresponding to 837 transcripts were found to change substantially in expression levels. The total 837 transcripts were classified into 6 categories according to the bioinformatic analysis. Percentages show the gene distributions in these 6 categories. Information for each transcript was collected in database dbMCd.

measure of similarity of each gene expression pattern, $x = (x_1, x_2, x_3, x_4)$ to each of the 14 idealized patterns $y = (y_1, y_2, y_3, y_4)$. The 4 entries of x and y corresponded to the 4-dimensional gene expression levels at 0, 24, 48, and 72 hours, respectively. Each gene was assigned to a cluster labeled by the idealized pattern that had the maximal correlation with that gene. We selected only genes that hybridized well compared with the background (considered "present" by GeneChip software) and had maximal AD amplitude greater than 200 U in at least 1 of the 4 stages. We further tabulated the 14 patterns according to whether the gene expression changed at early (0-hour), intermediate (24- and 48-hour), and late (72-hour) time points and whether gene expression monotonically increased (up-regulated), monotonically decreased (down-regulated), or was not monotonic (transient). Table 1 shows 8 clusters of 104 genes that had significant changes of mRNA levels, arranged according to the temporal stage and the monotonic/transient changes of expression levels.

Principal component analysis determined whether we could comprehensively present multidimensional data (4-dimensional in our case) in a simple 2-dimensional graph. First, we found the 4 principal components, which were the axes of the most compact 4-dimensional ellipsoid that encompassed the 4-dimensional cloud of data. Each axis was a different linear combination of the original 4 variables. Then we verified that the first 2 principal components (the first 2 largest axes of the ellipsoid) captured most (95.2%) of the variation of the data. Therefore, the data could be faithfully projected (with a minor loss of information) into a 2-dimensional graph, with the 2 largest principal components as the x - and y -axes. As shown in Figure 3, genes tend to coalesce in clusters, according to their labels determined by their similarity to an ideal expression pattern. In summary, a genomic (global) picture of the distribution of genes according to their similarity to predetermined idealized multidimensional expression patterns is concisely displayed in a 2-dimensional graph.

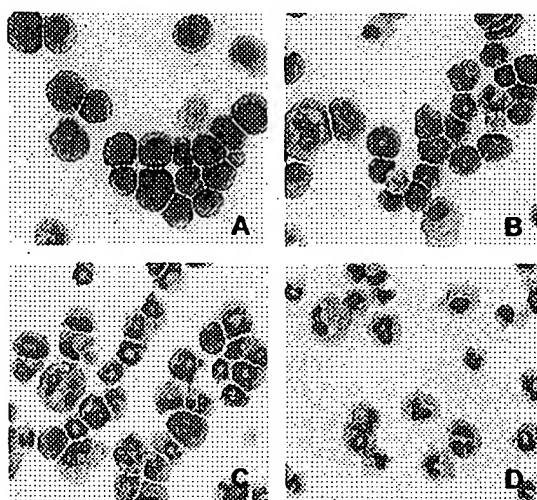


Figure 1. Morphology of MPRO cells during differentiation. MPRO cells were induced as described in "Materials and methods," concentrated by cyto spin, and Wright-Giemsa stained. (A) Uninduced MPRO cells. (B) MPRO cells induced with ATRA for 24 hours. (C) MPRO cells induced with ATRA for 48 hours. (D) MPRO cells induced with ATRA for 72 hours.

Table 1. Genes differently regulated during the different stages of mouse promyelocytic cell line differentiation process

| Time | | | |
|-----------------|---|--|--|
| Category | Early | Middle | Late |
| Up-regulation | LHHH (n = 10) <i>Mad</i> <i>P2rx1</i> <i>ttgb2</i> <i>Il1r2</i> <i>Lcn2</i> <i>Ilpr5</i> <i>Cebpb</i> <i>H2-D</i> <i>Etoh16</i> <i>Zyx</i> | LLHH (n = 6) <i>Pirai</i> <i>Cybb</i> <i>Pfc</i> <i>Pira5</i> <i>Cd53</i> <i>Ilngnr2</i> | LLLH (n = 13) <i>Il1a</i> <i>Csf1rli</i> <i>Cts1</i> <i>S100a8</i> <i>L-COR</i> <i>Ctss</i> <i>Aldo1</i> <i>Rac2</i> <i>Fpr1</i> <i>Ctsd</i> <i>Ubb</i> <i>Ptmb4</i> |
| Down-regulation | HLLL (n = 11) <i>Tcrg-V4</i> <i>Ly64</i> <i>Ctsg</i> <i>Spl2-1</i> <i>Mcp18</i> <i>Myc</i> <i>Myb</i> <i>Tlr4</i> <i>Npm1</i> <i>Erh</i> <i>Hsp60</i> | HHLL (n = 1) <i>Mpo</i> | HHHL (n = 37) <i>Actx</i> <i>Ir2</i> <i>EL2</i> <i>Rpl19</i> <i>Actb</i> <i>Ly6e</i> <i>Aif1</i> <i>Hist2</i> <i>Pesma2</i> <i>Gnas</i> <i>Zfp36</i> <i>Il4ra</i> <i>Ltbr</i> <i>Shfdg1</i> <i>Max</i> <i>Rps8</i> <i>Csf2rb1</i> <i>Sipi</i> <i>Tdex1</i> <i>Tpi</i> <i>B3f3</i> <i>Cnrf</i> <i>Gys3</i> <i>Sic10a1</i> <i>Ctsb</i> <i>Sepp1</i> <i>Rtn3</i> <i>Ccnb2</i> <i>S100a9</i> <i>Cf11</i> <i>Hist5-2ax</i> <i>Rela</i> <i>Copa</i> <i>Gstm1</i> <i>Gnb2-rs1</i> <i>Gm</i> <i>RPL8</i> |
| Transient | | LLHL (n = 9) <i>Sell</i> <i>Klf2</i> <i>Pira6</i> <i>Plrb</i> <i>Lst1</i> <i>Ltf</i> <i>Sema4d</i> <i>Stat6</i> <i>Mmp9</i> LHHL (n = 17) <i>Cebpa</i> <i>Lyza</i> <i>Fcgr3</i> <i>Arf5</i> <i>Lamp1</i> <i>Stat3</i> <i>Csf2ra</i> <i>Osi</i> <i>Actg</i> <i>Sipi1</i> <i>Gpx3</i> <i>Ptprc</i> <i>Prtm3</i> <i>Irf1</i> <i>Rps6ka1</i> <i>Ltb4r</i> <i>Myln</i> | |

Arrays of Affymetrix Mu11k containing 13103 probe sets corresponding to 12002 GenBank accessions were used for hybridization. Arrays were hybridized with streptavidin-phycoerythrin (Molecular Probes) biotin-labeled RNA and scanned. Intensity for each feature of the array was captured using Genechip software (Affymetrix), and a single raw expression level for each gene was derived from the 20 probe pairs representing each gene using a trimmed mean algorithm. For each gene, an AD of 24-, 48-, and 72-hour samples was calibrated by dividing the slope of the linear regression line for a graph with the x-axis the AD of 0-hour probe sets and the y-axis the AD of the respective time point (24, 48, or 72 hours). A threshold of 20 U was assigned to any gene with a calculated expression level below 20 because discrimination of expression below this level could not be performed with confidence.³⁰ Each gene expression profile was categorized as described in Tables 3, 4, and 5. For the 4 time points, the minimum AD of the relatively higher group (MIN-H) was divided by the maximum AD of the relatively low group (MAX-L), and those genes whose MIN-H/MAX-L greater than 2 were selected as meaningfully regulated. Genes were sorted in descending order based on the MIN-H/MAX-L. Genes in boldface are those whose expression level was in the top 20% (ie, maximum AD of 4 time points greater than 3000), and genes in italics are those in the bottom 20% (ie, maximum AD of 4 time points less than 300). The differentiation period was grouped into 3 stages: early (0-hour), middle (24-hour and 48-hour), and late (72-hour) stages.

AD indicates average difference; gene symbols are expanded in an Appendix at the end of this article.

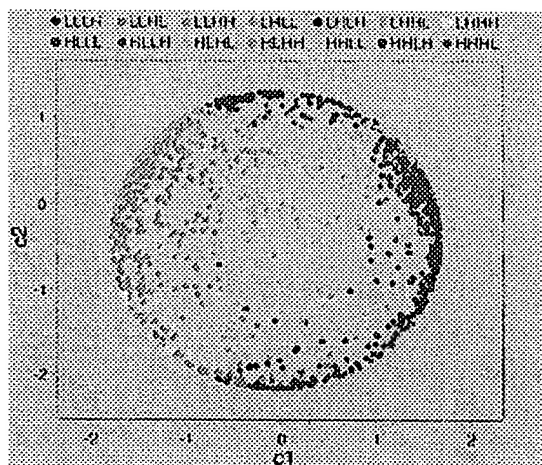


Figure 3. Gene clusters in the first 2 principal component spaces. Principal component analysis allowed us to present the multidimensional data (in this case, 4-dimensional data of each gene expression pattern) in a simple 2-dimensional graph. We derived the 4 principal components, which are a linear combination of the standardized expression intensities (zero mean and unit variance) at 0, 24, 48, and 72 hours. The first 2 principal components captured most of the variation of the data (approximately 85%). Therefore, the data can be displayed (with a minor loss of information) in a 2-dimensional graph. The first and second principal components, c_1 and c_2 , are given by the linear combinations $c_1 = 0.747 \cdot n_1 - 0.11 \cdot n_2 - 0.656 \cdot n_3 + 0 \cdot n_4$ and $c_2 = 0.278 \cdot n_1 + 0.353 \cdot n_2 + 0.233 \cdot n_3 - 0.863 \cdot n_4$, where n_1, n_2, n_3 , and n_4 are the rescaled and standardized expression levels at 0, 24, 48, and 72 hours, respectively. The axes legends c_1 and c_2 stand for the first 2 principal components. In this paper we used the Pearson correlation to measure the similarity of each gene with the idealized expression patterns, as opposed to the Euclidean distance we used in a previous work,¹⁹ because clusters were better separated using this measure. In both cases, we presented the data in the 2-dimensional space of the lowest principal components. The data had a tendency to be circularly distributed when we used the Pearson correlation as a distance measure.

We have previously demonstrated a correlation coefficient of 0.93 between visual estimates of changes in band intensity on DD and PhosphorImager System (Molecular Dynamics, Sunnyvale, CA) estimates of band intensity and a correlation coefficient of 0.88 between hybridization intensity changes of mRNA on Northern blot analyses and changes in band intensity on DD.¹⁹ In a few cases there were clear discrepancies in the pattern of expression of a gene, as estimated by DD and by oligonucleotide chip analysis. We chose the 6 most extreme cases and examined the levels of mRNA change for these genes by Northern blot analysis (Figure 4). In 5 cases, the Northern blot results agreed with the results of the DD analysis, whereas the results of Gnb2-rs1 disagreed with the oligonucleotide array but duplicate bands from DD showed a relatively high level of expression in the 0 time sample that did not correlate with the Northern blot (Table 2). One possible explanation for these findings was the change in the relative use of different polyadenylation sites after the addition of ATRA to the MPRO cells.

Constructing a database for mRNA level changes during myeloid differentiation

Based on the data obtained above, an in-house database (dbMC) was constructed that included 2 subdatabases, dbMCd and dbMCa, for collecting gene information from DD or oligonucleotide arrays, respectively. Each entry in dbMC is accompanied by a so-called executive summary. The linkage between dbMCd and dbMCa was established by UniGene ID and cluster ID. dbMC contains the temporal expression patterns of genes during the MPRO cell differentiation process, including not only products represented in public databases but also novel transcripts.

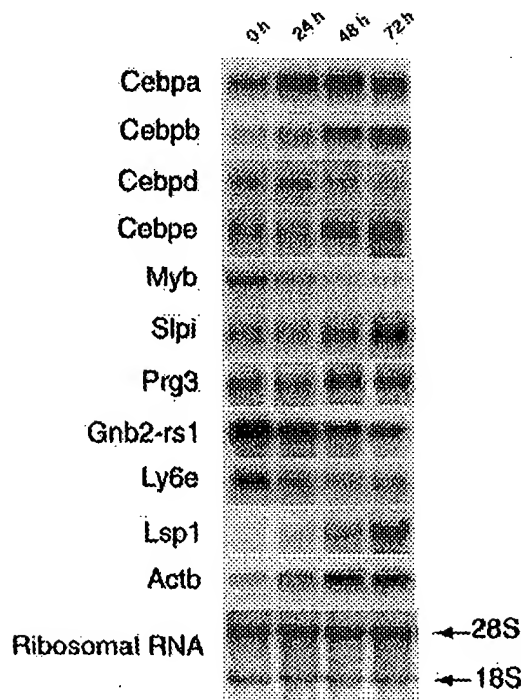


Figure 4. Northern blot analysis of selected mRNAs. Equivalent amounts of RNA from MPRO cells induced by ATRA at different time points (0 hour, 24 hours, 48 hours, and 72 hours) were resolved by formaldehyde-agarose gel electrophoresis, stained to verify the amount of loading. Eleven genes were separately probed on the RNA filters. The gene symbol of each probe was listed at the left of a related Northern blot result. Detailed information on these 11 probes was listed in Table 5. One of the RNA-blotted membrane photographs is shown with methylene blue-stained 28S and 18S RNA subunits demonstrating the quality and quantity of RNA loaded in individual lanes.

Analysis of gene expression patterns during MPRO differentiation

Many of the genes identified in this study were found in myeloid cells or were implicated in myeloid development for the first time. We detected 8 cytokines²⁵ and chemokines whose mRNA levels changed more than 5-fold by arrays and 2-fold by DD during the maturation of MPRO cells (see our Web site, <http://bioinfo.mbb>).

yale.edu/expression/neutrophil). Among these were 2 members of the CC chemokine family. Interleukin-1 α (IL-1 α) was up-regulated at the late stage of differentiation (LLLH pattern, Table 1).

mRNA for approximately 52 receptors was detected by one or the other method. A number of the receptors known to be present on mature neutrophils showed late induction of mRNA, and their levels of induction were high, indicating that the expression of these products is a prominent event late in neutrophil maturation (Table 3). Rarely was mRNA for receptors down-regulated, consistent with myeloid maturation being accompanied by increasing responsiveness of the cell to a variety of external stimuli.

Expression of mRNA for granule proteins

Neutrophils contain several types of granules that develop at different stages of myeloid maturation.^{3,17,26} Levels of mRNAs encoding secondary granule proteins, such as lactoferrin, increased as the cells matured (Table 4). The level of mRNA for Mmp9, reported as a tertiary granule protein, increased markedly between 24 and 48 hours after the induction of differentiation, whereas mRNAs for secondary granule proteins either increased less markedly or showed a maximum increase by 24 hours. mRNAs for several primary granule constituents, such as myeloperoxidase and cathepsin G, were present in unstimulated cells and decreased as the cells matured. There was a discrepancy in the measurements of proteoglycan mRNA by DD and oligonucleotide chips, but Northern blots showed that it reached a peak at 48 hours and then declined (Figure 4). Cathepsin D is reported as a primary granule protein, but its pattern of mRNA expression more closely resembled that of secondary granule constituents. In addition to known granule components, mRNAs for several other cathepsins were up-regulated during myeloid differentiation, in parallel with or later than the tertiary granule protein mRNAs.

mRNAs for transcription factors

Transcription factor genes, including several identified at the sites of consistent chromosome rearrangements in acute myeloid leukemia, have been implicated in normal myeloid differentiation and in the expression of neutrophil proteins.^{2,5,27} However comprehensive information concerning the expression of these transcription factors during myeloid development is not readily available. Therefore, we compared gene names and identifiers in our databases to those of the transcription factor database Transfac (<http://>

Table 2. Expression patterns of genes detected by Northern blot analysis

| Gene symbol | Gene accession | AD value by array | | | | Intensity by DD | | | |
|-------------|----------------|-------------------|------|------|------|-----------------|------|------|------|
| | | 0 h | 24 h | 48 h | 72 h | 0 h | 24 h | 48 h | 72 h |
| Cebpa | M62362 | 33 | 212 | 182 | 44 | — | — | — | — |
| Cebpb | X62600 | 390 | 1248 | 1380 | 1903 | — | — | — | — |
| Cebpd | X61800 | 157 | 262 | 168 | 430 | — | — | — | — |
| Cebpe | — | — | — | — | — | — | — | — | — |
| Myb | M12848 | 892 | 356 | 230 | 435 | — | — | — | — |
| Sipl | U73004 | 617 | 501 | 783 | 402 | 1 | 2 | 3 | 3 |
| Prg3 | W45834 | 153 | 259 | 339 | 345 | 5 | 1 | 1 | 2 |
| Gnb2-rs1 | X75313 | 4231 | 3623 | 3215 | 3403 | 4 | 4 | 1 | 1 |
| Ly6e | U04268 | 3061 | 5391 | 2844 | 1262 | 3 | 2 | 1 | 1 |
| Lsp1 | M90316 | 65 | 376 | 840 | 28 | 2 | 3 | 5 | 6 |
| Actb | X03765 | 3095 | 3588 | 3976 | 2434 | 1 | 2 | 3 | 2 |

Gene symbol and gene accession refer to National Center for Biotechnology Information databases and, in particular, to Locus Link. AD value is the average difference in the value of hybridization intensity between the set of perfectly matched oligonucleotides and the set of mismatched oligonucleotide in the oligonucleotide array. Band intensities from DD were semiquantified on a scale from 1 (+) to 6 (+++++). These estimates are shown as boldface numbers in this table.¹⁹ Both AD value and intensity of genes were studied at 4 time points corresponding to MPRO cells induced for the indicated times.

DD indicates differential display; MPRO, mouse promyelocytic cell line; for gene symbols, see the Appendix at the end of this article.

Table 3. Receptors expressed during myeloid differentiation process

| Maximal fold change | Gene symbol | Gene accession | AD value by array | | | |
|------------------------|-------------|----------------|-------------------|------|------|------|
| | | | 0 h | 24 h | 48 h | 72 h |
| Less than 2 | Bzrp | D21207 | 641 | 658 | 881 | 887 |
| | Cmkar4 | X99581 | 508 | 447 | 378 | 684 |
| | Crry | M34173 | 433 | 384 | 506 | 506 |
| | Csf2rb1 | M34397 | 318 | 345 | 410 | 241 |
| | Htr5a | Z18278 | 188 | 272 | 273 | 339 |
| | M6pr | X64068 | 536 | 409 | 408 | 649 |
| | MPPIR | AA116789 | 232 | 84 | 63 | 381 |
| | TCRGB | M28053 | 165 | 212 | 244 | 299 |
| | Trfrs1a | M59377 | 0 | 1 | 1 | 1 |
| 2 or more, less than 3 | Cmkbr1 | U28404 | 221 | 244 | 504 | 638 |
| | Crrr | X72305 | 121 | 200 | 250 | 355 |
| | Csf2ra | M85078 | 171 | 372 | 402 | 254 |
| | Ebl3 | AF013114 | 187 | 270 | 428 | 148 |
| | Grid1 | D10171 | 128 | 164 | 150 | 257 |
| | Ifngr | J05265 | 141 | 263 | 327 | 251 |
| | Il2rg | U21795 | 205 | 184 | 231 | 477 |
| | Ldr | X64414 | 1399 | 1653 | 1665 | 3968 |
| | P40-8 | J02870 | 849 | 677 | 381 | 640 |
| | Plaur | X62701 | 312 | 443 | 476 | 734 |
| | Rarg | M34476 | 102 | 113 | 114 | 218 |
| | Srb1 | U37799 | 126 | 232 | 132 | 258 |
| | Cr2 | M28281 | 83 | 138 | 243 | 77 |
| | Csf2rb2 | M29855 | 209 | 249 | 437 | 111 |
| 3 or more, less than 4 | Fcgr1g | J05020 | 2388 | 2766 | 3365 | 8751 |
| | Fcgr2b | X04648 | 1703 | 1652 | 1431 | 4605 |
| | Ifngr2 | U69599 | 1 | 2 | 2 | 3 |
| 4 or more, less than 5 | Nr4a1 | X16995 | 66 | 188 | 202 | 401 |
| 5 or more | Il1r2 | X58769 | 482 | 1796 | 2872 | 3818 |
| | C5r1 | L05630 | 185 | 434 | 808 | 1078 |
| | Drd2 | X55674 | 0 | 0 | 0 | 219 |
| | Fcgr3 | M14215 | 1 | 1 | 1 | 2 |
| | Fpr1 | L22161 | 0 | 69 | 141 | 671 |
| | GCR | AA240711 | 2 | 0 | 0 | 0 |
| | L-CCR | AA034646 | 48 | 175 | 314 | 2056 |
| | NMDARGB | AAB20211 | 2 | 2 | 0 | 0 |
| | P2rx1 | X84896 | 79 | 346 | 530 | 744 |
| | Pira1 | U96682 | 0 | 43 | 172 | 378 |
| | Pira5 | U96686 | 274 | 391 | 954 | 1874 |
| | Pira6 | U96687 | 122 | 635 | 2014 | 1716 |
| | Pirb | U96689 | 191 | 445 | 966 | 747 |
| | Sell | M25324 | 46 | 104 | 570 | 20 |
| | Tcrq-V4 | M54996 | 1650 | 78 | 65 | 315 |

Receptors are identified as present whose maximal AD values were more than or equal to 200 U in this study. Genes were sorted by their expression patterns as follows: first by the average difference value, then by the difference between minimum and maximum AD for the 4 time points, and last by the alphabetical order of gene symbols. Genes were ordered according to the maximal fold change of AD values. Abbreviations of gene names are taken from gene symbols listed in the Locus Link portion of the National Center for Biotechnology Information database where available. Numbers in bold denote those gene expression patterns obtained by differential display rather than by oligonucleotide array assays. The other information is presented as in the legend to Table 2.

AD indicates average difference; gene symbols are expanded in an Appendix at the end of this article.

www.transfac.gbf-braunschweig.de/TRANSFAC) and determined which factors contained in this database were present at detectable levels in MPRO cell mRNA, using Affymetrix software for the criteria for inclusion of mRNAs from approximately 200 murine transcription factors probe sets on the oligonucleotide chip. Of these, 54 were expressed and 13 showed changes of 3-fold or more in chip signal (Table 5).

The changes in certain transcription factors, such as the moderate down-regulation of *myb* and *myc* and the up-regulation of the Max dimerization protein MAD, were consistent with the shift of the cells

from a proliferative to a differentiated state.²⁸ Some changes are more difficult to explain, such as the up-regulation of DP1, a partner for E2f factors in the regulation of S-phase genes, and the mild up-regulation of the *Id* genes, commonly associated with an inhibition of differentiation by competition with bHLH transcriptional activators.²⁹

The C/EBP family has been extensively studied with respect to myeloid differentiation.^{2,30} Absolute levels of the C/EBP α and δ mRNAs were low, probably at the borderline of significance for the oligonucleotide chip assay, whereas the level of C/EBP β appeared higher. In addition, there were discrepancies between the chip

Table 4. Granule constituents expressed during mouse promyelocytic cell line cell differentiation

| Granule constituent | Gene symbol | Gene accession | AD value by array | | | |
|------------------------------|-------------|----------------|-------------------|------|------|------|
| | | | 0 h | 24 h | 48 h | 72 h |
| Azurophil (primary) granules | | | | | | |
| | Man2c1 | AA161860 | 178 | 134 | 99 | 164 |
| | Ctsb | M65270 | 442 | 480 | 595 | 389 |
| | Ctsd | X52886 | 214 | 1087 | 1828 | 2784 |
| | Ctsf | M96801 | 1509 | 405 | 46 | 286 |
| | E12 | U04962 | 658 | 1273 | 843 | 157 |
| | E1a2 | AA589016 | 47 | 159 | 134 | 163 |
| | Gus-s | M63836 | 544 | 226 | 266 | 254 |
| | Lyzs | M21050 | 0 | 1 | 1 | 3 |
| | Mcpt8 | X78545 | 831 | 268 | 66 | 491 |
| | Mpo | X15378 | 3788 | 3009 | 776 | 692 |
| | Prg | X16133 | 2621 | 2653 | 2920 | 9859 |
| Possible granule proteins | | | | | | |
| | Ctsc | AA144887 | 252 | 194 | 342 | 576 |
| | Ctse | X97399 | 1 | 3 | 4 | 5 |
| | Ctsh | U06119 | 45 | 124 | 195 | 156 |
| | Ctsl | X06086 | 16 | 11 | 31 | 237 |
| | Ctsa | AA089333 | 12 | 9 | 88 | 463 |
| Specific secondary granules | | | | | | |
| | Cpa3 | J05118 | 621 | 270 | 90 | 601 |
| | Cd36l2 | AB008553 | 113 | 93 | 157 | 187 |
| | Cnlp | X94353 | 80 | 479 | 704 | 626 |
| | Cybb | U43384 | 8 | 24 | 91 | 128 |
| | Ear2 | --- | 0 | 1 | 1 | 2 |
| | Fpr1 | L22181 | 178 | 220 | 235 | 846 |
| | Itgb2 | X14951 | 0 | 2 | 4 | 2 |
| | Lcn2 | W13166 | 916 | 3513 | 3831 | 6036 |
| | Ltf | J03298 | 19 | 162 | 333 | 138 |
| | MBP | W45834 | 5 | 1 | 1 | 2 |
| | Mmp13 | X66473 | 44 | 43 | 72 | 178 |
| | Ngp | L37297 | 2661 | 4782 | 2311 | 6912 |
| Tertiary granules | | | | | | |
| | Mmp9 | Z27231 | 0 | 1 | 2 | 2 |

Shown are the possible granule protein cDNAs represented on the oligonucleotide arrays, sorted by their expression patterns as follows: first by the average difference AD value, then by the granule types, and last by the alphabetical order of gene symbols. Data are presented as described in the legend to Table 3.

AD indicates average difference; gene symbols are expanded in an Appendix at the end of this article.

estimates and the mRNA levels observed by Northern blotting with specific probes for these genes. In particular, the latter method, more sensitive and specific, showed that C/EBP α began to decline in the most mature cells, whereas C/EBP δ mRNA declined progressively beginning at 24 hours after the onset of differentiation.

C/EBP ϵ is a more recently cloned C/EBP family member. Previous studies indicated it is expressed in a large array of human leukemia cell lines blocked at various stages of differentiation and that it is up-regulated during granulocytic differentiation.³¹ A C/EBP ϵ probe was not included in the oligonucleotide chips, and this mRNA was not detected by DD. Therefore, we examined the C/EBP ϵ expression patterns by quantitative PCR and Northern blot analysis (Figure 4). C/EBP ϵ exon 1 was PCR amplified from MPRO RNAs using primers RY48 (AGCCCCCGACACCCCTTGATGA) and RY49 (TGGCACACT-GCGGGCAGACAG).³² The results showed that C/EBP ϵ is expressed throughout myeloid differentiation, with expression levels increased moderately in the later stages.

We detected a number of other transcription factors that are broadly expressed or that have been reported in other studies of hematopoiesis (Table 5). Some of the factors that were most strongly induced during differentiation have been studied in other contexts but not previously implicated in hematopoiesis, such as a mammalian homologue to the *Drosophila* enhancer of split gene, a transcriptional silencer. The mammalian gene is expressed at relatively high levels as measured by the oligonucleotide chip and

is a candidate for mediation of the silencing of growth-related genes in the maturing neutrophil. Another candidate transcriptional silencer, Tif1b, may serve as a corepressor for the KRAB domain family of zinc finger transcription factors and also may mediate binding of the heterochromatin protein HP1 to DNA.³³

There were 26 transcription factors whose mRNAs showed no significant changes by oligonucleotide chip analysis and were not identified as differentially regulated genes by differential display assays. PU.1, a factor necessary for the production of neutrophils and the expression of several neutrophil genes,³⁴ showed less than a 3-fold increase in mRNA, below the threshold for a significant change. Other candidate hematopoietic transcription factors, such as PEBP1aB2 (AML1), GATA-1, and SP-2, were represented on the oligonucleotide chips, but their mRNA levels were so low that they were reported as absent in this study. The possibility that small changes in the levels or ratios of some transcription factors could produce marked changes in transcription potentially limits the ability of data generated by present methods to explain transcriptional changes during differentiation.

Protein expression patterns of MPRO cells during ATRA induction

We visually compared the 2DE patterns from MPRO cells at the same time points used for mRNA analysis. In most cases the

Table 5. Transcription modulators presented during myeloid differentiation

| Maximal fold change | Gene symbol | Gene accession | AD value by array | | | |
|------------------------|-------------|----------------|-------------------|------|------|------|
| | | | 0 h | 24 h | 48 h | 72 h |
| Less than 2-fold | | | | | | |
| | Zfp11-6 | AB020542 | 2630 | 2989 | 2795 | 2515 |
| | Btf3 | W13502 | 3 | 3 | 2 | 1 |
| | Gata2 | AB000096 | 562 | 770 | 472 | 730 |
| | Hmg1 | J04179 | 337 | 348 | 177 | 232 |
| | Ikb1 | M31885 | 455 | 787 | 721 | 637 |
| | Max | M63903 | 256 | 224 | 312 | 172 |
| | Nfatc2 | AA560093 | 2313 | 3218 | 2396 | 2542 |
| | Pm1 | U33626 | 173 | 281 | 329 | 306 |
| | Rarg | M34476 | 102 | 113 | 114 | 218 |
| | Rela | M61909 | 297 | 260 | 304 | 244 |
| | Sox15 | W53527 | 419 | 461 | 484 | 637 |
| | Ybx1 | M62867 | 643 | 469 | 472 | 496 |
| | Zfp162 | Y12838 | 671 | 734 | 720 | 992 |
| 2 or more, less than 3 | | | | | | |
| | Cebpd | X61800 | 157 | 262 | 168 | 430 |
| | Ikb2 | M69293 | 244 | 210 | 310 | 604 |
| | Jund1 | W29356 | 1274 | 2002 | 1434 | 3085 |
| | Lyf1 | X57687 | 399 | 342 | 347 | 891 |
| | Nfe2 | L09600 | 458 | 743 | 1042 | 505 |
| | Nfkb1 | L28117 | 953 | 2044 | 1876 | 2034 |
| | Pbx1 | AF020186 | 611 | 303 | 345 | 212 |
| | slp1 | A34693 | 375 | 784 | 891 | 529 |
| | Ttf1b | U67303 | 673 | 659 | 420 | 863 |
| | Trp53 | P10361 | 259 | 149 | 125 | 361 |
| | Usf2 | U12283 | 120 | 185 | 285 | 192 |
| | Ybx3 | L35549 | 96 | 169 | 210 | 119 |
| | Zfp216 | AA510137 | 82 | 151 | 204 | 106 |
| 3 or more, less than 4 | | | | | | |
| | Irf1 | M21065 | 85 | 207 | 278 | 198 |
| | Klf2 | U25096 | 62 | 86 | 246 | 77 |
| | Myb | M12848 | 692 | 356 | 230 | 435 |
| | Stat3 | AA396029 | 484 | 1057 | 1012 | 290 |
| | Tfdp1 | Q08539 | 307 | 560 | 505 | 1093 |
| 4 or more, less than 5 | | | | | | |
| | Cebpb | X62600 | 390 | 1248 | 1380 | 1903 |
| | Str14 | Y07836 | 223 | 383 | 510 | 836 |
| 5 or more | | | | | | |
| | Cebpa | M62362 | 33 | 212 | 182 | 44 |
| | Grg | X73359 | 89 | 565 | 916 | 1005 |
| | Mad | X83106 | 0 | 111 | 167 | 327 |
| | Myc | L00039 | 314 | 112 | 62 | 173 |
| | Etch16 | W89667 | 169 | 386 | 313 | 1003 |
| | TBX1 | AA542220 | 0 | 0 | 1 | 2 |

Shown are the transcription factors identified as present by the oligonucleotide array analysis whose maximal AD between perfect match and mismatch oligonucleotide sets was greater than or equal to 200 U in this study. Data are presented as described in the legend to Table 3.

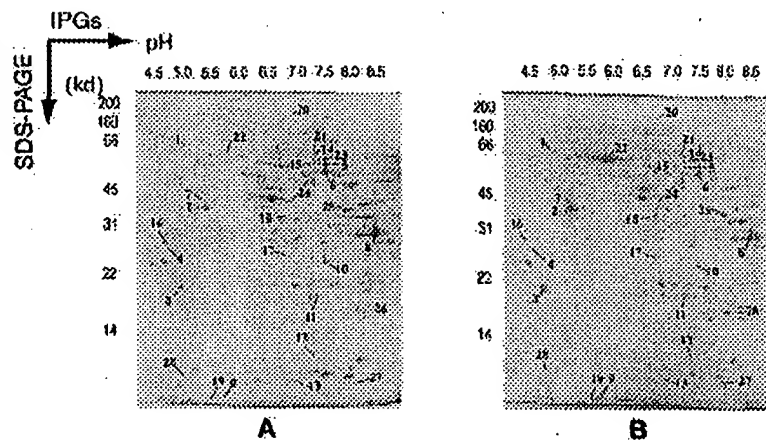
AD indicates average difference; gene symbols are expanded in an Appendix at the end of this article.

peptides identified for a given protein were derived from regions along the entire length of the protein, indicating the observed products were not the result of proteolytic degradation. These data must be considered with several caveats: membrane and other hydrophobic proteins and very basic proteins are not well displayed by the standard 2DE approach, and proteins present at low levels will be missed.³⁵ In addition, to simplify MS analysis, we used a Coomassie dye stain rather than silver to visualize proteins, and this decreased the sensitivity of detection of minor proteins. The MS method we used was sufficiently sensitive to identify proteins that could barely be visualized by colloidal blue staining. However, a limitation of the method for the mouse is that the current database lacks predicted amino acid sequences for a substantial fraction of murine genes. In addition, very small proteins give only a few peptides, making statistically confident identification difficult.

Figure 5 shows the analytical colloidal blue-stained 2DE IPG reference maps of differentiated MPRO cells. Expression patterns of more than 500 protein spots were detected and observed through the entire series of gels. Protein spots could easily be cross-matched to each other, indicating the reproducibility of the method. As marked on the gel pictures (Figure 5), 50 proteins with a wide range of molecular weights (1 to 200 kD), isoelectric points (4 to 9), and abundances were subjected to MS protein identification. The results are presented in Table 6.

Comparing the theoretical value of the molecular weight and *pI* of each protein to that of the observed value, we confidently identified 28 proteins in the expected position on the gels (spots 1 to 28). Some of the other proteins with strong matches to the murine databases migrated to a somewhat unexpected *pI* position. Nine spots gave clear peptide peaks on mass spectroscopy but did not match any known gene. Their identification will require amino acid

Figure 5. 2DE electrophoretograms of MPRO cells. MPRO cell lysate (2.5×10^6 cell/sample) was loaded for 2DE analysis. Gels were stained with brilliant blue G-collodial dye. (A) 2DE map of uninduced MPRO cell (0 hour). (B) 2DE map of matured MPRO cells (72 hours). Protein spots marked in the maps were considered differentially expressed and were subjected to MS analysis. The resultant protein information is listed in Table 6.



sequence analysis or availability of more extensive murine databases. We searched for the expression patterns of the genes cognate to the expressed proteins in dbMC (Table 6). Nineteen genes were found in dbMC, the mRNA for 5 genes was reported as absent, and 13 genes were present during MPRO differentiation. Comparison of the expression patterns showed only 4 genes of 18 present on the oligonucleotide chips whose expression was consistent at the RNA level and protein level. None of these was on the list of the genes

that were differentially expressed significantly (5-fold or greater change by array or 2-fold or greater change by DD).

Discussion

We explored the temporal patterns of gene expression during myeloid development. A database has been developed to provide a

Table 6. Correlation of expression patterns between mRNA level and protein level

| Spot | Protein definition | GI number | Predicted value | | Percentage (%) | 2DE pattern | | cDNA expression pattern | | Ag |
|------|--|-----------|-----------------|-----|----------------|-------------|------|-------------------------|--------|----|
| | | | kd | pI | | 0 h | 72 h | 0 h | 72 h | |
| 1 | GRP 78 | 2505545 | 72.4 | 5.1 | | 1 | 3 | 1321 | 1043.3 | N |
| 2 | Actin, gamma, cytoplasmic | 6752954 | 41.77 | 5.3 | 40 | 3 | 6 | 0 | 2 | Y |
| 3 | RHO GDI 2 | 2494703 | 22.83 | 4.9 | 33 | 3 | 3 | 341 | 441.6 | Y |
| 4 | Proliferating cell nuclear antigen | 7242171 | 28.77 | 4.7 | 42 | 1 | 0 | 544 | 430.9 | Y |
| 5 | APS kinase | 4038346 | 69.8 | 7.1 | 24 | 2 | 1 | 43 | 50.7 | N |
| 6 | Pyruvate kinase 3 | 6755074 | 57.9 | 7.2 | 48 | 6 | 4 | 3047 | 5880.3 | N |
| 7 | Melanoma X-actin | 6671509 | 41.72 | 5.3 | 39 | 1 | 3 | 2539 | 341.3 | N |
| 8 | Glyceraldehyde-3-phosphate dehydrogenase | 6579937 | 35.79 | 8.7 | 39 | 8 | 7 | 3073 | 5742.3 | N |
| 9 | Stein 3 | 461911 | 10.99 | 5.9 | 48 | 0 | 4 | N/A | N/A | — |
| 10 | Guanine nucleotide binding protein, beta-2, related sequence 1 | 6680047 | 35.06 | 7.9 | 21 | 4 | 2 | 139 | 303.1 | N |
| 11 | Triosephosphate isomerase | 6578413 | 26.69 | 6.9 | 26 | 3 | 3 | 3312 | 2650.1 | Y |
| 12 | Testis-derived c-abl protein | 1196524 | 17.19 | 7 | 51 | 2 | 3 | 152 | 126.9 | N |
| 13 | RNA binding motif protein 3 | 7945121 | 16.59 | 6.8 | 25 | 1 | 0 | 628 | 812.4 | N |
| 14 | Collapsin response mediator | 6681019 | 62.16 | 6.4 | 36 | 2 | 0 | Absent | Absent | N |
| 15 | Lamin A | 220474 | 47.52 | 6.6 | 35 | 2 | 0 | Absent | Absent | N |
| 16 | 47-kd keratin | 52783 | 35.82 | 4.8 | 29 | 3 | 0 | Absent | Absent | N |
| 17 | sid478p | 5931565 | 31.3 | 6.7 | 30 | 1 | 2 | Absent | Absent | N |
| 18 | MHC class II H2-IA-beta-5 | 3169662 | 28.6 | 7.1 | 39 | 1 | 2 | N/A | N/A | — |
| 19 | Androgen-binding protein: subunit alpha | 739346 | 8.04 | 6.4 | 68 | 0 | 2 | Absent | Absent | N |
| 20 | Neuronal apoptosis inhibitory protein | 5932010 | 158.7 | 6 | 17 | 1 | 0 | N/A | N/A | — |
| 21 | PAD type IV | 6755018 | 74.46 | 7.2 | 21 | 1 | 3 | N/A | N/A | — |
| 22 | Human serum albumin homologue | 3212625 | 66.45 | 5.7 | 24 | 0 | 6 | N/A | N/A | — |
| 23 | syncrtp | 6576815 | 62.53 | 7.2 | 33 | 2 | 1 | N/A | N/A | — |
| 24 | Transamidinase | 1730203 | 48.22 | 7.2 | 31 | 3 | 1 | N/A | N/A | — |
| 25 | PGK crgr phosphoglycerate | 1730519 | 44.54 | 8.3 | 47 | 5 | 4 | 1088 | 1402.3 | N |
| 26 | Proliferation-associated gene A | 6754976 | 22.16 | 8.6 | 53 | 3 | 1 | N/A | N/A | — |
| 27 | Putative peroxisomal antioxidant enzyme | 3913065 | 17 | 7.8 | 55 | 0 | 3 | N/A | N/A | — |
| 28 | IgE chain C2 region | 2137430 | 12.1 | 5.2 | 38 | 0 | 1 | N/A | N/A | — |

The proteins listed here are represented by the spots marked in the electrophoretograms shown in Figure 5.

Protein definition, GI number, and predicted value refer to the protein name, accession number, and properties derived from the National Center for Biotechnology Information protein database. The column labeled % shows the percentage of peptides predicted from the protein sequence that were detected by mass spectrometry. The expression level of protein spots expressed in mouse promyelocytic cell line cell induced by all-trans retinoic acid for 0 hours and 72 hours (Figure 5) were scored on a scale of 1 (+) to 8 (+++++). The cDNA expression patterns of the cognate mRNAs are listed in the cDNA expression pattern column abstracted from the dbMC database. The genes not represented on the oligonucleotide arrays were marked as N/A. Ag showed the correlation of gene patterns at mRNA level or protein level.

Y indicates agreement and N discrepancy between changes in cDNA and protein spot intensity. The numbers in bold were obtained with DD. 2DE indicates 2-dimensional gel electrophoresis; IgE, immunoglobulin E; DD, differential display.

reference for later research on the molecular mechanisms underlying normal myeloid development.

The MPRO cell system morphologically mimics normal myeloid differentiation and biochemically proceeds further toward mature neutrophils than most other *in vitro* systems. Because the arrest in differentiation of MPRO cells growing in the absence of ATRA is not physiologic, there is a theoretical risk that gene expression in these cells is not coordinated in the way that it is in normal differentiation. It is encouraging that, for the most part, the timing of expression of genes for proteins of the various neutrophil granules is consistent with the timing of the morphologic and biochemical appearance of these granule components during normal myeloid differentiation.

The DD technique provides certain advantages for detecting and comparing mRNA levels in different samples. First, the method is, in principle, similar to competitive RT-PCR, and, with the use of stringent PCR conditions, is expected to be about as reliable. Second, display patterns are reproducible. Third, the method detects the levels not only of RNAs already represented in the database but also of unknown RNA species that may represent "new" genes. Fourth, closely related genes can be distinguished regardless of cross-hybridization, provided there are some single nucleotide differences in the 3' end sequence. Limitations associated with this technique are that numerous gels are necessary to get complete information and that comparison of the levels of different mRNAs is only approximate because of the differential amplification of bands of different size or sequence.

Oligonucleotide chip analysis is a fast and effective means of accessing mRNA expression patterns.²⁰ Cluster analysis of groups of samples by this approach is effective. However, the present results indicate that alternative methods of verification are desirable before the data on an unexpected change in a particular gene are definitively accepted.

To obtain the broadest range of information from the myeloid differentiation process, both differential display and oligonucleotide chip techniques were applied in the current study. As a result, 65.3% of the observed changes in mRNA levels came from the differential display method and 41.5% came from oligonucleotide chip assays.

Our data showed in general that changes in expression pattern by the 2 methods agreed qualitatively but that there was some quantitative variation. Our results indicate that DD may be a more accurate way to detect changes in levels of gene expression than the oligonucleotide chip assay. However, improvements in the types of oligonucleotides used in arrays may close this gap in the future.

The mRNAs for a limited number of transcription factors vary in a pattern correlating with that of the mRNAs for primary or secondary granule proteins. However, more detailed information is needed, and the underlying mechanisms of granule gene regulation remain unclear. The number of potential positive and negative regulatory factors found here is sufficiently small as to make it feasible to perform *in vivo* studies, such as chromatin immunoprecipitation.

The oligonucleotide chip used in this study focused on known genes, whereas the DD method samples all polyadenylated transcripts. The latter method generated a large number of products not associated with known genes, in part because the mouse genome is not as well represented in the database as the human genome. However, our experience with DD and human mRNAs indicates that substantial fractions of the products represented as ESTs or not represented at all in the public databases are cDNA copies from introns, hnRNA, or other RNA with internal A runs.

Approximately 59 sequences obtained from gel-display bands had significant changes in the level of expression and a sequence that did not match that for any named gene in the public databases.

Of these, 38 had plausible or excellent polyA signals. This is only an approximate estimate of the number of new genes found³⁶ because a fraction of the mRNAs for known genes still had poor polyA signals. In addition, the full 3' untranslated region is often not known for characterized genes, and in some cases these new genes may prove to be identical to products identified by the oligonucleotide chips when more complete sequences are obtained. At the least, their presence indicates that a substantial fraction of the regulatory or functional circuitry of maturing myeloid cells remains unexplored and that valuable tools for their investigation will emerge from a combination of RNA expression studies and analysis of emerging genomic sequences.

The desired end point for the description of gene expression in a biologic system is not only the analysis of mRNA transcript levels but also the accurate measurement of protein abundance. The developments in 2DE and new MS instrumentation make it possible to accomplish this work rapidly and efficiently. In this study, we attempted to identify a number of the proteins differentially expressed between uninduced and ATRA-differentiated MPRO cells and to examine the relation between mRNA and protein expression levels for these genes representing the same state.

For protein levels based on estimated intensity of Coomassie dye staining in 2DE, there was poor correlation between changes in mRNA levels and estimated protein levels. Other groups have studied the correlation between mRNA and protein levels in yeast and liver cells.^{11,12,14} In the liver cell experiments,^{11,12} correlation coefficients of 0.4 to less than 0.5 were observed. In an extensive study in yeast,^{11,12} the correlation coefficient was high if the most abundant mRNAs and proteins were considered. If a handful of these products was omitted, the remaining correlation coefficient was 0.4 or less. However, one could restore some of the correlation by averaging individual data points into broad proteomic categories.³⁷

The discrepancies between mRNA and protein levels in MPRO cells appear to be substantially larger than those observed for yeast. Possible causes for the discrepancies include translational regulation, differential expression of certain mRNAs at various stages of cell growth *in vitro*, post-translational protein modification that varies with the stage of maturation of the cells, and selective degradation or excretion of proteins *in vivo*. Furthermore, here we are focusing on a developmental time-course, whereas the yeast study concentrated on the organism in vegetative growth. New techniques, equipment, and bioinformatic analysis tools must be developed to make such systematic, global, and quantitative analyses feasible.

The initial studies of protein expression presented here provide a cautionary note for efforts to interpret cell composition and function in relation to mRNA levels. Discrepancies we observed between gene expression and protein abundance suggest that selective post-transcriptional controls may be at least as important as changes in mRNA levels in determining the protein composition of neutrophils and that they are phenomena less well explored than transcriptional control. Analysis of mRNA expression patterns is itself only a small beginning toward a genome-wide description of cellular components.

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References

- Lawson ND, Berliner N. Neutrophil maturation and the role of retinoic acid. *Exp Hematol*. 1999; 27:1355-1367.
- Tenen DG, Hromas R, Licht JD, Zhang DE. Transcription factors, normal myeloid development, and leukemia. *Blood*. 1997;90:489-519.
- Sigurdsson F, Khanna-Gupta A, Lawson N, Berliner N. Control of late neutrophil-specific gene expression: insights into regulation of myeloid differentiation. *Semin Hematol*. 1997;34:303-310.
- Lenny N, Westendorf JJ, Hiebert SW. Transcriptional regulation during myelopoiesis. *Mol Biol Rep*. 1997;24:157-168.
- Yunis JJ, Tanzer J. Molecular mechanisms of hematologic malignancies. *Crit Rev Oncog*. 1993;4: 161-190.
- Velculescu VE, Zhang L, Vogelstein B, Kinzler KW. Serial analysis of gene expression. *Science*. 1995;270:484-487.
- Stollberg J, Urschitz J, Urban Z, Boyd CD. A quantitative evaluation of SAGE. *Genome Res*. 2000;10:1241-1248.
- Subrahmanyam YV, Baskaran N, Newburger PE, Weissman SM. A modified method for the display of 3'-end restriction fragments of cDNAs: molecular profiling of gene expression in neutrophils. *Methods Enzymol*. 1999;303:272-297.
- Subrahmanyam YVBK, Yamaga S, Newburger PE, Weissman SM. A modified approach for the efficient display of 3' end restriction fragments of cDNAs. In: Leslie RA, Robertson HA, eds. *Differential Display: A Practical Approach*. Practical Approach Series. Oxford, UK: Oxford University Press. 2000;101-129.
- Appella E, Arnot D, Sakaguchi K, Wirth PJ. Proteome mapping by two-dimensional polyacrylamide gel electrophoresis in combination with mass spectrometric protein sequence analysis. *EXS*. 2000;88:1-27.
- Anderson NL, Anderson NG. Proteome and proteomics: new technologies, new concepts, and new words. *Electrophoresis*. 1998;19:1853-1861.
- Gygi SP, Rochon Y, Franzos BR, Aebersold R. Correlation between protein and mRNA abundance in yeast. *Mol Cell Biol*. 1999;19:1726-1730.
- The Whitehead Institute for Biomedical Research/MIT Center for Genome Research. Molecular Pattern Recognition Web site. Available at: www.genome.wi.mit.edu/MPR/analysis.html#RS. Accessed May 4, 2001.
- Futcher B, Latter GI, Monardo P, McLaughlin CS, Gerrels JI. A sampling of the yeast proteome. *Mol Cell Biol*. 1999;19:7357-7368.
- Tsai S, Collins SJ. A dominant negative retinoic acid receptor blocks neutrophil differentiation at the promyelocyte stage. *Proc Natl Acad Sci U S A*. 1993;90:7153-7157.
- Johnson M, Calazzo T, Moline JM, Donahue R, Groopman J. Inhibition of bone marrow myelopoiesis and erythropoiesis in vitro by anti-retroviral nucleoside derivatives. *Br J Haematol*. 1988;70:137-141.
- Lawson ND, Krause DS, Berliner N. Normal neutrophil differentiation and secondary granule gene expression in the EML and MPRO cell lines. *Exp Hematol*. 1998;26:1178-1185.
- Prashar Y, Weissman SM. Analysis of differential gene expression by display of 3' end restriction fragments of cDNAs. *Proc Natl Acad Sci U S A*. 1995;93:659-663.
- Subrahmanyam YVBK, Yamaga S, Prashar Y, et al. RNA expression patterns change dramatically in human neutrophils exposed to bacteria. *Blood*. 2001;97:2455-2468.
- Chee M, Yang R, Hubbell E, et al. Accessing genetic information with high-density DNA arrays. *Science*. 1996;274:610-614.
- Lipshutz RJ, Chee M, Hubbell E, et al. Using oligonucleotide probe arrays to access genetic diversity. *Biotechniques*. 1995;19:442-447.
- Lipshutz RJ, Fodor SP, Gingeras TR, Lockhart DJ. High-density synthetic oligonucleotide arrays. *Nat Genet*. 1999;21:20-24.
- Mao M, Fu G, Wu JS, et al. Identification of genes expressed in human CD34(+) hematopoietic stem/progenitor cells by expressed sequence tags and efficient full-length cDNA cloning. *Proc Natl Acad Sci U S A*. 1998;95:8175-8180.
- Neuhoff V, Arnold N, Taube D, Ehrhardt W. Improved staining of proteins in polyacrylamide gels including isoelectric focusing gels with clear background at nanogram sensitivity using Coomassie Brilliant Blue G-250 and R-250. *Electrophoresis*. 1988;9:255-262.
- Wang Q, Miyakawa Y, Fox N, Kaushensky K. Interferon- α directly represses megakaryopoiesis by inhibiting thrombopoietin-induced signaling through induction of SOCS-1. *Blood*. 2000;96:2093-2099.
- Gullberg U, Bengtsson N, Bulow E, et al. Processing and targeting of granule proteins in human neutrophils. *J Immunol Methods*. 1999;232:201-210.
- Nichols J, Nimer SD. Transcription factors, translocations, and leukemia. *Blood*. 1992;80:2953-2963.
- Amati B, Land H. Myc-Max-Mad: a transcription factor network controlling cell cycle progression, differentiation and death. *Curr Opin Genet Dev*. 1994;4:102-108.
- Pagliuca A, Gello P, De Luca P, Lania L. Class A helix-loop-helix proteins are positive regulators of several cyclin-dependent kinase inhibitors' promoter activity and negatively affect cell growth. *Cancer Res*. 2000;60:1376-1382.
- Yamanaka R, Lekstrom-Himes J, Barlow C, Wynshaw-Boris A, Xanthopoulos KG. CCAAT/enhancer binding proteins are critical components of the transcriptional regulation of hematopoiesis (review). *Int J Mol Med*. 1998;1:213-221.
- Morosetti R, Park DJ, Chumakov AM, et al. A novel, myeloid transcription factor, C/EBP epsilon, is up-regulated during granulocytic, but not monocytic, differentiation. *Blood*. 1997;90:2591-2600.
- Yamanaka R, Barlow C, Lekstrom-Himes J, et al. Impaired granulopoiesis, myelodysplasia, and early lethality in CCAAT/enhancer binding protein epsilon-deficient mice. *Proc Natl Acad Sci U S A*. 1997;94:13187-13192.
- Nielsen AL, Ortiz JA, You J, et al. Interaction with members of the heterochromatin protein 1 (HP1) family and histone deacetylation are differentially involved in transcriptional silencing by members of the TIF1 family. *EMBO J*. 1999;18:6385-6395.
- Anderson KL, Smith KA, Perkin H, et al. PU.1 and the granulocyte- and macrophage colony-stimulating factor receptors play distinct roles in late-stage myeloid cell differentiation. *Blood*. 1999;94:2310-2318.
- Gong A, Obermaier C, Boguth G, Weiss W. Recent developments in two-dimensional gel electrophoresis with immobilized pH gradients: wide pH gradients up to pH 12, longer separation distances and simplified procedures. *Electrophoresis*. 1999;20:712-717.
- Wilusz J, Pettine SM, Shenk T. Functional analysis of point mutations in the AAUAAA motif of the SV40 late polyadenylation signal. *Nucleic Acids Res*. 1989;17:3899-3908.
- Jansen R, Gerstein M. Analysis of the yeast transcriptome with structural and functional categories: characterizing highly expressed proteins. *Nucleic Acids Res*. 2000;28:1481-1488.
- Tamayo P, Sionim D, Mesirov J, et al. Interpreting patterns of gene expression with self-organizing maps: methods and application to hematopoietic differentiation. *Proc Natl Acad Sci U S A*. 1999; 96:2907-2912.
- ProteinMetrics. ProFound search engine Web site. Available at: http://www.proteinmetrics.com/profound_bin/WebProFound.exe. Accessed May 4, 2001.

Appendix

Gene symbols used in tables: Actb: actin, beta, cytoplasmic; Actg: actin, gamma, cytoplasmic; Actr: melanoma X-actin; Aldol1: aldolase 1, A isoform; Arf5: ADP-ribosylation factor 5; Atf1: activating transcription factor 1; Atf2: activating transcription factor 2; Btl3: basic transcription factor 3a; Bzrp: peripheral-type benzodiazepine receptor; C5r1: complement component 5, receptor 1/G protein-coupled receptor (C5a); Cx2b2: cyclin B2; Cd36l2: CD36 antigen (collagen type I receptor, thrombospondin receptor)-like 2; Cd53: CDS3 antigen; Cebp: CCAAT/enhancer binding protein C/EBP, alpha; Cebp: CCAAT/enhancer binding protein (C/EBP), beta; Cebp: CCAAT/enhancer binding protein (C/EBP), delta; Cebp: CCAAT/enhancer binding protein (C/EBP), epsilon; Cfl1: cofilin 1, nonmuscle; Cmk2r: chemokine (C-X-C) receptor 4; Cmk2r1: chemokine (C-C) receptor 1/Mip1a receptor; Cnlp: cathelin-like protein; Cntr: ciliary neurotrophic factor/zinc finger protein PZF; Cpa3: carboxypeptidase A3, mast cell; Cr2: complement receptor 2; Crtr: corticotropin releasing hormone receptor; Crry: complement receptor-related protein; Csf1r: CSF 1 (M-CSF) receptor/c-fms/CD115; Csf2ra: CSF 2 (GM-CSF) receptor, alpha, low-affinity/CD116; Csf2rb1: CSF 2 (GM-CSF) receptor, beta 2, low-affinity/IL-3 receptor-like protein (AIC2B)/CDw131;

Csf2rb2: CSF 2 (GM-CSF) receptor, beta 2, low-affinity/IL-3 receptor (AIC2A); Ctsb: cathepsin B; Ctsc: cathepsin C; Ctsd: cathepsin D; Ctse: cathepsin E; Ctsf: cathepsin F; Ctsi: cathepsin G; Ctsj: cathepsin H; Ctsl: cathepsin L; Ctss: cathepsin S; Cybb: cytochrome b-245, beta; Drd2: dopamine receptor 2; E2f1: E2F transcription factor 1; E2f2: eosinophil-associated ribonuclease 2; Ebi3: Epstein-Barr virus-induced gene 3/cytokine receptor-like molecule (EBI3); El2: Balb/c neutrophil elastase; Ela2: elastase 2; Enh: enhancer of rudimentary homolog (Drosophila); Etah6: ethanol induced 6/steroid regulatory element binding transcription factor 1 (SREBF1) homolog; F2r12: coagulation factor II (thrombin) receptor-like 2; Fcgr1g: Fc receptor, IgE, high affinity I, gamma polypeptide; Fcgr2b: Fc receptor, IgG, low affinity IIb; Fcgr3: Fc receptor, IgG, low affinity III; Fpr1: formyl peptide receptor 1/MLP receptor; Gabpb1: GA repeat binding protein (GABP-beta1 subunit); Gata2: GATA-binding protein 2; Gnas: guanine nucleotide binding protein, alpha stimulating; Gmb2-rs1: guanine nucleotide binding protein, beta-2, related sequence 1; Gpx3: glutathione peroxidase 3; Grg: related to Drosophila groucho gene; Grid1: glutamate receptor channel subunit delta 1; Grn: granulins; Gstm1: glutathione-S-transferase, mu 1; Gus-s: beta-glucuronidase structural; Gys3: glycogen synthase 3, brain; H2-D: histocompatibility 2, D

region locus 1; Hist2: histone gene complex 2; Hist5-2ax: H2A histone family, member X; Hing1: high mobility group protein 1; Hsp60: heat shock protein, 60 kDa; Htr5a: 5-hydroxytryptamine (serotonin) receptor 5A; Idh1: inhibitor of DNA binding 1/helix-loop-helix DNA binding protein regulator (Id); Idh2: inhibitor of DNA binding 2; Ifngr: interferon gamma receptor; Ifngr2: interferon gamma receptor 2; Ii: Ia-associated invariant chain; Ili1a: IL1 alpha; Ili1r2: IL1 receptor, type II; Ili2rg: IL2 receptor, gamma chain; Ili4ra: IL4 receptor, alpha; Ili10rb: IL10 receptor, beta; Ili17r: IL17 receptor; Irfl: interferon regulatory factor 1; Irfl2: interferon regulatory factor-2; Itgb2: integrin beta 2 (Cd18); Itp5: inositol 1,4,5-trisphosphate receptor (type 2); Jund1: Jun proto-oncogene-related gene d1/transcription factor JUN-D; Klf2: Kruppel-like factor LKLF; L-CCR: lipopolysaccharide inducible C-C chemokine receptor-related; Lcn2: lipocalin 2; Ldlr: low density lipoprotein receptor; Lsp1: Lymphocyte-specific 1/S37/pp52; Lst1: leukocyte-specific transcript 1; Ltb4r: leukotriene B4 receptor; Ltr: lymphotxin-beta receptor; Ltf: lactotransferrin; Ly64: lymphocyte antigen 64; Ly6e: lymphocyte antigen 6 complex, locus E; Lyl1: lymphoblastic leukemia/bHLH factor; Lyzs: lysozyme; M6pr: mannose-6-phosphate receptor, cation dependent; Mad: Max dimerization protein; Man2c1: mannosidase, alpha, class 2C, member 1; Max: Max protein; Maz: MYC-associated zinc finger protein (purine-binding transcription factor); MBP: eosinophil granule major basic protein precursor; Mqpt8: mast cell protease 8; Mll: myeloid/lymphoid or mixed-lineage leukemia; Mmp13: matrix metalloproteinase 13/collagenase; Mmp9: matrix metalloproteinase 9/gelatinase B; Mpo: myeloperoxidase; Myb: myeloblastosis oncogene; Mybl2: myeloblastosis oncogene-like 2; Myc: myelocytomatosis oncogene; Myln: myosin light chain, alkali, nonmuscle; Nfatc2: nuclear factor of activated T cells, cytoplasmic 2; Nfe2: nuclear factor, erythroid-derived 2, 45 kDa; Nfkb1: NF-kappa-B (p105); Ngp: neutrophilic granule protein; NMDR8B: N-methyl-D-aspartate receptor glutamate-binding chain homolog; Npm1: nucleophosmin 1; Nr4a1: nuclear receptor subfamily 4, group A, member 1; Osi: oxidative stress induced; P2rx1: purinergic receptor P2X, ligand-gated ion channel, 1; P2ry2: purinergic receptor P2Y, G-protein-coupled 2; P40-8: P40-8, functional/laminin receptor; Pbx1: pre B-cell leukemia transcription factor 1; Pif: properdin factor, complement; Pira1: paired-Ig-like receptor A1; Pira5: paired-Ig-like receptor

A5; Pira6: paired-Ig-like receptor A6; Pirb: paired-Ig-like receptor B; Plaur: urokinase plasminogen activator receptor; PMI: putative receptor protein (SP: P17152); Pml: promyelocytic leukemia; Prg: proteoglycan, secretory granule; Prg3: proteoglycan 3/eosinophil major basic protein 2; Prtn3: proteinase 3; Psma2: proteasome (prosome, macropain) subunit, alpha type 2; Ptmb4: prothymosin beta 4; Ptpnc: protein tyrosine phosphatase, receptor type, C; Rac2: RAS-related C3 botulinum substrate 2; Rarg: retinoic acid receptor, gamma; Rela: avian reticuloendotheliosis viral (v-rel) oncogene homolog A/NF-kappa-B p65; Rpl19: ribosomal protein L19; RPL8: ribosomal protein L8; Rps6ka1: ribosomal protein S6 kinase polypeptide 1; Rps8: ribosomal protein S8; Rtn3: reticulon 3; S100a8: S100 calcium binding protein A8 (calgranulin A); S100a9: S100 calcium-binding protein A9 (calgranulin B); Sdf2: stromal cell-derived factor receptor 2; Sell: selectin L (lymphocyte adhesion molecule 1); Sema4d: semaphorin 4D; Sepp1: selenoprotein P, plasma, 1; Sfpi1: SFFV proviral integration 1; Shfdg1: split hand/foot deleted gene 1; Slc10a1: solute carrier family 10 (sodium/bile acid cotransporter family), member 1; Slpi: secretory leukocyte protease inhibitor; Sox15: SRY-box containing gene 15; Spi2-1: serine protease inhibitor 2-1; Srb1: scavenger receptor class B1; Stat3: signal transducer and activator of transcription 3; Stat5a: signal transducer and activator of transcription 5A; Stat6: signal transducer and activator of transcription 6; Stra14: basic-helix-loop-helix protein-retinoic acid induced; Thx1: TBX1 protein/LPS-induced TNF-alpha factor homolog; Tcrb: T-cell-receptor gamma chain beta-chain gene constant region; Tcrb-V4: T-cell-receptor gamma, variable 4; Tctex1: t-complex testis expressed 1; Tfdp1: transcription factor Dp 1; Tif1b: transcriptional intermediary factor 1, beta; Tlr4: toll-like receptor 4; Tnfisf1a: TNF receptor superfamily, member 1a; Tnfisf1b: TNF superfamily, member 1b; Tomm70a: translocase of outer mitochondrial membrane 70 (yeast) homolog A; Tpi: triosephosphate isomerase; Trp53: transformation-related protein 53; Ubb: ubiquitin B; Usf2: upstream transcription factor 2; Ybx1: Y box transcription factor; Ybx3: Y box binding protein; Zfp11-6: zinc finger protein s11-6; Zfp18: zinc finger protein 18 homolog; Zfp36: zinc finger protein 36; Zfp162: zinc finger protein 162; Zfp216: zinc finger protein 216; Zfpm1: zinc finger protein, multitype 1; Znfn1a1: zinc finger protein, subfamily 1A, 1 (Ikaro); Zyx: zyxin.

A Genomic and Proteomic Analysis of Activation of the Human Neutrophil by Lipopolysaccharide and Its Mediation by p38 Mitogen-activated Protein Kinase*

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Michael B. Fessler^{†§}, Kenneth C. Malcolm[§], Mark William Duncan[¶], and G. Scott Worthen^{†§¶}

From the [†]Department of Medicine, Division of Pulmonary Sciences and Critical Care Medicine, University of Colorado Health Sciences Center, the [§]Department of Medicine, National Jewish Medical and Research Center, and the [¶]Biochemical Mass Spectrometry Facility, School of Pharmacy, University of Colorado Health Sciences Center, Denver, Colorado 80262

Bacterial lipopolysaccharide (LPS) evokes several functional responses in the neutrophil that contribute to innate immunity. Although certain responses, such as adhesion and synthesis of tumor necrosis factor- α , are inhibited by pretreatment with an inhibitor of p38 mitogen-activated protein kinase, others, such as actin assembly, are unaffected. The aim of the present study was to investigate the changes in neutrophil gene transcription and protein expression following lipopolysaccharide exposure and to establish their dependence on p38 signaling. Microarray analysis indicated expression of 13% of the 7070 Affymetrix gene set in nonstimulated neutrophils, and LPS up-regulation of 100 distinct genes, including cytokines and chemokines, signaling molecules, and regulators of transcription. Proteomic analysis yielded a separate list of up-regulated modulators of inflammation, signaling molecules, and cytoskeletal proteins. Poor concordance between mRNA transcript and protein expression changes was noted. Pretreatment with the p38 inhibitor SB203580 attenuated 23% of LPS-regulated genes and 18% of LPS-regulated proteins by $\geq 40\%$. This study indicates that p38 plays a selective role in regulation of neutrophil transcripts and proteins following lipopolysaccharide exposure, clarifies that several of the effects of lipopolysaccharide are post-transcriptional and post-translational, and identifies several proteins not previously reported to be involved in the innate immune response.

Lipopolysaccharide (LPS),¹ a component of the outer cell wall of Gram-negative bacteria, evokes a variety of functional responses in the human neutrophil (PMN) after binding to a plasma membrane receptor complex that involves the Toll-like

receptors (TLRs) (1–5). These “immediate” functional responses, including actin assembly, adhesion, activation of nuclear factor-kappa B (NF- κ B), and priming for an enhanced secretory response and for release of reactive oxygen intermediates, appear to be central both to the innate immune response and to the pathogenesis of several inflammatory human diseases, including sepsis and the acute respiratory distress syndrome (6). p38 mitogen-activated protein kinase (p38 MAPK) has been shown to mediate LPS-induced PMN adhesion, NF- κ B activation, and TNF- α and IL-8 translation and release (7), and its blockade attenuates LPS-induced PMN accumulation in the airspace (8). However, other cascades almost certainly lead to downstream effectors of the LPS signal; for example, actin assembly appears to be p38 MAPK-independent (9). An improved understanding of the transcriptional and translational responses of the neutrophil to LPS and the modulation of these responses by p38 MAPK might carry pathogenetic and therapeutic implications.

Historically, it has been believed that the downstream PMN transcriptional response to LPS is static and that PMN functional responses to LPS that depend on *de novo* protein synthesis are primarily limited to the release of cytokines (10). However, recent studies indicate a robust transcriptional response (11). To date, most studies have relied upon and reported a short list of functional assays of the LPS-exposed PMN; therefore, no exhaustive investigation of either the transcriptional response or protein synthetic repertoire of the PMN has been reported. Although several techniques have been used to evaluate transcripts, the screening of global changes in mRNA by microarray analysis has only recently become possible. In this way, thousands of genes can be screened in an unbiased fashion for transcript abundance. Such genomic screens in mammalian cells have previously been applied to define altered expression profiles in response to agonists (12) and to drug action (13) and during cell cycle progression (14).

Although DNA microarray technology is expected to provide insight into the response of the human PMN to LPS (15), inhibition of LPS-stimulated IL-1 and TNF- α production by p38 MAPK inhibitors in THP-1 cells (16) and of TNF- α synthesis in human PMNs (9) occurs at a translational level and would therefore not be detected by DNA microarrays. Furthermore, in other systems, such as yeast and human liver, mRNA and protein levels show poor correlation (17, 18). Proteomics is a complementary tool for assessing global changes in cellular protein expression, thereby providing additional insight into cellular signal regulation. A proteomic approach has proven useful in different systems for dissecting signal transduction cascades and describing their output (19, 20) and has even

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[†] To whom correspondence should be addressed: Dept. of Medicine, D403, Neustadt Bldg., National Jewish Medical and Research Center, 1400 Jackson St., Denver, CO 80206. Tel.: 303-398-1171; Fax: 303-398-1381; E-mail: worthens@njc.org.

¹ The abbreviations used are: LPS, lipopolysaccharide; DTT, dithiothreitol; IEF, isoelectric focusing; IFN, interferon; IL, interleukin; MALDI-TOF, matrix-assisted laser desorption/ionization-time of flight; MAPK, mitogen-activated protein kinase; NF- κ B, nuclear factor-kappa B; pI, isoelectric point; PMN, neutrophil (polymorphonuclear leukocyte); TLR, Toll-like Receptor; TNF, tumor necrosis factor; CHCA, α -cyano-4-hydroxycinnamic acid; AERSEF, 4-(2-aminoethyl)-benzenesulfonyl fluoride hydrochloride; MS, mass spectrometry; CaM, Ca²⁺-calmodulin; ERK, extracellular signal-regulated kinase; E-64, epoxysuccinyl-64.

recently been used to detect novel upstream messengers involved in LPS signal transduction (21). We have applied DNA microarrays and proteomics to define and compare transcriptional and post-transcriptional alterations in the LPS-exposed PMN and to establish the dependence of these alterations on p38 MAPK signaling.

EXPERIMENTAL PROCEDURES

Materials—Endotoxin-free reagents and plastics were used in all experiments. Aprotinin, leupeptin, AEBSF, E-64, pepstatin, and bestatin protease inhibitors, spermine HCl, and α -cyano-4-hydroxycinnamic acid (CHCA) were all purchased from Sigma Chemical Co. (St. Louis, MO). SB203580, a p38 MAPK inhibitor, was purchased from Calbiochem-Novabiochem Corp. (San Diego, CA). For two-dimensional PAGE, rehydration buffer, equilibration buffers, vertical electrophoresis solutions, and 10% homogeneous polyacrylamide slab gels were purchased from Genomic Solutions, Inc. (GSI, Ann Arbor, MI). Sequencing grade porcine trypsin was purchased from Promega (Madison, WI).

LPS Incubation—PMNs were isolated by the plasma Percoll method (22), a technique that yields less than 5% monocytic contamination, and resuspended at a concentration of 15.4×10^6 /ml in RPMI 1640 culture medium (BioWhittaker, Walkersville, MD) supplemented with 10 mM HEPES (pH 7.6) and 1% heat-inactivated platelet-poor plasma. After addition of 100 ng/ml *Escherichia coli* 0111:B4 LPS (List Biological), incubation was carried out with continuous rotation (4 h, 37 °C) both in the presence and absence of SB203580. Both Affymetrix analysis and proteomic analysis utilized 75×10^6 cells. For microarray analysis, nonstimulated and 4-h-treated PMNs were collected from three separate donors. A more detailed time course following LPS exposure was performed using polymerase chain reaction. For proteomic analysis, LPS incubations from separate donors ($n = 6$) were performed and then analyzed individually. Control and post-LPS incubation PMNs were washed (0.34 M sucrose/1 mM EDTA/10 mM Tris) and then lysed in a modified rehydration buffer (GSI, Ann Arbor, MI) supplemented with 2 M thiourea, 50 mM dithiothreitol (DTT), 22.5 mM spermine HCl, and a mixture of six protease inhibitors (10 μ g/ml aprotinin, 10 μ g/ml leupeptin, 2 mM AEBSF, 5 μ M E-64, 1 μ M pepstatin, 10 μ M bestatin). DNA was pelleted by centrifugation at 250,000 $\times g$ for 60 min (23).

Affymetrix Oligonucleotide Array—Five micrograms of total RNA was isolated with TRIzol (Invitrogen) and RNeasy columns (Qiagen) and subsequently labeled with biotin as described by Affymetrix. Briefly, first-strand synthesis was accomplished with Superscript II reverse transcriptase (Invitrogen) using a T7-oligo(dT)₂₄ primer for 1 h at 42 °C followed by second-strand synthesis using *E. coli* DNA polymerase I and RNase H (Invitrogen) at 16 °C for 2 h. Double-stranded DNA was used as a template for *in vitro* transcription with T7 RNA polymerase in the presence of biotin-labeled UTP and CTP using the BioArray High Yield RNA transcript labeling kit (Enzo). Fifteen micrograms of cRNA was fragmented and used for hybridization to Affymetrix HuGene 6800FL Genechips. Each sample was hybridized initially using a Test2 Genechip to test for sample degradation and full-length *in vitro* translation. Data were analyzed using Affymetrix Genechip software. Results from three separate donors were analyzed.

Reverse Transcription and Polymerase Chain Reaction—cDNA was prepared by reverse transcription using 2 μ g total RNA, derived from 20×10^6 cells that were treated as indicated. Polymerase chain reactions were performed using specific primers for *Mx-1*, *TNF- α* , *MCP-1*, *p65*, *S100A4*, and glyceraldehyde-3-phosphate dehydrogenase.

Two-dimensional PAGE—The protein concentration of the lysates was measured as described by Bradford *et al.* (24). Poor isoelectric focusing (IEF) results were encountered unless the polycationic spermine was diluted (data not shown); therefore, lysates were diluted with rehydration buffer (GSI, Ann Arbor, MI) to achieve a final spermine concentration of 6 mM. Equal protein loads (1.5 mg) of control and LPS-stimulated neutrophils were used to rehydrate IEF gels overnight (18 cm, pH 3–10 nonlinear Immobiline DryStrip IEF gels, Amersham Biosciences; Piscataway, NJ). IEF was performed at 20 °C to 100-kVh (Phaser, GSI) under mineral oil, followed by two 10-min SDS equilibration steps (DTT and then iodoacetamide-containing equilibration buffers, GSI) and then by vertical electrophoresis on 10% homogeneous polyacrylamide slab gels (GSI) at 500 V. Protein spots were visualized by agitation in colloidal Coomassie Brilliant Blue G-250 (16 h) (25), followed by destaining in deionized water (20 h). In separate experiments, control and LPS-stimulated PMN lysates from three donors were pooled and then analyzed by two-dimensional PAGE using overlapping narrow isoelectric point (pI) ranges (18 cm, pH 5.0–6.0, 6.5–

6.7, and 6–11, Amersham Biosciences, Piscataway, NJ). Identical IEF and vertical electrophoresis parameters were used for all gels.

Image Analysis of Two-dimensional Gels—Colloidal Coomassie-stained gels were digitized using a Powerlook II (UMAX Data Systems, Inc., Taiwan) flatbed scanner with 8-bit dynamic range and 150-dpi resolution. BioImage (GSI, Ann Arbor, MI) 2D-Analyzer software was used to locate, quantitate, and match protein spots on the control and LPS gel images. Analysis was performed by assigning 50 common anchor spots between paired images; the remaining spots were compared by a constellation-matching algorithm. All data were then carefully reviewed by the operator to account for any discrepancies. Protein loading between control and experimental gels may have varied because of inconsistencies in rehydration of the different IEF gel strips; therefore, gel images were normalized so that the sum of the integrated intensities of all matched spots on paired gels was made equal. Control and LPS-stimulated gel images from individual donor experiments were matched to generate composite images; composite images were then matched into a master composite image to track the LPS response of protein spots among different donors (26). Only those spots that were common (image-matched) to all original 12 (pH 3.0–10.0) gels were considered for further analysis. For these spots, the LPS-induced change in integrated intensity in the six experiments was subjected to statistical analysis with a two-tailed Student's *t* test, and those spots with $p < 0.05$ were identified by peptide mass fingerprinting (described below). For the narrow range (pH 5.0–6.0, 5.5–6.7, and 6–11) two-dimensional PAGE experiments using pooled donors, only those spots with concordant regulation exceeding 1.5-fold or that appeared *de novo* in the LPS gel in two repeat experiments were further analyzed.

In-gel Tryptic Digestion—In-gel digestion of protein spots was performed with sequencing grade porcine-modified trypsin using the method of Hellman *et al.* (27). Tryptic peptides were then extracted (50 μ l of 50% acetonitrile/5% trifluoroacetic acid, 2 h), and the supernatant was taken to dryness in a vacuum centrifuge and then redissolved in trifluoroacetic acid (20 μ l, 0.5%). Peptides were then purified and concentrated using ZipTip_{C18} pipette tips (Millipore, Bedford, MA).

MALDI-TOF Mass Spectrometry—Analyses were performed on an Applied Biosystems matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) Voyager-DE PRO mass spectrometer (Framingham, MA) operated in delayed extraction mode. Samples (0.5 μ l) were spotted onto a sample plate to which matrix (0.5 μ l of 10 mg/ml CHCA) was added. The sample-matrix mixture was dried at room temperature and then analyzed in reflector mode. CHCA was also spotted alone as a negative control. Spectra were the sum of 100 laser shots, and those peaks with a signal-to-noise ratio of greater than 3:1 were selected for data base searching. Spectra were internally calibrated using autolytic trypsin peptides (m/z 842.51, 2211.10).

Data Base Searching Algorithm—The monoisotopic masses for each protonated peptide were: (a) entered into the program MS-Fit (available at prospector.ucsf.edu) for searches against the Swiss-Prot, NCBI, and GenPept databases, and (b) entered into Mascot (available at matrixscience.com), an algorithm testing statistical significance of peptide mass fingerprinting identifications. For MS-Fit searches, masses derived from trypsin, CHCA, keratin, and Coomassie Brilliant Blue G-250 were excluded. Search parameters included a maximum allowed peptide mass error of 0.1 Da (0.8 Da in the few instances in which linear mode was used), consideration of one incomplete cleavage per peptide, pI range of 3.0–10.0, and molecular mass range of 1–200 kDa. Accepted modifications included carbamidomethylation of cysteine residues (from iodoacetamide exposure following IEF) (28) and methionine oxidation, a common modification occurring during SDS-PAGE (29). Protein identifications were assigned when three criteria were met: 1) statistical significance ($p < 0.05$) of the match when tested by Mascot (matrixscience.com); 2) >20% sequence coverage by the tryptic peptides; and 3) concordance ($\pm 15\%$) with the molecular weight and pI of the parent two-dimensional PAGE protein spot. The following special exceptions were considered: (a) protein identifications not fulfilling criterion 2 were still assigned if criteria 1 and 3 were fulfilled and no other *Homo sapiens* proteins with peptide mass-matched p values < 0.05 were identified by Mascot; (b) if criterion 3 was not fulfilled (lower than expected molecular weight), a cleavage product of the identified protein was inferred, and the cumulative molecular weight of the tryptic peptides was compared with that of the two-dimensional-PAGE spot to ensure that it was not exceeded; (c) if criterion 3 was not fulfilled (isolated discordance between theoretical and observed pI), post-translational modification of an unrecovered peptide was inferred; and (d) if two or more *H. sapiens* protein assignments with >4 mutually exclusive matching peptides were identified, a protein mixture in the two-dimensional PAGE

spot was inferred and further analysis halted (quantitative conclusions regarding the individual protein constituents could not be drawn).

RESULTS

Genes Differentially Expressed in LPS-stimulated Neutrophils—Human PMNs were left untreated or incubated in the presence of 100 ng/ml LPS for 4 h. As a control to confirm that the PMNs were quiescent at baseline and that LPS resulted in normal stimulation, mRNA was isolated, cDNA was prepared, and PCR for TNF- α was performed. Little TNF- α expression was seen in nonstimulated cells, whereas LPS treatment led to an increase in expression in each of the donors subsequently used for microarray analysis (data not shown). No macrophage-colony stimulating factor receptor transcript was detected by oligonucleotide microarray analysis, confirming there was no significant monocytic contamination.

Human PMNs express a limited repertoire of mRNA transcripts at baseline but respond to LPS with differential expression of genes in many families. Considering only those genes present by microarray analysis in all three donors, unstimulated PMNs expressed 13.0% (923 of 7070 genes) of the Affymetrix gene set. Gene classes represented at baseline include metabolic enzymes, structural proteins, receptors, signaling proteins, and transcription factors. By comparison, human monocytes expressed ~40% and human fibroblasts ~35% of the represented genes (data not shown). By the criterion of a >3-fold increase in expression in all three donors on Affymetrix oligonucleotide array analysis, exposure of PMNs to LPS for 4 h resulted in the up-regulation of 100 genes (Table I).

Genes from several different functional classes were induced in PMNs following LPS exposure. Of interest, a number of transcriptional regulators were induced, including transcription factors of the NF- κ B family. The transcriptional NF- κ B complex has previously been implicated in the regulation of the genes induced by LPS (11). The genes for several cytokines and chemokines were also found to be up-regulated. These include TNF- α , IL-1 β , IL-6, MCP-1, MIP-3 α , and MIP-1 β (Table I). PCR was performed to confirm the results from the microarray analysis. PCR analysis on selected genes indicates that the time course for changes can be rapid or delayed but parallel the changes found in the array at the 4-h time point (data not shown). Other up-regulated genes included those for metabolic enzymes, immune response molecules, kinases, phosphatases, signaling molecules, adhesion and cytoskeletal components, interferon-stimulated genes, and those with unknown or miscellaneous function (Table I).

LPS stimulation of PMN also resulted in the down-regulation of 56 genes (Table II). Down-regulated genes were identified as transcriptional regulators, protein and lipid kinases and phosphatases, structural molecules, and signaling molecules. Genes for metabolic proteins were also evident, as were several uncharacterized genes.

Two-dimensional PAGE and Image Analysis—In contrast to the limited number of transcripts found at baseline, PMNs were found to express a large number and variety of proteins in the nonstimulated state (Fig. 1, A and C, and Tables III–V). Reproducible protein expression patterns were found on the pH 3.0–10.0 gels, and the majority of proteins fell in the pH 5.0–7.0 range (Fig. 1A). The basic region (pH > 7.0) consistently exhibited poor resolution, precluding meaningful image analysis and further workup (data not shown). Depending on the spot-finding parameters (minimum spot intensity, filter width) selected on the image analysis software, spot-by-spot manual editing was found to be necessary to avoid over- and under-detected spots; moreover, further manual editing was performed to screen for unmatched and mismatched spots following matching of paired control and LPS-stimulated gels. After spot

editing, ~1200 well-resolved spots were evident on each pH 3.0–10.0 gel. In an attempt to improve resolution of the pH range bearing the greatest number of well-resolved spots, overlapping narrow pH range gels (pH 5.0–6.0, 5.5–6.7, 6–11) were also run. Of interest, a similar number of well-resolved spots (~1200) were detected on the narrow pH range gels (Fig. 1, C and D). Assuming a detection limit for Coomassie of 15 ng (0.25 pmol, or 1.5×10^{11} molecules, for a 60-kDa protein) and a protein load per gel corresponding to 75×10^6 PMNs, we estimate a detection limit on our gels of 2000 molecules/cell for a 60-kDa protein. As investigators have suggested in other cell lines with the use of high resolution two-dimensional-PAGE methods (30), we estimate that >10,000 proteins are expressed in the resting PMN.

Human PMNs respond to LPS with the differential expression of a large number of proteins. In the six individual pH 3.0–10.0 experiments, the number of protein spots that increased in integrated intensity by at least 50% following LPS exposure was 185, 122, 104, 104, 96, and 131, respectively. The number of protein spots that decreased by at least 50% following LPS exposure was 72, 151, 102, 98, 128, and 97, respectively. Although gel-to-gel regional variability in resolution was expected to account for individual spots not being well visualized on particular gels, only those spots that were matched to all 12 original gels were analyzed further. Overall, the number of spots matched to all 12 original gels was 125. The numbers of spots that were both matched to all 12 original gels and that increased by at least 50% in integrated intensity in the individual experiments following LPS exposure were 46, 13, 17, 27, 22, and 20, respectively. The numbers of spots that were matched to all 12 gels and that decreased by at least 50% were 6, 22, 17, 22, 34, and 28, respectively. The LPS-induced change in integrated intensity of the 125 spots that were matched to all 12 original gels was subjected to statistical analysis with a two-tailed Student's *t* test, and those spots with statistically significant ($p < 0.05$) regulation among the six experiments were identified by peptide mass fingerprinting (Table III).

Identification of LPS-regulated Proteins—Several proteins were consistently up-regulated on the pH 3.0–10.0 gels (Table III), including regulators of inflammation (annexin III) and signaling molecules (Rab-GDP dissociation inhibitor β). Several actin fragments were seen to be consistently up-regulated in the six experiments following LPS exposure (Table III). Of interest, the proteasome β chain was also consistently up-regulated. Down-regulated proteins included other signaling molecules, such as Rho GTPase activating protein 1.

On the pH 5.0–6.0 and 5.5–6.7 gels, several proteins were found to show increases of greater than 1.5-fold following LPS exposure (Tables IV and V), including cytoskeletal proteins, such as moesin, nonmuscle myosin heavy chain, and a putative phosphorylated form of nonmuscle myosin heavy chain, and signaling molecules, such as protein phosphatase 1 and PO $_4$ -stathmin. The putative phosphorylated form of nonmuscle myosin heavy chain (spot #1101) was positioned 0.03 pH unit more acidic than the unmodified protein (spot #1102) (Fig. 1D) and was distinguished by a tryptic peptide (m/z 1366.74) not present in the unmodified protein, consistent with phosphorylation of serine 685. Serine 685 is predicted by NetPhos 2.0 Prediction Server (available at www.cbs.dtu.dk/services/NetPhos/31) to be a high probability phosphorylation residue and by ScanProsite (www.expasy.ch/tools/scnpsite.html) to be a substrate for protein kinase C. The tryptic phosphopeptide identified in PO $_4$ -stathmin, extending from residues 15 to 27 (1468.7 Da), is consistent with phosphorylation of either serine 16, a known substrate for Ca $^{2+}$ /calmodulin (CaM)-dependent kinases (32), or serine 25, a known substrate for p38 δ and ERK (Fig. 2A)

TABLE I
Human neutrophil genes induced after 4 h of LPS exposure

| Description | GenBank™ no. | Change-fold |
|--|---------------|-------------|
| Transcriptional regulation | | |
| <i>Pleiomorphic adenoma gene-like 2</i> | D83784 | 16.8 |
| <i>NFKB2</i> | S76638 | 12.3 |
| <i>NFKB1E</i> | U91616 | 11.5 |
| <i>p65</i> | L19067 | 8.4 |
| <i>BCL3</i> | U05681 | 7.7 |
| <i>X-box binding protein 1</i> | M31627 | 7.5 |
| <i>Metal-regulatory transcription factor 1</i> | X78710 | 7.4 |
| <i>Ets-2</i> | J04102 | 7.4 |
| <i>c-Rel</i> | X75042 | 6.2 |
| <i>NFKB1</i> | M58603 | 5.8 |
| <i>Basic leucine zipper transcription factor, ATF-like</i> | U15460 | 4.7 |
| <i>IKB</i> | M69043 | 3.8 |
| <i>MAX dimerization protein</i> | L06895 | 3.6 |
| <i>DIF2</i> | S81914 | 3.1 |
| Cytokines and receptors | | |
| <i>MCP-1</i> | M69203 | 78.7 |
| <i>MIP-1β</i> | M72885 | 48.8 |
| <i>αHelix coiled-coil rod homolog</i> | AF014958 | 20.8 |
| <i>IL-1β</i> | X04500 | 17.6 |
| <i>GRO3 (beta)</i> | M57731 | 17.3 |
| <i>TNF-α</i> | X02910 | 14.5 |
| <i>MIP-3α</i> | U64197 | 8.1 |
| <i>IL10RA</i> | U00672 | 7.3 |
| <i>IL-6</i> | Y00081 | 6.3 |
| <i>GROα</i> | X54489 | 4 |
| <i>HM74</i> | D10923 | 3.8 |
| Immune response | | |
| <i>Orosomucoid</i> | X02544 | 20.2 |
| <i>Complement component C3</i> | K02765 | 12.8 |
| <i>Protease inhibitor 9</i> | U71364 | 9.5 |
| <i>Complement component 3a receptor 1</i> | U28488 | 6.1 |
| <i>Protease inhibitor 3</i> | L10343 | 4.9 |
| <i>SLPI/antileukoprotease</i> | X04470 | 4.7 |
| <i>ELANH2/elastase inhibitor</i> | M93056 | 4.6 |
| <i>CD58</i> | Y00636 | 3.8 |
| <i>Complement component PFC</i> | M83652 | 3.5 |
| Kinases | | |
| <i>CNK/FNK/PLK-like</i> | U56998 | 16.2 |
| <i>Cot</i> | D14497 | 11.9 |
| <i>Pim-2</i> | U77735 | 9.5 |
| <i>LIMK2</i> | D45906 | 4.3 |
| Phosphatases | | |
| <i>PAC-1/DUSP2</i> | L11329 | 11.8 |
| <i>DUSP5</i> | U15932 | 5.3 |
| <i>PHA1</i> | U73477 | 3.4 |
| Signaling molecules | | |
| <i>TNFAIP1/A20</i> | M59465 | 10 |
| <i>TRAF1</i> | U19261 | 6.2 |
| <i>RanBP2</i> | D42063 | 5.6 |
| <i>GNA15</i> | M63904 | 5.2 |
| <i>PTAFR</i> | D10202 | 3.9 |
| Adhesion and cytoskeleton | | |
| <i>ICAM1</i> | M24283 | 22.4 |
| <i>CEACAM1 (biliary glycoprotein)</i> | X16354 | 6.3 |
| <i>LIMS1</i> | U09284 | 6.1 |
| <i>SNL/actin bundling protein</i> | U03057 | 5.9 |
| <i>Galectin-1/LGALS1</i> | M57710 | 4.7 |
| <i>MEMD/ALCAM</i> | U30999 | 4.2 |
| <i>CD44</i> | HG2981—HT3125 | 3.9 |
| <i>TSG-6</i> | M31165 | 3.7 |
| Metabolic | | |
| <i>GTP cyclohydrolase 1</i> | U19523 | 13.5 |
| <i>NDUFV2/ubiquinone reductase</i> | M22538 | 8.6 |
| <i>PSMA6/proteasome iota</i> | X59417 | 8.4 |
| <i>UDP-galactose transporter (SLC35A2)</i> | D84454 | 7.3 |
| <i>PLAU (urokinase)</i> | X02419 | 6.4 |
| <i>KYNU/kynurenine hydrolase</i> | U57721 | 5.5 |
| <i>AMPD3</i> | D12775 | 5 |
| <i>P4HA1/prolyl 4-hydroxylase</i> | M24486 | 4.7 |
| <i>γ Glutamylcysteine synthetase</i> | L35546 | 4.5 |
| <i>ATP6D</i> | J05682 | 4.2 |
| <i>ATP6S1</i> | D16469 | 4 |

TABLE I—continued

| Description | GenBank™ no. | Change-fold |
|---|--------------|-------------|
| <i>Glycerol kinase</i> | K68285 | 3.6 |
| <i>FACL1</i> | L09229 | 3.5 |
| <i>AK3</i> | X60673 | 3.3 |
| Interferon-inducible | | |
| <i>ISG15</i> | M13755 | 22.5 |
| <i>Mx1</i> | M33882 | 19.4 |
| <i>IFI56</i> | M24594 | 12.1 |
| <i>INDO</i> | M34455 | 5.2 |
| <i>GBP1</i> | M55542 | 4.3 |
| <i>PRKR</i> | U50648 | 3.7 |
| <i>IFIT4</i> | U52513 | 3.6 |
| <i>IFI54</i> | M14660 | 3.5 |
| <i>IFI58</i> | U34605 | 3.5 |
| <i>IFP35</i> | U72882 | 3 |
| Other | | |
| <i>Gos2</i> | M72895 | 48.8 |
| <i>MIHC/IAP1</i> | U37546 | 7.2 |
| <i>KIAA0105</i> | D14661 | 5.1 |
| <i>KIAA0118</i> | D42087 | 5 |
| <i>SNAP23</i> | U55936 | 5 |
| <i>CASP5</i> | U28015 | 4.8 |
| <i>KIAA0113</i> | D30755 | 4.8 |
| <i>KIAA0255</i> | D87444 | 4.7 |
| <i>Hepatitis-derived GF</i> | D16431 | 4.7 |
| <i>PTGS2</i> | D28235 | 4.6 |
| <i>CD48</i> | M37766 | 4.3 |
| <i>UNC119 homolog</i> | U40998 | 4.2 |
| <i>KIAA0151</i> | D63485 | 3.9 |
| <i>Rab1b</i> | XM035660 | 3.8 |
| <i>Annexin VII</i> | J04543 | 3.7 |
| <i>KIAA0110</i> | D14811 | 3.7 |
| <i>Adrenomedullin</i> | D14874 | 3.7 |
| <i>AIM1</i> | U83115 | 3.6 |
| <i>KIAA0250</i> | D87437 | 3.2 |
| <i>P5-1</i> | L06175 | 3.2 |
| Scavenger receptor expressed by endothelial cells | D63483 | 3.2 |
| <i>VHL</i> | L15409 | 3.1 |

(33). Assuming that no other multiply phosphorylated stathmin species had escaped detection, analysis of the integrated intensities of the PO_4 -stathmin and stathmin spots indicates that the percentage of the PO_4 form of total cellular stathmin increased from 11% to 38% with LPS stimulation (Fig. 2B). This is similar to a previous report of an increase from <10% to 35–40% of the Ser²⁶-phosphorylated form in Jurkat cells stimulated with anti-CD3 (34).

Effect of SB203580 on LPS-stimulated Gene Expression—Gene expression analysis of PMNs stimulated with LPS indicated that the majority of genes induced by LPS were unaffected by prior treatment of PMN with SB203580. Of the 100 genes up-regulated by LPS, the up-regulation of 23 was inhibited by greater than 40% (Table VI). The majority of these genes affected by SB203580 were inhibited by less than 60%, whereas only six were inhibited by greater than 80%, all of which represent previously identified interferon-stimulated genes. Induction of cytokine genes by LPS, with the exception of IL-6, was generally unaffected by SB203580.

Effect of SB203580 on LPS-stimulated Protein Expression—Similar to the effect of SB203580 on LPS-stimulated gene expression, little effect of SB203580 was seen on expression levels for the majority of LPS-regulated proteins (Table VII). Two exceptions are annexin III and α -enolase, for which LPS-stimulated expression was attenuated in the presence of the p38 MAPK inhibitor.

Comparison of Microarray and Proteomics Results—Of the LPS-regulated proteins identified by peptide mass fingerprinting for which probes were present on the oligonucleotide microarray, poor concordance was found at the mRNA level (Table VIII). For 13 LPS-up-regulated proteins, 2 corresponding

mRNA transcripts were up-regulated, 1 was down-regulated, 5 were unchanged, and 5 were not detected by the Affymetrix chip. For 5 down-regulated proteins, 3 corresponding transcripts were down-regulated, 1 was unchanged, and 1 was not detected. Varying patterns of LPS regulation emerge for those candidates detected at both the transcript and protein level. Proteasome β chain was up-regulated at both the transcript and protein levels (Table VIII), with no notable effect of SB203580 on expression at either level. Similarly, CAP1, RhoGAP1, and ficolin 1 were down-regulated at both the mRNA transcript and protein level (Table VIII), with no notable effect of SB203580. Annexin III was down-regulated at the transcript level and up-regulated at the protein level, with an inhibitory effect of SB203580 seen only at the protein level (Tables VII and VIII).

DISCUSSION

Interaction of bacterial LPS with the human PMN represents a model system for studying the activation and output of the innate immune system during infection and inflammation. A recent publication (35) describes the gene expression changes of a cultured monocytic cell line after infection by the Gram-positive bacterium *Listeria monocytogenes*. The cell wall components of Gram-positive bacteria, like Gram-negative-derived LPS (*i.e.* from *E. coli*), are known to signal through TLRs (36, 37). Importantly, many of the expression changes found in LPS-stimulated PMNs in the present study were also described in the bacteria-exposed monocytic cells, indicating that many of the gene expression changes seen in bacterial infection are likely mediated by TLRs (38, 39) and that the LPS model system accurately reflects exposure of immune cells to infec-

TABLE II
Human neutrophil genes repressed (>4-fold) after 4 h of LPS exposure

| Description | GenBank™ no. | Change |
|---|---------------|--------|
| | | -fold |
| Kinases | | |
| <i>CAMK, II, gamma</i> | U50360 | -4 |
| <i>Diacylglycerol kinase, delta</i> | D63479 | -4.2 |
| <i>PRKCL2/PRK2</i> protein kinase C-like 2 | U33052 | -4.3 |
| <i>MAPKAPK3</i> | U09578 | -6.3 |
| Protein kinase Ht31, cAMP-dependent | HG2167-HT2237 | -8 |
| <i>CAMK II</i> | L07044 | -9.8 |
| Transporters | | |
| <i>SLC25A5</i> /solute carrier family 25, member 5 | J02683 | -4.2 |
| <i>SLC19A1</i> ; folate transporter | U17566 | -4.4 |
| <i>SLC2A3</i> ; facilitated glucose transporter | M20681 | -5 |
| Metabolic | | |
| <i>Carbonic anhydrase IV</i> | L10955 | -4.4 |
| <i>RNase A family, k6</i> | U64998 | -4.5 |
| <i>Glycogen phosphorylase</i> ; liver | M14636 | -4.6 |
| <i>Inositol polyphosphate-5-phosphatase</i> | U57650 | -4.6 |
| <i>Inositol 1,3,4-trisphosphate 5/6-kinase</i> | U51336 | -4.7 |
| <i>Transketolase</i> | L12711 | -4.8 |
| <i>Protein phosphatase 4</i> , reg. subunit 1 (clone 23840) | U79267 | -4.9 |
| <i>Cytidine deaminase</i> | L27943 | -5.4 |
| <i>MGAT1</i> | M55621 | -5.4 |
| <i>HMOX1</i> | X06985 | -5.4 |
| <i>MAN2A2</i> | L28821 | -5.8 |
| <i>Glycogenin</i> (also represents U31525) | HG4334-HT4604 | -5.9 |
| Structural | | |
| <i>Fibrinogen-like protein (pT49 protein)</i> | Z36531 | -4.2 |
| <i>H2AFZ</i> | M37583 | -4.7 |
| <i>Paxillin</i> | U14588 | -4.9 |
| <i>Lamin B R</i> | L25931 | -5.9 |
| <i>Dynamin 2</i> | L36983 | -6.2 |
| <i>Actinin 1</i> | M95178 | -6.7 |
| α -Tubulin | X01703 | -10 |
| <i>Tubulin, α1</i> , isoform 44 | HG2259-HT2348 | -15 |
| Transcriptional regulators | | |
| <i>Lymphoblastic leukemia-derived sequence 1</i> | M22638 | -4.4 |
| <i>MAX-interacting protein 1</i> | L07648 | -4.5 |
| <i>Nuclear factor erythroid 2</i> isoform f | S77763 | -6 |
| <i>Transducer of ERBB2, 1</i> | D36305 | -6.9 |
| <i>NFATC4</i> | L41067 | -7.8 |
| <i>ATF-2 (CRE-Bpa)</i> | L05515 | -9.6 |
| Receptors | | |
| <i>Lymphotoxin β receptor</i> | L04270 | -4.4 |
| <i>Folate receptor 3 (gamma)</i> | U08471 | -5 |
| | U11875 | -5.3 |
| Signaling | | |
| <i>Pix-α; cool-2 (KIAA0006)</i> | D25304 | -4.5 |
| <i>ARHB/RhoB</i> | M12174 | -4.5 |
| <i>TNFSF10; TRAIL</i> | U37518 | -6.6 |
| Ca²⁺ binding | | |
| <i>ANXII</i> | L19605 | -4.3 |
| <i>S100A4</i> | M80563 | -4.8 |
| <i>ANXI</i> | X05908 | -4.8 |
| Other | | |
| <i>Proteolipid protein 2</i> | L09604 | -4.9 |
| <i>Protein phosphatase 1, α catalytic subunit</i> | HG1614-HT1614 | -5 |
| <i>TIMP2</i> | M32304 | -5.1 |
| <i>KIAA0199</i> | D83782 | -5.2 |
| <i>Lipin 2 (KIAA0249)</i> | D87436 | -5.6 |
| <i>LRMP (Jaw1)</i> | U10485 | -5.8 |
| <i>CUGBP2</i> | U69546 | -6.9 |
| Clone 23933 | U79273 | -7 |
| <i>PECAM1</i> | L34657 | -8 |
| <i>Delta sleep-inducing peptide</i> | Z50781 | -8.7 |
| <i>DiGeorge synd. critical region gene 2 (KIAA0163)</i> | D79985 | -9 |
| <i>SELPLG; CD162; selectin P ligand</i> | U25956 | -32 |

tion. Nevertheless, the reliance upon DNA microarrays alone affords insight only into the transcriptional response without corroboration at the protein level. In the present study, appli-

cation of both DNA microarray and proteomics technology to our model system provides unique insight into both the cellular biology of the activated PMN and the responsiveness and reg-

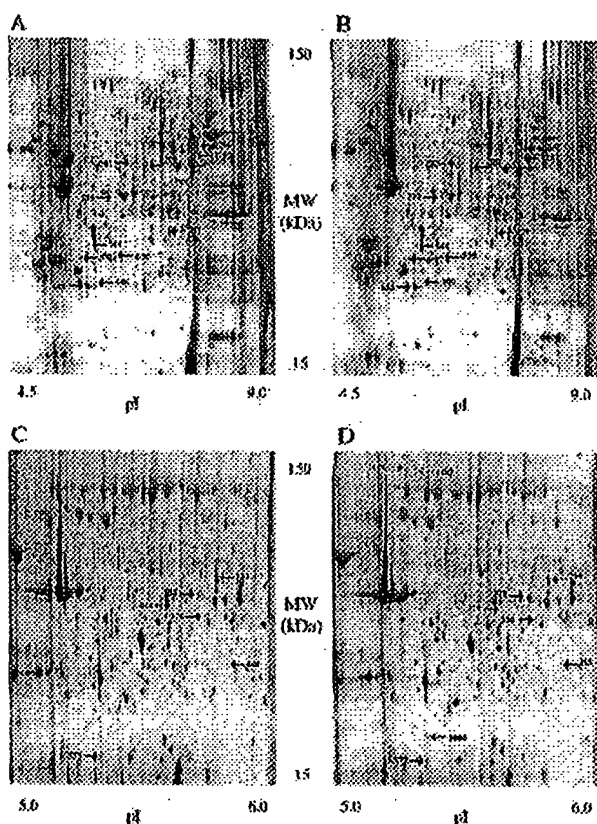


FIG. 1. Two-dimensional PAGE of LPS-exposed human PMNs. A and B, colloidal Coomassie Blue-stained pH 3.0–10.0, two-dimensional PAGE gels (A, control; B, LPS-exposed) with up-regulated (solid arrows) and down-regulated (hatched arrows) proteins indicated. These results are representative of six separate experiments. C and D, colloidal Coomassie Blue-stained pH 5.0–6.0, two-dimensional PAGE gels (C, control; D, LPS-exposed) with up-regulated (solid arrows), new (solid arrow, open arrowhead), and down-regulated (hatched arrows) proteins indicated. LPS-exposed PMNs from three blood donors were pooled.

ulation of its transcriptional and translational machinery. As will be discussed below, our study identifies, in particular, novel aspects of the LPS-stimulated PMN transcriptional regulation, activity in the innate immune response, signaling, cytoskeletal reorganization, and priming for granule release.

In the present study, the increase in NF- κ B transcript abundance (Table I) detected by the microarrays corroborates the findings of other studies of PMNs and monocytes (40) and indicates a mechanism for the responsiveness and scope of the PMN transcriptional machinery following LPS exposure. NF- κ B, recently described to be activated by LPS through the TLR/MyD88/interleukin-1 receptor-associated kinase pathway (1, 4), is the only transcriptional complex reported to be induced by LPS in the PMN. However, because the transcriptional NF- κ B complex has been implicated in the regulation of only a portion of the genes induced by LPS in this study (data not shown), the importance of alternative transcriptional regulators in the PMN is clear. Of interest, several other known and putative transcriptional regulators with less well defined functions were also up-regulated in the present study, including *PLAGL2*, a putative zinc-finger protein, *XPB-1*, *MTF-1*, *Ets-2*, *B-ATF*, and *DIF-2*. On the other hand, LPS-down-regulated genes include *ATF-2* (a known target of p38), *NEATC4*, *TOB-1*, *NF-E2*, *MXI-1*, and *LYL-1*. Although the exact role of these gene products in regulating cell function is unknown,

these data indicate that the range of transcriptional responses in the LPS-stimulated PMN is much broader than previously suggested and that the signaling capabilities of the PMN in the immune response are thereby likely extended in scope and specificity.

As expected from the literature, the genes for several cytokines and chemokines, including *IL-1 β* , *IL-6*, and *MIP-1 β* , were found to be up-regulated (Table I). On the other hand, the notable absence of up-regulated cytokines in the proteomics experiments reflects their removal in the post-LPS incubation wash performed prior to lysis for two-dimensional-PAGE. Up-regulation of these inflammatory mediators is well documented in PMNs exposed to LPS and in animal models of LPS-induced sepsis syndrome and acute respiratory distress syndrome, a PMN-mediated illness (41, 42). Several genes in this family were up-regulated that have not, to our knowledge, been described in LPS-stimulated cells, including *MCP-1*, *GRO3*, *IL-10RA*, and *HM74*, an orphan G protein-coupled receptor with homology to chemokine receptors. The down-regulation of *TNFSF10*, *lymphotoxin b receptor*, and *TNFAIP1* were also observed. The modulation of genes involved in cytokine signaling, including the adapter molecules *TRAF1* (LPS and TNF receptor signaling) and *TNFAIP1* (TNF receptor signaling) and several kinases and phosphatases, may indicate a change in cytokine responsiveness after LPS treatment. Relevant in this regard from the proteomics data are: 1) the up-regulation of protein phosphatase 1, which has been shown to regulate PMN NADPH oxidase activation and translocation (43, 44) and to regulate LPS-induced NF- κ B activation (45); 2) the down-regulation of Rho-GAP1, which has been shown to regulate NADPH oxidase activity in the PMN (46); and 3) the up-regulation of *PO₄-stathmin* (Table IV), a phosphoprotein postulated to function as a relay and integrator of multiple signal transduction pathways (34). Several noncytokine, nonchemokine genes involved in the immune response were also up-regulated, including the complement pathway members *C3*, *C3AR1*, and *PFC*; the protease inhibitors *ELANH2* (elastase inhibitor), *SLPI*, *PI-3*, and *PI-9*; and the acute phase protein *orosomucoid*. LPS regulation of *C3AR1* and *orosomucoid* expression have not previously been reported. In the proteomics experiments, the down-regulation of *ficolin-1* (Table III), a collectin-like cell surface protein reported to activate the complement system and to mediate adhesion and phagocytosis in monocytes but not previously reported in granulocytes (47), may represent negative modulation of the innate immune response. The finding that genes other than cytokines and chemokines are regulated by the PMN in response to LPS indicates that the PMN plays a more sophisticated role in host-defense and immunity than previously thought.

Treatment of the PMN with LPS lead to the induction of a set of genes associated with the anti-viral Type I interferons, IFN α/β . This induction occurs independently of the release of IFN or another unidentified soluble factor.² Furthermore, the set of genes expressed is smaller than that induced by IFN α/β , as described by Der *et al.* (12). This may be due to differences in the scope of the signaling systems activated by LPS and IFN α/β , or the time course of analysis of genes in the LPS-stimulated PMN. The implication that LPS treatment of PMN allows PMN to express anti-viral activity is currently being tested. Of interest was the finding that induction of interferon-stimulated genes was blocked by pretreatment of PMNs with SB203580. Work from our laboratory has indicated that signal transducers and activators of transcription activation does not occur in response to LPS in PMNs.² In addition, interferon-

² K. C. Malcolm and G. S. Worthen, manuscript in preparation.

TABLE III
Analysis of pH 3.0–10.0 two-dimensional PAGE gels

Mean change(fold) in expression level among six PMN donors is reported. The change in expression for the proteins listed was statistically significant ($p < 0.05$) as measured by a two-tailed Student's *t* test.

| Identification [spot no.] | Swiss-Prot no. | Estimated M_r /pI | Theoretical M_r /pI | Peptides matched/ submitted | Protein covered | Mean change |
|--|----------------|---------------------|-----------------------|--------------------------------|-----------------|-------------|
| | | | | % | % | fold |
| <i>Up-regulated</i> | | | | | | |
| Proteasome β chain [646] | P28070 | 27/5.7 | 29.2/5.72 | 9/12 (75%) | 36% | 1.51 |
| Annexin III [550] | P12429 | 31/5.7 | 36.4/5.6 | 14/18 (78%) | 42% | 1.37 |
| Actin fragment [544] ^a | P02570 | 32/5.5 | (41.7/5.29) | 13/15 (87%) | (34%) | 1.74 |
| Actin fragment [591] ^a | P02570 | 30/5.4 | (41.7/5.29) | 14/18 (78%) | (29%) | 1.60 |
| α -Enolase [380] | P06733 | 41/5.7 | 47.2/7.01 | 9/10 (90%) | 24% | 1.65 |
| Rab-GDP dissociation inhibitor β [289] | P50395 | 50/6.1 | 50.7/6.11 | 10/11 (91%) | 25% | 1.24 |
| Glutathione S-transferase P [648] | P09211 | 23/5.5 | 23.4/5.43 | 6/8 (75%) | 41% | 1.54 |
| Pre-B-cell colony enhancing factor [1152] | P43490 | 53/7.0 | 55.5/6.69 | 12/16 (75%) | 25% | 1.29 |
| <i>Down-regulated</i> | | | | | | |
| Adenylyl cyclase-associated protein 1 [256] | Q01518 | 55/7.3 | 51.7/8.07 | 16/22 (73%) | 34% | 0.53 |
| Rho-GAP1 [283] | Q07960 | 50/5.8 | 50.4/5.85 | 7/9 (78%) | 22% | 0.67 |
| Ficolin 1 [511] | O00602 | 33/6.5 | 35/6.39 | 10/12 (83%) | 25% | 0.74 |

^a The theoretical pI and M_r of native actin are indicated. Protein coverage indicates coverage of native actin.

TABLE IV
Analysis of pH 5.0–6.0 two-dimensional PAGE gels

Results are from pooled samples for control ($n=3$) and LPS-exposed ($n=3$) PMNs from human donors. Expression of the reported proteins was altered >1.5-fold following LPS exposure in two repeat experiments. "New" designates proteins seen in the LPS gel in two repeat experiments but not detectable in the corresponding control gels.

| Identification [spot no.] | Swiss-Prot no. | Estimated M_r /pI | Theoretical M_r /pI | Peptides matched/ submitted | Protein covered | Change |
|--|-----------------------|---------------------|-----------------------|--------------------------------|-----------------|------------------|
| | | | | % | % | fold |
| <i>Up-regulated</i> | | | | | | |
| Protein-tyrosine kinase 9-like [468] | Q9Y3F5 ^a | 34/5.81 | 39.5/6.37 | 10/14 (71%) | 34% | 1.8 |
| Protein phosphatase 1, catalytic subunit, β isoform [378] | P37140 | 38/5.73 | 37.2/5.84 | 7/10 (70%) | 22% | 2.0 |
| PO ₄ -stathmin [577] | P16949 ^b | 18/5.36 | 17.3/5.76 | 9/12 (75%) | 42% | 2.1 ^c |
| Nonmuscle myosin heavy chain [1102] | 189036 ^d | 145/5.32 | 145/5.23 | 20/21 (95%) | 17% | New |
| Putative PO ₄ -nonmuscle myosin heavy chain [1101] ^d | 189036 ^{b,c} | 145/5.29 | 145/5.23 | 14/16 (87%) | 13% | New |
| Leukocyte elastase inhibitor [318] | P30740 | 42/5.71 | 42.7/5.9 | 9/13 (69%) | 22% | 2.4 |
| Grancalcin [1004] | P28676 | 24/5.36 | 24.0/5.02 | 7/10 (70%) | 31% | New |
| <i>Down-regulated</i> | | | | | | |
| Adenosylhomocysteinase [324] | P23526 | 48/5.82 | 47.7/6.04 | 7/9 (78%) | 14% | 0.4 |
| PEST phosphatase interacting protein homolog [234] ^f | 4100162 ^f | 48/5.30 | 47.6/5.35 | 11/13 (85%) | 30% | 0.5 |

^a TrEMBL accession number.

^b Accession number and theoretical pI and M_r for the unmodified protein are indicated.

^c NCBI accession number.

^d See text for explanation.

^e Among three experiments, the ratio of PO₄-stathmin expression increase, following LPS exposure in the presence of SB203580 divided by that in the absence of SB203580, was 0.93.

^f Genpept accession number.

^g This search was performed using average masses measured by linear mode MALDI-TOF MS.

TABLE V
Analysis of pH 5.5–6.7 two-dimensional PAGE gels

Results are from pooled samples for control ($n=3$) and LPS-exposed ($n=3$) PMNs from human donors. Expression of the reported proteins was altered >1.5-fold following LPS exposure in two repeat experiments.

| Identification [spot no.] | Swiss-Prot no. | Estimated M_r /pI | Theoretical M_r /pI | Peptides matched/ submitted | Protein covered | Change |
|--------------------------------|----------------|---------------------|-----------------------|--------------------------------|-----------------|--------|
| | | | | % | % | fold |
| <i>Up-regulated</i> | | | | | | |
| Transaldolase [475] | P37837 | 38/5.95 | 37.5/6.36 | 13/17 (76%) | 33% | 2.5 |
| Isocitrate dehydrogenase [431] | O75874 | 46/6.25 | 46.7/6.35 | 7/7 (100%) | 13% | 2.3 |
| Moessin [201] | P26038 | 61/6.09 | 67.8/6.07 | 11/13 (85%) | 17% | 2.1 |
| α -Enolase [459] | P06733 | 43/5.64 | 47.2/7.01 | 7/10 (70%) | 17% | 3.8 |
| <i>Down-regulated</i> | | | | | | |
| Calponin H2 [240] | Q99439 | 34/6.65 | 33.7/6.94 | 10/11 (90%) | 27% | 0.5 |

regulatory factor 3, a known regulator of interferon-stimulated gene transcription, is not a direct target of p38 kinase.² Therefore, gene expression analysis of LPS-stimulated PMNs has uncovered a previously uncharacterized signal transduction system that is sensitive to inhibition of p38 MAPK.

Knowledge of the genes down-regulated by LPS permits the

development of further hypotheses addressing PMN function in the face of infection. Strikingly, several down-regulated genes and gene products are structural in nature (e.g. paxillin, actinin, calponin H2) (Tables II and V). A known consequence to the PMN of LPS exposure is decreased motility (48). Up-regulation of genes for adhesion molecules (ICAM-1, CD44, AL-

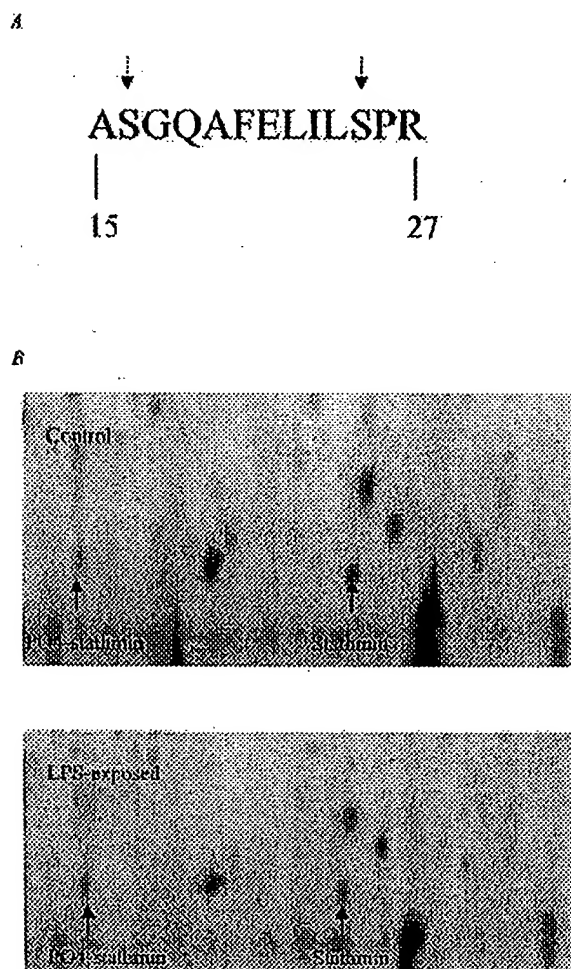


FIG. 2. A, the predicted sequence of the tryptic phosphopeptide in PO_4 -stathmin (1468.72 Da). The peptide mass measured by MALDI-TOF MS and the predicted mass differed by 14 ppm. As indicated, two alternate phosphorylation sites are possible: serine 16 and serine 25. B, PO_4 -stathmin and stathmin were identified on the control and LPS-exposed pH 5.0–6.0 gels. Consistent with phosphorylation, the PO_4 -stathmin spot was distinguished by a peptide of mass 1468.72 Da (i.e. 80 Da greater than the peptide of 1388.72 Da seen in the stathmin spot). Assuming that no other multiply phosphorylated stathmin species have escaped detection, analysis of the integrated intensities of the PO_4 -stathmin and stathmin spots indicates that the percentage of the PO_4 form of total cellular stathmin has increased from 11% to 38% with LPS stimulation. The decrease in integrated intensity for stathmin was equal in amount to the increase in PO_4 -stathmin following LPS exposure.

CAM, and TSG-6), and down-regulation of genes for structural proteins, indicates a genetic basis for this observation. Down-regulation of two genes implicated in cytoskeletal regulation, *Pix- α* and *RhoB*, was also observed. The calcium-binding protein S100A4, down-regulated in LPS-treated PMNs (Table II), has been implicated in cell motility and metastasis (49). Decreased motility may be beneficial in sustaining the inflammatory response at sites of infection. In addition, LPS treatment results in an inhibition of apoptosis (50). Therefore, the longer residence time of the PMN at sites of infection is consistent with the long term genetically coded changes seen in these gene-profiling experiments and indicates that the changes in gene expression are functionally relevant to host defense and immunity.

By providing information on post-translational modification, the proteomics data may provide further insights into the cy-

TABLE VI
Effect of SB203580 on LPS-stimulated gene expression
Genes are reported for which the SB203580/control expression ratio is ≤ 0.60 .

| Gene name | -fold change ratio (SB203580/control) | Change in absence of SB203580 |
|------------------------|--|----------------------------------|
| | | <i>-fold</i> |
| <i>ISG15</i> | 0.09 | 22.5 |
| <i>HCR</i> | 0.38 | 20.8 |
| <i>Mx-1</i> | 0 | 19.4 |
| <i>IFI56</i> | 0 | 12.1 |
| <i>PI-9</i> | 0.57 | 9.5 |
| <i>Ets-2</i> | 0.59 | 7.4 |
| <i>IL-6</i> | 0.45 | 6.3 |
| <i>Rel</i> | 0.50 | 6.2 |
| <i>LIMS1</i> | 0.58 | 6.1 |
| <i>C3AR1</i> | 0.49 | 6.1 |
| <i>INDO</i> | 0.35 | 5.2 |
| <i>KIAA0105</i> | 0.41 | 5.1 |
| <i>SNAP23</i> | 0.58 | 5.0 |
| <i>SLPI</i> | 0.58 | 4.7 |
| <i>ELNAH2</i> | 0.49 | 4.6 |
| <i>HM-74</i> | 0.57 | 3.8 |
| <i>PKR</i> | 0 | 3.7 |
| <i>MAD</i> | 0.21 | 3.6 |
| <i>IFIT4</i> | 0.12 | 3.6 |
| <i>Glycerol kinase</i> | 0 | 3.6 |
| <i>IFI54</i> | 0 | 3.5 |
| <i>IFI58</i> | 0.39 | 3.5 |
| <i>IPF35</i> | 0.46 | 3.0 |

TABLE VII
Effect of SB203580 on LPS-stimulated protein expression

| Protein name | -fold change ratio (SB203580/control) | Change in absence of SB203580 |
|--|--|----------------------------------|
| | | <i>-fold</i> |
| <i>Up-regulated</i> | | |
| Proteasome β chain | 0.8 | 1.51 |
| Annexin III | 0.6 | 1.37 |
| Actin fragment [544] | 0.8 | 1.74 |
| Actin fragment [591] | 0.8 | 1.60 |
| α -Enolase | 0.6 | 1.65 |
| Rab-GDP dissociation inhibitor β | 1.1 | 1.24 |
| Glutathione S-transferase P | 1.2 | 1.54 |
| Pre-B-cell colony enhancing factor | 1.2 | 1.29 |
| <i>Down-regulated</i> | | |
| Adenyl cyclase-associated protein 1 | 1.3 | 0.53 |
| Rho-GAP1 | 0.8 | 0.67 |
| Ficolin 1 | 1.0 | 0.74 |

toskeletal remodeling effects of LPS upon the PMN. We contend that the actin fragments identified (Table III) are unlikely to represent technical artifacts. Rather, their specificity (identical molecular weight/pI among different experiments), statistically significant up-regulation by LPS, as well as the use of a lysis buffer containing chaotropes and multiple protease inhibitors argue instead that these fragments are physiologic consequences of LPS exposure in the human PMN. More specifically, the up-regulation of these fragments following LPS exposure (Table III) suggests that LPS may activate an actin-cleaving enzyme, which, in turn, remodels the cytoskeleton. Intriguing in this vein, calpain has recently been reported to play an important role in cell migration and cytoskeletal organization of fibroblasts (51). The possibilities that LPS may induce calpain activation and that calpain activation may regulate cytoskeletal reorganization and motility are currently under investigation. An alternative possibility is that actin cleavage is a marker of neutrophil apoptosis (52).

Other LPS-regulated proteins may play important roles in cytoskeletal reorganization. The up-regulation of protein-tyrosine kinase 9-like (A6-related protein) may modulate LPS-

TABLE VIII
LPS-regulated proteins for which a probe was present on the Affymetrix chip
A comparison of corresponding protein and mRNA transcript changes following LPS exposure is shown.

| Protein | Protein change | mRNA change |
|--|----------------|-------------------|
| | | -fold |
| Up-regulated | | |
| Proteasome β chain | 1.5 | 1.9 \uparrow |
| Leukocyte elastase inhibitor | 2.4 | 4.6 \uparrow |
| Rab-GDI β | 1.24 | NC ^a |
| Grancalcin | New | NC |
| Transaldolase | 2.5 | NC |
| Moesin | 2.1 | NC |
| Nonmuscle myosin heavy chain | New | NC |
| Glutathione S-transferase P | 1.54 | Absent |
| Pre-B cell enhancing factor | 1.29 | Absent |
| Isocitrate dehydrogenase | 2.3 | Absent |
| PO ₄ -stathmin | 2.1 | Absent (stathmin) |
| Protein phosphatase 1, β catalytic subunit | 2 | Absent |
| Annexin III | 3.1 | 3.1 \downarrow |
| Down-regulated | | |
| Adenylyl cyclase-associated protein 1 | 1.9 | 2.1 \downarrow |
| Rho-GAP 1 | 1.5 | 2.7 \downarrow |
| Ficolin 1 | 1.4 | 1.7 \downarrow |
| Adenosylhomocysteinase | 2.5 | Absent |
| Calponin H2 | 2 | NC |

^a NC, no measureable change.

induced actin polymerization, because it bears a high degree of homology to twinfilin (A6), an actin monomer-binding protein that localizes to sites of rapid filament assembly in cells and is believed to regulate actin filament turnover (53). In turn, LPS-induced down-regulation of Rho-GTPase activating protein 1 (Table III) may regulate twinfilin (and protein-tyrosine kinase 9-like) activity, because twinfilin has been shown to colocalize with Rac1 and Cdc42 and to be regulated by active Rac1 in NIH 3T3 cells (53). Activation of Rho proteins may be facilitated by LPS up-regulation of moesin (Table V), because moesin reportedly induces the dissociation of Rho from GDI (54). Rac1 may, in turn, promote activation of the actin filament-nucleating Arp2/3 complex through interactions with WASP (Wiskott-Aldrich syndrome protein) family proteins (55) and, interestingly, is postulated to regulate the dynamics of both the actin and microtubule cytoskeletons via phosphorylation of stathmin (Table IV) (56). Calponin H2 is an actin-binding protein not previously reported in PMNs that is postulated to play a role in cytoskeletal organization (57). Its down-regulation by LPS (Table V) likely modulates LPS-induced cytoskeletal reorganization. The up-regulation of nonmuscle myosin heavy chain and a putative phosphorylated form of myosin heavy chain (putative protein kinase C substrate by prediction rules) in the LPS-exposed PMN (Table IV) is of uncertain significance; myosin has been implicated in multiple functions in the PMN, including locomotion, fluid pinocytosis, and phagocytosis (58). Of interest, however, S100A4 (down-regulated, Table II) has been reported to regulate cytoskeletal dynamics by inhibiting protein kinase C-mediated phosphorylation of nonmuscle myosin heavy chain (59).

LPS induction of stathmin phosphorylation (Table IV and Fig. 2) may represent another mechanism by which the cytoskeleton is remodeled. Stathmin is a phosphoprotein reportedly involved in both signal transduction and in regulation of the microtubulin filament network; furthermore, phosphorylation of stathmin has been reported to modulate its tubulin-binding avidity (60). Inferences can be made about both the phosphorylation site on PO₄-stathmin and the responsible kinase induced by LPS. Four phosphorylation sites in stathmin have been well described: Ser¹⁶, Ser²⁵, Ser³⁸, and Ser⁶³ (32, 33).

Ser¹⁶ has been reported as a substrate for Ca²⁺/calmodulin (CaM)-dependent kinases (32), and Ser²⁵ as primarily a substrate for p38 and ERK (33), with p34^{cdc2} also active but bearing a 5-fold preference for Ser³⁸ (34). As stated above, the phosphopeptide identified in PO₄-stathmin, extending from residues 15 to 27 (1468.7 Da), is consistent with phosphorylation of either Ser¹⁶ or Ser²⁵ (Fig. 2). Although both p38 δ and p38 α MAPK isoforms are expressed in the human PMN, LPS has been shown to selectively activate the p38 α isoform in human PMNs (9). The p38 α isoform, however, has been shown to be relatively inactive at Ser²⁵; in fact, p38 δ is ~100-fold more active at Ser²⁵, and selective p38 α inhibitors do not inhibit the stress-activated phosphorylation of stathmin in 293 cells (33). Further support for the lack of involvement of p38 signaling in phosphorylation of stathmin in our system is the apparent lack of effect of SB203580 (a selective p38 α and p38 β inhibitor) on LPS-induced expression of PO₄-stathmin (Table IV). Because p34^{cdc2} is relatively inactive at Ser²⁵ (34), we conclude that the phosphorylation site is likely to be Ser¹⁶, a reported substrate of CaM-dependent kinase. Although CaM kinases have previously been implicated in gene activation in LPS-exposed myelomonocytic HD11 cells (61), stathmin signaling has not, to our knowledge, been previously reported in either PMNs or lipopolysaccharide signal transduction.

Cytoskeletal reorganization, a well-described regulator of granule release (62), may underlie LPS-induced priming for PMN granule release, but several LPS-regulated proteins may provide more specific clues. LPS exposure led to increased levels of grancalcin, a calcium-binding protein previously detected in PMNs and shown to translocate to granules and plasma membrane in the presence of physiologic concentrations of calcium (63). Similarly, annexin III, a calcium-binding protein highly expressed in PMN granule membranes and implicated in calcium-mediated secretion (64) and in granule fusion (65), was also found to be up-regulated. Exocytosis of granule contents may also be facilitated by LPS up-regulation of Rab-GDP dissociation inhibitor (Table III), which has been proposed to recycle Rab after vesicle fusion by extracting it from the membrane and loading it onto newly formed transport intermediates (66).

Parallel use of DNA microarrays and proteomics affords a powerful strategy for comparison of corresponding mRNA transcripts and proteins, thereby affording new insight into the mechanisms by which the cell regulates its signaling responses to the external environment. Of interest, a poor correlation was found between corresponding transcripts and proteins (Table VIII), as reported in other systems (17, 18). The finding in some cases of unchanged transcript abundance in the face of regulated protein levels indicates post-transcriptional modulation following LPS exposure. The finding of undetected transcripts in the face of regulated levels of the corresponding proteins may indicate previous transcription of these genes in an earlier state of the myeloid maturation of the PMN, producing stable protein species that have undergone post-translational alteration following LPS exposure. The use of SB203580, a p38 inhibitor, adds further insights into the mechanisms of LPS regulation. At the level of mRNA expression, SB203580 inhibited 23% of LPS-stimulated genes by $\geq 40\%$ and 11% of genes by $\geq 60\%$; therefore, p38 plays a specific role in gene regulation in the PMN. In particular, proteasome β chain was up-regulated at both the mRNA transcript and protein level (Table VIII), with no notable effect of SB203580 on expression at either level, consistent with a non-p38-mediated pathway of primary transcriptional up-regulation induced by LPS. Similarly, CAP1, Rho-GAP1, and ficolin 1 were down-regulated at both the mRNA transcript and protein level (Table VIII), with

no notable effect of SB203580, consistent with a non-p38-mediated pathway of primary transcriptional down-regulation. Interestingly, annexin III was down-regulated at the transcript level and up-regulated at the protein level, with an inhibitory effect of SB203580 seen only at the protein level (Table VII), consistent with a p38-mediated post-transcriptional up-regulation induced by LPS.

Limitations of the present study should be noted. Gene expression analysis by cDNA microarrays does not distinguish between transcriptional regulation and mRNA stabilization; similarly, two-dimensional PAGE proteomics by itself does not distinguish among transcriptional, translational, or post-translational regulation of protein abundance. Transcript detection by microarray technology is limited to the probes included; protein identification by two-dimensional PAGE proteomics is limited to well-resolved regions of the gel, may perform less well with hydrophobic and high molecular weight proteins, and tends to select for more abundant protein species (30). Harvesting of the LPS-incubated PMNs at 4 h may have prevented detection of earlier, transient changes and may have thereby introduced artifactual transcript-protein discordance. Furthermore, the post-LPS incubation, pre-two-dimensional PAGE cell washes would be expected to remove secreted proteins from further analysis, with uncertain effects on detected protein abundance depending on such factors as the degree of *de novo* synthesis and extent of degranulation/exocytosis. Because protein binding of Coomassie Blue has a limited dynamic range and is typically not linear throughout the range of detection, image analysis of Coomassie Blue-stained protein spots should be considered semi-quantitative. For some protein spots, the apparent magnitude of regulation by LPS may have been blunted by the spot approaching staining saturation in the control gel. By limiting our analysis to those protein spots common to all twelve pH 3.0–10.0 two-dimensional gels, we likely excluded some LPS-regulated proteins that happened to be either poorly resolved on a subset of the gels or unmatched by the image analysis software. By further limiting the analysis to those matched spots on the pH 3.0–10.0 gels for which a two-tailed *t* test demonstrated $p < 0.05$, the list of regulated proteins was likely also limited by statistical power. In addition to those regulated proteins listed in Table III, three others were up-regulated and three down-regulated with $p < 0.09$ (data not shown).

Limiting our reported results to those changes that met statistical significance among the donors carries further important implications. We have encountered a two order of magnitude range of response in unselected donor LPS-induced PMN functions, such as TNF- α and superoxide anion release (data not shown). The sources of this physiologic heterogeneity remain uncertain but may possibly include such factors as natural mutations of the LPS receptor component, TLR4 (67). By selecting for LPS effects common to all donors, we may not have characterized the range of genomic and proteomic heterogeneity present in the population and thereby may have focused on only a narrow portion of a broader biological response to LPS. We contend that this reductionist approach is valid because it would be expected to enrich for biologically integral responses of the PMN to LPS. Nevertheless, correlation of genomic and proteomic profiles with functional phenotypes of the PMN may bear important diagnostic and therapeutic implications and will be pursued in future studies.

Widespread regulation of numerous noncytokine/chemokine genes and proteins in the LPS-stimulated human PMN is a novel finding. These data indicate that, despite a narrow scope of gene expression in the nonstimulated state, the terminally differentiated, short-lived PMN likely plays a role in the innate

immune response that is far more sophisticated and dynamic than the simple release of preformed inflammatory mediators. Although gene expression appears to be an important mechanism by which PMNs respond acutely to infection, mRNA transcript/protein concordance is limited, and post-transcriptional (and post-translational) modifications also play an important role. The alteration of multiple transcriptional regulators, G-protein regulators, PO_4 -stathmin, and protein phosphatase 1 indicates that one of the responses to LPS exposure is to modify subsequent signaling events by bacterial components or by other cytokines and chemokines. Finally, the finding that p38 MAPK mediates LPS regulation of a limited subset of transcripts and proteins underlines the continuing need to define signal transduction cascades in the neutrophil.

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REFERENCES

- Bowie, A., and O'Neill, L. A. (2000) *J. Leukocyte Biol.* 67, 508–514
- Chow, J. C., Young, D. W., Golenbock, D. T., Christ, W. J., and Gusovsky, F. (1999) *J. Biol. Chem.* 274, 10689–10692
- Kirschning, C. J., Wesche, H., Merrill Ayres, T., and Rothe, M. (1998) *J. Exp. Med.* 188, 2091–2097
- Muzio, M., Polentarutti, N., Bosio, D., Pahlavan, M. K., and Mantovani, A. (2000) *J. Leukocyte Biol.* 67, 450–456
- Yang, R. B., Mark, M. R., Gray, A., Huang, A., Xia, M. H., Zhang, M., Goddard, A., Wood, W. L., Gurney, A. L., and Godowski, P. J. (1998) *Nature* 385, 284–288
- Parsons, P. E., Worthen, G. S., Moore, E. E., Tate, R. M., and Henson, P. M. (1989) *Am. Rev. Respir. Dis.* 140, 294–301
- Nick, J. A., Avdi, N. J., Young, S. K., McDonald, P. P., Billstrom, M. A., Henson, P. M., Johnson, G. L., and Worthen, G. S. (1999) *Chest* 116, 54S–55S
- Nick, J. A., Young, S. K., Brown, K. K., Avdi, N. J., Arndt, P. G., Suratt, B. T., James, M. S., Henson, P. M., and Worthen, G. S. (2000) *J. Immunol.* 164, 2151–2159
- Nick, J. A., Avdi, N. J., Young, S. K., Lehman, L. A., McDonald, P. P., Frasch, S. C., Billstrom, M. A., Henson, P. M., Johnson, G. L., and Worthen, G. S. (1999) *J. Clin. Invest.* 103, 851–858
- Cassatella, M. A. (1995) *Immunol. Today* 16, 21–26
- McDonald, P. P., Bald, A., and Cassatella, M. A. (1997) *Blood* 89, 3421–3433
- Der, S. D., Zhou, A., Williams, B. R., and Silverman, R. H. (1998) *Proc. Natl. Acad. Sci. U. S. A.* 95, 15623–15628
- Karpf, A. R., Peterson, P. W., Rawlins, J. T., Dalley, B. K., Yang, Q., Albertsen, H., and Jones, D. A. (1999) *Proc. Natl. Acad. Sci. U. S. A.* 96, 14007–14012
- Iyer, V. R., Eisen, M. B., Ross, D. T., Schuler, G., Moore, T., Lee, J. C., Trent, J. M., Staudt, L. M., Hudson, J., Jr., Bogni, M. S., Lashkari, D., Shalon, D., Botstein, D., and Brown, P. O. (1999) *Science* 283, 83–87
- Han, J., Jiang, Y., Li, Z., Kravchenko, V. V., and Ulevitch, R. J. (1997) *Nature* 386, 296–299
- Lee, J. C., Leydon, J. T., McDonnell, P. C., Gallagher, T. F., Kumar, S., Green, D., McNulty, D., Blumenthal, M. J., Heys, J. R., Landvatter, S. W. et al. (1994) *Nature* 372, 739–746
- Anderson, L., and Selinger, J. (1997) *Electrophoresis* 18, 533–537
- Gygi, S. P., Rochon, Y., Franz, B. R., and Aebersold, R. (1999) *Mol. Cell. Biol.* 19, 1720–1730
- Lewis, T. S., Hunt, J. B., Aveline, L. D., Jonscher, K. R., Louie, D. F., Yeh, J. M., Nahrini, T. S., Resing, K. A., and Ahn, N. G. (2000) *Mol. Cell* 6, 1343–1354
- Soskic, V., Goriach, M., Poznanovic, S., Boehmer, F. D., and Godovac-Zimmermann, J. (1999) *Biochemistry* 38, 1757–1764
- Triantafyllou, K., Triantafyllou, M., and Dedrick, R. L. (2001) *Nat. Immunol.* 2, 338–345
- Nick, J. A., Avdi, N. J., Young, S. K., Knall, C., Gerwins, P., Johnson, G. L., and Worthen, G. S. (1997) *J. Clin. Invest.* 99, 975–986
- Rabilloud, T., Valette, C., and Lawrence, J. F. (1994) *Electrophoresis* 15, 1552–1558
- Bradford, M. M. (1976) *Anal. Biochem.* 72, 248–254
- Neuhoff, V., Arold, N., Taube, D., and Ehrhardt, W. (1988) *Electrophoresis* 9, 255–262
- Arnot, D., O'Connell, K. L., King, K. L., and Stults, J. T. (1998) *Anal. Biochem.* 258, 1–18
- Hellman, U., Wernstedt, C., Genez, J., and Heldin, C. H. (1995) *Anal. Biochem.* 224, 451–455
- Yan, J. X., Sanchez, J. C., Rouge, V., Williams, K. L., and Hochstrasser, D. F. (1999) *Electrophoresis* 20, 723–726
- Mardian, J. K., and Isenberg, I. (1978) *Anal. Biochem.* 81, 1–12
- Corthals, G. L., Wasinger, V. C., Hochstrasser, D. F., and Sanchez, J. C. (2000) *Electrophoresis* 21, 1104–1115
- Blom, N., Gammeltoft, S., and Brunak, S. (1999) *J. Mol. Biol.* 294, 1351–1362
- le Gouvello, S., Manceau, V., and Sobel, A. (1998) *J. Immunol.* 161, 1113–1122
- Parker, C. G., Hunt, J., Diener, K., McGinley, M., Soriano, B., Keesler, G. A., Bray, J., Yan, Z., Wang, X. S., Kohno, T., and Lichtenstein, H. S. (1998)

- Biochem. Biophys. Res. Commun.* 249, 791-796
34. Marklund, U., Brattsand, G., Shingler, V., and Gullberg, M. (1993) *J. Biol. Chem.* 268, 15039-15047
 35. Cohen, P., Bouaboula, M., Bellis, M., Baron, V., Jbilo, O., Poinot-Chazel, C., Galiegus, S., Hadibi, E. H., and Casellas, P. (2000) *J. Biol. Chem.* 275, 11181-11190
 36. Yoshimura, A., Lien, E., Ingalls, R. R., Tuomanen, E., Dziarski, R., and Golenbock, D. (1999) *J. Immunol.* 163, 1-5
 37. Lien, E., Sellati, T. J., Yoshimura, A., Flo, T. H., Rawadi, G., Finberg, R. W., Carroll, J. D., Espevik, T., Ingalls, R. R., Radolf, J. D., and Golenbock, D. T. (1999) *J. Biol. Chem.* 274, 33419-33425
 38. Underhill, D. M., Ozinsky, A., Hajjar, A. M., Stevens, A., Wilson, C. B., Bassetti, M., and Aderem, A. (1999) *Nature* 401, 811-815
 39. Brightbill, H. D., Libraty, D. H., Krutzik, S. R., Yang, R. B., Belisle, J. T., Bleharski, J. R., Maitland, M., Norgard, M. V., Plevy, S. E., Smale, S. T., Brennan, P. J., Bloom, B. R., Godowski, P. J., and Modlin, R. L. (1999) *Science* 285, 732-736
 40. de Wit, H., Dokter, W. H., Kuopmans, S. B., Lummen, C., van der Leij, M., Smit, J. W., and Vellenga, E. (1998) *Leukemia* 12, 363-370
 41. Johnston, C. J., Finkelstein, J. N., Gelein, R., and Oberdorster, G. (1998) *Toxicol. Sci.* 46, 300-307
 42. Ulich, T. R., Wateon, L. R., Yin, S. M., Guo, K. Z., Wang, P., Thang, H., and del Castillo, J. (1991) *Am. J. Pathol.* 138, 1485-1496
 43. Karlsson, A., Nixon, J. B., and McPhail, L. C. (2000) *J. Leukocyte Biol.* 67, 396-404
 44. Dorseuil, O., Quinn, M. T., and Bokoch, G. M. (1995) *J. Leukocyte Biol.* 58, 108-113
 45. Pahan, K., Sheikh, F. G., Nambodiri, A. M., and Singh, I. (1998) *J. Biol. Chem.* 273, 12219-12226
 46. Geiszt, M., Dagher, M. C., Molnar, G., Havasi, A., Faure, J., Paclet, M. H., Morel, F., and Ligeti, E. (2001) *Biochem. J.* 355, 851-858
 47. Teh, C., Le, Y., Lee, S. H., and Lu, J. (2000) *Immunology* 101, 225-232
 48. Wagner, J. G., and Roth, R. A. (1999) *J. Leukocyte Biol.* 66, 10-24
 49. Barraclough, R. (1998) *Biochim. Biophys. Acta* 1448, 190-199
 50. Lee, A., Whyte, M. K., and Haslett, C. (1993) *J. Leukocyte Biol.* 54, 283-288
 51. Dourdin, N., Bhatt, A. K., Dutt, P., Greer, P. A., Arthur, J. S., Elce, J. S., and Huttenlocher, A. (2001) *J. Biol. Chem.* 276, 48362-48366
 52. Brown, S. B., Bailey, K., and Savill, J. (1997) *Biochem. J.* 323, 233-237
 53. Vartiainen, M., Ojala, P. J., Auvinen, P., Peranen, J., and Lappalainen, P. (2000) *Mol. Cell. Biol.* 20, 1772-1783
 54. Takahashi, K., Sasaki, T., Mammoto, A., Takaishi, K., Kameyama, T., Tsukita, S., and Takai, Y. (1997) *J. Biol. Chem.* 272, 23371-23375
 55. Machesky, L. M., and Inaall, R. H. (1998) *Curr. Biol.* 8, 1347-1356
 56. Daub, H., Gevaert, K., Vandekerckhove, J., Sobel, A., and Hall, A. (2001) *J. Biol. Chem.* 276, 1677-1680
 57. Masuda, H., Tanaka, K., Takagi, M., Ohgami, K., Sakamaki, T., Shibata, N., and Takahashi, K. (1996) *J. Biochem. (Tokyo)* 120, 415-424
 58. Valerius, N. H., Stendahl, O. I., Hartwig, J. H., and Stossel, T. P. (1982) *Adv. Exp. Med. Biol.* 141, 19-28
 59. Krijavetska, M., Tarabyskina, S., Branstetter, L., Maitland, N., Lomonosov, M., Hansen, K., Georgiev, G., and Lukamidin, E. (1998) *J. Biol. Chem.* 273, 9852-9856
 60. Steinmetz, M. O., Jahnke, W., Towbin, H., Garcia-Echeverria, C., Vnshol, H., Muller, D., and van Oostrum, J. (2001) *EMBO Rep.* 2, 505-510
 61. Regenhard, P., Goeths, R., and Phi-van, L. (2001) *J. Leukocyte Biol.* 69, 651-658
 62. Valentijn, K., Valentijn, J. A., and Jamieson, J. D. (1999) *Biochem. Biophys. Res. Commun.* 266, 652-661
 63. Teahan, C. G., Totty, N. F., and Segal, A. W. (1992) *Biochem. J.* 286, 549-554
 64. Rosales, J. L., and Ernst, J. D. (1997) *J. Immunol.* 159, 6195-6202
 65. Le Cabec, V., and Maridonneau-Parini, I. (1994) *Biochem. J.* 303, 481-487
 66. Gilbert, P. M., and Burd, C. G. (2001) *J. Biol. Chem.* 276, 8014-8020
 67. Arbour, N. C., Lorenz, E., Schutte, B. C., Zabner, J., Kline, J. N., Jones, M., Frees, K., Watt, J. L., and Schwartz, D. A. (2000) *Nat. Genet.* 25, 187-191

Discordant Protein and mRNA Expression in Lung Adenocarcinomas*

Guoan Chen‡, Tarek G. Gharib‡, Chiang-Ching Huang§, Jeremy M. G. Taylor§, David E. Misek¶, Sharon L. R. Kardia||, Thomas J. Giordano**, Mark D. Iannettoni‡, Mark B. Orringer‡, Samir M. Hanash¶, and David G. Beer‡ ††

The relationship between gene expression measured at the mRNA level and the corresponding protein level is not well characterized in human cancer. In this study, we compared mRNA and protein expression for a cohort of genes in the same lung adenocarcinomas. The abundance of 165 protein spots representing 98 individual genes was analyzed in 76 lung adenocarcinomas and nine non-neoplastic lung tissues using two-dimensional polyacrylamide gel electrophoresis. Specific polypeptides were identified using matrix-assisted laser desorption/ionization mass spectrometry. For the same 85 samples, mRNA levels were determined using oligonucleotide microarrays, allowing a comparative analysis of mRNA and protein expression among the 165 protein spots. Twenty-eight of the 165 protein spots (17%) or 21 of 98 genes (21.4%) had a statistically significant correlation between protein and mRNA expression ($r > 0.2445$; $p < 0.05$); however, among all 165 proteins the correlation coefficient values (r) ranged from -0.467 to 0.442 . Correlation coefficient values were not related to protein abundance. Further, no significant correlation between mRNA and protein expression was found ($r = -0.025$) if the average levels of mRNA or protein among all samples were applied across the 165 protein spots (98 genes). The mRNA/protein correlation coefficient also varied among proteins with multiple isoforms, indicating potentially separate isoform-specific mechanisms for the regulation of protein abundance. Among the 21 genes with a significant correlation between mRNA and protein, five genes differed significantly between stage I and stage III lung adenocarcinomas. Using a quantitative analysis of mRNA and protein expression within the same lung adenocarcinomas, we showed that only a subset of the proteins exhibited a significant correlation with mRNA abundance. *Molecular & Cellular Proteomics* 1:304–313, 2002.

Lung cancer is the leading cause of cancer death for both men and women in the United States. Adenocarcinomas of the lung comprise ~40% of all new cases of non-small cell

lung cancer and are now the most common histologic type. Functional genomics, broadly defined as the comprehensive analysis of genes and their products, have become a recent focus of the life sciences (1). Application of these approaches to lung adenocarcinomas has the potential to aid in the identification of high risk patients with resectable early stage lung cancer that may benefit from adjuvant therapy, as well as to identify new therapeutic targets. In human lung cancer, however, little is currently understood regarding the relationship between gene expression as determined by measuring mRNA levels and the corresponding abundance of the protein products.

A number of powerful techniques for analysis of gene expression have been used including differential display (2), serial analysis of gene expression (3), DNA microarrays (4), and proteomics via two-dimensional polyacrylamide gel electrophoresis and mass spectrometry (5). Bioinformatics tools have also been developed to help determine quantitative mRNA/protein expression profiles of all types of cells and tissues (6) and now can be applied to benign and malignant tumors. DNA microarrays (cDNA and oligonucleotide) permit the parallel assessment of thousands of genes and have been utilized in gene expression monitoring (7), polymorphism analysis (8), and DNA sequencing (9). Recent studies have focused on classification or identification of subgroups of lung tumors using DNA microarrays (10, 11). The use of mRNA expression patterns by themselves, however, is insufficient for understanding the expression of protein products, as additional post-transcriptional mechanisms, including protein translation, post-translational modification, and degradation, may influence the level of a protein present in a given cell or tissue. Proteomic analyses, a complementary technology to DNA microarrays for monitoring gene expression, involves protein separation and quantitative assessment of protein spots using 2D¹-PAGE and protein identification using mass spectrometry. By combining proteomic and transcriptional analyses of the same samples, however, it may be possible to understand the complex mechanisms influencing protein expression in human cancer.

In this study, we determined mRNA and protein levels for 165 proteins (98 genes) in 76 lung adenocarcinomas and nine

From the Departments of ‡Surgery, §Biostatistics, ||Epidemiology, ¶Pathology, and ¶Pediatrics, University of Michigan, Ann Arbor, Michigan 48109

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¹ The abbreviations used are: 2D, two-dimensional; MALDI-MS, matrix-assisted laser desorption/ionization mass spectrometry.

Protein and mRNA Correlation in Lung Adenocarcinomas

TABLE I
Correlation coefficients of protein and mRNA where only one spot was present on 2D gels

r^* , correlation coefficient value > 0.2445 ; $p < 0.05$. Values in boldface are significant at $p < 0.05$.

| Spot | Unigene | Gene name | r^* | Protein name |
|------|-----------|-----------|---------------|--|
| 1104 | Hs.184510 | SFN | 0.4337 | 14-3-3 σ |
| 0994 | Hs.77840 | ANXA4 | 0.4219 | Annexin IV |
| 1314 | Hs.10958 | DJ-1 | 0.3982 | DJ-1 protein/MER5 |
| 1454 | Hs.75428 | SOD1 | 0.3863 | Superoxide dismutase (Cu-Zn) |
| 1638 | Hs.227751 | LGALS1 | 0.3318 | Galectin 1 |
| 0264 | Hs.129548 | HNRPK | 0.3034 | Transformation up-regulated nuclear protein |
| 1405 | Hs.111334 | FTL | 0.2849 | Ferritin light chain |
| 0963 | Hs.300711 | ANXA5 | 0.2468 | Annexin V |
| 1252 | Hs.4745 | PSMC | 0.2445 | 26 S proteasome p28 |
| 0906 | Hs.234489 | LDHB | 0.4420 | L-lactate dehydrogenase H chain (LDH-B) |
| 1171 | Hs.241515 | COX11 | 0.2310 | COX 11 |
| 1160 | Hs.181013 | PGAM1 | 0.2023 | Phosphoglycerate mutase |
| 0759 | Hs.74635 | DLD | 0.1965 | Dihydrolipoamide dehydrogenase precursor |
| 1193 | Hs.83383 | AOE372 | 0.1932 | Antioxidant enzyme AOE372 |
| 0172 | Hs.3069 | HSPA9B | 0.1872 | GRP75 |
| 0777 | Hs.979 | PDHB | 0.1855 | Pyruvate dehydrogenase E1- β subunit precursor |
| 1249 | Hs.226795 | GSTP1 | 0.1773 | Glutathione S-transferase pi (GST-pi) |
| 1685 | Hs.76136 | TXN | 0.1732 | Thioredoxin |
| 1205 | Hs.82314 | HPRT1 | 0.1588 | HG phosphoribosyltransferase |
| 1230 | Hs.279860 | TPT1 | 0.1466 | Translationally controlled tumor protein (TCTP) |
| 0603 | Hs.181357 | LAMR1 | 0.1463 | LAMR |
| 1358 | Hs.28914 | APRT | 0.1399 | Adenine phosphoribosyl transferase |
| 1410 | Hs.82113 | DUT | 0.1213 | dUTP pyrophosphatase (dUTPase) |
| 1825 | Hs.112378 | LIMS1 | 0.1213 | Pinch-2 protein |
| 0871 | Hs.250502 | CA8 | 0.1122 | Carbonic anhydrase-related protein; Syntaxin |
| 0289 | Hs.82916 | CCT6A | 0.1106 | Chaperonin-like protein |
| 1143 | Hs.11465 | GSTTLp28 | 0.0997 | Glutathione S-transferase homolog (GST homolog) |
| 1456 | Hs.118638 | NME1 | 0.0932 | Nm23 (NDPKA) |
| 1598 | Hs.278503 | RIG | 0.0905 | RIG (U32331) |
| 1354 | Hs.89761 | ATP5D | 0.0904 | F1F0-type ATP synthase subunit d |
| 1445 | Hs.155485 | HIP2 | 0.0843 | Huntingtin interacting protein 2 (HIP2) |
| 1479 | Hs.177486 | APP | 0.0746 | Amyloid B4A |
| 0608 | Hs.182265 | KRT19 | 0.0439 | Cytokeratin 19 |
| 1071 | Hs.10842 | RAN | 0.0277 | GTP-binding nuclear protein RAN(TC4) |
| 0991 | Hs.297939 | CTSB | 0.0254 | Cathepsin B |
| 0842 | Hs.77274 | PLAU | 0.0248 | Urokinase plasminogen activator |
| 0823 | Hs.198248 | B4GALT1 | 0.0183 | β 1,4-galactosyl transferase |
| 0613 | Hs.1247 | APOA4 | 0.0176 | Apolipoprotein A4 (ApoA4) |
| 1338 | Hs.104143 | CLTA | 0.0123 | Clathrin light chain A |
| 0902 | Hs.5123 | SID6-306 | 0.0117 | Cytosolic inorganic pyrophosphatase |
| 1688 | Hs.1473 | GRP | -0.0040 | Preprogastrin-releasing peptide |
| 0265 | Hs.274402 | HSPA1B | -0.0071 | Heat shock-induced protein |
| 1414 | Hs.77541 | ARF5 | -0.0096 | ADP-ribosylation factor 1 |
| 0710 | Hs.97206 | HIP1 | -0.0114 | Huntingtin interacting protein 1 (HIP1) |
| 0532 | Hs.170328 | MSN | -0.0132 | Moesin/E |
| 0525 | Hs.284255 | ALPP | -0.0148 | Alkaline phosphate, placental |
| 0513 | Hs.76901 | PDIR | -0.0289 | Protein disulfide isomerase-related protein 5 |
| 1659 | Hs.256697 | HINT | -0.0312 | Protein kinase C inhibitor |
| 1262 | Hs.7016 | RAB7 | -0.0362 | Rab 7 protein |
| 0190 | Hs.184411 | ALB | -0.0470 | Albumin |
| 0948 | Hs.2795 | LDHA | -0.0549 | Lactate dehydrogenase-A (LDHA) |
| 0502 | Hs.180532 | GPI | -0.0575 | Hsp89 |
| 0152 | Hs.75410 | HSPA5 | -0.0640 | GRP78 |
| 1054 | Hs.74276 | CLIC1 | -0.0686 | Nuclear chloride channel (RNCC protein) |
| 0709 | Hs.253495 | SFTPD | -0.0936 | Pulmonary surfactant protein D |
| 0867 | Hs.78996 | PCNA | -0.0982 | PCNA |
| 0165 | Hs.180414 | HSPA8 | -0.1014 | Heat shock cognate protein, 71 kDa |
| 1109 | Hs.75103 | YWHAZ | -0.1018 | 14-3-3 ζ/Δ |
| 0137 | Hs.554 | SSA2 | -0.1032 | Ro/ss-A antigen |

Protein and mRNA Correlation in Lung Adenocarcinomas

TABLE 1—continued

| Spot | Unigene | Gene name | r* | Protein name |
|------|-----------|-----------|---------|---------------------------------------|
| 0278 | Hs.4112 | TCP1 | -0.1237 | T-complex protein 1, α subunit |
| 1769 | Hs.9614 | NPM1 | -0.1738 | B23/numatrin |
| 0089 | Hs.74335 | HSPCB | -0.2049 | Hsp90 |
| 2511 | Hs.153179 | FABP5 | -0.2109 | E-FABP/FABP5 |
| 1739 | Hs.16488 | CALR | -0.2344 | Calreticulin 32 |
| 1138 | Hs.301961 | GSTM4 | -0.2438 | Glutathione S-transferase M4 (GST m4) |
| 2533 | Hs.77060 | PSMB6 | -0.2512 | Macropain subunit Δ |

non-neoplastic lung tissues. Protein levels were determined using quantitative 2D-PAGE analysis, and the separated protein polypeptides were identified using matrix-assisted laser desorption/ionization mass spectrometry (MALDI-MS). The corresponding mRNA levels for the identified proteins within the same samples were determined using oligonucleotide microarrays. Correlation analyses showed that protein abundance is likely a reflection of the transcription for a subset of proteins, but translation and post-translational modifications also appear to influence the expression levels of many individual proteins in lung adenocarcinomas.

EXPERIMENTAL PROCEDURES

Tissues—Fifty-seven stage I and 19 stage III lung adenocarcinomas, as well as nine non-neoplastic lung tissue samples, were used for protein and mRNA analyses. Patient consent was obtained, and the project was approved by the Institutional Review Board. All tissues were obtained after resection at the University of Michigan Health System between May 1991 and July 1998. Tissues were all snap-frozen in liquid nitrogen and then stored at -80°C . The patients included 46 females and 30 males ranging in age from 40.9 to 84.6 (average 63.8) years. Most patients (66/76) demonstrated a positive smoking history. Sixty-one tumor samples were classified as bronchial-derived, 14 were classified as bronchoalveolar, and one had both features. Eighteen tumor samples were classified as well differentiated, 38 were classified as moderate, and 19 were classified as poorly differentiated adenocarcinomas. Hematoxylin-stained cryostat sections (5 μm), prepared from the same tumor pieces to be utilized for protein and mRNA isolation, were evaluated by a pathologist and compared with hematoxylin- and eosin-stained sections made from paraffin blocks of the same tumors. Specimens were excluded from analysis if they showed unclear or mixed histology (e.g. adenosquamous), tumor cellularity less than 70%, potential metastatic origin as indicated by previous tumor history, extensive lymphocytic infiltration, or fibrosis or if the patient had received prior chemotherapy or radiotherapy.

Oligonucleotide Array Hybridization—The HuGeneFL oligonucleotide arrays (Affymetrix, Santa Clara, CA) containing 6800 genes were used in this study. Total RNA was isolated from all samples using Trizol reagent (Invitrogen). The resulting RNA was then subjected to further purification using RNeasy spin columns (Qiagen). Preparation of cRNA, hybridization, and scanning of the HuGeneFL arrays were performed according to the manufacturer's protocol (Affymetrix, Santa Clara, CA). Data analysis was performed using GeneChip 4.0 software. The gene expression profile of each tumor was normalized to the median gene expression profile for the entire sample. Details of data trimming and normalization are described elsewhere (11).

2D-PAGE and Quantitative Protein Analysis—Tissue for both protein and mRNA isolation came from contiguous areas of each sample. Protein separation using 2D-PAGE, silver staining, and digitization

were performed as described previously (12, 13). Our 2D-PAGE system allows us to run 20 gels at one time (one batch). Spot detection and quantification were accomplished utilizing Bio Image Visage System software (Bioimage Corp., Ann Arbor, MI). The integrated intensity of each spot was calculated as the measured optical density units $\times \text{mm}^2$. Of the total possible 2000 spots detectable on each gel, 820 spots on the gel of each sample were matched using a Gel-ed match program with the same spots on a chosen "master" gel. In each sample, 250 ubiquitously expressed reference spots were used to adjust for variations between gels, such as that created by subtle differences in protein loading or gel staining. Slight differences because of batch were corrected after spot-size quantification.

Mass Spectrometry and 2D Western Blotting—Preparative 2D gels were run using extracts from A549 lung adenocarcinoma cells (obtained from ATCC) and using the identical experimental conditions as the analytical 2D gels, except 30% more protein was loaded. The resolved protein gels were silver-stained using successive incubations in 0.02% sodium thiosulfate for 2 min, 0.1% silver nitrate for 40 min, and 0.014% formaldehyde plus 2% sodium carbonate for 10 min. For protein identification, protein polypeptides underwent trypsin digestion followed by MALDI-MS using a MALDI-TOF Voyager-DE mass spectrometer (Perseptive Biosystems, Framingham, MA). The masses were compared with known trypsin digest databases using the MS-FIT database (University of California, San Francisco; prospector.ucsf.edu/ucsfhtml3.2/msfit.htm). Some of the polypeptides included in the analysis had been identified prior to this study on the basis of sequencing (14). The identified protein spots used in this paper are shown in Fig. 1A. The method for 2D-PAGE Western blot verification was as described previously (15). The 2D Western blots of GRP58 and Op18 are shown in Fig. 1, C and E; the others, such as GRP78, GRP75, HSP70, HSC70, KRT8, KRT18, KRT19, Vimentin, ApoJ, 14-3-3, Annexin I, Annexin II, PGP9.5, DJ-1, GST-pi, and PGAM, are described elsewhere.²

Statistical Analysis—Missing values were replaced with the mean value of the protein spot. The transform $x \rightarrow \log(1 + x)$ was applied to normalize all protein expression values. The relationship between protein and mRNA expression levels within the same samples was examined using the Spearman correlation coefficient analysis (16). To identify potentially significant correlations between gene and protein expression, we used an analytical strategy similar to SAM (significance analysis of microarrays) (17), which uses a permutation technique to determine the significance of changes in gene expression between different biological states. To obtain permuted correlation coefficients between gene and protein expression, genes were exchanged first in such a way that permuted correlation coefficient were calculated based on pseudo pairs of genes and proteins. The distribution of permuted correlation coefficients became stable after 60 permutations. This procedure was then repeated 60 times to obtain 60 sets of permuted correlation coefficients. For each of the 60 permutations, the correlations of genes and proteins were ranked

² Chen et al., submitted for publication.

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TABLE II

Correlation coefficients of protein and mRNA where multiple isoforms were present on 2D gels

r^* , correlation coefficient value > 0.2445; $p < 0.05$. Values in boldface are significant at $p < 0.05$.

| Spot | Unigene | Gene name | r^* | Protein name |
|------|-----------|-----------|---------------|---|
| 1494 | Hs.81915 | LAP18 | 0.4003 | OP18 (Stathmin) |
| 0957 | Hs.77899 | TPM1 | 0.3930 | Tropomyosins 1-5 |
| 0353 | Hs.289101 | GRP58 | 0.3802 | Protease disulfide isomerase (GRP58) |
| 0855 | Hs.169476 | GAPD | 0.3693 | Glyceraldehyde-3-phosphate dehydrogenase |
| 1198 | Hs.41707 | HSPB3 | 0.3668 | Hsp27 |
| 1203 | Hs.83848 | TPI1 | 0.3395 | Triose phosphate isomerase (TPI) |
| 0523 | Hs.65114 | KRT18 | 0.3335 | Cytokeratin 18 |
| 1492 | Hs.81915 | LAP18 | 0.3234 | OP18 (Stathmin) |
| 1493 | Hs.81915 | LAP18 | 0.3154 | OP18 (Stathmin) |
| 1181 | Hs.78225 | ANXA1 | 0.3102 | Annexin variant I |
| 0439 | Hs.242463 | KRT8 | 0.3049 | Cytokeratin 8 |
| 0505 | Hs.297753 | VIM | 0.2939 | Vimentin |
| 0593 | Hs.297753 | VIM | 0.2809 | Vimentin |
| 1874 | Hs.75313 | AKR1B1 | 0.2790 | Aldose reductase |
| 0935 | Hs.75544 | YWHAH | 0.2775 | 14-3-3 η |
| 2524 | Hs.78225 | ANXA1 | 0.2612 | Annexin I |
| 2324 | Hs.65114 | KRT18 | 0.2601 | Cytokeratin 18 |
| 1192 | Hs.41707 | HSPB3 | 0.2558 | Hsp27 |
| 0350 | Hs.289101 | GRP58 | 0.2516 | Phospholipase C (GRP58) |
| 0992 | Hs.75313 | AKR1B1 | -0.2460 | Aldose reductase |
| 0861 | Hs.75313 | AKR1B1 | 0.0761 | Aldose reductase |
| 0853 | Hs.75313 | AKR1B1 | -0.0675 | Aldose reductase |
| 2503 | Hs.76392 | ALDH1 | -0.0565 | Aldehyde dehydrogenase |
| 0381 | Hs.76392 | ALDH1 | -0.0371 | Aldehyde dehydrogenase |
| 0371 | Hs.76392 | ALDH1 | -0.0680 | Aldehyde dehydrogenase |
| 1179 | Hs.78225 | ANXA1 | 0.2052 | Annexin variant I |
| 0762 | Hs.78225 | ANXA1 | -0.0739 | Annexin I |
| 0760 | Hs.78225 | ANXA1 | -0.0228 | Annexin I |
| 2506 | Hs.217493 | ANXA2 | 0.2223 | Lipocotin (annexin II) |
| 0772 | Hs.217493 | ANXA2 | 0.2080 | Lipocotin (annexin II) |
| 0723 | Hs.217493 | ANXA2 | 0.0701 | Lipocotin |
| 1239 | Hs.93194 | APOA1 | 0.1133 | Apolipoprotein A1 (ApoA1) |
| 1237 | Hs.93194 | APOA1 | -0.0373 | Apolipoprotein A1 (ApoA1) |
| 1234 | Hs.93194 | APOA1 | -0.0894 | Apolipoprotein A1 (ApoA1) |
| 0428 | Hs.25 | ATP5B | 0.0080 | ATP synthase β subunit precursor |
| 0427 | Hs.25 | ATP5B | 0.0122 | ATP synthase β subunit precursor |
| 0424 | Hs.25 | ATP5B | -0.0992 | ATP synthase β subunit precursor |
| 0863 | Hs.75106 | CLU | -0.0483 | Apolipoprotein J (ApoJ) |
| 0780 | Hs.75106 | CLU | -0.0443 | Apolipoprotein J (ApoJ) |
| 1527 | Hs.119140 | EIF5A | -0.0726 | eIF-5A |
| 1484 | Hs.119140 | EIF5A | -0.0376 | eIF-5A |
| 1728 | Hs.5241 | FABP1 | -0.1916 | L-FABP |
| 1712 | Hs.5241 | FABP1 | -0.0473 | L-FABP |
| 0947 | Hs.169476 | GAPD | 0.1745 | Glyceraldehyde-3-phosphate dehydrogenase |
| 1232 | Hs.75207 | GLO1 | 0.2249 | Glyoxalase-I |
| 1229 | Hs.75207 | GLO1 | 0.0450 | Glyoxalase-I |
| 1595 | Hs.158300 | HAP1 | -0.0137 | Huntingtin-associated protein 1 (neuroan 1) |
| 1810 | Hs.75990 | HP | -0.4672 | α -Haptoglobin |
| 1459 | Hs.75990 | HP | 0.0802 | α -Haptoglobin |
| 1458 | Hs.75990 | HP | -0.0305 | α -Haptoglobin |
| 0619 | Hs.75990 | HP | 0.0461 | B-haptoglobin |
| 0615 | Hs.75990 | HP | -0.0034 | B-haptoglobin |
| 1250 | Hs.41707 | HSPB3 | -0.1024 | Hsp27 |
| 0549 | Hs.79037 | HSPD1 | 0.1074 | Hsp60 |
| 0338 | Hs.79037 | HSPD1 | 0.2265 | Hsp60 |
| 0333 | Hs.79037 | HSPD1 | 0.1383 | Hsp60 |
| 0331 | Hs.79037 | HSPD1 | 0.1603 | Hsp60 |
| 2381 | Hs.65114 | KRT18 | 0.2016 | Cytokeratin 18 |
| 0535 | Hs.65114 | KRT18 | 0.1106 | Cytokeratin 18 |

Protein and mRNA Correlation in Lung Adenocarcinomas

TABLE II—continued

Correlation coefficients of protein and mRNA where multiple isoforms were present on 2D gels

r^* , correlation coefficient value > 0.2445 ; $p < 0.05$. Values in boldface are significant at $p < 0.05$.

| Spot | Unigene | Gene name | r^* | Protein name |
|------|-----------|-----------|---------|--|
| 0529 | Hs.65114 | KRT18 | 0.1279 | Cytokeratin 18 |
| 0528 | Hs.65114 | KRT18 | 0.0414 | Cytokeratin 18 |
| 0527 | Hs.65114 | KRT18 | 0.0436 | Cytokeratin 18 |
| 0514 | Hs.65114 | KRT18 | 0.0733 | Cytokeratin 18 |
| 0451 | Hs.242463 | KRT8 | -0.0111 | Cytokeratin 8 |
| 0446 | Hs.242463 | KRT8 | 0.0347 | Cytokeratin 8 |
| 0444 | Hs.242463 | KRT8 | -0.1311 | Cytokeratin 8 |
| 0443 | Hs.242463 | KRT8 | 0.0942 | Cytokeratin 8 |
| 1488 | Hs.81915 | LAP18 | 0.0495 | OP18 (Stathmin) |
| 0321 | Hs.75655 | P4HB | -0.0546 | PDI (proly-4-OH-B) |
| 0320 | Hs.75655 | P4HB | -0.0041 | PDI (proly-4-OH-B) |
| 1063 | Hs.75323 | PHB | 0.0441 | Prohibitin |
| 0837 | Hs.75323 | PHB | 0.1402 | Prohibitin |
| 0326 | Hs.297681 | SERPINA1 | -0.0227 | α -1-Antitripsin |
| 0322 | Hs.297681 | SERPINA1 | -0.0277 | α -1-Antitripsin |
| 0241 | Hs.297681 | SERPINA1 | -0.0148 | α -1-Antitripsin |
| 1280 | Hs.301254 | SFTPA1 | -0.1488 | Pulmonary surfactant-associated protein |
| 1278 | Hs.301254 | SFTPA1 | -0.2040 | Pulmonary surfactant-associated protein |
| 0866 | Hs.73980 | TNNT1 | 0.1162 | Troponin T |
| 0778 | Hs.73980 | TNNT1 | 0.0740 | Troponin T |
| 1213 | Hs.83848 | TPI1 | 0.0024 | Triose phosphate isomerase (TPI) |
| 1210 | Hs.83848 | TPI1 | 0.0490 | Triose phosphate isomerase (TPI) |
| 1207 | Hs.83848 | TPI1 | -0.1615 | Triose phosphate isomerase (TPI) |
| 1204 | Hs.83848 | TPI1 | 0.0209 | Triose phosphate isomerase (TPI) |
| 1202 | Hs.83848 | TPI1 | 0.0721 | Triose phosphate isomerase (TPI) |
| 1161 | Hs.83848 | TPI1 | 0.2265 | Triose phosphate isomerase (TPI) |
| 1052 | Hs.77899 | TPM1 | -0.1040 | Tropomyosin clean-product |
| 1039 | Hs.77899 | TPM1 | -0.2999 | Cytoskeletal tropomyosin |
| 1035 | Hs.77899 | TPM1 | -0.3821 | Tropomyosin |
| 0783 | Hs.77899 | TPM1 | 0.0757 | Tropomyosins 1-5 |
| 1574 | Hs.194366 | TTR | -0.0065 | Transthyretin |
| 0809 | Hs.194366 | TTR | 0.0399 | Transthyretin multimer |
| 2202 | Hs.76118 | UCHL1 | -0.0220 | Ubiquitin carboxyl-terminal hydrolase isozyme L1 |
| 1246 | Hs.76118 | UCHL1 | -0.1261 | Ubiquitin carboxyl-terminal hydrolase isozyme L1 |
| 1242 | Hs.76118 | UCHL1 | 0.1473 | Ubiquitin carboxyl-terminal hydrolase isozyme L1 |
| 0606 | Hs.297753 | VIM | 0.0951 | Vimentin |
| 0594 | Hs.297753 | VIM | -0.2664 | Vimentin-derived protein (vid4) |
| 0508 | Hs.297753 | VIM | 0.1008 | Vimentin-derived protein (vid2) |
| 0419 | Hs.297753 | VIM | 0.0032 | Vimentin-derived protein (vid1) |
| 1279 | Hs.75544 | YWHAH | 0.0059 | 14-3-3 η |

such that $\rho_p(i)$ denotes the i th largest correlation coefficient for p th permutation. Hence, the expected correlation coefficient, $\rho_E(i)$, was the average over the 60 permutations, $\rho_E(i) = \sum_{p=1}^{60} \rho_p(i)/60$. A scatter plot of observed correlations ($\rho(i)$) versus the expected correlations is shown in Fig. 2D. For this study, we chose threshold $\Delta = 0.115$ so that correlation would be considered significant if absolute value of difference between $\rho(i)$ and $\rho_E(i)$ was greater than the threshold. Twenty-nine (including one with observed correlation coefficient -0.4672) of 165 pairs of gene and protein expression were called significant in such criteria, and the permuted data generated an average of 5.1 falsely significant pairs of gene and protein expression. This provided an estimated false discovery rate (the percentage of pairs of gene and protein expression identified by chance) for our data set.

RESULTS

Correlation of Individual Proteins and mRNA Expression within Each Tumor—We have examined quantitatively 165

protein spots on 2D gels representing 98 genes and compared protein levels with mRNA levels for a cohort of 85 lung adenocarcinomas and normal lung samples. Of the 165 protein spots, 69 proteins were represented by only one known spot on 2D gels for an individual gene, whereas 96 protein spots showed multiple protein products from 29 different genes. 2D Western blotting verified the proteins identified by mass spectrometry when specific antibodies were available. Spearman correlation coefficients of the proteins and their associated mRNA for each protein spot were generated using all 76 lung adenocarcinomas and nine non-neoplastic lung tissues (see Tables I and II, and see Figs. 1 and 2). The correlation coefficients (r) ranged from -0.467 to 0.442 (Fig. 2D). A total of 28 protein spots (21 genes) were found to have a statistically significant correlation between expression of

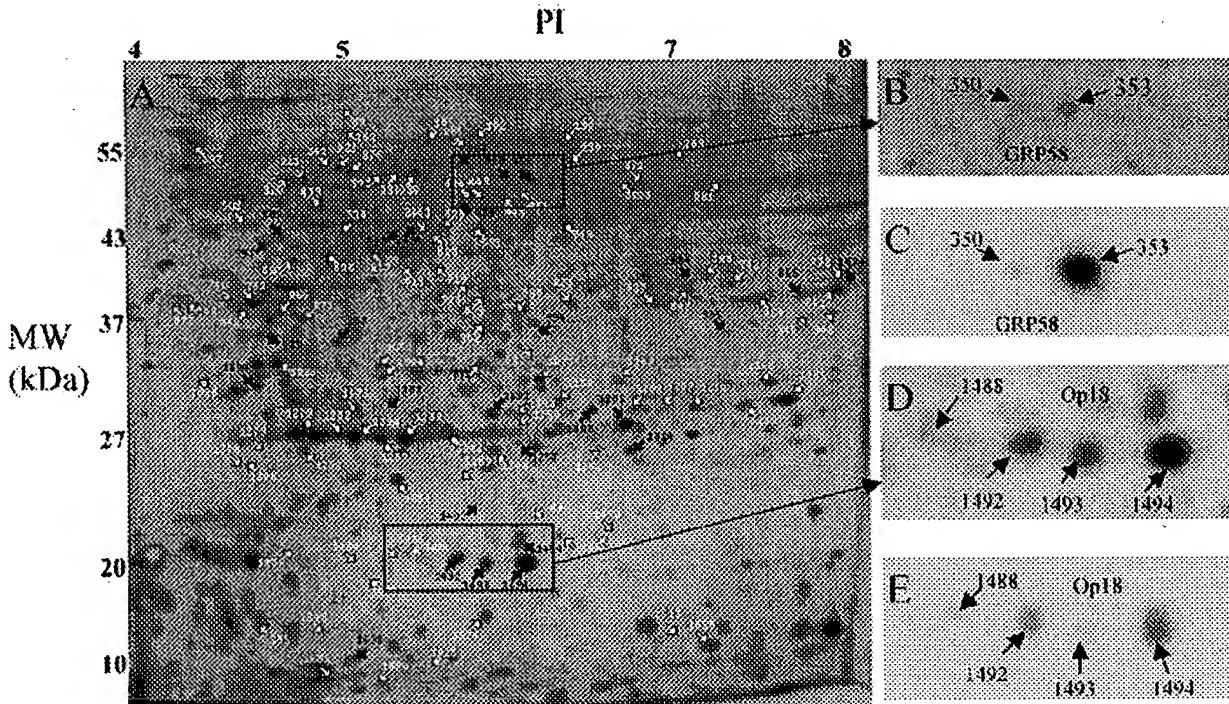


FIG. 1. A, digital image of a silver-stained 2D-PAGE separation of a stage I lung adenocarcinoma showing protein spots separated by molecular mass (MW) and isoelectric point (PI). Twenty-eight protein spots whose expression levels are correlated with mRNA abundance are indicated by the black arrows. B, the outlined areas of A showing protein GRP58. C, 2D Western blot of GRP58 from the A549 lung adenocarcinoma cell line. D, the outlined areas of A showing the protein isoforms of Op18. E, 2D Western blot of Op18 from A549 cells.

their protein and mRNA ($r > 0.2445$; $p < 0.05$). This accounts for 17% (28/165) of the 165 protein spots. Among the 69 genes for which only a single protein spot was known (Table I), nine genes (9/69, 13%) were observed to show a statistically significant relationship between protein and mRNA abundance ($r > 0.2445$; $p < 0.05$). The proteins whose expression levels were correlated with their mRNA abundance included those involved in signal transduction, carbohydrate metabolism, apoptosis, protein post-translational modification, structural proteins, and heat shock proteins (Table III).

Individual Isoforms of the Same Protein Have Different Protein/mRNA Correlation Coefficients—Of the 165 protein spots, 96 represent protein products of 29 genes with at least two isoforms. Among these 96 protein spots, 19 (19/96 protein spots, 20%) showed a statistically significant correlation between their protein and mRNA expression ($r > 0.2445$; $p < 0.05$) (Table II) and represented 12 genes (12/29, 41%). Individual isoforms of the same protein demonstrated different protein/mRNA correlation coefficients. For example, 2D-PAGE/Western analysis revealed four isoforms of OP18 differing in regards to isoelectric point but similar in molecular weight. Three of the four isoforms (spots 1492, 1493, and 1494) showed a statistically significant correlation between their protein and mRNA abundance ($r = 0.3234$, 0.3154 , and 0.4003 , respectively). The fourth isoform (spot 1488) showed no correlation be-

tween protein and mRNA expression ($r = 0.0495$). Similarly, just one of five quantified isoforms of cytokeratin 8 (spot 439) demonstrated a statistically significant correlation between protein and mRNA abundance ($r = 0.3049$; $p < 0.05$) (Table II).

In addition to differences in the relationship between mRNA levels and protein expression among separate isoforms, some genes with very comparable mRNA levels showed a 24-fold difference in their protein expression. Genes with comparable protein expression levels also showed up to a 28-fold variance in their mRNA levels.

Lack of Correlation for mRNA and Protein Expression when Using Average Tumor Values across All 165 Protein Spots (98 Genes)—The relationship between mRNA and protein expression was also examined by using the average expression values for all samples. To analyze this relationship using this approach, the average value for each protein or mRNA was generated using all 85 lung tissue samples. The range of normalized average protein values ranged from -0.0646 to 0.0979 (raw value 0.0036 to 4.1947), and the range for mRNA was from 0 to 15260.5 for all 165 individual protein spots. The Spearman correlation coefficient for the whole data set (165 protein spots/98 genes) was -0.025 (Fig. 3A). Even for the 28 protein spots (Fig. 2D) that were found to have a statistically significant correlation between their mRNA and protein, use of

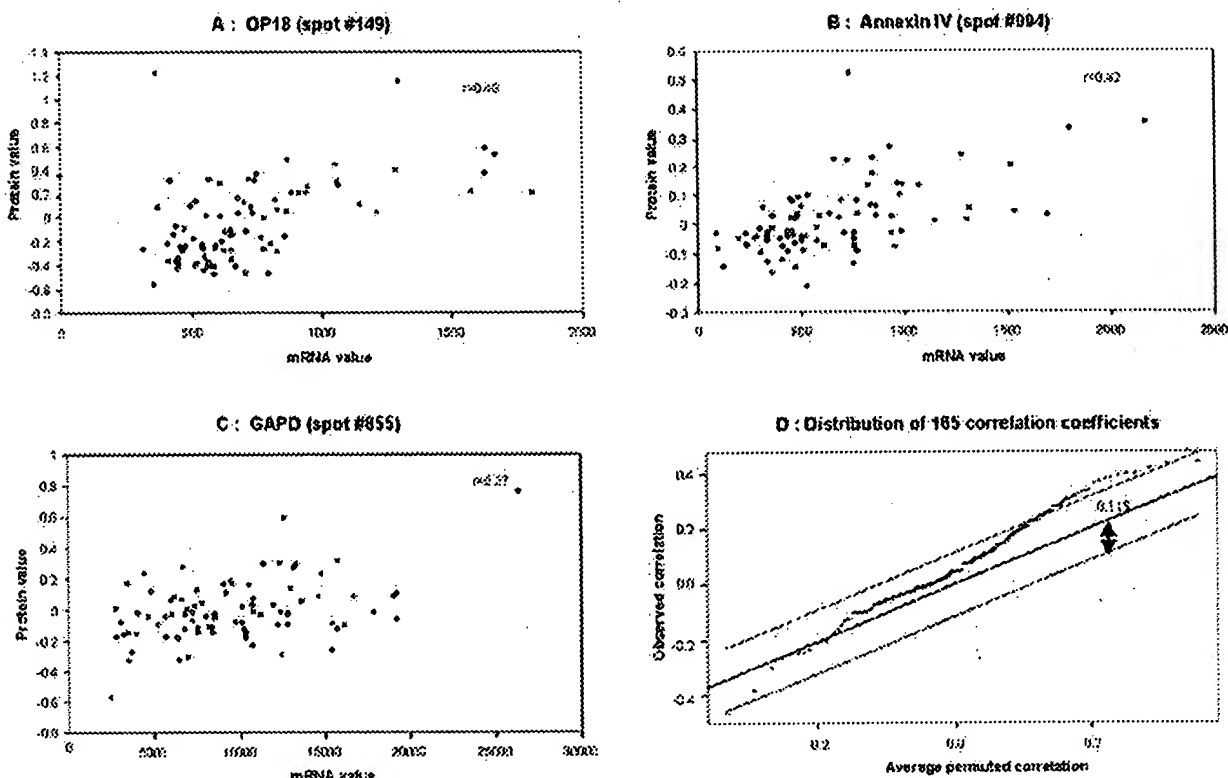


FIG. 2. A–C, plots showing the correlation between mRNA and protein for the three selected genes Op18, Annexin IV, and GAPD for all 76 lung adenocarcinomas and nine non-neoplastic lung samples ($p < 0.05$). D, distribution of all 165 Spearman correlation coefficients (r) and verification analysis using SAM. A more detailed description of the method is provided under "Experimental Procedures." Approximately 17% of the 165 proteins demonstrate a significant correlation between mRNA and protein levels as demonstrated by the values shown beyond the outer range of threshold $\Delta = 0.115$. Normalized protein values were used, thus negative values for some proteins are observed.

the average value resulted in a correlation coefficient value of -0.035 , which was not significant (Fig. 3B).

Lack of a Relationship between Protein/mRNA Correlation Coefficients and Average Protein Abundance—To determine whether an absolute protein level might influence the correlation with mRNA, the mean value of each protein (relative abundance) and the Spearman protein/mRNA correlation coefficients among all 85 samples were examined. No relationship between the protein abundance and the correlation coefficients was observed ($r = 0.039$; $p > 0.05$). A detailed analysis of separate subsets of proteins with differing levels of abundance (less than -0.0014 , larger than -0.0014 , or larger than 0.0077) also showed a lack of correlation between mRNA and protein expression among the 83 (50%), 82 (50%), and 41 (25%) of 165 total protein spots, respectively ($r = 0.016$, 0.08 , and 0.172 , respectively).

Stage-related Changes in the Protein/mRNA Correlation Coefficients—To determine whether the 21 genes (28 protein spots) showing a significant correlation between the protein and mRNA expression among all samples demonstrate changes in this relationship during tumor progression, the correlations were examined separately for stage I ($n = 57$) and

stage III ($n = 19$) lung adenocarcinomas (Table III). The number of non-neoplastic lung samples ($n = 9$) was insufficient for a separate correlation analysis of this group. Many of the protein spots represent one of several known protein isoforms for a given gene. The majority of genes (16/21) did not differ in the protein/mRNA correlation between stage I and stage III tumors indicating a similar regulatory relationship between the mRNA and protein spot. GRP-58, PSMC, SOD1, TPI1, and VIM, however, were found to demonstrate significant differences in the correlation coefficients between stage I and stage III lung adenocarcinomas. For GRP-58, PSMC, and VIM the change in the correlation coefficient was because of a relative increase in protein expression in stage III tumors. For SOD and TPI the change resulted from a relative decrease in expression of this specific protein in stage III tumors.

DISCUSSION

Relatively little is known about the regulatory mechanisms controlling the complex patterns of protein abundance and post-translational modification in tumors. Most reports concerning the regulation of protein translation have focused on

TABLE III
Stage-dependent analysis of protein-mRNA correlation coefficients

r, correlation coefficient. Values in boldface indicate a significant difference between stage I and stage III.

| Spot | Gene name | r (Stage I) | r (Stage III) | Function |
|------|-----------|---------------|---------------|--|
| 1874 | AKR1B1 | 0.269 | 0.106 | Carbohydrate metabolism; electron transporter |
| 2524 | ANXA1 | 0.184 | 0.572 | Phospholipase inhibitor; signal transduction |
| 0994 | ANXA4 | 0.660 | 0.362 | Phospholipase inhibitor |
| 0963 | ANXA5 | 0.241 | 0.390 | Phospholipase inhibitor; calcium binding; phospholipid binding |
| 1314 | DJ-1 | 0.363 | 0.354 | Signal transduction |
| 1405 | FTL | 0.126 | 0.358 | Iron storage protein |
| 0855 | GAPD | 0.243 | 0.581 | Carbohydrate metabolism (glycolysis regulation) |
| 0350 | GRP58 | 0.327 | -0.087 | Signal transduction; protein disulfide isomerase |
| 0264 | HNRPK | 0.360 | 0.243 | RNA-binding protein (RNA processing/modification) |
| 1192 | HSPB3 | 0.457 | 0.633 | Heat shock protein |
| 0523 | KRT18 | 0.115 | 0.371 | Structural protein |
| 0439 | KRT8 | 0.323 | 0.436 | Structural protein |
| 1492 | LAP18 | 0.483 | 0.663 | Signal transduction; cell growth and maintenance |
| 1638 | LGALS1 | 0.200 | 0.528 | Apoptosis; cell adhesion; cell size control |
| 1252 | PSMC | 0.253 | 0.060 | Protein degradation |
| 1104 | SFN | 0.465 | 0.475 | Signal transduction (protein kinase C inhibitor) |
| 1454 | SOD1 | 0.352 | 0.079 | Oxidoreductase |
| 1203 | TPI1 | 0.378 | 0.009 | Carbohydrate metabolism |
| 0957 | TPM1 | 0.475 | 0.225 | Structural protein (muscle); control of heart |
| 0593 | VIM | -0.054 | 0.556 | Structural protein |
| 0935 | YWHAH | 0.283 | 0.210 | Signal transduction |

one or several protein products (18). Celis *et al.* (19) found a good correlation between transcript and protein levels among 40 well resolved, abundant proteins using a proteomic and microarray study of bladder cancer. By comparing the mRNA and protein expression levels within the same tumor samples, we found that 17% (28/165) of the protein spots (21/98 genes) show a statistically significant correlation between mRNA and protein. These proteins appear to represent a diverse group of gene products and include those involved in signal transduction, carbohydrate metabolism, protein modification, cell structure, heat shock, and apoptosis. These results suggest that expression of this subset of 165 proteins is likely to be regulated at the transcriptional level in these tissues. The majority of the protein isoforms, however, did not correlate with mRNA levels, and thus their expression is regulated by other mechanisms. We also observed a subset of proteins that demonstrated a negative correlation with the mRNA expression values; for example α -haptoglobin demonstrated a strong negative correlation with its mRNA expression values. This may reflect negative feedback on the mRNA or the protein or the presence of other regulatory influences that are not understood currently.

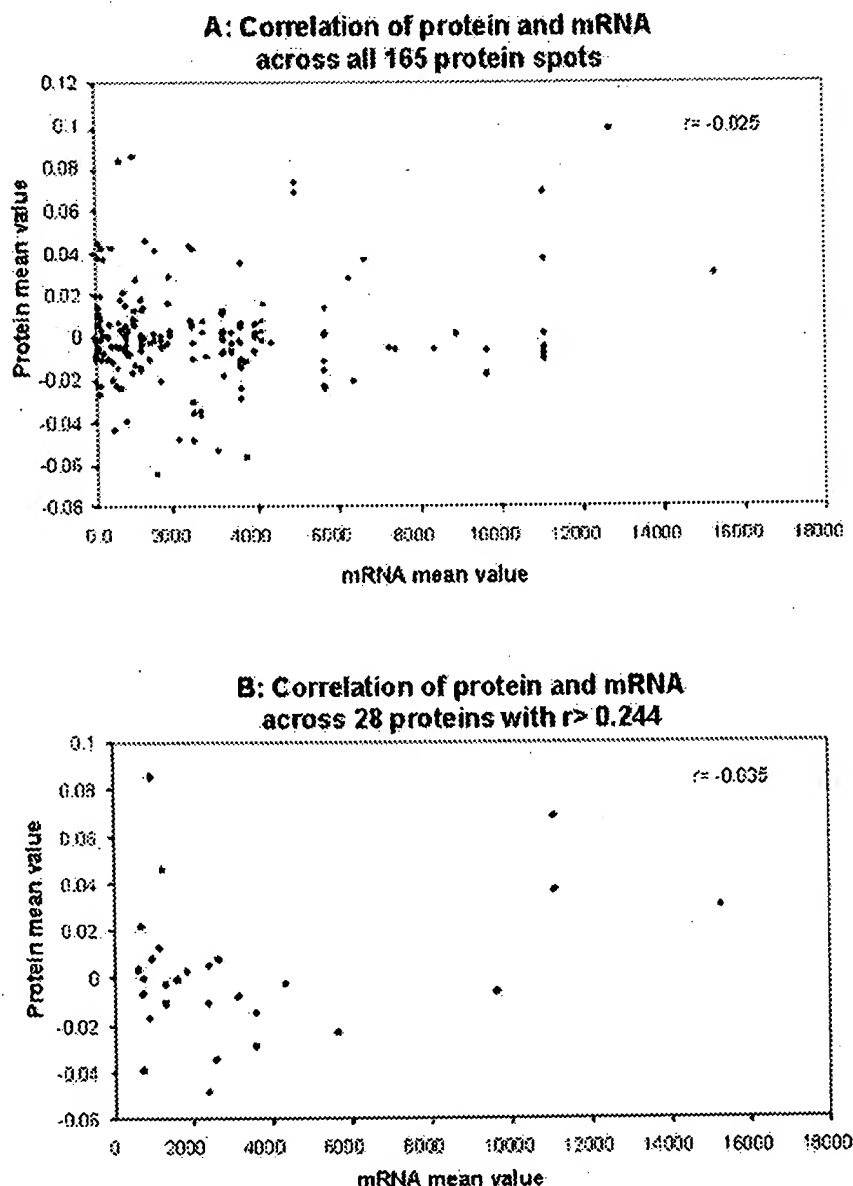
Post-translational modification or processing will result in individual protein products of the same gene migrating to different locations on 2D-PAGE gels (20). Because the identity of all possible isoforms for each protein examined has not been characterized completely, this may influence the correlation analyses performed in this study. This is partly because of limitations of the 2D-PAGE and mass spectrometry technologies (21, 22). Potential inconsistencies between mRNA and protein correlations that have been reported may also be because of differences, even in the same gene, in the mech-

anisms of protein translation among different cells or as measured in different laboratories (23).

In this study, we examined 165 protein spots identified in lung adenocarcinomas. Ninety-six protein spots, representing the products of 29 genes, contained at least two protein isoforms. Nineteen of 96 protein spots, representing 12 genes, were shown to have a statistically significant correlation between their protein and mRNA expression, suggesting that the levels of these proteins reflects the transcription of the corresponding genes. Differences in protein/mRNA correlations were found among the individual isoforms of a given protein. For example, of the four OP18 isoforms, three showed a statistically significant correlation between the protein and mRNA expression levels. The lack of relationship for the one isoform, however, indicates that individual protein isoforms of the same gene product can be regulated differentially. This is not unexpected and likely reflects other post-translational mechanisms that can influence isoform abundance in tissues and cancer.

In addition to the analyses of the correlation of mRNA/protein within the same tumor samples, we also tested the global relationship between mRNA and the corresponding protein abundance across all 165 protein spots in the lung samples. A protein and mRNA average value for each gene was generated using all 85 lung tissues samples. We observed a very wide range of normalized average protein and mRNA values. The correlation coefficient generated using this average value data set was -0.025 , and even for the 28 protein spots that showed a statistically significant correlation between individual mRNA and proteins, the correlation value was only -0.035 . This suggests that it is not possible to predict overall protein expression levels based on average

FIG. 3. The overall correlation of mRNA and protein levels across all 165 protein spots (A) and across 28 protein spots that contained individual r values larger than 0.244 (B) are shown. Each protein or mRNA mean value was calculated based on all 76 lung adenocarcinomas and nine non-neoplastic lung samples using quantitative 2D-PAGE and Affymetrix oligonucleotide microarrays. The Spearman correlation coefficients for the two data sets (A and B) were -0.025 and -0.035 , respectively, indicating a lack of correlation if mean values for mRNA and protein for all samples is used.



mRNA abundance in lung cancer samples. This conclusion is also supported by previous results from Anderson and Seilhamer (24), who examined 19 genes in human liver cells, and by Gygi *et al.* (25), who examined 106 genes in yeast. Both studies found a lack of correlation between mRNA and protein expression when average or overall levels were used.

A good correlation was reported when the 11 most abundant proteins were examined in yeast (25), suggesting that the level of protein abundance may be a factor that may influence the correlation between mRNA and protein. In the present study, a fairly wide range of mean protein values among 165 protein spots in lung adenocarcinomas was observed, and the correlation coefficients also varied from -0.467 to 0.442 .

A comparison between the mean value of each protein and the correlation coefficient generated using all 85 tissue samples did not reveal a strong relationship between the overall protein abundance and the correlation coefficients ($r = 0.039$; $p > 0.05$). Detailed analysis of different subsets of protein abundance also failed to show a correlation between mRNA and protein expression. Thus in contrast to yeast, a relationship between mRNA/protein correlation coefficient and protein abundance in human lung adenocarcinomas was not observed.

The results of this study indicate that the level of protein abundance in lung adenocarcinomas is associated with the corresponding levels of mRNA in 17% (28 proteins) of the total 165 protein spots examined. This was substantially

higher than the amount predicted to result by chance alone (which was 5.1) and suggests that a transcriptional mechanism likely underlies the abundance of these proteins in lung adenocarcinomas. We also demonstrate that the expression of individual isoforms of the same protein may or may not correlate with the mRNA, indicating that separate and likely post-translational mechanisms account for the regulation of isoform abundance. These mechanisms may also account for the differences in the correlation coefficients observed between stage I and stage III tumors, indicating that specific protein isoforms show regulatory changes during tumor progression. Further studies in lung adenocarcinomas will examine the relationship between the expression of individual protein isoforms and specific clinical-pathological features of these tumors, such as the presence of angiolymphatic invasion, and nodal or pleural surface involvement. The potential to identify specific protein isoforms associated with biological behavior in lung adenocarcinomas would be of considerable interest and will add to our understanding of the regulation of gene products by transcriptional, translational, and post-translational mechanisms.

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† To whom correspondence should be addressed: General Thoracic Surgery, SRB II, B560, Box 0686, University of Michigan Medical School, Ann Arbor, MI 48109-0886; E-mail: dgbeer@umich.edu.

REFERENCES

- Ideker, T., Thorsson, V., Raniish, J. A., Christmas, R., Buhler, J., Eng, J. K., Bumgarner, R., Goodlett, D. R., Aebersold, R., Hood, L. (2001) Integrated genomic and proteomic analyses of a systematically perturbed metabolic network. *Science* **292**, 929–934.
- Liang, P., Pardee, A. B. (1998) Differential display. A general protocol. *Mol. Biotechnol.* **10**, 261–267.
- Porter, D. A., Krop, I. E., Nasser, S., Sgroi, D., Kaelin, C. M., Marks, J. R., Riggins, G., Polyak, K. (2001) A sage (serial analysis of gene expression) view of breast tumor progression. *Cancer Res.* **61**, 5697–5702.
- Bittnner, M., Meltzer, P., Chen, Y., Jiang, Y., Seftor, E., Hendrix, M., Radmacher, M., Simon, R., Yakhini, Z., Ben-Dor, A., Sampas, N., Dougherty, E., Wang, E., Marincola, F., Gooden, C., Lueders, J., Glatfelter, A., Pollock, P., Carpen, J., Gillanders, E., Leja, D., Dietrich, K., Beaudry, C., Berens, M., Alberts, D., Sondak, V. (2000) Molecular classification of cutaneous malignant melanoma by gene expression profiling. *Nature* **406**, 536–540.
- Fung, E. T., Wright, G. L., Jr., Dalmasso, E. A. (2000) Proteomic strategies for biomarker identification: progress and challenges. *Curr. Opin. Mol. Ther.* **2**, 643–650.
- Davidson, D., Baldock, R. (2001) Bioinformatics beyond sequence: mapping gene function in the embryo. *Nat. Rev. Genet.* **2**, 409–417.
- Chee, M., Yang, R., Hubbell, E., Berno, A., Huang, X. C., Stern, D., Winkler, J., Lockhart, D. J., Morris, M. S., Fodor, S. P. (1996) Accessing genetic information with high-density DNA arrays. *Science* **274**, 610–614.
- Wang, D. G., Fan, J. B., Siao, C. J., Berno, A., Young, P., Sapolsky, R., Ghandour, G., Perkins, N., Winchester, E., Spencer, J., Kruglyak, L., Stein, L., Hsie, L., Topaloglou, T., Hubbell, E., Robinson, E., Mittmann, M., Morris, M. S., Shen, N., Kilburn, D., Rioux, J., Nusbaum, C., Rozen, S., Hudson, T. J., Lander, E. S. (1998) Large-scale identification, mapping, and genotyping of single-nucleotide polymorphisms in the human genome. *Science* **280**, 1077–1082.
- Pease, A. C., Solas, D., Sullivan, E. J., Cronin, M. T., Holmes, C. P., Fodor, S. P. (1994) Light-generated oligonucleotide arrays for rapid DNA sequence analysis. *Proc. Natl. Acad. Sci. U. S. A.* **91**, 5022–5026.
- Bhattacharjee, A., Richards, W. G., Staunton, J., Li, C., Monti, S., Vasa, P., Ladd, C., Beheshti, J., Bueno, R., Gillette, M., Loda, M., Weber, G., Mark, E. J., Lander, E. S., Wong, W., Johnson, B. E., Golub, T. R., Sugarbaker, D. J., Meyerson, M. (2001) Classification of human lung carcinomas by mRNA expression profiling reveals distinct adenocarcinoma subclasses. *Proc. Natl. Acad. Sci. U. S. A.* **98**, 13790–13795.
- Giordano, T. J., Shedden, K. A., Schwartz, D. R., Kuick, R., Taylor, J. M. G., Lee, N., Misek, D. E., Greenon, J. K., Kardias, S. L. R., Beer, D. G., Rennert, G., Cho, K. R., Gruber, S. B., Fearon, E. R., Hanash, S. (2001) Organ-specific molecular classification of lung, colon and ovarian adenocarcinomas using gene expression profiles. *Am. J. Pathol.* **159**, 1231–1238.
- Strahler, J. R., Kuick, R., Hanash, S. M. (1989) in *Protein Structure: A Practical Approach* (Creighton, T., ed) pp. 65–92, IRL Press, Oxford.
- Merril, C. R., Dunau, M. L., Goldman, D. (1981) A rapid sensitive silver stain for polypeptides in polyacrylamide gels. *Anal. Biochem.* **101**, 201–207.
- Hanash, S. M., Strahler, J. R., Chan, Y., Kuick, R., Teichroew, D., Neel, J. V., Hailat, N., Keim, D. R., Gratiot-Deans, J., Ungar, D., Richardson, B. C. (1993) Data base analysis of protein expression patterns during T-cell ontogeny and activation. *Proc. Natl. Acad. Sci. U. S. A.* **90**, 3314–3318.
- Brichory, F. M., Misek, D. E., Yim, A. M., Krause, M. C., Giordano, T. J., Beer, D. G., Hanash, S. M. (2001) An immune response manifested by the common occurrence of annexins I and II autoantibodies and high circulating levels of IL-6 in lung cancer. *Proc. Natl. Acad. Sci. U. S. A.* **98**, 9824–9829.
- Lavens-Phillips, S. E., MacGlashan, D. W., Jr. (2000) The tyrosine kinases p53/56lyn and p72syk are differentially expressed at the protein level but not at the messenger RNA level in nonreleasing human basophils. *Am. J. Respir. Cell Mol. Biol.* **23**, 566–571.
- Tusher, V. G., Tibshirani, R., Chu, G. (2001) Significance analysis of microarrays applied to the ionizing radiation response. *Proc. Natl. Acad. Sci. U. S. A.* **98**, 5116–5121.
- Tew, K. D., Monks, A., Barone, L., Rosser, D., Akerman, G., Montali, J. A., Wheatley, J. B., Schmidt, D. E., Jr. (1996) Glutathione-associated enzymes in the human cell lines of the National Cancer Institute Drug Screening Program. *Mol. Pharmacol.* **50**, 149–159.
- Celis, J. E., Kruhoffer, M., Gromova, I., Frederiksen, C., Ostergaard, M., Thykjaer, T., Gromov, P., Yu, J., Palsdottir, H., Magnusson, N., Orntoft, T. F. (2000) Gene expression profiling: monitoring transcription and translation products using DNA microarrays and proteomics. *FEBS Lett.* **480**, 2–16.
- Anderson, N. L., Anderson, N. G. (1998) Proteome and proteomics: new technologies, new concepts, and new words. *Electrophoresis* **19**, 1853–1861.
- Gygi, S. P., Corthals, G. L., Zhang, Y., Rochon, Y., Aebersold, R. (2000) Evaluation of two-dimensional gel electrophoresis-based proteome analysis technology. *Proc. Natl. Acad. Sci. U. S. A.* **97**, 9390–9395.
- Fey, S. J., Larsen, P. M. (2001) 2D or not 2D. Two-dimensional gel electrophoresis. *Curr. Opin. Chem. Biol.* **5**, 26–33.
- McBride, S., Walsh, D., Meleady, P., Daly, N., Clynes, M. (1999) Bromodeoxyuridine induces keratin protein synthesis at a posttranscriptional level in human lung tumor cell lines. *Differentiation* **64**, 185–193.
- Anderson, L., Seilhamer, J. (1997) A comparison of selected mRNA and protein abundances in human liver. *Electrophoresis* **18**, 533–537.
- Gygi, S. P., Rochon, Y., Franz, B. R., Aebersold, R. (1999) Correlation between protein and mRNA abundance in yeast. *Mol. Cell. Biol.* **19**, 1720–1730.



An increased high-mobility group A2 expression level is associated with malignant phenotype in pancreatic exocrine tissue.

Abe N, Watanabe T, Suzuki Y, Matsumoto N, Masaki T, Mori T, Sugiyama M, Chiappetta G, Fusco A, Atomi Y.

First Department of Surgery, Kyorin University School of Medicine, 6-20-2, Shinkawa, Mitaka, Tokyo 181-8611, Japan. abenbtg@kyorin-u.ac.jp

The altered form of the high-mobility group A2 (HMGA2) gene is somehow related to the generation of human benign and malignant tumours of mesenchymal origin. However, only a few data on the expression of HMGA2 in malignant tumour originating from epithelial tissue are available. In this study, we examined the HMGA2 expression level in pancreatic carcinoma, and investigated whether alterations in the HMGA2 expression level are associated with a malignant phenotype in pancreatic tissue. High-mobility group A2 mRNA and protein expression was determined in eight surgically resected specimens of non-neoplastic tissue (six specimens of normal pancreatic tissue and two of chronic pancreatitis tissue) and 27 pancreatic carcinomas by highly sensitive reverse transcriptase-polymerase chain reaction (RT-PCR) techniques and immunohistochemical staining, respectively. Reverse transcriptase-polymerase chain reaction analysis revealed the expression of the HMGA2 gene in non-neoplastic pancreatic tissue, although its expression level was significantly lower than that in carcinoma. Immunohistochemical analysis indicated that the presence of the HMGA2 gene in non-neoplastic pancreatic tissue observed in RT-PCR reflects its abundant expression in islet cells, together with its focal expression in duct epithelial cells. Intense and multifocal or diffuse HMGA2 immunoreactivity was noted in all the pancreatic carcinoma examined. A strong correlation between HMGA2 overexpression and the diagnosis of carcinoma was statistically verified. Based on these findings, we propose that an increased expression level of the HMGA2 protein is closely associated with the malignant phenotype in the pancreatic exocrine system, and accordingly, HMGA2 could serve as a potential diagnostic molecular marker for distinguishing pancreatic malignant cells from non-neoplastic pancreatic exocrine cells.

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Bruce Alberts • Dennis Bray
Julian Lewis • Martin Raff • Keith Roberts
James D. Watson



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Bruce Alberts received his Ph.D. from Harvard University and is currently President of the National Academy of Sciences and Professor of Biochemistry and Biophysics at the University of California, San Francisco. *Dennis Bray* received his Ph.D. from the Massachusetts Institute of Technology and is currently a Medical Research Council Fellow in the Department of Zoology, University of Cambridge.

Julian Lewis received his D.Phil. from the University of Oxford and is currently a Senior Scientist in the Imperial Cancer Research Fund Developmental Biology Unit, University of Oxford. *Martin Raff* received his M.D. from McGill University and is currently a Professor in the MRC Laboratory for Molecular Cell Biology and the Biology Department, University College, London. *Keith Roberts* received his Ph.D. from the University of Cambridge and is currently Head of the Department of Cell Biology, the John Innes Institute, Norwich. *James D. Watson* received his Ph.D. from Indiana University and is currently Director of the Cold Spring Harbor Laboratory. He is the author of *Molecular Biology of the Gene* and, with Francis Crick and Maurice Wilkins, won the Nobel Prize in Medicine and Physiology in 1962.

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Front cover: The photograph shows a rat nerve cell in culture. It is labeled with a fluorescent antibody that stains its cell body and dendritic processes (yellow). Nerve terminals (green) from other neurons (not visible), which have made synapses on the cell, are labeled with a different antibody (Courtesy of Old Mondigli and Pietro de Camilli)

Dedication page: Gavin Borden, late president of Garland Publishing, weathered in during his mid-1980s climb near Mount McKinley with AlRat (author Bruce Alberts and famous mountaineer guide Edogs Stamp) 1940-1992

Back cover: The authors, in alphabetical order, crossing Abbey Road in London on their way to lunch. Much of this third edition was written in a house just around the corner. (Photograph by Richard Oliver)

extracts. If these minor cell proteins differ among cells to the same extent as the more abundant proteins, as is commonly assumed, only a small number of protein differences (perhaps several hundred) suffice to create very large differences in cell morphology and behavior.

A Cell Can Change the Expression of Its Genes in Response to External Signals³

Most of the specialized cells in a multicellular organism are capable of altering their patterns of gene expression in response to extracellular cues. If a liver cell is exposed to a glucocorticoid hormone, for example, the production of several specific proteins is dramatically increased. Glucocorticoids are released during periods of starvation or intense exercise and signal the liver to increase the production of glucose from amino acids and other small molecules; the set of proteins whose production is induced includes enzymes such as tyrosine aminotransferase, which helps to convert tyrosine to glucose. When the hormone is no longer present, the production of these proteins drops to its normal level.

Other cell types respond to glucocorticoids in different ways. In fat cells, for example, the production of tyrosine aminotransferase is reduced, while some other cell types do not respond to glucocorticoids at all. These examples illustrate a general feature of cell specialization—different cell types often respond in different ways to the same extracellular signal. Underlying this specialization are features that do not change, which give each cell type its permanently distinctive character. These features reflect the persistent expression of different sets of genes.

Gene Expression Can Be Regulated at Many of the Steps in the Pathway from DNA to RNA to Protein⁴

If differences between the various cell types of an organism depend on the particular genes that the cells express, at what level is the control of gene expression exercised? There are many steps in the pathway leading from DNA to protein, and all of them can in principle be regulated. Thus a cell can control the proteins it makes by (1) controlling when and how often a given gene is transcribed (**transcriptional control**), (2) controlling how the primary RNA transcript is spliced or otherwise processed (**RNA processing control**), (3) selecting which completed mRNAs in the cell nucleus are exported to the cytoplasm (**RNA transport control**), (4) selecting which mRNAs in the cytoplasm are translated by ribosomes (**translational control**), (5) selectively destabilizing certain mRNA molecules in the cytoplasm (**mRNA degradation control**), or (6) selectively activating, inactivating, or compartmentalizing specific protein molecules after they have been made (**protein activity control**) (Figure 9-2).

For most genes transcriptional controls are paramount. This makes sense because, of all the possible control points illustrated in Figure 9-2, only transcriptional control ensures that no superfluous intermediates are synthesized. In the

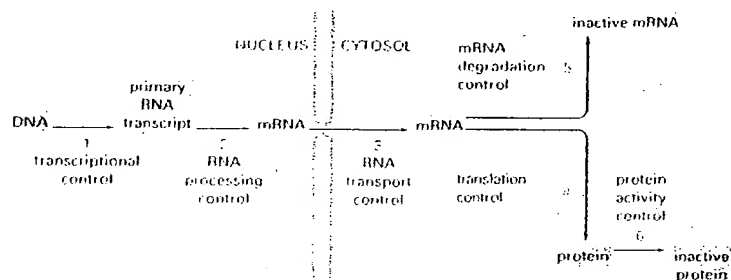


Figure 9-2 Six steps at which eucaryote gene expression can be controlled. Only controls that operate at steps 1 through 5 are discussed in this chapter. The regulation of protein activity (step 6) is discussed in Chapter 5; this includes reversible activation or inactivation by protein phosphorylation as well as irreversible inactivation by proteolytic degradation.

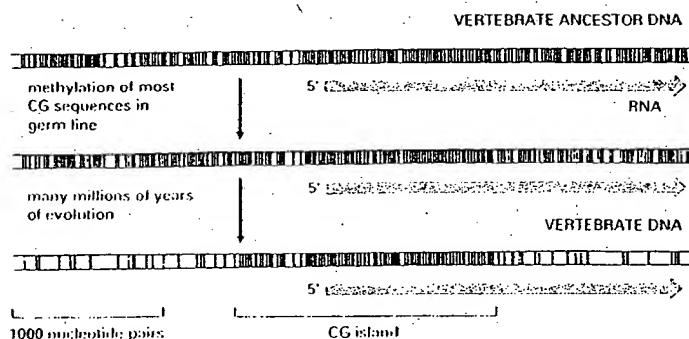


Figure 9-71 A mechanism to explain both the marked deficiency of CG sequences and the presence of CG islands in vertebrate genomes. A black line marks the location of an unmethylated CG dinucleotide in the DNA sequence, while a red line marks the location of a methylated CG dinucleotide.

Summary

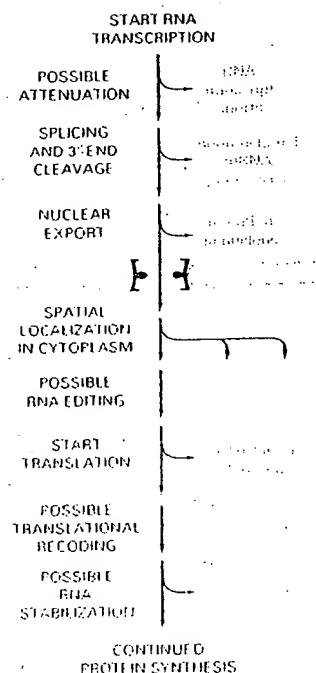
The many types of cells in animals and plants are created largely through mechanisms that cause different genes to be transcribed in different cells. Since many specialized animal cells can maintain their unique character when grown in culture, the gene regulatory mechanisms involved in creating them must be stable once established and heritable when the cell divides, endowing the cell with a memory of its developmental history. Prokaryotes and yeasts provide unusually accessible model systems in which to study gene regulatory mechanisms, some of which may be relevant to the creation of specialized cell types in higher eucaryotes. One such mechanism involves a competitive interaction between two (or more) gene regulatory proteins, each of which inhibits the synthesis of the other; this can create a flip-flop switch that switches a cell between two alternative patterns of gene expression. Direct or indirect positive feedback loops, which enable gene regulatory proteins to perpetuate their own synthesis, provide a general mechanism for cell memory.

In eucaryotes gene transcription is generally controlled by combinations of gene regulatory proteins. It is thought that each type of cell in a higher eucaryotic organism contains a specific combination of gene regulatory proteins that ensures the expression of only those genes appropriate to that type of cell. A given gene regulatory protein may be expressed in a variety of circumstances and typically is involved in the regulation of many genes.

In addition to diffusible gene regulatory proteins, inherited states of chromatin condensation are also utilized by eucaryotic cells to regulate gene expression. In vertebrates DNA methylation also plays a part, mainly as a device to reinforce decisions about gene expression that are made initially by other mechanisms.

Although controls on the initiation of gene transcription are the predominant form of regulation for most genes, other controls can act later in the pathway from RNA to protein to modulate the amount of gene product that is made. Although these posttranscriptional controls, which operate after RNA polymerase has bound to the gene's promoter and begun RNA synthesis, are less common than transcriptional control, for many genes they are crucial. It seems that every step in gene expression that could be controlled in principle is likely to be regulated under some circumstances for some genes.

We consider the varieties of posttranscriptional regulation in temporal order, according to the sequence of events that might be experienced by an RNA molecule after its transcription has begun (Figure 9-72).



Possible post-transcriptional controls on gene expression. Only a few of these controls are likely to be used for any one gene.

MOLECULAR BIOLOGY OF
THE CELL

fourth edition

Bruce Alberts

Alexander Johnson

Julian Lewis

Martin Raff

Keith Roberts

Peter Walter

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Bruce Alberts received his Ph.D. from Harvard University and is President of the National Academy of Sciences and Professor of Biochemistry and Biophysics at the University of California, San Francisco. **Alexander Johnson** received his Ph.D. from Harvard University and is a Professor of Microbiology and Immunology at the University of California, San Francisco. **Julian Lewis** received his D.Phil. from the University of Oxford and is a Principal Scientist at the Imperial Cancer Research Fund, London. **Martin Raff** received his M.D. from McGill University and is at the Medical Research Council Laboratory for Molecular Cell Biology and Cell Biology Unit and in the Biology Department at University College London. **Keith Roberts** received his Ph.D. from the University of Cambridge and is Associate Research Director at the John Innes Centre, Norwich. **Peter Walter** received his Ph.D. from The Rockefeller University in New York and is Professor and Chairman of the Department of Biochemistry and Biophysics at the University of California, San Francisco, and an Investigator of the Howard Hughes Medical Institute.

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Cell Biology Interactive

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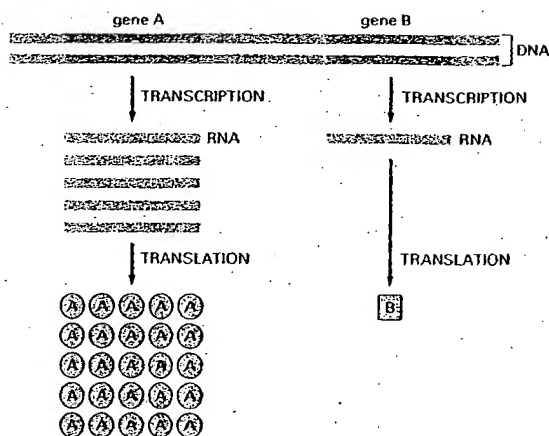


Figure 6-3 Genes can be expressed with different efficiencies. Gene A is transcribed and translated much more efficiently than gene B. This allows the amount of protein A in the cell to be much greater than that of protein B.

FROM DNA TO RNA

Transcription and translation are the means by which cells read out, or express, the genetic instructions in their genes. Because many identical RNA copies can be made from the same gene, and each RNA molecule can direct the synthesis of many identical protein molecules, cells can synthesize a large amount of protein rapidly when necessary. But each gene can also be transcribed and translated with a different efficiency, allowing the cell to make vast quantities of some proteins and tiny quantities of others (Figure 6-3). Moreover, as we see in the next chapter, a cell can change (or regulate) the expression of each of its genes according to the needs of the moment—most obviously by controlling the production of its RNA.

Portions of DNA Sequence Are Transcribed into RNA

The first step a cell takes in reading out a needed part of its genetic instructions is to copy a particular portion of its DNA nucleotide sequence—a gene—into an RNA nucleotide sequence. The information in RNA, although copied into another chemical form, is still written in essentially the same language as it is in DNA—the language of a nucleotide sequence. Hence the name **transcription**.

Like DNA, RNA is a linear polymer made of four different types of nucleotide subunits linked together by phosphodiester bonds (Figure 6-4). It differs from DNA chemically in two respects: (1) the nucleotides in RNA are *ribonucleotides*—that is, they contain the sugar ribose (hence the name *ribonucleic acid*) rather than deoxyribose; (2) although, like DNA, RNA contains the bases adenine (A), guanine (G), and cytosine (C), it contains the base uracil (U) instead of the thymine (T) in DNA. Since U, like T, can base-pair by hydrogen-bonding with A (Figure 6-5), the complementary base-pairing properties described for DNA in Chapters 4 and 5 apply also to RNA (in RNA, G pairs with C, and A pairs with U). It is not uncommon, however, to find other types of base pairs in RNA: for example, G pairing with U occasionally.

Despite these small chemical differences, DNA and RNA differ quite dramatically in overall structure. Whereas DNA always occurs in cells as a double-stranded helix, RNA is single-stranded. RNA chains therefore fold up into a variety of shapes, just as a polypeptide chain folds up to form the final shape of a protein (Figure 6-6). As we see later in this chapter, the ability to fold into complex three-dimensional shapes allows some RNA molecules to have structural and catalytic functions.

Transcription Produces RNA Complementary to One Strand of DNA

All of the RNA in a cell is made by DNA transcription, a process that has certain similarities to the process of DNA replication discussed in Chapter 5.

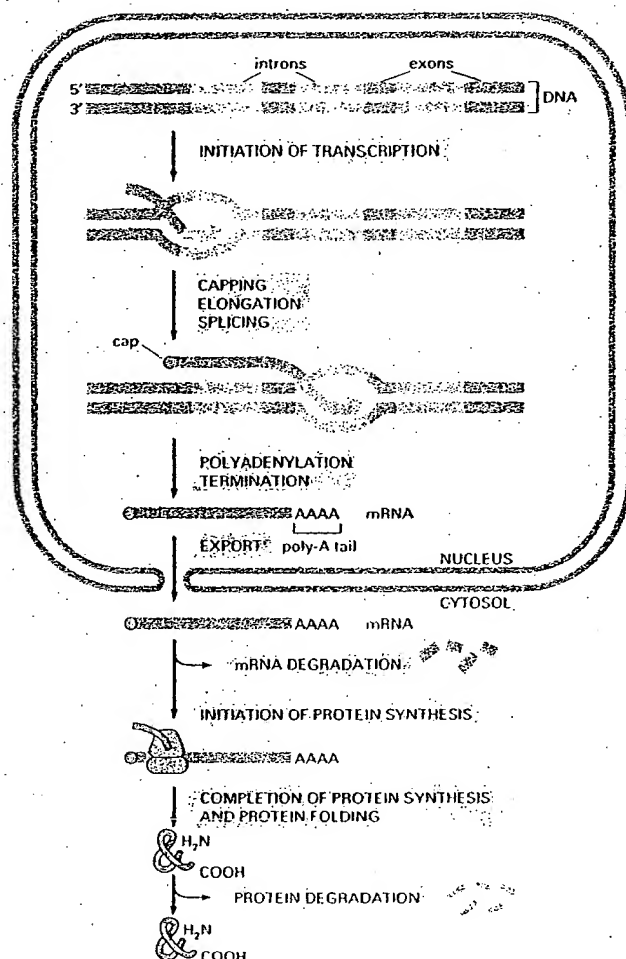


Figure 6-90 The production of a protein by a eucaryotic cell. The final level of each protein in a eucaryotic cell depends upon the efficiency of each step depicted.

ure 6-90) could be regulated by the cell for each individual protein. However, as we shall see in Chapter 7, the initiation of transcription is the most common point for a cell to regulate the expression of each of its genes. This makes sense, inasmuch as the most efficient way to keep a gene from being expressed is to block the very first step—the transcription of its DNA sequence into an RNA molecule.

Summary

The translation of the nucleotide sequence of an mRNA molecule into protein takes place in the cytoplasm on a large ribonucleoprotein assembly called a ribosome. The amino acids used for protein synthesis are first attached to a family of tRNA molecules, each of which recognizes, by complementary base-pair interactions, particular sets of three nucleotides in the mRNA (codons). The sequence of nucleotides in the mRNA is then read from one end to the other in sets of three according to the genetic code.

To initiate translation, a small ribosomal subunit binds to the mRNA molecule at a start codon (AUG) that is recognized by a unique initiator tRNA molecule. A large ribosomal subunit binds to complete the ribosome and begin the elongation phase of protein synthesis. During this phase, aminoacyl tRNAs—each bearing a specific amino acid bind sequentially to the appropriate codon in mRNA by forming complementary base pairs with the tRNA anticodon. Each amino acid is added to the C-terminal end of the growing polypeptide by means of a cycle of three sequential

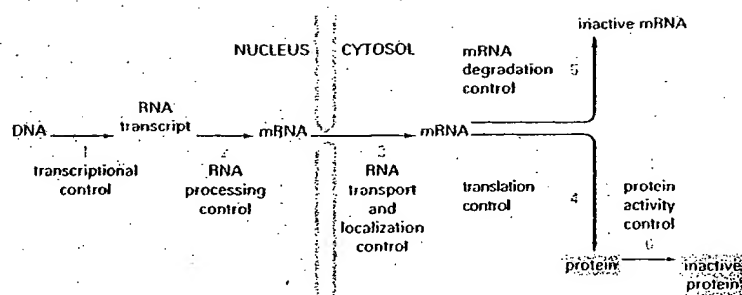


Figure 7-5 Six steps at which eucaryotic gene expression can be controlled. Controls that operate at steps 1 through 5 are discussed in this chapter. Step 6, the regulation of protein activity, includes reversible activation or inactivation by protein phosphorylation (discussed in Chapter 3) as well as irreversible inactivation by proteolytic degradation (discussed in Chapter 6).

Gene Expression Can Be Regulated at Many of the Steps in the Pathway from DNA to RNA to Protein

If differences among the various cell types of an organism depend on the particular genes that the cells express, at what level is the control of gene expression exercised? As we saw in the last chapter, there are many steps in the pathway leading from DNA to protein, and all of them can in principle be regulated. Thus a cell can control the proteins it makes by (1) controlling when and how often a given gene is transcribed (**transcriptional control**), (2) controlling how the RNA transcript is spliced or otherwise processed (**RNA processing control**), (3) selecting which completed mRNAs in the cell nucleus are exported to the cytosol and determining where in the cytosol they are localized (**RNA transport and localization control**), (4) selecting which mRNAs in the cytoplasm are translated by ribosomes (**translational control**), (5) selectively destabilizing certain mRNA molecules in the cytoplasm (**mRNA degradation control**), or (6) selectively activating, inactivating, degrading, or compartmentalizing specific protein molecules after they have been made (**protein activity control**) (Figure 7-5).

For most genes transcriptional controls are paramount. This makes sense because, of all the possible control points illustrated in Figure 7-5, only transcriptional control ensures that the cell will not synthesize superfluous intermediates. In the following sections we discuss the DNA and protein components that perform this function by regulating the initiation of gene transcription. We shall return at the end of the chapter to the additional ways of regulating gene expression.

Summary

The genome of a cell contains in its DNA sequence the information to make many thousands of different protein and RNA molecules. A cell typically expresses only a fraction of its genes, and the different types of cells in multicellular organisms arise because different sets of genes are expressed. Moreover, cells can change the pattern of genes they express in response to changes in their environment, such as signals from other cells. Although all of the steps involved in expressing a gene can in principle be regulated, for most genes the initiation of RNA transcription is the most important point of control.

How does a cell determine which of its thousands of genes to transcribe? As mentioned briefly in Chapters 4 and 6, the transcription of each gene is controlled by a regulatory region of DNA relatively near the site where transcription begins. Some regulatory regions are simple and act as switches that are thrown by a single signal. Many others are complex and act as tiny microprocessors, responding to a variety of signals that they interpret and integrate to switch the neighboring gene on or off. Whether complex or simple, these switching devices



Selective apoptosis of natural killer-cell tumours by l-asparaginase.

Ando M, Sugimoto K, Kitoh T, Sasaki M, Mukai K, Ando J, Egashira M, Schuster SM, Oshimi K.

Department of Haematology, Juntendo University School of Medicine, Tokyo, Japan.

We examined the effectiveness of various anti-tumour agents to natural killer (NK)-cell tumour cell lines and samples, which are generally resistant to chemotherapy, using flow cytometric terminal deoxynucleotidyl-transferase-mediated dUTP-biotin nick end-labelling (TUNEL) assay. Although NK-YS and NK-92 were highly resistant to various anti-tumour agents, l-asparaginase induced apoptosis in these two NK-cell lines. NK-cell leukaemia/lymphoma and acute lymphoblastic leukaemia (ALL) samples were selectively sensitive to l-asparaginase and to doxorubicin (DXR) respectively. Samples of chronic NK lymphocytosis, an NK-cell disorder with an indolent clinical course, were resistant to both drugs. Our study clearly separated two major categories of NK-cell disorders and ALL according to the sensitivity to DXR and l-asparaginase. We examined asparagine synthetase levels by real-time quantitative polymerase chain reaction (RQ-PCR) and immunostaining in these samples. At least in nasal-type NK-cell lymphoma, there was a good correlation among asparagine synthetase expression, in vitro sensitivity and clinical response to l-asparaginase. In aggressive NK-cell leukaemia, although asparagine synthetase expression was high at both mRNA and protein levels, l-asparaginase induced considerable apoptosis. Furthermore, samples of each disease entity occupied a distinct area in two-dimensional plotting with asparagine synthetase mRNA level (RQ-PCR) and in vitro l-asparaginase sensitivity (TUNEL assay). We confirmed rather specific anti-tumour activity of l-asparaginase against NK-cell tumours in vitro, which provides an experimental background to the clinical use of l-asparaginase for NK-cell tumours.

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Human thyroid carcinoma cell lines and normal thyrocytes: expression and regulation of matrix metalloproteinase-1 and tissue matrix metalloproteinase inhibitor-1 messenger-RNA and protein.

Aust G, Hofmann A, Laue S, Rost A, Kohler T, Scherbaum WA.

Institut of Anatomy, University of Leipzig, Germany.

Matrix metalloproteinase-1 (MMP-1) and tissue matrix metalloproteinase inhibitor 1 (TIMP-1) play an important role in remodeling the extracellular matrix in normal and pathological processes. The effect of phorbol-myristate acetate (PMA), interleukin-1 (IL-1), and tumor necrosis factor-alpha (TNF-alpha) on MMP-1 and TIMP-1 expression was studied on highly purified thyrocytes and undifferentiated 8505 C, C 643, HTh 74, SW 1736 thyroid carcinoma cells compared with thyroid-derived fibroblasts. Messenger RNA (mRNA) levels were monitored by competitive semiquantitative reverse transcriptase polymerase chain reaction (RT-PCR) after 24 hours. Culture supernatants were assayed for free and/or complexed MMP-1 and TIMP-1 after 48 hours using enzyme-linked immunosorbent assay (ELISA) systems (detection limit: <2 ng/mL). MMP-1 and TIMP-1 mRNA were present in all cell types, although thyrocytes showed MMP-1 mRNA levels near the detection limit. 8505 C expressed MMP-1 mRNA levels of up to 10(6) times those of the other cells analyzed. PMA and IL-1 increased MMP-1 mRNA in most cell types. TIMP-1 mRNA increased after treatment with PMA in all cells except 8505 C, whereas only slight effects were shown after IL-1 stimulation. MMP-1 protein was undetectable in normal thyrocyte cultures, but was secreted spontaneously by all cell lines ([ng/mL]; C 643: 15+/-7; HTh 74: 81+/-1; SW 1736: 13+/-2; 8505 C: 2097+/-320). There was a strong correlation between levels of MMP-1 mRNA and protein ($r = 0.99$, $p < .0001$). PMA and IL-1 increased MMP-1 secretion in all cell types after 48 hours. Fibroblasts ([ng/mL] 517+/-55) and the cell lines (C 643: 142+/-48; HTh 74: 115+/-13; SW 1736: 202+/-14; 8505C: 120+/-19) secreted TIMP-1 in unstimulated cultures, whereas only a trace amount was detected in thyrocyte cultures, even after PMA treatment. IL-1 upregulated TIMP-1 secretion after 48 hours in SW 1736, HTh 74, and C 643 cells. Our data suggest that in contrast to normal thyrocytes, dedifferentiated thyroid carcinoma cell lines are potential producers of MMP-1 as well as TIMP-1. High MMP-1 or MMP-1/TIMP-1 expression may play a role in tissue invasion of undifferentiated thyroid cancer cells.

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CLINICAL RESEARCH

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Human Thyroid Carcinoma Cell Lines and Normal Thyrocytes: Expression and Regulation of Matrix Metalloproteinase-1 and Tissue Matrix Metalloproteinase Inhibitor-1 Messenger-RNA and Protein

G. AUST,¹ A. HOFMANN,² S. LAUE,² A. ROST,³ T. KÖHLER,³ and W.A. SCHERBAUM⁴

ABSTRACT

Matrix metalloproteinase-1 (MMP-1) and tissue matrix metalloproteinase inhibitor 1 (TIMP-1) play an important role in remodeling the extracellular matrix in normal and pathological processes. The effect of phorbol-myristate acetate (PMA), interleukin-1 (IL-1), and tumor necrosis factor- α (TNF- α) on MMP-1 and TIMP-1 expression was studied on highly purified thyrocytes and undifferentiated 8505 C, C 643, HTh 74, SW 1736 thyroid carcinoma cells compared with thyroid-derived fibroblasts. Messenger RNA (mRNA) levels were monitored by competitive semiquantitative reverse transcriptase polymerase chain reaction (RT-PCR) after 24 hours. Culture supernatants were assayed for free and/or complexed MMP-1 and TIMP-1 after 48 hours using enzyme-linked immunosorbent assay (ELISA) systems (detection limit: <2 ng/mL). MMP-1 and TIMP-1 mRNA were present in all cell types, although thyrocytes showed MMP-1 mRNA levels near the detection limit. 8505 C expressed MMP-1 mRNA levels of up to 10^6 times those of the other cells analyzed. PMA and IL-1 increased MMP-1 mRNA in most cell types. TIMP-1 mRNA increased after treatment with PMA in all cells except 8505 C, whereas only slight effects were shown after IL-1 stimulation. MMP-1 protein was undetectable in normal thyrocyte cultures, but was secreted spontaneously by all cell lines ([ng/mL]: C 643: 15 ± 7 ; HTh 74: 81 ± 1 ; SW 1736: 13 ± 2 ; 8505C: 2097 ± 320). There was a strong correlation between levels of MMP-1 mRNA and protein ($r = 0.99$, $p < .0001$). PMA and IL-1 increased MMP-1 secretion in all cell types after 48 hours. Fibroblasts ([ng/mL] 517 ± 55) and the cell lines (C 643: 142 ± 48 ; HTh 74: 115 ± 13 ; SW 1736: 202 ± 14 ; 8505 C: 120 ± 19) secreted TIMP-1 in unstimulated cultures, whereas only a trace amount was detected in thyrocyte cultures, even after PMA treatment. IL-1 upregulated TIMP-1 secretion after 48 hours in SW 1736, HTh 74, and C 643 cells. Our data suggest that in contrast to normal thyrocytes, dedifferentiated thyroid carcinoma cell lines are potential producers of MMP-1 as well as TIMP-1. High MMP-1 or MMP-1/TIMP-1 expression may play a role in tissue invasion of undifferentiated thyroid cancer cells.

INTRODUCTION

MATRIX METALLOPROTEINASES, (MMPs) constitute a family of structurally related proteolytic enzymes responsible for the proteolytic degradation of extracellular matrix (ECM) components. They are important participants in normal tissue remodeling and contribute to the phenotype of several pathological conditions that are associated with progressive ECM degradation. MMPs are highly regulated at different levels (1). At the transcriptional level, MMP expression can be directly induced or

suppressed on external stimulation, ie, with cytokines, phorbol 12-myristate 13-acetate (PMA), lipopolysaccharide (LPS), or retinoic acid (2,3). After secretion at post-transcriptional level, latent MMP proenzymes are regulated by proteolytic activation and interaction with tissue inhibitors of matrix metalloproteinase (TIMPs), their specific inhibitors. Any imbalance between the proteolytic MMPs activities and the TIMPs that could be influenced and caused by cytokines could potentially lead to pathological conditions (4).

MMP-1, although known as an interstitial collagenase,

¹Institut of Anatomy, ²Department of Internal Medicine III, and ³Institute of Clinical Chemistry and Pathobiochemistry, University of Leipzig, Germany. ⁴Department of Endocrinology, University of Duesseldorf.

is the only enzyme active at neutral pH that can degrade extracellular fibers comprised of collagen types I, II, and III. With this initial step, MMP-1 provides the cleavage products to other collagenase types (5). The major specific inhibitor of MMP-1 is TIMP-1, a 28.5-kd glycoprotein, which forms 1:1 stoichiometric complexes with the protease (6). Cytokines and growth factors have been shown to regulate the expression of both MMP-1 and TIMP-1 (1,7,8).

Although the participation of MMP-1 as the initial collagenase in tissue breakdown during tumor development is well documented (9–11), only one study has described the expression (12) but no study has as yet investigated the regulation of this enzyme in different thyroid tumors. Few studies have been published investigating the role of other MMPs in normal and pathological thyroid tissue by *in situ* hybridization and immunohistochemistry (13–16). Furthermore, tissue remodeling includes both the action of MMPs and their inhibitors; thus, these enzymes could be involved in autoimmune and other nonautoimmune thyroid diseases during morphological changes (17,18). It is still unknown whether or not thyrocytes are able to express MMPs and TIMPs. Although type IV collagenases (MMP-2 and MMP-9) were detected in various human epithelial cells of different tissue origin (19,20), only one study described the secretion of MMP-1 by epithelial cells (21).

Highly purified normal thyrocytes and four thyroid carcinoma cell lines were included in this study to investigate the involvement of these cells in MMP-1 and TIMP-1 production during thyroid tissue remodeling processes and in malignant thyroid neoplasms. MMP-1 and TIMP-1 expression were studied at both the mRNA and protein level by semiquantitative RT-PCR and ELISA measurement, respectively.

In unstimulated carcinoma cell lines both MMP-1 and TIMP-1 mRNA were expressed, partly at a high level, followed by the spontaneous secretion of the proteins. The various conditions for the stimulation of the different cell lines by cytokines and PMA were defined. In contrast to the cell lines, normal thyrocytes did not secrete MMP-1 and only trace amount of TIMP-1, even after stimulation with PMA.

MATERIALS AND METHODS

Preparation of tissues, thyroid-derived cells, and cell lines

Thyrocytes were prepared from surgical thyroid specimens from 3 patients (1 Graves' disease, 2 nontoxic goiter; mean age 54.3 ± 5.0 years). Fibroblasts were separated from thyroid tissue of 5 other patients (3 Graves' disease, 2 nontoxic goiter; mean age 43.6 ± 6.4 years). Graves' disease and nontoxic goiter were diagnosed on the strength of clinical, biochemical, and immunologic features as well as thyroid scintiscans.

Thyroid tissue was trimmed of fat and connective tissue immediately after surgery. Thyroid-derived cells were enriched after gradual enzymatic digestion of tissue and cultured over a period of 16 hours as described. Thyrocytes were obtained from the adherent fraction by incubating

the cell monolayer with phosphate buffered saline (PBS) without $\text{Ca}^{2+}/\text{Mg}^{2+}$ for 45 minutes (22). Residual fibroblasts were removed after subsequent incubation of the cells with the fibroblast-specific mab FibAS01 (22) and goat-anti-mouse IgG-DYNABEADS® M450 (DYNAL, Hamburg, Germany) according to the manufacturer's protocol.

Thyroid-derived fibroblasts were obtained after culturing small pieces of thyroid tissue in Dulbeccos's Modified Eagle's Medium (DMEM) with 10% fetal calf serum (FCS) and harvested in the 5th to 7th passage. The purity of the thyrocytes and fibroblasts was determined by using indirect immunofluorescence technique on a FACS-Scan (Becton Dickinson GmbH, Heidelberg, Germany) as described (22).

The following human anaplastic thyroid carcinoma cell lines were cultured in DMEM with 10% FCS: C 643 (23); SW 1736 (23); and HTh 74 (24). The cell line 8505 C (25) was purchased from the German Collection of Microorganisms and Animal Cell Cultures (DSMZ ACC219). This cell line was established from a primary thyroid tumor characterized histologically as a undifferentiated carcinoma that was partially composed of poorly differentiated papillary cells (25). This is a feature of a subgroup of anaplastic carcinoma (26). The majority of these coexistent better differentiated carcinoma foci in anaplastic carcinoma were papillary (26).

In vitro cultures

Using 24-well plates, 1×10^5 cells were cultured for 24 hours. The medium was aspirated and replaced with 500 μL OPTI-MEM (GIBCO BRL, Grand Island, NY) without FCS to eliminate possible stimulation of MMP-1 and TIMP-1 production by FCS. The medium contained the desired concentration of human IL-1 α (10 U/ml; Pepro Tech EC Ltd., London, UK), TNF- α (100 U/ml; Pepro Tech EC Ltd.), interferon- γ (IFN- γ) (500 U/ml; Pepro Tech EC Ltd.), or 10 ng/mL PMA (SIGMA).

Triplicate cultures of each stimulator were analyzed after 3, 6, and 24 hours at the mRNA and after 24 and 48 hours at the protein level. The supernatants were removed and stored at -80°C for further use. First, a collagenolytic assay based on the digestion of type I collagen was performed. This method showed direct evidence of free pro-MMP-1 enzyme in the cell culture supernatants of unstimulated and IL-1 α stimulated 8505 C, HTh 74, and C634 cells (data not shown). However, the method does not allow quantitation of MMP-1 enzyme activity. Thus, the cell culture supernatants were assayed for MMP-1, TIMP-1, and MMP1/TIMP-1 complex by ELISA (Amersham Life Sciences, Braunschweig, Germany). The MMP-1 assay (sensitivity: 1.7 ng/mL) detected only total human MMP-1, ie, free MMP-1 and MMP-1 complexed with inhibitors such as TIMP-1. It did not detect MMP-1 bound by the nonspecific protease inhibitor α_2 -macroglobulin. The MMP-1/TIMP-1 assay (sensitivity: 1.5 ng/mL) detected MMP-1/TIMP-1 complex, ie, activated MMP-1 that has been subsequently complexed with the specific MMP-1 inhibitor TIMP-1. It did not detect free active MMP-1, free TIMP-1, or pro-MMP-1. There was no cross-reactivity with active MMP-1 bound by the nonspecific protease inhibitor α_2 -macroglobulin. The TIMP-1 assay (sensitivity:

1.25 ng/mL) detected total human TIMP-1, i.e., free TIMP-1 and that complexed with MMPs. The assay did not fully cross-react with TIMP-1 in complexes with other MMP. It did not cross-react with TIMP-2.

RNA extraction and cDNA synthesis

For gene expression studies, 5 mL RNAzol™ B (Biotex Laboratories Inc., Houston, TX) was added to the cell culture wells. The content of three wells of any cell type was pooled and then stored frozen for further mRNA analysis in liquid nitrogen. Total cellular RNA (cDNA) was isolated from the probes according to the manufacturer's protocol. RNA was fractionated on a denaturing 1.0% agarose gel and stained with ethidium bromide to confirm that spectrophotometric measurements were accurate and that the RNA had not been degraded. Five micrograms total RNA was reverse-transcribed to cDNA using the First-strand cDNA synthesis kit of Pharmacia (Uppsala, Sweden) in a total reaction volume of 15 μ L.

mRNA analysis by competitive RT-PCR

To correct for variations across different cDNA preparations, all samples were first adjusted to contain equal input glyceraldehyde-3-phosphate dehydrogenase (GAPDH) cDNA concentrations. Semi-quantitative GAPDH RT-PCR was used with a heterologous synthetic competitor fragment. The generation of the specific PCR products from the competitor and the cDNA with the GAPDH primers were published earlier (22,27).

We then estimated the MMP-1 and TIMP-1 cDNA in these adjusted samples. The primers were selected using the DNAsis computer program (Hitachi Software Engineering Co, Yokohama, Japan). The primer pairs span one or more introns to allow unambiguous discrimination between cDNA and unwanted contaminating genomic DNA. In quantitating MMP-1 and TIMP-1 cDNA, a rapid one-step method was introduced to synthesize an internal homologous competitor (plan diagram of procedure: Fig. 1, exemplary for MMP-1 [28]). A hybrid primer was synthesized (MMP-1hy) that consisted of two segments (seg₁, seg₂). It

had a length of 40 nucleotides, in which 20 nucleotides (seg₁) at the 3' end corresponded to the opposite strand of the target sequence a predetermined distance from primer MMP-1f, and 20 nucleotides at the 5' end (seg₂ = MMP-1r) that corresponded to the target sequence upstream from the segment seg₁. Amplification with the primers MMP-1f and MMP-1hy from the cDNA resulted in a 478-base pair (bp) (polymerase chain reaction (PCR) product. It was freed from excess primers and deoxynucleoside-triphosphates (dNTPs) using the Qiaquick Gel Extraction Kit (Qiagen GmbH, Hilden, Germany) and quantified. A known number of copies of the competitor was introduced in the GAPDH-adjusted samples and amplified with the primers MMP-1f and MMP-1r. With this approach, two products were generated, one derived from the cDNA (560 bp) and another, 82 bp smaller in size derived from the internal competitor (Fig. 1). PCR products were resolved by gel electrophoresis (1.5% agarose gel). The relative amounts of sample cDNA and competitor were quantified by measuring the intensity of ethidium fluorescence with a CCD image sensor and analyzing the data with the EASY program (Herolab, Wiesloch, Germany). The initial amounts of sample cDNA and competitor were assumed to be equal in those reactions where the ratio of the two products was judged to be equal. This was expressed in arbitrary units (AU) (22,29). One AU was defined as the lowest concentration of competitor yielding a detectable amplification product when added to PCR alone. For example, if equivalence between sample cDNA and competitor was reached using a 100-fold concentrated competitor the relative sample cDNA concentration was 100 AU. Thyrocytes and the cell lines were analyzed for the expression of thyroid-specific and cytokine receptor mRNAs in a simple RT-PCR. The sequences of the TPO and cytokine receptor primer pairs have been published by Watson et al. (30) and Tada et al. (31) and gave the following product sizes: TPO: 506 bp; IL-1R type I (p80): 300 bp; IL-1R type II (p68): 392 bp; TNF- α R (p75): 324 bp; TNF- α R (p55): 587 bp and IFN- γ R: 899 bp. The thyroglobulin (Tg) and thyroid stimulating hormone receptor (TSH-R) primer pairs were selected according to the published sequences using the DNAsis program (Table 1).

Each 25- μ L amplification reaction contained 2.5 μ L 10 \times concentrated PCR buffer (15 mM MgCl₂, Boehringer Mannheim, Germany), 0.3 U Taq DNA polymerase (Boehringer Mannheim, Germany), 100 μ M dNTPs (Perkin Elmer, Weiterstadt, Germany), 0.1 μ M of each primer (IMB, Jena, Germany), and 1 μ L cDNA and competitor in adjusted dilution. Furthermore, restriction mapping (restriction enzymes: Boehringer Mannheim GmbH, Germany) was carried out to confirm the originality of the PCR product (Fig. 1, Table 1).

Statistics

Protein levels of thyrocyte or fibroblast cultures from the different patients and of the thyroid carcinoma cell lines obtained from three separate experiments were presented as mean \pm SEM values. Statistical comparisons between unstimulated and stimulated cell cultures were performed by the alternate (Welch) t-test. The correlation between basal mRNA levels and the unstimulated protein secretion in all cell types was calculated according to the Spearman method.

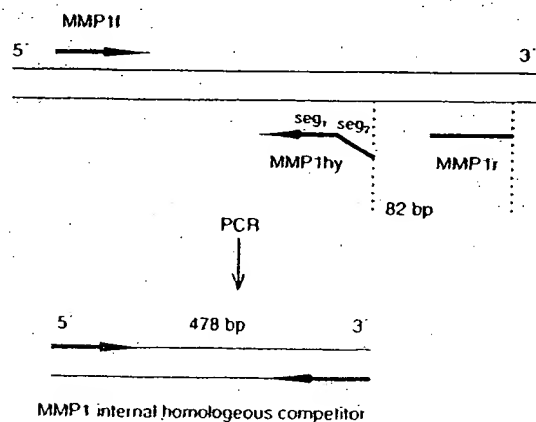


FIG. 1. General scheme for generating homologous competitors used for quantitative PCR.

TABLE 1. PRIMERS, LENGTH OF AMPLIFIED TEMPLATES, RESTRICTION MAPPING AND ASSAY CONDITIONS FOR RT-PCR

| | | Primer | Length of cDNA (bp) | Length of competitor (bp) | Annealing temperature | Number of cycles |
|---------------------|---------|---|------------------------|------------------------------|--------------------------|---------------------|
| Tg | forward | GCAGATCTTACTGAGTGGCT | 416 | | 60 | 35 |
| | reverse | TGTCAGCACAGTGGCAATAC | | | | |
| TSH-R exons 1-4 | forward | ACTTGCTGCAGCTGGTGCT | 354 | | 65 | 35 |
| | reverse | TGAGGGCATCAGGGTCTATG | | | | |
| TSH-R exons 4-10 | forward | GAAATTCGGAATACCAGGAACCTTA ACT | 896 | | 53 | 35 |
| | reverse | AATCATCGGACTTGGGGGTACA | | | | |
| MMP-1 | forward | TGGGAGCAAAACACATCTGAC | 560 | 478 | 64 | 33 |
| | reverse | ATCACTTCTCCCGAATCGT | | | | |
| | hybrid | ATCACTTCTCCCGAATCGT CCATATATGGGTTGGATGCC CTTAGGGGATGCCGCTGACA | | | | |
| TIMP-1 | forward | GGCAGGCAGCAAGGTGACG | 351 | 274 | 64 | 30 |
| | reverse | GGCAGGCAGGCAAGGTGACG | | | | |
| | hybrid | GGATGGATAAACAGGGAAAC | | | | |

Tg indicates thyroglobulin; TSH-R, thyroid stimulating hormone receptor; MMP-1, matrix metalloproteinase-1; TIMP-1, tissue inhibitor of metalloproteinase-1; bp, base pair.

RESULTS

Thyroid specific and cytokine receptor mRNA expression

Isolated thyrocytes as well as 8505 C cells expressed Tg and thyroperoxidase (TPO) mRNA, whereas transcripts of the TSH-R (exons 1-4, 354 bp, exons 4-10, 896 bp) were present only in the thyrocytes. The three anaplastic thyroid carcinoma cell lines SW 1736, C 634, and HTh 74 were completely negative for the Tg, TPO, and TSH-R mRNAs (Fig. 2). All cell lines and thyrocytes expressed IL-1R (type I and type II), TNF- α (p75 and p55) and IFN- γ R mRNA (Fig. 2).

Basal MMP-1 and TIMP-1 mRNA and protein expression

In most stimulation experiments, mRNA levels did not increase until 24 hours of incubation. The 24-hour mRNA levels are shown in Figures 3 and 4. The 3- and 6-hour levels are demonstrated in those experiments where the mRNA levels reached their peak before 24 hours of stimulation. If not otherwise indicated, the MMP-1 levels were measured using the ELISA system, which recognizes free/complexed MMP-1.

MMP-1 and TIMP-1 mRNA were found during unstimulated culture in all investigated cell types, although the mRNA levels varied over a great range. 8505 C showed a basal MMP-1 mRNA level 20 times as high as those of the HTh 74 cells, 6×10^4 times as high as C 643, and 2×10^6 times as high as SW 1736 cells. In thyrocytes, MMP-1 mRNA levels were found near the detection limit (Figs. 3 and 4).

Generally, when analyzing the noted cell types, the measured basal MMP-1 or TIMP-1 mRNA levels correlated well with the basal protein expression (MMP-1: $r = 0.99$, $p < .0001$; TIMP-1: $r = 0.98$, $p < .002$). Corresponding to the high MMP-1 mRNA level, 8505 C cells secreted extremely high levels of MMP-1. No MMP-1 or TIMP-1 was

detected in unstimulated thyrocyte cultures at any time-point examined. All other cell types showed a spontaneous MMP-1 and TIMP-1 secretion (Figs. 5 and 6). Thyroid-derived fibroblasts produced basal TIMP-1 levels of up to 4 times higher in the four carcinoma cell lines, which secreted nearly the same amounts of basal TIMP-1 protein. Nevertheless, TIMP-1 secretion of fibroblasts was found at lower levels than expected after TIMP-1 mRNA measurement in 4 of 5 analyzed patients. The results of the fibroblast cultures from patient five showing a higher TIMP-1 expression than those from the 4 other patients (basal 24 hour: 50 ± 2 ; PMA 24 hour: 90 ± 6 ng/mL TIMP-1) was omitted in Figure 6.

Comparing the basal amount of free/complexed and TIMP-1 complexed MMP-1 after 24 hours of stimulation, a significant level of MMP-1 was not complexed with TIMP-1 in 8505 C cultures, whereas in fibroblast cultures most of the MMP-1 activity was inhibited by TIMP-1. The anaplastic carcinoma cell line HTh 74 did not show such a great discrepancy between free/complexed and TIMP-1 complexed MMP-1 level as 8505 C cells (Fig. 7).

Effects of IL-1 α on MMP-1 and TIMP-1 mRNA and protein expression

Experiments were performed to determine whether human thyroid epithelial cells and thyroid carcinoma cell lines could produce or increase basal MMP-1 and TIMP-1 secretion after exposure to various stimuli. The results from these stimulation experiments are summarized in Figures 3 through 5. Generally, there was a delay in protein secretion level in comparison to the mRNA expression level. At the protein level, the cytokine-mediated stimulating or inhibiting effect is more distinct after 48 hours compared with 24 hours, even when the mRNA level had already decreased after 6 hours.

IL-1 upregulated MMP-1 mRNA in SW 1736-cells up to 100 times and, in thyroid-derived fibroblasts, up to 12 times after 24 hours of incubation (Fig. 3). This increased mRNA level was accompanied by a significantly enhanced

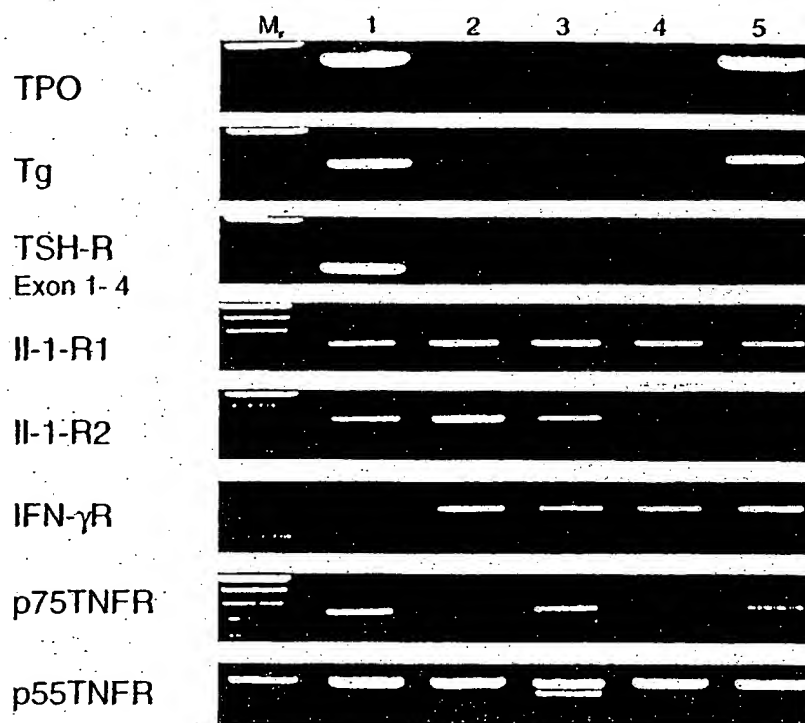


FIG. 2. Amplification of thyroid specific and interleukin-receptor mRNA in thyrocytes (1), SW 1736 (2), C 643 (3), HTh 74 (4) and 8505 C (5) cells using RT-PCR; M_r = 100-bp ladder (GIBCO).

MMP-1 secretion after 48 hours. Furthermore, IL-1 α increased MMP-1 mRNA expression in thyrocytes up to seven times after 6 hours, but no MMP-1 protein could be detected in thyrocyte cultures. IL-1 had no stimulatory effect on MMP-1 mRNA expression in C 643, HTh 74, and 8505 C cells after 24 hours, although a significant increase of MMP-1 secretion was found in HTh-74 and SW 1736 cells after 48 hours of incubation (Fig. 5). This discrepancy may be explained by a possible increase in MMP-1 mRNA level after 24 hours of stimulation. The same effect could also be observed in the IL-1 stimulated TIMP-1 at the mRNA as well as the protein level: the only slight effect of IL-1 on TIMP-1 mRNA expression in carcinoma cell lines after 24 hours was accompanied by a significant increase of TIMP-1 secretion in 8505 C and HTh 74 cells after 48 hours (Figs. 4 and 6).

Effects of TNF- α on both MMP-1/TIMP-1 mRNA and protein expression

In contrast to IL-1, TNF- α did not stimulate the MMP-1 and TIMP-1 mRNA and protein levels in all carcinoma cell lines and thyrocytes. Only thyroid-derived fibroblasts responded with a slight upregulation of MMP-1 and TIMP-1 mRNA expression after TNF- α stimulation, which was not accompanied by an increase of MMP-1 and TIMP-1 secretion.

Effects of PMA, and IFN- γ on MMP-1 and TIMP-1 mRNA and protein expression

PMA was included in our study as a positive control because it is known to upregulate or induce both MMP-1 and TIMP-1 secretion in various cell types (1,32). Indeed, PMA was able to induce or enhance MMP-1 mRNA levels in all cell types investigated, although the detected levels varied to a large extent (Fig. 3). This result is in good correlation with the significantly increased MMP-1 protein levels that were already detectable after 24 hours of stimulation (Fig. 5). PMA upregulated TIMP-1 mRNA levels by up to 20 times in C 643, and up to 2 times in SW 1736 and HTh 74 cells, fibroblasts and thyrocytes, but it did not change the TIMP-1 mRNA content in 8505 cells (Fig. 4). At the protein level, we found a significant stimulation of TIMP-1 secretion in C 643 and HTh 74 cells, as well as in thyroid-derived fibroblasts (Fig. 6).

In contrast to PMA, IFN- γ was without effect on stimulation or downregulation of MMP-1 and TIMP-1 mRNA or protein in any of the cell types investigated (Figs 5 and 6).

The main inhibitor of MMP-1 is TIMP-1, which forms 1:1 stoichiometric complexes with MMP-1, although some other inhibitors can also bind MMP-1. On the other hand, TIMP-1 can bind other MMP types.

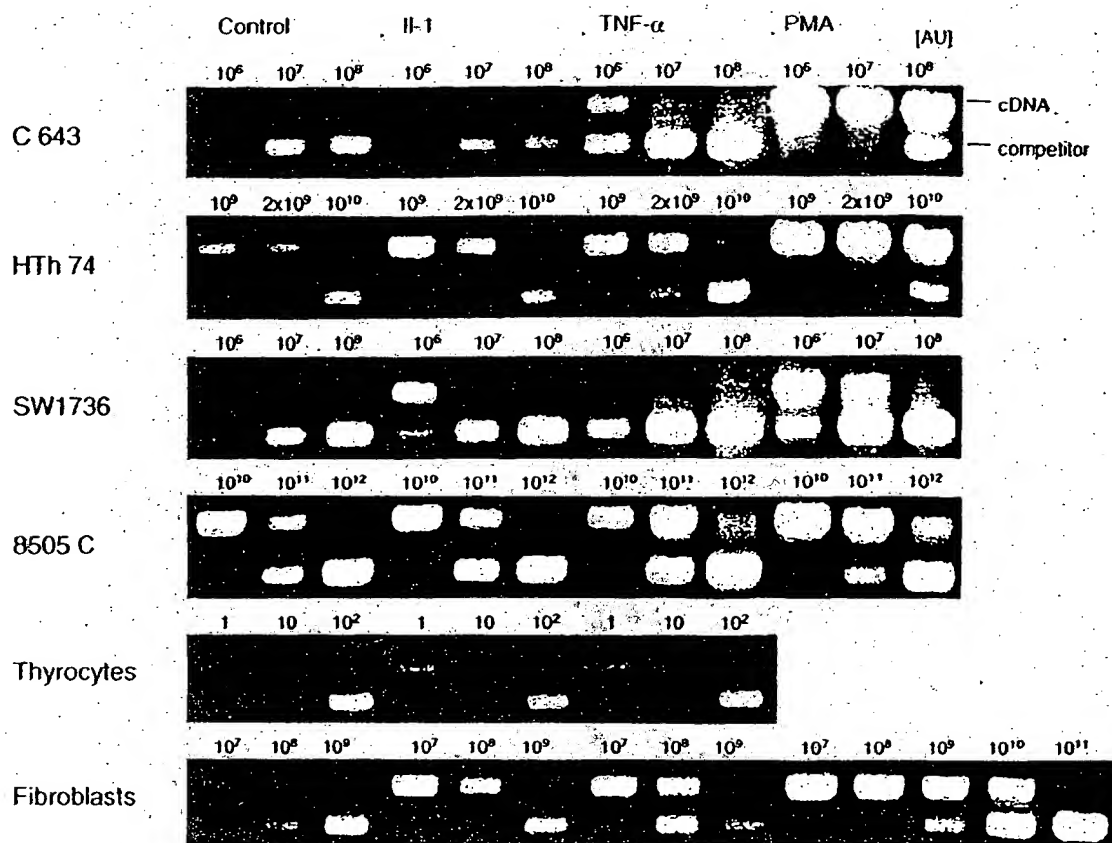


FIG. 3. Representative samples of competitive amplified MMP-1 mRNA of thyrocytes, thyroid-derived fibroblasts and thyroid carcinoma cell lines without stimulation (control) and after stimulation with 10 U/mL IL-1 α and 100 U/mL TNF- α and 10 ng/mL PMA after 24 hours. Serial dilutions of known amounts of the competitor fragment were coamplified with identical aliquots of cDNA. The 560-bp (cDNA) and 478-bp (competitor) PCR products were visualized by agarose gel electrophoresis and ethidiumbromide staining. The relative concentration of the added competitor was given in arbitrary units (AU) in the figure. One AU was defined as the lowest concentration of the competitor yielding a detectable amplification for MMP-1 mRNA. The ratio of competitor to cDNA fragments was determined by measuring the intensity of ethidium fluorescence with a CCD image sensor and analysis of data. Measured cDNA concentration can be expressed in AU.

DISCUSSION

Our findings demonstrate for the first time that thyroid carcinoma cell lines are able to express MMP-1 and TIMP-1 mRNA and protein at significant levels *in vitro*. The observation of spontaneous release of MMP-1 and TIMP-1 corresponds well with earlier studies covering the secretion of these proteins by several carcinoma cell lines (33,34).

However, in contrast to its clear physiological function in extracellular matrix breakdown, the role of MMP-1 in tumor growth and metastases is still controversial (9-11,35). Recently, Murray et al. (10) demonstrated that MMP-1 is associated with poor prognosis in colorectal cancer, and has a prognostic value independent of the Dukes stage. Therefore, MMP-1 could be a target for therapeutic intervention in such tumors. Furthermore, the hypothesis of whether or not cancer cells themselves are able to

produce MMP, or whether cancer cells stimulate the surrounding stromal cells to secrete MMP *in vivo*, is disputed. MMP-1 mRNA and protein were detected by both *in situ* hybridization and immunohistochemistry in stromal as well as tumor cells of head, neck, gastric, colorectal, and mammary carcinomas (9,10,36,37). In contrast, Kameyama (12) demonstrated by *in situ* hybridization that the MMP-1 mRNA was not expressed in the cancer cells but in the surrounding fibrous capsules of strongly differentiated papillary thyroid carcinoma tissue. Highly differentiated follicular carcinomas and follicular adenomas were depleted for MMP-1 transcripts. Undifferentiated follicular, papillary, and aggressive anaplastic carcinomas that showed poor prognosis and strong tumor invasive and metastatic potential and that can be compared in their morphological, genetic and growth features with undifferentiated thyroid carcinoma cell lines were not included this

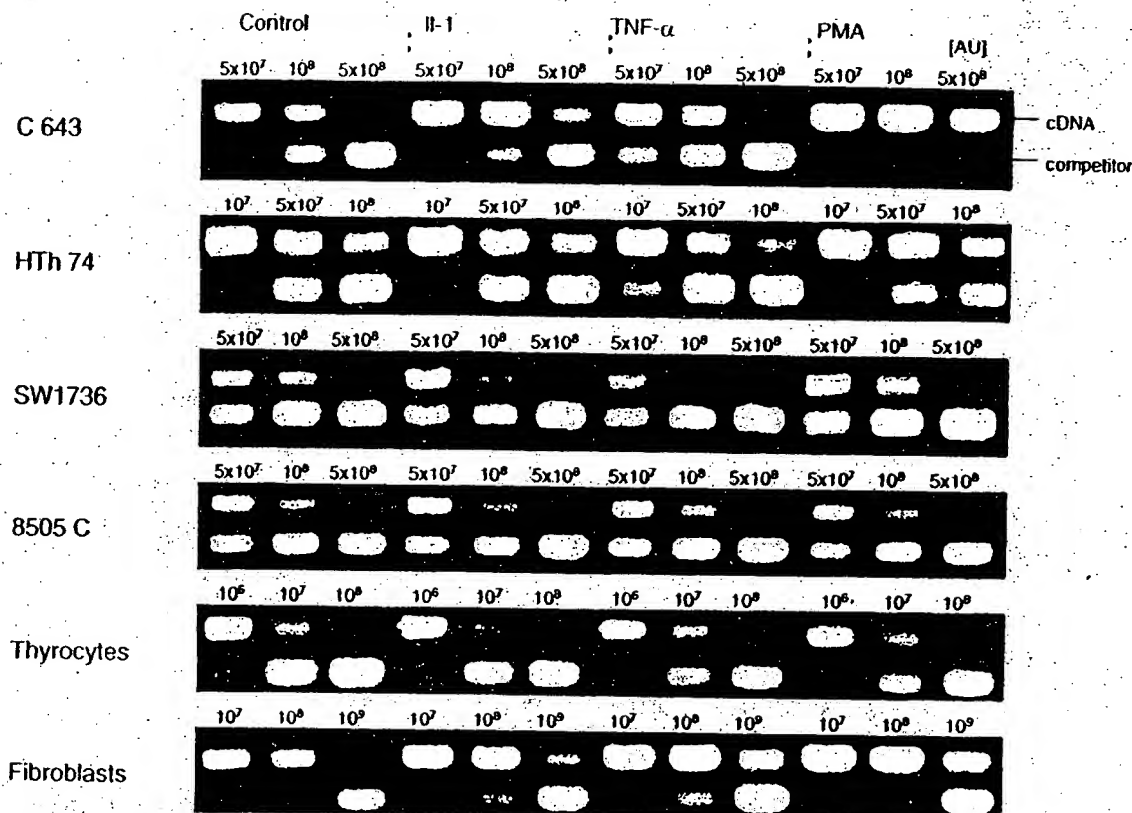


FIG. 4. Competitive TIMP-1 mRNA RT-PCR yielding a 351-bp (cDNA) and a 274-bp (competitor) PCR product. For further details see Figure 3.

study. However, the missing expression of MMP-1 by normal thyrocytes and the spontaneous secretion of this protein by highly malignant thyroid carcinoma cell lines, as demonstrated in our study, indicate the involvement of MMP-1 secretion of transformed thyrocytes in aggressive thyroid tumors.

Although all cell lines analyzed in our study spontaneously secreted MMP-1, we observed marked differences in the basal secretion capacity. The highest MMP-1 levels were determined in cultures of 8505 C cells. Only 8505 C cells expressed TPO and Tg mRNA that may be put down to residual differentiated components in the cell line (see *Materials*). However, none of the analyzed cell lines expressed TSH-R mRNA. The cell population doubling times were less than 40 hours. All cell lines had accumulations of multiple genetic events. These facts indicate the undifferentiated pathology of the studied lines. It is well known that anaplastic carcinoma cell lines well retain the malignant characteristics of their parental tumors (38–40).

Furthermore, we found a distorted proportion between MMP-1 and TIMP-1 mRNA/protein for carcinoma cell lines but not for normal thyroid-derived fibroblasts. The most disadvantageous constellation between MMP-1 and TIMP-1 was found in 8505 C cells. Similar to other studies (41), these results suggest the influence of an altered MMP/TIMP relation on tumor progression. However, it

should be mentioned that most studies, including the present one, do not take into consideration that a number of inhibitors distinct from TIMP-1 may regulate MMP-1 activity. Taking into account that the balance of active enzyme and TIMP-1 concentration strongly influence the extent of local matrix degradation, a number of studies showed unexpectedly high levels of TIMP-1 in malignant neoplasms (9,42,43). There is a great discussion as to whether the overall expression of MMP-1 and TIMP-1 or the amount of noncomplexed MMP-1 could be critical in aggressive tumor development. This fact underlines the nature of tissue breakdown, reflecting the complicated network of selective and coordinated production of individual proteinases and inhibitors under normal and pathophysiological conditions. Thus, the invasive and metastatic potential of thyroid tumors depends on the local net level of active MMPs.

The synthesis of MMP-1 and TIMP-1 is influenced by a variety of biochemical stimuli. The recent findings on MMP-1 and TIMP-1 gene promoters are useful in understanding the complex mechanisms implied in the regulation of MMP synthesis modulated by cytokines and tumor promoters (34,44,45). The promoter regions contain tumor promoter responsive elements (TRE) and binding motifs for the transcription factor PEA-3, which are recognized by proto-oncogenic transcription factors, such as the

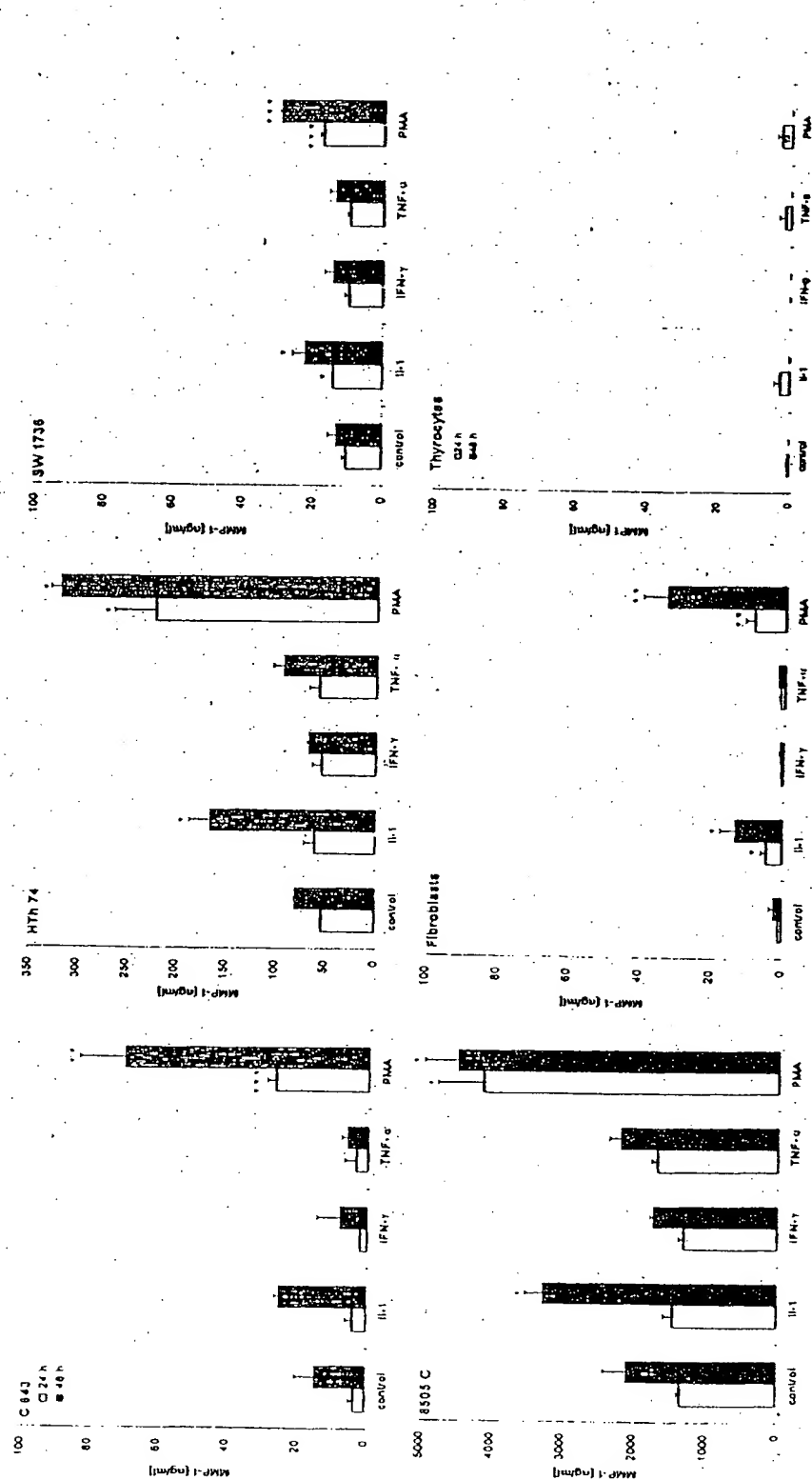


FIG. 5. MMP-1 protein levels (mean \pm SEM) in supernatants of unstimulated and stimulated cultures of thyrocytes ($n = 3$), thyroid-derived fibroblasts ($n = 4$) and the thyroid carcinoma cell lines 8505 C, SW 1736, C 643, HTH 74 ($n = 3$) detected by MMP-1 ELISA, which recognizes total MMP-1, ie, free MMP-1 and that complexed with inhibitors such as TIMP-1, but not α_2 -macroglobulin. Cells were stimulated with 10 U/mL IL-1 α , 100 U/mL TNF- α , 500 U/mL IFN- γ , or 10 ng/ml PMA for 24 and 48 hours. For fibroblasts, each point represents the mean \pm SEM of four different donors, each experiment performed in triplicate. For cell lines, each point represents the mean \pm SEM of three separate experiments each performed in triplicate. Significant differences between the basal and stimulated MMP-1 levels are indicated by asterisks (* $p < .05$; ** $p < .01$; *** $p < .005$). Please note the differences in scale between 8505 C, HTH 74, and the other cell types.

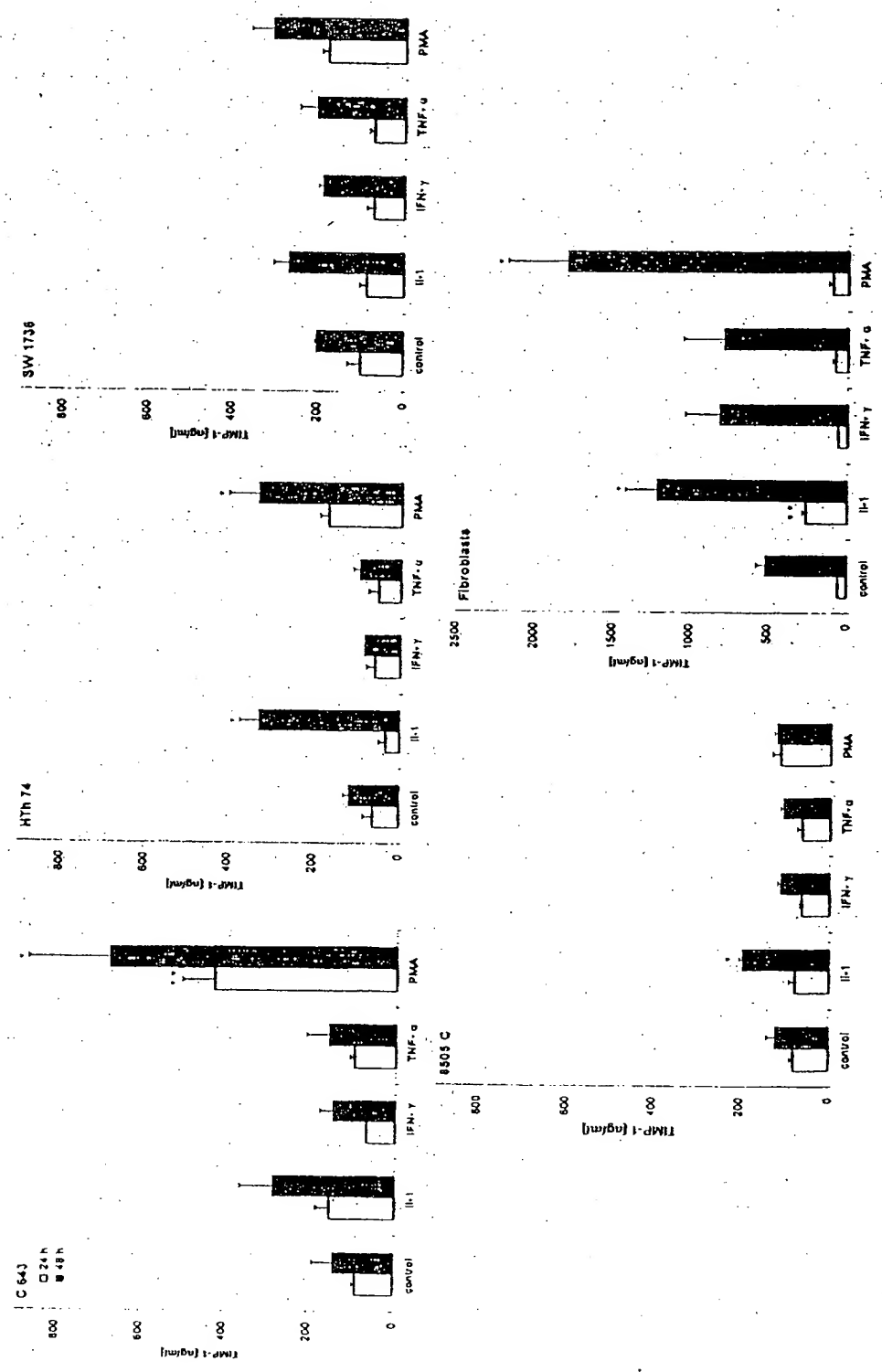


FIG. 6. ELISA detection of free and MMP-complexed TIMP-1 (mean \pm SEM). Asterisks indicate significant differences between the basal and stimulated TIMP-1 levels (* $p < .05$; ** $p < .01$; *** $p < .005$). For further details see Figure 5. Please note the differences in scale between fibroblasts and the other cell types.

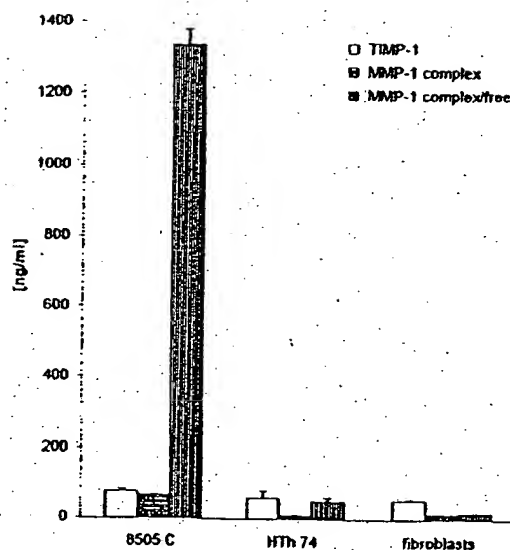


FIG. 7. Comparison between (i) free/complex and (ii) TIMP-1 complexed MMP-1 levels, and (iii) TIMP-1 levels in supernatants of unstimulated 8505 C and HTh 74 cells, and thyroid-derived fibroblasts after 24 hours using a (i) MMP-1 ELISA that recognizes total MMP-1 (see Figure 5). The (ii) MMP-1/TIMP-1 assay recognizes MMP-1/TIMP-1 complexes, ie, activated MMP-1 that has subsequently been complexed with the specific MMP inhibitor TIMP-1. The (iii) TIMP-1 ELISA recognizes total TIMP-1, ie, free TIMP-1 and that complexed with MMPs.

fos and *jun* family (45–47). IL-1, TNF- α , and PMA up-regulate proto-oncogenes like *fos* and *jun*, resulting in the stimulation of MMP-1 and TIMP-1 (45,48). The action of the cytokines is mediated by their specific receptors. In our study, IL-1R (type I and type II), TNF- α R (p75 and p55) and IFN- γ R mRNAs were demonstrated in all investigated cell types. PMA and IL-1 were shown to elevate MMP-1 and TIMP-1 in nearly all cell types investigated, thus confirming the results of several studies on other epithelial cells (reviewed in refs. 1,7,49). In the majority of experiments, we found a concordant expression of MMP-1 and TIMP-1 after stimulation, possibly achieved by the coordinated actions of the nuclear transcription factors, although MMP-1 and TIMP-1 expression can also be independently or even reciprocally regulated (1). The effect of TNF- α was not as distinct as in the case of PMA and IL-1, although several investigators found a pronounced effect of TNF- α particularly on TIMP-1 secretion (4,34). In contrast to studies performed with other cell types (863,864,819), IFN- γ did not influence MMP-1 and TIMP-1 expression in thyroid carcinoma cell lines. In summary, the involvement of the intrathyroidal physiological and pathological cytokine microenvironment in the regulation of MMP-1 and TIMP-1 induction activation and inhibition is strongly suggested.

Furthermore, the data demonstrate that regular human thyrocytes did not produce MMP-1, even after powerful stimulation with PMA. Investigating other mammalian epithelial cells, only one study revealed the production of

MMP-1 by rabbit corneal cells (21). It is yet not clear whether the MMP-1 mRNA detected in thyrocytes is due to a low level of constitutive transcription of the MMP-1 gene (illegitimate transcription), an existing pool of stable MMP-1 mRNA, or *in vitro* induction of MMP-1 mRNA. But it seems more likely that residual fibroblasts contained in the purified thyrocyte preparation (<0.2%) are responsible for the slightly positive RT-PCR results. Another explanation could be that thyrocytes are indeed MMP-1 producers, but the ELISA detection system used was not sensitive enough to measure extremely low MMP-1 secretion levels. Furthermore, the discrepancy between elevated TIMP-1 mRNA levels of thyrocytes and the extremely low TIMP-1 protein secretion by these cells is difficult to explain. Post-transcriptional regulatory events may be responsible for this confounding result.

Taken together, the present study suggests that the intrathyroidal cytokine microenvironment is involved in the regulation of MMP-1 and its inhibitor TIMP-1 in the thyroid, and that both proteins may be secreted by dedifferentiated thyroid carcinoma cells and involved in aggressive thyroid tumors *in vivo*.

REFERENCES

1. Ries C, Petrides PE 1995 Cytokine regulation of matrix metalloproteinase activity and its regulatory dysfunction in disease. *Biol Chem Hoppe Seyler* 376:345–355.
2. Pierce RA, Sandefur S, Doyle GA, Welgus HG 1996 Monocytic cell type-specific transcriptional induction of collagenase. *J Clin Invest* 97:1890–1899.
3. Pan L, Eckhoff C, Brinckerhoff CE 1995 Suppression of collagenase gene expression by all-trans and 9-cis retinoic acid is ligand dependent and requires both RARs and RXRs. *J Cell Biochem* 57:575–589.
4. Shingu M, Nagai Y, Isayama T, Naono T, Nobunaga M 1993 The effects of cytokines on metalloproteinase inhibitors (TIMP) and collagenase production by human chondrocytes and timp production by synovial cells and endothelial cells. *Clin Exp Immunol* 94:145–149.
5. Goldberg GL, Wilhelm SM, Kronberger A, Bauer EA, Grant GA, Eisen AZ 1986 Human fibroblast collagenase. Complete primary structure and homology to an oncogene transformation-induced rat protein. *J Biol Chem* 261:6600–6605.
6. Docherty AJ, Lyons A, Smith BJ, Wright EM, Stephens PE, Harris TJ, Murphy G, Reynolds JJ 1985 Sequence of human tissue inhibitor of metalloproteinases and its identity to erythroid-potentiating activity. *Nature* 318:66–69.
7. Mauviel A 1993 Cytokine regulation of metalloproteinase gene expression. *J Cell Biochem* 53:288–295.
8. Murphy G 1995 Matrix metalloproteinases and their inhibitors. *Acta Orthop Scand Suppl* 266:55–60.
9. Nomura H, Fujimoto N, Seiki M, Mai M, Okada Y 1996 Enhanced production of matrix metalloproteinases and activation of matrix metalloproteinase 2 (gelatinase A) in human gastric carcinomas. *Int J Cancer* 69:9–16.
10. Murray GI, Duncan ME, O'Neil P, Melvin WT, Fothergill JE 1996 Matrix metalloproteinase-1 is associated with poor prognosis in colorectal cancer. *Nat Med* 2:461–462.
11. Onisto M, Garbisa S, Caenazzo C, Freda MP, Di Francesco C, Nitti D, Lionta LA, Stetler-Stevenson WG 1993 Reverse transcription-polymerase chain reaction phenotyping of met-

- alloproteinases and inhibitors involved in tumor matrix invasion. *Diagn Mol Pathol* 2:74-80.
12. Kameyama K 1996 Expression of MMP-1 in the capsule of thyroid cancer—Relationship with invasiveness. *Pathol Res Pract* 192:20-26.
 13. Campo E, Merino MJ, Liotta L, Neumann R, Stetler-Stevenson W 1992 Distribution of the 72-kd type IV collagenase in nonneoplastic and neoplastic thyroid tissue. *Hum Pathol* 23:1395-1401.
 14. Demeure MJ, Damsky CH, Elfman F, Goretzki PE, Wong MG, Clark O 1992 Invasion by cultured human follicular thyroid cancer correlates with increased beta 1 integrins and production of proteases. *World J Surg* 16:770-776.
 15. Nakano T, Kusunoki T, Funasaka K, Murata K, Nishida S, Tomura T 1995 Study of type I and IV collagenase activity in human thyroid diseases. *Nippon Jibiinkoka Gakkai Kaiho* 98:937-941.
 16. Zedenius J, Stahle-Backdahl M, Enberg U, Grimelius L, Larsson C, Wallin G, Backdahl M 1996 Stromal fibroblasts adjacent to invasive thyroid tumors: expression of gelatinase A but not stromelysin 3 mRNA. *World J Surg* 20:101-106.
 17. Arai M, Niiooka M, Maruyama K, Wada N, Fujimoto N, Nomiyama T, Tanaka S, Okazaki I 1996 Changes in serum levels of metalloproteinases and their inhibitors by treatment of chronic hepatitis C with interferon. *Dig Dis Sci* 41:995-1000.
 18. Nikkari ST, O'Brien KD, Ferguson M, Hatsukami T, Welgus HG, Alpers CE, Clowes AW 1995 Interstitial collagenase (MMP-1) expression in human carotid atherosclerosis. *Circulation* 92:1393-1398.
 19. Knowlden J, Martin J, Davies M, Williams JD 1995 Metalloproteinase generation by human glomerular epithelial cells. *Kidney Int* 47:1682-1689.
 20. Buisson AC, Zahm JM, Polette M, Pierrot D, Bellon G, Puchelle E, Birembaut P, Tournier JM 1996 Gelatinase B is involved in the in vitro wound repair of human respiratory epithelium. *J Cell Physiol* 166:413-426.
 21. Tao Y, Bazan HE, Bazan NG 1995 Platelet-activating factor induces the expression of metalloproteinases-1 and -9, but not -2 or -3, in the corneal epithelium. *Invest Ophthalmol Vis Sci* 36:345-354.
 22. Aust G, Heuer M, Laue S, Lehmann I, Hofmann A, Heldin N-E, Scherbaum WA 1996 Expression of TNA-alpha mRNA and protein in pathological thyroid tissue and carcinoma cell lines. *Clin Exp Immunol* 105:148-154.
 23. Mark J, Ekedahl C, Dahlenfors R, Westermarck B 1987 Cytogenetical observations in five human anaplastic thyroid carcinomas. *Hereditas* 107:163-174.
 24. Heldin NE, Cvejic D, Smeds S, Westermarck B 1991 Coexpression of functionally active receptors for thyrotropin and platelet-derived growth factor in human thyroid carcinoma cells. *Endocrinology* 129:2187-2193.
 25. Ito T, Seyama T, Hayashi Y, Hayashi T, Dohi K, Mizuno T, Iwamoto KS, Tsuyama N, Nakamura N, Akiyama M 1994 Establishment of two human thyroid carcinoma cell lines (8305C, 8505C) bearing p53 gene mutations. *International Journal of Oncology* 4:583-586.
 26. Wallin G, Backdahl M, Tallroth-Ekman E, Lundell G, Auer G, Lowhagen T 1989 Co-existent anaplastic and well differentiated thyroid carcinomas: a nuclear DNA study. *Eur J Surg Oncol* 15:43-48.
 27. Heuer M, Aust G, Ode-Hakim S, Scherbaum WA 1996 Different cytokine mRNA profiles in Graves' disease, Hashimoto's thyroiditis and non-autoimmune thyroid disorders determined by quantitative reverse transcriptase chain reaction (RT-PCR). *Thyroid* 6:97-106.
 28. Celi FS, Zenilman ME, Shuldiner AR 1993 A rapid and versatile method to synthesize internal standards for competitive PCR. *Nucleic Acids Res* 21:1047.
 29. Platzer C, Ode-Hakim S, Reinke P, Docke WD, Ewert R, Volk HD 1994 Quantitative PCR analysis of cytokine transcription patterns in peripheral mononuclear cells after anti-CD3 rejection therapy using two novel multispecific competitor fragments. *Transplantation* 58:264-268.
 30. Watson PF, Pickerill AP, Davies R, Weetman AP 1994 Analysis of cytokine gene expression in Graves' disease and multinodular goiter. *J Clin Endocrinol Metab* 79:355-360.
 31. Tada M, Diserens AC, Desbaillets J, de Tribolet N 1994 Analysis of cytokine receptor messenger RNA expression in human glioblastoma cells and normal astrocytes by reverse-transcription polymerase chain reaction. *J Neurosurg* 80:1063-1073.
 32. Nakano A, Tani E, Miyazaki K, Yamamoto Y, Furuyama J 1995 Matrix metalloproteinases and tissue inhibitors of metalloproteinases in human gliomas. *J Neurosurg* 83:298-307.
 33. Whitelock JM, O'Grady RL, Gibbins JR 1991 Interstitial collagenase (matrix metalloproteinase 1) associated with the plasma membrane of both neoplastic and nonneoplastic cells. *Invasion Metastasis* 11:139-148.
 34. Mackay AR, Ballin M, Pelina MD, Farina AR, Nason AM, Hartzler JL, Thorgerisson UP 1992 Effect of phorbol ester and cytokines on matrix metalloproteinase and tissue inhibitor of metalloproteinase expression in tumor and normal cell lines. *Invasion Metastasis* 12:168-184.
 35. Urbanski SJ, Edwards DR, Mailand A, Leco KJ, Watson A, Kossakowska AE 1992 Expression of metalloproteinases and their inhibitors in primary pulmonary carcinomas. *Br J Cancer* 66:1188-1194.
 36. Polette M, Clavel C, Muller D, Abecassis J, Binniger I, Birembaut P 1991 Detection of mRNAs encoding collagenase 1 and stromelysin 2 in carcinomas of the head and neck by in situ hybridization. *Invasion Metastasis* 11:76-83.
 37. Clavel C, Polette M, Doco M, Binniger I, Birembaut P 1992 Immunolocalization of matrix metalloproteinases and their tissue inhibitor in human mammary pathology. *Bull Cancer (Paris)* 79:261-270.
 38. Boghaert ER, Aijn K, Taylor K, Greenberg VL, Fowler C, Zimmer SG 1996 Quantitative and qualitative differences in growth, invasion and lung colonization of an anaplastic and a papillary human thyroid cancer cell line in vitro and in vivo. *Clin Exp Metastasis* 14:440-450.
 39. Asakawa H, Kobayashi T, Komoike Y, Yamagawa T, Takahashi M, Wakasugi E, Maruyama H, Tamaki Y, Matsuzawa Y, Monden M 1996 Establishment of anaplastic thyroid carcinoma cell lines useful for analysis of chemosensitivity and carcinogenesis. *J Clin Endocrinol Metab* 81:3547-3552.
 40. Viglietto G, Maglione D, Rambaldi M, Cerutti J, Romano A, Trapasso F, Fedele M, Ippolito P, Chiappetta G, Borti G, et al 1995 Upregulation of vascular endothelial growth factor (VEGF) and downregulation of placenta growth factor (PlGF) associated with malignancy in human thyroid tumors and cell lines. *Oncogene* 11:1569-1579.
 41. Nuovo GJ, MacConnell PB, Simsir A, Valea F, French DL 1995 Correlation of the in situ detection of polymerase chain reaction-amplified metalloproteinase complementary DNAs and their inhibitors with prognosis in cervical carcinoma. *Cancer Res* 55:267-275.
 42. Naruo S, Kanayama H, Takigawa H, Kagawa S, Yamashita K, Hayakawa T 1994 Serum levels of a tissue inhibitor of

- metalloproteinases-1 (TIMP-1) in bladder cancer patients. *Int J Urol* 1:228-231.
43. Kossakowska AE, Urbanski SJ, Watson A, Hayden LJ, Edwards DR 1993 Patterns of expression of metalloproteinases and their inhibitors in human malignant lymphomas. *Oncol Res* 5:19-28.
 44. Edwards DR, Rocheleau H, Sharma RR, Wills AJ, Cowie A, Hassell JA, Heath JK 1992 Involvement of AP1 and PEA3 binding sites in the regulation of murine tissue inhibitor of metalloproteinases-1 (TIMP-1) transcription. *Biochim Biophys Acta* 1171:41-55.
 45. Schonthal A, Herrlich P, Rahmsdorf HJ, Ponta H 1988 Requirement for fos gene expression in the transcriptional activation of collagenase by other oncogenes and phorbol esters. *Cell* 54:325-334.
 46. Gutman A, Wasylyk B 1990 The collagenase gene promoter contains a TPA and oncogene responsive unit encompassing the PEA3 and AP-1 binding sites. *EMBO J* 9:2241-2246.
 47. Wasylyk C, Gutman A, Nicholson R, Wasylyk B 1991 The c-Ets oncoprotein activates the stromelysin promoter through the same elements as several non-nuclear oncoproteins. *EMBO J* 10:1127-1134.
 48. Brenner DA, O'Hara M, Angel P, Chojkier M, Karin M 1989 Prolonged activation of jun and collagenase genes by tumour necrosis factor-alpha. *Nature* 337:661-663.
 49. Opdenakker G, Van Damme J 1992 Cytokines and proteases in invasive processes: molecular similarities between inflammation and cancer. *Cytokine* 4:251-258.

Address reprint requests to:
Dr. Gabriela Aust
Institute of Anatomy
University of Leipzig
Liebigstr. 13
Leipzig, D-04103, Germany



Expression of embryonic fibronectin isoform EIIIA parallels alpha-smooth muscle actin in maturing and diseased kidney.

Barnes VL, Musa J, Mitchell RJ, Barnes JL.

Department of Medicine, Division of Nephrology, University of Texas Health Science Center, San Antonio, Texas, USA.

In this study we examined if an association exists between expression of an alternatively spliced "embryonic" fibronectin isoform EIIIA (Fn-EIIIA) and alpha-smooth muscle actin (alpha-SMA) in the maturing and adult rat kidney and in two unrelated models of glomerular disease, passive accelerated anti-glomerular basement membrane (GBM) nephritis and Habu venom (HV)-induced proliferative glomerulonephritis, using immunohistochemistry and in situ hybridization. Fn-EIIIA and alpha-SMA proteins were abundantly expressed in mesangium and in periglomerular and peritubular interstitium of 20-day embryonic and 7-day (D-7) postnatal kidneys in regions of tubule and glomerular development. Staining was markedly reduced in these structures in maturing juvenile (D-14) kidney and was largely lost in adult kidney. Expression of Fn-EIIIA and alpha-SMA was reinitiated in the mesangium and the periglomerular and peritubular interstitium in both models and was also observed in glomerular crescents in anti-GBM nephritis. Increased expression of Fn-EIIIA mRNA by in situ hybridization corresponded to the localization of protein staining. Dual labeling experiments verified co-localization of Fn-EIIIA and alpha-SMA, showing a strong correlation of staining between location and staining intensity during kidney development, maturation, and disease. Expression of EIIIA mRNA corresponded to protein expression in developing and diseased kidneys and was lost in adult kidney. These studies show a recapitulation of the co-expression of Fn-EIIIA and alpha-SMA in anti-GBM disease and suggest a functional link for these two proteins.

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BMI-1 gene amplification and overexpression in hematological malignancies occur mainly in mantle cell lymphomas.

Bea S, Tort F, Pinyol M, Puig X, Hernandez L, Hernandez S, Fernandez PL, van Lohuizen M, Colomer D, Campo E.

The Hematopathology Section, Laboratory of Anatomic Pathology, Hospital Clinic, Institut d'Investigacions Biomediques August Pi i Sunyer (IDIBAPS), University of Barcelona, Spain.

The BMI-1 gene is a putative oncogene belonging to the Polycomb group family that cooperates with c-myc in the generation of mouse lymphomas and seems to participate in cell cycle regulation and senescence by acting as a transcriptional repressor of the INK4a/ARF locus. The BMI-1 gene has been located on chromosome 10p13, a region involved in chromosomal translocations in infant leukemias, and amplified in occasional non-Hodgkin's lymphomas (NHLs) and solid tumors. To determine the possible alterations of this gene in human malignancies, we have examined 160 lymphoproliferative disorders, 13 myeloid leukemias, and 89 carcinomas by Southern blot analysis and detected BMI-1 gene amplification (3- to 7-fold) in 4 of 36 (11%) mantle cell lymphomas (MCLs) with no alterations in the INK4a/ARF locus. BMI-1 and p16INK4a mRNA and protein expression were also studied by real-time quantitative reverse transcription-PCR and Western blot, respectively, in a subset of NHLs. BMI-1 expression was significantly higher in chronic lymphocytic leukemia and MCL than in follicular lymphoma and large B cell lymphoma. The four tumors with gene amplification showed significantly higher mRNA levels than other MCLs and NHLs with the BMI-1 gene in germline configuration. Five additional MCLs also showed very high mRNA levels without gene amplification. A good correlation between BMI-1 mRNA levels and protein expression was observed in all types of lymphomas. No relationship was detected between BMI-1 and p16INK4a mRNA levels. These findings suggest that BMI-1 gene alterations in human neoplasms are uncommon, but they may contribute to the pathogenesis in a subset of malignant lymphomas, particularly of mantle cell type.

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BMI-1 Gene Amplification and Overexpression in Hematological Malignancies Occur Mainly in Mantle Cell Lymphomas¹

Silvia Beà, Frederic Tort, Magda Pinyol, Xavier Puig, Luis Hernández, Silvia Hernández, Pedro L. Fernández, Maarten van Lohuizen, Dolors Colomer,² and Elias Campo²

The Hematopathology Section, Laboratory of Anatomic Pathology, Hospital Clinic, Institut d'Investigacions Biomèdiques "August Pi i Sunyer" (IDIBAPS), University of Barcelona, 08036 Barcelona, Spain [S. B., F. T., M. P., X. P., L. H., S. H., P. L. F., D. C., E. C.], and Division of Molecular Carcinogenesis, The Netherlands Cancer Institute, 1066 CX Amsterdam, Netherlands [M. v. L.]

Abstract

The *BMI-1* gene is a putative oncogene belonging to the Polycomb group family that cooperates with *c-myc* in the generation of mouse lymphomas and seems to participate in cell cycle regulation and senescence by acting as a transcriptional repressor of the *INK4a/ARF* locus. The *BMI-1* gene has been located on chromosome 10p13, a region involved in chromosomal translocations in infant leukemias, and amplified in occasional non-Hodgkin's lymphomas (NHLs) and solid tumors. To determine the possible alterations of this gene in human malignancies, we have examined 160 lymphoproliferative disorders, 13 myeloid leukemias, and 89 carcinomas by Southern blot analysis and detected *BMI-1* gene amplification (3- to 7-fold) in 4 of 36 (11%) mantle cell lymphomas (MCLs) with no alterations in the *INK4a/ARF* locus. *BMI-1* and *p16^{INK4a}* mRNA and protein expression were also studied by real-time quantitative reverse transcription-PCR and Western blot, respectively, in a subset of NHLs. *BMI-1* expression was significantly higher in chronic lymphocytic leukemia and MCL than in follicular lymphoma and large B cell lymphoma. The four tumors with gene amplification showed significantly higher mRNA levels than other MCLs and NHLs with the *BMI-1* gene in germline configuration. Five additional MCLs also showed very high mRNA levels without gene amplification. A good correlation between *BMI-1* mRNA levels and protein expression was observed in all types of lymphomas. No relationship was detected between *BMI-1* and *p16^{INK4a}* mRNA levels. These findings suggest that *BMI-1* gene alterations in human neoplasms are uncommon, but they may contribute to the pathogenesis in a subset of malignant lymphomas, particularly of mantle cell type.

Introduction

The *BMI-1*³ gene is a putative oncogene of the Polycomb group originally identified by retroviral insertional mutagenesis in Eμ-*c-myc* transgenic mice infected with the Moloney murine leukemia virus (1, 2). These animals had a rapid development of pre-B cell lymphomas showing frequent proviral insertions near the *BMI-1* gene. This integration resulted in *BMI-1* overexpression suggesting a cooperative effect between *C-MYC* and *BMI-1* genes in the development of these tumors (3, 4). Recent studies have indicated that the *BMI-1* gene may also participate in cell cycle control and senescence through the

INK4a/ARF locus by acting as an upstream negative regulator of *p16^{INK4a}* and *p14/p19^{ARF}* gene expression (5). The human *BMI-1* gene has been mapped to chromosome 10p13 (6), a region involved in chromosomal translocations in infant leukemias (7) and rearrangements in malignant T cell lymphomas (8, 9). More recently, high-level DNA amplifications of this region have been found by comparative genomic hybridization in NHLs and solid tumors (10, 11). However, the possible implication of the *BMI-1* gene in these alterations and its role in the pathogenesis of human tumors is not known. The aim of this study was to analyze the possible *BMI-1* gene alterations and expression in a large series of human neoplasms and to determine the relationship with *INK4a/ARF* locus aberrations.

Materials and Methods

Case Selection. A series of 262 human tumors, including 173 hematological malignancies and 89 carcinomas (Table 1), matched normal tissues from all carcinomas, 11 samples of normal peripheral mononuclear cells, and 5 reactive lymph nodes and tonsils, were selected based on the availability of frozen samples for molecular analysis.

DNA Extraction and Southern Blot Analysis. Genomic DNA was obtained using Proteinase K/RNase treatment. 15 μg were digested with *EcoRI* and *HindIII* restriction enzymes (Life Technologies, Inc., Gaithersburg, MD), for Southern blot analysis and hybridized with a 1.5-kb *PstI* fragment of the partial *BMI-1* cDNA (6).

RNA Extraction and Real-time Quantitative RT-PCR. Total RNA was obtained from 67 lymphoid neoplasms (10 CLLs, 27 MCLs, 8 FLs, and 22 LCLs) using guanidine/isothiocyanate extraction and cesium/chloride gradient centrifugation. One μg of total RNA was transcribed into cDNA using MMLV-reverse transcriptase (Life Technologies, Inc.) and random hexamers, following manufacturer's directions. Sequences of the *BMI-1* and the *p16* detection probes and primers were designed using the Primer Express program (Applied Biosystems, Foster City) as follows: *BMI-1* sense, 5'-CTGGTTCG-CCATTGACAGC-3'; *BMI-1* antisense, 5'-CAGAAAATGAATGCGAG-CCA-3'; *p16* sense, 5'-CAACGCACCGAATAGTTACGG-3'; *p16* antisense, 5'-AACTTCGTCTCCAGAGTCGC-3'. The probes *BMI-1*, 5'-CAGTCC-GCTTCAAGATGGCCGC-3', and *p16*, 5'-CGGAGGCCGATCCAGGTGG-GTA-3', were labeled with 6-carboxy-fluorescein as the reporter dye. The TaqMan-GAPDH Control Reagents (Applied Biosystems) were used to amplify and detect the *GAPDH* gene, as recommended by the manufacturer. The quantitative assay amplified 1 μl of cDNA in two to four replicates using the primers and probes described above and the standard master mix (Applied Biosystems). All reactions were performed in an ABI PRISM 7700 Sequence Detector System (Applied Biosystems). *GAPDH*, *BMI-1*, and *p16^{INK4a}* expression was related to a standard curve derived from serial dilutions of Raji cDNA. The RUs of *BMI-1* and *p16^{INK4a}* expression were defined as the mRNA levels of these genes normalized to the *GAPDH* expression level in each case.

Protein Analysis. Whole-cell protein extracts were obtained from additional frozen tissue available in 31 cases (7 CLLs, 12 MCLs, 8 FLs, and 4 LCLs), loaded onto a 10% SDS-polyacrylamide gel, and electrophoreted to a nitrocellulose membrane (Amersham). Blocked membranes were incubated sequentially with the monoclonal antibody BMI-F6 (12), antimouse conju-

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²To whom requests for reprints should be addressed, at the Department of Pathology, Hospital Clinic, University of Barcelona, Villarroel 170, 08036-Barcelona, Spain. Phone: 34 93 227 5450; Fax: 34 93 227 5572; E-mail: campo@medicina.ub.es.

³The abbreviations used are: *BMI-1*, B cell-specific Moloney murine leukemia virus integration site 1; NHL, non-Hodgkin's lymphoma; CLL, chronic lymphocytic leukemia; FL, follicular lymphoma; LCL, large B cell lymphoma; MCL, mantle cell lymphoma; RT-PCR, reverse transcription-PCR; RU, relative units.

Table 1 Hematological malignancies and solid tumor samples analyzed for BMI-1 gene alterations

| Tissue samples | No. of cases |
|--------------------------------------|--------------|
| Hematological malignancies | |
| Hodgkin's disease | 2 |
| B cell lymphoproliferative disorders | |
| B-Acute lymphoblastic leukemia | 14 |
| CLL | 29 |
| Hairy cell leukemia | 4 |
| FL | 15 |
| MCL | 36 |
| LCL | 40 |
| T cell lymphoproliferative disorders | |
| T-Acute lymphoblastic leukemia | 8 |
| Large granular cell leukemia | 4 |
| Peripheral T-cell lymphoma | 8 |
| Myeloproliferative disorders | |
| Acute myeloid leukemia | 7 |
| Chronic myeloid leukemia | 6 |
| Solid tumors | |
| Colon carcinoma | 26 |
| Breast carcinoma | 29 |
| Laryngeal squamous cell carcinoma | 34 |
| Total | 262 |

gated to horseradish peroxidase (Amersham), and detected by enhanced chemiluminescence (Amersham) according to the manufacturer's recommendations.

Statistical Analysis. Because of the non-normal distribution of the samples and the small size of some subsets of tumors, the statistical evaluation was performed using nonparametric tests (SPSS, version 9.0). Comparison between mRNA expression levels in the different groups of NHLs was performed using the Kruskal-Wallis Test, with a *P* for significance set at 0.05. For differences between particular groups, the conservative Bonferroni procedure was performed, and the *P* was set at 0.005. The remaining statistical analyses were carried out using the Mann-Whitney nonparametric *U* test (significance, *P* < 0.05). The comparison between BMI-1 and p16^{INK4a} quantitative mRNA levels was also performed using the Pearson's correlation coefficient.

Results

BMI-1 Gene Amplification. The BMI-1 gene was examined by Southern blot in a large series of human tumors and normal samples (Table 1). The cDNA probe used in the study detected three *Eco*RI fragments of 7.3, 3.8, and 2.6 kb and three *Hind*III fragments of 6.2, 4, and 3.5 kb. BMI-1 gene amplification (3- to 7-fold) was detected in 4 of 36 (11%) MCLs (Fig. 1). The amplifications were confirmed with both restriction enzymes. The amplified MCLs were two blastoid and two typical variants. No amplifications were observed in any of the solid tumors when compared with their respective matched non-neoplastic mucosa. No BMI-1 gene rearrangements were observed in any of the samples examined.

BMI-1 mRNA Expression. To determine the BMI-1 expression pattern in NHL we analyzed BMI-1 mRNA levels by real-time quantitative RT-PCR in 67 lymphomas (10 CLLs, 27 MCLs, 8 FLs, and 22 LCLs), including the four tumors with gene amplification. A distinct BMI-1 mRNA expression pattern was observed in the different types of lymphomas (Fig. 2; Kruskal-Wallis Test; *P* < 0.001). The BMI-1 mRNA levels in CLLs (mean, 2.2 RU; SD, 1.3) and MCLs with no BMI-1 gene amplification (mean, 2.5 RU; SD, 2.3) were significantly higher than in FLs (mean, 0.9 RU; SD, 0.8) and LCLs (mean, 0.6 RU; SD, 0.4; Mann-Whitney nonparametric *U* test; *P* < 0.01). The 4 MCLs with BMI-1 gene amplification showed significantly higher levels of expression than all other groups of tumors (mean, 5.1 RU; SD, 1.6; *P* < 0.005). In addition, five typical MCLs with no structural alterations of the gene also showed very high levels of BMI-1 mRNA expression ranging from 4 to 9.8 RU, similar to cases with gene amplification (Fig. 2A).

BMI-1 Protein Expression. BMI-1 protein expression was examined by Western blot in 31 tumors (7 CLLs; 12 MCLs, including two

cases with BMI-1 gene amplification and 4 cases with mRNA overexpression and no structural alteration of the gene; 8 FLs, and 4 LCLs) in which additional frozen tissue was available. The monoclonal antibody against BMI-1 detected three closely migrating proteins of *M*_r 45,000–48,000 (2). The two more slowly migrating bands probably represent phosphorylated isoforms of the protein (12). The two MCLs with gene amplification and three of four cases with mRNA overexpression without amplification of the gene showed very high levels of protein expression. The remaining MCLs and CLLs showed intermediate levels of protein expression, whereas low- or no-expression signals were detected in the LCLs and FLs included in the study (Fig. 3). These results indicate that BMI-1 protein expression in NHL is concordant with the mRNA levels observed by real-time quantitative RT-PCR.

Relationship between BMI-1 and p16^{INK4a} Gene Alterations. The *INK4a/ARF* locus has been recently identified as a downstream target of the transcriptional repressing activity of the BMI-1 gene, suggesting that this gene may contribute to human neoplasias with wild type *INK4a/ARF* (5). Most of the lymphoproliferative disorders analyzed in the present study, including the four cases with BMI-1 gene amplification, had been previously examined for p53 gene mutations and *INK4a/ARF* locus alterations, including gene deletions, mutations, hypermethylation, and expression (13, 14). The four MCLs with BMI-1 gene amplification and mRNA overexpression and the five tumors with BMI-1 mRNA overexpression with no structural alterations of the gene showed a wild-type configuration of the *INK4a/ARF* locus (13). However, one case with BMI-1 gene amplification and one case with mRNA overexpression with no alteration of the gene showed p53 gene mutations associated with allelic deletions.

To determine the possible relationship between BMI-1 and p16^{INK4a} mRNA expression, p16^{INK4a} mRNA levels were evaluated by real-time quantitative RT-PCR in 50 tumors (10 CLLs, 27 MCLs, and 13 LCLs), including 6 cases with alterations in the *INK4a/ARF* locus (2 MCLs and 1 LCL with p16^{INK4a} gene deletion, 2 LCLs with p16 promoter hypermethylation, and 1 CLL with p16^{INK4a} gene mutation), and the 4 lymphomas with BMI-1 amplification. Negative or negligible levels of p16^{INK4a} were observed in the 6 tumors with *INK4a/ARF* locus alterations. These cases were not included in the comparisons between BMI-1 and p16^{INK4a} mRNA expression. The p16^{INK4a} expression levels were relatively similar in the different types of tumors. Only LCLs tended to have lower levels of expression, but the differences did not reach statistical significance (Fig. 2B). No differences were observed in the p16^{INK4a} mRNA levels between tumors with BMI-1 gene amplification and overexpression and lymphomas with germline configuration of the gene.

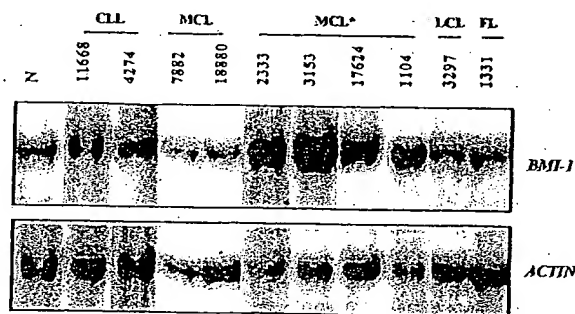


Fig. 1. Southern blot analysis of BMI-1 gene. Four MCLs (MCL*) showed BMI-1 gene amplification (3- to 7-fold) compared with non-neoplastic tissues (N) and other NHLs. No amplifications or gene rearrangements were detected in the remaining NHLs and carcinomas included in the study.

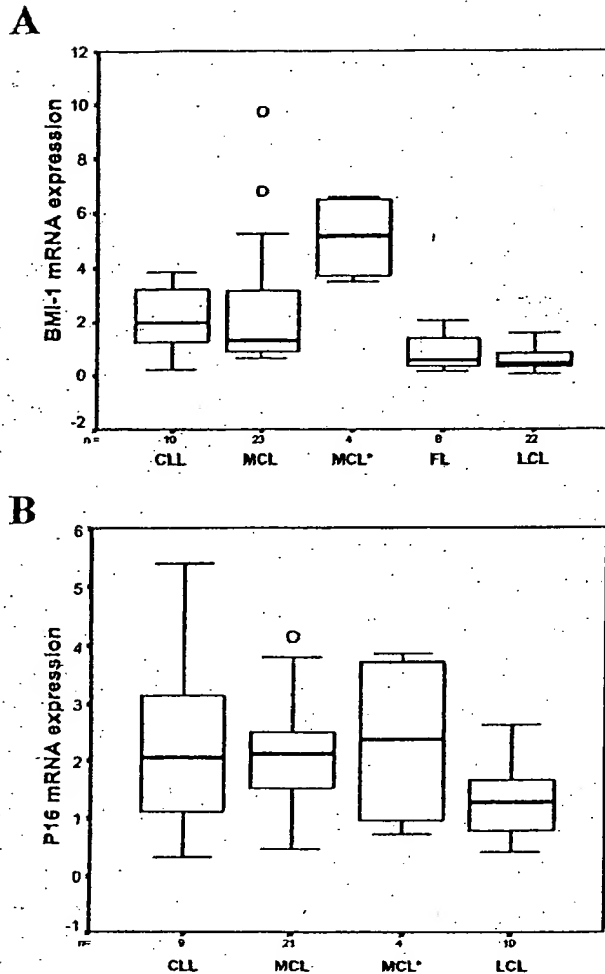


Fig. 2. A, quantitative BMI-1 mRNA transcript analysis (median and range) using real-time RT-PCR in a series of NHLs. MCLs with *BMI-1* gene amplification (*MCL**) revealed significantly higher overall BMI-1 mRNA levels than all other types of NHLs, including MCLs with no structural alterations of the gene ($P < 0.005$). MCLs and CLLs expressed significantly higher levels than FLs and LCLs ($P < 0.001$). Results are depicted as the ratio of absolute BMI-1:GADPH mRNA transcript numbers (RU). Bars, SD. B, quantitative p16^{INK4a} mRNA transcript analysis (median and range) using real-time RT-PCR in a series of NHLs. Expression levels were relatively similar in the different types of tumors. Results are depicted as the ratio of absolute p16^{INK4a}:GADPH mRNA transcript numbers (RU). Bars, SD.

Discussion

In the present study, we have examined a large series of human tumors for the presence of gene alterations and mRNA expression of the *BMI-1* gene. Gene amplification was identified in four MCLs. These tumors showed significantly higher levels of mRNA and protein expression compared with other lymphomas with *BMI-1* in germline configuration. BMI-1 expression levels were also highly up-regulated in a subset of MCLs with no apparent structural alterations of the gene. No alterations were detected in any of the different types of carcinomas included in the study. *BMI-1* is considered an oncogene belonging to the Polycomb group family of genes. These proteins mainly act as transcriptional regulators, controlling specific target genes involved in development, cell differentiation, proliferation, and senescence. Different studies have shown the implication of BMI-1 overexpression in the development of lymphomas in murine and feline animal models (3, 4). The findings of the present study indicate

for the first time that *BMI-1* gene alterations in human neoplasms are an uncommon phenomenon, but they seem to occur mainly in a subset of NHLs, particularly of mantle cell type.

The human *BMI-1* gene has been mapped to chromosome 10p13. High-level DNA amplifications and gains in this region have been identified by comparative genomic hybridization in occasional solid tumors and NHLs (10, 11). Different chromosomal translocations involving the 10p13 region have also been identified in infant leukemias and T cell lymphoproliferative disorders (7, 8, 15). Most acute leukemias with this chromosomal alteration occur in children <12 months of age, whereas it seems to be extremely rare in adults. 10p translocations in T-cell lymphoproliferative disorders have been observed mainly in adult T cell leukemia/lymphomas and occasional cutaneous T cell lymphomas. In our study, we did not observe *BMI-1* rearrangements or amplifications in any of the acute leukemias or T cell lymphomas. However, all of the acute leukemias in this study were diagnosed in patients over 16 years, and no adult T cell leukemia/lymphomas or cutaneous lymphomas could be included in the series. Similarly, high-level DNA amplifications at the 10p13 region have been detected in head and neck carcinomas and other solid tumors. Although we found no evidence for *BMI-1* gene rearrangements or amplifications in a substantial set of carcinomas, this does not exclude the possibility of increased gene expression or protein levels in these tumors. Additional studies are required to elucidate the possible involvement of *BMI-1* in these particular groups of human neoplasms.

In human hematopoietic cells, BMI-1 is preferentially expressed in primitive CD34+ bone marrow cells, whereas it is negative or very low in more mature CD34- cells (16). In peripheral lymphocytes, and particularly in follicular B cells, BMI-1 protein expression has been detected in resting cells of the mantle zone, whereas it is down-regulated in proliferating germinal center cells (17, 18). These observations indicate that BMI-1 expression in normal hematopoietic cells is tightly regulated in relation with cell differentiation in bone marrow and antigen-specific response in peripheral lymphocytes. BMI-1 expression in human tumors has not been examined previously. In this study, we have demonstrated that BMI-1 mRNA and protein expression show a distinct pattern in different types of lymphomas. Thus, BMI-1 levels were low in LCLs and FLs and significantly higher in MCLs and CLLs. These findings suggest that BMI-1 expression patterns in B cell lymphomas maintain in part the expression profile of their normal cell counterparts; because FLs and at least a subgroup of LCLs are considered lymphomas derived from follicular germinal center cells, whereas MCLs and CLLs are tumors mainly derived from naive pregerminal center cells. However, the four MCLs with *BMI-1* gene amplification expressed significantly higher mRNA levels than all other tumors. In addition, five MCLs with no structural alterations of the gene showed high mRNA levels similar to those observed in tumors with *BMI-1* gene amplification, suggesting that other mechanisms may be involved in up-regulation of the gene in these lymphomas. Different studies using animal models have shown a dose-dependent effect of *BMI-1* gene expression on skeleton development

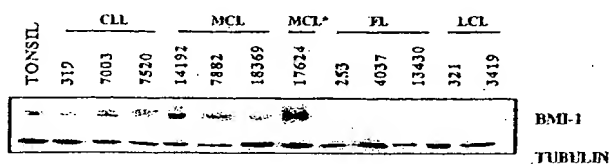


Fig. 3. Western blot analysis of BMI-1 protein in NHLs. The amplified MCL (17624) showed the highest BMI-1 protein levels, whereas other MCLs and CLLs had intermediate levels of expression. Very low or negative signal was observed in FLs and LCLs.

and lymphomagenesis (1, 3). These observations suggest that the high mRNA and protein levels detected in a subset of MCLs may play a role in the pathogenesis of these neoplasms.

Recent studies have identified the *INK4a/ARF* locus as a downstream target of the BMI-1 transcriptional repressor activity, suggesting that BMI-1 overexpression may contribute to human neoplasias that retain the wild-type *INK4a/ARF* locus (5). Interestingly, in our study, BMI-1 amplification and overexpression appeared in tumors with no alterations in *p16^{INK4a}* and *p14^{ARF}* genes. However, we could not detect differences in the expression levels of *p16^{INK4a}* in tumors with and without BMI-1 gene alterations. The reasons for this apparent discrepancy with experimental observations are not clear. One possibility may be that genes other than *INK4a/ARF* are the main targets of BMI-1 repressor activity in these tumors. Particularly, different genes of the HOX family are regulated by BMI-1 and may also be involved in lymphomagenesis (19, 20).

In conclusion, the findings of this study indicate that BMI-1 gene expression is differentially regulated in B cell lymphomas. Alterations of the gene seem to be an uncommon phenomenon in human neoplasms, but they may contribute to the pathogenesis in a subset of MCLs. Although, BMI-1 gene alterations occurred in tumors with wild-type *INK4a/ARF* locus, the possible cooperation between these genes and the oncogenic mechanisms of BMI-1 in human neoplasms require additional analysis.

Acknowledgments

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References

- Haupt, Y., Alexander, W. S., Barri, G., Klinken, S. P., and Adams, J. M. Novel zinc finger gene implicated as myc collaborator by retrovirally accelerated lymphomagenesis in E μ -myc transgenic mice. *Cell*, 65: 753-763, 1991.
- van Lohuizen, M., Verbeek, S., Scheijen, B., Wientjens, E., van der Gulden, H., and Berns, A. Identification of cooperating oncogenes in E μ -myc transgenic mice by provirus tagging. *Cell*, 65: 737-752, 1991.
- Alkema, M. J., Jacobs, H., van Lohuizen, M., and Berns, A. Perturbation of B and T cell development and predisposition to lymphomagenesis in E μ Bmi1 transgenic mice require the Bmi1 RING finger. *Oncogene*, 15: 899-910, 1997.
- Haupt, Y., Bath, M. L., Harris, A. W., and Adams, J. M. Bmi-1 transgene induces lymphomas and collaborates with myc in tumorigenesis. *Oncogene*, 8: 3161-3164, 1993.
- Jacobs, J. J., Kieboom, K., Marino, S., DePinho, R. A., and van Lohuizen, M. The oncogene and Polycomb-group gene *bmi-1* regulates cell proliferation and senescence through the *ink4a* locus. *Nature (Lond.)*, 397: 164-168, 1999.
- Alkema, M. J., Wiegant, J., Raap, A. K., Berns, A., and van Lohuizen, M. Characterization and chromosomal localization of the human proto-oncogene BMI-1. *Hum. Mol. Genet.*, 2: 1597-1603, 1993.
- Pui, C. H., Raimondi, S. C., Murphy, S. B., Ribeiro, R. C., Kalwinsky, D. K., Dahl, G. V., Crist, W. M., and Williams, D. L. An analysis of leukemic cell chromosomal features in infants. *Blood*, 69: 1289-1293, 1987.
- Berger, R., Baranger, L., Bernheim, A., Valensi, F., Flandrin, G., and Berheimin, A. T. Cytogenetics of T-cell malignant lymphoma. Report of 17 cases and review of the chromosomal breakpoints. *Cancer Genet. Cytogenet.*, 36: 123-130, 1988.
- D'Alessandro, E., Paterlini, P., Lo Re, M. L., Di Cola, M., Ligas, C., Quagliaro, D., and Del Porto, G. Cytogenetic follow-up in a case of Sezary syndrome. *Cancer Genet. Cytogenet.*, 45: 231-236, 1990.
- Bea, S., Ribas, M., Hernandez, J. M., Bosch, F., Pinyol, M., Hernandez, L., Garcia, J. L., Flores, T., Gonzalez, M., Lopez-Guillermo, A., Piris, M. A., Cardesa, A., Montserrat, E., Miro, R., and Campo, E. Increased number of chromosomal imbalances and high-level DNA amplifications in mantle cell lymphoma are associated with blastoid variants. *Blood*, 93: 4365-4374, 1999.
- Knuutila, S., Bjorkqvist, A. M., Autio, K., Tarkkanen, M., Wolf, M., Monni, O., Szymanska, J., Larramendy, M. L., Tapper, J., Pirc, H., el-Rifai, W., Hemmer, S., Wasenius, V. M., Vidgren, V., and Zhu, Y. DNA copy number amplifications in human neoplasms: review of comparative genomic hybridization studies. *Am. J. Pathol.*, 152: 1107-1123, 1998.
- Alkema, M. J., Bronk, M., Verhoeven, E., Otte, A., van't Veer, L. J., Berns, A., and van Lohuizen, M. Identification of Bmi1-interacting proteins as constituents of a multimeric mammalian polycomb complex. *Genes Dev.*, 11: 226-240, 1997.
- Pinyol, M., Hernandez, L., Martinez, A., Cobo, F., Hernandez, S., Bea, S., Lopez-Guillermo, A., Nayach, I., Palacin, A., Nadal, A., Fernandez, P., Montserrat, E., Cardesa, A., and Campo, E. *INK4a/ARF* locus alterations in human non-Hodgkin's lymphomas mainly occur in tumors with wild type *p53* gene. *Am. J. Pathol.*, 156: 1987-1996, 2000.
- Pinyol, M., Cobo, F., Bea, S., Jares, P., Nayach, I., Fernandez, P. L., Montserrat, E., Cardesa, A., and Campo, E. *p16^{INK4a}* gene inactivation by deletions, mutations, and hypermethylation is associated with transformed and aggressive variants of non-Hodgkin's lymphomas. *Blood*, 91: 2977-2984, 1998.
- Foot, A. B., Oakhill, A., and Kitchen, C. Acute monoblastic leukemia of infancy in Klinefelter's syndrome. *Cancer Genet. Cytogenet.*, 61: 99-100, 1992.
- Lessard, J., Baban, S., and Sauvageau, G. Stage-specific expression of polycomb group genes in human bone marrow cells. *Blood*, 91: 1216-1224, 1998.
- Raaphorst, F. M., van Kemenade, F. J., Fieret, E., Hamer, K. M., Satijn, D. P., Otte, A. P., and Meijer, C. J. Cutting edge: polycomb gene expression patterns reflect distinct B cell differentiation stages in human germinal centers. *J. Immunol.*, 164: 1-4, 2000.
- Raaphorst, F. M., van Kemenade, F. J., Blokzijl, T., Fieret, E., Hamer, K. M., Satijn, D. P., Otte, A. P., and Meijer, C. J. Coexpression of BMI-1 and EZH2 polycomb group genes in Reed-Sternberg cells of Hodgkin's disease. *Am. J. Pathol.*, 157: 709-715, 2000.
- Gould, A. Functions of mammalian Polycomb group and trithorax group related genes. *Curr. Opin. Genet. Dev.*, 7: 488-494, 1997.
- van Oostveen, J., Bijl, J., Raaphorst, F., Walboomers, J., and Meijer, C. The role of homeobox genes in normal hematopoiesis and hematological malignancies. *Leukemia*, 13: 1675-1690, 1999.

Gene-expression profiles predict survival of patients with lung adenocarcinoma

DAVID G. BEER¹, SHARON L.R. Kardia², CHIANG-CHING HUANG³, THOMAS J. GIORDANO⁴, ALBERT M. LEVIN², DAVID E. MISEK⁵, LIN LIN¹, GUOAN CHEN¹, TAREK G. GHARIB¹, DAFYDD G. THOMAS⁴, MICHELLE L. LIZYNESS⁴, RORK KUIICK⁵, SATORU HAYASAKA³, JEREMY M.G. TAYLOR¹, MARK D. IANNETTONI¹, MARK B. ORRINGER¹ & SAMIR HANASH⁵

Departments of ¹Surgery, ²Epidemiology, ³Biostatistics, ⁴Pathology and ⁵Pediatrics, University of Michigan, Ann Arbor, Michigan, USA

Correspondence should be addressed to D.G.B.; email: dgbeer@umich.edu.

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Histopathology is insufficient to predict disease progression and clinical outcome in lung adenocarcinoma. Here we show that gene-expression profiles based on microarray analysis can be used to predict patient survival in early-stage lung adenocarcinomas. Genes most related to survival were identified with univariate Cox analysis. Using either two equivalent but independent training and testing sets, or 'leave-one-out' cross-validation analysis with all tumors, a risk index based on the top 50 genes identified low-risk and high-risk stage I lung adenocarcinomas, which differed significantly with respect to survival. This risk index was then validated using an independent sample of lung adenocarcinomas that predicted high- and low-risk groups. This index included genes not previously associated with survival. The identification of a set of genes that predict survival in early-stage lung adenocarcinoma allows delineation of a high-risk group that may benefit from adjuvant therapy.

Lung cancer remains the leading cause of cancer death in industrialized countries. Most patients with non-small cell lung cancer (NSCLC) present with advanced disease, and despite recent advances in multi-modality therapy, the overall 10-year survival rate remains a dismal 8–10%¹. However, a significant minority of patients (~25–30%) with NSCLC have stage I disease and receive surgical intervention alone. Although 35–50% of patients with stage I disease will relapse within 5 years^{2,3}, it is not currently possible to identify specific high-risk patients.

Adenocarcinoma is currently the predominant histological subtype of NSCLC (refs. 1,5,6). Although morphological assessment of lung carcinomas can roughly stratify patients, there is a need to identify patients at high risk for recurrent or metastatic disease. Preoperative variables that affect survival of patients with NSCLC have been identified^{7–10}. Tumor size, vascular invasion, poor differentiation, high tumor-proliferative index and several genetic alterations, including *K-ras* (refs. 11,12) and *p53* (refs. 10,13) mutations, have prognostic significance. Multiple independently assessed genes or gene products have also been investigated to better predict patient prognosis in lung cancer^{14–16}. Technologies that simultaneously analyze the expression of thousands of genes¹⁷ can be used to correlate gene-expression patterns with numerous clinical parameters—including patient outcome—to better predict tumor behavior in individual patients¹⁸. Analyses of lung cancers using array technologies have identified subgroups of tumors that differ according to tumor type and histological subclasses and, to a lesser extent, survival among adenocarcinoma patients^{19,20}. Here we correlated gene-expression profiles with clinical outcome in a cohort of patients with lung adenocarcinoma and identified specific genes that

predict survival among patients with stage I disease. For further validation, we also show that the risk index predicted survival in an independent cohort of stage I lung adenocarcinomas.

Hierarchical profile clustering yields three tumor subsets

Using oligonucleotide arrays, we generated gene-expression profiles for 86 primary lung adenocarcinomas, including 67 stage I and 19 stage III tumors, as well as 10 non-neoplastic lung samples. Selected sample replicates showed high correlation among coefficients and reliable reproducibility. We determined transcript abundance using a custom algorithm and the data set was trimmed of genes expressed at extremely low levels, that is, genes were excluded if the measure of their 75th percentile value was less than 100. Although potentially resulting in the loss of some information, trimming in this manner decreased the possibility that the clustering algorithm would be strongly influenced by genes with little or no expression in these samples. Hierarchical clustering with the resulting 4,966 genes yielded 3 clusters of tumors (Fig. 1). All 10 non-neoplastic samples clustered tightly together within Cluster 1 (data not shown). We examined the relationships between cluster and patient and tumor characteristics (Fig. 1 and Supplementary Figure A online). There were associations between cluster and stage ($P = 0.030$) and between cluster and differentiation ($P = 0.01$). Cluster 1 contained the greatest percentage (42.8%) of well differentiated tumors, followed by Cluster 2 (27%) and Cluster 3 (4.7%). Cluster 3 contained the highest percentage of both poorly differentiated (57.6%) and stage III tumors (42.8%), yet contained 3 (14.3%) moderately differentiated and 1 (5%) well differentiated stage I tumor. Notably, 11 stage I tumors were present in Cluster 3, sug-

gesting a common gene-expression profile for this subset of stage I and stage III tumors.

For patients with stage I and stage III tumors, the average ages were 68.1 and 64.5 years and the percentage of smokers was 88.9% and 89.5%, respectively. Marginally significant associations between cluster and smoking history were observed ($P = 0.06$). A significant relationship between histopathological classification and cluster was only discernable for bronchioloalveolar adenocarcinomas (BAs), which were only present in Clusters 1 and 2 ($P = 0.0055$) and comprised 35.7% and 12.3% of tumors for Clusters 1 and 2, respectively.

We examined the heterogeneity in gene-expression profiles based on the trimmed data set among normal lung samples and stage I and stage III adenocarcinomas by calculating correlation coefficients between all pairs of samples. In contrast to normal lung samples that displayed highly similar gene-expression profiles (median correlation, 0.9), both stage I and III lung tumors demonstrated much greater heterogeneity in their expression profiles with lower correlation coefficients (median values, 0.82 and 0.79, respectively).

Northern-blot and immunohistochemistry analyses

Of the 4,966 genes examined, 967 differed significantly between stage I and III adenocarcinomas; a number in excess of that expected by chance alone (248 at alpha level (α) = 0.05). Three genes were arbitrarily selected to verify the microarray expression data. The mRNA from 20 of the normal lung and tumor samples was examined by northern-blot hybridization with probes for insulin-like growth factor-binding protein 3 (IGFBP3), cystatin C

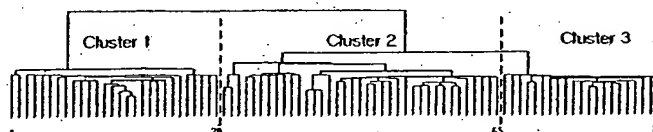


Fig. 1 Unsupervised classification analysis of lung adenocarcinomas. 3 classes of tumors identified by agglomerative hierarchical clustering of gene-expression profiles using the 4,966 expressed genes. Patient and histopathological information for each lung adenocarcinoma case by cluster designation and methods for K-ras 12/13th-codon mutational status and nuclear p53 protein accumulation are provided (Supplementary Figure A online). TN classification denotes information regarding patient tumor size and nodal involvement. Associations between cluster membership and patient or histopathological variables are indicated at significance level ($P \leq 0.05$).

and lactate dehydrogenase A (LDH-A) (Fig. 2a). Two gene probes not represented on the microarrays were used as controls, including histone H4, a potential index of overall cell proliferation, and 28S ribosomal RNA, a control for sample loading and transfer. The relative amounts of IGFBP3, cystatin C and LDH-A mRNA strongly correlated with microarray-based measurements (Fig. 2b). In both assays, IGFBP3 and LDH-A mRNA levels increased from stage I to stage III adenocarcinomas and were higher than those in normal lung. Cystatin C mRNA levels were more variable but relatively greater in normal lung than tumors. These results suggest that the oligonucleotide microarrays provided reliable measures of gene expression. The tumors showed slightly greater histone H4 expression than the normal lung, likely reflecting increased proliferation of tumor cells.

Immunohistochemistry was performed for IGFBP3, cystatin C and HSP-70 to determine whether mRNA overexpression was reflected by an increase of their corresponding proteins in tumors.

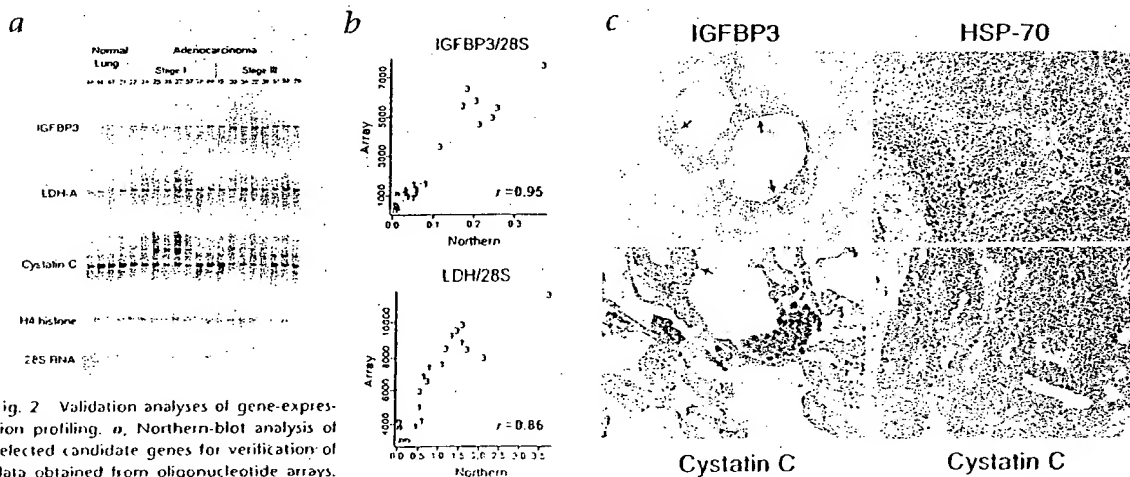


Fig. 2 Validation analyses of gene-expression profiling. **a**, Northern-blot analysis of selected candidate genes for verification of data obtained from oligonucleotide arrays. The same sample RNA for the 4 uninvolved lung, 8 stage I and 8 stage III tumors was used for the northern-blot and oligonucleotide array analyses. **b**, Correlation analysis of quantitative data obtained from oligonucleotide arrays and northern blots measured by integrated phosphorimager-based signals for the IGFBP3 and LDH-A genes. The ratio of IGFBP3, cystatin C and LDH-A mRNA to 28S rRNA was determined. The relative values for each gene from each sample are shown. n, non-neoplastic normal lung; 1, stage I tumors; 3, stage III tumors. **c**, Immunohistochemical analysis of IGFBP3, HSP-70 and cystatin C in lung and lung adenocarcinomas. Cytoplasmic IGFBP3 immunoreactivity in a neoplastic gland (tumor 127) with prominent apical staining (blue reactant staining, arrow, upper left). Diffuse cytoplasmic HSP-70 immunoreactivity (tumor 127), yet stromal elements show no reactivity (upper right). Normal lung parenchyma (lower left) shows cytoplasmic cystatin C immunoreactivity in alveolar pneumocytes (arrow) and intra-alveolar macrophages but tumor (L90) shows diffuse cytoplasmic cystatin C immunoreactivity with prominent apical staining (lower right). Magnification, $\times 200$.

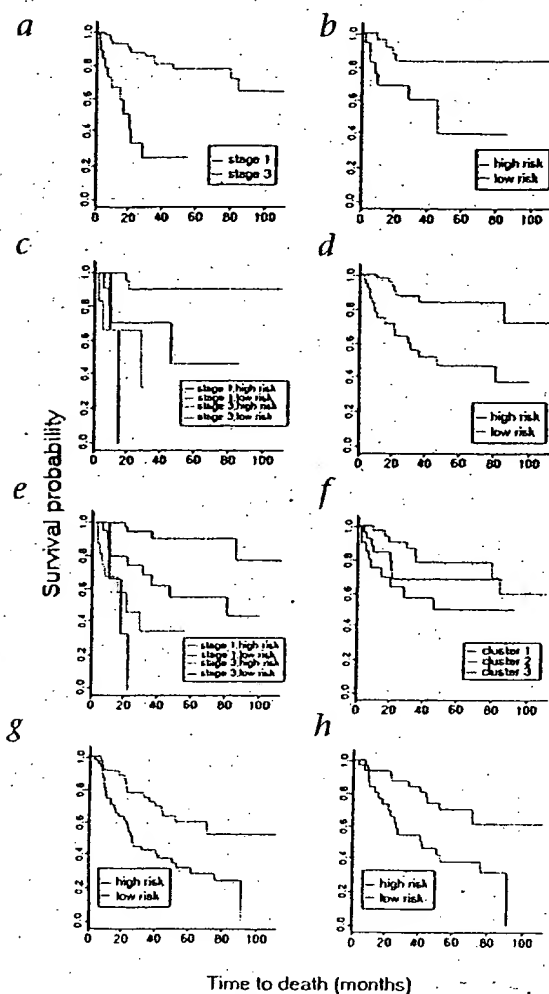


Fig. 3 Gene-expression profiles and patient survival. **a**, Relationship between tumor stage and patient survival (stage I and stage III differ significantly, $P < 0.0001$). **b**, Relationship between the survival in the 43 test samples and their risk assignments based on the 50-gene risk index estimated in the 43 training samples. The high- and low-risk groups differ significantly ($P = 0.024$). **c**, Relationship between patient survival and the risk assignments in test samples (in **b**) conditional for tumor stage. The high- and low-risk stage I groups differ significantly ($P = 0.028$), whereas stage III low- and high-risk groups did not ($P = 0.634$). **d**, Relationship between survival in the test cases and their risk assignments based on the 86 'leave-one-out' cross-validation of the 50-gene risk index. The high- and low-risk groups differ significantly ($P = 0.0006$). **e**, Relationship between test case's risk assignment and survival (in **d**) conditional on tumor stage. The high- and low-risk stage I lung adenocarcinoma groups differ significantly from each other ($P = 0.003$), whereas low- and high-risk stage III tumors do not. **f**, Relationship between tumor class identified by hierarchical clustering and patient survival. Survival for patients in Cluster 3 differed relative to the tumors in Cluster 2 ($P = 0.037$) and approached significance for Cluster 1 and 2 combined ($P = 0.06$). **g**, Analysis of the Michigan-based risk index using top cross-validated survival genes identify a low- and high-risk group in an independent cohort of 84 Massachusetts-based lung adenocarcinomas that are significantly different ($P = 0.003$). **h**, Among the 62 stage I lung adenocarcinomas in the Massachusetts sample, the high- and low-risk groups differed significantly ($P = 0.006$).

After conservatively choosing the 60th percentile cutoff point from the training set, we then applied this risk index and cutoff point to the testing set. The risk index of the top 50 genes correctly identified low- and high-risk individuals within the independent testing set ($P = 0.024$) (Fig. 3b and Supplementary Methods online). Notably, 11 stage I tumors were included in the high-risk subgroup. When this risk assignment was then conditionally examined for stage progression (Fig. 3c), low- and high-risk groups among stage I tumors were found to differ ($P = 0.028$) in their survival.

Identification of a robust set of survival genes

Although predictive of patient survival, a single training-testing set may not provide the most robust set of genes due to random sampling issues. Therefore, a 'leave-one-out' cross-validation approach was used to identify genes associated with survival from all 86-tumor samples. We first developed a 50-gene risk index in each training set, and then applied the risk index to the test case held out from the full set of tumors and assigned the held out tumor to the high- or low-risk groups (Fig. 3d). The high and low-risk subgroups determined in the test cases differed significantly in their overall survival ($P = 0.0006$). Among the larger group of stage I lung adenocarcinomas, the low-risk ($n = 46$) and high-risk ($n = 21$) groups had markedly different survival ($P = 0.003$) (Fig. 3e). Table 1 lists selected examples of the cumulative top 100 genes derived from this cross-validation procedure (complete list in Supplementary Table A online).

It was also noted that many of the stage I patients in the high-risk subgroup (Fig. 3e) were present in Cluster 3 (Fig. 1). Kaplan-Meier analysis (Fig. 3f) demonstrated a significantly worse survival ($P = 0.037$) for patients in Cluster 3 relative to patients in Cluster 2 and approaching significance for Cluster 1 and 2 combined ($P = 0.06$). This further indicates the important relationship between gene-expression profiles and patient survival, independent of disease stage.

Consistent with previous analyses of lung adenocarcinomas²³, 40% of stage I and 57.8% of stage III tumors had 12th or 13th codon *K-ras* gene mutations. Those patients with tumors containing *K-ras* mutations showed a trend of poorer survival, but

Immunoreactivity for both *IGFBP-3* and *HSP-70* (Fig. 2c) was detected in the cytoplasm of the adenocarcinomas, with little detectable reactivity in the stromal or inflammatory cells. Cystatin C was detected in alveolar pneumocytes and intra-alveolar macrophages in non-neoplastic lung parenchyma and also consistently in the cytoplasm of neoplastic cells.

Gene-expression profiles predict survival

As expected, Kaplan-Meier survival curves (Fig. 3a) and log-rank tests indicated poorer survival among stage III compared with stage I adenocarcinomas ($P = <0.0001$). Two statistical approaches were used to determine whether gene-expression profiles could predict survival using the data set of 4,966 genes. In one approach, equal numbers of randomly assigned stage I and stage III tumors constituted training ($n = 43$) and testing ($n = 43$) sets. In the training set, the top 10, 20, 50 or 75 genes were used to create risk indices that were evaluated for their association with survival using the 50th, 60th or 70th percentile cutoff points to categorize patients into high or low groups. The results were similar across cutoff points but the 50-gene risk index had the best overall association with survival in the training set.



Table 1 Selected examples of the top 100 genes from cross-validation

| Gene name | P (normal versus tumor t-test) | % Change in tumor | P (stage I versus stage III t-test) | % Change in stage III | Coefficient β | Unigene comment |
|-----------|--------------------------------------|----------------------|---|--------------------------|------------------------|--|
| CASP4 | 0.56 | -6% | 0.02 | 57% | 0.0022 | Apoptosis-related Caspase 4, apoptosis- related cysteine protease |
| P63 | 9.73E-04 | 37% | 0.03 | 43% | 0.0010 | Transmembrane protein (63 kD), endoplasmic reticulum/ Golgi intermediate compartment |
| KRT17 | 8.02E-08 | 126% | 0.11 | 55% | 0.0003 | Cell adhesion and structure Keratin 7 |
| LAMB1 | 0.14 | -20% | 0.01 | 60% | 0.0027 | Laminin, β 1 |
| BMP2 | 0.54 | -21% | 0.27 | 47% | 0.0044 | Cell cycle and growth regulators Bone morphogenetic protein 2 |
| CDC6 | 1.31E-05 | 1070% | 0.05 | 148% | 0.0124 | CDC6 (cell division cycle 6, <i>Saccharomyces cerevisiae</i> homolog) |
| S100P | 2.10E-08 | 1572% | 0.19 | 77% | 0.0001 | S100 calcium-binding protein P |
| SERPINE1 | 2.89E-03 | 72% | 0.25 | 30% | 0.0008 | Serine (or cysteine) proteinase inhibitor, clade E (nexin) |
| STX1A | 8.65E-08 | 54% | 0.07 | 26% | 0.0031 | Syntaxin 1A (brain) |
| ADM | 0.05 | 39% | 0.04 | 117% | 0.0016 | Cell signaling adrenomedullin |
| AKAP 12 | 8.53E-03 | -47% | 0.05 | 214% | 0.0010 | A kinase (PRKA) anchor protein (gravin) 12 |
| ARHE | 0.06 | -39% | 0.05 | 87% | 0.0092 | ras homolog gene family, member E |
| GRB7 | 2.02E-03 | 38% | 0.63 | 15% | 0.0030 | Growth factor receptor-bound protein 7 |
| VEGF | 6.50E-08 | 174% | 0.02 | 85% | 0.0013 | Vascular endothelial growth factor |
| WNT10B | 0.05 | 31% | 0.48 | 20% | 0.0022 | Wingless-type MMTV integration site family, member 10B |
| HSPA8 | 0.36 | 8% | 9.01E-04 | 51% | 0.0008 | Chaperones Heat-shock 70 kD protein 8 |
| ERBB2 | 0.04 | 92% | 0.37 | 120% | 0.0013 | Receptors v-erb-b2 avian erythroblastic leukemia viral oncogene homolog 2 |
| FXRD3 | 0.10 | 111% | 0.31 | 73% | 0.0046 | FXRD domain-containing ion transport regulator 3 |
| SLC20A1 | 1.34E-03 | 58% | 0.02 | 66% | 0.0021 | Solute carrier family 20 (phosphate transporter), member 1 |
| CSTB | 1.57E-04 | 50% | 0.15 | 34% | 0.0001 | Enzymes, cellular metabolism Cystatin B (stefin B) |
| CTSL | 0.48 | -10% | 0.03 | 67% | 0.0007 | Cathepsin L |
| CYP24 | 3.16E-06 | N/A | 0.97 | 2% | 0.0008 | Cytochrome P450, subfamily XXIV (vitamin D 24-hydroxylase) |
| FUT3 | 1.07E-07 | 114% | 0.97 | -1% | 0.0033 | Fucosyltransferase 3 (galactoside 3(4)-L- fucosyltransferase, Lewis blood group included) |
| MLN64 | 0.20 | 32% | 0.42 | 80% | 0.0007 | Steroidogenic acute regulatory protein related |
| PDE7A | 0.12 | 33% | 0.01 | -35% | -0.0187 | Phosphodiesterase 7A |
| PLG1 | 0.04 | -68% | 0.35 | -170% | -0.0011 | Plasminogen-like |
| SLC1A6 | 0.07 | -32% | 0.12 | 86% | 0.0069 | Solute carrier family 1 (high-affinity aspartate/ glutamate transporter), member 6 |
| COPEB | 0.10 | -33% | 0.26 | 25% | 0.0016 | Transcription and translation Core promoter element binding protein |
| CRK | 0.10 | 32% | 0.03 | 48% | 0.0098 | v-crk avian sarcoma virus CT10 oncogene homolog |
| RELA | 0.26 | -7% | 0.01 | 20% | 0.0034 | v-rel avian reticuloendotheliosis viral oncogene homolog A |
| KIAA0005 | 2.21E-04 | 40% | 0.02 | 45% | 0.0010 | Unknown function KIAA0005 gene product |
| MCB1 | 0.27 | 125% | 0.33 | 459% | 0.0018 | Mammaglobin 1 |

Bolded genes were also significant for survival in 43 tumor training set (Fig. 3b).

Table 1 Selected examples of the cumulative top 100 genes identified using training-testing, cross-validation of all 86 lung tumor samples. The percent change, as well as the direction, for the average values of the 10 non-neoplastic lung to all tumors, and for the 67 stage I to the 19 stage III tumors are shown. A positive coefficient β value is indicative of a relationship of gene expression to a

poorer patient outcome. The genes are listed in potential functional categories. Genes that were also present in the top 50 survival genes using the 43-tumor training set (Fig. 3b) are indicated in bold type. Complete listing of the gene probe sets and annotated gene and unigene identifiers can be found in the Supplementary Methods.

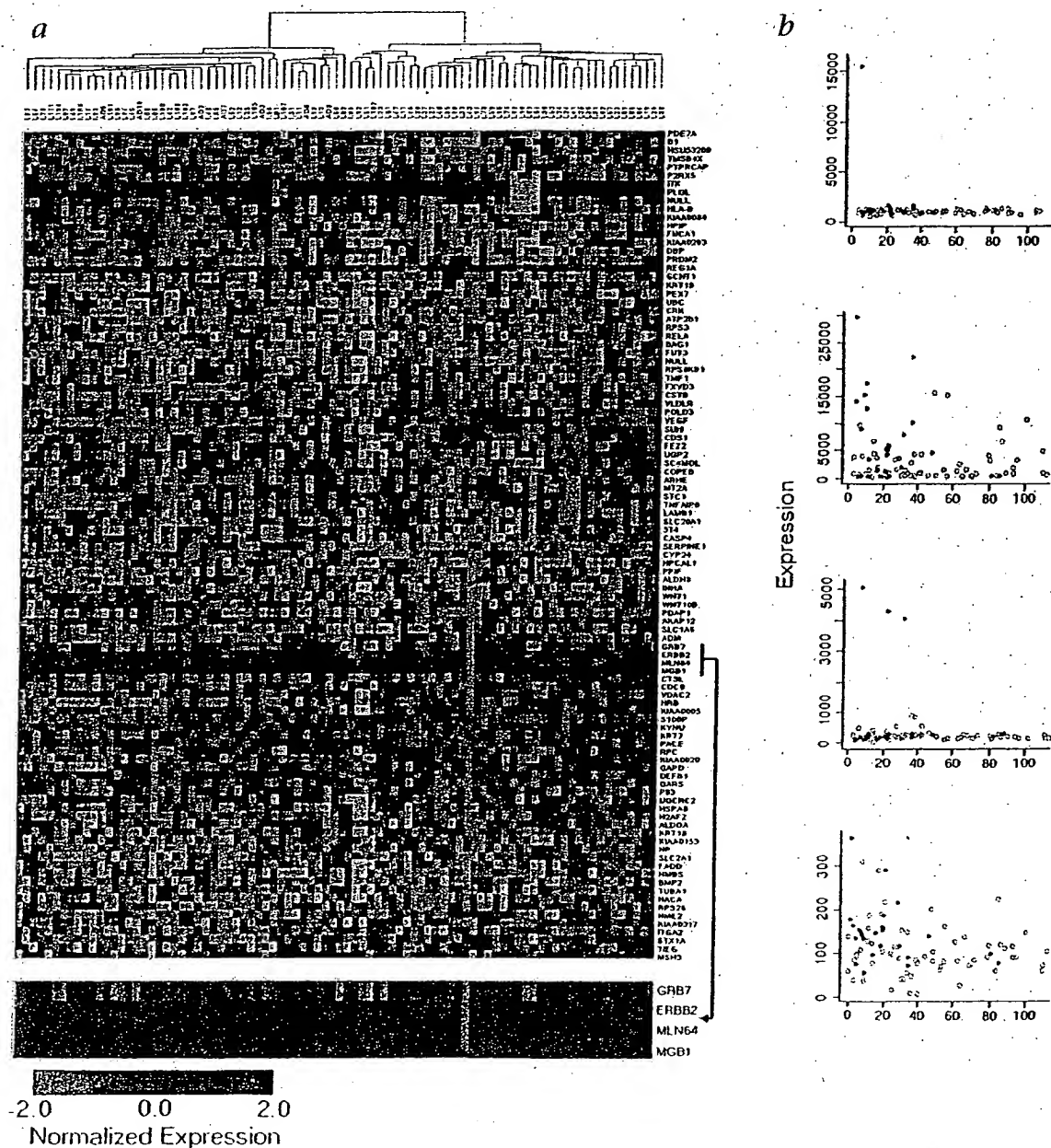


Fig. 4 Gene expression patterns of top survival genes. **a**, Gene-expression patterns determined using agglomerative hierarchical clustering of the 86 lung adenocarcinomas against the 100 survival-related genes (Table 1) identified by the training-testing, cross-validation analysis. Substantially elevated (red) or decreased (green) expression of the genes is observed in individual tumors. Some tumors (black arrow and expanded area) show extremely elevated expression of specific genes. **b**, An outlier gene-expression pattern (>5 times the interquartile range among all samples) is observed for the *erbB2* and *Reg1A* genes (top left and right, respectively). The *SLC20A1* and *erk* genes (bottom left and right, respectively) show a graded pattern of expression related to patient survival. \circ , alive; \bullet , dead (also in **c**). **c**, The number of outliers per person identified in the top 100 genes plotted by survival distribution.

this difference did not reach statistical significance among all patients ($P = 0.25$), between patients within tumor clusters ($P = 0.41$) or when analyzed separately among stage I ($P = 0.22$) and stage III ($P = 0.53$) patients. Nuclear accumulation of p53 was detected in 17.9% stage I and in 22.2% stage III tumors. No significant relationship was observed for p53 staining and patient survival, cluster or tumor stage.

Confirmation using an independent set of adenocarcinomas

The robustness of our 50-gene risk index in predicting survival in lung adenocarcinomas was tested using oligonucleotide gene-expression data obtained from a completely independent (Massachusetts-based) sample of 84 lung adenocarcinomas (62 stage I, 14 stage II and 8 stage III; ref. 21, and dataset A at www.genome.wi.mit.edu/MPR/lung). To ensure equivalent power for testing and comparability of samples, the criteria for including tumors in the analysis were 40% or greater tumor cellularity, no mixed histology (that is, adenosquamous) and patient survival information. To obtain comparative gene-expression measures between the two data sets, gene sequences present on the U95A and HuGeneFL array were examined, and expression data for our top 50 cross-validation genes for all 84 Massachusetts samples were obtained and processed²⁴ (see also Supplementary Methods online). When we examined the risk assignment of these 84 samples, employing the identical cutoff point used for the 86 Michigan-based lung samples, we observed low- and high-risk groups (Fig. 3g; $P = 0.003$). Notably, among the 62 stage I tumors, high- and low-risk groups were observed that differed significantly ($P = 0.006$) in their survival (Fig. 3h).

Survival genes had graded and outlier expression patterns

A statistical and graphical analysis of the 100 survival-related

genes (Table 1) clustered against all 86 tumors revealed individual tumors with substantially elevated expression in both a limited and larger number of genes (Fig. 4a). Among these genes, we observed two distinct patterns of expression related to patient survival. One pattern, designated 'outlier', included genes showing substantially elevated expression (greater than five times the interquartile range among all samples), whereas the other pattern, designated 'graded', was characterized by continuously distributed expression with patient survival (Fig. 4b). The *erbB2* and *Reg1A* genes are examples of outlier expression patterns and *S100P* and *crk* genes of graded patterns. The number of outliers per person in the top 100 genes was identified and plotted according to survival times and events (Fig. 4c). Both stage I and stage III lung adenocarcinomas showed outlier gene patterns and 10 tumors contained 3 or more outlier genes.

Because gene amplification may result in increased gene expression, the nine genes with outlier expression patterns (*erbB2*, *SLC1A6*, *Wnt 1*, *MGB1*, *Reg1A*, *AKAP12*, *PACE*, *CYP24*, *KYNU*) and one gene with a graded expression pattern (*KRT18*) were examined using quantitative genomic PCR to evaluate genomic copy number (Fig. 5a). Gene amplification of *erbB2* (17q12) was detected in tumor L94, which had the highest *erbB2* mRNA expression (Fig. 4a). Gene amplification was not detected for any of the other seven tested genes in tumor L94, as well as in other tumors. The two genes most frequently demonstrating the outlier pattern in these lung adenocarcinomas were *KYNU* and *CYP24*, and were present in 10 and 9 tumors, respectively. *CYP24* has been described as a gene amplified and overexpressed in breast cancer²⁵, and these results indicate elevated expression in lung adenocarcinoma.

To determine whether the graded or outlier gene-expression patterns also occur at the protein-expression level, 10 of the 100

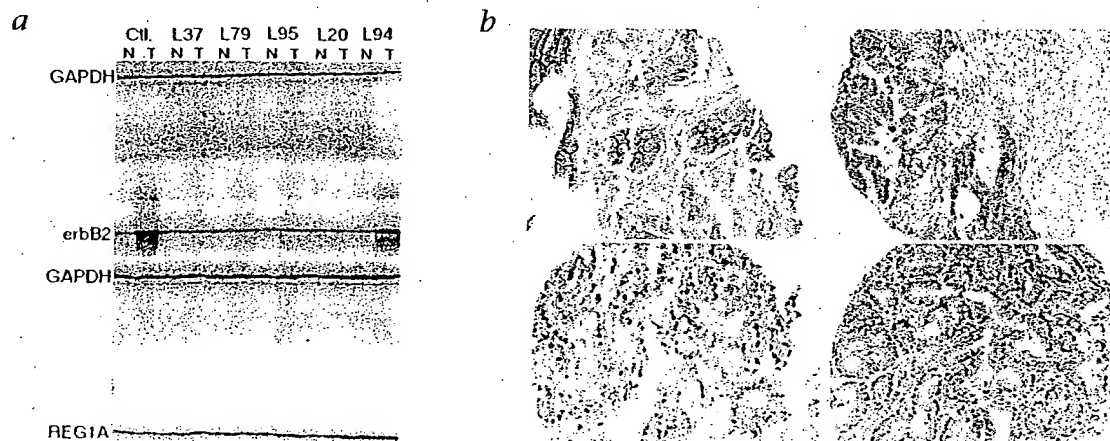


Fig. 5 Gene amplification and protein expression of survival-related genes. **a**, Analysis of potential gene amplification for 9 genes showing outlier expression patterns in the lung tumors (*erbB2*, *SLC1A6*, *Wnt 1*, *MGB1*, *Reg1A*, *AKAP12*, *PACE*, *CYP24* and *KYNU*) and examined using quantitative genomic PCR. A gene showing graded expression pattern (*KRT18*), and one gene (*PACE4*) with a similar chromosome location as *PACE1*, were used as controls. Only *erbB2* and *Reg1A* are shown. An esophageal adenocarcinoma with known high-level genomic amplification of *erbB2* was used as a positive control and normal esophagus DNA was used as a negative control (C11). PCR fragments sizes were 343 bp for *GAPDH*, 166 bp for *erbB2* and 126 bp for

Reg1A. DNA is from normal lung (N) and tumor (T) from each patient (for example L37). **b**, Immunohistochemical analysis of survival related genes with lung adenocarcinoma microarrays using the tumors from this study. The transmembrane *erbB2* protein (top left) expression is substantially increased in tumor L94 containing the amplified *erbB2* gene (Fig. 4a and b). Expression of *VEGF* (top right) and *S100P* (bottom left) was located within the neoplastic cells and the pattern of immunoreactivity was consistent with the graded expression pattern demonstrated by their mRNA profiles. Expression of the oncogene *crk* (bottom right) was abundantly expressed in neoplastic lung cells. Magnification, $\times 400$ (*erbB2*); $\times 200$ (*VEGF*, *S100P* and *crk*).

top survival genes (Table 1) for which specific antibodies were available were chosen for immunohistochemical analysis using lung-tumor arrays from this study (Fig. 5b). Expression of membrane *erbB2* protein was substantially increased in the *erbB2*-amplified tumor L94 and very low levels of expression were present in other tumors, consistent with mRNA-expression measurements (Fig. 4a and b). CDC6 protein expression was also substantially higher in tumor L94; consistent with mRNA levels (data not shown). Expression of vascular endothelial growth factor (VEGF) and S100P (Fig. 5b), as well as cytokeratin 18 (KRT18), cytokeratin 7 (KRT7) and fas-associated death domain (FADD) protein (data not shown), was located within the lung tumor cells and consistent with the graded expression pattern of the mRNA profiles. The oncogene *crk* showed both graded mRNA as well as a graded protein-expression pattern with survival, and was abundantly expressed in the tumor cells (Fig. 5b). These results indicate that many survival-associated genes are expressed at the protein level and demonstrate similar mRNA and protein-expression patterns.

Discussion

We used several approaches for the analysis of gene-expression data related to clinicopathological variables and patient survival. One approach, hierarchical clustering, was used to examine similarities among lung adenocarcinomas in their patterns of gene expression. Previous studies of lung tumors^{21,22} have also used this method to describe subclasses of lung tumors. Here, we found three clusters that showed significant differences with respect to tumor stage and tumor differentiation. This suggests, as expected, that tumors with similar histological features of differentiation demonstrate similarities in gene expression. This feature also partly underlies the observed statistical association of tumor stage and cluster, as many of the higher-stage tumors, often poorly differentiated and previously associated with a reduced survival¹⁰, were located in Cluster 3. Although this cluster contained the highest percentage of stage III tumors, it also contained a nearly equal mixture of stage I and stage III tumors and not all tumors were poorly differentiated. This indicates that a subset of stage I lung adenocarcinomas share gene-expression profiles with higher-stage tumors. Notably, 10 of the 11 stage I tumors found in Cluster 3 were the high-risk stage I tumors identified using the risk index in the 'leave-one-out' cross-validation.

In contrast to previous analyses of lung adenocarcinomas^{21,22}, we validated the expression data from the arrays. The strong correlation of northern-blot analysis and oligonucleotide-array data for gene expression in the same samples (Fig. 2b) indicates that these studies provide robust gene-expression estimates. Immunohistochemistry using the same tumor samples in tissue arrays demonstrates protein expression within the lung tumor cells. Together, these studies indicate that many of the genes identified using gene-expression profiles are likely relevant to lung adenocarcinoma. For example, *IGFBP3* gene expression is increased in lung adenocarcinomas (Fig. 2c). *IGFBP3* protein modulates the autocrine or paracrine effects of insulin-like growth factors; elevated *IGFBP3* expression is observed in colon cancer²⁸, and increased serum *IGFBP3* is associated with progression in breast cancer²⁷. Heat-shock protein 70 (HSP-70) is increased in lung adenocarcinomas of smokers²⁹ and is associated with increased metastatic potential in breast cancer²⁹. Increased serum lactate dehydrogenase is correlated with tumor stage and tumor burden³⁰, and cystatin C, a cysteine protease inhibitor ex-

pressed in human lung cancers³¹, is prognostic in some cancers³². The decreased expression of this protease inhibitor may affect the invasive properties of the tumor cell.

The cross-validation analytical strategy we used is particularly informative for these types of gene-expression analyses for disease outcome^{33,34}, and identification of cross-validated genes with a larger tumor cohort may help refine this risk index for use in a clinical setting. The gene-expression data also provide opportunities to observe overarching patterns that advance our understanding of associations between genes and disease. For example, the top 100 survival genes include those involved in signaling, cell cycle and growth, transcription, translation and metabolism. Expression of many of these genes is likely a function of increased proliferation and metabolism in the more aggressive tumors. Some genes, such as *erbB2* and *Reg1A* (Fig. 4a and b), were highly overexpressed in a few patients having poor survival. In one tumor, the *erbB2* gene was amplified (Fig. 5a), demonstrating that genomic changes may underlie the overexpression of a subset of these outlier genes. Immunohistochemistry confirmed protein overexpression in this patient's tumor (Fig. 5b). Notably, seven of the eight outlier genes were not amplified, indicating that other mechanisms underlie the increased mRNA expression of these survival-related genes.

Most genes showed a graded relationship between expression and patient survival. Genes such as that encoding VEGF, known to be strongly associated with survival in lung cancer^{35,36} were identified as related to patient survival in our study. VEGF demonstrated a graded expression pattern, as did the S100P and *crk* oncogene (Fig. 5b). S100P is a calcium-regulated protein not previously reported in lung cancer. The *crk* gene, the cellular homolog of the *v-crak* oncogene, is a member of a family of adaptor proteins involved in signal transduction and interacts directly with c-jun N-terminal kinase 1 (JNK1)³⁷. Although *crk* has not been shown to have a role in lung cancer, its role in the MAP-kinase pathway, which leads to activation of matrix metalloproteinase secretion and cell invasion³⁸, indicates potential involvement in the tumor cell invasion or metastasis of some lung adenocarcinomas. Among the many genes identified in this study, like *crk*, that may be causally involved in lung cancer progression (Table 1), some were related to survival in many patients, and others in only smaller subsets of patients. This result is consistent with the complex molecular architecture of tumors in general, the heterogeneity of lung adenocarcinomas in particular and the multiple mechanisms underlying tumor-cell survival, invasion and metastasis³⁹.

Our results demonstrate that a gene-expression risk profile—based on the genes most associated with patient survival—can distinguish stage I lung adenocarcinomas and differentiate prognoses. The particular genes that define the clusters, or are associated with survival, likely reflect the characteristics of the particular tumors included in the analysis. Current therapy for patients with stage I disease usually consists of surgical resection without adjuvant treatment²². Clearly, the identification of a high-risk group among patients with stage I disease would lead to consideration of additional therapeutic intervention for this group, possibly leading to improved survival of these patients.

Methods

Patient population. Sequential patients seen at the University of Michigan Hospital between May 1994 and July 2000 for stage I or stage III lung adenocarcinoma were evaluated for this study. Consent was received and the project was approved by the local Institutional Review Board. Primary tumors and adjacent non-neoplastic lung tissue were obtained at the time of

surgery. Peripheral portions of resected lung carcinomas were sectioned, evaluated by a study pathologist and compared with routine H&E sections of the same tumors, and utilized for mRNA isolation. Regions chosen for analysis contained a tumor cellularity greater than 70%, no mixed histology, potential metastatic origin, extensive lymphocytic infiltration or fibrosis. Tumors were histopathologically divided into two categories based on their growth pattern: bronchial-derived, if they exhibited invasive features with architectural destruction, and bronchioloalveolar, if they exhibited preservation of the lung architecture. All stage I patients received only surgical resection with intra-thoracic nodal sampling and no other treatments. Stage III patients received surgical resection plus chemotherapy and radiotherapy.

Gene-expression profiling and K-ras mutation analysis. RNA isolation, cRNA synthesis and gene-expression profiling were performed as described²⁴. Details of gene annotation and K-ras mutation analysis are provided in supplementary information.

Northern-blot analysis. Total cellular RNA (10 µg) was separated in 1.2% agarose-formaldehyde gels and vacuum-transferred to Gene Screen Plus (NEN Life Science Products, Boston, Massachusetts). Hybridization conditions and probe labeling were as described²⁵. Individual sequence-validated cDNA image clones for human IGF1BP3 (clone 1407750), LDH-A (clone 2420241), cystatin C (CTS3; clone 949938) were from Research Genetics (Huntsville, Alabama). The human histone H4 cDNA and the 28S ribosomal RNA 26-mer oligonucleotide probe were prepared and labeled as described²⁶.

Gene-amplification analysis. 11 genes were selected for the analysis of genomic alterations. Primers were designed using PrimerSelect 4.05 Windows 32 software (DNASTAR, Madison, Wisconsin), avoiding pseudogenes or potential homologous regions. Forward and reverse primers for the genes are provided (Supplementary Methods online). Quantitative genomic-PCR was then applied and analyzed as described²⁷.

Immunohistochemical staining. The H&E-stained slides of all primary lung tumors were used to identify the most representative regions of each tumor and a tissue microarray (TMA) block was constructed as described²⁸. Immunohistochemistry (IHC) was performed using both routine and sections from the TMA block as described²⁹. Detailed methods and the concentrations used for all antibodies are provided in the Supplementary Methods.

Statistical methods. *t*-tests were used to identify differences in mean gene-expression levels between comparison groups. Agglomerative hierarchical clustering³⁰ was applied using the average linkage method to investigate whether there was evidence for natural groupings of tumor samples based on correlations between gene-expression profiles. To investigate the robustness of the clustering inference, gene-expression values were perturbed by adding random Gaussian error of magnitude obtained from a duplicate sample to each data point and then reclustered to determine concordance in the tumor's class membership. Pearson, χ^2 and Fisher's exact tests were used to assess whether cluster membership was associated with physical and genetic characteristics of the tumors.

To determine whether gene-expression profiles were associated with variability in survival times, 2 separate but complementary approaches were used. In the first approach, the 86 tumors were randomly assigned to equivalent training and testing sets consisting of equal numbers of stage I and III tumors in order to validate a novel risk-index function that captured the effect of many genes at once. In the second approach, cross-validation³¹ was used to more robustly identify the genes associated with survival. Briefly, a 'leave-one-out' cross-validation procedure in which 85 of the 86 tumors (the training set) was used to identify genes that were univariately associated with survival. The risk index was defined as a linear combination of the gene-expression values for the top genes identified by univariate Cox proportional-hazard regression modeling³², weighted by their estimated regression coefficients. Kaplan-Meier survival plots and log-rank tests were then used to assess whether the risk-index assignment to high/low categories was validated in the test set. A more detailed description is provided (Supplementary Methods online).

Note: Supplementary information is available on the Nature Medicine website.

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Competing interests statement

The authors declare that they have no competing financial interests.

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1. Fry, W.A., Phillips, J.L. & Menck, H.R. Ten-year survey of lung cancer treatments and survival in hospitals in the United States. *Cancer* 66, 1867–1876 (1999).
2. Williams, D.E. et al. Survival of patients surgically treated for stage I lung cancer. *J. Thorac. Cardiovasc. Surg.* 82, 70–76 (1981).
3. Pairello, P.C. et al. Postsurgical stage I bronchogenic carcinoma: Morbid implications of recurrent disease. *Ann. Thorac. Surg.* 38, 331–338 (1984).
4. Naruke, T. et al. Prognosis and survival in resected carcinoma based on the new international staging system. *J. Thorac. Cardiovasc. Surg.* 96, 440–447 (1988).
5. Kaisermann, M.C. et al. Evolving features of lung adenocarcinoma in Rio de Janeiro, Brazil. *Oncol. Rep.* 8, 189–192 (2001).
6. Roggli, V.L. et al. Lung cancer heterogeneity: A blinded and randomized study of 100 consecutive cases. *Hum. Pathol.* 16, 569–579 (1985).
7. Gail, M.H. et al. Prognostic factors in patients with resected stage I non-small cell lung cancer: A report from the Lung Cancer Study Group. *Cancer* 54, 1802–1813 (1984).
8. Takise, A. et al. Histopathologic prognostic factors in adenocarcinomas of the peripheral lung less than 2 cm in diameter. *Cancer* 61, 2083–2088 (1988).
9. Ichinose, Y. et al. Is T factor of the TMN staging system a predominant prognostic factor in pathologic stage I non-small cell lung cancer. *J. Thorac. Cardiovasc. Surg.* 106, 90–94 (1993).
10. Harpole, D.H. et al. A prognostic model of recurrence and death in stage I non-small cell lung cancer utilizing presentation, histopathology, and oncoprotein expression. *Cancer Res.* 55, 51–56 (1995).
11. Rodenhuis, S. et al. Mutational activation of the K-ras oncogene: A possible pathogenic factor in adenocarcinoma of the lung. *N. Engl. J. Med.* 317, 929–935 (1987).
12. Slebos, R.J.C. et al. K-ras oncogene activation as a prognostic marker in adenocarcinoma of the lung. *N. Engl. J. Med.* 323, 561–565 (1990).
13. Horio, Y. et al. Prognostic significance of p53 mutations and 3p deletions in primary resected non-small cell lung cancer. *Cancer Res.* 53, 1–4 (1993).
14. Kern, J.A. et al. C-erbB-2 expression and codon 12 K-ras mutations both predict shortened survival for patients with pulmonary adenocarcinomas. *J. Clin. Invest.* 93, 516–520 (1994).
15. Ebina, M. et al. Relationship of p53 overexpression and up-regulation of proliferating cell nuclear antigen with the clinical course of non-small cell lung cancer. *Cancer Res.* 54, 2496–2503 (1994).
16. Mehdi, S.A. et al. Prognostic markers in resected stage I and II non-small cell lung cancer: an analysis of 260 patients with 5 year follow-up. *Clin. Lung Cancer* 1, 59–67 (1997).
17. Schneider, P.M. et al. Multiple molecular marker testing (p53, c-Ki-ras, c-erbB-2) improves estimation of prognosis in potentially curative resected non-small cell lung cancer. *Br. J. Cancer* 83, 473–479 (2000).
18. Herbst, R.S. et al. Differential expression of E-cadherin and type IV collagenase genes predicts outcome in patients with stage I non-small cell lung carcinoma. *Clin. Oncol.* 6, 790–797 (2000).
19. Liotta, L. & Petricion, E. Molecular profiling of human cancer. *Nature Rev. Genet.* 1, 48–56 (2000).
20. Golub, T.R. Editorial: Genome-wide views of cancer. *N. Engl. J. Med.* 344, 601–602 (2001).
21. Bhattacharjee, A. et al. Classification of human lung carcinomas by mRNA expression profiling reveals distinct adenocarcinoma subclasses. *Proc. Natl. Acad. Sci. USA* 98, 13790–13795 (2001).
22. Garber, M.E. et al. Diversity of gene expression in adenocarcinoma of the lung. *Proc. Natl. Acad. Sci. USA* 98, 13784–13789 (2001).
23. Mills, N.E. et al. Increased prevalence of K-ras oncogene mutations in lung adenocarcinoma. *Cancer Res.* 55, 1444–1447 (1995).
24. Giordano, J. et al. Organ-specific molecular classification of lung, colon and ovarian adenocarcinomas using gene expression profiles. *Am. J. Pathol.* 159, 1231–1238 (2001).
25. Albertson, D.G. et al. Quantitative mapping of amplicon structure by array CGH identifies CYP24 as a candidate oncogene. *Nature Genet.* 25, 144–146 (2000).
26. Kanstra, S. et al. IGF1BP3 mediates IGF1 proliferative response in colon cancer cells. *Int. J. Cancer* 87, 373–378 (2000).
27. Vadgama, J.V. et al. Plasma insulin-like growth factor-I and serum IGF-binding protein 3 can be associated with the progression of breast cancer, and predict the risk of recurrence and the probability of survival in African-American and Hispanic

ARTICLES

- women. *Oncology* 57, 330-340 (1999).
28. Volm, M., Mattern, J. & Stämmler, G. Up-regulation of heat shock protein 70 in adenocarcinoma of the lung in smokers. *Anticancer Res.* 15, 2607-2609 (1995).
 29. Ciocka, D.R. et al. Heat shock protein hsp70 in patients with axillary lymph node-positive breast cancer: prognostic implications. *J. Natl. Cancer. Inst.* 85, 570-574 (1993).
 30. Rotenberg, Z. et al. Total lactate dehydrogenase and its isoenzymes in serum of patients with non-small cell lung cancer. *Clin. Chem.* 34, 668-670 (1988).
 31. Krepela, E. et al. Cysteine proteases and cysteine protease inhibitors in non-small cell lung cancer. *Neoplasia* 45, 318-331 (1998).
 32. Kos, J. et al. Cysteine proteinases and their inhibitors in extracellular fluids: Markers for diagnosis and prognosis in cancer. *Int. J. Biol. Markers* 15, 84-89 (2000).
 33. Golub, T.R. et al. Molecular classification of cancer: Class discovery and class prediction by gene expression monitoring. *Science* 286, 531-537 (1999).
 34. Hedenfalk, I. et al. Gene-expression profiles in hereditary breast cancer. *N. Engl. J. Med.* 344, 539-548 (2001).
 35. Ohla, Y. et al. Vascular endothelial growth factor and lymph node metastasis in primary lung cancer. *Br. J. Cancer* 76, 1041-1045 (1997).
 36. Shibusa, T., Shijubo, N. & Abe, S. Tumor angiogenesis and vascular endothelial growth factor expression in stage I lung adenocarcinoma. *Clin. Cancer Res.* 4, 1483-1487 (1998).
 37. Girardin, S.E. & Yaniv, M. A direct interaction between JNK1 and Crkl is critical for Rac1-induced JNK activation. *EMBO J.* 20, 3437-3446 (2001).
 38. Liu, E. et al. The Ras-mitogen-activated protein kinase pathway is critical for the activation of matrix metalloproteinase secretion and the invasiveness in v-crk-transformed 3Y1. *Cancer Res.* 60, 2361-64 (2000).
 39. Hanahan, D. & Weinberg, R.A. The hallmarks of cancer. *Cell* 100, 57-70 (2000).
 40. Hanson, L.A. et al. Expression of the glucocorticoid receptor and K-ras genes in urethan-induced mouse lung tumors and transformed cell lines. *Exp. Lung. Res.* 17, 371-387 (1991).
 41. Lin, L. et al. A minimal critical region of the 8p22-23 amplicon in esophageal adenocarcinomas defined using STS-amplification mapping and quantitative PCR includes the GATA-4 gene. *Cancer Res.* 60, 1341-1347 (2000).
 42. Kononen, J. et al. Tissue microarrays for high throughput molecular profiling of tumor specimens. *Nature Med.* 4, 844-847 (1998).
 43. Johnson, R. & Wichern, D.W. *Applied Multivariate Statistical Analysis*. 543-578 (Prentice Hall, New Jersey, 1988).
 44. Stone, M. Asymptotics for and against cross-validation. *Biometrika* 64, 29-38 (1977).
 45. Cox, D.R. Regression models and life tables. *J.R. Stat. Soc.* 34, 187-220 (1972).





Rapid quantitation of proinflammatory and chemoattractant cytokine expression in small tissue samples and monocyte-derived dendritic cells: validation of a new real-time RT-PCR technology.

Blaschke V, Reich K, Blaschke S, Zipprich S, Neumann C.

Department of Dermatology, von-Siebold-Str. 3, D-37075, Goettingen, Germany.
vblasch@gwdg.de

The analysis of cytokine profiles plays a central part in the characterization of disease-related inflammatory pathways and the identification of functional properties of immune cell subpopulations. Because tissue biopsy samples are too small to allow the detection of cytokine protein, the detection of mRNA by RT-PCR analysis is often used to investigate the cytokine milieu in inflammatory lesions. RT-PCR itself is a qualitative method, indicating the presence or absence of specific transcripts. With the use of internal or external standards it may also serve as a quantitative method. The most widely accepted method is quantitative competitive RT-PCR, based on internal shortened standards. Recently, online real-time PCR has been introduced (LightCycler), which allows quantitation in less than 30 min. Here, we have tested its use for the analysis of cytokine gene expression in different experimental in vitro and ex vivo settings. First, we compared quantitative competitive RT-PCR with real-time RT-PCR in the quantitation of transcription levels of the CD4(+) cell-specific chemoattractant Interleukin-16 during the maturation of monocyte-derived dendritic cells, and found a good correlation between both methods. Second, differences in the amounts of IL-16 mRNA in synovial tissue from patients with rheumatoid arthritis and osteoarthritis as assessed by real-time RT-PCR paralleled differences in the level of IL-16 protein in the synovial fluid. Finally, we employed real-time RT-PCR to study the cutaneous expression of several cytokines during experimental immunomodulatory therapy of psoriasis by Interleukin-10, and demonstrate that the technique is suitable for pharmacogenomic monitoring. In summary, real-time RT-PCR is a sensitive and rapid tool for quantifying mRNA expression even with small quantities of tissue. The results obtained do not differ from those generated by quantitative competitive RT-PCR.

Publication Types:

- Evaluation Studies

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Butyrate-induced reversal of dexamethasone resistance in autonomous rat Nb2 lymphoma cells.

Buckley AR, Krumenacker JS, Buckley DJ, Leff MA, Magnuson NS, Reed JC, Miyashita T, de Jong C, Gout PW.

Department of Pharmacology and Toxicology, University of North Dakota School of Medicine and Health Sciences, Grand Forks 58202-9037, USA.

abuckley@mail.med.und.nodak.edu

The parental rat Nb2 lymphoma is a prolactin (PRL)-dependent T cell line. Exposure of a PRL-independent subline, Nb2-SFJCD1, to sodium butyrate (NaBT) causes transient reversal of their growth factor-independent proliferation in association with constitutive expression of protooncogenes pim-1 and c-myc. In the present study, we investigated the effect of NaBT treatment on the sensitivity of Nb2-SFJCD1 cells to dexamethasone (DEX)-induced apoptosis. Pretreatment with NaBT (2 mM, 72 h) partially reversed resistance to apoptosis in Nb2-SFJCD1 cells exposed to DEX (100 nM) for 12 h, assessed by flow cytometric analyses of DNA fragmentation. However, the cytolytic effect of DEX was abrogated by PRL in a time- and concentration-dependent manner. Evaluation of apoptosis-associated gene expression in NaBT-pre-treated cultures incubated with DEX or DEX+PRL indicated that the apoptosis resistance did not stem from altered bcl-2 or bax expression. However, there was a strong correlation between the resistance to DEX-activated apoptosis and their enhanced expression of pim-1 mRNA and protein. The results show that it is possible to reverse DEX-induced apoptosis of Nb2 pre-T cells and suggest the pim-1 gene product has an important role as a suppressor of this process, perhaps functioning as a mediator of PRL action.

PMID: 14646523 [PubMed]



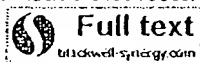
Alterations in neuropeptide Y levels and Y1 binding sites in the Flinders Sensitive Line rats, a genetic animal model of depression.

Caberlotto L, Jimenez P, Overstreet DH, Hurd YL, Mathe AA, Fuxe K.

Department of Neuroscience, Karolinska Institute, Stockholm, Sweden.

Previously, we observed specific alterations of neuropeptide Y (NPY) and Y1 receptor mRNA expression in discrete regions of the Flinders Sensitive Line rats (FSL), an animal model of depression. In order to clarify the correlation between mRNA expression and protein content, radioimmunoassay and receptor autoradiography were currently performed. In the FSL rats, NPY-like immunoreactivity (NPY-LI) was decreased in the hippocampal CA region, while Y1 binding sites were increased; NPY-LI was increased in the arcuate nucleus. Fluoxetine treatment elevated NPY-LI in the arcuate and anterior cingulate cortex and increased Y1 binding sites in the medial amygdala and occipital cortex in both strains. No differences were found regarding the Y2 binding sites. The results demonstrate a good correlation between NPY peptide and mRNA expression, and sustain the possible involvement of NPY and Y1 receptors in depression.

PMID: 10327163 [PubMed - indexed for MEDLINE]



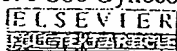
Neurokinin 1 receptor and relative abundance of the short and long isoforms in the human brain.

Caberlotto L, Hurd YL, Murdock P, Wahlin JP, Melotto S, Corsi M, Carletti R.

Department of Biology, Psychiatry CEDD, GlaxoSmithKline Medicine Research Centre, Verona, Italy. Laura.L.Caberlotto@gsk.com

Substance P exerts its various biochemical effects mainly via interactions through neurokinin-1 receptors (NK1). Recently, the NK1 receptor has attracted considerable interest for its possible role in a variety of psychiatric disorders including depression and anxiety. However, little is known regarding the anatomical distribution of NK1 in the human central nervous system (CNS). Riboprobe in situ hybridization, quantitative PCR and in vitro autoradiography were performed. Highest NK1 mRNA levels were localized in the locus coeruleus and ventral striatum, while moderate hybridization signals were observed in the cerebral cortex (most abundant in the visual cortex), hippocampus and different amygdaloid nuclei. Very low levels of the NK1 mRNA were detected in the cerebellum and thalamus. In view of the existence of a long and short isoform of the NK1 receptor, it was of interest to assess whether there was a differential distribution of the two splice variants in the human CNS and peripheral tissues. A quantitative TaqMan PCR analysis showed that the long NK1 isoform was the most prevalent throughout the human brain, while in peripheral tissues the truncated form was the most represented. 3H-Substance P autoradiography revealed a good correlation between receptor binding sites and NK1 mRNA expression throughout the brain, with the highest levels of binding in the locus coeruleus. These results provide the anatomical evidence that the NK1 receptors have a strong association with neuronal systems relevant to mood regulation and stress in the human brain, but do not suggest a region-specific role of the two isoforms in the CNS.

PMID: 12752772 [PubMed - indexed for MEDLINE]



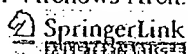
Characterization of cyclin D2 expression in human endometrium.

Choi D, Yoon S, Lee E, Hwang S, Song S, Kim J, Yoon BK, Lee JH.

Department of Obstetrics and Gynecology, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, South Korea. dschoi@smc.samsung.co.kr

OBJECTIVE: This study was undertaken to investigate cyclin D2 mRNA and protein expression in human endometrium during the menstrual cycle. **METHODS:** Endometrial samples were obtained from 15 premenopausal nonpregnant women who had hysterectomies for benign gynecologic reasons. They were divided into the following five groups according to histologic dating: early proliferative ($n = 3$), mid to late proliferative ($n = 3$), early secretory ($n = 3$), mid secretory ($n = 3$), and late secretory ($n = 3$). Cyclin D2 mRNA and protein expression were analyzed using reverse transcriptase-polymerase chain reaction, Western blotting, and immunohistochemistry. **RESULTS:** Cyclin D2 mRNA and protein were expressed in human endometrial tissue throughout the menstrual cycle. Cyclin D2 mRNA and protein expression of proliferative phase endometrium were significantly higher than those of secretory phase endometrium ($P < .05$). The staining intensity of cyclin D2 in proliferative phase endometrium was higher than that in secretory phase ($P < .05$). Cyclin D2 mRNA level showed good correlation with cyclin D2 protein level ($R = 0.579$, $P < .03$), and cyclin D2 protein also showed good correlation with immunohistochemical staining intensity ($R = 0.562$, $P < .03$). **CONCLUSION:** Cyclin D2 was expressed in human endometrium throughout the menstrual cycle. Cyclin D2 mRNA and protein were expressed at high levels in proliferative phase endometrium, especially in the early proliferative phase, and then decreased in the secretory phase.

PMID: 11839508 [PubMed - indexed for MEDLINE]



Human chorionic gonadotrophin beta expression in malignant Barrett's oesophagus.

Couvelard A, Paraf F, Vidaud D, Dubois S, Vidaud M, Flejou JE, Degott C.

Service d'Anatomie Pathologique, Hopital Beaujon, 92118 Clichy cedex, France.
anne.couvelard@bjn.ap-hop-paris.fr

BACKGROUND: Human chorionic gonadotrophin beta (hCGbeta) is expressed in several non-trophoblastic tumours, and this is usually associated with aggressive behaviour. Little is known about hCGbeta expression in Barrett's adenocarcinoma. **MATERIALS AND METHODS:** We determined the hCGbeta profile in a large series of surgically resected Barrett's adenocarcinoma (a) at mRNA level using real-time quantitative reverse-transcription polymerase chain reaction analysis and (b) at protein level using immunohistochemistry with a polyclonal antibody and with a monoclonal antibody specific for free hCGbeta. We then sought links between the hCGbeta protein expression pattern and clinical and pathological parameters, including patient outcome as well as vascular endothelial growth factor (VEGF) expression. **RESULTS:** hCGbeta protein expression was observed in 43 of 76 (57%) Barrett's adenocarcinomas. We showed a strong correlation between hCGbeta protein abundance and CGB mRNA level. We observed a statistical link between hCGbeta protein expression and infiltrative tumour type ($P=0.023$), perineural neoplastic invasion ($P=0.007$) and VEGF protein expression ($P=0.016$). hCGbeta expression tended to be associated with a poor outcome (16% versus 36% survival 8 years after resection). **CONCLUSION:** Expression of hCGbeta correlates with specific infiltrative characteristics and is associated with higher VEGF expression. Both molecules may play a co-ordinated role in the development of Barrett's adenocarcinomas.

PMID: 15309632 [PubMed - indexed for MEDLINE]



Downregulation of ENaC activity and expression by TNF-alpha in alveolar epithelial cells.

Dagenais A, Frechette R, Yamagata Y, Yamagata T, Carmel JF, Clermont ME, Brochiero E, Masse C, Berthiaume Y.

Centre de recherche, CHUM-Hotel-Dieu, 3850 St-Urbain, Montreal, Quebec, Canada H2W 1T7, andre.dagenais.chum@ssss.gouv.qc.ca

Sodium absorption by an amiloride-sensitive channel is the main driving force of lung liquid clearance at birth and lung edema clearance in adulthood. In this study, we tested whether tumor necrosis factor-alpha (TNF-alpha), a proinflammatory cytokine involved in several lung pathologies, could modulate sodium absorption in cultured alveolar epithelial cells. We found that TNF-alpha decreased the expression of the alpha-, beta-, and gamma-subunits of epithelial sodium channel (ENaC) mRNA to 36, 43, and 16% of the controls after 24-h treatment and reduced to 50% the amount of alpha-ENaC protein in these cells. There was no impact, however, on alpha(1) and beta(1) Na(+)-K(+)-ATPase mRNA expression. Amiloride-sensitive current and ouabain-sensitive Rb(+) uptake were reduced, respectively, to 28 and 39% of the controls. A strong correlation was found at different TNF-alpha concentrations between the decrease of amiloride-sensitive current and alpha-ENaC mRNA expression. All these data show that TNF-alpha, a proinflammatory cytokine present during lung infection, has a profound influence on the capacity of alveolar epithelial cells to transport sodium.

PMID: 14514522 [PubMed - indexed for MEDLINE]

Involvement of the CCND1 gene in hairy cell leukemia.

de Boer CJ, Kluin-Nelemans JC, Dreef E, Kester MG, Kluin PM, Schuurung E, van Krieken JH.

Department of Pathology, University of Leiden, The Netherlands.

BACKGROUND: Previous results suggested increased mRNA expression of CCND1 in hairy cell leukemia (HCL). The CCND1 gene is involved in the t(11;14)(q13;q32) chromosomal rearrangement, a characteristic abnormality in mantle cell lymphoma (MCL). We and others reported that, in contrast to other B-cell lymphomas, almost all MCL have over-expression of the CCND1 gene with a good correlation between RNA and protein analysis. Recent studies showed that overexpression of the cyclin D1 protein can be easily detected by immunohistochemistry (IHC) on formalin-fixed, paraffin embedded tissues. **PATIENTS AND METHODS:** To investigate whether the CCND1 gene is involved in HCL, we performed IHC on a series of 22 cases using formalin-fixed paraffin embedded splenectomy specimens. For IHC the sections were boiled in citrate buffer. The presence of rearrangements within the BCL-1 locus and the CCND1 gene was analyzed in 13 of 22 cases by Southern blot analysis using all available break-point probes. Expression of CCND1 was analyzed at the mRNA level (Northern blot) and protein level (IHC). **RESULTS:** Overexpression of the cyclin D1 protein using IHC was observed in all cases, with strong expression in 5 cases. Pre-existing B- and T-cell areas of the spleen did not express significant levels of the cyclin D1 protein. Seven of 9 cases analyzed by both IHC and Northern blotting showed overexpression of the CCND1 gene with both methods. No genomic abnormalities were observed in any of the 13 cases studied by Southern blot analysis. Additionally, no 11q13 abnormalities were detected by banding analysis of 19 of 22 cases. **CONCLUSIONS:** The elevated levels of CCND1 mRNA and protein in conjunction with the absence of overt rearrangements within the BCL-1 locus distinguish HCL from MCL and other B-cell malignancies. This suggests that activation of the CCND1 gene in HCL is due to mechanisms other than chromosomal rearrangement.

PMID: 8740788 [PubMed - indexed for MEDLINE]



Inhibin and activin production and subunit expression in human placental cells cultured in vitro.

Debieve F, Pampfer S, Thomas K

Department of Obstetrics and Gynecological Endocrinology, Universite Catholique de Louvain, 1200 Brussels, Belgium.

Inhibins and activins are dimeric proteins, with each subunit being one of three related protein subunits (alpha, betaA or betaB). The mRNA levels of these subunits were studied quantitatively during in-vitro differentiation of human cytotrophoblast cells into syncytium, using Northern blot analysis and semi-quantitative reverse transcription-polymerase chain reaction (RT-PCR) analysis. The corresponding protein concentrations were determined by specific enzyme-linked immunosorbent assays for inhibin A, B, pro alphaC and activin A in cellular protein extracts and culture medium (n = 5).

Immunofluorescence studies showed syncytium formation after 48 h. The alpha subunit was present before plating and increased at 48 h ($P < 0.001$) while the betaA subunit was weak before plating and increased at 24 h. The betaB subunit was not detected. With respect to corresponding protein synthesis, inhibin A (alpha + betaA) had risen after 48 h in cellular protein extract and after 72 h in culture medium, while activin A (betaA + betaB) was detected after 24 h, with no significant variations in culture medium. There was a good correlation between inhibin A and alpha subunit expression ($r = 0.736$, $P < 0.001$), as well as between activin A and betaA subunit expression ($r = 0.755$, $P < 0.001$). This study showed that mRNA expression parallels protein synthesis of inhibin and activin in trophoblast cells. Inhibin A synthesis appears to be dependent on alpha subunit mRNA expression, rather than on the betaA subunit which controls activin A synthesis. This study has also shown that isolated cytotrophoblast cells do not produce dimeric inhibin. However, during the transformation of cytotrophoblast cells into syncytium, betaA subunit mRNA expression may be an indicator of cell aggregation, while alpha subunit mRNA expression may be an indicator of cell fusion.

PMID: 10908285 [PubMed - indexed for MEDLINE]



Expression of membrane-type matrix metalloproteinases 4, 5, and 6 in mouse corneas infected with *P. aeruginosa*.

Dong Z, Katar M, Alousi S, Berk RS.

Department of Immunology and Microbiology, Wayne State University School of Medicine, 540 E. Canfield, Detroit, MI 48201, USA.

PURPOSE: To investigate the expression and regulation of membrane-type matrix metalloproteinases (MT-MMPs) 4, 5, and 6 in the mouse corneas infected with *Pseudomonas aeruginosa*. **METHODS:** C57BL/6J mice were intracorneally infected with *P. aeruginosa*. The expression of MT4-, MT5-, and MT6-MMP was detected at both the mRNA and protein levels by RT-PCR and immunoblot analysis. Immunohistochemical staining was performed to localize the expression of MT4- and MT5-MMP in the mouse corneas. **RESULTS:** Expression of MT4- and MT5-MMP was detected in the normal (uninfected) cornea by RT-PCR and immunoblot analysis. When infected with *P. aeruginosa*, the corneas showed significant induction of each MT-MMP. Localization of MT4- and MT5-MMP revealed that the expression of MT5-MMP was restricted to the epithelial tissue in the normal cornea, whereas the induced expression of MT4- and MT5-MMP was predominantly in the substantia propria, which contained most of the infiltrating cells. MT6-MMP expression was not detected in the uninfected cornea but was upregulated in the infected corneas. **CONCLUSIONS:** Expression of MT4-, MT5-, and MT6-MMP was induced in corneas infected with *P. aeruginosa*. Immunohistochemistry showed predominant immunoreactivity of MT4- and MT5-MMP in the substantia propria. Previous histologic studies have revealed different patterns of inflammatory cell infiltration with an increased number of polymorphonuclear neutrophils (PMNs) during the early stage of inflammation and increased macrophages during the late stage. These results indicate a good correlation between the overexpression of the MT-MMPs in the infected corneas and the inflammatory response—that is, leukocyte infiltration—indicating that inflammatory cells such as macrophages and PMNs may play a role in the upregulation of MT-MMPs during corneal infection, which in turn can cause the destruction of corneal tissue.

PMID: 11726626 [PubMed - indexed for MEDLINE]

Assessment of proliferative activity in colorectal carcinomas by quantitative reverse transcriptase-polymerase chain reaction (RT-PCR).

Duchrow M, Hasemeyer S, Broll R, Bruch HP, Windhovel U.

Surgical Research Laboratory, Surgical Clinic, Medical University of Lubeck, Ratzeburger Allee 160, D-23538 Lubeck, Germany.

The monoclonal antibody Ki-67 and the isospecific monoclonal antibody MIB-1 are routinely used in oncology to assess the proliferation index of tumor cells. A more objective and sensitive method is the determination of the of Ki-67 protein-specific mRNA by quantitative reverse transcriptase-polymerase chain reaction (RT-PCR). In 25 resected colorectal adenocarcinomas of different stages and grades we determined between 0.2 and 4.4 amol (10(-18) mol) Ki-67 protein-specific mRNA per microgram total RNA (median = 0.88 amol). The corresponding Ki-67 indices (expressing the percentage of Ki-67/MIB-1 positive tumor cells) ranged from 41 to 81% (median = 61%). We found a good correlation between Ki-67 index and mRNA expression ($r = 0.75$), a significant correlation between both data and tumor stage (primary tumor, regional nodes, metastasis [pTNM] staging classification) ($p < 0.001$), but not between both data and tumor grade. Both Ki-67 indices ($p = 0.05$) and mRNA levels ($p = 0.014$) correlated significantly to the patients' survival. These results demonstrate that the Ki-67 protein-specific quantitative RT-PCR is a useful method for the characterization of tumor cell proliferation.

PMID: 11486701 [PubMed - indexed for MEDLINE]

Comment in:

- [Equine Vet J. 2002 Jul;34\(4\):326-7.](#)

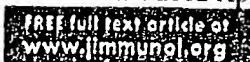
Molecular characterisation of carbohydrate digestion and absorption in equine small intestine.

Dyer J, Fernandez-Castano Merediz E, Salmon KS, Proudman CJ, Edwards GB, Shirazi-Beechey SP.

Department of Veterinary Preclinical Sciences, University of Liverpool, UK.

Dietary carbohydrates, when digested and absorbed in the small intestine of the horse, provide a substantial fraction of metabolisable energy. However, if levels in diets exceed the capacity of the equine small intestine to digest and absorb them, they reach the hindgut, cause alterations in microbial populations and the metabolite products and predispose the horse to gastrointestinal diseases. We set out to determine, at the molecular level, the mechanisms, properties and the site of expression of carbohydrate digestive and absorptive functions of the equine small intestinal brush-border membrane. We have demonstrated that the disaccharidases sucrase, lactase and maltase are expressed diversely along the length of the intestine and D-glucose is transported across the equine intestinal brush-border membrane by a high affinity, low capacity, Na⁺/glucose cotransporter type 1 isoform (SGLT1). The highest rate of transport is in duodenum > jejunum > ileum. We have cloned and sequenced the cDNA encoding equine SGLT1 and alignment with SGLT1 of other species indicates 85-89% homology at the nucleotide and 84-87% identity at the amino acid levels. We have shown that there is a good correlation between levels of functional SGLT1 protein and SGLT1 mRNA abundance along the length of the small intestine. This indicates that the major site of glucose absorption in horses maintained on conventional grass-based diets is in the proximal intestine, and the expression of equine intestinal SGLT1 along the proximal to distal axis of the intestine is regulated at the level of mRNA abundance. The data presented in this paper are the first to provide information on the capacity of the equine intestine to digest and absorb soluble carbohydrates and has implications for a better feed management, pharmaceutical intervention and for dietary supplementation in horses following intestinal resection.

PMID: 12117106 [PubMed - indexed for MEDLINE]



Suppressors of cytokine signaling proteins are differentially expressed in Th1 and Th2 cells: implications for Th cell lineage commitment and maintenance.

Egwuagu CE, Yu CR, Zhang M, Mahdi RM, Kim SJ, Gery I.

Laboratory of Immunology, National Eye Institute, National Institutes of Health, Bethesda, MD 20892, USA. emeka@helix.nih.gov

Positive regulatory factors induced by IL-12/STAT4 and IL-4/STAT6 signaling during T cell development contribute to polarized patterns of cytokine expression manifested by differentiated Th cells. These two critical and antagonistic signaling pathways are under negative feedback regulation by a multimember family of intracellular proteins called suppressor of cytokine signaling (SOCS). However, it is not known whether these negative regulatory factors also modulate Th1/Th2 lineage commitment and maintenance. We show here that CD4(+) naive T cells constitutively express low levels of SOCS1, SOCS2, and SOCS3 mRNAs. These mRNAs and their proteins increase significantly in nonpolarized Th cells after activation by TCR signaling. We further show that differentiation into Th1 or Th2 phenotype is accompanied by preferential expression of distinct SOCS mRNA transcripts and proteins. SOCS1 expression is 5-fold higher in Th1 than in Th2 cells, whereas Th2 cells contain 23-fold higher levels of SOCS3. We also demonstrate that IL-12-induced STAT4 activation is inhibited in Th2 cells that express high levels of SOCS3 whereas IL-4/STAT6 signaling is constitutively activated in Th2 cells, but not Th1 cells, with high SOCS1 expression. These results suggest that mutually exclusive use of STAT4 and STAT6 signaling pathways by differentiated Th cells may derive in part, from SOCS3- or SOCS1-mediated repression of IL-12/STAT4- or IL-4/STAT6 signaling in Th2 and Th1 cells, respectively. Given the strong correlation between distinct patterns of SOCS expression and differentiation into the Th1 or Th2 phenotype, SOCS1 and SOCS3 proteins are therefore Th lineage markers that can serve as therapeutic targets for immune modulation therapy.

PMID: 11907070 [PubMed - indexed for MEDLINE]

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Intravitreal invading cells contribute to vitreal cytokine milieu in proliferative vitreoretinopathy.

El-Ghrably IA, Dua HS, Orr GM, Fischer D, Tighe PJ.

Larry A Donoso Laboratory for Eye Research, Department of Ophthalmology, University of Nottingham, UK.

AIM: To examine the contribution of infiltrating cells in the local production of cytokines within the vitreous of patients with proliferative vitreoretinopathy (PVR). **METHODS:** The presence of mRNA coding for IL-6, IL-8, IL-1beta, IL-1alpha, TNFalpha, IFNgamma, IL-12, and HPRT was investigated in 25 vitreous samples from patients with PVR, 11 vitreous samples from patients with retinal detachment (RD) not complicated by PVR, and 10 vitreous samples from patients with macular hole (MH). A quantitative reverse transcriptase polymerase chain reaction (RT-PCR) using an internal competitor was used to investigate these samples. From these samples, 15 PVR, 8 RD, and 8 MH were analysed for the protein levels of the same cytokines using enzyme linked immunosorbent assay (ELISA). Spearman correlation was used to test any association between mRNA and cytokine protein levels as an indicator of the contribution these cells make to the intravitreal cytokine milieu. **RESULTS:** A strong correlation was found between mRNA and their respective cytokine levels (protein products) for IL-6, IL-8, IL-1beta, IL-1alpha, TNFalpha, IFNgamma (Spearman $r = 0.83, 0.73, 0.67, 0.91, 0.73,$ and 0.73 respectively), but not for IL-12. The median levels of IL-6, IL-8, IL-1beta, and IFNgamma mRNA and their respective cytokines were significantly higher ($p < 0.05$) in patients with PVR than in those with macular hole. There was no statistically significant difference in the median levels of IL-1alpha mRNA between PVR and MH but the cytokine IL-1alpha was detected at a significantly higher level in PVR compared with MH patients. Between PVR and RD patients, there was no statistically significant difference in mRNA levels for all the investigated cytokines ($p > 0.05$) except for IL-6 where there was a statistical significance ($p = 0.038$). In contrast, the median levels of IL-6, IL-8, and IL-1beta cytokines were significantly higher ($p < 0.05$) in patients with PVR than in those with RD, whereas for IL-1alpha and IFNgamma no significant statistical difference was detected between PVR and RD patients ($p > 0.05$). When results of RD and MH patients were compared, a statistical difference was only detected in mRNA levels of IFNgamma ($p = 0.008$). However, no difference was detected for IFNgamma (protein product) or for any of the other cytokines between RD and MH patients. **CONCLUSION:** Levels of both protein and mRNA encoding IL-6, IL-8, IL-1beta, and IFNgamma is significantly increased in vitreous samples from patients with PVR. The strong correlation between ELISA detectable cytokines (protein products) and their respective mRNA levels suggest that intravitreal, invasive cells are the major source of these cytokines, with the exception of IL-12. Cells invading the vitreous do not appear to locally produce IL-12 mRNA. This would appear to implicate cells peripheral to the

vitreal mass as the major source of this cytokine.

PMID: 11264138 [PubMed - indexed for MEDLINE]



Modulation of the glutamatergic receptors (AMPA and NMDA) and of glutamate vesicular transporter 2 in the rat facial nucleus after axotomy.

Eleore L, Vassias I, Vidal PP, de Waele C.

LNRS (CNRS-Paris V), ESA 7060, Centre Universitaire des Saints-Peres, 45 rue des Saints-Peres, 75270 Paris Cedex 06, France.

Facial nerve axotomy is a good model for studying neuronal plasticity and regeneration in the peripheral nervous system. We investigated in the rat the effect of axotomy on the different subunits of excitatory glutamatergic AMPA (GLuR1-4), NMDA (NR1, NR2A-D) receptors, post-synaptic density 95, vesicular glutamate transporter 2, beta catenin and cadherin. mRNA levels and/or protein production were analyzed 1, 3, 8, 30 and 60 days after facial nerve axotomy by in situ hybridization and immunohistofluorescence. mRNAs coding for the GLuR2-4, NR1, NR2A, B, D subunits of glutamatergic receptors and for post-synaptic density 95, were less abundant after axotomy. The decrease began as early as 1 or 3 days after axotomy; the mRNAs levels were lowest 8 days post-lesion, and returned to normal or near normal 60 days after the lesion. The NR2C subunit mRNAs were not detected in either lesioned or intact facial nuclei.

Immunohistochemistry using specific antibodies against GLuR2-3 subunits and against NR1 confirmed this down-regulation. There was also a large decrease in vesicular glutamate transporter 2 immunostaining in the axotomized facial nuclei at early stages following facial nerve section. In contrast, no decrease of NR2A subunit and of post-synaptic density 95 could be detected at any time following the lesion. beta Catenin and cadherin immunoreactivity pattern changed around the cell body of facial motoneuron by day 3 after axotomy, and then, tends to recover at day post-lesion 60 days. Therefore, our results suggest a high correlation between restoration of nerve/muscle synaptic contact, synaptic structure and function in facial nuclei. To investigate the mechanisms involved in the change of expression of these proteins following axotomy, the facial nerve was perfused with tetrodotoxin for 8 days. The blockade of action potential significantly decreased GLuR2-3, NR1 and NR2A mRNAs in the ipsilateral facial nuclei. Thus, axotomy-induced changes in mRNA abundance seemed to depend partly on disruption of activity.

PMID: 16182453 [PubMed - in process]

Altered levels of scavenging enzymes in embryos subjected to a diabetic environment.

Forsberg H, Borg LA, Cagliero E, Eriksson UJ.

Department of Medical Cell Biology, University of Uppsala, Sweden.

Maternal diabetes during pregnancy is associated with an increased rate of congenital malformations in the offspring. The exact molecular etiology of the disturbed embryogenesis is unknown, but an involvement of radical oxygen species in the teratological process has been suggested. Oxidative damage presupposes an imbalance between the activity of the free oxygen radicals and the antioxidant defence mechanisms on the cellular level. The aim of the present study was to investigate if maternal diabetes *in vivo*, or high glucose *in vitro* alters the expression of the free oxygen radical scavenging enzymes superoxide dismutase (CuZnSOD and MnSOD), catalase and glutathione peroxidase in rat embryos during late organogenesis. We studied offspring of normal and diabetic rats on gestational days 11 and 12, and also evaluated day-11 embryos after a 48 hour culture period in 10 mM or 50 mM glucose concentration. Both maternal diabetes and high glucose culture caused growth retardation and increased rate of congenital malformations in the embryos. The CuZnSOD and MnSOD enzymes were expressed on gestational day 11 and both CuZnSOD, MnSOD and catalase were expressed on day 12 with increased concentrations of MnSOD transcripts when challenged by a diabetic milieu. There was a good correlation between mRNA, protein, and activity levels, suggesting that the regulation of these enzymes occurs primarily at the pretranslational level. Maternal diabetes *in vivo* and high glucose concentration *in vitro* induced increased MnSOD expression, concomitant with increased total SOD activity, and a tentative decrease in catalase expression and activity in the embryos. These findings support the notion of enhanced oxidative stress in the embryo as an etiologic agent in diabetic teratogenesis.

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Induction of the estrogen receptor by growth hormone and glucocorticoid substitution in primary cultures of rat hepatocytes.

Freyschuss B, Stavreus-Evers A, Sahlin L, Eriksson H.

Department of Reproductive Endocrinology, Karolinska Hospital, Stockholm, Sweden.

Hepatic estrogen receptors (ER) mediate estrogenic effects on mammalian liver metabolism and are thereby involved in the regulation of important physiological/pathological processes, such as coagulation, atherosclerosis, and hypertension. The regulation of the formation of the ER in primary cultures of rat hepatocytes was studied by assaying ER and ER mRNA under different endocrine conditions. The ER concentration was measured using two different methods, a ligand-binding technique and an ER enzyme immunoassay. The results obtained by the two methods showed good correlation, and linear regression analysis gave a correlation coefficient of 0.95. ER concentrations fell to low steady state levels within 16 h after establishing the cell culture and remained low in the absence of hormonal substitution. Upon medium supplementation with pituitary GH and the glucocorticoid dexamethasone (DEX) in combination, the ER concentration increased 6-fold from 4.2 ± 1.0 to 25.8 ± 7.0 fmol/mg cytosolic protein. ER mRNA was measured by solution hybridization. Substitution with GH and DEX in combination increased ER mRNA to $210 \pm 14\%$ of control levels. No effect on ER mRNA stability was seen after hormone treatment. It is concluded that the regulatory effects of GH and DEX on the hepatic ER in this *in vitro* system are very similar to the effects of these hormones under *in vivo* conditions. The inducible expression of the ER has never before, to our knowledge, been demonstrated in any mammalian liver cell culture system.

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Cyclin D1-negative mantle cell lymphoma: a clinicopathologic study based on gene expression profiling.

Fu K, Weisenburger DD, Greiner TC, Dave S, Wright G, Rosenwald A, Chiorazzi M, Iqbal J, Gesk S, Siebert R, De Jong D, Jaffe ES, Wilson WH, Delabie J, Ott G, Dave BJ, Sanger WG, Smith LM, Rimsza L, Braziel RM, Muller-Hermelink HK, Campo E, Gascoyne RD, Staudt LM, Chan WC; Lymphoma/Leukemia Molecular Profiling Project.

Department of Pathology and Microbiology, University of Nebraska Medical Center, 983135 Nebraska Medical Center, Omaha, NE 68198-3135, USA. kfu@unmc.edu

Cyclin D1 overexpression is believed to be essential in the pathogenesis of mantle cell lymphoma (MCL). Hence, the existence of cyclin D1-negative MCL has been controversial and difficult to substantiate. Our previous gene expression profiling study identified several cases that lacked cyclin D1 expression, but had a gene expression signature typical of MCL. Herein, we report the clinical, pathologic, and genetic features of 6 cases of cyclin D1-negative MCL. All 6 cases exhibited the characteristic morphologic features and the unique gene expression signature of MCL but lacked the t(11;14)(q13; q32) by fluorescence in situ hybridization (FISH) analysis. The tumor cells also failed to express cyclin D1 protein, but instead expressed either cyclin D2 (2 cases) or cyclin D3 (4 cases). There was good correlation between cyclin D protein expression and the corresponding mRNA expression levels by gene expression analysis. Using interphase FISH, we did not detect chromosomal translocations or amplifications involving CCND2 and CCND3 loci in these cases. Patients with cyclin D1-negative MCL were similar clinically to those with cyclin D1-positive MCL. In conclusion, cases of cyclin D1-negative MCL do exist and are part of the spectrum of MCL. Up-regulation of cyclin D2 or D3 may substitute for cyclin D1 in the pathogenesis of MCL.

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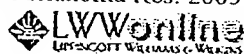
Oxytocin receptors in bovine cervix: distribution and gene expression during the estrous cycle.

Fuchs AR, Ivell R, Fields PA, Chang SM, Fields MJ.

Department of Obstetrics and Gynecology, Cornell University Medical College, New York, New York 10021, USA.

Oxytocin (OT) receptor (OTR) concentrations were determined in the cervix of nonpregnant cows on cycle Days 0, 3, 7-8, 17, and 19 (n = 3-4 cows each day); [³H]OT was used as the labeled ligand. Mucosal and muscle layers of the cervix were also analyzed separately for both ligand binding and expression of the OTR gene using a newly developed RNase protection assay (RAP). Cellular localization of OTR protein was determined by immunohistochemistry. All regions of cervix from cows at estrus had high concentrations of OTR; in the luteal phase, all were sharply down-regulated. At estrus the mucosal layer had about 30-fold higher concentrations than the muscle layer. OTR mRNA was readily detected by RAP in the mucosa from estrous cows, while much weaker signals were found in the muscle. On Days 7-17, the OTR mRNA signals in both mucosa and muscle were very faint or nondetectable. Thus, there was a good correlation between ligand binding and mRNA expression, which suggests that OTR concentrations are mainly regulated at the transcriptional level. The epithelial cells at the luminal surface of the mucosa were the principal site of immunoreactive OTR; muscle cells showed significantly weaker signals. Previously, OT was found to stimulate prostaglandin (PG) E₂ output in vitro in bovine cervical tissues. Since PGE₂ is capable of softening the cervix, our findings suggest that OT may have a novel physiological function to cause softening of the bovine cervix mediated by the release of PGE₂.

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Silencing of the thrombomodulin gene in human malignant melanoma.

Furuta J, Kameda A, Umebayashi Y, Otsuka F, Sugimura T, Ushijima T.

Carcinogenesis Division, National Cancer Center Research Institute, Tokyo, Japan.

The loss of thrombomodulin (TM) expression is associated with tumour growth, infiltration and lymph node metastasis in human tumours. In melanoma cell lines, TM is reported to mediate cell adhesion, and its introduction into TM-negative melanoma cell lines suppresses their growth. In this study, we analysed TM expression in surgical melanoma specimens and the role of its promoter methylation in the loss of its expression. In 15 (75%) of the 20 specimens (five from a primary site and 15 from metastatic sites), melanoma cells lacked TM immunoreactivity. Methylation of the TM promoter region was detected in 10 (67%) of the 15 TM-negative specimens by methylation-specific polymerase chain reaction, whereas methylation was detected in two (40%) of the five TM-positive specimens. In cell lines, complete methylation of the TM promoter CpG island was detected in six (46%) of 13 melanoma cell lines, whereas no methylation was detected in two cultured normal melanocytes. There was a good correlation between the methylated status of the CpG island and the loss of TM messenger RNA (mRNA) expression. Treatment of melanoma cell lines with a demethylating agent, 5-aza-2'-deoxycytidine, induced demethylation of the promoter CpG island and the restoration of mRNA and protein expression. These findings suggest that most human melanomas lack TM expression, and that methylation of the promoter CpG island is one of the mechanisms responsible.

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A sampling of the yeast proteome.

Fletcher B, Latter GI, Monardo P, McLaughlin CS, Garrels JJ.

Cold Spring Harbor Laboratory, Cold Spring Harbor, New York 11724, USA.
fletcher@cshl.org

In this study, we examined yeast proteins by two-dimensional (2D) gel electrophoresis and gathered quantitative information from about 1,400 spots. We found that there is an enormous range of protein abundance and, for identified spots, a good correlation between protein abundance, mRNA abundance, and codon bias. For each molecule of well-translated mRNA, there were about 4,000 molecules of protein. The relative abundance of proteins was measured in glucose and ethanol media. Protein turnover was examined and found to be insignificant for abundant proteins. Some phosphoproteins were identified. The behavior of proteins in differential centrifugation experiments was examined. Such experiments with 2D gels can give a global view of the yeast proteome.

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A Sampling of the Yeast Proteome

B. FUTCHER,^{1*} G. I. LATTER,¹ P. MONARDO,¹ C. S. McLAUGHLIN,² AND J. I. GARRELS³

Cold Spring Harbor Laboratory, Cold Spring Harbor, New York 11724¹; Department of Biological Chemistry, University of California, Irvine, California 92717²; and Proteome, Inc., Beverly, Massachusetts 01915³

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In this study, we examined yeast proteins by two-dimensional (2D) gel electrophoresis and gathered quantitative information from about 1,400 spots. We found that there is an enormous range of protein abundance and, for identified spots, a good correlation between protein abundance, mRNA abundance, and codon bias. For each molecule of well-translated mRNA, there were about 4,000 molecules of protein. The relative abundance of proteins was measured in glucose and ethanol media. Protein turnover was examined and found to be insignificant for abundant proteins. Some phosphoproteins were identified. The behavior of proteins in differential centrifugation experiments was examined. Such experiments with 2D gels can give a global view of the yeast proteome.

The sequence of the yeast genome has been determined (9). More recently, the number of mRNA molecules for each expressed gene has been measured (27, 30). The next logical level of analysis is that of the expressed set of proteins. We have begun to analyze the yeast proteome by using two-dimensional (2D) gels.

2D gel electrophoresis separates proteins according to isoelectric point in one dimension and molecular weight in the other dimension (21), allowing resolution of thousands of proteins on a single gel. Although modern imaging and computing techniques can extract quantitative data for each of the spots in a 2D gel, there are only a few cases in which quantitative data have been gathered from 2D gels. 2D gel electrophoresis is almost unique in its ability to examine biological responses over thousands of proteins simultaneously and should therefore allow us a relatively comprehensive view of cellular metabolism.

We and others have worked toward assembling a yeast protein database consisting of a collection of identified spots in 2D gels and of data on each of these spots under various conditions (2, 7, 8, 10, 23, 25). These data could then be used in analyzing a protein or a metabolic process. *Saccharomyces cerevisiae* is a good organism for this approach since it has a well-understood physiology as well as a large number of mutants, and its genome has been sequenced. Given the sequence and the relative lack of introns in *S. cerevisiae*, it is easy to predict the sequence of the primary protein product of most genes. This aids tremendously in identifying these proteins on 2D gels.

There are three pillars on which such a database rests: (i) visualization of many protein spots simultaneously, (ii) quantification of the protein in each spot, and (iii) identification of the gene product for each spot. Our first efforts at visualization and identification for *S. cerevisiae* have been described elsewhere (7, 8). Here we describe quantitative data for these proteins under a variety of experimental conditions.

MATERIALS AND METHODS

Strains and media. *S. cerevisiae* W303 (MATa *ade2-1 his3-11,15 leu2-3,112 trp1-1 ura3-1 can1-100*) was used (26). –Met YNB (yeast nitrogen base) medium was 1.7 g of YNB (Difco) per liter, 5 g of ammonium sulfate per liter, and

adenine, uracil, and all amino acids except methionine; –Met. –Cys YNB medium was the same but without methionine or cysteine. Medium was supplemented with 2% glucose (for most experiments) or with 2% ethanol (for ethanol experiments). Low-phosphate YEPD was described by Warner (28).

Isotopic labeling of yeast and preparation of cell extracts. Yeast strains were labeled and proteins were extracted as described by Garrels et al. (7, 8). Briefly, cells were grown to 5×10^6 cells per ml. at 30°C; 1 ml of culture was transferred to a fresh tube, and 0.3 mCi of [³⁵S]methionine (e.g., Express protein labeling mix; New England Nuclear) was added to this 1-ml culture. The cells were incubated for a further 10 to 15 min and then transferred to a 1.5-ml microcentrifuge tube, chilled on ice, and harvested by centrifugation. The supernatant was removed, and the cell pellet was resuspended in 100 µl of lysis buffer (20 mM Tris-HCl [pH 7.6], 10 mM NaF, 10 mM sodium pyrophosphate, 0.5 mM EDTA, 0.1% deoxycholate; just before use, phenylmethylsulfonyl fluoride was added to 1 mM, leupeptin was added to 1 µg/ml, pepstatin was added to 1 µg/ml, tosyl-sulfonyl phenylalanyl chloromethyl ketone was added to 10 µg/ml, and soybean trypsin inhibitor was added to 10 µg/ml).

The resuspended cells were transferred to a screw-cap 1.5-ml polypropylene tube containing 0.28 g of glass beads (0.5-mm diameter; Biospec Products) or 0.40 g of zirconia beads (0.5-mm diameter; Biospec Products). After the cap was secured, the tube was inserted into a MiniBeadbeater 8 (Biospec Products) and shaken at medium high speed at 4°C for 1 min. Breakage was typically 75%. Tubes were then spun in a microcentrifuge for 10 s at 5,000 × g at 4°C.

With a very fine pipette tip, liquid was withdrawn from the beads and transferred to a prechilled 1.5-ml tube containing 7 µl of DNase I (0.5 mg/ml; Cooper product no. 6330)–RNase A (0.25 mg/ml; Cooper product no. 5679)–Mg (50 mM MgCl₂) mix. Typically 70 µl of liquid was recovered. The mixture was incubated on ice for 10 min to allow the RNase and DNase to work.

Next, 75 µl of 2× SDS (2× SDS is 0.6% sodium dodecyl sulfate [SDS], 2% mercaptoethanol, and 0.1 M Tris-HCl [pH 8]) was added. The tube was plunged into boiling water, incubated for 1 min, and then plunged into ice. After cooling, the tube was centrifuged at 4°C for 3 min at 14,000 × g. The supernatant was transferred to a fresh tube and frozen at –70°C. About 5 µl of this supernatant was used for each 2D gel.

2D polyacrylamide gels. 2D gels were made and run as described elsewhere (6–8).

Image analysis of the gels. The Quest II software system was used for quantitative image analysis (20, 22). Two techniques were used to collect quantitative data for analysis by Quest II software. First, before the advent of phosphorimagers, gels were dried and fluorographed. Each gel was exposed to film for three different times (typically 1 day, 2 weeks, and 6 weeks) to increase the dynamic range of the data. The films were scanned along with calibration strips to relate film optical density to disintegrations per minute in the gels and analyzed by the software to obtain a linear relationship between disintegrations per minute in the spots and optical densities of the film images. The quantitative data are expressed as parts per million of the total cellular protein. This value is calculated from the disintegrations per minute of the sample loaded onto the gel and by comparing the film density of each data spot with density of the film over the calibration strips of known radioactivity exposed to the same film. This yields the disintegrations per minute per millimeter for each spot on the gel and thence its parts-per-million value.

After the advent of phosphorimaging, gels bearing ³⁵S-labeled proteins were exposed to phosphorimager screens and scanned by a Fuji phosphorimager, typically for two exposures per gel. Calibration strips of known radioactivity were exposed simultaneously. Scan data from the phosphorimager was assimilated by Quest II software, and quantitative data were recorded for the spots on the gels.

* Corresponding author. Mailing address: Cold Spring Harbor Laboratory, Cold Spring Harbor, NY 11724. Phone: (516) 367-8828. Fax: (516) 367-8369. E-mail: fletcher@cschl.org.

Measurements of protein turnover. Cells in exponential phase were pulse-labeled with [35 S]methionine, excess cold Met and Cys were added, and samples of equal volume were taken from the culture at intervals up to 90 min (in one experiment) or up to 160 min (in a second experiment). Incorporation of 35 S into protein was essentially 100% by the first sample (10 min). Extracts were made, and equal fractions of the samples were loaded on 2D gels (i.e., the different samples had different amounts of protein but equal amounts of 35 S). Spots were quantitated with a phosphorimaging and Quest software.

The software was queried for spots whose radioactivity decreased through the time course. The algorithm examined all data points for all spots, drew a best-fit line through the data points, and looked for spots where this line had a statistically significant negative slope. In one of the experiments, there was one such spot. To the eye, this was a minor, unidentified spot seen only in the first two samples (10 and 20 min). In the other experiment, the Quest software found no spots meeting the criteria. Therefore, we concluded that none of the identified spots (and all but one of the visible spots) represented proteins with long half-lives.

Centrifugal fractionation. Cells were labeled, harvested, and broken with glass beads by the standard method described above except that no detergent (i.e., no deoxycholate) was present in the lysis buffer. The crude lysate was cleared of unbroken cells and large debris by centrifugation at $300 \times g$ for 30 s. The supernatant of this centrifugation was then spun at $16,000 \times g$ for 10 min to give the pellet used for Fig. 6B. The supernatant of the $16,000 \times g$, 10-min spin was then spun at $100,000 \times g$ for 30 min to give the supernatant used for Fig. 6A.

Protein abundance calculations. A haploid yeast cell contains about 4×10^{-12} g of protein (1, 15). Assuming a mean protein mass of 50 kDa, there are about 50×10^6 molecules of protein per cell. There are about 1.8 methionines per 10 kDa of protein mass, which implies 4.5×10^8 molecules of methionine per cell (neglecting the small pool of free Met). We measured (i) the counts per minute in each spot on the 2D gels, (ii) the total number of counts on each gel (by integrating counts over the entire gel), and (iii) the total number of counts loaded on the gel (by scintillation counting of the original sample). Thus, we know what fraction of the total incorporated radioactivity is present in each spot. After correcting for the methionine (and cysteine [see below]) content of each protein, we calculated an absolute number of protein molecules based on the fraction of radioactivity in each spot and on 50×10^6 total molecules per cell.

The labeling mixture used contained about one-fifth as much radioactive cysteine as radioactive methionine. Therefore, the number of cysteine molecules per protein was also taken into account in calculating the number of molecules of protein, but Cys molecules were weighted one-fifth as heavily as Met molecules.

mRNA abundance calculations. For estimation of mRNA abundance, we used SAGE (serial analysis of gene expression) data (27) and Affymetrix chip hybridization data (29a, 30). The mRNA column in Table 1 shows mRNA abundance calculated from SAGE data alone. However, the SAGE data came from cells growing in YEPD medium, whereas our protein measurements were from cells growing in YNB medium. In addition, SAGE data for low-abundance mRNAs suffers from statistical variation. Therefore, we also used chip hybridization data (29a, 30) for mRNA from cells grown in YNB. These hybridization data also had disadvantages. First, the amounts of high-abundance mRNAs were systematically underestimated, probably because of saturation in the hybridizations, which used 10 μ g of cRNA. For example, the abundance of *ADH1* mRNA was 197 copies per cell by SAGE but only 32 copies per cell by hybridization, and the abundance of *ENO2* mRNA was 248 copies per cell by SAGE but only 41 by hybridization. When the amount of cRNA used in the hybridization was reduced to 1 μ g, the apparent amounts of mRNA were similar to the amounts determined by SAGE (29a, 29b). However, experiments using 1 μ g of cRNA have been done for only some genes (29a). Because amounts of mRNA were normalized to 15,000 per cell, and because the amounts of abundant mRNAs were underestimated, there is a 2.2-fold overestimate of the abundance of nonabundant mRNAs. We calculated this factor of 2.2 by adding together the number of mRNA molecules from a large number of genes expressed at a low level for both SAGE data and hybridization data. The sum for the same genes from hybridization data is 2.2-fold greater than that from SAGE data.

To take into account these difficulties, we compiled a list of "adjusted" mRNA abundance as follows. For all high-abundance mRNAs of our identified proteins, we used SAGE data. For all of these particular mRNAs, chip hybridization suggested that mRNA abundance was the same in YEPD and YNB media. For medium-abundance mRNAs, SAGE data were used, but when hybridization data showed a significant difference between YEPD and YNB, then the SAGE data were adjusted by the appropriate factor. Finally, for low-abundance mRNAs, we used data from chip hybridizations from YNB medium but divided by 2.2 to normalize to the SAGE results. These calculations were completed without reference to protein abundance.

CAI. The codon adaptation index (CAI) was taken from the yeast proteome database (YPD) (13), for which calculations were made according to Sharp and Li (24). Briefly, the index uses a reference set of highly expressed genes to assign a value to each codon, and then a score for a gene is calculated from the frequency of use of the various codons in that gene (24).

Statistical analysis. The JMP program was used with the aid of T. Tully. The JMP program showed that neither mRNA nor protein abundances were normally distributed; therefore, Spearman rank correlation coefficients (r_s) were

calculated. The mRNA (adjusted and unadjusted) and protein data were also transformed so that Pearson product-moment correlation coefficients (r_p) could be calculated. First, this was done by a Box-Cox transformation of log-transformed data. This transformation produced normal distributions, and an r_p of 0.76 was achieved. However, because the Box-Cox transformation is complex, we also did a simpler logarithmic transformation. This produced a normal distribution for the protein data. However, the distribution for the mRNA and adjusted mRNA data was close to, but not quite, normal. Nevertheless, we calculated the r_p and found that it was 0.76, identical to the coefficient from the Box-Cox transformed data. We therefore believe that this correlation coefficient is not misleading, despite the fact that the log(mRNA) distribution is not quite normal.

RESULTS

Visualization of 1,400 spots on three gel systems. Yeast proteins have isoelectric points ranging from 3.1 to 12.8, and masses ranging from less than 10 kDa to 470 kDa. It is difficult to examine all proteins on a single kind of gel, because a gel with the needed range in pI and mass would give poor resolution of the thousands of spots in the central region of the gel. Therefore, we have used three gel systems: (i) pH "4 to 8" with 10% polyacrylamide; (ii) pH "3 to 10" with 10% polyacrylamide; and (iii) nonequilibrium with 15% polyacrylamide (7, 8). Each gel system allows good resolution of a subset of yeast proteins.

Figure 1 shows a pH 4–8, 10% polyacrylamide gel. The pH at the basic end of the isoelectric focusing gel cannot be maintained throughout focusing, and so the proteins resolved on such gels have isoelectric points between pH 4 and pH 6.7. For these pH 4–8 gels, we see 600 to 900 spots on the best gels after multiple exposures.

The pH 3–10 gels (not shown) extend the pI range somewhat beyond pH 7.5, allowing detection of several hundred additional spots. Finally, we use nonequilibrium gels with 15% acrylamide in the second dimension. These allow visualization of about 100 very basic proteins and about 170 small proteins (less than 20 kDa). In total, using all three gel systems, about 1,400 spots can be seen. These represent about 1,200 different proteins, which is about one-quarter to one-third of the proteins expressed under these conditions (27, 30). Here, we focus on the proteins seen on the pH 4–8 gels.

Although nearly all expressed proteins are present on these gels, the number seen is limited by a problem we call coverage. Since there are thousands of proteins on each gel, many proteins comigrate or nearly comigrate. When two proteins are resolved, but are close together, and one protein spot is much more intense than the other, a problem arises in visualizing the weaker spot: at long exposures when the weak signal is strong enough for detection, the signal from the strong spot spreads and covers the signal from the weaker spot. Thus, weak spots can be seen only when they are well separated from strong spots.

For a given gel, the number of detectable spots initially rises with exposure time. However, beyond an optimal exposure, the number of distinguishable spots begins to decrease, because signals from strong spots cover signals from nearby weak spots. At long exposures, the whole autoradiogram turns black. Thus, there is an optimum exposure yielding the maximum number of spots, and at this exposure the weakest spots are not seen.

Largely because of the problem of coverage, the proteins seen are strongly biased toward abundant proteins. All identified proteins have a CAI of 0.18 or more, and we have identified no transcription factors or protein kinases, which are nonabundant proteins. Thus, this technology is useful for examining protein synthesis, amino acid metabolism, and glycolysis but not for examining transcription, DNA replication, or the cell cycle.

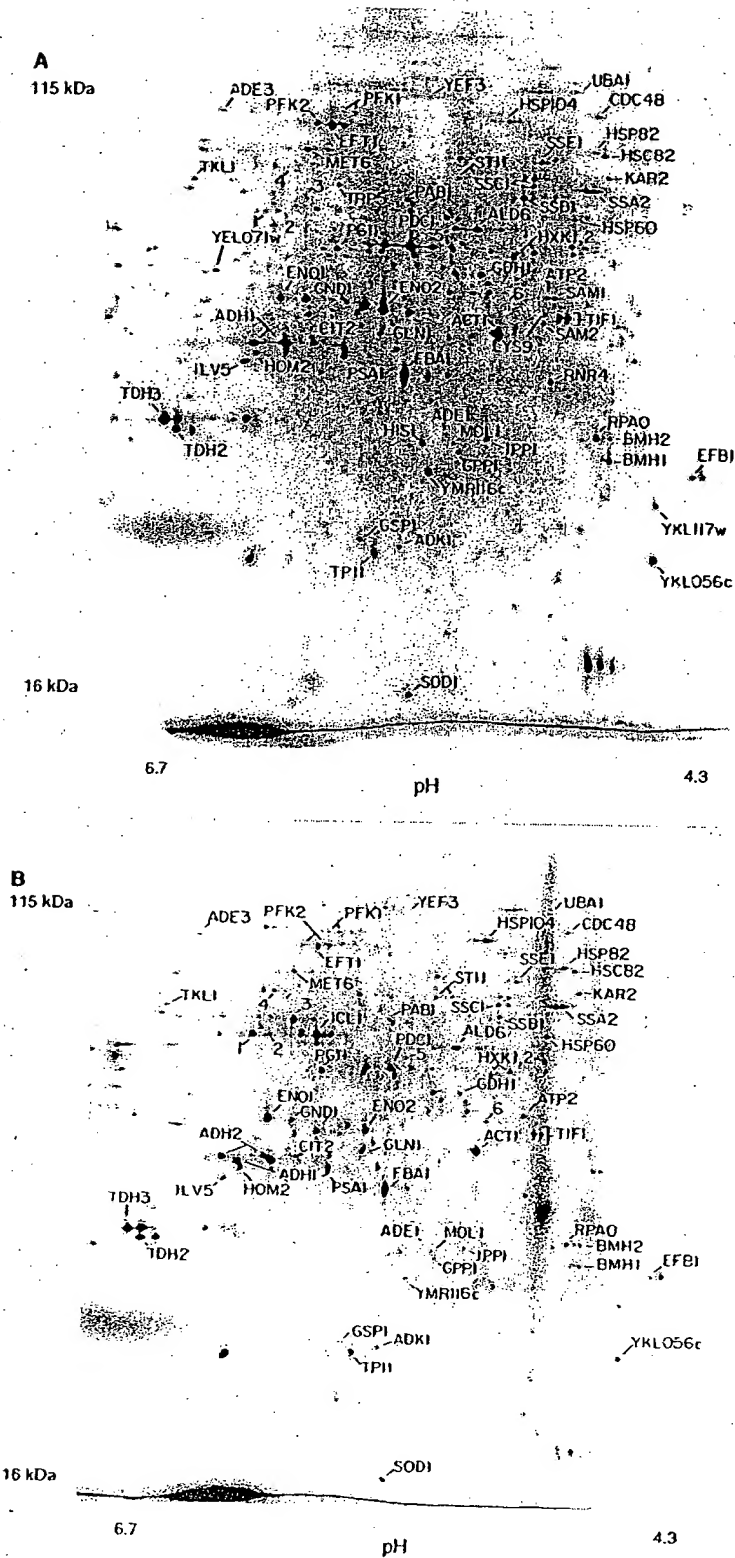


FIG. 1. 2D gels. The horizontal axis is the isoelectric focusing dimension, which stretches from pH 6.7 (left) to pH 4.3 (right). The vertical axis is the polyacrylamide gel dimension, which stretches from about 15 kDa (bottom) to at least 130 kDa (top). For panel A, extract was made from cells in log phase in glucose; for panel B, cells were grown in ethanol. The spots labeled 1 through 6 are unidentified proteins highly induced in ethanol.

Spot identification. The identification of various spots has been described elsewhere (7, 8). At present, 169 different spots representing 148 proteins have been identified. Many of these spots have been independently identified (2, 10, 23, 25). The main methods used in spot identification have been analysis of amino acid composition, gene overexpression, peptide sequencing, and mass spectrometry.

Pulse-chase experiments and protein turnover. Pulse-chase experiments were done to measure protein half-lives (Materials and Methods). Cells were labeled with [35 S]methionine for 10 min, and then an excess of unlabeled methionine was added. Samples were taken at 0, 10, 20, 30, 60, and 90 min after the beginning of the chase. Equal amounts of 35 S were loaded from each sample; 2D gels were run, and spots were quantitated. Surprisingly, almost every spot was nearly constant in amount of radioactivity over the entire time course (not shown). A few spots shifted from one position to another because of post-translational modifications (e.g., phosphorylation of Rpa0 and Efb1). Thus, the proteins being visualized are all or nearly all very stable proteins, with half-lives of more than 90 min. Gygi et al. (10) have come to a similar conclusion by using the N-end rule to predict protein half-lives. This result does not imply that all yeast proteins are stable. The proteins being visualized are abundant proteins; this is partly because they are stable proteins.

Protein quantitation. Because all of the proteins seen had effectively the same half-life, the abundance of each protein was directly proportional to the amount of radioactivity incorporated during labeling. Thus, after taking into account the total number of protein molecules per cell, the average content of methionine and cysteine, and the methionine and cysteine content of each identified protein, we could calculate the abundance of each identified protein (Tables 1 and 2; Materials and Methods). About 1,000 unidentified proteins were also quantified, assuming an average content of Met and Cys.

Many proteins give multiple spots (7, 8). The contribution from each spot was summed to give the total protein amount. However, many proteins probably have minor spots that we are not aware of, causing the amount of protein to be underestimated.

When the proteins on a pH 4–8 gel were ordered by abundance, the most abundant protein had 8,904 ppm, the 10th most abundant had 2,842 ppm, the 100th most abundant had 314 ppm, the 500th most abundant had 57 ppm, and the 1,000th most abundant (visualized at greater than optimum exposure) had 23 ppm. Thus, there is more than a 300-fold range in abundance among the visualized proteins. The most abundant 10 proteins account for about 25% of the total protein on the pH 4–8 gel, the most abundant 60 proteins account for 50%, and the most abundant 500 proteins account for 80%. Since it seems likely that the pH 4–8 gels give a representative sampling of all proteins, we estimate that half of the total cellular protein is accounted for by fewer than 100 different gene products, principally glycolytic enzymes and proteins involved in protein synthesis.

Correlation of protein abundance with mRNA abundance. Estimates of mRNA abundance for each gene have been made by SAGE (27) and by hybridization of cRNA to oligonucleotide arrays (30). These two methods give broadly similar results, yet each method has strengths and weaknesses (Materials and Methods). Table 1 lists the number of molecules of mRNA per cell for each gene studied. One measurement (mRNA) uses data from SAGE analysis alone (27); a second incorporates data from both SAGE and hybridization (30) (adjusted mRNA) (Table 1; Materials and Methods). We correlated protein abundance with mRNA abundance (Fig. 2). For ad-

justed mRNA versus protein, the Spearman rank correlation coefficient, r_s , was 0.74 ($P < 0.0001$), and the Pearson correlation coefficient, r_p , on log transformed data (Materials and Methods) was 0.76 ($P < 0.00001$). We obtained similar correlations for mRNA versus protein and also for other data transformations (Materials and Methods). Thus, several statistical methods show a strong and significant correlation between mRNA abundance and protein abundance. Of course, the correlation is far from perfect; for mRNAs of a given abundance, there is at least a 10-fold range of protein abundance (Fig. 2). Some of this scatter is probably due to posttranscriptional regulation, and some is due to errors in the mRNA or protein data. For example, the protein Yef3 runs poorly on our gels, giving multiple smeared spots. Its abundance has probably been underestimated, partly explaining the low protein/mRNA ratio of Yef3. It is the most extreme outlier in Fig. 2.

These data on mRNA (27, 30) and protein abundance (Table 1) suggest that for each mRNA molecule, there are on average 4,000 molecules of the cognate protein. For instance, for Act1 (actin) there are about 54 molecules of mRNA per cell and about 205,000 molecules of protein. Assuming an mRNA half-life of 30 min (12) and a cell doubling time of 120 min, this suggests that an individual molecule of mRNA might be translated roughly 1,000 times. These calculations are limited to mRNAs for abundant proteins, which are likely to be the mRNAs that are translated best.

A full complement of cell protein is synthesized in about 120 min under these conditions. Thus, 4,000 molecules of protein per molecule of mRNA implies that translation initiates on an mRNA about once every 2 s. This is a remarkably high rate; it implies that if an average mRNA bears 10 ribosomes engaged in translation, then each ribosome completes translation in 20 s; if an average protein has 450 residues; this in turn implies translation of over 20 amino acids per s, a rate considerably higher than estimated for mammals (3 to 8 amino acids per s) (18). These estimates depend on the amount of mRNA per cell (11, 27).

The large number of protein molecules that can be made from a single mRNA raises the issue of how abundance is controlled for less abundant proteins. Many nonabundant proteins may be unstable, and this would reduce the protein/mRNA ratio. In addition, many nonabundant proteins may be translated at suboptimal rates. We have found that mRNAs for nonabundant proteins usually have suboptimal contexts for translational initiation. For example, there are over 600 yeast genes which probably have short open reading frames in the mRNA upstream of the main open reading frame (17a). These may be devices for reducing the amount of protein made from a molecule of mRNA.

Correlation of codon bias with protein abundance. The mRNAs for highly expressed proteins preferentially use some codons rather than others specifying the same amino acid (14). This preference is called codon bias. The codons preferred are those for which the tRNAs are present in the greatest amounts. Use of these codons may make translation faster or more efficient and may decrease misincorporation. These effects are most important for the cell for abundant proteins, and so codon bias is most extreme for abundant proteins. The effect can be dramatic—highly biased mRNAs may use only 25 of the 61 codons.

We asked whether the correlation of codon bias with abundance continues for medium-abundance proteins. There are various mathematical expressions quantifying codon bias; here, we have used the CAI (24) (Materials and Methods) because it gives a result between 0 and 1. The r_s for CAI versus protein abundance is 0.80 ($P < 0.0001$), similar to the mRNA-protein

TABLE 1. Quantitative data*

| Function | Name | CAI | mRNA | Adjusted mRNA | Protein (Glu) (10 ³) | Protein (Eth) (10 ³) | E/G ratio |
|-------------------------|--------|-------|---------------|---------------|----------------------------------|----------------------------------|-----------|
| Carbohydrate metabolism | Adh1 | 0.810 | 197 | 197 | 1,230 | 972 | 0.79 |
| | Adh2 | 0.504 | 0 | | 0 | 963 | >20 |
| | Cit2 | 0.185 | 1 | 2.8 | 23 | 288 | 12 |
| | Eno1 | 0.870 | No <i>Nla</i> | | 410 | 974 | 2.4 |
| | Eno2 | 0.892 | 248 | 248 | 650 | 215 | 0.33 |
| | Fba1 | 0.868 | 179 | 179 | 640 | 608 | 0.95 |
| | Hxk1,2 | 0.500 | 13 | 10.5 | 62 | 46 | |
| | Icl1 | 0.251 | 0 | | 0 | 671 | >20 |
| | Pdb1 | 0.342 | 5 | 5 | 41 | 33 | |
| | Pdc1 | 0.903 | 226 | 226 | 280 | 205 | 0.73 |
| | Pfk1 | 0.465 | 5 | 5 | 75 | 53 | 0.71 |
| | Pgi1 | 0.681 | 14 | 14 | 160 | 120 | 0.75 |
| | Pyc1 | 0.260 | 1 | 0.7 | 37 | 34 | |
| | Tal1 | 0.579 | 5 | 5 | 110 | 35 | |
| | Tdh2 | 0.904 | 63 | 63 | 430 | 876 | NR |
| | Tdh3 | 0.924 | 460 | 460 | 1,670 | 1,927 | NR |
| | Tpi1 | 0.817 | No <i>Nla</i> | | No Met | No Met | |
| Protein synthesis | Efb1 | 0.762 | 33 | 16.5 | 358 | 362 | |
| | Eft1,2 | 0.801 | 26 | 26 | 99 | 54 | 0.55 |
| | Prt1 | 0.303 | 4 | 0.7 | 12 | 6 | |
| | Rpa0 | 0.793 | 246 | 246 | 277 | 100 | 0.36 |
| | Tif1,2 | 0.752 | 29 | 29 | 233 | 106 | 0.46 |
| | Yef3 | 0.777 | 36 | 36 | 14 | ND | |
| Heat shock | Hsc82 | 0.581 | 2 | 2.9 | 112 | 75 | 0.67 |
| | Hsp60 | 0.381 | 9 | 2.3 | 35 | 82 | 2.3 |
| | Hsp82 | 0.517 | 2 | 1.3 | 52 | 135 | 2.6 |
| | Hsp104 | 0.304 | 7 | 7 | 70 | 161 | 2.3 |
| | Kar2 | 0.439 | 5 | 10.1 | 43 | 102 | 2.4 |
| | Ssa1 | 0.709 | 2 | 4.3 | 303 | 421 | 1.4 |
| | Ssa2 | 0.802 | 10 | 5 | 213 | 324 | 1.5 |
| | Ssb1,2 | 0.850 | 50 | 50 | 270 | 85 | |
| | Ssc1 | 0.521 | 2 | 2.6 | 68 | 80 | 1.2 |
| | Sse1 | 0.521 | 8 | 8 | 96 | 48 | |
| | Sti1 | 0.247 | 1 | 1.1 | 25 | 44 | 1.7 |
| Amino acid synthesis | Ade1 | 0.229 | 4 | 4 | 14 | 27 | |
| | Ade3 | 0.276 | 2 | 1.7 | 12 | 9 | |
| | Ade5,7 | 0.257 | 2 | 1.4 | 14 | 4 | |
| | Arg4 | 0.229 | 1 | 8.1 | 41 | 41 | |
| | Gdh1 | 0.585 | 10 | 27 | 148 | 55 | |
| | Gln1 | 0.524 | 11 | 11 | 77 | 104 | 1.3 |
| | His4 | 0.267 | 3 | 3 | 15 | 23 | 1.5 |
| | Ilv5 | 0.801 | 6 | 6 | 152 | 109 | 0.7 |
| | Lys9 | 0.332 | 4 | 4 | 32 | 17 | 0.52 |
| | Met6 | 0.657 | No <i>Nla</i> | 22 | 190 | 80 | 0.42 |
| | Pro2 | 0.248 | 3 | 3 | 30 | 12 | |
| | Ser1 | 0.258 | 2 | 1.2 | 15 | 8 | |
| | Trp5 | 0.319 | 5 | 5 | 28 | 12 | |
| Miscellaneous | Act1 | 0.710 | 54 | 54 | 205 | 164 | 0.78 |
| | Adk1 | 0.531 | No <i>Nla</i> | | 47 | 43 | |
| | Ald6 | 0.520 | 3 | 3 | 181 | 159 | |
| | Atp2 | 0.424 | 1 | 4.1 | 76 | 109 | 1.4 |
| | Bmh1 | 0.322 | 46 | 46 | 191 | 137 | 0.72 |
| | Bmh2 | 0.384 | 1 | 1.4 | 134 | 147 | |
| | Cdc43 | 0.306 | 2 | 2.4 | 32 | 26 | |
| | Cdc60 | 0.299 | 2 | 0.86 | 6 | 2 | |
| | Erg20 | 0.373 | 5 | 5 | 92 | 39 | |
| | Gpp1 | 0.603 | 16 | 5 | 234 | 158 | |
| | Gsp1 | 0.621 | 3 | 3 | 115 | 39 | |
| | Ipp1 | 0.620 | 4 | 4 | 254 | 147 | 0.34 |
| | Lcb1 | 0.173 | 0.3 | 0.8 | 19 | 40 | 0.58 |
| | Mof1 | 0.423 | 0 | 0.45 | 20 | 16 | |
| | Pab1 | 0.488 | 3 | 3 | 41 | 19 | |
| | Psa1 | 0.600 | 15 | 15 | 148 | 56 | 0.47 |
| | Rnr4 | 0.497 | 6 | 6 | 44 | 37 | |
| | Sam1 | 0.494 | 5 | 5 | 59 | 21 | |
| | Sam2 | 0.497 | 3 | 15 | 63 | 20 | |
| | Sod1 | 0.376 | 36 | 36 | 631 | 618 | |
| | Uba1 | 0.212 | 2 | 2 | 14 | 20 | |
| | YKL056 | 0.731 | 62 | 62 | 253 | 112 | 0.44 |
| | YLR109 | 0.549 | 21 | 21 | 930 | | |
| | YMR116 | 0.777 | 41 | 41 | 184 | 40 | 0.20 |

* CAI, a measure of codon bias, is taken from the YPD. mRNA, number of mRNA molecules per cell from SAGE data (27); adjusted mRNA, number of mRNA molecules per cell based on both SAGE and chip hybridization (30) (see Materials and Methods); Protein (Glu), number of molecules of protein per cell in YNB-glucose; Protein (Eth), number of molecules of protein per cell in YNB-ethanol; E/G ratio, ratio of protein abundance in ethanol to glucose. The E/G ratio is not given if it was close to 1 or if it was not repeatable (NR) in multiple gels. Some gene products (e.g., Tif1 and Tif2 [Tif1,2]) were difficult to distinguish on either a protein or an mRNA basis; these are pooled. No *Nla*, there was no suitable *Nla*III site in the 3' region of the gene, and so there are no SAGE mRNA data; No Met, the mature gene product contains no methionines, and so there are no reliable protein data.

TABLE 2. Functions of proteins listed in Table 1

| Name ^a | YPD title lines ^b |
|-------------------|--|
| Adh1 | Alcohol dehydrogenase I; cytoplasmic isozyme reducing acetaldehyde to ethanol, regenerating NAD ⁺ |
| Adh2 | Alcohol dehydrogenase II; oxidizes ethanol to acetaldehyde, glucose repressed |
| Cit2 | Citrate synthase, peroxisomal (nonmitochondrial); converts acetyl-CoA and oxaloacetate to citrate plus CoA |
| Eno1 | Enolase 1 (2-phosphoglycerate dehydratase); converts 2-phospho-D-glycerate to phosphoenolpyruvate in glycolysis |
| Eno2 | Enolase 2 (2-phosphoglycerate dehydratase); converts 2-phospho-D-glycerate to phosphoenolpyruvate in glycolysis |
| Fba1 | Fructose biphosphate aldolase II; sixth step in glycolysis |
| Hxk1 | Hexokinase I; converts hexoses to hexose phosphates in glycolysis; repressed by glucose |
| Hxk2 | Hexokinase II; converts hexoses to hexose phosphates in glycolysis and plays a regulatory role in glucose repression |
| Icl1 | Isocitrate lyase, peroxisomal; carries out part of the glyoxylate cycle; required for gluconeogenesis |
| Pdb1 | Pyruvate dehydrogenase complex, E1 beta subunit |
| Pdc1 | Pyruvate decarboxylase isozyme 1 |
| Pfk1 | Phosphofructokinase alpha subunit, part of a complex with Pfk2p which carries out a key regulatory step in glycolysis |
| Pgi1 | Glucose-6-phosphate isomerase, converts glucose-6-phosphate to fructose-6-phosphate |
| Pyc1 | Pyruvate carboxylase 1; converts pyruvate to oxaloacetate for gluconeogenesis |
| Tal1 | Transaldolase; component of nonoxidative part of pentose phosphate pathway |
| Tdh2 | Glyceraldehyde-3-phosphate dehydrogenase 2; converts D-glyceraldehyde 3-phosphate to 1,3-dephosphoglycerate |
| Tdh3 | Glyceraldehyde-3-phosphate dehydrogenase 3; converts D-glyceraldehyde 3-phosphate to 1,3-dephosphoglycerate |
| Tpi1 | Triosephosphate isomerase; interconverts glyceraldehyde-3-phosphate and dihydroxyacetone phosphate |
| Efb1 | Translation elongation factor EF-1 β ; GDP/GTP exchange factor for Tef1p/Tef2p |
| Eft1 | Translation elongation factor EF-2; contains diphthamide which is not essential for activity; identical to Eft2p |
| Eft2 | Translation elongation factor EF-2; contains diphthamide which is not essential for activity; identical to Eft1p |
| Prt1 | Translation initiation factor eIF3 beta subunit (p90); has an RNA recognition domain |
| Rpa0 (RPPO) | Acidic ribosomal protein A0 |
| Tif1 | Translation initiation factor 4A (eIF4A) of the DEAD box family |
| Tif2 | Translation initiation factor 4A (eIF4A) of the DEAD box family |
| Yef3 | Translation elongation factor EF-3A; member of ATP-binding cassette superfamily |
| Hsc82 | Chaperonin homologous to <i>E. coli</i> HtpG and mammalian HSP90 |
| Hsp60 | Mitochondrial chaperonin that cooperates with Hsp10p; homolog of <i>E. coli</i> GroEL |
| Hsp82 | Heat-inducible chaperonin homologous to <i>E. coli</i> HtpG and mammalian HSP90 |
| Hsp104 | Heat shock protein required for induced thermotolerance and for resolubilizing aggregates of denatured proteins; important for [psi ⁻]-to-[psi ⁺] prion conversion |
| Kar2 | Heat shock protein of the endoplasmic reticulum lumen required for protein translocation across the endoplasmic reticulum membrane and for nuclear fusion; member of the HSP70 family |
| Ssa1 | Cytoplasmic chaperone; heat shock protein of the HSP70 family |
| Ssa2 | Cytoplasmic chaperone; member of the HSP70 family |
| Ssb1 | Heat shock protein of HSP70 family involved in the translational apparatus |
| Ssb2 | Heat shock protein of HSP70 family, cytoplasmic |
| Ssc1 | Mitochondrial protein that acts as an import motor with Tim44p and plays a chaperonin role in receiving and folding of protein chains during import; heat shock protein of HSP70 family |
| Sse1 | Heat shock protein of the HSP70 family; multicopy suppressor of mutants with hyperactivated Ras/cyclic AMP pathway |
| Sti1 | Stress-induced protein required for optimal growth at high and low temperature; has tetratricopeptide repeats |
| Ade1 | Phosphoribosylamidoimidazole-succinocarboxamide synthase; catalyzes the seventh step in de novo purine biosynthesis pathway |
| Ade3 | C ₄ tetrahydrofolate synthase (trifunctional enzyme), cytoplasmic |
| Ade5,7 | Phosphoribosylamine-glycine ligase plus phosphoribosylformylglycinamide cyclo-ligase; bifunctional protein |
| Arg4 | Argininosuccinate lyase; catalyzes the final step in arginine biosynthesis |
| Gdh1 | Glutamate dehydrogenase (NADP ⁺); combines ammonia and α -ketoglutarate to form glutamate |
| Gln1 | Glutamine synthetase; combines ammonia to glutamate in ATP-driven reaction |
| His4 | Phosphoribosyl-AMP cyclohydrolase/phosphoribosyl-ATP pyrophosphohydrolase/histidinol dehydrogenase; 2nd, 3rd, and 10th steps of his biosynthesis pathway |
| Ily5 | Ketol-acid reductoisomerase (acetohydroxy, acid reductoisomerase) (alpha-keto- β -hydroxyacyl) reductoisomerase; second step in Val and Ily biosynthesis pathway |
| Lys9 | Saccharopine dehydrogenase (NADP ⁺ , L-glutamate forming) (saccharopine reductase), seventh step in lysine biosynthesis pathway |
| Met6 | Homocysteine methyltransferase; (5-methyltetrahydropteroyl triglutamate-homocysteine methyltransferase), methionine synthase, cobalamin independent |
| Pro2 | γ -Glutamyl phosphate reductase (phosphoglutamate dehydrogenase), proline biosynthetic enzyme |
| Ser1 | Phosphoserine transaminase; involved in synthesis of serine from 3-phosphoglycerate |
| Trp5 | Tryptophan synthase, fast (5th) step in tryptophan biosynthesis pathway |
| Act1 | Actin; involved in cell polarization, endocytosis, and other cytoskeletal functions |
| Adk1 | Adenylate kinase (GTP:AMP phosphotransferase), cytoplasmic |
| Ald6 | Cytosolic acetaldehyde dehydrogenase |
| Atp2 | Beta subunit of F1-ATP synthase; 3 copies are found in each F1 oligomer |
| Bmh1 | Homolog of mammalian 14-3-3 protein; has strong similarity to Bmh2p |
| Bmh2 | Homolog of mammalian 14-3-3 protein; has strong similarity to Bmh1p |
| Cdc48 | Protein of the AAA family of ATPases; required for cell division and homotypic membrane fusion |
| Cdc60 | Leucyl-tRNA synthetase, cytoplasmic |
| Erg20 | Farnesyl pyrophosphate synthetase; may be rate-limiting step in sterol biosynthesis pathway |
| Gpp1 (Rhr2) | DL-Glycerol phosphate phosphatase |
| Gsp1 | Ran, a GTP-binding protein of the Ras superfamily involved in trafficking through nuclear pores |
| Ipp1 | Inorganic pyrophosphatase, cytoplasmic |
| Lcb1 | Component of serine C-palmitoyltransferase; first step in biosynthesis of long-chain base component of sphingolipids |
| Mol1 (Thi4) | Thiamine-repressed protein essential for growth in the absence of thiamine |
| Pab1 | Poly(A)-binding protein of cytoplasm and nucleus; part of the 3'-end RNA-processing complex (cleavage factor I); has 4 RNA recognition domains |
| Psa1 | Mannose-1-phosphate guanylttransferase; GDP-mannose pyrophosphorylase |
| Rnr4 | Ribonucleotide reductase small subunit |
| Sam1 | S-Adenosylmethionine synthetase 1 |
| Sam2 | S-Adenosylmethionine synthetase 2 |
| Sod1 | Copper-zinc superoxide dismutase |
| Uba1 | Ubiquitin-activating (E1) enzyme |
| YKL056 | Resembles translationally controlled tumor protein of animal cells and higher plants |
| YLR109 (Ahp1) | Alkyl hydroperoxide reductase |
| YMR116 (Asc1) | Abundant protein with effects on translational efficiency and cell size, has two WD (WD-40) repeats |

^a Accepted name from the *Saccharomyces* genome database and YPD. Names in parentheses represent recent changes.

^b Courtesy of Proteome, Inc., reprinted with permission.

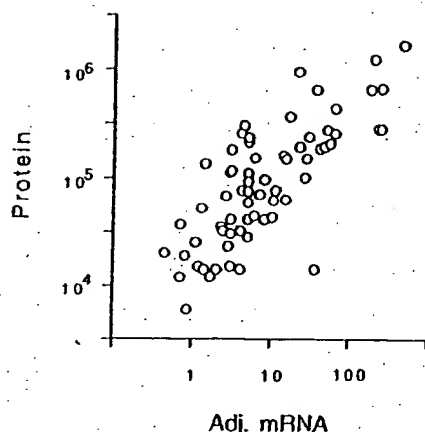


FIG. 2. Correlation of protein abundance with adjusted mRNA abundance. The number of molecules per cell of each protein is plotted against the number of molecules per cell of the cognate mRNA, with an r_p of 0.76. Note the logarithmic axes. Data for mRNA were taken from references 27 and 30 and combined as described in Materials and Methods.

correlation, confirming a strong correlation between CAI and protein abundance (Fig. 3). The relationship between CAI and protein abundance is log linear from about 1,000,000 to about 10,000 molecules per cell. We have no data for rarer proteins.

It is not clear whether CAI reflects maximum or average levels of protein expression. The proteins used for the CAI-protein correlation included some proteins which were not expressed at maximum levels under the condition of the experiment (Hsc82, Hsp104, Ssa1, Ade1, Arg4, His4, and others). When these proteins were removed from consideration and the correlation between CAI and the remaining (presumably constitutive) proteins was recalculated, the r_p was essentially unchanged (not shown).

The equation describing the graph in Fig. 3 is $\log(\text{protein molecules/cell}) = (2.3 \times \text{CAI}) + 3.7$. Thus, under certain conditions (a CAI of 0.3 or greater; a constitutively expressed gene), a very rough estimate of protein abundance can be made by raising 10 to the power of $[(2.3 \times \text{CAI}) + 3.7]$.

The distribution of CAI over the genome (Fig. 4) consists of a lower, bell-shaped distribution, possibly indicating a region where there is no selection for codon bias, and an upper, flat distribution, starting at a CAI of about 0.3, possibly indicating a region where there is selection for codon bias. Almost all of the proteins whose abundance we have measured are in the upper, flat portion of the distribution. In the lower, bell-shaped region, we do not know whether there is a correlation between CAI and protein abundance.

Changes in protein abundance in glucose and ethanol. A comparison of cells grown in glucose (Fig. 1A) with cells grown in ethanol (Fig. 1B) is shown in Table 1. As is well known, some proteins are induced tremendously during growth on ethanol. Two striking examples are the peroxisomal enzymes Icl1 (isocitrate lyase) and Cit2 (citrate synthase), which are induced in ethanol by more than 100- and 12-fold, respectively (Fig. 1; Table 1). These enzymes are key components of the glyoxylate shunt, which diverts some acetyl coenzyme A (acetyl-CoA) from the tricarboxylic acid cycle to gluconeogenesis. *S. cerevisiae* requires large amounts of carbohydrate for its cell wall; in ethanol medium, this carbohydrate comes from gluconeogenesis, which depends on the glyoxylate shunt and on the glycolytic pathway running in reverse. The need for

gluconeogenesis also explains why glycolytic enzymes are abundant even in ethanol medium. Thus, 2D gel analysis shows the prominence of the glycolytic and glyoxylate shunt enzymes in cells grown on ethanol, emphasizing that gluconeogenesis, presumably largely for production of the cell wall, is a major metabolic activity under these conditions.

During gluconeogenesis, substrate-product relationships are reversed for the glycolytic enzymes. One might expect that not all glycolytic enzymes would be well adapted to the reverse reaction. Indeed, 2D gels show that in ethanol, Adh2 (alcohol dehydrogenase 2) is strongly induced (16), while its isozyme Adh1 is not greatly affected. Adh1 and Adh2 each interconvert acetaldehyde and ethanol. Adh1 has a relatively high K_m for ethanol (17 mM), while Adh2 has a lower K_m (0.8 mM) (5). Thus, it is thought that Adh1 is specialized for glycolysis (acetaldehyde to ethanol), while Adh2 is specialized for respiration (ethanol to acetaldehyde) (5, 29). Similarly, Eno1 (enolase 1) is induced in ethanol, while its isozyme Eno2 (enolase 2) decreases in abundance (Table 1) (4, 19). Eno1 is inhibited by 2-phosphoglycerate (the glycolytic substrate), while Eno2 is inhibited by phosphoenolpyruvate (the gluconeogenic substrate) (4). Perhaps Eno1 has a lower K_m for phosphoenolpyruvate than does Eno2, though to our knowledge this has not been tested. Thus, the 2D gels distinguish isozymes specialized for growth on glucose (Adh1 and Eno2) from isozymes specialized for ethanol (Adh2 and Eno1).

Many heat shock proteins (e.g., Hsp60, Hsp82, Hsp104, and Kar2) were about twofold more abundant in ethanol medium than in glucose medium. This is consistent with the increased heat resistance of cells grown in ethanol (3).

Enzymes involved in protein synthesis (Eft1, Rpa0, and Tif1) were about twice as abundant in glucose medium as in ethanol medium. This may reflect the higher growth rate of the cells in glucose.

Phosphorylation of proteins. To examine protein phosphorylation, we labeled cells with ^{32}P and ran 2D gels to examine phosphoproteins. About 300 distinct spots, probably representing 150 to 200 proteins, could be seen on pH 4–8 gels (Fig. 5B). We then aligned autoradiograms of three gels, each with a different kind of labeled protein (^{32}P only [Fig. 5B], ^{32}P plus ^{35}S [Fig. 5A], and ^{35}S only [not shown, but see Fig. 1 for example]). In this way, we made provisional identification of

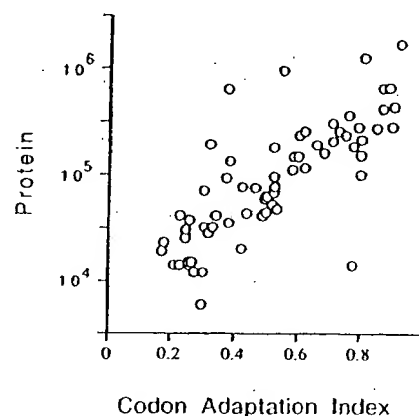


FIG. 3. Correlation of protein abundance with CAI. The number of molecules per cell of each protein is plotted against the CAI for that protein. Note the logarithmic scale on the protein axis. Data for the CAI are from the YPD database (13).

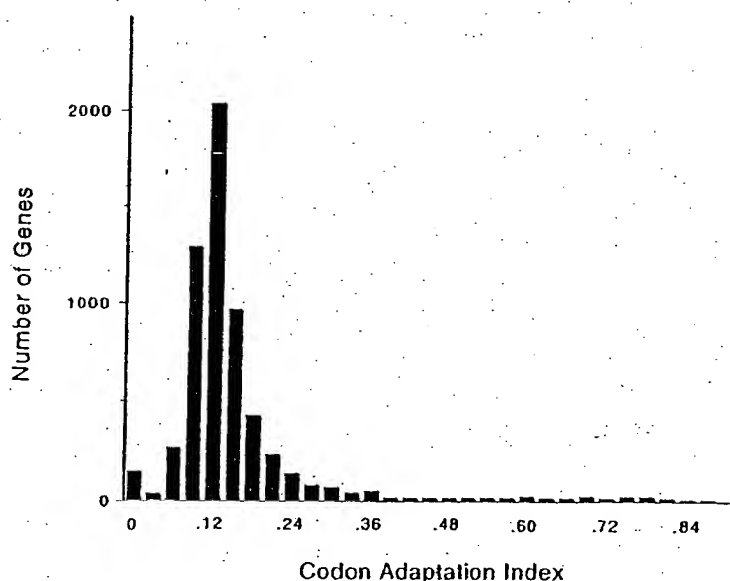


FIG. 4. Distribution of CAI over the whole genome, shown in intervals of 0.030 (i.e., there are 150 genes with a CAI between 0.000 and 0.030, inclusive; 31 genes with a CAI between 0.031 and 0.060; 269 genes with a CAI between 0.061 and 0.090; 1,296 genes with a CAI between 0.091 and 0.120; etc.). The distribution peaks with 2,028 genes with a CAI between 0.121 and 0.150.

some of the ^{32}P -labeled spots as particular ^{35}S -labeled spots. All such identifications are somewhat uncertain, since precise alignments are difficult, and of course multiple spots may exactly comigrate. Nevertheless, we believe that most of the provisional identifications are probably correct. Among the major ^{32}P -labeled proteins are the hexokinases Hxk1 and Hxk2, the acidic ribosome-associated protein Rpa0, the translation factors Yef3 and Efb1, and probably Hsp70 heat shock proteins of the Ssa and Ssb families. Rpa0 and Efb1 are quantitatively monophosphorylated.

Many yeast proteins resolve into multiple spots on these 2D gels (7). Yef3 has five or more spots, at least four of which comigrate with ^{32}P . Tpi1 has a major spot showing no ^{32}P labeling and a minor, more acidic spot which overlaps with some ^{32}P label. Tif1 has at least seven spots (7); two of these overlap with some ^{32}P label, but five do not (Fig. 5). Eft1 has at least three spots (7), and none of these overlap with ^{32}P , although there are three nearby, unidentified ^{32}P -labeled spots (a, c, and d in Fig. 5). Spots that seem to be extra forms of Met6, Pdc1, Eno2, and Fba1 can be seen in Fig. 6A, but there is little ^{32}P at these positions in Fig. 5. Thus, phosphorylation explains some but not all of the different protein isoforms seen.

The cell cycle is regulated in part by phosphorylation. We compared ^{32}P -labeled proteins from cells synchronized in G_1 with α -factor, in cells synchronized in G_1 by depletion of G_1 cyclins, and in cells synchronized in M phase with nocodazole. Only very minor differences were seen, and these were difficult to reproduce. The cell cycle proteins regulated by phosphorylation may not be abundant enough for this technique to be applied easily.

Centrifugal fractionation. We fractionated ^{35}S -labeled extracts by centrifugation (Materials and Methods). Figure 6A shows the proteins in the supernatant of a high-speed ($100,000 \times g$, 30 min) centrifugation, while Fig. 6B shows the proteins in the pellet of a low-speed ($16,000 \times g$, 10 min) centrifugation. Many proteins are tremendously enriched in one fraction or the other, while others are present in both.

Most glycolytic enzymes (e.g., Tdh2, Tdh3, Eno2, Pdc1, Adh1, and Fba1) are enriched in the supernatant fraction. The only exception is Pfk1 (not indicated), which is found in both pellet and supernatant fractions. Many proteins involved in protein synthesis (Eft1, Yef3, Prt1, Tif1, and Rpa0) are in the pellet, possibly because of the association of ribosomes with the endoplasmic reticulum. However, Efb1 is in the supernatant, as is a substantial portion of the Eft1. Perhaps surprisingly, several mitochondrial proteins (Atp2 [not shown] and Ily5) are largely in the supernatant. Perhaps glass bead breakage of cells releases mitochondrial proteins. The nuclear protein Gsp1 is in the pellet fraction. The enrichment produced by centrifugation makes it possible to see minor spots which are otherwise poorly resolved from surrounding proteins. Figure 6B shows that the previously identified Tif1 spot is surrounded by as many as six other spots that cofractionate. We observed six identical or very similar additional spots when we overexpressed Tif1 from a high-copy-number plasmid (not shown). Signal overlaps only one or two of these spots in ^{32}P -labeling experiments (Fig. 5), and so the different forms are not mainly due to different phosphorylation states.

DISCUSSION

Our experience with developing a 2D gel protein database for *S. cerevisiae* is summarized here. With current technology, we can see the most abundant 1,200 proteins, which is about one-third to one-quarter of the proteins expressed. The remaining proteins will be difficult to see and study with the methods that we have used, not because of a lack of sensitivity but because weak spots are covered by nearby strong spots.

Of the 1,200 proteins seen, we have identified 148, with a bias toward the most abundant proteins. Steady application of the methods already used would allow identification of most of the remaining proteins. Gene overexpression will be particularly useful, since it is not affected by the lower abundance of the remaining visible proteins.

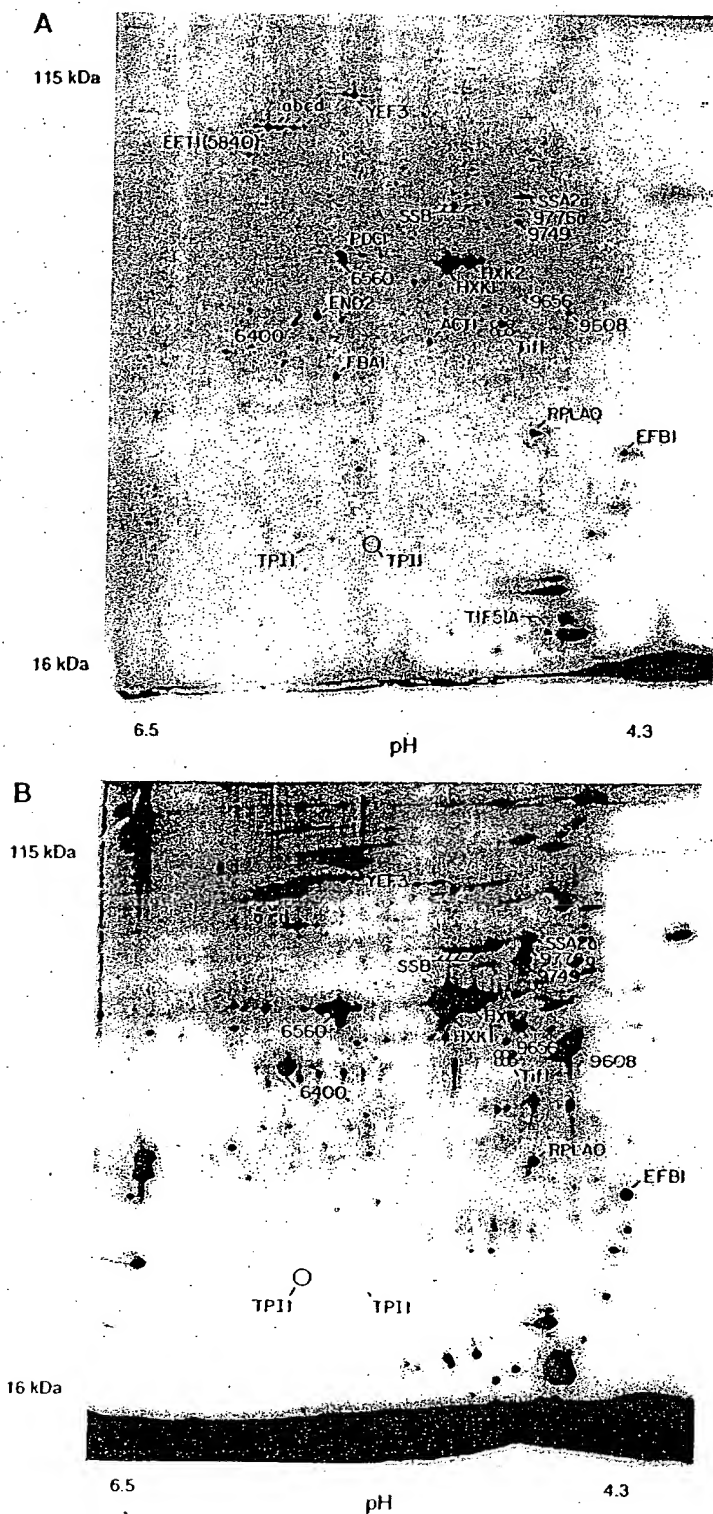


FIG. 5. Phosphorylated proteins. (A) Mixture of ^{32}P -labeled proteins and ^{35}S -labeled proteins. Two separate labeling reactions were done, one with ^{32}P and one with ^{35}S , and extracts were mixed and run on a 2D gel. Spots marked with numbers rather than gene names represent spots noted on ^{35}S gels but unidentified. Spots labeling with ^{32}P were identified by (i) increased labeling compared to the ^{35}S -only gel (not shown); (ii) the characteristic fuzziness of a ^{32}P -labeled spot; and (iii) the decay of signal intensity seen on exposures made 4 weeks later (not shown). A minor form of Tpi1 and at least six minor forms of Tif1 have been noted in overexpression experiments (see also Fig. 6B); positions of the minor forms are indicated by circles. (B) ^{32}P -only labeling. The major form of Tpi1, which is not labeled with ^{32}P , is indicated by a large circle; positions of seven forms of Tif1 are indicated by smaller circles.

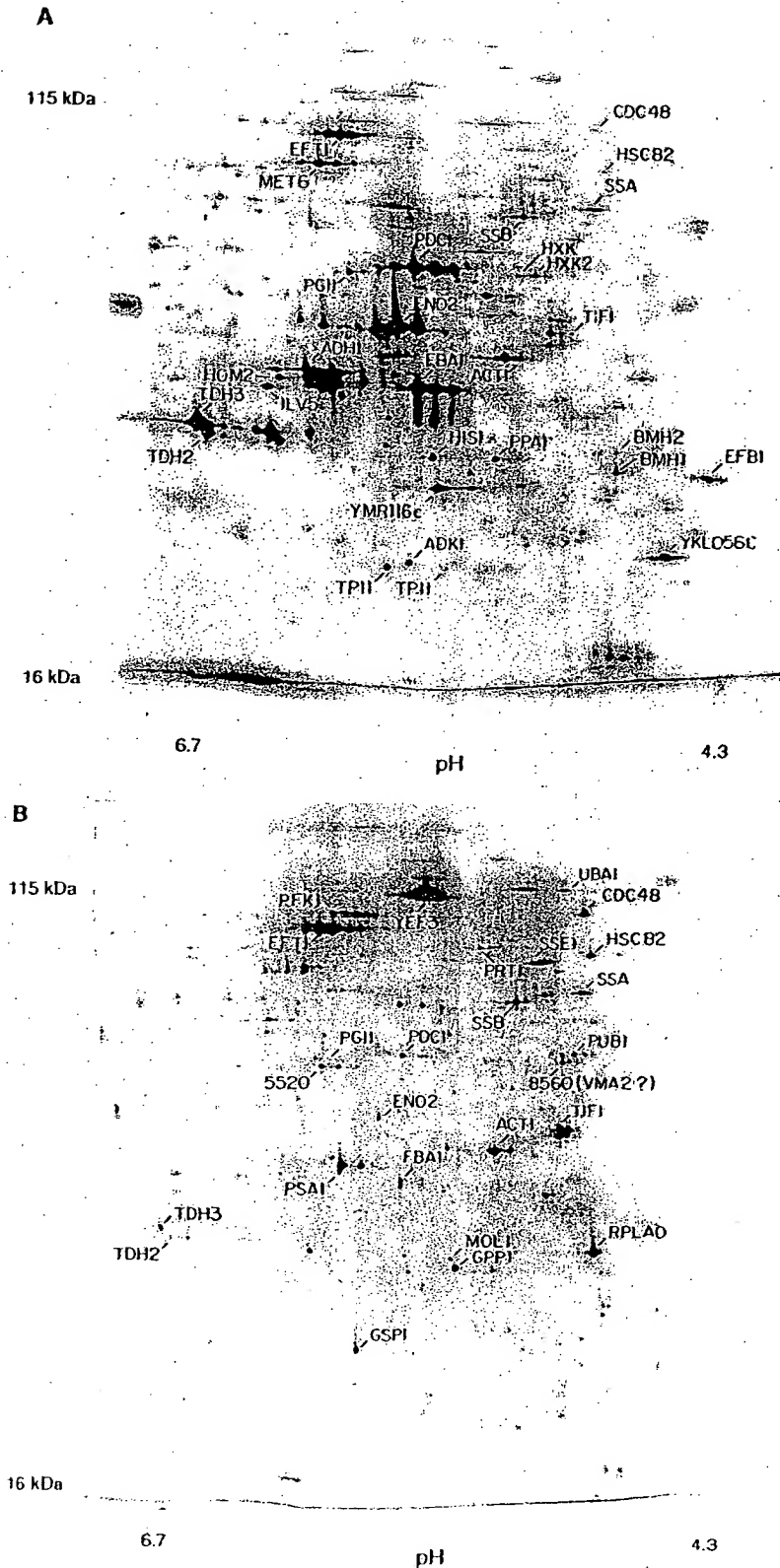


FIG. 6. Fractionation by centrifugation. (A) Proteins in the supernatant of a $100,000 \times g$, 30-min spin; proteins in the pellet of a $16,000 \times g$, 10-min spin. Supernatant fractions examined in multiple experiments done over a wide range of g forces looked similar to each other, as did the pellet fractions.

2D gels of the kind that we have used are not suitable for visualization of rare proteins. However it will be possible to study on a global basis metabolic processes involving relatively abundant proteins, such as protein synthesis, glycolysis, gluconeogenesis, amino acid synthesis, cell wall synthesis, nucleotide synthesis, lipid metabolism, and the heat shock response.

Gygi et al. (10) have recently completed a study similar to ours. Despite generating broadly similar data, Gygi et al. reached markedly different conclusions. We believe that both mRNA abundance and codon bias are useful predictors of protein abundance. However, Gygi et al. feel that mRNA abundance is a poor predictor of protein abundance and that "codon bias is not a predictor of either protein or mRNA levels" (10). These different conclusions are partly a matter of viewpoint. Gygi et al. focus on the fact that the correlations of mRNA and codon bias with protein abundance are far from perfect, while we focus on the fact that, considering the wide range of mRNA and protein abundance and the undoubted presence of other mechanisms affecting protein abundance, the correlations are quite good.

However, the different conclusions are also partly due to different methods of statistical analysis and to real differences in data. With respect to statistics, Gygi et al. used the Pearson product-moment correlation coefficient (r_p) to measure the covariance of mRNA and protein abundance. Depending on the subset of data included, their r_p values ranged from 0.1 to 0.94. Because of the low r_p values with some subsets of the data, Gygi et al. concluded that the correlation of mRNA to protein was poor. However, the r_p correlation is a parametric statistic and so requires variates following a bivariate normal distribution; that is, it would be valid only if both mRNA and protein abundances were normally distributed. In fact, both distributions are very far from normal (data not shown), and so a calculation of r_p is inappropriate. There was no statistical backing for the assertion that codon bias fails to predict protein abundance.

We have taken two statistical approaches. First, we have used the Spearman rank correlation coefficient (r_s). Since this statistic is nonparametric, there is no requirement for the data to be normally distributed. Using the r_s , we find that mRNA abundance is well correlated with protein abundance ($r_s = 0.74$), and the CAI is also well correlated with protein abundance ($r_s = 0.80$) (and also with mRNA abundance [data not shown]). For the data of Gygi et al. (10), we obtained similar results, though with their data the correlation is not as good; $r_s = 0.59$ for the mRNA-to-protein correlation, and $r_s = 0.59$ for the codon bias-to-protein correlation.

In a second approach, we transformed the mRNA and protein data to forms where they were normally distributed, to allow calculation of an r_p (Materials and Methods). Two transformations, Box-Cox and logarithmic, were used; both gave good correlations with our data [e.g., $r_p = 0.76$ for $\log(\text{adjusted RNA})$ to $\log(\text{protein})$]. We were not able to transform the data of Gygi et al. to a normal distribution.

Finally, there are also some differences in data between the two studies. These may be partly due to the different measurement techniques used: Gygi et al. measured protein abundance by cutting spots out of gels and measuring the radioactivity in each spot by scintillation counting, whereas we used phosphorimaging of intact gels coupled to image analysis. We compared our data to theirs for the proteins common between the studies (but excluding proteins whose mRNAs are known to differ between rich and minimal media, and excluding Tif1, which was anomalous in differing by 100-fold between the two data sets). The r_s between the two protein data sets was 0.88 ($P < 0.0001$). Although this is a strong correlation, the fact that

it is less than 1.0 suggests that there may have been errors in measuring protein abundance in one or both studies. After normalizing the two data sets to assume the same amount of protein per cell, we found a systematic tendency for the protein abundance data of Gygi et al. to be slightly higher than ours for the highest-abundance proteins and also for the lowest-abundance proteins but slightly lower than ours for the middle-abundance proteins. These systematic differences suggest some systematic errors in protein measurement. Although we do not know what the errors are, we suggest the following as a reasonable speculation. For the highest-abundance proteins, we may have underestimated the amount of protein because of a slightly nonlinear response of the phosphorimager screens. For the lowest-abundance proteins, Gygi et al. may have overestimated the amount of protein because of difficulties in accurately cutting very small spots out of the gel and because of difficulties in background subtraction for these small, weak spots. The difference in the middle abundance proteins may be a consequence of normalization, given the two errors above.

The low-abundance proteins in the data set of Gygi et al. have a poor correlation with mRNA abundance. We calculate that the r_s is 0.74 for the top 54 proteins of Gygi et al. but only 0.22 for the bottom 53 proteins, a statistically significant difference. However, with our data set, the r_s is 0.62 for the top 33 proteins and 0.56 (not significantly different) for the bottom 33 proteins (which are comparable in abundance to the bottom 53 proteins of Gygi et al.). Thus, our data set maintains a good correlation between mRNA and protein abundance even at low protein abundance. This is consistent with our speculation that protein quantification by phosphorimaging and image analysis may be more accurate for small, weak spots than is cutting out spots followed by scintillation counting. Our relatively good correlations even for nonabundant proteins may also reflect the fact that we used both SAGE data and RNA hybridization data, which is most helpful for the least abundant mRNAs. In summary, we feel that the poor correlation of protein to mRNA for the nonabundant proteins of Gygi et al. may reflect difficulty in accurately measuring these nonabundant proteins and mRNAs, rather than indicating a truly poor correlation *in vivo*. It is not surprising that observed correlations would be poorer with less-abundant proteins and mRNAs, simply because the accuracy of measurement would be worse.

How well can mRNA abundance predict protein abundance? With $r_p = 0.76$ for logarithmically transformed mRNA and protein data, the coefficient of determination, $(r_p)^2$, is 0.58. This means that more than half (in log space) of the variation in protein abundance is explained by variation in mRNA abundance. When converted back to arithmetic values, protein abundances vary over about 200-fold (Table 1), and $(r_p)^2 = 0.58$ for the log data means that of this 200-fold variation, about 20-fold is explained by variation in the abundance of mRNA and about 10-fold is unexplained (but could be due partly to measurement errors). For proteins much less abundant than those considered here, we imagine the *in vivo* correlation between mRNA and protein abundance will be worse, and other regulatory mechanisms such as protein turnover will be more important.

Some important conclusions can be drawn from this sampling of the proteome. First, there is an enormous range of protein abundance, from nearly 2,000,000 molecules per cell for some glycolytic enzymes to about 100 per cell for some cell cycle proteins (26a). Second, about half of all cellular protein is found in fewer than 100 different gene products, which are mostly involved in carbohydrate metabolism or protein synthe-

sis. Third, the correlation between protein abundance and CAI is log linear as far as we can see, which is from about 10,000 protein molecules per cell to about 1,000,000. This is somewhat surprising, because it implies that selective forces for codon bias are significant even at moderate expression levels. It also means that codon bias is a useful predictor of protein abundance even for moderately low bias proteins. Fourth, there is a good correlation between protein abundance and mRNA abundance for the proteins that we have studied. This validates the use of mRNA abundance as a rough predictor of protein abundance, at least for relatively abundant proteins. Fifth, for these abundant proteins, there are about 4,000 molecules of protein for each molecule of mRNA. This last conclusion raises questions as to how the levels of nonabundant proteins are regulated and suggests that protein instability, regulated translation, suboptimal rates of translation, and other mechanisms in addition to transcriptional control may be very important for these proteins.

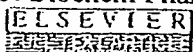
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REFERENCES

- Baroni, M. D., E. Martegani, P. Monti, and L. Alberghina. 1989. Cell size modulation by *CDC25* and *RA52* genes in *Saccharomyces cerevisiae*. *Mol. Cell. Biol.* 9:2715-2723.
- Boucherie, H., F. Sagliocco, R. Joubert, J. Maillet, J. Labarre, and M. Perrot. 1996. Two-dimensional gel protein database of *Saccharomyces cerevisiae*. *Electrophoresis* 17:1683-1699.
- Elliott, B., and B. Futcher. 1993. Stress resistance of yeast cells is largely independent of cell cycle phase. *Yeast* 9:33-42.
- Entian, K. D., B. Meurer, H. Kohler, K. H. Mann, and D. Mecke. 1987. Studies on the regulation of enolases and compartmentation of cytosolic enzymes in *Saccharomyces cerevisiae*. *Biochim. Biophys. Acta* 923:214-221.
- Ganzhorn, A. J., D. W. Green, A. D. Hershey, R. M. Gould, and B. V. Plapp. 1987. Kinetic characterization of yeast alcohol dehydrogenases. Amino acid residue 294 and substrate specificity. *J. Biol. Chem.* 262:3754-3761.
- Garrels, J. I. 1989. The Quest system for quantitative analysis of two-dimensional gels. *J. Biol. Chem.* 264:5269-5282.
- Garrels, J. I., B. Futcher, R. Kobayashi, G. I. Latter, B. Schwender, T. Volpe, J. R. Warner, and C. S. McLaughlin. 1994. Protein identifications for a *Saccharomyces cerevisiae* protein database. *Electrophoresis* 15:1466-1486.
- Garrels, J. I., C. S. McLaughlin, J. R. Warner, B. Futcher, G. I. Latter, R. Kobayashi, B. Schwender, T. Volpe, D. S. Anderson, R. Mesquita-Fuentes, and W. E. Payne. 1997. Proteome studies of *S. cerevisiae*: identification and characterization of abundant proteins. *Electrophoresis* 18:1347-1360.
- Goffrau, A., B. G. Barrell, H. Bussey, R. W. Davis, B. Dujon, H. Feldmann, F. Galibert, J. D. Hoebisel, C. Jacq, M. Johnston, E. J. Louis, H. W. Mewes, Y. Morakami, P. Philippsen, H. Tettelin, and S. G. Oliver. 1996. Life with 6000 genes. *Science* 274:563-567.
- Gygi, S. P., Y. Rochon, B. R. Franz, and R. Aebersold. 1999. Correlation between protein and mRNA abundance in yeast. *Mol. Cell. Biol.* 19:1720-1730.
- Hereford, L. M., and M. Rosbash. 1977. Number and distribution of polyadenylated RNA sequences in yeast. *Cell* 10:453-462.
- Herrick, D., R. Parker, and A. Jacobson. 1990. Identification and comparison of stable and unstable mRNAs in *Saccharomyces cerevisiae*. *Mol. Cell. Biol.* 10:2269-2284.
- Hodges, P. E., A. H. McKee, B. P. Davis, W. E. Payne, and J. I. Garrels. 1999. The Yeast Proteome Database (YPD): a model for the organization of genome-wide functional data. *Nucleic Acids Res.* 27:69-73.
- Ikemura, T. 1985. Codon usage and tRNA content in unicellular and multicellular organisms. *Mol. Biol. Evol.* 2:13-34.
- Johnston, G. C., F. R. Pringle, and L. H. Hartwell. 1977. Coordination of growth with cell division in the yeast *S. cerevisiae*. *Exp. Cell Res.* 105:79-98.
- Johnston, M., and M. Carlson. 1992. Regulation of carbon and phosphate utilization, p. 193-281. In E. Jones, J. Pringle, and J. Broach (ed.), *The molecular and cellular biology of the yeast Saccharomyces*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.
- Kornblatt, M. J., and A. Klugerman. 1989. Characterization of the enolase isozymes of rabbit brain: kinetic differences between mammalian and yeast enolases. *Biochem. Cell. Biol.* 67:103-107.
- Latter, G., and B. Futcher. Unpublished data.
- Mathews, B., N. Sonenberg, and J. W. B. Hershey. 1996. Origins and targets of translational control, p. 1-29. In J. W. B. Hershey, M. B. Mathews, and N. Sonenberg (ed.), *Translational control*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.
- McAlister, L., and M. J. Holland. 1982. Targeted deletion of a yeast enolase structural gene. Identification and isolation of yeast enolase isozymes. *J. Biol. Chem.* 257:7181-7188.
- Monardo, P. J., T. Boutell, J. I. Garrels, and G. I. Latter. 1994. A distributed system for two-dimensional gel analysis. *Comput. Appl. Biosci.* 10:137-143.
- O'Farrell, P. H. 1975. High resolution two-dimensional electrophoresis of proteins. *J. Biol. Chem.* 250:4007-4021.
- Patterson, S. D., and G. I. Latter. 1993. Evaluation of storage phosphor imaging for quantitative analysis of 2-D gels using the Quest II system. *BioTechniques* 15:1076-1083.
- Sagliocco, F., J. C. Guillemot, C. Monribot, J. Capdevielle, M. Perrot, E. Ferran, P. Ferrara, and H. Boucherie. 1996. Identification of proteins of the yeast protein map using genetically manipulated strains and peptide-mass fingerprinting. *Yeast* 12:1519-1533.
- Sharp, P. M., and W. H. Li. 1987. The Codon Adaptation Index—a measure of directional synonymous codon usage bias, and its potential applications. *Nucleic Acids Res.* 15:1281-1295.
- Shevchenko, A., O. N. Jensen, A. V. Podilejnikov, F. Sagliocco, M. Wilm, O. Vorm, P. Mortensen, A. Shevchenko, H. Boucherie, and M. Mann. 1996. Linking genome and proteome by mass spectrometry: large-scale identification of yeast proteins from two dimensional gels. *Proc. Natl. Acad. Sci. USA* 93:14440-14445.
- Thomas, B. J., and R. Rothstein. 1989. Elevated recombination rates in transcriptionally active DNA. *Cell* 56:619-630.
- Tyres, M., and B. Futcher. Unpublished data.
- Velculescu, V. E., L. Zhang, W. Zhou, J. Vogelstein, M. A. Basrai, D. E. Bassett, Jr., P. Hieter, B. Vogelstein, and K. W. Kinzler. 1997. Characterization of the yeast transcriptome. *Cell* 88:243-251.
- Warner, J. 1991. Labeling of RNA and phosphoproteins in *S. cerevisiae*. *Methods Enzymol.* 194:423-428.
- Wills, C. 1976. Production of yeast alcohol dehydrogenase isoenzymes by selection. *Nature* 261:26-29.
- Wodicka, L. Personal communication.
- Wodicka, L. Unpublished data.
- Wodicka, L., H. Dong, M. Mittmann, M.-H. Ho, and D. J. Lockhart. 1997. Genome-wide expression monitoring in *Saccharomyces cerevisiae*. *Nat. Biotechnol.* 15:1359-1367.



Pre-translational regulation of cytochrome P450 genes is responsible for disease-specific changes of individual P450 enzymes among patients with cirrhosis.

George J. Liddle C, Murray M, Byth K, Farrell GC.

Department of Gastroenterology and Hepatology, University of Sydney at Westmead Hospital, NSW, Australia.

We have recently reported that disease-specific differential alterations in the hepatic expression of xenobiotic-metabolizing cytochrome P450 (CYP P450) enzymes occur in patients with advanced liver disease. In order to determine whether the observed changes in CYP proteins are modulated at pre- or post-translational levels, we have now examined the hepatic levels of mRNA for CYPs 1A2, 2C9, 2E1 and 3A4 by solution hybridization in the same livers of 20 controls (surgical waste from histologically normal livers), 32 cases of hepatocellular and 18 of cholestatic severe chronic liver disease. CYP1A2 mRNA and CYP1A immunoreactive protein were both reduced in livers with hepatocellular and cholestatic types of cirrhosis. In contrast, CYP3A4 mRNA and protein were reduced only in livers from patients with hepatocellular diseases. For 1A2 and 3A4 there were significant correlations between mRNA species and the respective protein contents ($r_{S1A2} = 0.74$, $r_{S3A4} = 0.64$, $P < 0.0001$). CYP2C9 mRNA was reduced in patients with both cholestatic and hepatocellular types of liver disease, but 2C protein was reduced only in patients with cholestatic dysfunction. The correlation between CYP2C9 mRNA and protein, was also significant ($r_s = 0.36$, $P < 0.005$) but mRNA levels accounted for only 13% of the variability in protein rankings. This is probably a consequence of other CYP2C proteins apart from 2C9 being detected by the anti-2C antibody. CYP2E1 mRNA and protein were reduced in patients with cholestatic liver disease, but in hepatocellular disease the expression of only CYP2E1 mRNA was decreased. CYP2E1 mRNA was significantly correlated with CYP2E1 protein but accounted for only 18% of the variability in protein rankings ($r_s = 0.43$, $P < 0.0005$). Taken collectively these data indicate that the disease-specific alterations of xenobiotic-metabolizing CYP enzymes among patients with cirrhosis is due, at least in part, to pre-translational mechanisms. The lack of a strong correlation between CYP2E1 mRNA and protein suggests that this gene, like its rat orthologue, may be subject to pre-translational as well as translational and/or post-translational regulation.

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Cyclooxygenase-2 expression in macrophages: modulation by protein kinase C- α .

Giroux M, Descoteaux A.

Institut National de la Recherche Scientifique-Institut Armand-Frappier, Université du Québec, Laval, Canada.

Cyclooxygenase-2 (COX-2) is an inducible enzyme responsible for high levels of PG production during inflammation and immune responses. Previous studies with pharmacological inhibitors suggested a role for protein kinase C (PKC) in PG production possibly by regulating COX-2 expression. In this study, we addressed the role of PKC- α in the modulation of COX-2 expression and PGE2 synthesis by the overexpressing of a dominant-negative (DN) mutant of this isoenzyme in the mouse macrophage cell line RAW 264.7. We investigated the effect of various stimuli on COX-2 expression, namely, LPS, IFN- γ , and the intracellular parasite *Leishmania donovani*. Whereas LPS-induced COX-2 mRNA and protein expression were down-regulated in DN PKC- α -overexpressing clones, IFN- γ -induced COX-2 expression was up-regulated in DN PKC- α -overexpressing clones with respect to normal RAW 264.7 cells. Measurements of PGE2 levels revealed a strong correlation between PGE2 secretion and IFN- γ -induced COX-2 mRNA and protein levels in DN PKC- α -overexpressing clones. Taken together, these results suggest a role for PKC- α in the modulation of LPS- and IFN- γ -induced COX-2 expression, as well as in IFN- γ -induced PGE2 secretion.

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Transcript profiling of human platelets using microarray and serial analysis of gene expression.

Gnatenko DV, Dunn JJ, McCorkle SR, Weissmann D, Perrotta PL, Bahou WF.

Department of Medicine, Program in Genetics, State University of New York, Stony Brook 11794-8151, USA.

Human platelets are anucleate blood cells that retain cytoplasmic mRNA and maintain functionally intact protein translational capabilities. We have adapted complementary techniques of microarray and serial analysis of gene expression (SAGE) for genetic profiling of highly purified human blood platelets. Microarray analysis using the Affymetrix HG-U95Av2 approximately 12 600-probe set maximally identified the expression of 2147 (range, 13%-17%) platelet-expressed transcripts, with approximately 22% collectively involved in metabolism and receptor/signaling, and an overrepresentation of genes with unassigned function (32%). In contrast, a modified SAGE protocol using the Type IIS restriction enzyme Mmel (generating 21-base pair [bp] or 22-bp tags) demonstrated that 89% of tags represented mitochondrial (mt) transcripts (enriched in 16S and 12S ribosomal RNAs), presumably related to persistent mt-transcription in the absence of nuclear-derived transcripts. The frequency of non-mt SAGE tags paralleled average difference values (relative expression) for the most "abundant" transcripts as determined by microarray analysis, establishing the concordance of both techniques for platelet profiling. Quantitative reverse transcription-polymerase chain reaction (PCR) confirmed the highest frequency of mt-derived transcripts, along with the mRNAs for neurogranin (NGN, a protein kinase C substrate) and the complement lysis inhibitor clusterin among the top 5 most abundant transcripts. For confirmatory characterization, immunoblots and flow cytometric analyses were performed, establishing abundant cell-surface expression of clusterin and intracellular expression of NGN. These observations demonstrate a strong correlation between high transcript abundance and protein expression, and they establish the validity of transcript analysis as a tool for identifying novel platelet proteins that may regulate normal and pathologic platelet (and/or megakaryocyte) functions.

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Transcript profiling of human platelets using microarray and serial analysis of gene expression

Dmitri V. Gnatenko, John J. Dunn, Sean R. McCorkle, David Weissmann, Peter L. Perrotta, and Wadie F. Bahou

Human platelets are anucleate blood cells that retain cytoplasmic mRNA and maintain functionally intact protein translational capabilities. We have adapted complementary techniques of microarray and serial analysis of gene expression (SAGE) for genetic profiling of highly purified human blood platelets. Microarray analysis using the Affymetrix HG-U95Av2 approximately 12 600-probe set maximally identified the expression of 2147 (range, 13%-17%) platelet-expressed transcripts, with approximately 22% collectively involved in metabolism and receptor/signaling, and an overrepresentation of genes with unassigned function (32%). In contrast, a modified SAGE protocol using the Type IIS restriction enzyme

MmeI (generating 21-base pair [bp] or 22-bp tags) demonstrated that 89% of tags represented mitochondrial (mt) transcripts (enriched in 16S and 12S ribosomal RNAs), presumably related to persistent mt-transcription in the absence of nuclear-derived transcripts. The frequency of non-mt SAGE tags paralleled average difference values (relative expression) for the most "abundant" transcripts as determined by microarray analysis, establishing the concordance of both techniques for platelet profiling. Quantitative reverse transcription-polymerase chain reaction (PCR) confirmed the highest frequency of mt-derived transcripts, along with the mRNAs for neurogranin (NGN, a protein kinase C substrate) and

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Introduction

Human blood platelets play critical roles in normal hemostatic processes and pathologic conditions such as thrombosis, vascular remodeling, inflammation, and wound repair. Generated as cytoplasmic buds from precursor bone marrow megakaryocytes, platelets are anucleate and lack nuclear DNA, although they retain megakaryocyte-derived mRNAs.^{1,2} Platelets contain rough endoplasmic reticulum and polyribosomes, and they retain the ability for protein biosynthesis from cytoplasmic mRNA.³ Quiescent platelets generally display minimal translational activity, although newly formed platelets such as those found in patients with immune thrombocytopenic purpura (ITP) synthesize various α -granule and membrane glycoproteins (GPs), including GPIb and GPIIb/IIIa ($\alpha_{IIb}\beta_3$). Furthermore, stimulation of quiescent platelets by agonists such as α -thrombin increases protein synthesis of various platelet proteins, including Bcl-3.⁴ Like nucleated cells, the rapid translation of preexisting mRNAs may be regulated by integrin ligation to extracellular matrices.⁵ In the case of platelets, the primary integrin involved in this process appears to be $\alpha_{IIb}\beta_3$ with cooperative signals mediated by the collagen receptor $\alpha_2\beta_1$.^{6,7}

Integrin-mediated platelet protein synthesis appears to be regulated at the level of translation initiation involving the eukaryotic initiation factor 4E (eIF4E). Instead of directly influencing eIF4E activity via posttranslational modifications (ie, phosphorylation), platelet eIF4E activity best correlates with its spatial redistribution to the mRNA-enriched cytoskeleton.⁸ Furthermore, because protein translation is partially inhibited by the immunosuppressant rapamycin, it suggests that adhesion- and/or aggregation-induced outside-in-signaling function to regulate protein synthesis through the mTOR (mammalian target of rapamycin) pathway.^{6,8,9}

Despite the biologic importance of platelets and their intact protein synthetic capabilities, remarkably little is known about platelet mRNAs. Younger platelets contain larger amounts of mRNA with a greater capacity for protein synthesis, as determined by using fluorescent nucleic acid dyes such as thiazole orange.¹⁰ This assay has been used as a quantitative determinant of younger or "reticulated" platelets (RPs). Indeed increased reticulated platelets are typically found in patients with conditions associated with rapid platelet turnover such as ITP; typically RP percentages in

From the Department of Medicine, Department of Pathology, and Program in Genetics, State University of New York, Stony Brook; Biology Department, Brookhaven National Laboratory, Upton, NY; and Department of Pathology, Robert Wood Johnson Medical Center, New Brunswick, NJ.

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Reprints: Wadie F. Bahou, Division of Hematology, HSCT15-040, State University of New York at Stony Brook, Stony Brook, NY 11794-8151; e-mail: wbahou@notes.cc.sunysb.edu.

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such patients approach 10% to 20% of all platelets, considerably higher than in healthy control subjects.¹¹ Interestingly, high RPs have been associated with enhanced thrombotic risk when identified in patients with thrombocytosis,¹⁰ suggesting that quantitatively increased mRNA levels may be associated with the prothrombotic phenotype. Whether this is related to globally altered gene expression profiles or to select changes more evident during situations of rapid platelet turnover remains unknown. Certainly, technical limitations of this assay limit its utility in defining prothrombotic genotypes,¹⁰⁻¹² and it cannot identify differentially expressed genes that may be causally implicated in disordered platelet phenotypes.

Toward the goal of defining the molecular anatomy of the platelet genome, we have adapted complementary techniques of microarray and serial analysis of gene expression (SAGE) for genetic profiling of highly purified human blood platelets. Microarray technology represents a "closed" profiling strategy limited by the target genes imprinted onto gene chips. In contrast, SAGE is an "open" architectural system that can be used to identify novel genes and to quantify differentially expressed mRNAs.¹³⁻¹⁵ The sequence of each tag along with its positional location uniquely identifies the gene from which it is derived, and differentially expressed genes can be identified in a quantitative manner because the tag frequency reflects the mRNA level at the time of cellular harvest and analysis. By using both technologies, we have identified a number of previously uncharacterized genes that appear to be expressed in human platelets, while simultaneously establishing the dominant frequency of mitochondrial-expressed genomes comprising the platelet mRNA pool. These observations provide a panoramic overview of the platelet transcriptome, while additionally providing insights into the molecular pathways regulating platelet (and/or megakaryocyte) function in normal and pathologic conditions.

Materials and methods

Reagents and supplies

Thermus aquaticus (Taq) polymerase was purchased from (Roche, Indianapolis, IN). T4 DNA ligase was purchased from Invitrogen (Carlsbad, CA), and restriction enzymes were from New England Biolabs (Beverly, MA), except for *MmeI*, which was obtained from the Center for Technology Transfer (Gdansk, Poland). All oligonucleotides were synthesized on an Applied Biosystems (Foster City, CA) 3-channel synthesizer and are listed in Table 1. Monoclonal antibodies used for flow cytometric analysis included the FITC (fluorescein isothiocyanate)-conjugated anti-CD41 ($\alpha_{IIb}\beta_3$) immunoglobulin G1 (IgG1; Immunotech, Miami, FL); phycoerythrin (PE)-conjugated anti-glycophorin (IgG2; Becton Dickinson Pharmingen, San Diego, CA); and peridinin chlorophyll protein (PERCP)-conjugated anti-CD45 (IgG1; Becton Dickinson Pharmingen).

Platelet isolation, purification, and immunodetection

All human subjects provided informed consent for an IRB (Institutional Review Board)-approved protocol completed in conjunction with the General Clinical Research Center at Stony Brook University Hospital. Peripheral blood (20 mL) from healthy volunteers drawn into 2 mL of 4% sodium citrate (0.4% vol/vol final concentration) was used to isolate erythrocytes by differential centrifugation (1500g) or to isolate pure leukocytes by density-gradient centrifugation as previously described.¹⁶ Platelets collected from healthy volunteers by apheresis were used within 24 hours of collection. After addition of 2 mM EDTA (ethylenediaminetetraacetic acid), apheresis-derived platelets from a single donor were centrifuged at 140g for 15 minutes at 25°C. To minimize leukocyte contamination, only the upper 9/10 of the platelet-rich plasma (PRP) was

used for gel filtration over a BioGel A50M column (1000 mL total volume) equilibrated with HBMT (HEPES-buffered modified Tyrodes buffer: 10 mM HEPES (*N*-2-hydroxyethylpiperazine-*N'*-2-ethanesulfonic acid) pH 7.4, 150 mM NaCl, 2.5 mM KCl, 0.3 mM NaH_2PO_4 , 12 mM NaHCO_3 , 0.2% bovine serum albumen [BSA], 0.1% glucose, 2 mM EDTA). Gel-filtered platelets (GFPs) were subsequently filtered through a 5- μm nonwetting nylon filament filter (BioDesign, Carmel, NY) at 25°C and harvested by centrifugation at 1500g for 10 minutes at 25°C. Platelets were gently and thoroughly resuspended in 10 mL HBMT buffer and incubated with 120 μL murine monoclonal anti-CD45 antibody conjugated to magnetic microbeads (Miltenyi Biotec, Bergisch Gladbach, Germany) on a rotating platform for 45 minutes at 25°C. Magnetic separation columns were used to capture CD45⁺ cells (leukocyte fraction) by positive selection (MACS II; Miltenyi Biotec). Purified platelets were concentrated by centrifugation at 1500g and immediately used for total RNA isolation.

The efficiency of platelet purification was documented at each step by flow cytometry.¹⁷ Briefly, aliquots containing 2×10^6 platelets were incubated with saturating concentrations of FITC-conjugated anti-CD41, PE-conjugated anti-glycophorin, and PERCP-conjugated anti-CD45 for 15 minutes in the dark at 25°C, washed with phosphate-buffered saline (PBS), and fixed in PBS/1% formalin. Samples were analyzed using a FACScan (fluorescence-activated cell sorter scan) flow cytometer (Becton Dickinson) using CELLQuest software designed to quantify the number of CD45⁺ and glycophorin-positive events in the sample (expressed as the number of events per 100 000 CD41⁺ events). For some experiments, fixed platelets were permeabilized with 0.1% Triton-X/PBS for 30 minutes at 25°C prior to the addition of primary antibodies, all as previously described.¹⁷

Platelet protein detection was completed by sodium dodecyl sulfate (SDS)-polyacrylamide gel electrophoresis (PAGE) and immunoblot analysis as previously described, using the species-specific horseradish peroxidase-conjugated secondary antibody and enhanced chemiluminescence.¹⁸ Antibodies included the anti-clusterin monoclonal antibody (Quidel, Santa Clara, CA; 1:1000 primary and 1:10 000 secondary) and the antineurogranin rabbit polyclonal antibody (Chemicon International, Temecula, CA; 1:1000 primary and 1:10 000 secondary).

Molecular analyses and microarray profiling

Purified, individual cell fractions were resuspended in 10 mL Trizol reagent (Invitrogen), transferred into diethylpyrocarbonate (DEPC)-treated Corex (Springfield, MA) tubes, and serially purified and precipitated by using isopropanol essentially as previously described.¹⁸ Total cellular RNA was harvested by centrifugation at 12 500g for 20 minutes at 4°C, washed 2 times with 75% ethanol (10 mL/tube), and resuspended in 100 μL DEPC-treated water. Platelet mRNA quantitation was performed by using fluorescence-based real-time PCR (polymerase chain reaction) technology (TaqMan Real-Time PCR; Applied Biosystems, Foster City, CA). Oligonucleotide primer pairs were generated by using Primer3 software (www.genome.wi.mit.edu), designed to generate approximately 200-base pair (bp) PCR products at the same annealing temperature, and are outlined in Table 1. Purified platelet mRNA (4 μg) was used for first-strand cDNA synthesis using oligo(dT) and SuperScript II reverse transcriptase (Invitrogen). For real-time reverse transcription (RT)-PCR analysis, the RT reaction was equally divided among primer pairs and used in a 40-cycle PCR reaction for each target gene by using the following cycle: 94°C for 30 seconds, 55°C for 30 seconds, 72°C for 1 minute, and 71°C for 10 seconds (40 cycles total). mRNA levels were quantified by monitoring real-time fluorometric intensity of SYBR green I. Relative mRNA abundance was determined from triplicate assays performed in parallel for each primer pair and was calculated by using the comparative threshold cycle number ($\Delta\text{-Ct}$ method) as previously described.¹⁹

Gene expression profiles were completed by using the approximately 12 600-probe set HG-U95Av2 gene chip (Affymetrix, Santa Clara, CA). Total cellular RNA (5 μg) was used for cDNA synthesis by using SuperScript Choice system (Life Technologies, Rockville, MD) and an oligo(dT) primer containing the T7 polymerase recognition sequence (Primer S1; Table 1), followed by cDNA purification using GFX spin columns. In vitro transcription was completed in the presence of biotinylated ribonucleotides by using a BioArray HighYield RNA Transcript

Table 1. Oligonucleotide primers

| Primer | Gene and primer direction | Sequence (5' - 3') | Nucleotide Position |
|------------|-----------------------------|---|---------------------|
| S1 | Oligo (dT) | 5'-Bn-GGCCAGTGAATTGTAATACGACTCACTATAGGAGGCGG- (dT) ₂₄ -3' | — |
| Cassette A | SAGE | 5'-TTTGATTTCGTGGTCGAGTACAAGTGGCTTAATCCGACATG-3' 3'-*CCTAAAGCAGCAGCTCATGTTGATCGAATAGGCTp-5' | — |
| Cassette B | SAGE | 5'-pTTCATGCGGAGACGTCGCCCACTAGTGTCCGAAGTACTA*-3' 3'-NNAAGTACCGCTCTGACGGCGGTGATCACAGCGTTGACTGAT-5' | — |
| S2 | SAGE | 5'-Bn-GGATTTCGTGGTCGAGTACA-3' | — |
| S3 | SAGE | 5'-Bn-TAGTCAGGTGCGACACTAGTGGC-3' | — |
| GP4 | Glycoprotein IIB [F] | 5'-AGGGCTTTGAGAGACTCATCTGTA-3' | 2094-2117 |
| GP5 | Glycoprotein IIB [R] | 5'-ACAATCTTGCTGTTTGATTCTG-3' | 2301-2279 |
| GP6 | Glycoprotein IIIA [F] | 5'-TATAAGAGGCCAGCTCACTTC-3' | 2335-2358 |
| GP7 | Glycoprotein IIIA [R] | 5'-CACTCCACATCTGACATTCTCC-3' | 2532-2509 |
| PAR18 | PAR1 [F] | 5'-AATGTGCTCTGATATGGAAGCA-3' | 2585-2608 |
| PAR19 | PAR1 [R] | 5'-CCCAATGTTCAAACTTCCTTACC-3' | 2776-2753 |
| SR8 | 16S rRNA [F] | 5'-TGCAAGGTAGCATAATCACTTGT-3' | 2586-2609 |
| SR9 | 16S rRNA [R] | 5'-GTTTAGGACCTGTGGTTTGTTAG-3' | 2785-2762 |
| NADH10 | NADH2 [F] | 5'-CTAGCCCCATCTCAATCATATAC-3' | 4875-4898 |
| NADH11 | NADH2 [R] | 5'-AATGGTTATGTTAGGGTTGACGG-3' | 5075-5052 |
| THYM12 | Thymosin β4 [F] | 5'-AAGACAGAGACGCAAGAGAAAAAT-3' | 135-158 |
| THYM13 | Thymosin β4 [R] | 5'-GCAGCAGTCATTAACTTGTAT-3' | 336-313 |
| CLUS14 | Clusterin [F] | 5'-CCAACAGATTCTACGAGAGG-3' | 1006-1028 |
| CLUS15 | Clusterin [R] | 5'-CGTTATATTCTCGTCAACTCT-3' | 1222-1199 |
| NRG16 | Neurogranin [F] | 5'-GCCCTTTAGTTAGTCTGCACTC-3' | 1351-1374 |
| NRG17 | Neurogranin [R] | 5'-TTTCTTTAAGTAGTGTGCTTGG-3' | 1567-1544 |
| TCR18 | T-cell receptor β-chain [F] | 5'-CCACAATATGTTTGGTATCGT-3' | 131-153 |
| TCR19 | T-cell receptor β-chain [R] | 5'-CTAGCACTGCAGATGTAGAAGCT-3' | 332-310 |
| CD4520 | CD45 [F] | 5'-GCTCAGATGGACAAGTA-3' | 3771-3788 |
| CD4521 | CD45 [R] | 5'-CACACCCATACACATACA-3' | 4280-4261 |

[F] indicates forward (sense) strand; [R], reverse (antisense) strand; Bn, biotin; p, a phosphorylated 5' end (cassettes A and B); underlining, *Mall* sites in cassettes A and B; arrows, corresponding sequence for S2 and S3 within cassettes A and B, respectively; bold, the *Mme* site; and N, A, C, T, or G, nucleotide position based on the following accession numbers: glycoprotein IIB (J02764), glycoprotein IIIA (M35999), PAR1 (M62424), 16S rRNA and NADH2 (NC_001807), thymosin β4 (M17733), clusterin (M25915), neurogranin (X99076), TCR β-chain (AF043182), CD45 (Y00638).

*Indicates an amino-modified 3' end in both cassettes; —, not applicable.

Labeling Kit (Enzo Diagnostics, Farmingdale, NY), and, after metal-induced fragmentation, 15 μg biotinylated cRNA was hybridized to the HG-U95Av2 oligonucleotide probe array for 16 hours at 45°C. After washing, the cRNA was detected with streptavidin-phycoerythrin (Molecular Probes, Eugene, OR) and analysis was completed by using a Hewlett-Packard Gene Array Scanner (Affymetrix). The average difference value (AD) for each probe set was quantified using MAS 4.01 software (Affymetrix), calculated as an average of fluorescence differences for perfectly matched versus single-nucleotide mismatched 25-mer oligonucleotides (16 to 20 oligonucleotide pairs per probe set). The software is designed to exclude "positive calls" in the presence of high average differences with associated high mismatch intensities.

SAGE profiles

Platelet SAGE libraries were generated essentially as previously described,¹⁵ modified as outlined in Figure 1 for the use of *Mme* as the tagging enzyme.¹⁹ This type IIS restriction enzyme cleaves 20 of 18 bp past its nonpalindromic (TCCRAC) recognition sequence, thereby generating longer tags (21- or 22-mer) than those obtained using *Bsm*F1 as the standard tagging enzyme (13-14 bp tags). These longer *Mme*-generated tags potentially provide for more definitive "tag-to-gene" identification and are particularly useful in characterizing expression patterns in the absence of complete genomic sequence data (comprehensive methods detailed in Dunn et al¹⁵). Briefly, poly(A) mRNA was isolated from 10 μg total platelet RNA using the oligo-dT S1 primer conjugated to magnetic beads (Dyna Beads, Lake Success, NY), followed by cDNA synthesis using SuperScript II reverse transcriptase (Invitrogen). The cDNA was then digested

with the restriction enzyme *Mall* (anchoring enzyme), ligated to cassette A using T4 DNA ligase, and, after the beads were extensively washed, the cDNA was digested with *Mme* to release the tags from the beads. After purification, tags were ligated to degenerate cassette B linkers (specifically

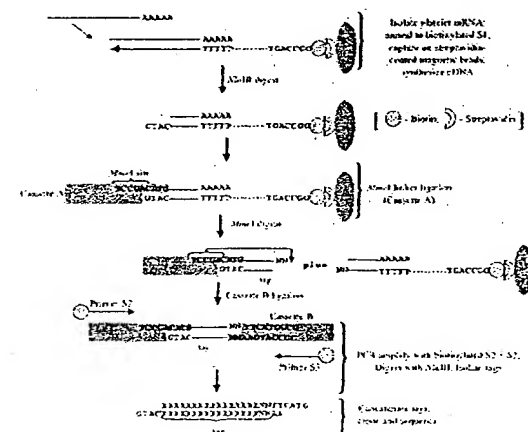


Figure 1. Schema outlining the modified SAGE protocol used in platelet analyses. The final tags are flanked by the *Mall* (anchoring enzyme) CATG sequence, thereby providing tag-to-gene identification when exported to a relational database (refer to "Bioinformatic analyses" and Table 1 for details).

designed to anneal to the nonuniform *MmeI* overhangs), and PCR-amplified using biotinylated primers S2 and S3 for 30 cycles (95°C for 30 seconds; 58°C for 30 seconds; 72°C for 30 seconds) using Platinum Taq DNA polymerase (Gibco BRL). A fraction (20%) of the pooled PCR products were then subjected to one round of linear amplification using primer pair S2/S3, followed by a second round of 25 amplifications using primer S2 alone (95°C for 30 seconds; 58°C for 30 seconds; 72°C for 30 seconds). Primer S3 was subsequently added for one cycle (95°C for 2.5 minutes; 58°C for 30 seconds; 72°C for 5 minutes); the latter steps were collectively adapted to exclude heteroduplex formation.¹⁸ Unincorporated primers were removed by incubation with 200 U *Escherichia coli* exonuclease I for 60 minutes at 37°C. PCR products were then pooled and digested with *NlaIII* to release tags, and biotinylated linker arms were cleared using streptavidin-coated immunoaffinity magnetic beads (Dyna Beads). Tags were concatamerized using 5 U/μL T4 DNA ligase, and products more than 100 bp were isolated by size-fractionation in low-melting agarose gels. The DNA was purified by GFX spin columns, and the concatamers were cloned into the *SphI* site of pZero (Invitrogen). After transformation into *E. coli* TOP10 cells, recombinant clones were isolated and sequenced in 96-well microtiter plates using an ABI 377 sequencer and ABI Prism BigDye terminator chemistry (Perkin-Elmer Applied Biosystems, Branchburg, NJ).

Bioinformatic analyses

Functional grouping of genes determined to be present by Affymetrix MAS 4.01 software was performed using a dChip program linked to the National Center for Biotechnology LocusLink, which is an annotated reference database for genes and their postulated functions.²⁰ Of the approximately 12 600-probe sets represented on the Affymetrix HG-U95Av2 Gene chip, functional annotations exist for approximately 8100 with the remainder categorized as unknown. Microarray data were visualized and analyzed using BRB-ArrayTools software (Version 2.1), kindly developed and provided by Dr Richard Simon and Amy Peng (linus.nci.nih.gov/BRB-ArrayTools.html). A logarithmic (base 2) transformation was applied to the average difference values for individual data sets for determination of microarray concordancies. Discordancy was defined as a 2-log difference in the maximum log intensities between individual experiments.

SAGE tags were extracted by using in-house SAGE software uniquely modified to identify *MmeI* tags. The software ensures that only unambiguous 21- to 22-bp tag sequences are extracted for transcript profiling. Tags with ambiguities (Ns), lengths other than 21 or 22 bp, or with ambiguous orientations were extracted to separate files for manual editing or further examination. Finalized data were exported to a relational database for tag quantification and genetic identification.²⁰

Results

Platelet purification

To ensure that the RNA profiles accurately represented those of circulating blood platelets, a number of complementary methods were implemented to remove contaminating nucleated leukocytes. Purification methods incorporating gel filtration, a 5-μm leukocyte reduction filter, and magnetic CD45 immunodepletion allowed for the cumulative enrichment of highly purified platelets. The efficacy of this purification method was initially established by using peripheral blood platelet-rich plasma as the starting material. The final product contained no more than 3 to 5 leukocytes per 1×10^5 platelets as determined by parallel flow cytometric analysis, representing an approximate 450-fold reduction of nucleated leukocytes. These results correlated well with molecular evidence for leukocyte depletion as determined by RT-PCR using both CD45 and T-cell receptor β-chain (TCRβ) primers (see Figure 2). Because the total RNA yield from peripheral blood platelets was insufficient for microarray studies, we adapted the protocol to platelet apheresis donors with nearly identical final purity (Figure

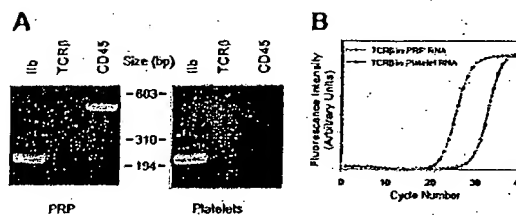


Figure 2. Determination of platelet purity. (A) Total cellular RNA (1.8 μg) from platelet-rich plasma (PRP) or purified platelets from a single apheresis donor were analyzed by RT-PCR (35 cycles) using oligonucleotide primers specific for glycoprotein 1b (GPIIb), T-cell receptor β-chain (TCRβ), or CD45; 10 μL of the 50 μL reactions were analyzed by ethidium-stained agarose gel electrophoresis. Minimal to no TCRβ gene product was visually evident only in PRP. Size markers corresponding to *HaeIII*-restricted 4X174 DNA are shown. (B) Real-time RT-PCR was completed by using 1.8 μg total RNA and TCRβ-specific oligonucleotide primers optimized for quantitative analysis by real-time PCR.¹⁸ On the basis of parallel determinations using RNA isolated from known amounts of purified leukocyte standards, the leukocyte-depletion protocol represents an approximate 2.5-log purification from the starting PRP. Results are representative of one complete set of experiments repeated on 2 separate occasions, and data points represent the mean from triplicate wells, with standard errors of the mean (SEM) less than 1% (not shown).

2). The platelet recovery was nearly 65% of the starting material, yielding approximately 2.3×10^{11} platelets from an initial apheresis pack containing approximately 3.6×10^{11} platelets. The bulk of the losses occurred during the initial centrifugation and filtration steps. The purification protocol was less effective at removing erythrocytes, although there were less than 50 glycophorin-positive cells per 1×10^5 platelets after the final purification step. Nonetheless, these cells represent unlikely sources for contaminating cellular RNA (see "Cellular microarray analysis" below).

Cellular microarray analysis

The purified platelet RNA was sufficient for microarray studies and was used for cRNA generation and hybridization to the Affymetrix HG-U95Av2 GeneChip. The anatomic profile of platelet RNAs from 3 healthy male donors was determined by using Affymetrix software. Of the 12 599 probe sets imprinted onto the chip, a maximum of 2147 (17%) transcripts were computationally identified as "present" by the Affymetrix software, 152 (1.2%) were equivocal, and nearly 82% were absent. As a fraction of the total genes present on the chip, the percentage of platelet-expressed genes (15%-17%) was generally lower than that obtained from other human cell types in which 30% to 50% of genes are present as determined by Affymetrix software (J. Schwedes, personal communication, May 2002). The "limited number" of platelet-expressed transcripts presumably reflects the lack of ongoing gene transcription in the anucleate platelet. Because less than 1% of circulating red blood cells contain residual RNA, it is unlikely that any of these transcripts are erythrocyte derived, although this was formally addressed by isolating total cellular RNA from 20 mL of whole blood (corresponding to an ~3-log fold excess of erythrocytes than that identified in our final sample). The total cellular yield of RNA from this starting material was approximately 250 ng, suggesting that less than 1 ng erythrocyte-derived RNA was present in the purified platelet preparations. Despite this, however, both α- and β-globin transcripts—along with both the ferritin heavy and light chains—were identified as abundant transcripts (Table 2). Although the most parsimonious explanation would be residual contaminating reticulocytes, this is not supported by our erythrocyte contamination estimates, and their significance remains unresolved.

As a means of better dissecting the molecular anatomy of the platelet, expressed genes were grouped on the basis of assigned

Table 2. Top 50 human platelet-expressed genes

| Accession no. | Gene symbol | AD values, range* | Gene transcript† | Leukocyte expression‡ |
|---------------|-------------|-------------------|---|-----------------------|
| M17733 | TMSB4X | 140 142-307 852 | Thymosin β 4 mRNA, complete cds | + |
| X99076 | NRGN | 101 510-148 279 | Neurogranin gene | + |
| M25079 | HBB | 40 839-229 556 | β -globin mRNA, complete cds | + |
| M25915 | CLU | 84 720-140 246 | Complement cytolysis inhibitor (clusterin) complete cds | - |
| J04755 | FTHP1 | 82 980-148 621 | Fertilin H processed pseudogene, complete cds | - |
| D78361 | OAZ1 | 73 098-118 140 | mRNA for ornithine decarboxylase antizyme | - |
| X04409 | GNAS | 77 761-94 781 | mRNA for coupling protein G(s) α -subunit (alpha-S1) | - |
| M25897 | PF4 | 62 811-126 908 | Platelet factor 4 mRNA, complete cds | - |
| AB021288 | B2M | 61 689-108 921 | β 2-microglobulin | + |
| X00351 | ACTB | 25 143-73 775 | mRNA for β -actin | - |
| D21261 | TAGLN2 | 76 687-101 931 | mRNA for KIAA0120 gene | + |
| AL031670 | FTLL1 | 69 865-99 966 | Fertilin, light polypeptide 1 | + |
| U59632 | GPIIB | 41 404-110 328 | Platelet glycoprotein IIb chain mRNA | - |
| M21121 | CCL5 | 47 308-106 399 | T-cell-specific protein (RANTES) mRNA, complete cds | - |
| X13710 | GFX1 | 41 318-96 878 | Unspliced mRNA for glutathione peroxidase | - |
| J00153 | HBA1 | 21 326-144 201 | Alpha globin gene cluster on chromosome 16 | + |
| M22919 | MYL6 | 46 337-106 833 | Nonmuscle/smooth muscle alkali myosin light chain gene | + |
| L20941 | FTH1 | 52 787-74 763 | Fertilin heavy chain mRNA, complete cds | - |
| J03040 | SPARC | 51 156-74 261 | SPARC/osteonectin mRNA, complete cds | - |
| X56009 | GNAS | 45 543-72 096 | GSA mRNA for α subunit of GsGTP binding protein | - |
| X58536 | HLA | 31 183-82 613 | mRNA for major HLA class I locus C heavy chain | + |
| M54995 | PPBP | 46 571-67 169 | Connective tissue activation peptide III mRNA | - |
| U34995 | GAPD | 35 095-70 250 | Normal keratinocyte subtraction library mRNA, clone H22a | + |
| L40399 | MLM3 | 32 107-73 364 | Clone zap112 (mult. protein homolog 3) mRNA | - |
| X77548 | NCOA4 | 31 452-61 036 | cDNA for RFG (RET proto-oncogene RET/PTC3) | - |
| U90551 | H2AFL | 35 086-51 892 | Histone 2A-like protein (H2Afl) mRNA | - |
| M11353 | H3F3A | 31 614-55 813 | H3.3 histone class C mRNA | - |
| Z12962 | RPL41 | 36 003-54 853 | mRNA for homologue to yeast ribosomal protein L41 | + |
| X06956 | TUBA1 | 20 988-61 798 | HALPHA 44 gene for α -tubulin | - |
| AB028950 | TLN1 | 24 571-58 611 | mRNA for KIAA 1027 protein | - |
| Y12711 | PGRMC1 | 33 680-43 174 | mRNA for putative progesterone binding protein | - |
| M16279 | MIC2 | 30 894-48 166 | Integrated membrane protein (MIC2) mRNA | - |
| D78577 | YWHAH | 24 785-50 437 | Brain 14-3-3 protein β -chain | - |
| AF070585 | TOP3B | 20 027-67 945 | Clone 24675, unknown cDNA | + |
| AA524802 | Unknown | 23 846-39 481 | CDNA, IMAGE clone 954213 | - |
| AB009010 | UBC | 28 745-38 389 | mRNA for polyubiquitin UbC | + |
| X57985 | H2AFO | 21 678-52 108 | Genes for histones H2B, 1 and H2A | - |
| X54304 | MLCB | 25 733-34 109 | mRNA for myosin regulatory light chain | - |
| M14539 | F13A1 | 23 691-48 474 | Factor XIII subunit α -polypeptide mRNA, 3' end | - |
| AJ540958 | Unknown | 24 872-41 118 | cDNA, PEC 1.2_15_HOI.1 5' end/clone | - |
| AL050396 | FLNA | 13 634-55 235 | cDNA DKF2p 586K1720 | - |
| X56841 | HLA-E | 12 890-49 327 | Nonclassical MHC class I antigen gene | - |
| M26252 | PKM2 | 15 450-47 786 | TCB (cytosolic thyroid hormone-binding protein) | - |
| M14630 | PTMA | 19 314-45 088 | Prothymosin alpha mRNA | - |
| AF045229 | RGS10 | 19 156-34 243 | Regulator of G protein signaling 10 mRNA | - |
| AA477898 | Unknown | 16 863-44 756 | cDNA, Z834108.11 5' end | - |
| X95404 | FLI1 | 15 216-37 456 | mRNA for nonmuscle type coflin | - |
| M34480 | ITGA2B | 8 627-45 495 | Platelet glycoprotein IIb (GPIIb) mRNA | - |
| Z83738 | H2BFE | 18 001-31 306 | H2BFE gene | - |
| L19779 | H2AFO | 17 319-38 951 | Histone H2A.2 mRNA, complete cds | - |

*Gene expression quantifications were calculated as the average difference (AD) value (matched versus mismatched oligonucleotides) for each probe set using Affymetrix GeneChip software, version 4.01. The range of values from 3 distinct platelet microarrays is shown; the normalization value for all microarray analyses was 250.

†Transcripts are rank-ordered (highest to lowest) using BRB-ArrayTools software by log-intensities of AD values obtained from 3 different healthy donors; 33 of the top 40 transcripts were listed among the top 50 in all 3 microarray sets.

‡Leukocyte expression was determined by microarray analysis using purified peripheral blood leukocytes, followed by construction of rank-intensity plots for comparison to platelet top 50 transcripts.²⁰ Top leukocyte-derived transcripts identified within the ranked top 50 platelet transcripts are depicted by a (+) present, or (-) absent, cds indicates coding sequence.

gene annotations, and this analysis was used to provide a panoramic definition of the platelet transcriptome. Of the genes that could be cataloged within assigned "clusters," those involved in metabolism (11%) and receptor/signaling (11%) represented the largest groups. Also evident in these analyses is the relatively large percentage of genes involved in functions unrelated to these key groups (ie, miscellaneous, 25%), and the overrepresentation of genes with unknown function (32%) as annotated by Affymetrix

and RefSeq databases.²¹ These results identify a vast array (nearly one half) of platelet genes (and gene products) that presumably have important, but poorly characterized functions, in platelet and/or megakaryocyte biology.

Although microarray analysis is not truly quantitative, rank-ordering using the mean log-intensities from 3 independent microarray analyses allowed for the categorization of the top platelet transcripts (Table 2). Computational analyses demonstrated that

only 10 of the top 100 genes were discordant among the 3 platelet microarrays, although 71 of 100 genes were discordant between platelet and leukocyte arrays. An inventory of the top 50 platelet genes is listed in Table 2, which also delineates those found to be highly expressed in peripheral blood leukocytes by parallel microarray experiments with this purified cellular fraction (data not shown). Further analysis of these cell subsets demonstrated that approximately 25% ($n = 547$) of the total platelet transcripts were platelet restricted. Furthermore, only 10 of the 50 most highly expressed genes were found to overlap, confirming the distinct cellular profiles of each transcriptome. Of the 12 overlap genes, 3 corresponded to globin or ferritin chains (again suggesting the presence of contaminating reticulocytes in both purified fractions), and another 4 were involved in actin cytoskeletal reorganization and human leukocyte antigen (HLA) expression, gene products that regulate critical functions in both cell types. Given the importance of cytoskeletal reorganization in downstream platelet activation events, it is not unexpected that components of the actin machinery system would demonstrate prominent transcript expression. Previous estimates suggest that 20% to 30% of the total platelet proteome is comprised of actin with other components such as actin-binding protein, myosin, and talin accounting for an additional 2% to 5% of the total protein.^{1,22} The mRNAs encoding the actin-related machinery are overrepresented in our microarray analysis, with 8 such transcripts found among the 50 highest platelet-expressed genes. Interestingly thymosin $\beta 4$ demonstrated the highest expression pattern. In unstimulated platelets, 30% to 40% of actin is polymerized as F-actin,²² whereas the balance of actin monomers (G-actin) are polymerization inhibited by sequestering proteins such as profilin (100 μM) and thymosin $\beta 4$ (600 μM).²³ The high thymosin $\beta 4$ transcript expression not only correlates with its known abundance in platelets but also supports the importance of actin inhibitory proteins in maintaining the nonstimulated state of circulating platelets.

Platelet SAGE analyses

Although these initial studies identified the distribution and relative expression patterns of the genes within the Affymetrix data set, they do not allow for analyses of genes that are unrepresented by these oligonucleotide chips. Unlike closed microarray profiling strategies, SAGE is an open architectural system that is ideally suited for novel gene and pathway identification. Accordingly, the platelet RNA used for microarray studies was used for platelet SAGE. A total of 2033 tags were initially cataloged, of which 1800 (89%) corresponded to mitochondrial-derived genes. These results were quite different from those obtained by microarray analyses, but the discrepancy can be resolved by the nonrepresentation of the mitochondrial genome on the gene chip. The mitochondrial genome is a compact approximately 16.6-kilobase (kb) sequence encoding 13 genes and 2 ribosomal subunits.²⁴ Primary mitochondrial transcripts are polycistronic and typically contain premature termination or unpredictable splice sites, resulting in multiple polyadenylated transcripts from individual genes.^{24,25} Indeed, the overall distribution of platelet-derived mitochondrial SAGE tags is quite similar to that found in muscle.²⁵ All 13 genes containing *Nla*III sites were detected, whereas neither of the non-*Nla*III-containing genes were identified (nicotinamide adenine dinucleotide [NADH] dehydrogenase subunit 4L and adenosine triphosphatase [ATPase] 8). Most of the tags were from the 16S and 12S ribosomal RNAs—which collectively accounted for 68% of the total mitochondrial tags—with the fewest tags represented by NADH dehydrogenase subunits 3, 5, 6, and cytochrome c oxidase 1

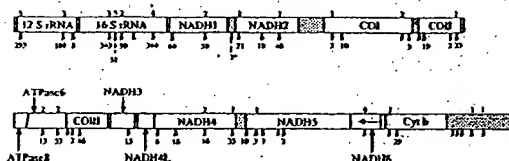


Figure 3. Schema of the mitochondrial genome with SAGE tag distributions (only tags with identical matches are displayed). The abundance of the SAGE tags ($n = 1800$) at individual *Nla*III sites (arrows) within the mitochondrial heavy strand is shown on the bottom, whereas those tags corresponding to the mitochondrial light strand are delineated above the arrows (the presence of an unaccompanied arrow implies no SAGE tags at that *Nla*III site). The gene products of mt-DNA (RefSeq accession no. NC_001807) are delineated by the open rectangles, whereas stippled boxes represent tRNA genes and control regions (the single tag represented by the [•] refers to mitochondrial transfer RNA-serine). Note that NADH6 is encoded by the light strand and that there are no *Nla*III sites within the ATPase8 gene segment. COX1, cytochrome c oxidase subunit; Cyt b, cytochrome b.

(Figure 3). The NADH dehydrogenase subunit 6 RNA is the only mRNA encoded by the light (L) strand of mitochondrial DNA and was the least abundantly detected transcript.

The unusually high preponderance of mitochondrial-derived genes is not inconsistent with the known enrichment of these genomes in human platelets,^{1,24} and presumably reflects persistent transcription from the mitochondrial (mt) genome in the absence of nuclear-derived transcripts. This overrepresentation of mtDNA in platelets is considerably greater than that of its closest cell type (skeletal muscle), in which mt genomes represent approximately 20% to 25% of all SAGE tags.²⁵ Interestingly, the energy metabolism of platelets is not dissimilar from that of skeletal muscle, both cell types actively using glycolysis and large amounts of glycogen for ATP generation.²⁶ Like muscle, platelets are metabolically adapted to rapidly expend large amounts of energy required for aggregation, granule release, and clot retraction. Similar to the situation in all eukaryotic cells, platelet mitochondria represent the primary source of ATP, which is generated from oxidative phosphorylation reactions occurring within these organelles. Mitochondria are also responsible for most of the toxic reactive oxygen species generated as by-products of oxidative phosphorylation and are central regulators of the apoptotic process in other cellular types. The mtDNA encodes polypeptides found within 4 of the 5 multifunctional complexes that regulate oxidative phosphorylation within the platelet mitochondria.²⁷ Whether the continued generation of these polypeptides has a role in platelet energy metabolism and/or the apoptotic mechanisms regulating platelet survival remains speculative, although not inconsistent with our observations.

Comparative analysis of SAGE and microarray transcript abundance

Complete SAGE libraries require the sequencing of up to 30 000 tags for an exhaustive cataloging of individual mRNAs, especially those with limited copy numbers.^{13,28} Given the preponderance of mt-derived transcripts, comparable sampling would have required sequence analysis of nearly 300 000 SAGE tags, an inordinate number for comprehensive analysis of the platelet transcriptome. For platelets, alternative methodologies incorporating subtractive SAGE will be required for more comprehensive transcript profiling.²⁹ Our initial sampling of nonmitochondrial genes remains informative, however, and entirely consistent with the results of platelet microarray studies. As shown in Table 3, SAGE tags for the genes encoding thymosin $\beta 4$, $\beta 2$ -microglobulin, neurogranin, and the platelet glycoprotein Ib β polypeptide were among the most frequently identified platelet genes, similar to the rank-ordered results determined by microarray analysis. To formally confirm the

Table 3. SAGE-identified nonmitochondrial tags

| Frequency | CATG + SAGE tags* | Accession no.† | Gene | Microarray‡ |
|-----------|--------------------|----------------|---|-------------|
| 26 | GTTGTGGTTAATCTGGT | NM_004048.1 | β 2-microglobulin (B2M), mRNA | PPP |
| 21 | TTGGTGAAGGAAGAAGT | NM_021109.1 | Thymosin β 4; X chromosome (TMSB4X), mRNA | P |
| 8 | AGCTCCGCAGCCAGGTC | NM_002620.1 | Platelet factor 4 variant 1 (PF4V1), mRNA | P |
| 8 | AGCTCCGCAGCCGGGTT | NM_002619.1 | Platelet factor 4 (PF4), mRNA | P |
| 7 | TGTATAAGACAACCTC | NM_002704.1 | Proplatelet basic protein (β -thromboglobulin) | Pp |
| 5 | GGGCACAATGCGGTCCA | NM_000407.1 | Glycoprotein Ibb polypeptide, mRNA | P |
| 3 | AGGTAATAAAAGGTAAT | NM_003512.1 | H2A histone family, member L (H2AFL), mRNA | P |
| 3 | AGTGGCAAGTAAATGGC | NM_021914.2 | Cofilin 2 (muscle) (CFL2), mRNA | N/A |
| 3 | TGACTGTCTGGGTTGG | NM_006176.1 | Neurogranin (protein kinase C substrate, RC3) mRNA | P |
| 3 | TTGGGGTTTCCTTTACC | NM_002032.1 | Ferritin, heavy polypeptide 1 (FTH1), mRNA | P |
| 2 | CCCTTGACTACTACCTAT | NM_025158.1 | Hypothetical protein FLJ22251 (FLJ22251), mRNA | N/A |
| 2 | CCTGTAAACCCAGCTAC | NM_032779.1 | Hypothetical protein FLJ14397 (FLJ14397), mRNA | N/A |
| 2 | CTTGTAAGTCCAGCTAC | NM_017962.1 | Hypothetical protein FLJ20825 (FLJ20825), mRNA | N/A |

*Unique tags identified more than once.

†Refers to the RefSeq accession no.²¹ Note that this number does not necessarily correspond to the accession no. provided by Affymetrix software annotations (Table 1).

‡Presence (P) or absence (A) is based on results from 3 distinct platelet microarray experiments. Capitalized "P" designates a gene that is in the top 50 on all 3 microarray experiments, whereas small "p" designates those transcripts not in the top 50. Two of the genes (β 2-microglobulin and β -thromboglobulin) are represented by 3 and 2 probe sets, respectively, on the HG-U95Av2 gene chip; for β 2-M, all 3 probe sets were in the top 50 genes, whereas for thymosin β 4 1 of 2 was in the top 50 for all experiments (the other probe set was in the top 75 for all experiments). N/A indicates oligonucleotide not present on Affymetrix HG-U95Av2 gene chip.

results independently obtained by SAGE and microarray analysis, quantitative RT-PCR was completed by using oligonucleotide primers specific for 2 abundant mitochondrial transcripts, 16S rRNA and NADH2 thymosin β 4 (high-abundance by microarray and SAGE), 2 incompletely characterized high-abundance transcripts (neurogranin and clusterin; see "Protein immunoblot analysis of platelet clusterin and neurogranin"), a low-abundant transcript (T-cell receptor β -polypeptide), and the genes encoding proteins with well-established quantitative determinations (ie, glycoprotein $\alpha_{IIb}\beta_3$ [\sim 50 000 receptors/platelet]; protease-activated receptor-1 (PAR1) [\sim 1800 receptors/platelet]).¹ As shown in Figure 4, these analyses reveal excellent concordance between SAGE and microarray studies, demonstrating the predominant frequency of the mitochondrial-derived 16S rRNA/NADH2 transcripts, with incrementally lower expression of other transcripts as initially demonstrated by microarray (16S > NADH2 > thymosin β 4 > neurogranin > clusterin > $\alpha_{IIb}\beta_3$ > PAR1 > TCR β).

Given the small number of nonmitochondrial SAGE tags available for analysis ($n = 233$), limited conclusions can be drawn using traditional (nonsubtraction) platelet SAGE libraries as pre-

sented here. Overall, a total of 126 unique tags were identified, the majority of which (94) were represented only once. Of the total unique tags, nearly one half represented novel genes not present on the Affymetrix U95Av2 GeneChip. Of the genes with unique tags identified more than once, there was excellent concordance with microarray expression analysis, with nearly all of the SAGE tags in Table 3 corresponding to platelet top 75 microarray transcripts. The platelet factor (PF) 4 variant represents a single aberration because this was rank-ordered approximately 350 by microarray, although its SAGE tag frequency was identical to that of the predominant PF4 transcript. The lack of extensive nonmitochondrial SAGE sampling precludes any further extrapolations from this apparent aberration. Of note, a subset of these tags had long poly(A) tracts, although they all corresponded to genes identified in the RefSeq database.²¹ We cannot exclude the possibility of a SAGE artifact for this small subset of tags (\sim 2%, representing 46 of 2033 tags), although the authenticity of the vast majority of tags (\sim 98%) clearly validates the methodology. These tags are most likely explained by the unique biology of the platelet (ie, mRNA decay in the absence of de novo transcription) or to mRNA degradation occurring during the extensive purification methods. In summary, even with a remarkably limited sampling, the power of this approach in gene identification of relatively abundant and less abundant transcripts is evident. It is clear, however, given the unique molecular anatomy of the platelet (ie, abundance of mitochondrial transcripts), that SAGE adaptations will be required for more comprehensive genetic profiling.²⁹

Protein immunoblot analysis of platelet clusterin and neurogranin

Although most of the "most abundant" transcripts would conform to a priori predictions for platelet-expressed mRNAs, a number of transcripts were identified that had been poorly characterized in human platelets. To further establish the authenticity of highly expressed transcripts such as clusterin and neurogranin, confirmatory protein analyses were completed. As shown in Figure 5, both proteins were clearly detected in purified platelet lysates; furthermore, their cellular platelet distributions conformed to those predicted based on previously proposed functions. Note for example that clusterin—functionally characterized as a complement lysis inhibitor able to block the terminal complement reaction—is primarily expressed on the extracellular platelet membrane.³⁰

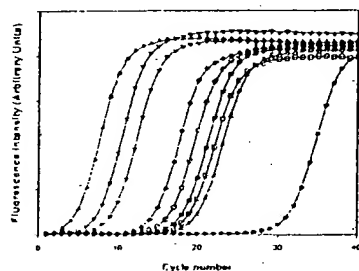


Figure 4. Quantitative real-time RT-PCR analysis of platelet transcripts. Real-time RT-PCR was completed by using purified platelet RNA and oligonucleotide primer pairs specifically designed using Primer3 software to generate similarly-sized (\sim 200-bp) PCR products, optimized to the same annealing temperature. In graph, (□) represents $\alpha_{IIb}\beta_3$, (■) represents $\alpha_{IIb}\beta_3$, (△) represents PAR1, (▲) represents 16S rRNA, (▼) represents NADH2, (▽) represents thymosin, (○) represents clusterin, (●) represents neurogranin, and (●) represents TCR β . Curves are representative of one complete set of experiments (repeated twice), and line plots reflect average determinations from 3 wells performed in parallel with SEM less than 1% for all data points.

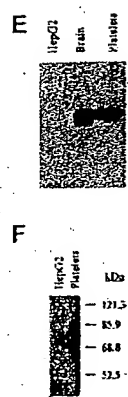
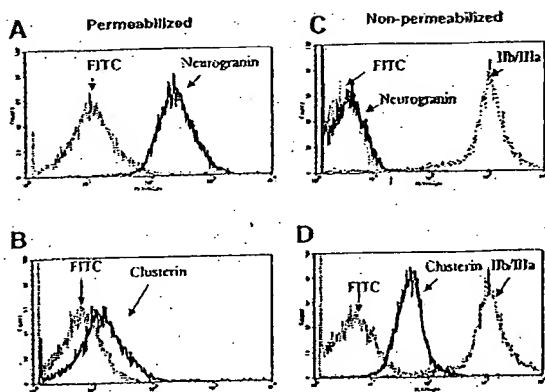


Figure 5. Immunocytology of platelet neurogranin and clusterin. (A-D) Gel-filtered platelets were either fixed in 3.7% formaldehyde (non-permeabilized) or fixed with permeabilization in the presence of 0.1% Triton-X, followed by flow cytometric analysis using anti-neurogranin, anti-IIs/IIa, or anti-clusterin antibodies and the FITC-conjugated species-specific secondary antibody (in C, the FITC-conjugated anti-rabbit and anti-mouse controls are essentially superimposed). (E-F) Ten micrograms of solubilized HepG2 cells (hepatocyte cell line), human brain, or purified platelet lysates were analyzed by SDS-PAGE, and immunoblot analysis were completed by using 1:1000 dilutions of either anti-neurogranin (18% SDS-PAGE) or anti-clusterin (8% SDS-PAGE) antibodies. The anti-clusterin antibody recognized 2 platelet immunoreactive species under shorter exposure. Although the relative neurogranin and clusterin protein abundances are suboptimally quantified by these analyses, platelet clusterin appears to demonstrate considerable expression when compared with that previously identified in hepatocytes.³¹

Given the importance of complement activation in platelet destruction, the prominent expression of cell-surface clusterin might suggest a role for this protein in normal and pathologic events regulating platelet survival. Interestingly, a clusterin-deficient knockout mouse has been generated that demonstrates enhanced cardiac dysfunction in a model of autoimmune myocarditis.³¹ Although these mice apparently have normal baseline hemograms (B. Aronow, personal communication, October 2002), it remains unestablished if they would be predisposed to immune-type thrombocytopenia in systemic models of autoimmunity.

Similarly, the gene encoding an intracellular effector protein that may have key roles in downstream platelet activation events has now been demonstrated to have abundant transcript expression in human platelets. Neurogranin is a highly expressed platelet transcript with its gene product demonstrating a primarily intracellular pattern of distribution. Neurogranin is generally described as a brain-specific, Ca^{2+} -sensitive calmodulin-binding phosphoprotein that is preferentially expressed in neuronal cell bodies and dendrites.^{32,33} It is a specific protein kinase C (PKC) substrate that can also be modified by nitric oxide and other oxidants to form intramolecular disulfide bonds. Both its phosphorylation and oxidation state attenuate its binding affinity for calmodulin.³³ In stimulated platelets, PKC generation is linked to various activation pathways such as calcium-regulated kinases, mitogen-activated protein (MAP) kinases, and receptor tyrosine kinases.¹ Thus, these observations suggest that platelet neurogranin may function as a previously unidentified component of a PKC-dependent activation pathway coupled to one (or more) of these effector proteins.

Discussion

These data provide documentation for a unique platelet mRNA profile that may provide a tool for analyzing platelet molecular networks. Nonetheless, the molecular analysis of the platelet transcriptome may be confounded by the constant decay of mRNAs in the absence of new gene transcription, a situation that may, for example, limit the identification of low-abundance transcripts. Similarly, because the circulating platelet pool contains

a mixed population of variably aged platelets, a "static" mRNA profile represents an average of this heterogeneous blood pool. Despite these potential limitations, the combination of genomic and proteomic technologies are likely to provide powerful tools for the global analysis of platelet function. Current strategies for cataloging "whole cellular proteomes" are generally accomplished by using 2 developing methodologies: (1) high resolution 2-dimensional polyacrylamide gel electrophoresis (2-DE) with mass spectrometric sequence identification,³⁴ and (2) microcapillary liquid chromatography with tandem mass spectrometry ($\mu\text{LC-MC/MC}$).³⁵ Further modifications of both procedures have been devised for direct comparative studies between 2 cellular proteomes. The introduction of 2-DE differential gel electrophoresis has now made it possible to detect and quantify differences between experimental sample pairs resolved on the same 2-dimensional gel.³⁶ Likewise, the application of isotope-coded affinity tags to $\mu\text{LC-MC/MC}$ represent a novel means of quantitative analyses between cellular proteomes.³⁷ The success of both approaches relies on the availability of comprehensive genomic databases and mathematical algorithms for optimal protein identification. Indeed, mathematical modeling studies have demonstrated the need to delineate both protein and mRNA expression levels for optimal definition of intracellular networks.³⁸ Our data present an initial framework for delineating platelet function by defining the molecular anatomy of human platelets, information that is likely to provide important clues into the dynamic protein interactions regulating normal and pathologic platelet functions. Furthermore, because the platelet transcriptome mirrors the mRNAs derived from precursor megakaryocytes, these analyses may provide insights into the biochemical and molecular events regulating megakaryocytopoiesis and/or proplatelet formation.

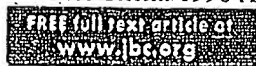
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References

- Steinberg P, Hill R. Platelets and megakaryocytes. In: Le R, et al, eds. *Wintrobe's Clinical Hematology*. Baltimore, MD: Williams & Wilkins; 1999.
- Newman P, Gorski J, White G, Gidwitz S, Cretney C, Aster R. Enzymatic amplification of platelet-specific messenger RNA using the polymerase chain reaction. *J Clin Invest*. 1988;82:739-743.
- Kieffer N, Guichard J, Farci J, Vainchenker W, Breton-Gorius J. Biosynthesis of major platelet proteins in human blood platelets. *Eur J Biochem*. 1987;164:189-195.
- Weyrich A, Dixon D, Pablo R, et al. Signal-dependent translation of a regulatory protein, Bcl-2, in activated human platelets. *Proc Natl Acad Sci U S A*. 1998;95:5556-5561.
- Benecke BJ, Ben Ze'ev A, Penman S. The control of mRNA production, translation and turnover in

- suspended and reattached anchorage-dependent fibroblasts. *Cell*. 1978;14:931-939.
6. Pabla R, Weyrich AS, Dixon DA, et al. Integrin-dependent control of translation: engagement of integrin $\alpha 5 \beta 1$ regulates synthesis of proteins in activated human platelets. *J Cell Biol*. 1999;144:175-184.
 7. Chicurel ME, Singer RH, Meyer CJ, Ingber DE. Integrin binding and mechanical tension induce movement of mRNA and ribosomes to focal adhesions. *Nature*. 1998;392:730-733.
 8. Lindemann S, Tolley N, Eyre J, Kraiss L, Mahoney T, Weyrich A. Integrins regulate the intracellular distribution of eukaryotic initiation factor 4E in platelets. *J Biol Chem*. 2001;276:33947-33951.
 9. Brown EJ, Schreiber SL. A signaling pathway to translational control. *Cell*. 1996;86:517-520.
 10. Rinder H, Schuster J, Rinder C, Wang C, Schweidter H, Smith B. Correlation of thrombosis with increased platelet turnover in thrombocytosis. *Blood*. 1998;91:1288-1294.
 11. Richards E, Baglin T. Quantitation of reticulated platelets: methodology and clinical application. *Br J Haematol*. 1995;91:445-451.
 12. Robinson M, Mackie I, Khai K, et al. Flow cytometric analysis of reticulated platelets: evidence for a large proportion of non-specific labelling of dense granules by fluorescent dyes. *Br J Haematol*. 1998;100:351-357.
 13. Velculescu V, Zhang L, Vogelstein B, Kinzler K. Serial analysis of gene expression. *Science*. 1995;270:484-487.
 14. Zhang L, Zhou W, Velculescu V, et al. Gene expression profiles in normal and cancer cells. *Science*. 1997;276:1268-1272.
 15. Morin PJ, Sparks AB, Korinek V, et al. Activation of beta-catenin-Tcf signaling in colon cancer by mutations in beta-catenin or APC. *Science*. 1997;275:1787-1790.
 16. Bahou W, Campbell A, Wicha M. cDNA cloning and molecular characterization of MSE55: a novel human serum constituent protein that displays bone marrow stromal endothelial cell-specific expression. *J Biol Chem*. 1992;267:13986-13992.
 17. Bahou W, Collier B, Potter C, Norton K, Kutok J, Goligorsky M. The thrombin receptor extracellular domain contains sites crucial for peptide ligand-induced activation. *J Clin Invest*. 1993;91:1405-1413.
 18. Heid C, Stevens J, Livak K, Williams P. Real-time quantitative PCR. *Genome Res*. 1996;6:986-994.
 19. Dunn J, McCorkle S, Praissman L, et al. Genome signature tags (GSTs): a system for profiling genomic DNA. *Nucleic Acid Res*. 2001;29:137-140.
 20. Kroll T, Wolf S. Ranking: a closer look on globalisation methods for normalization of gene expression arrays. *Nucleic Acids Res*. 2002;30:e50.
 21. Pruitt KD, Maglott DR. RefSeq and LocusLink: NCBI gene-centered resources. *Nucleic Acids Res*. 2001;29:137-140.
 22. Fox JE, Boyles JK, Reynolds CC, Phillips DR. Actin filament content and organization in unstimulated platelets. *J Cell Biol*. 1984;98:1985-1991.
 23. Safer D, Etzinger M, Nachmias VT. Thymosin beta 4 and Fx, an actin-sequestering peptide, are indistinguishable. *J Biol Chem*. 1991;266:4029-4032.
 24. Wallace DC. Mouse models for mitochondrial disease. *Am J Med Genet*. 2001;106:71-93.
 25. Welle S, Bhatt K, Thornton C. Inventory of high-abundance mRNAs in skeletal muscle of normal men. *Genome Res*. 1999;9:506-513.
 26. Karparkin S, Charnatz A, Langer RM. Glycogenesis and glycconeogenesis in human platelets. Incorporation of glucose, pyruvate, and citrate into platelet glycogen; glycogen synthetase and fructose-1,6-diphosphatase activity. *J Clin Invest*. 1970;49:140-149.
 27. Raha S, Robinson BH. Mitochondria, oxygen free radicals, and apoptosis. *Am J Med Genet*. 2001;106:62-70.
 28. Yu J, Zhang L, Hwang P, Rago C, Kinzler K, Vogelstein B. Identification and classification of p53-regulated genes. *Proc Natl Acad Sci U S A*. 1999;96:14517-14522.
 29. Wang E, Miller L, Ohnmacht G, Liu E, Marincola F. High-fidelity mRNA amplification for gene profiling. *Nat Biotechnol*. 2000;18:157-159.
 30. Kirschbaum L, Sharpe JA, Murphy B, et al. Molecular cloning and characterization of the novel, human complement-associated protein, SP-40,40: a link between the complement and reproductive systems. *EMBO J*. 1989;8:711-718.
 31. McLaughlin L, Zhu G, Mistry M, et al. Apolipoprotein J/chapterin limits the severity of murine autoimmune myocarditis. *J Clin Invest*. 2000;105:1113.
 32. Martinez DA, Perez JL, Benjal J, Coloma A. Structure, organization, and chromosomal mapping of the human neurogranin gene (NRGN). *Genomics*. 1997;41:243-249.
 33. Wu J, Li J, Huang K, Huang F. Attenuation of PKC and PKA signal transduction in the neurogranin knockout mouse. *J Biol Chem*. 2002;277:19498-19505.
 34. Gygi S, Rochon Y, Franza B, Aebersold R. Correlation between protein and mRNA abundance in yeast. *Mol Cell Biol*. 1999;19:1720-1730.
 35. Link A, Eng J, Schieltz DM, et al. Direct analysis of protein complexes using mass spectrometry. *Nat Biotechnol*. 1999;17:676-682.
 36. Unlu M, Morgan M, Minden J. Difference gel electrophoresis: a single gel method for detecting changes in protein extracts. *Electrophoresis*. 1997;18:2071-2077.
 37. Gygi S, Rist B, Gerber SA, Turecek F, Gelb MH, Aebersold R. Quantitative analysis of complex protein mixtures using isotope-coded affinity tags. *Nat Biotechnol*. 1999;17:994-999.
 38. Halizmanikatis V, Lee K. Dynamical analysis of gene networks requires both mRNA and protein expression information. *Metabol Eng*. 1999;1:275-281.



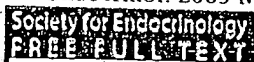
Overexpression of a DEAD box protein (DDX1) in neuroblastoma and retinoblastoma cell lines.

Godbout R, Packer M, Bie W.

Department of Oncology, Cross Cancer Institute and University of Alberta, 11560 University Ave., Edmonton, Alberta T6G1Z2, Canada.

The DEAD box gene, DDX1, is a putative RNA helicase that is co-amplified with MYCN in a subset of retinoblastoma (RB) and neuroblastoma (NB) tumors and cell lines. Although gene amplification usually involves hundreds to thousands of kilobase pairs of DNA, a number of studies suggest that co-amplified genes are only overexpressed if they provide a selective advantage to the cells in which they are amplified. Here, we further characterize DDX1 by identifying its putative transcription and translation initiation sites. We analyze DDX1 protein levels in MYCN/DDX1-amplified NB and RB cell lines using polyclonal antibodies specific to DDX1 and show that there is a good correlation with DDX1 gene copy number, DDX1 transcript levels, and DDX1 protein levels in all cell lines studied. DDX1 protein is found in both the nucleus and cytoplasm of DDX1-amplified lines but is localized primarily to the nucleus of nonamplified cells. Our results indicate that DDX1 may be involved in either the formation or progression of a subset of NB and RB tumors and suggest that DDX1 normally plays a role in the metabolism of RNAs located in the nucleus of the cell.

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Modulation of gap junction mediated intercellular communication in TM3 Leydig cells.

Goldenberg RC, Fortes FS, Crist  ncho JM, Morales MM, Franci CR, Varanda WA, Campos de Carvalho AC.

Institute of Biophysics Carlos Chagas Filho, UFRJ, Brazil.

Long-term modulation of intercellular communication via gap junctions was investigated in TM3 Leydig cells, under low and high confluence states, and upon treatment of the cells for different times with activators of protein kinase A (PKA) and protein kinase C (PKC). Cells in low confluence were readily coupled, as determined by transfer of the dye Lucifer Yellow; on reaching confluence, the cells uncoupled. Western blots and RT-PCR revealed that connexin 43 (Cx43) was abundantly expressed in TM3 Leydig cells and its expression was decreased after the cells achieved confluence. Stimulation of PKA or PKC induced a decrease in cell-cell communication. Staurosporin, an inhibitor of protein kinases, increased coupling and was able to prevent and reverse the uncoupling actions of dibutyryl cAMP and 12-O-tetradecanoyl-phorbol-13-acetate (TPA). Under modulation by confluence, Cx43 was localized to the appositional membranes when cells were coupled and was mainly in the cytoplasm when they were uncoupled. In addition, cAMP and TPA reduced the surface membrane labeling for Cx43, whereas staurosporin increased it. These data show a strong correlation between functional coupling and the membrane distribution of Cx43, implying that this connexin has an important role in intercellular communication between TM3 cells. Furthermore, increased testosterone secretion in response to luteinizing hormone was accompanied by a decrease in intercellular communication, suggesting that gap junction mediated coupling may be a modulator of hormone secretion in TM3 cells.

PMID: 12740021 [PubMed - indexed for MEDLINE]

Expression level of Ubc9 protein in rat tissues.

Golebiowski E, Szulc A, Sakowicz M, Szutowicz A, Pawelczyk T.

Department of Molecular Medicine, Medical University of Gdansk, 80-211 Gdansk, Poland.

Ubc9 is a homologue of the E2 ubiquitin conjugating enzyme and participates in the covalent linking of SUMO-1 molecule to the target protein. In this report we describe a simple and efficient method for obtaining pure human recombinant Ubc9 protein. The purified Ubc9 retained its native structure and was fully active in an in vitro sumoylation assay with the promyelocytic leukaemia (PML) peptide as a substrate. In order to better understand the physiology of Ubc9 protein we examined its levels in several rat tissues. Immunoblot analyses performed on tissue extracts revealed quantitative and qualitative differences in the expression pattern of Ubc9. The Ubc9 protein was present at a high level in spleen and lung. Moderate level of Ubc9 was detected in kidney and liver. Low amount of Ubc9 was observed in brain, whereas the 18 kDa band of Ubc9 was barely visible or absent in heart and skeletal muscle. In heart and muscle extracts the Ubc9 antibodies recognized a 38 kDa protein band. This band was not visible in extracts of other rat tissues. A comparison of the relative levels of Ubc9 mRNA and protein indicated that the overall expression level of Ubc9 was the highest in spleen and lung. In spleen, lung, kidney, brain, liver and heart there was a good correlation between the 18 kDa protein and Ubc9 mRNA levels. In skeletal muscle the Ubc9 mRNA level was unproportionally high comparing to the level of the 18 kDa protein. The presented data indicate that in the rat the expression of the Ubc9 protein appears to have some degree of tissue specificity.

PMID: 14739995 [PubMed - indexed for MEDLINE]



Analysis of mRNA expression and protein abundance data: an approach for the comparison of the enrichment of features in the cellular population of proteins and transcripts

Dov Greenbaum^{3,†}, Ronald Jansen^{1,†} and Mark Gerstein^{1,2,*}

¹Departments of Molecular Biophysics & Biochemistry, ²Computer Science and ³Genetics, 266 Whitney Avenue, Yale University, PO Box 208114, New Haven, CT 06520, USA

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ABSTRACT

Motivation: Protein abundance is related to mRNA expression through many different cellular processes. Up to now, there have been conflicting results on how correlated the levels of these two quantities are. Given that expression and abundance data are significantly more complex and noisy than the underlying genomic sequence information, it is reasonable to simplify and average them in terms of broad proteomic categories and features (e.g. functions or secondary structures), for understanding their relationship. Furthermore, it will be essential to integrate, within a common framework, the results of many varied experiments by different investigators. This will allow one to survey the characteristics of highly expressed genes and proteins.

Results: To this end, we outline a formalism for merging and scaling many different gene expression and protein abundance data sets into a comprehensive reference set, and we develop an approach for analyzing this in terms of broad categories, such as composition, function, structure and localization. As the various experiments are not always done using the same set of genes, sampling bias becomes a central issue, and our formalism is designed to explicitly show this and correct for it. We apply our formalism to the currently available gene expression and protein abundance data for yeast. Overall, we found substantial agreement between gene expression and protein abundance, in terms of the enrichment of structural and functional categories. This agreement, which was considerably greater than the simple correlation between these quantities for individual genes, reflects the way broad categories collect many individual measurements into simple, robust averages. In particular, we found

that in comparison to the population of genes in the yeast genome, the cellular populations of transcripts and proteins (weighted by their respective abundances, the transcriptome and what we dub the translome) were both enriched in: (i) the small amino acids Val, Gly, and Ala; (ii) low molecular weight proteins; (iii) helices and sheets relative to coils; (iv) cytoplasmic proteins relative to nuclear ones; and (v) proteins involved in 'protein synthesis,' 'cell structure,' and 'energy production.'

Supplementary information: <http://genecensus.org/expression/translatome>

Contact: mark.gerstein@yale.edu

INTRODUCTION

High throughput experimentation, measuring mRNA (Schena *et al.*, 1995; Eisen and Brown, 1999; Ferea and Brown, 1999; Lipshutz *et al.*, 1999) and protein expression (Anderson and Seilhauer, 1997; Futcher *et al.*, 1999; Gygi *et al.*, 1999a; Ross-Macdonald *et al.*, 1999; Lopez, 2000; MacBeath and Schreiber, 2000; Nelson *et al.*, 2000; Zhu *et al.*, 2000) are currently the single richest source of genomic information. However, how to best interpret this data is still an open question (Bassett *et al.*, 1996; Wittes and Friedman, 1999; Zhang, 1999; Gerstein and Jansen, 2000; Searls, 2000; Sherlock, 2000; Claverie, 1999; Einarson and Golemis, 2000; Epstein and Butow, 2000; Shapiro and Harris, 2000). Understanding how protein abundance is related to mRNA transcript levels is essential for interpreting gene expression, protein interactions, structures and functions in a cellular system (Hatzimanikatis *et al.*, 1999). Moreover, as protein concentration is the more relevant variable with respect to enzyme activity, it connects genomics to the physical chemistry of the cell (Kidd *et al.*, 2001). Protein abundance may also be invaluable for diagnostics and for determining drug targets (Corthals *et al.*, 2000).

*To whom correspondence should be addressed.

†These authors contributed equally to this work.

Previously, we surveyed the population of protein features—such as folds, amino acid composition, and functions—in yeast, and other recently sequenced genomes (Gerstein, 1997, 1998a,b; Gerstein and Hegyi, 1998; Hegyi and Gerstein, 1999; Das and Gerstein, 2000; Lin and Gerstein, 2000), and we extended this concept to compare the population of features in the yeast transcriptome to that in the genome (Drawid *et al.*, 2000; Jansen and Gerstein, 2000). Others have also done related work (Frishman and Mewes, 1997; Tatusov *et al.*, 1997; Jones, 1998; Wallin and von Heijne, 1998; Frishman and Mewes, 1999; Wolf *et al.*, 1999). Here, we present a new methodology to compare the features of the mRNA expression population with the protein abundance population.

Precise terminology is essential for this comparison. Unfortunately, 'proteome' is used inconsistently. Proteome can logically be used to describe all the distinct proteins in the genome (Qi *et al.*, 1996; Cavalcoli *et al.*, 1997; Fey *et al.*, 1997; Garrels *et al.*, 1997; Gaasterland, 1999; Jones, 1999; Sali, 1999; Tekala *et al.*, 1999; Bairoch, 2000; Cambillau and Claverie, 2000; Doolittle, 2000; Pandey and Mann, 2000; Rubin *et al.*, 2000) and, in this context, it is equivalent to what others may refer to as the coding part of the genome. However, in papers on two-dimensional (2D) electrophoresis, it is often used to describe the sum total of proteins in a cell, taking into account the different levels of protein abundance (Shevchenko *et al.*, 1996; Gygi *et al.*, 2000a; Lopez, 2000; Washburn and Yates, 2000). In an effort to be clear, we propose the term 'translatome' for this second usage of proteome.

With this definition, we are able to refer compactly to three different cellular populations. These are illustrated in Figure 1.

- (i) We use the term *genome* when we refer to the population of open reading frames, where each ORF counts once.
- (ii) We use the term *transcriptome* when we refer to the population of mRNA transcripts. This term was originally coined by Velculescu *et al.* (1997). Note that each ORF may give rise to different numbers of transcripts. Consequently, the transcriptome is essentially the same as the genome but with each ORF weighted by its expression level.
- (iii) The next level is the cellular population of proteins. As each protein represents a translated transcript, we make an analogy with the term transcriptome and use the term *translatome* as described above to describe this third population. Thus, the translatome is a subset of the genome where each ORF is weighted by its associated level of protein abundance.

Note that one could also, less compactly call the translatome a 'weighted proteome.' However, doing so assumes one of the two aforementioned definitions of proteome. To avoid ambiguity, we studiously avoid the use of proteome altogether in the paper.

Differences between the transcriptome and the translatome exist given that transcripts from different genes can give rise to different numbers of proteins, due to different rates of translation and protein degradation. Post-transcriptional modifications further affect the translatome.

In our analysis of the transcriptome and translatome, we focus on global protein features rather than the comparison of individual genes. Previous analyses have shown that differences between mRNA expression and protein abundance levels can be quite dramatic for individual genes. This may either be due to the noise in the data or to fundamental biological processes. However, our analyses show that the variation between transcriptome and translatome is much smaller for global properties that are computed by averaging over the properties of many individual genes.

METHODS

Data sources used

For our analysis we culled many divergent data sets, representing protein abundance and mRNA expression experiments and also other sources of genome annotation. These are all summarized in Table 1.

Biases in the data

The databases that annotate the specific genes may not always be accurate (Ishii *et al.*, 2000). Gene Chip experiments suffer with regard to cross hybridization and the saturation of probes. SAGE data degrades for lowly expressed mRNAs. 2D gels are unable to resolve membrane proteins (approximately 30% of the genome) and basic proteins (Gerstein, 1998c; Krögh *et al.*, 2001). In addition, the procedures for identification and quantification of the protein spots are subject to uncertainties (Haynes and Yates, 2000). Human biases include the lack of low abundance proteins (Fey and Larsen, 2001; Gygi *et al.*, 2000b; Harry *et al.*, 2000) and the differences between laboratories in sample preparation. Our reference expression data set attempts to resolve these problems.

Data set scaling

A reference set for mRNA expression. With many different mRNA expression data sets available, it is worthwhile to integrate them into a single unified reference set, with the intention of reducing the noise and errors contained in the individual data sets and to obtain a unified estimate of the normal expression state in a cell.

We adopt an iterative scaling and merging formalism,

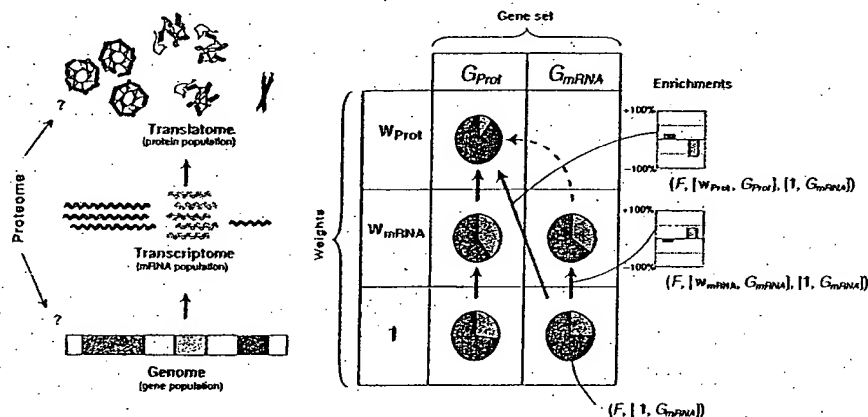


Fig. 1. Schematic overview of the analysis. On the left-side we outline the terms we use to describe the process of gene expression. The coding section of the genome is transcribed into a population of mRNA transcripts called the 'transcriptome.' The transcripts in turn are translated to a population of proteins; we use the term 'translatome' for this protein population rather than the alternative 'proteome' because the latter term may be confounded with the protein complement of the genome (which is not necessarily associated with a quantitative abundance level).

The matrix in the middle schematically shows an analysis of the three stages of expression. In general, we define a protein 'population' as a set of genes associated with a corresponding number of expression or abundance levels ('weights'). In the matrix each row represents a weight and each column a gene set. In particular, we differentiate between the mRNA reference expression set ($G_{mRNA} = G_{Gen}$), which essentially covers the complete genome, and the reference protein abundance set (G_{Prot}) which contains the proteins in data sets 2-DE #1 and 2-DE #2 (see Table 1) because the protein abundance set is a significantly smaller subset of the genome. By definition, this subset contains only proteins that can be identified by 2-D gel electrophoresis and is therefore biased in this sense. The enrichment figures throughout this paper, through a comparison of the right- and left-sides of this figure, show the results of the experimental biases of 2D gels on the data set. Each pie chart represents a composition of a particular protein feature F (for instance, an amino acid composition) in a population (represented by the symbol μ). We can further look at the 'enrichment' of this feature in one population relative to another (represented by the symbol Δ , see Section 'Methods' for an explanation of the formalism).

which we summarize below. We present a more detailed review of the methods on our web site.

We start with the values of one gene chip data set U_i where i is used throughout as a subscript to denote gene number. We then transform the values of the next Gene Chip data set X_i to Y_i with the following non-linear regression: $\min \sum_i (Y_i - U_i)^2$ with $Y_i = AX_i^B$ where A and B are the parameters of the regression. Note that two Gene Chip sets may not be defined for the same set of genes, so we have to perform the fit only over the genes common to both sets. The motivation for scaling is that the dynamic range of observed expression levels varies somewhat between different data sets, although cell types and growth conditions are very similar. Reasons for disparity may include different calibration procedures for relating fluorescence intensity to a cellular concentration (measured in copies of transcripts per cell) or different protocols for harvesting and reverse-transcribing the cellular mRNA.

We then merge and average the data to create a new

reference set V as follows:

$$\text{If } U_i \text{ and } Y_i \text{ are both defined for gene } i \text{ and } \frac{|Y_i - U_i|}{Y_i + U_i} < \alpha$$

$$\text{Then } V_i = \frac{1}{2}(Y_i + U_i)$$

$$\text{Else if only } Y_i \text{ exists, } V_i = Y_i$$

$$\text{Else } V_i = U_i$$

As presented above, where only one data set has a value for the corresponding ORF, we incorporated that value and did not exclude it. When both data sets have values for an ORF, we averaged the values if they were within 15% of each other; otherwise, we just stayed with the original chip data set U_i . We used $\alpha = 15\%$ in order to prevent outliers from skewing the result. This 15% value is a reasonable threshold for excluding outliers though other values (e.g. 10 or 20%) would give similar results (data not shown). Other data sets are subsequently included in the same procedure, continuing the iteration from the new

expression values V_i . The initial iteration starts with the Young Expression Set, as U_i , since we have the highest confidence in its accuracy.

The SAGE data (Velculescu *et al.*, 1997) was not included in the above procedure since it is of a fundamentally different nature. An advantage of the SAGE technology over Gene Chips is that there is no possible signal saturation for high expression levels, as is possible for chips (Futcher *et al.*, 1999). Conversely, SAGE values are less reliable for lowly expressed genes since there is a chance that one might not sequence a SAGE tag corresponding to such a gene altogether. Therefore, if after the last iteration, the average Gene Chip expression level V_i was both above a certain threshold β and below the SAGE expression level S_i for the same gene, it was replaced with the SAGE value; otherwise the average Gene Chip value was kept. This gave us our final expression set w_{mRNA} . Our treatment of the SAGE data is modeled after that in Futcher *et al.* (1999), and like them, we used $\beta = 16$.

This incorporation of the SAGE data into the reference data set ensures that the highly expressed outliers are as accurate as possible.

Rather than plain arithmetic averaging, this overall scaling procedure with the α cutoff avoids 'artificial averages' that combine very different values for a particular gene. Some expression values might be statistical outliers. In addition, it may be possible that the expression levels of a variety of genes can only be within mutually exclusive ranges or modes, such as when two alternative pathways are switched on or off. Simply averaging these would give values that are less representative of the particular mode values. This situation is analogous to that in averaging together an ensemble of protein structures (i.e. from NMR structure determination). Each structure could be stereochemically correct, with all side-chain atoms in predefined rotamer configurations. However, an average of all structures could yield one that is stereochemically incorrect if this involved averaging over particular side-chains in different rotameric states.

With regard to our regression analysis, we have investigated both non-linear and linear fits but found a non-linear procedure to be more advantageous. The non-linear relationship between different expression data sets perhaps reflects saturation in one or more of the Gene Chips—not an uncommon phenomenon. This non-linearity is immediately evident on scatter plots of two data sets against one another (see website). Accordingly, the non-linear fit produces a smaller residual than the linear fit: 98 297 (non-linear) versus 122 182 (linear) for the scaling of the Church data set and 59 828 (non-linear) versus 67 462 (linear) for the Samson data set.

A reference set for protein abundance. We followed a similar procedure to calculate a reference protein abundance set from the two gel electrophoresis data sets. We first scaled the two data sets against the mRNA expression reference data set, getting regression parameters C_j and D_j :

$$\min \sum_i (P_{i,j} - C_j w_{\text{mRNA},i}^{D_j})^2$$

where the subscript j indicates the data set 2-DE #1 or 2-DE #2 respectively; $P_{i,j}$ is the protein abundance value in data set j , and $w_{\text{mRNA},i}$ the corresponding reference expression value, and C_j and D_j are the parameters of the non-linear regression.

Using these parameters, we transformed the values of set 2-DE #2 onto 2-DE #1. Then we combined both sets into the reference protein set w_{Prot} by averaging them, if both values existed. Otherwise, by using the existing value, viz:

$$Q_{i,2} \equiv C_1 \left(\frac{P_{i,2}}{C_2} \right)^{D_1/D_2}$$

$w_{\text{Prot},i} = (P_{i,1} + Q_{i,2})/2$ if both $P_{i,1}$ and $Q_{i,2}$ exist.

Else if only $P_{i,1}$ exists, $w_{\text{Prot},i} = P_{i,1}$

Else if $Q_{i,2}$ exists, $w_{\text{Prot},i} = Q_{i,2}$.

Enrichment of features

Formalism. In the next part of our analysis, we want to group a number of proteins together into various categories based on common features and characterize those features that are enriched in one population relative to another, i.e. the translome population of proteins as measured by 2D gels relative to the transcriptome population of transcripts or the genome population of genes. To this end, we set up a formalism that could be applied universally to all the attributes that we were interested in. Due to the limitations of the experiments, the translome, transcriptome, and genome populations are defined on different sets of genes, and sometimes we want to remove this 'selection bias' by forcing them to be compared on exactly the same set of genes. This is a key aspect of our formalism as presented in Figure 1.

We call an entity like $[w, G]$ a 'population,' where G is a set describing a particular selection of genes from the genome and w is vector of weights associated with each element of this population. In particular, we focus on three main populations here:

- (i) $[1, G_{\text{Gen}}]$ is the population of genes in the genome, all 6280 genes weighted once ($w = 1$);
- (ii) $[w_{\text{mRNA}}, G_{\text{mRNA}}]$ is the observed population of the transcripts in the transcriptome, i.e. the 6249 genes in the reference expression set weighted by their reference expression value;

- (iii) $[w_{\text{Prot}}, G_{\text{Prot}}]$ is the observed cellular population of the proteins in the translome, i.e. the 181 genes in the reference abundance set weighted by their reference abundance value.

(The set of genes in the genome G_{Gen} is approximately equal to the genes in set G_{mRNA} , such that we can use both symbols interchangeably.) We can also use this notation to describe specific experiments—e.g. $[w_{\text{lacZ}}, G_{\text{lacZ}}]$ describes the gene set and weights relating to the transposon abundance set.

Furthermore, we define F_j as the value of a feature F in ORF j . For example, F could be the composition of leucine (a real number) or a binary value (0 or 1) indicating whether an ORF contains a trans-membrane segment. Given these definitions, the weighted average of feature F in population $[w, G]$ is:

$$\mu(F, [w, G]) \equiv \frac{\sum_{j \in G} w_j F_j}{\sum_{j \in G} w_j}$$

The weighted averages of two populations $[w, G]$ and $[v, S]$ can be compared by simply looking at their relative difference Δ :

$$\Delta(F, [v, S], [w, G]) = \frac{\mu(F, [v, S]) - \mu(F, [w, G])}{\mu(F, [w, G])}$$

where v and w are weights for the sets of ORFs S and G respectively. We call Δ the 'enrichment' of feature F because it indicates whether F is enriched (if Δ is positive) or depleted (if Δ is negative) in population $[v, S]$ relative to $[w, G]$.

Usually, the gene set G is defined by the particular experiment, for which the weight w was measured. However, it is also possible to combine the gene set associated with one experiment with expression levels from another set. One may want to do this to compute the enrichment only on the genes common to both populations, for which there are defined values for both w and v , viz: $\Delta(F, [v, S \cap G], [w, S \cap G])$. In practice, this is most relevant for comparing G_{Prot} and G_{mRNA} . Since G_{Prot} is completely a subset of G_{mRNA} , we need not explicitly deal with intersections, if we calculate all statistics directly over G_{Prot} .

One can adjust the weight vectors to take into account different types of averaging. For instance, when computing the amino acid composition ($F = aa$) from the amino acid compositions of individual ORFs $F_j = aa_j$ ($\forall j \in G$), we weight by ORF length. In the case of expression weights, we have:

$$w_j = N_j w_{\text{mRNA}, j} \quad \forall j \in G$$

where N_j is a measure of the length of ORF j (such as the number of amino acids).

On the other hand, when computing the average molecular weight per amino acid, we need to normalize by the number of amino acids per ORF, which is equivalent to choosing the following weights:

$$w_j = \frac{w_{\text{mRNA}, j}}{N_j} \quad \forall j \in G$$

Application of methodology to quantitative abundance sets

Having defined our formalism, we applied it to a diverse set of protein features in yeast.

Amino acid enrichment. As shown in Figure 2a, we used our methodology to measure the enrichment of individual amino acids in both the translome and the transcriptome relative to the genome. We found that three amino acids—valine, glycine and alanine—were consistently enriched in both transcriptome and translome populations.

In Figure 2a we compare different gene sets. In Figure 2b we focus mainly on the variation in enrichments when all the comparisons are restricted to the set of 181 genes ($G_{\text{Prot}} \cap G_{\text{mRNA}} = G_{\text{Prot}}$) common to all data sets. Thus, the differences between the populations now only reflect the effects of differential transcription of certain genes and differential translation of certain transcripts. We find here an enrichment specifically of cysteine in the translome in relation to the transcriptome.

To measure the statistical significance of the results on amino acid enrichment, we have performed a control analysis on a randomized data set (Figure 2d). We randomly permuted the expression values of the ORFs 1000 times and then recomputed the enrichments. This allowed us to compute distributions for the amino acid enrichments and, from integrating these, one-sided p -values indicating the significance of the observed enrichments.

Amino acid enrichment in Transposon data set. We also tried to extend our methodology, ineffectively, to cope with the semi-quantitative Transposon set. We used only those 450 ORFs that consistently yielded either no expression or high expression, as binary data, on or off. We show the enrichments of amino acids computed from this filtered Transposon abundance set in Figure 2a. Overall, the enrichments from this set seemed to be attenuated in comparison to other data.

Biomass enrichment. A corollary to amino acid enrichments is the determination of the average biomass of the transcriptome and translome populations (shown in Figure 2c). We found that the average molecular weight of a protein in both populations was, on average, lower than in the genome population. These preliminary observations suggest a cell preference to use less energetically expensive proteins for those that are highly transcribed or trans-

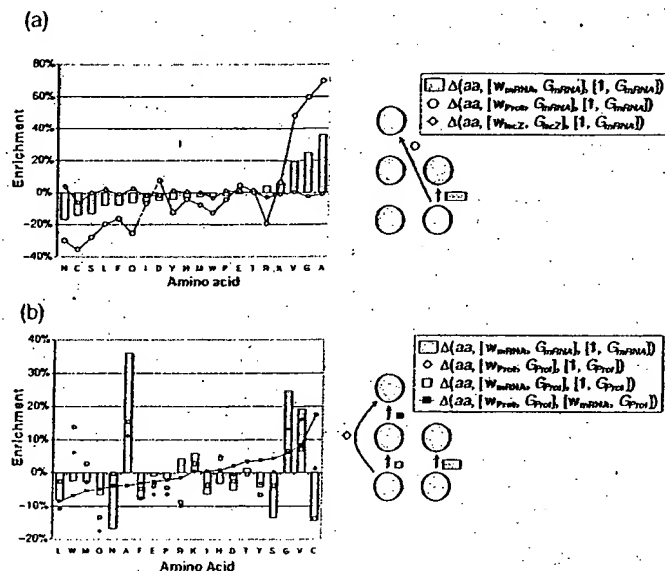


Fig. 2. Amino acid and biomass enrichment. (a) Shows the amino acid enrichments between different populations as indicated by the legend to the right of the plot (the legend is ordered in the same way as the schematic illustration in Figure 1). The bars indicate the enrichment of the transcriptome relative to the genome, whereas the circles indicate the enrichment of the translome relative to the genome. In addition, we also show the enrichment for protein abundance from the Transposon abundance set, represented by the circles with the line through them. (b) Shows a different view of amino acid enrichment from that contained in (a), now focusing on changes, and thus restricting the comparison to the genes common to all the data sets. The graph is ordered according to the enrichment from transcriptome to translome (black squares). We focus here only on the changes for the abundance gene set (G_{Prot}) to exclude the effects that arise from looking at different subsets. In this view the enrichments from genome to transcriptome (white squares) and from genome to translome (white diamonds) look more similar than do the analogous sets in (a). To make comparison with (a) easier we again show the enrichment from genome to the transcriptome for the complete gene set (G_{Gen} , shown in bars). (c) Shows biomass enrichment. The left panel depicts the average molecular weight per ORF (in units of kDa) and the right panel, the average molecular weight per amino acid (in units of Daltons) in each of the three stages of gene expression. The numbers inside the circles indicate the average molecular weights. The values next to the arrows indicate the enrichments in biomass between different populations. Both the circle diameters and the arrow widths are functions of the corresponding values (the hollow arrow indicates a positive value). It is very clear that the average molecular weight per ORF is much lower in the translome (by 20 or 15%) and transcriptome (by 29%) than in the genome. This relative depletion of biomass mainly takes place as a result of transcription; the effect of translation is less clear, depending on the populations compared. On the other hand, the depletion in the average molecular weight per amino acid (-3.3% from genome to translome) is an order of magnitude smaller than in the average weight per ORF. This shows that the yeast cell favors the expression of shorter ORFs over longer ones, and agrees with our earlier observation that there is a negative correlation between maximum ORF length and mRNA expression (Jansen and Gerstein, 2000); it seems that this effect mainly takes place during transcription rather than translation. (d) This plot shows that the amino acid enrichments are statistically significant. We have assessed significance by randomly permuting the expression levels among the genes and then recomputing the amino acid enrichments. This procedure can be repeated and used to generate distributions of random enrichments that can then be compared against the observed enrichments. In the plot the gray bars represent the observed enrichments already shown in Figure 3a. On top of the gray bars we show standard boxplots of enrichment distributions based on 1000 random permutations. (The middle line represents the distribution median. The upper and lower sides of the box coincide with the upper and lower quartiles. Outliers are shown as dots and defined as data points that are outside the range of the whiskers, the length of which is 1.5 the interquartile distance.) Based on the random distributions, we can compute one-sided p -values for the observed enrichments. Amino acids for which the p -values are less than 10^{-2} are shown in bold font.

lated. However, we also found that the average molecular weight per amino acid differed much less between the transcriptome and the translome on the one hand, and the

genome on the other hand (though it was still slightly less). This finding indicates that lower molecular weights in the translome and transcriptome relative to the genome are

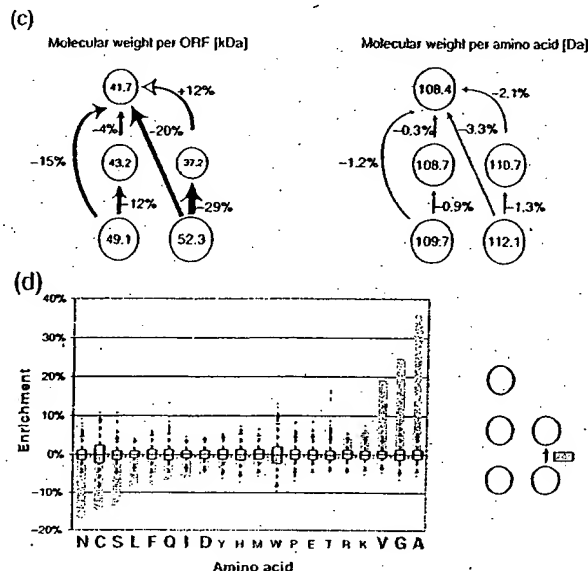


Fig. 2. Cont.

predominantly due to greater expression of shorter proteins rather than the incorporation of smaller amino acids.

Secondary structure composition. We also used our methodology to study the enrichment of secondary-structural features. Secondary structural annotation was derived from structure prediction applied uniformly to all the ORFs in the yeast genome as described in Table 1. As shown in Figure 3a, all three populations—genome, transcriptome, and translome—had a fairly similar composition of secondary structures—sheets, helices, and coils. The differences between populations were marginal and based only on the small subset of genes.

We also found that Transmembrane (TM) proteins were significantly depleted in the transcriptome (see website and caption). These results are consistent with our previous analyses (Jansen and Gerstein, 2000). The protein abundance data does not have any membrane proteins.

Subcellular localization. Figure 3c shows the enrichment of proteins associated with the various subcellular compartments. For clarity, we divided the cell into five distinct subcellular compartments, (see Table 1). We found that, in comparison to the genome, both the transcriptome and translome are enriched in cytoplasmic proteins. This is true whether we make our comparisons in

relation to the relatively large reference mRNA expression set or the smaller reference protein abundance set. As Figure 3c shows, the 2D gel experiments are clearly biased towards proteins from the cytoplasm. However, in the biased subset G_{Prot} transcription and translation lead to an even higher fraction of cytoplasmic proteins in the translome.

Functional categories. Finally, we compared the enrichment of various functional categories in both the translome and the transcriptome (see Figure 3b). This gives us a broad yet informative view of the cell as a whole. As described in Table 1, we used the top-level of the MIPS scheme for the functional category definitions. We found broad differences between the various populations, with some of the functional categories showing strikingly high enrichments.

DISCUSSION AND CONCLUSION

We developed: (i) a methodology for integrating many different types of gene expression and protein abundance into a common framework and applied this to a preliminary analysis; (ii) a procedure for scaling and merging different mRNA and protein sets together; and (iii) an approach for computing the enrichment of various proteomic features in the population of transcripts and proteins. We showed that by analyzing broad categories instead of individual noisy

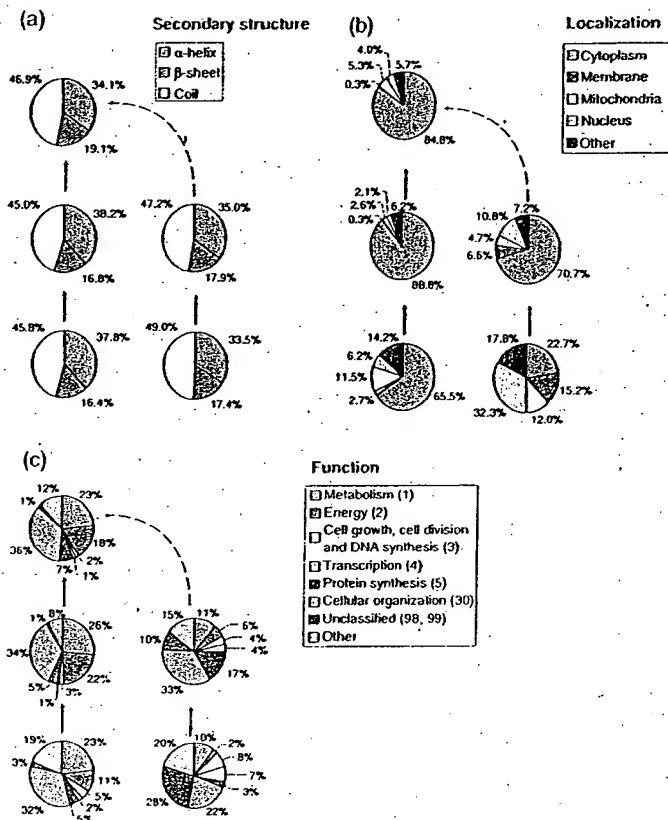


Fig. 3. Breakdown of the transcriptome and translome in terms of broad categories relating to structure, localization, and function. All of the subfigures are analogous to the schematic illustration in Figure 1. (a) Represents the composition of secondary structure in the different populations. (b) Represents the distribution of subcellular localizations associated with proteins in the various populations. We used standardized localizations developed earlier (Drawid and Geisstein, 2000), which, in turn, were derived from the MIPS, YPD, and SwissProt databases (Bairoch and Apweiler, 2000; Costanzo et al., 2000; Mewes et al., 2000). The subcellular localization has been experimentally determined for less than half of the yeast proteins, so our analysis applies only to this subset. (c) Shows the division of ORFs into different functional categories (according to the MIPS classification) in the various populations. Only the largest functional categories of the top level of the MIPS classification are shown. The group 'other' contains the smaller top-level categories lumped together. This 'other' group is different from the group 'unclassified,' which contains genes without any functional description.

data points, we could find logical trends in the underlying data. For example, individual transcription factors might have higher or lower protein abundance than one expects from their mRNA expression, but the category 'transcription factors' as a whole has a similar representation in the transcriptome and translome.

We found, as previously described (Futcher et al., 1999; Gygi et al., 1999b; Greenbaum et al., 2001), a weak correlation between individual measurements of mRNA

and protein abundance. The outliers of this correlation tend to be associated with cellular organization. One might conceive of using these outliers (i.e. those with significantly different transcriptional and translational behavior) to find consensus regulatory sequences. One possible method would involve using predicted mRNA structures (Jager et al., 1990; Zuker, 2000) to find and investigate consensus structural elements in these outliers to which the yeast translational machinery is known to be

Table 1. Data sets

| Data set | Description | Size [ORFs] | Reference |
|--------------------------|--|----------------|-------------------------------------|
| mRNA expression | | | |
| Young | Gene chip profiles yeast cells with mutations that affect transcription | 5455 | Holstege <i>et al.</i> (1998) |
| Church | Gene chip profiles of yeast cells under four different conditions | 6263 | Roth <i>et al.</i> (1998) |
| Samson | Comparing gene chip profiles for yeast cells subjected to alkylating agent | 6090 | Jelinsky and Samson (1999) |
| SAGE | Yeast cells during vegetative growth | 3778 | Velculescu <i>et al.</i> (1997) |
| Reference expression | Scaling and integrating the mRNA expression set into one data source | 6249 | — |
| Protein abundance | | | |
| 2-DE #1 | Measurement of yeast protein abundance by 2D gel electrophoresis and mass spectrometry | 156 | Gygi <i>et al.</i> (1999a,b) |
| 2-DE #2 | Similar to 2-DE set #1 | 71 | Futcher <i>et al.</i> (1999) |
| Transposon | Large-scale fusions of yeast genes with <i>lacZ</i> by transposon insertion | 1410 | Ross-Macdonald <i>et al.</i> (1999) |
| Reference abundance | Scaling and integrating the 2-DE data sets into one data source | 181 | — |
| Annotation | | | |
| Annotated localization | Subcellular localizations of yeast proteins | 2133 (6280) | Drawid and Gerstein (2000) |
| TM segments | Predicted TM and soluble proteins in yeast | 2710 (6280) | Gerstein (1998a,b,c) |
| MIPS functions | Functional categories for yeast ORFs | 3519 (6194) | Mewes <i>et al.</i> (2000) |
| GOR secondary structure | Predicted secondary structure yeast ORFs | 6280 | Gerstein (1998a,b,c) |

This table provides an overview of the data sets used in our analysis. The table is divided into three sections. The top section lists different mRNA expression sets. The middle section shows the protein abundance data sets used. The bottom section contains different annotations of protein features. The column 'Data set' lists a shorthand reference to each data set used throughout this paper. The next columns contain a brief description of the data sets, the number of ORFs contained in each of them, and the literature reference. In contrast to the other data we investigated, the reference expression and abundance data sets have been calculated for the purpose of our analysis (see text). An expanded version of the table is available on our web site.

Some further information on the genome annotations:

Localization. Protein localization information from YPD, MIPS and SwissProt were merged, filtered and standardized (Bairoch and Apweiler, 2000; Costanzo *et al.*, 2000; Mewes *et al.*, 2000) into five simplified compartments—cytoplasm, nucleus, membrane, extracellular (including proteins in ER and golgi), and mitochondrial—according to the protocol in Drawid *et al.* (2000). This yielded a standardized annotation of protein subcellular localization for 2133 out of 6280 ORFs.

TM segments. In 2710 out of 6280 yeast ORFs TM segments are predicted to occur, ranging from low to high confidence (732 ORFs). The TM prediction was performed as follows: the values from the scale for amino acids in a window of size 20 (the typical size of a TM helix) were averaged and then compared against a cutoff of -1 kcal mol^{-1} . A value under this cutoff was taken to indicate the existence of a TM helix. Initial hydrophobic stretches corresponding to signal sequences for membrane insertion were excluded. (These have the pattern of a charged residue within the first seven, followed by a stretch of 14 with an average hydrophobicity under the cutoff.) These parameters have been used, tested, and refined on surveys of membrane protein in genomes. 'Sure' membrane proteins had at least two TM-segments with an average hydrophobicity less than -2 kcal mol^{-1} (Rost *et al.*, 1995; Gerstein *et al.*, 2000; Santoni *et al.*, 2000; Senes *et al.*, 2000).

Functions. MIPS functional categories have been assigned to 3519 out of 6194 ORFs. (The remainder are assigned to category '98' or '99,' which corresponds to unclassified function.)

sensitive (McCarthy, 1998).

In relation to functional categories, we found three trends that were particularly notable: (i) the 'cellular

organization,' 'protein synthesis,' and 'energy production' categories were increasingly enriched as we moved from genome to transcriptome to translatome. In the transcrip-

tome and translome population relative to the genome; (ii) proteins with 'unclassified function' are significantly depleted, perhaps reflecting a bias against studying them; (iii) proteins in the 'transcription' and 'cell growth, cell division, and DNA synthesis' categories were consistently depleted. This reflects the fact that many of these proteins, such as transcription factors, act as 'switches' such that only small quantities of the protein are necessary to activate or deactivate a process. These results concur with previous calculations (Jansen and Gerstein, 2000) wherein we found the transcriptome is enriched specifically with proteins involved in protein synthesis and energy.

Limitations given the small size of the protein abundance data

Even with the extended coverage made possible by merging many data sets together into reference sets, the analysis is still limited by the minimal data. This was most applicable to the protein abundance measurements, potentially biasing our statistical results towards certain protein families. Moreover, the 181 proteins in G_{Prot} do not represent a random sample. They are skewed towards highly expressed, well-studied proteins. Our methodology attempts to control for this gene-selection bias through our enrichment formalism, which allows one to rather precisely gauge various aspects of the bias. Conversely, many protein features in both the translome and the transcriptome are dominated by highly expressed proteins. Under these circumstances, it is often sufficient to look at this smaller number of dominating proteins to characterize the whole population. This is similar to the development of the codon adaptation index for yeast (Sharp and Li, 1987). While based on only 24 highly expressed proteins, it has proven to be robust in predicting expression levels for the entire genome.

We believe that the essential formalism and approach that we develop will remain quite relevant for future data sets (Smith, 2000).

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REFERENCES

- An, H., Scopes, R.K. et al. (1991) Gel electrophoretic analysis of *Zymomonas mobilis* glycolytic and fermentative enzymes: identification of alcohol dehydrogenase II as a stress protein. *J. Bacteriol.*, **173**, 5975–5982.
- Anderson, L. and Seilhamer, J. (1997) A comparison of selected mRNA and protein abundances in human liver. *Electrophoresis*, **18**, 533–537.
- Bairoch, A. (2000) Serendipity in bioinformatics, the tribulations of a Swiss bioinformatician through exciting times! *Bioinformatics*, **16**, 48–64.
- Bairoch, A. and Apweiler, R. (2000) The SWISS-PROT protein sequence database and its supplement TrEMBL in 2000. *Nucleic Acids Res.*, **28**, 45–48.
- Bassett, D.E. Jr., Basrai, M.A. et al. (1996) Exploiting the complete yeast genome sequence. *Curr. Opin. Genet. Dev.*, **6**, 763–766.
- Batke, J., Benito, V.A. et al. (1992) A possible *in vivo* mechanism of intermediate transfer by glycolytic enzyme complexes: steady state fluorescence anisotropy analysis of an enzyme complex formation. *Arch. Biochem. Biophys.*, **296**, 654–659.
- Cambillau, C. and Claverie, J.M. (2000) Structural and genomic correlates of hyperthermostability. *J. Biol. Chem.*, **275**, 32383–32386.
- Cavalcoli, J.D., VanBogelen, R.A. et al. (1997) Unique identification of proteins from small genome organisms: theoretical feasibility of high throughput proteome analysis. *Electrophoresis*, **18**, 2703–2708.
- Claverie, J.M. (1999) Computational methods for the identification of differential and coordinated gene expression [in process citation]. *Hum. Mol. Genet.*, **8**, 1821–1832.
- Corthals, G., Wasinger, V.C., Hochstrasser, D.F. and Sanchez, J.C. (2000) The dynamic range of protein expression: a challenge for proteomic research. *Electrophoresis*, **21**, 1104–1115.
- Costanzo, M.C., Hogan, J.D. et al. (2000) The Yeast Proteome Database (YPD) and *Caenorhabditis elegans* Proteome Database (WormPD): comprehensive resources for the organization and comparison of model organism protein information. *Nucleic Acids Res.*, **28**, 73–76.
- Das, R. and Gerstein, M. (2000) The stability of thermophilic proteins: a study based on comprehensive genome comparison. *Funct. Int. Genom.*, **1**, 33–45.
- Doolittle, W.F. (2000) The nature of the universal ancestor and the evolution of the proteome. *Curr. Opin. Struct. Biol.*, **10**, 355–358.
- Drawid, A. and Gerstein, M. (2000) A Bayesian system integrating expression data with sequence patterns for localizing proteins: comprehensive application to the yeast genome. *J. Mol. Biol.*, **301**, 1059–1075.
- Drawid, A., Jansen, R. et al. (2000) Gene expression levels are correlated with protein subcellular localization. *Trends Genet.*, **10**, 426–430.
- Einarson, M. and Golemis, E. (2000) Encroaching genomics: adapting large-scale science to small academic laboratories. *Physiol. Genom.*, **2**, 85–92.
- Eisen, M.B. and Brown, P.O. (1999) DNA arrays for analysis of gene expression. *Meth. Enzymol.*, **303**, 179–205.
- Epstein, C. and Butow, R. (2000) Microarray technology—enhanced versatility, persistent challenge. *Curr. Opin. Biotechnol.*, **11**, 36–41.
- Ferea, T. and Brown, P. (1999) Observing the living genome. *Curr. Opin. Genet. Dev.*, **9**, 715–722.
- Fey, S.J., Nawrocki, A. et al. (1997) Proteome analysis of *Saccharomyces cerevisiae*: a methodological outline. *Electrophoresis*, **18**, 1361–72.
- Fey, S.J. and Larsen, P.M. (2001) 2D or not 2D. Two-dimensional gel electrophoresis. *Curr. Opin. Chem. Biol.*, **5**, 26–33.
- Frishman, D. and Mewes, H.W. (1997) Protein structural classes in five complete genomes [letter]. *Nat. Struct. Biol.*, **4**, 626–628.
- Frishman, D. and Mewes, H.W. (1999) Genome-based structural biology. *Prog. Biophys. Mol. Biol.*, **72**, 1–17.

- Futcher, B., Latter, G. *et al.* (1999) A sampling of the yeast proteome. *Mol. Cell Biol.*, **19**, 7357-7368.
- Gaasterland, T. (1999) Archaeal genomics. *Curr. Opin. Microbiol.*, **2**, 542-547.
- Garrels, J.I., McLaughlin, C.S. *et al.* (1997) Proteome studies of *Saccharomyces cerevisiae*: identification and characterization of abundant proteins. *Electrophoresis*, **18**, 1347-1360.
- Gerstein, M. (1997) A structural census of genomes: comparing bacterial, eukaryotic, and archaeal genomes in terms of protein structure. *J. Mol. Biol.*, **274**, 562-576.
- Gerstein, M. (1998a) How representative are the known structures of the proteins in a complete genome? A comprehensive structural census. *Fold. Des.*, **3**, 497-512.
- Gerstein, M. (1998b) Patterns of protein-fold usage in eight microbial genomes: a comprehensive structural census. *Proteins*, **33**, 518-534.
- Gerstein, M. (1998c) Patterns of protein-fold usage in eight microbial genomes: a comprehensive structural census. *Proteins*, **33**, 518-534.
- Gerstein, M. and Hegyi, H. (1998) Comparing genomes in terms of protein structure: surveys of a finite parts list. *FEMS Microbiol. Rev.*, **22**, 277-304.
- Gerstein, M. and Jansen, R. (2000) The current excitement in bioinformatics, analysis of whole-genome expression data: how does it relate to protein structure and function. *Curr. Opin. Struct. Biol.*, **10**, 574-584.
- Gerstein, M., Lin, J. *et al.* (2000) Protein folds in the worm genome. *Pac. Symp. Biocomput.*, 30-41.
- Greenbaum, D., Luscombe, N. *et al.* (2001) Interrelating different types of genomic data, from proteome to secretome: coming in on function. *Genome Res.*, **11**, 1463-1468.
- Gygi, S.P., Rist, B. *et al.* (1999a) Quantitative analysis of complex protein mixtures using isotope-coded affinity tags. *Nature Biotechnol.*, **17**, 994-999.
- Gygi, S.P., Rochon, Y. *et al.* (1999b) Correlation between protein and mRNA abundance in yeast. *Mol. Cell Biol.*, **19**, 1720-1730.
- Gygi, S.P., Conhals, G.L. *et al.* (2000a) Evaluation of two-dimensional gel electrophoresis-based proteome analysis technology. *Proc. Natl Acad. Sci. USA*, **97**, 9390-9395.
- Gygi, S.P., Rist, B. *et al.* (2000b) Measuring gene expression by quantitative proteome analysis. *Curr. Opin. Biotechnol.*, **11**, 396-401.
- Harry, J.L., Wilkins, M.R. *et al.* (2000) Proteomics: capacity versus utility. *Electrophoresis*, **21**, 1071-1081.
- Hatzinikitis, V., Choe, L.H. *et al.* (1999) Proteomics: theoretical and experimental considerations. *Biotechnol. Prog.*, **15**, 312-318.
- Haynes, P.A. and Yates, J.R. (2000) Proteome profiling: pitfalls and progress. *Yeast*, **17**, 81-87.
- Hegyi, H. and Gerstein, M. (1999) The relationship between protein structure and function: a comprehensive survey with application to the yeast genome. *J. Mol. Biol.*, **288**, 147-164.
- Holstege, F.C., Jennings, E.G. *et al.* (1998) Dissecting the regulatory circuitry of a eukaryotic genome. *Cell*, **95**, 717-728.
- Ishii, M., Hashimoto, S. *et al.* (2000) Direct comparison of genechip and SAGE on the quantitative accuracy in transcript profiling analysis. *Genomics*, **68**, 136-143.
- Ito, T., Tashiro, K. *et al.* (2000) Toward a protein-protein interaction map of the budding yeast: a comprehensive system to examine two-hybrid interactions in all possible combinations between the yeast proteins. *Proc. Natl Acad. Sci. USA*, **97**, 1143-1147.
- Jaeger, J.A., Turner, D.H. *et al.* (1990) Predicting optimal and suboptimal secondary structure for RNA. *Meth. Enzymol.*, **183**, 281-306.
- Jansen, R. and Gerstein, M. (2000) Analysis of the yeast transcriptome with structural and functional categories: characterizing highly expressed proteins. *Nucleic Acids Res.*, **28**, 1481-1488.
- Jeliosky, S.A. and Samson, L.D. (1999) Global response of *Saccharomyces cerevisiae* to an alkylating agent. *Proc. Natl Acad. Sci. USA*, **96**, 1486-1491.
- Jones, D.T. (1998) Do transmembrane protein superfolds exist? *FEBS Lett.*, **423**, 281-285.
- Jones, D.T. (1999) GenTHREADER: an efficient and reliable protein fold recognition method for genomic sequences. *J. Mol. Biol.*, **287**, 797-815.
- Kidd, D. *et al.* (2001) Profiling serine hydrolase activities in complex proteomes. *Biochemistry*, **40**, 4005-4015.
- Klose, J. (1975) Protein mapping by combined isoelectric focusing and electrophoresis of mouse tissues. A novel approach to testing for induced point mutations in mammals. *Humangenetik*, **26**, 231-243.
- Krogh, A. *et al.* (2001) Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J. Mol. Biol.*, **305**, 567-580.
- Lin, J. and Gerstein, M. (2000) Whole-genome trees based on the occurrence of folds and orthologs: implications for comparing genomes on different levels. *Genome Res.*, **10**, 808-818.
- Lipshutz, R.F. S., Gingeras, T.R. and Lockhart, D.J. (1999) High density synthetic oligonucleotide arrays. *Nature Genet.*, **21**, 20-24.
- Lopez, M.F. (2000) Better approaches to finding the needle in a haystack: optimizing proteome analysis through automation. *Electrophoresis*, **21**, 1082-1093.
- MacBeath, G. and Schreiber, S.L. (2000) Printing proteins as microarrays for high-throughput function determination. *Science*, **289**, 1760-1763.
- Matton, D.P., Constabel, P. *et al.* (1990) Alcohol dehydrogenase gene expression in potato following elicitor and stress treatment. *Plant Mol. Biol.*, **14**, 775-783.
- McCarthy, J.E. (1998) Posttranscriptional control of gene expression in yeast. *Microbiol. Mol. Biol. Rev.*, **62**, 1492-1553.
- Mewes, H.W., Frishman, D. *et al.* (2000) MIPS: a database for genomes and protein sequences. *Nucleic Acids Res.*, **28**, 27-40.
- Millar, A.A., Olive, M.R. *et al.* (1994) The expression and anaerobic induction of alcohol dehydrogenase in cotton. *Biochem. Genet.*, **32**, 279-300.
- Molloy, M.P. (2000) Two-dimensional electrophoresis of membrane proteins using immobilized pI gradients. *Anal. Biochem.*, **280**, 1-10.
- Nauchitel, V.V. and Somorjai, R.L. (1994) Spatial and free energy distribution patterns of amino acid residues in water soluble proteins. *Biophys. Chem.*, **51**, 327-336.
- Nelson, R.W., Nedelkov, D. *et al.* (2000) Biosensor chip mass spectrometry: a chip-based proteomics approach. *Electrophoresis*, **21**, 1155-1163.
- O'Farrell, P.H. (1975) High resolution two-dimensional elec-

- trophoresis of proteins. *J. Biol. Chem.*, 250, 4007-4021.
- Pandey, A. and Mann, M. (2000) Proteomics to study genes and genomes. *Nature*, 405, 837-846.
- Qi, S.Y., Moir, A. et al. (1996) Proteome of *Salmonella typhimurium* SL1344: identification of novel abundant cell envelope proteins and assignment to a two-dimensional reference map. *J. Bacteriol.*, 178, 5032-5038.
- Ross-Macdonald, P., Coelho, P.S. et al. (1999) Large-scale analysis of the yeast genome by transposon tagging and gene disruption. *Nature*, 402, 413-418.
- Rost, B., Casadio, R. et al. (1995) Transmembrane helices predicted at 95% accuracy. *Protein Sci.*, 4, 521-533.
- Roth, F.P., Hughes, J.D. et al. (1998) Finding DNA regulatory motifs within unaligned noncoding sequences clustered by whole-genome mRNA quantitation. *Nature Biotechnol.*, 16, 939-945.
- Rubin, G.M., Yandell, M.D. et al. (2000) Comparative genomics of the eukaryotes. *Science*, 287, 2204-2215.
- Sali, A. (1999) Functional links between proteins. *Nature*, 402, 25-26.
- Santoni, V., Molloy, M. et al. (2000) Membrane proteins and proteomics: an amour impossible? *Electrophoresis*, 21, 1054-1070.
- Schena, M., Shalon, D. et al. (1995) Quantitative monitoring of gene expression patterns with a complementary DNA microarray. *Science*, 270, 467-470.
- Searls, D.B. (2000) Using bioinformatics in gene and drug discovery. *Drug Discov. Today*, 5, 135-143.
- Senes, A., Gerstein, M. et al. (2000) Statistical analysis of amino acid patterns in transmembrane helices: the GxxxG motif occurs frequently and in association with beta-branched residues at neighboring positions. *J. Mol. Biol.*, 296, 921-936.
- Shapiro, L. and Harris, T. (2000) Finding function through structural genomics. *Curr. Opin. Biotechnol.*, 11, 31-35.
- Sharp, P.M. and Li, W.H. (1987) The codon adaptation index—a measure of directional synonymous codon usage bias, and its potential applications. *Nucleic Acids Res.*, 15, 1281-1295.
- Sherlock, G. (2000) Analysis of large-scale gene expression data. *Curr. Opin. Immunol.*, 12, 201-205.
- Shevchenko, A., Jensen, O.N. et al. (1996) Linking genome and proteome by mass spectrometry: large-scale identification of yeast proteins from two dimensional gels. *Proc. Natl Acad. Sci. USA*, 93, 14440-14445.
- Smith, R.D. (2000) Probing proteomes-seeing the whole picture? *Nature Biotechnol.*, 18, 1041-1042.
- Tatusov, R.L., Koonin, E.V. et al. (1997) A genomic perspective on protein families. *Science*, 278, 631-637.
- Tekaia, F., Lazcano, A. et al. (1999) The genomic tree as revealed from whole proteome comparisons. *Genome Res.*, 9, 550-557.
- Velculescu, V.E., Zhang, L. et al. (1997) Characterization of the yeast transcriptome. *Cell*, 88, 243-251.
- Wallin, E. and von Heijne, G. (1998) Genome-wide analysis of integral membrane proteins from eubacterial, archaean, and eukaryotic organisms. *Protein Sci.*, 7, 1029-1038.
- Washburn, M.P., Wolters, D. et al. (2001) Large-scale analysis of the yeast proteome by multidimensional protein identification technology. *Nature Biotechnol.*, 19, 242-247.
- Washburn, M.P. and Yates, J.R. 3rd (2000) Analysis of the microbial proteome. *Curr. Opin. Microbiol.*, 3, 292-297.
- Wittes, J. and Friedman, H.P. (1999) Searching for evidence of altered gene expression: a comment on statistical analysis of microarray data [editorial; comment]. *J. Natl Cancer. Inst.*, 91, 400-401.
- Wolf, Y.I., Brenner, S.E. et al. (1999) Distribution of protein folds in the three superkingdoms of life. *Genome Res.*, 9, 17-26.
- Young, K.H. (1998) Yeast two-hybrid: so many interactions, (in) so little time *Biol. Reprod.*, 58, 302-311.
- Zhang, M.Q. (1999) Large-scale gene expression data analysis: a new challenge to computational biologists (published erratum appears in *Genome Res.*, 1999, 9, 1156). *Genome Res.*, 9, 681-688.
- Zhu, H., Klemic, J.F. et al. (2000) Analysis of yeast protein kinases using protein chips. *Nature Genet.*, 26, 283-289.
- Zuker, M. (2000) Calculating nucleic acid secondary structure. *Curr. Opin. Struct. Biol.*, 10, 303-310.

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Thymidine kinase, thymidylate synthase, and dihydropyrimidine dehydrogenase profiles of cell lines of the National Cancer Institute's Anticancer Drug Screen.

Grem JL, Danenberg KD, Behan K, Parr A, Young L, Danenberg PV, Nguyen D, Drake J, Monks A, Allegra CJ.

Developmental Therapeutics Department, Medicine Branch, Division of Clinical Sciences, National Cancer Institute at the National Naval Medical Center, Bethesda, Maryland 20889, USA.

PURPOSE: To determine the expression of three targets of 5-fluorouracil (5-FU) and 5-fluoro-2'-deoxyuridine (FdUrd) in human tumor cell lines and to compare these with the 50% growth inhibition concentrations (GI(50)) from the National Cancer Institute database. **EXPERIMENTAL DESIGN:** Thymidine kinase (TK) activity was assessed by conversion of [(3)H]thymidine to [(3)H]TMP. Thymidylate synthase (TS) protein expression was determined by Western analysis. TS and dihydropyrimidine dehydrogenase (DPD) mRNA expression were measured by quantitative reverse transcription-PCR. **RESULTS:** The median (range) for the targets were as follows: 5-FU GI(50), 20.8 microM (0.8-536); FdUrd GI(50), 0.75 microM (0.25-237); TK, 0.93 nmol/min/mg (0.16-5.7); in arbitrary units: TS protein, 0.41 (0.05-2.95); TS mRNA, 1.05 (0.12-6.41); and DPD mRNA, 1.09 (0.00-24.4). A moderately strong correlation was noted between 5-FU and FdUrd GI(50)s ($r = 0.60$), whereas a weak-moderate correlation was seen between TS mRNA and protein expression ($r = 0.45$). Neither TS expression nor TK activity correlated with 5-FU or FdUrd GI(50)s, whereas lines with lower DPD expression tended to be more sensitive to 5-FU. Cell lines with faster doubling times and wild-type p53 were significantly more sensitive to 5-FU and FDURD. **CONCLUSIONS:** The lack of correlation may in part be attributable to the influence of downstream factors such as p53, the observation that the more sensitive cell lines with faster doubling times also had higher TS levels, and the standard procedure of the screen that uses a relatively short (48-h) drug exposure.

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Galanin in pituitary adenomas.

Grenback E, Bjellerup P, Wallerman E, Lundblad L, Anggard A, Ericson K, Aman K, Landry M, Schmidt WE, Holdfelt T, Hulting AL.

Department of Molecular Medicine, Endocrine and Diabetes Unit, Karolinska Hospital, S-17176 Stockholm, Sweden. Eva.Grenback@ks.se

Tumor galanin content was measured in extracts from human pituitary adenomas using a specific RIA method for monitoring human galanin. Twenty-two out of twenty-four tumors contained galanin with notably high levels in corticotroph adenomas, varying levels in clinically inactive tumors, and low levels in GH secreting adenomas. Tumor galanin and ACTH contents were closely correlated in all tumors. In four young patients with microadenomas and highly active Mb Cushing tumor galanin was inversely related to tumor volume. The molecular form of tumor galanin, studied with reverse-phase HPLC, was homogeneous with the majority of tumor galanin coeluting with standard human galanin. In the tumors analysed with in situ hybridization there was a good correlation between galanin peptide levels and galanin mRNA expression. In some tumors galanin mRNA and POMC levels coexisted, in others they were essentially in different cell populations. Levels of plasma galanin-LI were not related to tumor galanin concentration, and galanin levels were in the same range in sinus petrosus close to the pituitary venous drainage as in peripheral blood. Corticotrophin releasing-hormone injections in two patients caused ACTH, but no detectable galanin release into sinus petrosus. Our results demonstrate that corticotroph, but not GH adenomas, express high levels of galanin, in addition to ACTH, and that in some tumors both polypeptides are synthesised in the same cell population. However, galanin levels in plasma were not influenced by the tumor galanin content.

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Protein abundancy and mRNA levels of the adipocyte-type fatty acid binding protein correlate in non-invasive and invasive bladder transitional cell carcinomas.

Gromova I, Gromov P, Wolf H, Celis JE.

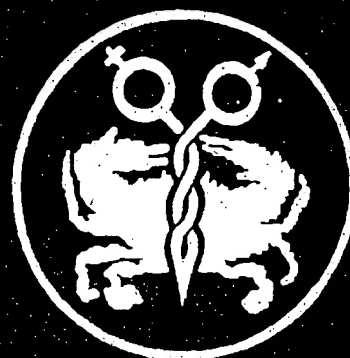
Department of Medical Biochemistry and Danish Centre for Human Genome Research,
The University of Aarhus, Aarhus C, Denmark.

The adipocyte type fatty acid-binding protein (A-FABP) is a small molecular weight fatty acid-binding protein whose expression correlates both with the grade of atypia and the stage of bladder transitional cell carcinomas (TCCs). To determine if the protein abundancy correlates with the mRNA levels in non-invasive and invasive lesions, we have analysed fresh TCCs (grade II, Ta; grade III, T2-4) by two-dimensional polyacrylamide gel electrophoresis (2D-PAGE) and measured the mRNA levels using the reverse transcription linked polymerase chain reaction (RT-PCR). Overall, the results showed a good correlation between protein abundancy and mRNA levels, indicating that the lack of expression of the protein observed in some lesions reflects low levels of transcription of the A-FABP gene rather than translational regulation. In addition, our studies showed that the loss of A-FABP protein observed in some tumors is not compensated by an increase in the skin fatty acid-binding-protein PA-FABP, as is the case in the A-FABP knockout mice.

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Protein abundance and mRNA levels of the adipocyte-type fatty acid binding protein correlate in non-invasive and invasive bladder transitional cell carcinomas

IRINA GROMOVA¹, PAVEL GROMOV¹, HANS WOLF² and JULIO E. CELIS¹

¹Department of Medical Biochemistry and Danish Centre for Human Genome Research, The University of Aarhus, DK-8000, Aarhus C; ²Department of Urology, Skejby Hospital, Aarhus, Denmark

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Abstract. The adipocyte type fatty acid-binding protein (A-FABP) is a small molecular weight fatty acid-binding protein whose expression correlates both with the grade of atypia and the stage of bladder transitional cell carcinomas (TCCs). To determine if the protein abundance correlates with the mRNA levels in non-invasive and invasive lesions, we have analysed fresh TCCs (grade II, Ta; grade III, T₂₋₄) by two-dimensional polyacrylamide gel electrophoresis (2D-PAGE) and measured the mRNA levels using the reverse transcription linked polymerase chain reaction (RT-PCR). Overall, the results showed a good correlation between protein abundance and mRNA levels, indicating that the lack of expression of the protein observed in some lesions reflects low levels of transcription of the A-FABP gene rather than translational regulation. In addition, our studies showed that the loss of A-FABP protein observed in some tumors is not compensated by an increase in the skin fatty acid-binding protein PA-FABP, as is the case in the A-FABP knockout mice.

Introduction

Bladder cancer accounts for 4.7% of all human cancers diagnosed. The spectrum of bladder tumors is broad with various histological types that include transitional cell carcinomas (TCCs), squamous cell carcinomas (SCCs), and adenocarcinomas (1-3). TCCs are by far the most prevalent tumors as they represent nearly 95% of all bladder cancers in

the Western Hemisphere. At first presentation, about 70% of the urinary bladder TCCs are diagnosed as differentiated superficial lesions that are confined either to the mucosa (Ta), or to the underlying connective tissue (T₁). The rest correspond to highly invasive, poorly differentiated tumors.

Non-invasive TCCs occur as two distinct growth patterns, papillary and non-papillary (flat) lesions (1,2), that display significant differences in their malignant potential and that are believed to originate from different genetic alterations (4-6). Papillary carcinomas usually correspond to low-grade lesions which frequently recur multiple times. These tumors begin as areas of hyperplasia that later undergo a process of dedifferentiation (grades I-IV). Invasive tumors may arise from these lesions, but often are derived from non-papillary carcinoma *in situ* that usually is of high grade at presentation and tend to invade and progress to muscle invasion and metastatic disease.

To date, many attempts have been made to pinpoint genetic changes that underly progression of bladder cancer as well as to identify molecular markers that correlate with tumor progression. Cytogenetic studies and molecular genetic data have shown that chromosomes 3p, 4p, 4q, 5q, 8p, 9p, 9q, 11p, 13q, 14q, 17p and 18q are frequently altered in bladder urothelial tumors (4,5 and refs. therein), and as a whole they have supported the notion that bladder cancer is a multistep process. Recently, Spruck *et al* (6) showed that chromosome 9 alterations occur early during development, while p53 mutations appear later in the process and confer invasive properties. The situation however is reverse in the case of Cis, as a large fraction of these lesions contain p53 mutations (5,6,8,9). Besides pointing towards two divergent pathways of bladder tumor progression, these studies implied that the order in which the genetic changes occur is important in determining the outcome of the lesion.

Assessment of bladder cancer is based on a thorough pathological examination of biopsy material which establishes the histological type of the tumor, its degree of differentiation (grade), and depth of invasion of the bladder wall (staging) (10-12). In spite of strict criteria for the pathological assessment of these lesions, there exist a significant inter-pathologist variation, a fact that emphasises the need for objective markers that may aid their classification. With this in mind, we are exploring the possibility of using proteome (13)

Correspondence to: Dr Irina Gromova, Department of Medical Biochemistry and Danish Centre for Human Genome Research, The University of Aarhus, DK-8000, Aarhus C, Denmark

Abbreviations: A-FABP, adipocyte type fatty acid-binding protein; PA-FABP, psoriasis associated fatty acid-binding protein; RT-PCR, reverse transcription linked polymerase chain reaction; 2D PAGE, two-dimensional polyacrylamide gel electrophoresis

Key words: progression, proteome, protein profiling, A-FABP protein and mRNA levels

expression profiles of these lesions as fingerprints to define their grade of atypia and eventually their stage (3,14). So far, more than 400 tumors of various grades and stages have been analysed by two-dimensional polyacrylamide gel electrophoresis (2D PAGE), and preliminary experiments have shown that even though the overall protein expression profiles of tumors of the same grade and stage are very similar, there are important differences suggesting that morphologically 'identical' TCCs may be further subdivided (1). Of the biomarkers of TCC progression identified so far, the adipocyte-type fatty acid binding protein (A-FABP) is perhaps one of the most interesting as the levels of this polypeptide have been shown to correlate both with the grade of atypia as well as with the stage of the disease (3). Given the putative importance of A-FABP as a progression marker, and since Anderson and Seilheimer (15) recently showed that post-transcriptional regulation of gene expression is a frequent phenomena in higher organisms, we have compared the levels of A-FABP mRNA and protein in non-invasive and invasive bladder TCCs expressing and lacking this protein.

Materials and methods

Tumors. Fresh bladder tumors were obtained immediately after transurethral resection. The grade and clinical stage of the tumors were determined by the pathologist at the Aarhus Municipal hospital. Clean tumors devoid of blood clots were divided into small pieces for 2D PAGE and DNA, and RNA preparation. The latter were immediately frozen in liquid nitrogen and store at -80°C until use.

[³⁵S]-methionine labeling and 2D-PAGE. In a few cases, small tumor pieces were labeled with [³⁵S]-methionine as previously described (3). 2D-PAGE was performed according to published procedures (16; see also <http://biobase.dk/cgi-bin/celis>).

RT-PCR. Frozen tumor samples were ground to powder in liquid nitrogen and total RNA was isolated using the acid guanidium isothiocyanate/phenol chloroform extraction procedure (17). The samples were treated with RNase-free DNases I (Pharmacia) to eliminate contaminating genomic DNA using the protocols recommended by the supplier. Poly(A)⁺ RNA was prepared using Poly (A)⁺ Quick columns according to the manufacturer's instructions (Stratagene). The synthesis of cDNA for RT-PCR reactions were carried out using the Gibco BRL SuperScript Kit. Two µg of total RNAs was mixed with oligo -dT11 primer, PCR buffer, MgCl₂ (25 mM), 0.1 M DDT and 10 mM dNTP. The mixture was incubated at 42°C for 5 min followed by the addition of SuperScript II reverse transcriptase and further incubation at 42°C for 50 min. The reaction was terminated by raising the temperature to 70°C for 15 min, followed by additional incubation at 37°C for 20 min in the presence of RNase H to deplete the RNA. Primers for known and cloned genes were purchase from Pharmacia as follows: A-FABP, Upper (from 186-208 bp) 5'-GATCATCAGTGTGAATGGGGAT-3'/lower (from 374-397 bp) 5'-CATCCTCTCGTTTCTCTTTATG-3'; B-actin upper 5'-GAGGTGGCTCTGACTGTACCAC-3'/lower 5'-CTCATTACGCTCTCGGAACATCTCG-3'.

Table I. Expression of A-FABP in non-invasive and invasive bladder TCCs.

| TCC | Grade/ stage | Level of A-FABP protein ^a | Level of A-FABP mRNA ^b |
|-------|--------------------------------------|---|--------------------------------------|
| 154 | GrlI/Ta | + | ++ |
| 166-5 | GrlI/Ta | - | + |
| 532-1 | GrlI/Ta | ++++ | ++++ |
| 533-1 | GrlI/Ta | + | + |
| 607-1 | GrlI/Ta | - | - |
| 692-1 | GrlI/Ta | +++ | +++ |
| 709-1 | GrlI/Ta | - | - |
| 763-1 | GrlI/Ta | ++ | ++ |
| 581-1 | GrlI/Ta | + | + |
| 616-1 | GrlI/Ta | + | ++ |
| 428-5 | GrlII/T ₂ -T ₃ | - | - |
| 570-2 | GrlII/T ₂ -T ₃ | - | - |
| 612-3 | GrlII/T ₂ -T ₃ | - | - |
| 711-1 | GrlII/T ₂ -T ₃ | - | - |
| 712-1 | GrlII/T ₂ -T ₃ | - | - |
| 727-1 | GrlII/T ₂ -T ₃ | - | - |

^aThe levels of A-FABP protein were determined based on the visual analysis of Coomassie Brilliant Blue stained gels and represent the average estimate of at least two different runs. Tumors scored as positive differed significantly with respect to the levels of the protein, and therefore are indicated with either four (very high), three (high), two (medium) and one (low) cross (see also Fig. 1);

^bThe mRNA levels were determined based on the intensity of Ethidium Bromide stained cDNA bands separated on agarose gel using the Bio-Rad Gel Doc 1000 system and represent the average estimate of at least three independent experiments. Corresponding mRNA levels are represented by crosses as described above.

PCR reactions were performed in a Progene thermal cycler using the Advantage Klen Tag Polymerase (Clontech). The cycling parameters consisted of 30 sec of denaturation at 94°C, with annealing of 30 sec at 60°C for B-actin or at 64°C for A-FABP. The extension time was for 2 min at 68°C for 29-40 cycles with the final extension of 7 min at 68°C. The PCR products were separated on 1.5% agarose gel electrophoresis followed by ethidium bromide staining and photography under UV light.

Results

A-FABP protein levels in non-invasive and invasive TCCs. One hundred suspected TCCs removed at the Department of Urology, Skejby Hospital, were analysed by high resolution 2D PAGE and Coomassie Brilliant Blue staining. Of these, 10 grade II, Ta TCCs (Table I) were chosen to correlate A-FABP protein and mRNA levels as these lesions yielded acceptable protein profiles both in terms of their purity as

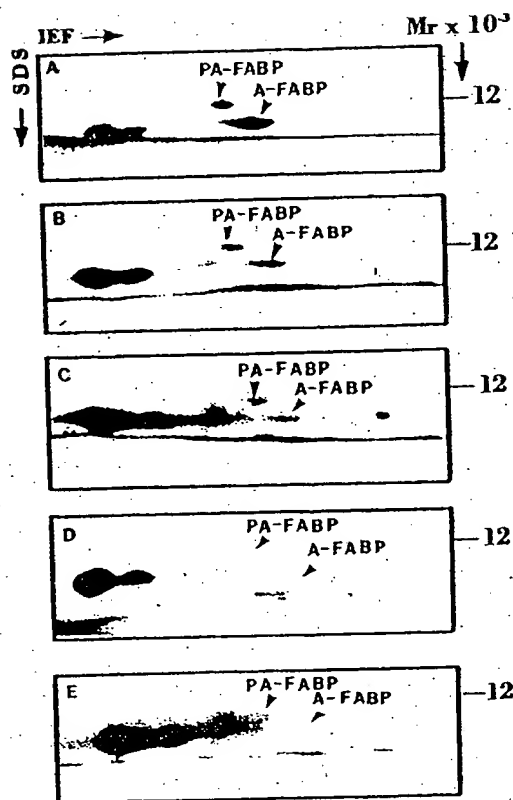


Figure 1. IEF 2D gels of whole cellular extracts from non-invasive and invasive TCCs. A, TCC 532-1; B, TCC 692-1; C, TCC 763-1; D, TCC 709-1 and E, TCC 711-1. Only the relevant area of the gels are shown.

assessed by monitoring for the absence of vimentin (contamination with connective tissue) and desmin (contamination with smooth muscle cells), as well as polypeptide resolution. In addition, reasonable amounts of these tumors were available for mRNA preparation.

Table I shows the levels of A-FABP protein expression in the 10 tumors analysed by 2D PAGE. The data were scored entirely based on the visual analysis of Coomassie Brilliant Blue stained gels and represent an average estimate of at least two different runs. Tumors scored as positive differed significantly with respect to the levels of this protein, and therefore are indicated with either four (very high), three (high), two (medium) and one cross (low). Representative examples of Coomassie stained 2D gels of tumors exhibiting very high (TCC 532-1, Fig. 1A), high (TCC 692-1; Fig. 1B), medium (TCC 763-1, Fig. 1C) and undetectable levels (TCC 709-1 and TCC 711-1 Fig. 1D-E) of A-FABP are shown in Fig. 1 (only the relevant area of the gels are shown).

A-FABP mRNA levels in non-invasive grade II, Ta TCCs. Since in many instances only a limited amount of fresh tumor was available, we used RT-PCR to determine the levels of A-FABP mRNA in the ten TCCs analysed by 2D PAGE (Fig. 1). Following amplification, the PCR products were analysed by conventional 1.5% agarose gel electrophoresis and visualised by ethidium bromide staining as shown in Fig. 2. The amount

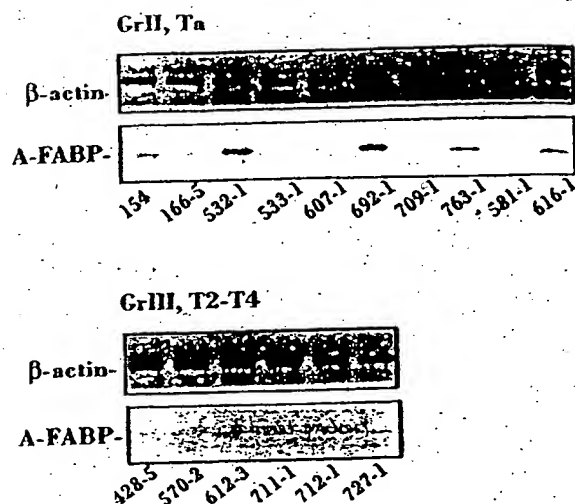


Figure 2. RT-PCR analysis of A-FABP mRNA expression in non-invasive (Gr II, Ta) and invasive TCCs (Gr III, T₂₋₄). For RT-PCR analysis, the ss c-DNA was synthesized by Reverse Transcriptase using total RNA, and used for RT-PCR amplification. The PCR products were resolved on 1.5% agarose gels and visualised under UV light following ethidium bromide staining. The A-FABP panels show the results of amplifications where the pair of gene specific primers was used to generate the 220 bp DNA fragment. Amplification of A-FABP was obtained after 30 cycles of PCR. The beta-actin panels represent the amplification of the beta-actin gene, which was used as an internal control to confirm that equal amounts of c-DNA were used in each reaction.

of cDNA in each lane was normalised using several house-keeping genes so as to achieve a more accurate assessment of the expression of the A-FABP mRNA. As shown in Fig. 2, TCC 532-1 exhibited the highest amount of A-FABP mRNA, followed by TCCs 692-1, 763-1, 616-1, 581-1, 154-1, 166-5 and 533-1. Undetectable levels of A-FABP mRNA were observed in the case of TCCs 607-1 and 709-1 (Fig. 2). Relative mRNA levels for the ten TCCs are given in Table I.

A-FABP protein and mRNA levels in invasive grade III, T₂₋₄ TCCs. Of the invasive TCCs (grade III, T₂₋₄) analysed by 2D PAGE only six yielded reasonable protein profiles for further study. As shown in Table I, none of these lesions expressed detectable levels of A-FABP as determined by Coomassie Brilliant Blue staining (Fig. 1E, TCC 711-1). In line with these results, the RT-PCR analysis of these tumors also revealed undetectable level of A-FABP mRNA (Fig. 2, Gr III T₂₋₄, Table I).

Loss of A-FABP protein is not compensated by an increase in PA-FABP. Recent studies of A-FABP knockout mice have shown that the loss of A-FABP in fat tissue is compensated by an increase in the skin fatty acid-binding protein mall (18). Our studies, however, indicated that the human homologue of mall, PA-FABP (19), did not compensate for the loss of A-FABP either in the non-invasive or the invasive tumors analysed in this study (Fig. 1D and E). In addition, Fig. 3 shows 2D gels of [³⁵S]-methionine labeled proteins from two grade II, Ta TCCs (192-4, T₁; Fig. 3A and 192-4, T₁; Fig. 3B).

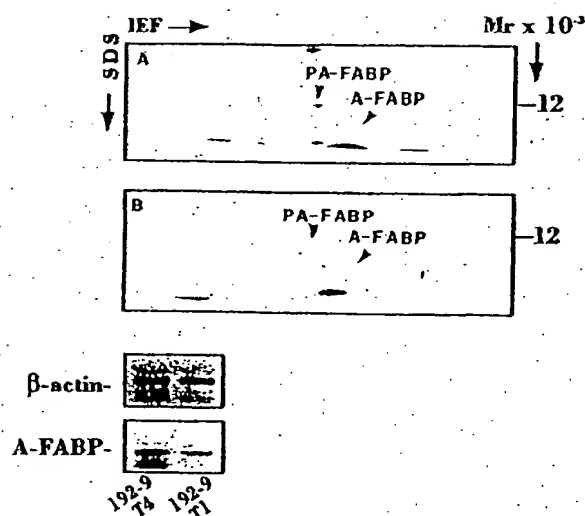


Figure 3. Levels of A-FABP and PA-FABP protein in grade II, T₂ tumors resected from the same patient. The two upper panels show the 2D gel autoradiograms of [³⁵S]-methionine labeled proteins from TCCs (grade II, T₂) resected from the same patient. A, TCC 192-9 tumor 4 and B, TCC 192-9 tumor 1. Only the relevant area of the autoradiograms are shown. The low panel shows the RT-PCR analysis of A-FABP mRNA expression in the same tumors (see also legend to Fig. 2).

which differ significantly in their levels of A-FABP protein and mRNA (Fig. 3, low panel). As shown in Fig. 3, the decrease in A-FABP observed in TCC 129-4, T₁ is not accompanied by an increase in the PA-FABP protein (Fig. 1E).

Discussion

Of the TCC progression markers identified to date, A-FABP is perhaps one of the most interesting as its presence correlates both with the grade of atypia ($p=0.0006$) and the stage of the disease ($p=0.0269$) (3). A-FABP is a low molecular weight protein belonging to a cytosolic multigene family of lipid-binding proteins that include heart, liver, intestinal, muscle, brain, skin and epithelial isoforms (20). Members of the FABP family are highly expressed in differentiated cells and show narrow tissue distribution. Their precise function is at present unknown, although there is evidence suggesting that they may play roles in intracellular lipid transport and metabolism, signal transduction (21,22) as well as growth control and differentiation (23). The role in signal transduction has been inferred from the fact that long-chain fatty acids and their metabolites can act as primary and second messengers in specific signalling pathways (24). Recently, it has been shown that A-FABP may play a central role in the pathway that links obesity with insulin resistance, most likely by connecting the fatty acid metabolism with the expression of TNF- α (18). Furthermore, there is evidence indicating that the A-FABP gene contains sequence information necessary for differentiation-dependent expression in adipocytes (25). Our own data derived from the study of TCCs and normal urothelium suggest that A-FABP may be required for normal urothelium differentiation (1), as may be the case for PA-FABP in the skin (19).

Considering the potential prognostic value of A-FABP protein and/or mRNA in TCC progression it was important to determine if the levels of both type of macromolecules correlated both in the non-invasive and the invasive lesions expressing and lacking A-FABP. The need for such correlation was underlined by recent studies of Anderson and Scilhamer (15), who reported a lack of overall correlation between the mRNA and protein levels of 45 rat proteins analysed by 2D PAGE in combination with cDNA arrays. Their data yielded a correlation coefficient of 0.45 which is half way between weak and perfect correlation. Clearly, our data showed a very good correlation between the protein and mRNA levels of A-FABP in all tumors analysed indicating that the loss of A-FABP protein observed in some tumors is not due to post-transcriptional regulation.

Recently, knockout mice carrying a null mutation in the ap2 gene encoding for A-FABP was produced (18). These animals do not show an obvious morphological or metabolic phenotype, but exhibit a 20-fold increase in the levels of the keratinocyte type FABP (*mal1*), which may compensate for the loss of the deleted gene (18). The human homologue of the *mal1* gene, PA-FABP, was cloned in our laboratory and has been shown to be highly upregulated in psoriatic skin as well as in abnormally differentiated primary keratinocytes (19). PA-FABP is expressed in normal urothelium together with A-FABP (3), and ongoing studies in the laboratory have shown that its level decreases significantly as tumors progress. Interestingly, the studies reported in this article did not revealed a compensatory up- or down-regulation of PA-FABP in the TCCs analysed, supporting the contention that PA-FABP may also play a role in cell growth and differentiation (19).

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References

1. Friedell GH, Nagy GK and Cohen SM: The pathology of human bladder cancer and related lesions. In: *The Pathology of Bladder Cancer*. I. Bryan GT and Cohen SM (eds). CRC Press Inc., Boca Raton FL, pp11-42, 1983.
2. Pauli BU, Alroy J and Weinstein RS: The ultrastructure and pathology of urinary bladder cancer. In: *The Pathology of Bladder Cancer*. II. Bryan GT and Cohen SM (eds). CRC Press Inc., Boca Raton FL, pp41-140, 1983.
3. Celis JE, Ostergaard M, Basse B, Celis A, Lauridsen JB, Ratz GP, Andersen I, Hein B, Wolf H, Ørntoft TF and Rasmussen HH: Loss of adipocyte-type fatty acid binding protein and other protein biomarkers is associated with progression of human bladder transitional cell carcinomas. *Cancer Res* 56: 4782-4790, 1996.
4. Heney NM, Ahmed S, Flanagan MJ, Frable W, Corder MP, Hafermann MD and Hawkins JR: Superficial bladder cancer: progression and recurrence. *J Urol* 130: 1083-1086, 1983.
5. Simoneau AR and Jones PA: Bladder cancer: the molecular progression to invasive disease (Review). *World J Urol* 12: 89-95, 1994.
6. Spruck CH, Ohniseit PF, Gonzalez-Zulueta M, Esrig D, Miyao N, Tsaim YC, Lerner SP, Schutte C, Yang AS and Cote R: Two molecular pathways to transitional cell carcinoma of the bladder. *Cancer Res* 54: 784-788, 1994.

7. Rosin MP, Cairns P, Epstein JI, Schoenberg MP and Sidransky D: Partial allelotype of carcinoma *in situ* of the human bladder. *Cancer Res* 55: 5213-5216, 1995.
8. Birchmeier W and Birchmeier C: Epithelial-mesenchymal transitions in development and tumour progression. In: *Epithelial-Mesenchymal Interactions in Cancer*. Goldberg ID and Rosen EM (eds). Birkhäuser, Basel, pp1-15, 1995.
9. Sidransky D, von Eschenbach A, Tsai YC, Jones P, Summerhayes I, Marshall F, Paul M, Green P, Hamilton SR, Frost P, *et al*: Identification of p53 gene mutations in bladder cancers and urine samples. *Science* 252: 706-709, 1991.
10. Lopez-Beltran A, Croghan GA, Croghan I, Huben RP, Mettlin C and Gatta JF: Prognostic factors in survival of bladder cancer. *Cancer* 70: 799-807, 1992.
11. Kern WH: The grade and pathologic stage of bladder cancer. *Cancer* 53: 1185-1189, 1984.
12. Jordan AM, Weingarten J and Murphy WM: Transitional cell neoplasms of the urinary bladder. Can biologic potential be predicted from histologic grading? *Cancer* 60: 2766-2774, 1987.
13. Wilkins MR, Sanchez JC, Gooley AA, Appel RD, Humphery-Smith I, Hochstrasser DF and Williams KL: Progress with proteome projects: why all proteins expressed by a genome should be identified and how to do it. *Biotechnol Genet Eng Rev* 13: 19-50, 1996.
14. Ostergaard M, Rasmussen HH, Nielsen HV, Vorum H, Ørntoft TF, Wolf H and Celis JE: Proteome profiling of bladder squamous cell carcinomas: identification of markers that define their degree of differentiation. *Cancer Res* 57: 4111-4117, 1997.
15. Anderson L and Seilhamer J: A comparison of selected mRNA and protein abundances in human liver. *Electrophoresis* 18: 533-537, 1997.
16. Celis JE, Ratz G, Basse B, Lauridsen JB and Celis A: High resolution two-dimensional gel electrophoresis of proteins: isoelectric focusing and non-equilibrium pH gradient electrophoresis (NEPHGE). In: *Cell Biology: A Laboratory Handbook*. III. Celis JE (ed). Academic Press, pp222-230, 1994.
17. Chomczynski P and Sacchi N: Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal Biochem* 162: 156-159, 1987.
18. Hotamisligil GS, Johnson RS, Distel RJ, Ellis R, Papaioannou VE and Spiegelman BM: Uncoupling of obesity from insulin resistance through a targeted mutation in aP2, the adipocyte fatty acid binding protein. *Science* 274: 1377-1379, 1996.
19. Madsen P, Rasmussen HH, Leffers H, Honore B and Celis JE: Molecular cloning and expression of a novel keratinocyte protein psoriasis-associated fatty acid-binding protein (PA-FABP) that is highly up-regulated in psoriatic skin and that shares similarity to fatty acid-binding proteins. *J Invest Dermatol* 99: 299-305, 1992.
20. Veerkamp JH, Paulussen RJ, Peeters RA, Maatman RG, van Moerkerk HT and van Kuppevelt TH: Detection, tissue distribution and (sub)cellular localization of fatty acid-binding protein types. *Mol Cell Biochem* 98: 11-18, 1990.
21. Glatz JF, Vork MM, Cistola DP and van der Vusse GJ: Cytoplasmic fatty acid binding protein: significance for intracellular transport of fatty acids and putative role on signal transduction pathways. *Prostaglandins Leukot Essent Fatty Acids* 48: 33-41, 1993.
22. Spitsberg VL, Matlashvili E and Gorewit RC: Association and coexpression of fatty-acid-binding protein and glycoprotein CD36 in the bovine mammary gland. *Eur J Biochem* 230: 872-878, 1995.
23. Yang Y, Spitzer E, Kenney N, Zschiesche W, Li M, Kromminga A, Muller T, Spener F, Lezius A, Veerkamp JH, *et al*: Members of the fatty acid binding protein family are differentiation factors for the mammary gland. *J Cell Biol* 127: 1097-1109, 1994.
24. Glatz JF, Borchers T, Spener F and van der Vusse GJ: Fatty acids in cell signalling: modulation by lipid binding proteins. *Prostaglandins Leukot Essent Fatty Acids* 52: 121-127, 1995.
25. Hunt CR, Ro JH, Dobson DE, Min HY and Spiegelman BM: Adipocyte P2 gene: developmental expression and homology of 5'-flanking sequences among fat cell-specific genes. *Proc Natl Acad Sci USA* 83: 3786-3790, 1986.

[The pathogenic role of macrophage migration inhibitory factor in acute respiratory distress syndrome]

[Article in Chinese]

Guo Y, Xie C.

Department of Respiratory Medicine, First Affiliated Hospital of Zhongshan University, Guangzhou 510080 China.

OBJECTIVE To investigate the expression and pathogenic role of macrophage migration inhibitory factor (MIF) in human acute respiratory distress syndrome (ARDS). **METHODS** The serum level of MIF in ARDS patients and normal persons were measured by ELISA method. Peripheral blood mononuclear cell (PBMC) MIF expression was determined by flow-cytometry. The expression of MIF mRNA and protein in the lung tissues were detected by using double immunohistochemistry labeling and in situ hybridization. **RESULTS** The serum level of MIF increased significantly in ARDS patients as compared with normal persons ($P < 0.01$). The percentage of PBMC MIF expression was higher in ARDS patients than in normal controls ($P < 0.01$). In situ hybridization and immunohistochemistry showed undetectable or weak MIF mRNA and protein expression in normal lungs. In contrast, there was marked upregulation of MIF mRNA and protein expression in the ARDS lungs. In ARDS macrophages infiltrated the alveolar space and interstitium, most of which also expressed MIF. Infiltrating macrophages were almost restricted to the areas of severe tissue damage. The MIF expression level showed a strong correlation with the number of infiltrating macrophages. **CONCLUSIONS** The serum level of MIF and PBMC MIF expression increased in ARDS patients with enhanced pulmonary MIF expression and macrophage infiltration, which suggests that MIF plays a pivotal role in the pathogenesis of ARDS.

PMID: 12126556 [PubMed - indexed for MEDLINE]

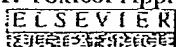
Restored expression and activity of organic ion transporters rOAT1, rOAT3 and rOCT2 after hyperuricemia in the rat kidney.

Habu Y, Yano I, Okuda M, Fukatsu A, Inui K.

Department of Pharmacy, Kyoto University Hospital, Faculty of Medicine, Kyoto University, Sakyo-ku, Kyoto 606-8507, Japan.

We previously reported that in hyperuricemic rats, renal impairment occurred and organic ion transport activity decreased, accompanied with a specific decrease in the expression of rat organic anion transporters, rOAT1 and rOAT3, and organic cation transporter, rOCT2. In the present study, we investigated the reversibility of the organic ion transport activity and expression of organic ion transporters (slc22a) during recovery from hyperuricemia. Hyperuricemia was induced by the administration of a chow containing uric acid and oxonic acid, an inhibitor of uric acid metabolism. Four days after discontinuance of the chow, the plasma uric acid concentration returned to the normal level, and renal functions such as creatinine clearance and BUN levels were restored, although the recovery of tubulointerstitial injury was varied in sites of the kidney. Basolateral uptake of p-aminohippurate (PAH) and tetraethylammonium (TEA), and both protein and mRNA levels of rOAT1, rOAT3 and rOCT2 in the kidney gradually improved during 14 days of recovery from hyperuricemia. Basolateral PAH transport showed a higher correlation with the protein level of rOAT1 ($r(2)=0.80$) than rOAT3 ($r(2)=0.34$), whereas basolateral TEA transport showed a strong correlation with rOCT2 protein ($r(2)=0.91$). The plasma testosterone concentration, which is a dominant factor in the regulation of rOCT2, was gradually restored during the recovery from hyperuricemia, but the correlation between the plasma testosterone level and rOCT2 protein expression in the kidney was not significant. These results suggest that the regulation of organic ion transporters, rOAT1, rOAT3 and rOCT2, by hyperuricemia is reversible, and the organic ion transport activity restores according to the expression levels of these transporters.

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Regulation of cytochrome P4501A1 in teleosts: sustained induction of CYP1A1 mRNA, protein, and catalytic activity by 2,3,7,8-tetrachlorodibenzofuran in the marine fish *Stenotomus chrysops*.

Hahn ME, Stegeman JJ.

Biology Department, Woods Hole Oceanographic Institution, Massachusetts 02543.

Cytochrome P4501A1 (CYP1A1) is known to play important roles in the activation and detoxification of carcinogens and other toxicants in vertebrate animals, including fish. Although extensively studied in mammalian systems, the regulation of CYP1A forms in other vertebrates is less well understood. We examined the time course and dose-response relationships for induction of CYP1A1 mRNA, protein, and catalytic activity by 2,3,7,8-tetrachlorodibenzofuran (TCDF) in the marine fish *Stenotomus chrysops* (scup). The time course of CYP1A1 induction was determined following a single ip dose (10 nmol/kg) of 2,3,7,8-TCDF. Hepatic ethoxyresorufin O-deethylase activity was increased after 1 day, reached a maximum by 8 days, and was still elevated 14 days after treatment. The content of immunodetectable CYP1A1 protein in liver was elevated on Day 1 and continued to increase through 14 days. CYP1A1 protein content was also strongly induced in heart and gill beginning at 2 days after treatment and extending through Day 14. Hepatic CYP1A1 mRNA was strongly induced by 1 day after dosing and remained elevated through 14 days. The sustained induction of CYP1A1 mRNA by 2,3,7,8-TCDF contrasts with the transient induction seen previously in fish treated with nonhalogenated inducers and most likely reflects differences in persistence of the inducers. Dose-response studies indicated that induction of CYP1A1 mRNA, protein, and catalytic activity occurred following doses of 2,3,7,8-TCDF as low as 0.4 nmol/kg (120 ng/kg), within the range of whole-body contents of this congener measured in fish from contaminated environments. The estimated dose producing half-maximal CYP1A1 induction in scup was approximately 2-10 nmol/kg, suggesting that the sensitivity of these fish to induction may be as great as or greater than that of rats. In contrast to previous results obtained with 3,3',4,4'-tetrachlorobiphenyl (TCB) and beta-naphthoflavone, which appear to inhibit or inactivate CYP1A1 in fish and other vertebrates, there was a good correlation among levels of CYP1A1 mRNA, protein, and catalytic activity in individual fish following various doses of 2,3,7,8-TCDF. The difference in response to 2,3,7,8-TCDF versus 3,3',4,4'-TCB may reflect differences in the inducing potencies of the two compounds relative to their similar potencies as inhibitors of CYP1A1 catalytic activity. In additional studies to evaluate structure-activity relationships for CYP1A1 induction by chlorinated dibenzofurans in fish, scup were treated with 2,3,6,8-tetrachlorodibenzofuran (2,3,6,8-TCDF). At 10 or 50 nmol/kg, 2,3,6,8-TCDF was inactive as an inducer of CYP1A1 mRNA, protein, or catalytic activity. (ABSTRACT TRUNCATED AT 400 WORDS)

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Expression of the pS2 gene in breast tissues assessed by pS2-mRNA analysis and pS2-protein radioimmunoassay.

Hahnel E, Robbins P, Harvey J, Sterrett G, Hahnel R.

Department of Pathology, University of Western Australia, Queen Elizabeth II Medical Centre, Nedlands.

The expression of the pS2 gene in breast tissues was assessed by measuring pS2-protein using a radioimmunoassay, and by determining pS2-mRNA using Northern blotting. There was a good correlation between the two measurements, indicating that expression of the pS2 gene in breast tissues may be assessed by either method. Since radioimmunoassay is technically easier and more efficient than Northern blotting, radioimmunoassay will be the method of choice in routine applications.

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Brief communication

Expression of the pS2 gene in breast tissues assessed by pS2-mRNA analysis and pS2-protein radioimmunoassay

Erika Hähnel, Peter Robbins, Jennet Harvey, Gregory Sterrett and Roland Hähnel
Department of Pathology, University of Western Australia, Queen Elizabeth II Medical Centre, Nedlands, 6009, Western Australia

Key words: breast tissue, pS2-mRNA, pS2 protein, radioimmunoassay

Summary

The expression of the pS2 gene in breast tissues was assessed by measuring pS2-protein using a radioimmunoassay, and by determining pS2-mRNA using Northern blotting. There was a good correlation between the two measurements, indicating that expression of the pS2 gene in breast tissues may be assessed by either method. Since radioimmunoassay is technically easier and more efficient than Northern blotting, radioimmunoassay will be the method of choice in routine applications.

Introduction

Expression of the pS2 gene is controlled by estrogen. This was first described in the MCF-7 breast cancer cell line [1]. pS2 expression has since been reported to be useful as a prognostic indicator [2, 3], although this was not confirmed in another series [4].

pS2 expression may be assessed in tissue homogenates by analysis of pS2-mRNA [5], by radioimmunoassay of the pS2-protein [2], or by immunocytochemical detection of the pS2 protein in tissue sections [5]. It was the aim of this study to establish the correlation between pS2-mRNA and pS2-protein by radioimmunoassay in a series of tissues obtained from mastectomy specimens performed for carcinoma of the breast. Primary breast carcinoma tissue, metastatic carcinoma within axillary nodes, and macroscopically benign breast tissue were examined.

Materials and methods

Breast tissues

Tissue specimens from mastectomies performed for carcinoma of the breast were examined. 32 primary breast carcinomas, 10 axillary lymph nodes containing metastatic breast carcinoma, and 20 samples of uninvolved breast tissue were analyzed for pS2 expression.

The primary breast carcinomas were histologically classified using a conventional subclassification. The presence or absence of primary tumour was assessed. The presence of metastatic carcinoma within lymph nodes studied was verified by histological examination of the node remnant after sampling.

'Uninvolved' breast tissue was sampled from sites well removed from the primary breast tumour (usually in another quadrant of the breast), and was selected only if the tissue appeared macroscopically unremarkable. Tissue sampling occurred imme-

diately upon arrival of the mastectomy specimen in the laboratory, with minimal delays between removal and sampling.

Tissues for pS2 analysis were snap frozen in liquid nitrogen and stored at -70°C until processed.

Extraction of RNA and determination of pS2-mRNA

Details of the procedure have been described in our previous paper [6]. Briefly, the deep-frozen tissue was homogenized in a micro-dismembrator. The homogeneous powder was extracted with guanidiniumisothiocyanatephenolchloroformisoamylalcohol, and RNA was precipitated with isopropanol. The washed RNA pellet was dissolved in SDS and glyoxylated, and the RNA preparation loaded onto agarose gel. After electrophoresis the gel was capillary blotted onto Zeta-probe membranes. Membranes were hybridized overnight with cDNA probes pS2 and 36B4, which were labeled with $[\alpha^{32}\text{P}]$ dCTP by nick translations. Washed membranes were exposed to Kodak X-omat AR film. Relative intensities of the mRNA bands were assessed visually as not detectable, very weak, weak, medium, strong, and very strong, taking the intensities of the ubiquitous 36B4 bands into account.

Radioimmunoassay of pS2-protein

Deep frozen specimens were pulverized with a microdismembrator. The tissue powder was suspended in 10 volumes of pH 7.5 phosphate buffer. The homogenate was centrifuged in a refrigerated centrifuge at 4°C for 60 minutes at 2600xg. The supernatant was removed with a Pasteur pipette, carefully avoiding the fat layer on the top. The protein concentration in the supernatant was estimated by use of the Coomassie dye-binding method [7]. An aliquot of the supernatant was diluted to a protein concentration between 1 and 2 mg/ml before assay of the pS2-protein. In one case the protein concentration of the supernatant was well below 1 mg/ml.

The estimation of the pS2-protein was performed using a solid phase, two-site radioimmunoassay. The kits were bought from CIS Biointernational, Gif-sur-Yvette, France (ELSA-PS2). In this method the molecules of pS2 are sandwiched between two monoclonal antibodies; the first one is coated on the ELSA solid phase, the second one is radiolabeled with 125-iodine. The radioactivity bound to the ELSA is proportional to the concentration of pS2-protein. Details of the procedure are supplied with the kit [8].

Results and discussion

32 primary breast carcinomas, metastatic breast carcinoma in 10 lymph nodes, and 20 samples of benign breast tissue from mastectomies were investigated. Two of the carcinomas were of the infiltrating lobular type, two were ductal carcinomas *in situ*, one was a multicentric invasive ductal carcinoma, all others were invasive ductal carcinomas.

Examples of pS2 Northern blots have been shown in our previous paper [6] which demonstrate that undegraded pS2-mRNA can be isolated by the method used.

The results of the pS2-protein and pS2-mRNA assays are shown on Fig. 1. There was a good correlation between the two types of results. When pS2-mRNA could not be detected by Northern blot, pS2-protein results were usually below 1 ng/mg protein (22 of 30), or between 1 and 3.7 ng/mg (6 of 30). Two were exceptions (7.7 and 14.6); one of them could have been due to the very low protein content in the cytosol which would lead to a large pS2 value and an associated error. There was no explanation for the other high result. Very weak pS2-mRNA signals on Northern blots corresponded to pS2-protein values between 1.1 and 19.2 with an average of 6.6 ng/mg protein (median 5.7). The mean and median pS2-protein concentration in the tissue with weak pS2-mRNA signals were 14.3 and 10.7 ng/mg protein, respectively. The average pS2-protein concentration increased to 32.7 (median 31.5) ng/mg protein for tissues assessed as medium pS2-mRNA intensity, and to 43.3 (median 53.8) ng/mg protein for tissues with strong or very strong

pS2-mRNA signals. These values should be used as an approximate guide only, since the number of samples in the various groups was fairly small. One-way analysis of variance confirmed that the means of the pS2-protein values in the groups made up according to their pS2-mRNA signal intensity, were significantly different ($p < 10^{-6}$).

If the pS2 gene is expressed, its expression is on average greater in breast carcinomas than in uninvolved breast tissue. If one takes pS2-protein values above 4 ng/mg protein as cut-off, the average pS2-protein in 14 breast cancers was 34.3 (median 35.2), while it was only 18.1 (median 13.8) in 12 uninvolved breast tissue samples. If the cut-off is taken at 10 ng/mg protein, average pS2-protein in breast carcinoma is also about twice the level of uninvolved tissue. There were not enough lymph node metastases which expressed the pS2 gene to allow a comparison with carcinoma or uninvolved breast tissue.

Recent preliminary results of pS2 by radioimmunoassay [9] are similar to ours for breast cancer but considerably lower than our results for normal breast tissue.

The values of the pS2-protein measured obviously depend on the protein used for calibration. We used the pS2-protein standards supplied with the CIS kit, which according to the supplier gave values from 0 to 740 ng/mg protein in a series of 205 breast cancer cytosols. Previously, a different standard had been used for presumably the same series of breast carcinomas [2], and a conversion factor to current standards is given as 2.8 [8].

It was noticed that the correlation between pS2-protein and pS2-mRNA was better in breast carcinoma specimens than in uninvolved breast tissue. This is unexplained, though it could be due to the variable content of cell or tissue types in adjoining parts of a specimen, a variation more likely to occur in our sampling of non-malignant breast tissue compared to sampling of carcinomas. A similar variability in breast carcinoma specimens will probably have a smaller influence on the pS2 results, since the malignant cells – if they do express the pS2 gene – contain more pS2-protein than normal breast.

CORRELATION OF pS2-mRNA AND pS2-PROTEIN

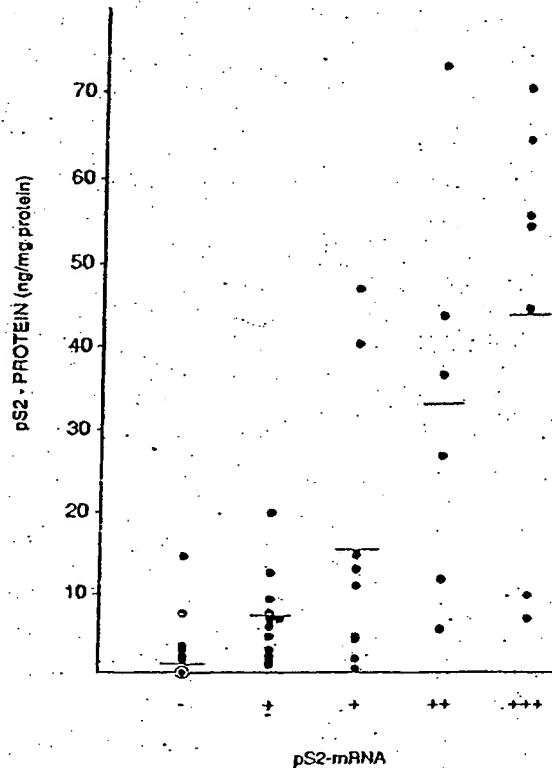


Fig. 1. Correlation between pS2-protein by radioimmunoassay and pS2-mRNA by Northern blot. \odot = 22 results below 1. The horizontal lines indicate the mean values.

Acknowledgements

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References

1. Masiakowski P, Breathnach R, Bloch J, Gannon F, Krust A, Cham-

1. bon P: Cloning of cDNA sequences of hormone-regulated genes from the MCF-7 human breast cancer cell lines. *Nucleic Acids Res* 10: 7895-7903, 1982
2. Foekens JA, Rio MC, Seguin P, Van Putten WLJ, Fauque J, Nap M, Klijn JGM, Chambon P: Predication of relapse and survival in breast cancer patients by pS2 protein status. *Cancer Res* 50: 3832-3837, 1990
3. Foekens JA, Van Putten WLJ, Portengen H, Rodenburg CJ, Reubi JC, Berns PMJJ, Henzen-Logmans SC, Van Der Burg MEL, Alexieva-Figusch J, Klijn JGM: Prognostic value of pS2 protein and receptors for epidermal growth factor, insulin-like growth factor, and somatostatin in patients with breast and ovarian cancer. *J Steroid Biochem Molec Biol* 37: 815-821, 1990
4. Henry JA, Piggott NH, Mallick UK, Nicholson S, Farndon JR, Westley BR, May FEB: pNR-2/pS2 immunohistochemical staining in breast cancer: Correlation with prognostic factors and endocrine response. *Br J Cancer* 63: 615-622, 1991
5. Rio MC, Bellocq JP, Gairard B, Rasmussen UB, Krust A, Koehl C, Calderoli H, Schiff V, Renaud R, Chambon P: Specific expression of the pS2 gene in subclasses of breast cancer in comparison with expression of the estrogen and progesterone receptors and the oncogene ERBB2. *Proc Natl Acad Sci USA* 84: 9243-9247, 1987
6. Hähnel E, Joyce R, Sterren GF, Harvey JM, Hähnel R: Detection of estradiol-induced messenger RNA (pS2) in uninvolved breast tissue from mastectomies for breast cancer. *Breast Cancer Res Treat* 20: 167-176, 1991
7. Bradford M: A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 72: 248-254, 1976
8. CIS ELSA-pS2: Immunoradiometric assay of pS2 protein. Package insert, December, 1990
9. Kouyoumdjian JC, Boissier B, Rymet JC, Bagnard G, Rotten D, Levaillant JP, Constancis B, Philippon C, Floury C, Thirion B: Determination of several prognostic parameters in human normal breast, benign mastopathies, and adenocarcinomas. *J Tumour Marker Oncol* 6: 111, 1991



The role of the epidermal growth factor receptor in sustaining neutrophil inflammation in severe asthma.

Hamilton LM, Torres-Lozano C, Puddicombe SM, Richter A, Kimber I, Dearman RJ, Vrugt B, Aalbers R, Holgate ST, Djukanovic R, Wilson SJ, Davies DE.

Division of Infection, Inflammation & Repair, School of Medicine, University of Southampton, UK.

BACKGROUND: The extent of epithelial injury in asthma is reflected by expression of the epidermal growth factor receptor (EGFR), which is increased in proportion to disease severity and is corticosteroid refractory. Although the EGFR is involved in epithelial growth and differentiation, it is unknown whether it also contributes to the inflammatory response in asthma. **OBJECTIVES:** Because severe asthma is characterized by neutrophilic inflammation, we investigated the relationship between EGFR activation and production of IL-8 and macrophage inhibitory protein-1 alpha (MIP-1alpha) using in vitro culture models and examined the association between epithelial expression of IL-8 and EGFR in bronchial biopsies from asthmatic subjects. **METHODS:** H292 or primary bronchial epithelial cells were exposed to EGF or H2O2 to achieve ligand-dependent and ligand-independent EGFR activation; IL-8 mRNA was measured by real-time PCR and IL-8 and MIP-1alpha protein measured by enzyme-linked immunosorbent assay (ELISA). Epithelial IL-8 and EGFR expression in bronchial biopsies from asthmatic subjects was examined by immunohistochemistry and quantified by image analysis. **RESULTS:** Using H292 cells, EGF and H2O2 increased IL-8 gene expression and release and this was completely suppressed by the EGFR-selective tyrosine kinase inhibitor, AG1478, but only partially by dexamethasone. MIP-1alpha release was not stimulated by EGF, whereas H2O2 caused a 1.8-fold increase and this was insensitive to AG1478. EGF also significantly stimulated IL-8 release from asthmatic or normal primary epithelial cell cultures established from bronchial brushings. In bronchial biopsies, epithelial IL-8, MIP-1alpha, EGFR and submucosal neutrophils were all significantly increased in severe compared to mild disease and there was a strong correlation between EGFR and IL-8 expression ($r = 0.70$, $P < 0.001$). **CONCLUSIONS:** These results suggest that in severe asthma, epithelial damage has the potential to contribute to neutrophilic inflammation through enhanced production of IL-8 via EGFR-dependent mechanisms.

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ELSEVIER

Human hepatic microsomal epoxide hydrolase: comparative analysis of polymorphic expression.

Hassett C, Lin J, Carty CL, Laurenzana EM, Omiecinski CJ.

Department of Environmental Health, University of Washington, Seattle 98105-6099, USA.

Interindividual variation in the expression of human microsomal epoxide hydrolase (mEH) may be an important risk factor for chemically induced toxicities, including cancer and teratogenesis. In this study, phenotypic variability and mEH genetic polymorphisms were examined in a bank of 40 transplant-quality human liver samples. Immunochemically determined protein content, enzymatic activities, polymorphic amino acids, as well as mEH RNA levels were evaluated in parallel. Enzymatic activity was assessed using (+/-)-benzo[a]pyrene-4,5-epoxide at 2 substrate concentrations. The relative hydrolyzing activities obtained using saturating substrate levels were highly correlated ($r = 0.85$) with results derived from limiting substrate concentrations and exhibit approximately an 8-fold range in activity levels across the panel of 40 liver samples. mEH enzyme activity also demonstrated strong correlation ($r > \text{or} = 0.74$) with an 8.4-fold variation determined for mEH protein content within the same samples. However, these protein/activity measurements were poorly correlated ($r < \text{or} = 0.23$) with mEH RNA levels, which exhibited a 49-fold variation. Two common polymorphic amino acid loci in the mEH protein did not exclusively account for variation in enzymatic activity, although this conclusion is confounded by heterozygosity in the samples. These data demonstrate the extent of hepatic mEH functional variability in well-preserved human tissues and suggest that polymorphism of mEH protein expression is regulated in part by posttranscriptional controls, which may include nonstructural regulatory regions of the mEH transcript.

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Review

Early Detection of Lung Cancer: Clinical Perspectives of Recent Advances in Biology and Radiology¹

Fred R. Hirsch,² Wilbur A. Franklin,
Adi F. Gazdar, and Paul A. Bunn, Jr.

Lung Cancer Program and Departments of Medicine and Pathology, University of Colorado Cancer Center, Denver, Colorado 80262 [F. R. H., W. A. F., P. A. B.]; Department of Pathology, University of Texas, Southwestern Medical Center, Dallas, Texas [A. F. G.]; and Department of Oncology, Finsen Center, National University Hospital, Copenhagen, Denmark [F. R. H.]

Abstract

Lung cancer is the most common cause of cancer death in developed countries. The prognosis is poor, with less than 15% of patients surviving 5 years after diagnosis. The poor prognosis is attributable to lack of efficient diagnostic methods for early detection and lack of successful treatment for metastatic disease. Most patients (>75%) present with stage III or IV disease and are rarely curable with current therapies. Within the last decade, rapid advances in molecular biology, pathology, bronchology, and radiology have provided a rational basis for improving outcome. These advancements have led to a better documentation of morphological changes in the bronchial epithelium before development of clinical evident invasive carcinomas. This has changed our concept of lung carcinogenesis and emphasized the multistep carcinogenesis approach on several levels. Combined with the technical developments in bronchoscopic techniques, e.g., laser-induced fluorescence endoscope (LIFE) bronchoscopy, we now have improved methods to localize preinvasive and early-invasive bronchial lesions. With the LIFE bronchoscope, a new morphological entity (angiogenic squamous dysplasia) has been recognized, which might be an important biomarker and target for antiangiogenic chemopreventive agents. To reduce the mortality of lung cancer, these new technologies have been taken into the clinic in different scientific settings. The use of low-dose spiral computed tomography in the screening of a high-risk population has demonstrated the possibility of diagnosing small peripheral tumors that are not seen on conventional X-ray. A shift in the therapeutic paradigm from targeting advanced clinically

manifest lung cancer toward asymptomatic preinvasive and early-invasive cancer is occurring. The present article reviews the recent advances in the diagnosis of preinvasive and early-invasive cancer to identify biomarkers for early detection of lung cancer and for chemoprevention studies.

Introduction

Lung cancer is the most common cause of cancer deaths in the countries of North America and other developed countries, accounting for 29% of all cancer deaths and more deaths than from prostate, breast, and colorectal cancer combined in the United States (1). Lung cancer will be diagnosed in ~170,000 new patients in the United States in the year 2000, and <15% of them will survive 5 years after diagnosis (1). The prognosis for the patients with lung cancer is strongly correlated to the stage of the disease at the time of diagnosis. Whereas patients with clinical stage IA disease have a 5-year survival of about 60%, the clinical stage II-IV disease 5-year survival rate ranges from 40% to less than 5% (2). Over two-thirds of the patients have regional lymph-node involvement or distant disease at the time of presentation (3). The poor prognosis is largely attributable to the lack of effective early detection methods and the inability to cure metastatic disease. The unsatisfactory cure rates supports efforts aimed at early identification and intervention in lung cancer.

Historically, the only diagnostic tests available for the detection of lung cancer in its early stages were chest radiography and sputum cytology. The efficacy of these tests as mass screening tools was evaluated in controlled trials sponsored by the NCI² and conducted at Johns Hopkins University, Memorial Sloan-Kettering Cancer Center, and the Mayo Clinic during the 1970s (4-6). The principal goal of these studies was to determine whether a reduction in lung cancer mortality could be achieved by adding sputum cytology testing to annual screening by chest radiography. Results from these trials showed that both tests could detect presymptomatic, early-stage carcinoma, particularly of squamous cell type. Resectability and survival rates were found to be generally higher in the study groups than in the control groups. However, improvements in resectability and survival did not lead to a reduction in overall lung cancer mortality, the most critical end point. A subsequent study of 6346 Czechoslovakian male smokers also found no reduction in lung cancer mortality after dual screening by chest radiography

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²To whom requests for reprints should be addressed, at University of Colorado Cancer Center, Department of Pathology, University of Colorado Health Sciences Center, 4200 East Ninth Avenue, B 216, Denver, Colorado 80262. E-mail: Fred.Hirsch@UCHSC.edu

²The abbreviations used are: NCI, National Cancer Institute; CIS, carcinoma *in situ*; CT, computed tomography; ASD, angiogenic squamous dysplasia; TSG, tumor suppressor gene; LOH, loss of heterozygosity; hnRNP, heterogeneous nuclear ribonucleoprotein; SPECT, second primary lung cancer; BAL, bronchoalveolar lavage; SCLC, small cell lung carcinoma; WLB, white light bronchoscopy; LIFE, laser-induced fluorescence endoscope; ELCAP, Early Lung Cancer Action Project; PET, positron emission tomography; FDG, [¹⁸F]fluoro-2-deoxyglucose.

and sputum cytology (7). The negative results from these screening studies lead the NCI and other health policy and research groups to conclude that mass screening programs involving periodic sputum cytological evaluation and chest radiographs could not be justified. However, controversies in the methodology and interpretation of the data from these studies have later been extensively discussed (8, 9). One additional study of annual chest X-ray screening is currently being conducted by the NCI; The Prostate-, Lung-, Colorectal-, and Ovarian (PLCO) screening trial. This trial includes individuals 55–74 years old, but they are not selected for this trial on the basis of high risk for lung cancer (e.g., smoking history with >20 pack-years).

The failure of clinical trials to demonstrate the efficacy of sputum cytology and chest radiography as mass screening tools has resulted in a search for better diagnostic approaches for early lung cancer detection that take advantage of recent developments in molecular biology, gene technology, and radiology (10). Furthermore, as has been the case for mammography screening for breast cancer, it has also been important to identify risk groups for lung cancer.

Although, much is known about predisposing factors, natural history, and the outcome based on histology and stage, our understanding remains very incomplete in many areas. What are the early premalignant changes molecularly, biochemically, and morphologically? Which changes are reversible and which are not? What research tools are available to provide answers to these questions? The identification of preinvasive lesions allows for developing promising methods for early intervention (11). The therapeutic paradigm and focus are today shifting from targeting only clinically verified lung cancer as previously toward targeting the premalignant and early-malignant lesions. Furthermore, the prospect of lung cancer screening has today become more meaningful as a consequence of recent developments in biology and radiology and better possibilities to define high-risk populations most suitable for lung cancer screening (12).

The present article will focus on the clinical perspectives of our biological knowledge of premalignant and early-malignant lesions and the potential of the recent technological advancement for early diagnosis of lung cancer.

Pathology of Preinvasive and Early Invasive Bronchial Lesions

Most of the efforts to classify lung cancer have been directed toward invasive carcinoma (13). However, better understanding of the pathogenesis of lung cancer aroused renewed interest in morphological abnormalities that fall short of invasive carcinoma but may indicate initiation of carcinogenesis. These morphological abnormalities are referred to as preinvasive lesions and are shown in Fig. 1. The last edition of the WHO classification of lung tumors included the classification of preinvasive lesions as a separate section. Numerous recent studies have indicated that lung cancer is not the result of a sudden transforming event in the bronchial epithelium but a multistep process in which gradually accruing sequential genetic and cellular changes result in the formation of an invasive (i.e., malignant) tumor. Mucosal changes in the large airways that

may precede or accompany invasive squamous carcinoma include hyperplasia, metaplasia, dysplasia, and CIS (14). Hyperplasia of the bronchial epithelium and squamous metaplasia have generally been considered reversible, and not premalignant in the sense of squamous dysplasia and CIS (15).

Squamous metaplasia is a common finding, especially as a response to cigarette smoking. Peters *et al.* (16) studied bronchoscopic biopsies from six sites in 106 heavy cigarette smokers; Squamous metaplasia was noted at one or more biopsy sites in approximately two-thirds of the group, and one-fourth showed squamous metaplasia in three or more biopsy sites. The incidence of squamous metaplasia increased with smoking history and was highest in individuals who had smoked more than two packs of cigarettes a day. Auerbach *et al.* (17) noted similar findings in autopsy tissues: basal cell hyperplasia and squamous metaplasia are increased in smokers in proportion to smoking history. Hyperplasia and metaplasia are believed to be reactive changes in the bronchial epithelium, as opposed to true preneoplastic changes (17, 18). The reasons for this include: (a) they are frequently found in association with chronic inflammation, and may be induced by mechanical trauma; (b) they spontaneously regress after smoking cessation; (c) in chronic smokers, the molecular changes present in these lesions are similar to those present in histologically normal epithelium; and (d) there are no reports linking their presence to increased risk for developing lung cancer. In contrast, moderate-to-severe dysplasia and CIS lesions seldom regress after smoking cessation (19).

Dysplasia and CIS are changes that frequently precede squamous cell carcinoma of the lung. Saccomanno *et al.* (20) studied more than 50,000 samples from 6,000 men, many of whom had worked in the uranium mining industry. Both smoking and uranium mining (radon exposure) were found to be associated with increased incidence of dysplasia, CIS, and invasive cancer. The studies of Saccomanno *et al.* established that increasing degrees of sputum atypia may be recognized an average of 4–5 years before the development of frank lung carcinoma.

Another question is: which grades of sputum atypia progress to cancer? From the Johns Hopkins cohort of the NCI chest X-ray/sputum screening trial, we know that among individuals with moderate atypia on sputum screening, ~10% developed known cancer up to 9 years later. Among individuals with severe atypia on the sputum screening, >40% developed known cancer during the same time period (21). Although there are data in the literature showing the relationship between sputum atypia and subsequent invasive cancer, there is still very little information about the histological progression in the bronchial mucosa in the high risk populations. In a recent publication, nine patients with CIS were followed with autofluorescence bronchoscopy at regular intervals, and 5 (56%) had progression to invasive cancer despite endobronchial therapy (22). The number of invasive cancers might even have been higher if treatment had not been given. Ongoing studies of high-risk subjects (e.g., the Colorado sputum cohort study) including serial follow-up bronchoscopies will provide evidence related to the frequency of development of invasive lung cancer as it relates to smoking history, airflow obstruction, and sputum atypia.

Since the previous WHO classification was published in

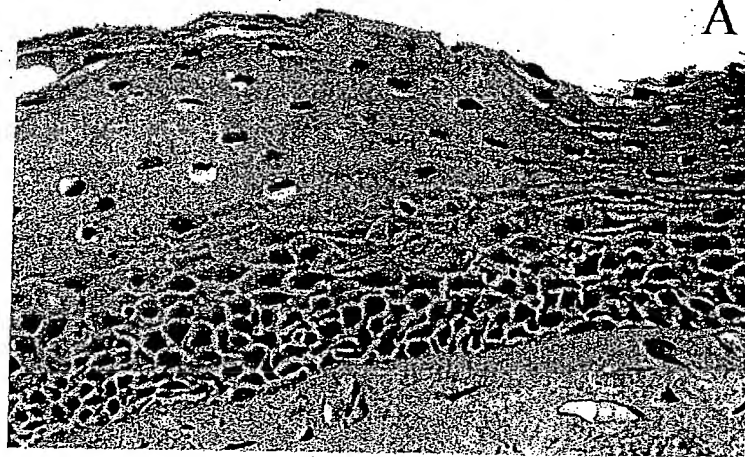
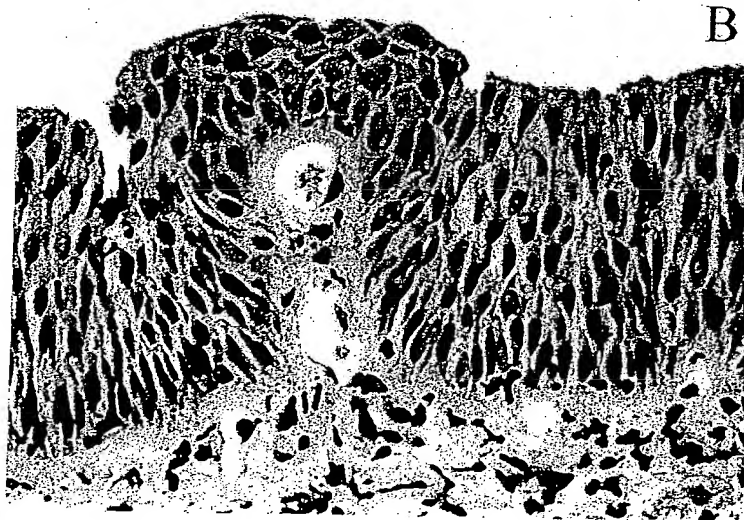


Fig. 1 A, squamous metaplasia. The cells are widely dispersed, with a regular maturation from the basal region to the top. There is keratinization, and the nuclei/cytoplasmic ratio is low. B, moderate dysplasia with ASD. Hypercellularity of the epithelium with incomplete maturation and micropapillary invasion of capillaries are seen. The nuclei/cytoplasmic ratio is high. C, severe dysplasia. There is marked pleomorphism of the cells with irregularity and prominent nucleoli.



1981, two nonsquamous lesions have been added to the WHO classification of premalignant lesions: atypical alveolar hyperplasia and diffuse idiopathic neuroendocrine cell hyperplasia (13). Both of these lesions are diagnosed rarely. The former consists of lesions <5 mm in diameter and composed of a peripheral epithelial cell proliferation with minimal cytological atypia or stromal response and resembles bronchioloalveolar carcinoma. The lesion has been seen in lung specimens resected for lung cancer, but no prospective significance of this lesion has been reported. However, this morphological lesion may play a role for the pathogenesis of peripheral lung adenocarcinomas (23, 24). The resolution of spiral CT (currently about 3 mm) approaches the diameter of these lesions, and it is anticipated that atypical alveolar hyperplasia will be increasingly encountered in subjects undergoing this procedure (25). Diffuse idiopathic neuroendocrine cell hyperplasia consists of a patchy increase in the number of well-differentiated neuroendocrine cells in the bronchioles. This process may result in the formation of small carcinoid tumors, and for this reason it is considered "preinvasive." To date, small cell carcinomas have not been associated with this lesion (13).

Recently, the use of fluorescence bronchoscopy (see below) has increased the recognition of dysplastic lesions in the large airways and a new morphological entity, ASD, was identified (26). Dysplasia of bronchial epithelium in "micropapillomatosis" and the possible link between angiogenesis and preinvasive bronchial epithelial dysplasia were recognized as early as 1983 by Muller and Muller (27), who also described the ultrastructure of these lesions. It has been suggested that this angiogenesis, which is recognized as capillary loops projecting into the dysplastic bronchial lining, is responsible for the reduced fluorescence seen in dysplastic lesions by LIFE bronchoscopes (Figs. 1 and 3; Ref. 26). Future prospective studies will show whether this morphological entity is correlated with a progression to lung cancer so as to be a target for the use of antiangiogenic agents for chemoprevention.

In general, there are several questions/problems relating to premalignant lesions, which will be addressed in future studies:

(a) The morphological criteria for premalignant and early-malignant changes, both on sputum cytology and in bronchial biopsies, have to be validated for intra- and interobserver reproducibility.

(b) Uniform and reproducible morphological/cytological criteria have to be published more extensively, and a training set of slides should be available. By the use of Internet technology, this could be more easily facilitated (28).

(c) The correlation of sputum atypia and histological changes in the bronchi in high-risk population is not well defined.

(d) The natural course of preinvasive changes in the bronchi from the high risk subjects needs to be clarified through longitudinal, prospective studies with reference to histological changes in the bronchi. Ongoing longitudinal studies with fluorescence bronchoscopy and multiple biopsies with histology and other biomarkers will define the ability of these markers to assess for risk.

(e) What is the pathology/biology of the small, often peripherally located, tumors (3 mm in diameter), which are more

often diagnosed with newer radiological techniques (e.g., low-dose spiral CT)?

(f) Optimization of the tissue procurement and processing techniques are important. Distinction of reactive from neoplastic processes is usually straightforward, but diagnostic difficulties may arise in the case of (a) inadequate or poorly prepared histological material to evaluate and (b) the presence of cytological atypia in epithelium stimulated by inflammation, viral infection, radiation, or chemotherapy.

(g) DNA array analyses of gene expression: will it be useful? How to collect proper mRNA? Can mRNA extracted from microdissected cells obtained at bronchoscopy be globally amplified and still remain representative of RNA present *in situ*?

Biology of Lung Carcinogenesis and Potential Early Detection Markers.

Lung cancer is the end-stage of multiple-step carcinogenesis, in most cases driven by genetic and epigenetic damage caused by chronic exposure to tobacco carcinogens. The genetic instability in human cancers appears to exist at two levels: at the chromosomal level, including large scale losses and gains; and at the nucleotide level including single or several base changes (29). Lung cancers harbor many numerical chromosomal abnormalities (aneuploidy) and structural cytogenetic abnormalities including deletions and nonreciprocal translocations (30). At least three classes of cellular genes are involved: proto-oncogenes, TSGs, and DNA repair genes. Oncogenic activation often occurs via point mutations, gene amplification, or chromosomal rearrangement, whereas TSGs are classically inactivated by the loss of one parental allele combined with a point or small mutation or aberrant methylation of a target TSG in the remaining allele. Additionally, dysregulated gene expression (either increased or decreased expression) can occur by other, as yet unknown, mechanisms (30). Present studies have not yet confirmed a prominent role for abnormalities of DNA repair genes in lung cancer.

Preneoplastic cells contain several molecular genetic abnormalities identical to some of the abnormalities found in overt lung cancer cells (Fig. 2). These include allele loss at several loci (3p, 9p, 8p, and 17p), *myc* and *ras* up-regulation, cyclin D1 overexpression, p53 mutations, and increased immunoreactivity, bcl-2 overexpression and DNA aneuploidy (31-35). Allelotyping of precisely microdissected, preneoplastic foci of cells suggests that the earliest changes in the bronchial epithelium is allele loss at chromosome regions 3p, then 9p, 8p, 17p, 5q, and then *ras* mutations (36-39). The biological meaning of LOH is only vaguely understood. Recent evidence suggests that LOH may be a consequence of mitotic recombination, that there is only infrequent physical loss of genetic loci, and that LOH probably precedes chromosomal duplication (40). Allelic loss would thus be significant primarily in the presence of mutation in the retained allele, and gene dosage would not be expected to exert a phenotypic effect in LOH. Some reports have indicated that *ras* activation occurs at early carcinoma stages (34). Histologically normal bronchial epithelium adjacent to cancers has also been shown to have certain genetic losses. Atypical adenomatous hyperplasia, the potential precursor lesion of adenocarcinomas, often have *Ki-ras* mutations (41).

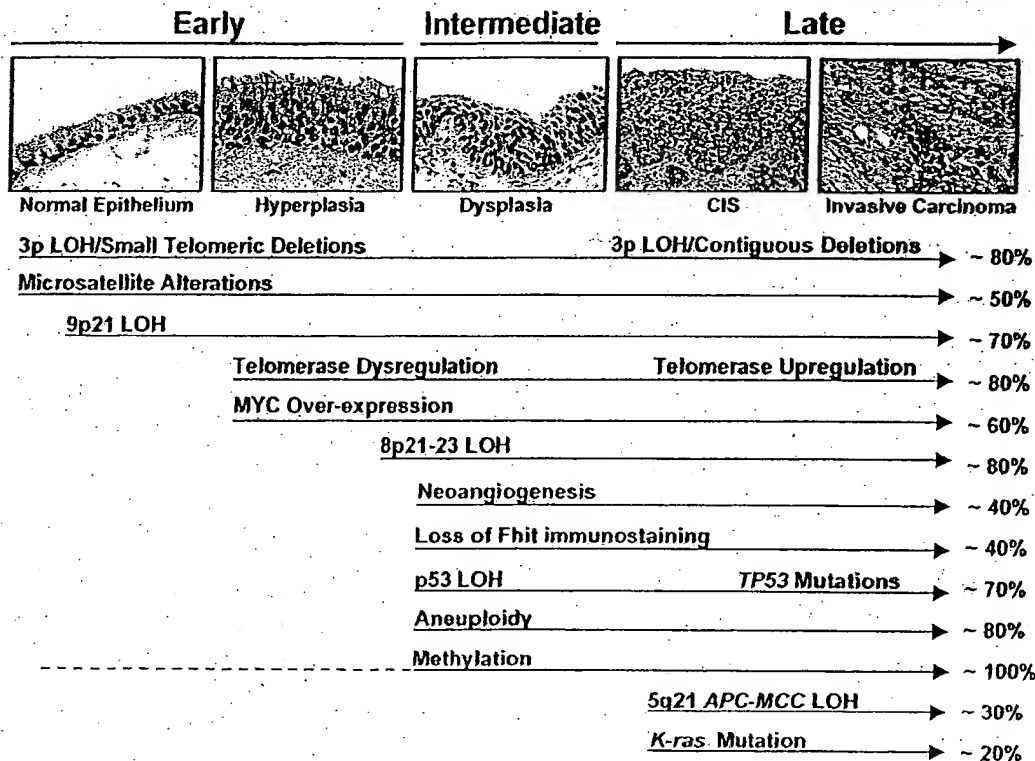


Fig. 2 Sequential changes during lung cancer pathogenesis. Although multiple genetic markers are abnormal in lung cancers, their appearance during the lengthy preneoplastic process varies. The timing of the appearance of these changes has been investigated in bronchial preneoplasia, because sequential sampling of the peripheral lung is technically difficult. Several alterations have been described in histologically normal bronchial epithelium of smokers. Other changes have been detected in slightly abnormal epithelium (hyperplasia, metaplasia) which we do not consider to be true premalignant lesions. These changes are regarded as early changes. Molecular changes detected frequently in dysplasia are regarded as intermediate in timing, whereas those usually detected at the CIS or invasive stages are regarded as late changes. It should be stressed that although there is a usual order, exceptions regarding the timing of onset may occur. Some changes are progressive, such as chromosome 3p deletions. Thus small discrete changes are present early, progressively become more extensive during pathogenesis, and frequently involve all or almost all of the arm in CIS samples. Although allelic loss at the TP53 locus may precede the onset of mutations, data on this sequence are scant. Dysregulation of the RNA component of telomerase (with its appearance in nonbasal cells) is an early event, whereas up-regulation of the gene is a relatively late event.

Molecular changes have been found not only in the lungs of patients with lung cancer, but also in the lungs of current and former smokers without lung cancer (18, 42, 43). These observations are consistent with the multistep model of carcinogenesis and "field cancerization" process, whereby the whole region is repeatedly exposed to carcinogenic damage (tobacco smoke) and is at risk for developing multiple, separate, clonally unrelated foci of neoplasia. The widespread aneuploidy that occurs throughout the respiratory tree of smokers supports this theory (44). However, the presence of the same somatic p53 point mutation at widely dispersed preneoplastic lesions in a smoker without invasive lung cancer indicates that expansion of a single progenitor clone may spread throughout the respiratory tree (45). These molecular alterations might thus be important targets for use in the early detection of lung cancer and for use as surrogate biomarkers in the follow-up of chemoprevention

studies. Detection of these mutant cells should be possible with the different molecular techniques in accessible specimens. The prospects of diagnosing these biological abnormalities in multiple types of clinical specimens are discussed below.

Specimens for Clinical Testing: Sputum

Since the 1930s, cytological examination of sputum has been used for the diagnosis of lung cancer (46). Cytological examination of sputa, especially multiple samples, is helpful for the detection of central tumors arising from the larger bronchi (e.g., squamous cell- and small cell carcinomas). Exfoliated cells from peripheral tumors, such as adenocarcinomas, arising from the smaller airways (small bronchi, bronchioles, and alveoli), especially those less than 2 cm in diameter, can be detected only occasionally in sputum samples. This has become of greater importance because the changes in cigarette exposition

(filters and decreased nicotine content) have created an increase in adenocarcinomas and a decrease in squamous carcinomas (47-49). The sensitivity of sputum cytology for early lung cancer is only in the 20%-30% range from screening studies, but by adhering to proper specimen collection, and processing and interpreting criteria, the yield can be substantially improved (50, 51). The data on the reliability of the sputum are conflicting (52-54). Browman *et al.* (52) reported interobserver agreement of 68% for exact and 82% for within - 1-category. Holliday *et al.* (54) reported low agreement within observers (27-60%) and across observers (13-50%). Within - 1-category intraobserver agreement underwent a two- or 3-fold increase in agreement, which was also the case for interobserver agreement. The variation in intra- and interobserver agreement seems to depend on experience among the cytotechnicians/cytopathologists and the composition of categories studied. A higher degree of agreement is obtained for higher grades of dysplasia (54). Risse *et al.* (55) showed that the ability to detect premalignant conditions is dependent on the number and type of cells present in the deeper airways, suggesting a mode of improvement that is unrelated to observer reliability. MacDougall *et al.* (56) concluded that sputum cytology was too insensitive and insufficiently accurate to be included in the routine work-up of any patient suspected of having lung cancer. To improve the reliability of sputum cytology examinations a simplification of the diagnostic categories from 6 (normal; squamous metaplasia; mild, moderate, and severe atypia; and carcinoma) to 2-3 categories have been proposed (54). Future clinicopathological studies will be required to validate this concept.

To improve the sensitivity of sputum examination as a population-screening tool for the detection of early lung cancer, several approaches are currently under development.

Immunostaining. Annual sputum specimens obtained from individuals screened at Johns Hopkins were obtained, and the patients were monitored for 8 years (57). Because the clinical outcome of these patients was known, archival sputum specimens were screened for the presence of biomarkers that could indicate the presence of lung tumors in an early, preinvasive stage. In an attempt to distinguish the pattern of marker expression Tockman *et al.* (58) studied two monoclonal antibodies. Positive staining predicted subsequent lung cancer approximately 2 years before clinical recognition of the disease, with a sensitivity of 91% and a specificity of 88% (58). One of these antibodies (703 D4) had a higher sensitivity and was later identified as recognizing hnRNP A2/B1 (59). The role of hnRNP A2/B1 overexpression for detecting preclinical lung cancer has been studied in a large high-risk population including 6000 Chinese tin miners who were heavy smokers and who had an extraordinary rate of lung cancer (60). The results from this study indicated that detection of hnRNP A2/B1 overexpression in sputum epithelium cells was 2- to 3-fold more sensitive for detection of lung cancer than standard chest X-ray and sputum cytology methods. The method was particularly effective in identifying early disease (60). The sensitivity was 74% versus 21% for cytology and 42% for chest X-ray. However, the biomarker had a lower specificity (70%) compared with cytology (100%) and chest radiograph (90%). An ongoing clinical trial is evaluating the performance of the A2/B1 protein as a biomarker for the early detection of SPLC. The patients at risk

for SPLC have the highest incidence of lung cancer (2-5%) among asymptomatic populations (61). In this trial, 13 SPLCs were identified by A2/B1, and the sensitivity and specificity were 77-82% and 65-81%, respectively. Among the cases identified as positive by immunocytochemistry and image cytometry, 67% developed SPLC within 1 year (62). Whereas the previous immunocytochemistry studies on material from the older screening material from the NCI-supported screening studies were made on sputum cells cytologically classified with moderately or gravely atypical metaplastic appearance, the latter studies have been done on cytologically "normal appearing" cells. More recently Sueoka *et al.* (63) reported the confirmation of the value of overexpression of hnRNP A2/B1 to detect preclinical lung cancer in Japan. Efforts to improve the sensitivity of hnRNP markers are ongoing (64).

PCR Techniques. PCR techniques have been used for the evaluation of molecular biomarkers for early lung cancer detection. In a pilot study with selected patients from the Johns Hopkins Lung Project (JHLP), 8 (53%) of 15 patients with adenocarcinoma or large cell carcinoma were detected by mutations in sputum cells from 1 to 13 months before clinical diagnosis (65). However, the method seemed to be less sensitive than the protein marker described above, and the identification of specific gene abnormalities is further limited by the need to know the specific mutation sequence with which to probe the sputum specimens. Currently, this approach is not practical for screening undiagnosed individuals. Future advances in gene chip technology may permit testing for all possible mutations of common oncogenes and TSGs in clinical specimens of asymptomatic individuals (62).

Microsatellite markers are small repeating DNA sequences found in the noncoding regions of a gene. PCR amplification of these repeat sequences provides a rapid method for assessment of LOH and facilitates the mapping of suppressor genes (66, 67). Microsatellite alterations are extension or deletions of these repeated elements. Detection of microsatellite alterations in histological or cytological specimens may facilitate the detection of clonal preneoplastic or neoplastic cell populations. Although the detection of microsatellite alterations does not indicate the specific genetic change in the tumor, detection of clonal cell populations might serve as a cancer screening marker (65). Identical alterations have been found in lung cancers and corresponding sputum samples demonstrating minimal atypia (68). The *p16* gene is located on the short arm of chromosome 9(9p21) and is frequently mutated or inactivated in tumors and cell lines derived from lung cancer (69, 70). Belinsky *et al.* (71) measured hypermethylation of the CpG islands in the sputum of lung cancer patients and demonstrated a high correlation with early stages of non-small cell lung cancer, which indicated that *p16* CpG hypermethylation could be useful in the prediction of future lung cancer. However, prospective studies are needed to evaluate the role of *p16* hypermethylation as a marker for early lung cancer detection. Multiple other genes are inactivated by hypermethylation in lung cancer (72), and the detection of hypermethylation may be useful for risk assessment and early diagnosis.

Computer-assisted Image Analysis. Computer-assisted image analysis was initially used to detect malignancy-associated changes (e.g., subvisual or nonobvious changes in the

distribution of DNA in the nuclei of histologically normal cells in the vicinity of preinvasive or invasive cancer; 73). In a retrospective analysis of sputum cytology slides, malignancy-associated changes alone correctly identified 74% of the subjects who later developed squamous cell carcinoma (74). The technique has been improved, and recent data showed sensitivities of 75% for stage 0/I lung cancer and 85% for adenocarcinomas with a specificity of 90% (75). This quantitative microscopy technique allows the examining of thousands of cells per slide within a relative short time. Similar techniques have been approved in the United States for cervical cancer screening, and might, in the future, play a role for lung cancer screening. However, no prospective clinical studies has evaluated this technology in a larger lung cancer screening setting.

High Throughput Technology. With future advances in gene chip technology, it might become feasible to probe for expression of multiple genes in sputum specimens of asymptomatic individuals. However, this requires a large amount of undegraded RNA from respiratory tract cells. With the high throughput technology, a higher sensitivity might be achieved by using multiple markers at the cost of achieving a lower specificity, which would be undesirable for a screening study.

In conclusion, we need to reevaluate the role of sputum cytology for screening and early detection of lung cancer because of advances in biomarkers and technology. Ongoing studies with standard and biomarker analysis in high-risk groups might change the previous negative attitude and provide a new perspective on sputum cytology as a mass screening tool when applied in a high-risk population. Adding different molecular diagnostic tests gives the possibility for early diagnosis far in advance of clinical presentation. However, validation of the tests in larger prospective studies is necessary, and the individual tests have to be compared with each other to define the role of early diagnosis in the overall management of high-risk subjects. Furthermore, health economic issues have to be considered.

Specimens for Clinical Testing: BAL

BAL involves the infusion and reaspiration of a sterile saline solution in distal segments of the lung via a fiberoptic bronchoscope. Ahrendt *et al.* (76) examined a series of 50 resected non-SCLC tumor patients and compared the tumor and BAL with regard to molecular markers including p53 mutations, K-ras mutation, the methylation status of the CpG island of the p16 gene, and microsatellite alteration (Tables 1 and 2). With the possible exception of the test for microsatellite alteration, all of the tests had relatively high sensitivity and could detect mutant cells in the presence of a large excess of normal cells. The frequencies of these changes in the tumors ranged from 27% (for K-ras mutations) to 56% (for p53 mutations). As expected, p53 mutations were more frequent in central (predominantly squamous cell) tumors, and K-ras mutations were more frequent in peripheral (predominantly adenocarcinoma) tumors. The specificity was high (nearly 100%) because, with the exception of microsatellite alterations, the same genetic change in BAL sample as in tumors was always found, but the sensitivity was low, and in only 53% of tumors that contained molecular lesions were the same abnormalities detected in corresponding BAL fluids. Specifically, the tests were least helpful in the

group of patients in whom improved diagnostic abilities are most needed, those with small, peripherally located tumors (77). Unfortunately, the investigators were not able to compare the molecular tests with routine cytopathological analysis of the BAL specimens. The sensitivity of the molecular tests in BAL specimens has to be improved, and we need to know the results from subjects at increased risk (current and former smokers without lung cancer and survivors of previous cancer of the upper respiratory tract) and subjects with chronic lung diseases as well as results from healthy never smokers.

A European group has previously shown that genetic alterations detected in DNA from bronchial lavage of individuals with lung cancer were also found in individuals with no evidence of malignant disease (78), which raises the question about the specificity of such molecular damage in neoplastic conditions. To improve the sensitivity and specificity of detecting allelic imbalance in lung tumors, high throughput PCR-based microsatellite assays have been established (79). In a recent study by Fielding *et al.* (80), the up-regulation of hnRNP A2/B1 was found to be a promising marker in BAL for the detection of premalignant and malignant bronchial lesions with a diagnostic sensitivity of 96% and a specificity of 82%.

It is too early yet to make conclusions as to whether BAL examinations will add to other pathological/molecular biological clinical studies. To obtain diagnostic material for BAL bronchoscopy is required, and we do not have any data that compare BAL examinations with biopsies. Thus, we do not know whether BAL is a valuable adjunct to the biopsies taken under the same bronchoscopy procedure.

Specimens for Clinical Testing: Peripheral Blood

For many years scientists have searched for a lung cancer-specific tumor marker that could be detected in peripheral blood. Optimism was raised in the "early" immunocytochemistry era by the use of monoclonal antibodies raised against more-or-less specific epithelial epitopes. In the search for epithelial cells in peripheral blood and bone marrow, monoclonal antibodies against cytokeratin have been used. However, these reactions are clearly not cancer-specific, and some antibodies have been shown to cross-react with normal blood or bone marrow elements (81, 82). Another explanation could be that cells from the macrophage/monocyte system may contain proteins derived from the primary tumor that have undergone necrosis and apoptosis and that these processed proteins are recognized by the antibodies (82). On the basis of "traditional" immunocytochemistry, no markers have been able to detect premalignant or early-malignant disorders based on a peripheral blood sample. However, with the development of DNA technologies, new possibilities have been raised, and, with the use of PCR techniques, some promising reports have been published.

Nanogram quantities of DNA circulating in blood are present in healthy individuals (83, 84). Tumor DNA is also released into the plasma component in increased quantities (85, 86). Thus, the plasma and serum of cancer patients is enriched in DNA, an average four times the amount of free DNA as compared with normal controls (87). In a study by Chen *et al.* (88), a comparison of microsatellite alterations in tumor and plasma DNA was done in SCLC patients, and 93% of the patients with

Table 1 Tissues and other resources for the study of molecular markers

| Specimen | Ref. | Comments |
|--|-----------------------|---|
| Tumor tissue | Numerous | Mixture of cell types, may require microdissection (139). Extensively used for most studies. Alcohol-fixed fine-needle aspirates may be used for mutational and other studies. |
| Sputum | 65, 68, 71, 74 | Respiratory cells usually in small minority. Most samples fixed in Saccomanno's fixative. Several studies have been performed on these specimens many years later. |
| Surrogate organ | 140 | Predominantly squamous epithelial cells. Buccal smears, brushings of tongue or tonsil may be explored as potential surrogate organs resulting from the field effect of tobacco damage of the entire upper aerodigestive tract. This concept needs to be confirmed. |
| Bronchial brush/wash | 141, 142, 143 | Predominantly respiratory cells. Fresh, frozen, or alcohol-fixed samples are suitable for multiple studies including FISH.* |
| Bronchial tissues | 42, 43, 45, 144, 145 | Usually from bronchial biopsies, but may be obtained from surgical resection specimens. Formalin fixation and paraffin embedding required for histological diagnosis, although EASI preps may permit identification and isolation of subpopulations. Paraffin sections may be used for genotyping polymorphisms, for allelotyping, and for <i>in situ</i> hybridization. |
| BAL fluids | 76, 78, 146, 147, 148 | BAL fluids are useful for examining the peripheral airway cells, which are the precursor cells of most adenocarcinomas. Numerous mononuclear cells present. Enrichment of epithelial cells desirable. |
| Blood components | 72, 92, 149 | Analysis of circulating tumor cells and genetic material shed by dying tumor cells into the plasma component may yield useful biological and diagnostic information. Gene mutations and presence of epithelial cell markers have been used to detect circulating tumor cells. Gene mutations, allelic loss, microsatellite alterations, and aberrant methylation have been used to identify tumor cell DNA released into the fluid compartment. |
| Tissue for molecular staging | 150, 151 | Although little data exists for lung cancers, regional lymph nodes, sentinel lymph nodes, and surgical resection margins have been used in other tumor types for molecular staging. |
| Tumor cell lines | 152, 153 | Provide an unlimited self-replicating source of high-quality molecular reagents and have been used for numerous studies. Cell lines may or may not reflect the properties of the tumors from which they were derived (26), although they probably represent cellular subpopulations (27). Aggressive metastatic tumors are more likely to be successfully cultured (28) resulting in skewed data. |
| Cultures of nonmalignant tissues | 154, 155 | Epithelial cultures may be useful for studying molecular changes during multistage pathogenesis. Temporary as well as a few immortalized cultures from nonmalignant epithelial cells have been established. B-lymphoblastoid cultures are useful for linkage analysis, for genetic susceptibility studies, and for allelotyping corresponding tumors. |
| Nonmalignant tissue from patients and from cancer-free relatives | 156, 157, 158 | Tissues such as buccal smears, tumor-free lymph nodes, and peripheral blood mononuclear cells are useful as controls for linkage analysis, for genetic susceptibility studies, and for allelotyping corresponding tumors. |

* FISH, fluorescence *in situ* hybridization; EASI, epithelial aggregate separation and isolation.

microsatellite alterations in tumor DNA also had modifications in the plasma DNA. However, some patients had LOH only in the tumor DNA. Because most of the microsatellite alterations were similar in tumor DNA and plasma DNA, they concluded that some of the DNA circulating in the blood comes from the tumor. Thus, modifications of circulating DNA can be used as an early detection marker. Detection of aberrant DNA methylation in serum DNA in patients with non-SCLC has been reported (72). Although the number of patients was small and the hypermethylated DNA was found in all stages, it opens up for the possibility to be used as an early lung cancer detection marker. Furthermore, *p53* and *ras* gene mutations have been

detected in the plasma and serum of patients with colorectal cancers (89-91), pancreatic carcinomas (92, 93), and hematological malignancies (94).

In conclusion, the limited direct accessibility of lung carcinomas has led to efforts to identify tumor-associated soluble markers in serum or plasma. Many of the currently recognized soluble markers were first identified as "tumor" markers but, when evaluated in nonneoplastic tissue, have often been found in normal cells as well as in tumors. For early detection of lung cancer, we need more clinical data evaluating these new molecular biological markers from multiple sites, especially in high-risk groups.

Table 2 Molecular approaches for lung cancer investigation

| Specimen | Ref. | Comment |
|--|------------------------------|--|
| Gene mutations | 159, 160, 161 | Widely used technique, especially for <i>p53</i> and <i>ras</i> genes. Often used to determine the role of a newly discovered gene in the pathogenesis of lung cancer. May be of diagnostic and prognostic significance. Multiple methodologies available. |
| Allelotyping | 18, 158 | Useful as a partial substitute for mutational analysis and for determining the chromosomal locations of putative tumor suppressor genes. Widely used to study multistage pathogenesis. Readily performed on formalin-fixed and microdissected tissues. Increasing use of genotyping using automatic sequencers. |
| Gene expression at RNA and protein level | 145, 162, 163, 164, 165, 166 | Northern blotting and reverse transcription-PCR are widely used to investigate gene expression. Western blotting often used for detection of protein expression. <i>In situ</i> hybridization for message expression can be performed on paraffin-embedded tissues and, thus, can be used to investigate multistage pathogenesis. Microarray techniques offer promise of examining all or most of the genome but currently require relatively large amount of high-quality RNA from purified cell populations. Sage technique useful for investigation and identification of expressed genes. Similarly, advances in proteomics will permit simultaneous detection of multiple proteins. Numerous immunohistochemical studies of oncogene expression have been used to study multistage pathogenesis. Of particular interest, hnRNP expression on exfoliated epithelial cells in sputum samples may predict for development of cancer. |
| Molecular cytogenetics | 40, 167, 168, 169, 170 | <i>In situ</i> hybridization studies of fixed materials or using smears has provided considerable information about numerical and structural changes. |
| Comparative genomic hybridization | 171, 172 | Useful for detection of gene amplifications. Less sensitive for the detection of regions of allelic loss. |
| Morphometric studies | 74, 173, 174 | May be applied to paraffin-embedded tissues. Useful for determining aneuploidy and for measuring a number of nuclear and cytoplasmic parameters. |

Specimens for Clinical Testing: Bronchoscopy

WLB is the most commonly used diagnostic tool for obtaining a definite histological diagnosis of lung cancer. Bronchoscopy has major diagnostic limitations for premalignant lesions. Because these lesions are only a few cells thick (0.2–1 mm) and have a surface diameter of only a few millimeters, they rarely are observed as visual abnormalities. Woolner (95) reported that squamous cell CIS was visible to experienced bronchoscopists in only 29% of cases. To address this limitation, fluorescence bronchoscopy was developed. Early studies of fluorescence bronchoscopy entailed the use of fluorescent drugs (hematoporphyrin dyes) that were preferentially retained in malignant tissue (96). Although, studies evaluating this approach did, in fact, show that early invasive and *in situ* cancers could be localized, the detection of dysplasia remained problematic (97–100). Furthermore, the development of photodynamic diagnostic systems was hampered by problems including skin photosensitizing and interference with tissue autofluorescence. To overcome these problems, a new laser photodynamic diagnostic system was developed (101). This system detected tumor-specific drug fluorescence at 630 nm wavelength, which is far from normal tissue autofluorescence (500–580 nm), and interference by autofluorescence from normal tissue should then have been eliminated, but it remained a significant problem (102).

Another approach was developed by Palcic *et al.* (103), who noticed the lack of autofluorescence in the tumor lesions by using blue light (442 nm) rather than white light to illuminate the bronchial surface. They amplified the difference in autofluorescence between normal, premalignant, and tumor tissue for clinical use (103, 104). Using a high-quality-charge coupled device and special algorithm, the LIFE was developed, taking advantage of the principle that dysplastic and malignant tissues reduce autofluorescent signals compared with normal tissue (Fig. 3).

Several studies have been performed comparing the diagnostic specificity and sensitivity of LIFE bronchoscopy versus WLB in diagnosing preinvasive and early-invasive lesions (105–109; Table 3). Most of the studies reported a higher diagnostic sensitivity of LIFE bronchoscopy in the detection of premalignant and early-malignant lesions at the cost of lower specificity (i.e., more false-positive results). In most of these studies, lesions with moderate dysplasia or worse were the target of the study and rated as "positive." The prevalence of preinvasive and early lung cancer varies widely from one study to another, from 20.2% (105) to 65.8% (102). The explanation might be beyond the risk profile of genetic variations or different levels of experience among the endoscopists as well as the pathologists involved. Furthermore, there seems to be a training effect in using the LIFE bronchoscope, which has been demon-

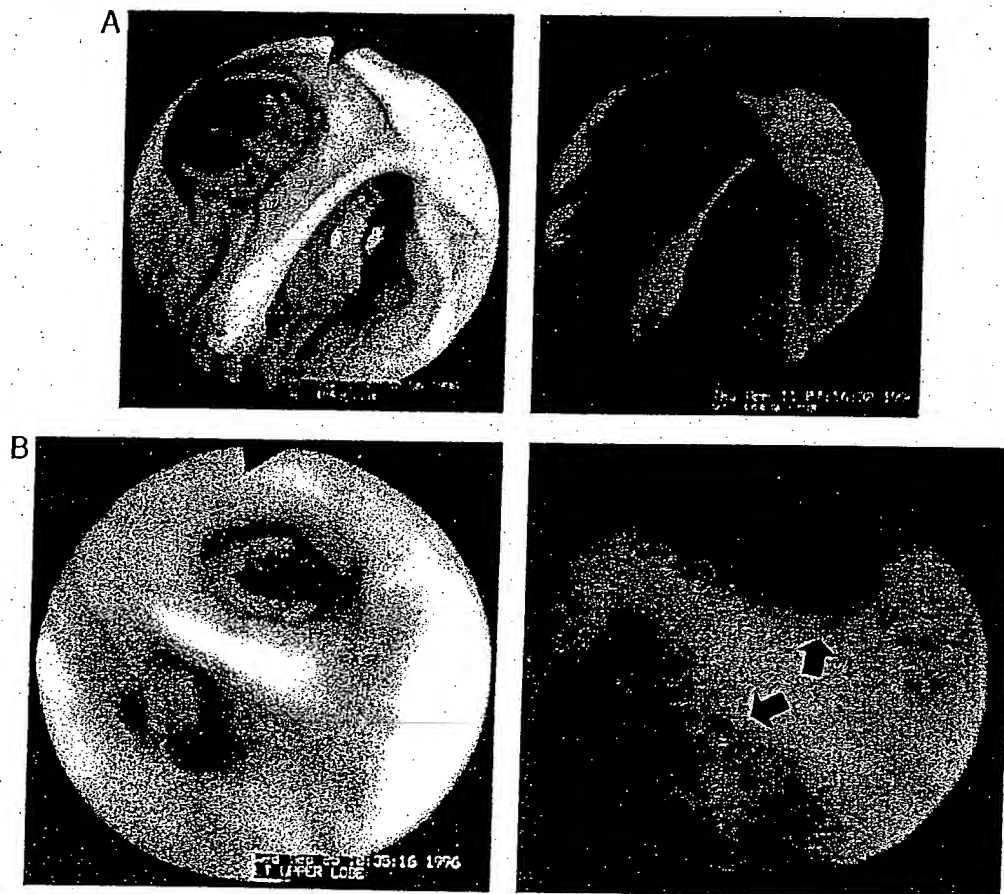


Fig. 3 A, normal WLB and normal LIFE bronchoscopy. B, WLB shows inflammatory changes in the bronchial mucosa but no suspicion of malignancy (left). LIFE bronchoscopy shows diffuse reduced autofluorescence (visualized by diffuse red-brownish colorization; arrows). Biopsy demonstrated diffuse severe dysplasia.

strated by Venmans *et al.* (107). In their study, the diagnostic sensitivity increased from 67 to 80% when comparing the first and the second half of the study. The use of the LIFE device in conjunction with WLB improved the detection rate of preneoplastic lesions and CIS significantly (Table 3). Kurie *et al.* (106) looked for more subtle tissue transformation, but their study included few patients with moderate dysplasia or worse. No improvement in the evaluation of metaplasia index was observed by the use of LIFE bronchoscopy. Thus, differences in the study population might explain the different conclusion. There are still no clinical studies with sufficient long-term data showing that moderate dysplasia is the most relevant clinical predictor of eventual malignancy. Limitations in making conclusions from the existing studies are also the potential methodological bias related to the order in which the different bronchoscopy procedures are done and whether the same examiner has performed both procedures. To address these issues, a

prospective randomized study between LIFE bronchoscopy and WLB was done at the University of Colorado Cancer Center. The study design included a randomization with regard to the order of procedure as well as the order of the individual bronchoscopist (109). The order of the procedure and of the individual bronchoscopist did not affect the results. The study also demonstrated a significantly higher sensitivity in detecting premalignant lesions visualized by the LIFE, but at the cost of a lower specificity (109). The reason for the low diagnostic specificity found with the LIFE bronchoscopy in the different studies might be attributable to the visualization of more abnormal foci with the LIFE bronchoscope, with the consequence that a larger number of biopsies were taken and, thus, there was a higher risk of more false-positive results. The use of LIFE bronchoscopy has led to the identification of a new morphological entity, the ASD, which is described above. In a recent morphological study angiodysplastic changes were frequently found in preneoplastic

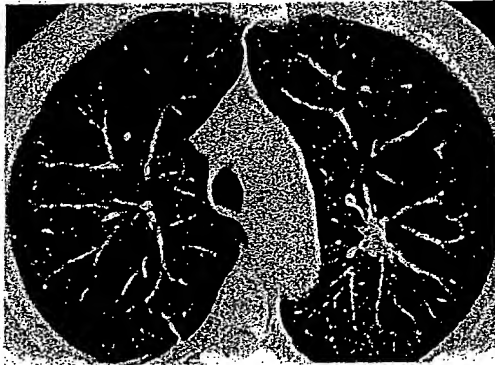


Fig. 4 Seventy-one-year-old man with a spicular nodule in upper left lobe demonstrated on low-dose helical CT (picture), but not visible on chest X-radiography. CT-guided biopsy showed adenocarcinoma.

and early-malignant lesions in the bronchi (26). The morphological entity has been confirmed in preneoplasias among smokers, and the perspectives of this finding have been extensively discussed (110). The prognostic significance of this morphological entity is currently studied in ongoing long-term follow-up studies. Future studies have to evaluate the role of ASD as a biomarker for early lesions and whether it can be used as a marker for treatment effect or therapeutic target for chemoprevention.

The LIFE bronchoscope may play an important role in the screening and follow-up of subjects at high risk of developing lung cancer. At this stage, however, it is unknown whether the LIFE bronchoscope will lead to a reduction in lung cancer mortality. There are also no data on cost-effectiveness and cost-benefit analyses available for this new diagnostic procedure. The use of the LIFE bronchoscope may also in the future be extended to other indications, e.g., patients staged as having resectable lung cancer on one side. Whether LIFE bronchoscopy of the contralateral lung will disclose abnormalities, which would change the therapeutic decision, is not yet reported.

Recent Advances in Radiology

The previous NCI-sponsored screening trials failed to demonstrate any reduction in the lung cancer mortality by sputum cytology and yearly chest radiography as mass screening tools for lung cancer screening. Limitations of design and execution of the studies, however, have been discussed extensively (8, 111, 112). An extended follow-up (median, 20.5 years) of the Mayo Lung Project was recently published (113). There was still no difference in lung cancer mortality between the intervention arm and the control arm (4.4 versus 3.9 deaths per 1000 person-years). However, the median survival for patients with resected early-stage disease was 16.0 years in the intervention arm versus 5.0 years in the usual-care arm ($P < 0.05$). The latter findings have raised the question as to whether some small lesions with limited clinical relevance may have been identified in the intervention arm, and the question of "overdiagnosis" was discussed in accompanying editorials (114).

Mass screening for lung cancer has been performed in Japan for many years and has been performed in over 500,000 people in about 80% of the local communities (115). Sobue *et al.* (116) observed that annual clinic-based chest X-ray screening for lung cancer in Japan showed reduced lung cancer mortality by about one-fourth among individuals who underwent screening once a year. In this screening program, the relative odds ratio of dying from lung cancer within 12 months was 0.535 and in the 12–24-month period was 0.638 (117). However, many studies have focused on the pitfalls in the detection of abnormalities by radiography (118–122). The limit of chest radiographic sensitivity for nodule detection is roughly 1 cm in diameter, by which time the tumor has over 10^9 cells and may already have violated bronchial epithelium and vascular epithelium. CT has been shown to be more effective in the detection of peripheral lung lesions compared with plain radiography or conventional tomography of the whole lung (123, 124).

Spiral CT scan is a relatively new technology with the ability to continuously acquire data resulting in a shorter scanning time, a lower radiation exposure, and improved diagnostic accuracy compared with those of plain radiography (125–127). Spiral CT allows the whole chest to be imaged in one or two breath-holds, reducing motion artifacts and eliminating respiratory misregistration or missing nodules. Although there is greater radiation exposure with CT than with chest radiography, low-dose techniques (lower mA of 30–50 compared with 200 for conventional CT) have achieved calculated exposure doses that are 17% that of conventional CT and 10 times that of chest radiographs. Further reduction in radiation dose while maintaining diagnostic accuracy is a topic of current research. Furthermore, for the baseline screening, low-dose spiral-CT-scan i.v. contrast is not administered. Nodules as small as 1–5 mm can be shown with modern spiral CT technology (25, 128). The obvious advantages with this new technology led some groups in Japan and in the United States to look to low-dose spiral CT as a tool for screening (Refs. 129–131; Tables 4 and 5).

In a Japanese report, spiral CT scans and chest radiographs were done twice a year in 1369 individuals (129). Peripheral lung cancer was detected in 15 (0.3%) of 3457 examinations, and, among the 15 lung cancer cases detected, the results of chest X-ray were negative in 11 (73%), and the tumors were detected only by low-dose spiral CT. The detection rates of low-dose spiral CT and chest X-ray were 0.43% (15 of 3457 examinations) and 0.12% (4 of 3457 examinations), respectively. Furthermore, 14 (93%) of the 15 lung cancers were stage I disease. The histology showed that 11 of the 15 lung cancer cases were adenocarcinoma, and 4 had squamous cell carcinoma. The effective exposure dose with spiral CT scan in that study was calculated to about one-sixth that of conventional CT.

The ELCAP in New York was designed to determine: (a) the frequency with which nodules were detected; (b) the frequency with which detected nodules represent malignant disease; and (c) the frequency with which malignant nodules are curable (131). In the ELCAP study, 27 lung cancers were found among 1000 subjects screened. Among the 27 patients with cancer, 85% had stage I disease (Table 5).

Another population-based study on low-dose CT screening has been published by Sone *et al.* (130), using a mobile low-dose spiral CT scanner. The detection rate was 0.48% (*i.e.*, 4–5

Table 3 Bronchoscopy versus WLB in diagnosing premalignant and early-malignant lesions

| Author | Sensitivity | | | | Specificity | | | | Predictive values | | | | | | | |
|--|-----------------|-------|------|----------------------|------------------------|------|----------------------|-------|-------------------|------|------|------|------|------|------|-----|
| | No. of biopsies | LIFE+ | | Relative sensitivity | LIFE+ | | Relative specificity | LIFE+ | PPV* | NPV | PPV | NPV | PPV | NPV | PPV | NPV |
| | | WLB | LIFE | | WLB | LIFE | | | | | | | | | | |
| Lam <i>et al.</i> (105) | 700 | 0.67 | NR | 0.25 | 6.3 (2.7) ^c | 0.66 | NR | 0.90 | NR | 0.33 | 0.89 | NR | NR | 0.39 | 0.83 | NR |
| Kurie <i>et al.</i> ^b (106) | 234 | NR | 0.38 | NR | NR | 0.56 | NR | NR | NR | NR | 0.16 | 0.81 | NR | NR | NR | NR |
| Vennans <i>et al.</i> (107) | 139 | NR | 0.89 | 0.78 | 1.43 | NR | 0.61 | 0.88 | NR | 0.20 | NR | 0.14 | 0.99 | 0.32 | 0.98 | NR |
| Vermulen <i>et al.</i> (108) | 172 | 0.93 | NR | 0.25 | 3.75 | 0.21 | NR | 0.87 | NR | 0.13 | 0.96 | NR | NR | 0.19 | 0.90 | NR |
| Kennedy <i>et al.</i> (109) | 394 | 0.79 | 0.72 | 0.18 | 4.4 | 0.3 | 0.43 | 0.78 | 0.38 | 0.21 | 0.85 | 0.25 | 0.87 | 0.17 | 0.80 | NR |

* PPV, positive predictive value; NPV, negative predictive value; NR, not reported.

^b Based on reference pathologist.^c If invasive carcinoma is included.

Table 4 Results from three population-based screening studies with low-dose spiral CT (LDCT)

| Authors | No. of individuals studied | True positive <i>n</i> | False positive ^a % | Predictive value % | Detection rate % | | Pack-yr | Age incl. yr |
|------------------------------|----------------------------|------------------------|-------------------------------|--------------------|------------------|-------|------------------|--------------|
| | | | | | LDCT | X-ray | | |
| Kaneko <i>et al.</i> (129) | 1369 | 15 | 15.6 | 6.6 | 0.43 | 0.12 | >20 | >50 |
| Sone <i>et al.</i> (130) | 3967 | 19 | 5.0 | 8.8 | 0.46-0.5 | | >30 ^b | 40-74 |
| Henschke <i>et al.</i> (131) | 1000 | 27 | 20.1 | 11.6 | 2.7 | 0.70 | >10 ^c | >60 |

^a Defined as individuals with "test-positive," in whom further workup gave no suspicion of malignancy.^b The study also included a group of nonsmokers.^c Average = 45 (not reported in the other studies).

Table 5 Histology, stage, and size of primary lung cancer detected by low-dose spiral CT

| Author | No. of cancers/ No. screened | Histology % | | | TNM % | | | | Size (mm) | | | | |
|------------------------------|---------------------------------|--------------------|--------|-------|-------|----|-----|----|-----------|-------|-----|-------|-----|
| | | Adeno ^a | Squam. | Other | I | II | III | IV | Average | Range | <10 | 11-20 | >21 |
| Kaneko <i>et al.</i> (129) | 15/1369 (1.1%) | 73 | 17 | | 93 | | 7 | | 12 | 8-18 | | | |
| Sone <i>et al.</i> (130) | 19/5483 (0.3%) | 63 | 5 | 32 | 84 | | | 16 | 17 | 6-47 | 4 | 14 | 3 |
| Henschke <i>et al.</i> (131) | 27/1000 (2.7%) | 67 | 3 | 30 | 85 | 4 | 11 | | | | 15 | 8 | 4 |

^a Adeno, adenocarcinoma; Squam., squamous cell carcinoma; TNM, tumor-node-metastasis.

cases per 1000 examinations). Surprisingly, there was no difference in the detection rate among smokers (0.52%) versus nonsmokers (0.46%). The results from the three population-based studies are summarized in Tables 4 and 5. The conclusion from these studies is that 85% of the lung cancers detected by low-dose CT were in stage I, offering improved possibility for curative treatment and better prognosis in general. However, the issue of "false-positive" scans has to be taken into consideration. Thus far, up to 20% of the participants with nodules on the scan had no malignancy during the follow-up period. The possibility that the cancers found represent incidental cancers as in the Mayo Lung Project must also be considered (114). The results from these studies confirm the expectation that low-dose CT increases the detection of small noncalcified nodules and, that lung cancer at an earlier and more curable stage are detected. The mobile CT screening study by Sone *et al.* (130) showed that low-dose CT increased the likelihood of detection of malignant disease 10 times as compared with radiography. The overall rate of malignant disease was lower in the Japanese studies (129, 130) compared with the ELCAP study (Ref. 131; detection rates 0.43-0.48% versus 2.7%). This could be because the Japanese studies screened individuals from the general population ages

40-74, whereas ELCAP screened people at high risk, ages ≥ 60 , with a tobacco history of at least 10 pack-years. Thus, as expected, the risk of the population to be screened affects the rate of cancer detection.

Questions remaining to be answered include: (a) what are the diagnostic sensitivity and specificity of this procedure; and (b) does screening reduce lung cancer mortality? The spiral CT has not been as sensitive for small central cancers as it is for small peripheral cancers (129, 131). Minute nodules of lung cancer that are near the threshold of detectability may be overlooked at spiral CT screening (132). A prospective study of the diagnostic sensitivity of spiral CT has recently shown that the diagnostic sensitivity exceeded the sensitivity of conventional CT in previous reports (25). However, there were limitations in the detection of intrapulmonary nodules smaller than 6 mm and of pleural lesions. Compared with surgery (thoracotomy with palpation of deflated lung, resection, and histology), the sensitivity of spiral CT was 60% for intrapulmonary nodules of <6 mm and 95% for nodules of ≥ 6 mm and was 100% for neoplastic lesions ≥ 6 mm. Furthermore, a marked difference in the sensitivities of two independent observers was found for nodules smaller than 6 mm, whereas agreement was much better for

6–10-mm nodules (25). Given these promising preliminary clinical results, further research is needed to determine the optimal technique for spiral CT screening, which includes collimation, reconstruction interval, pitch, and viewing methods. Decreasing the slice thickness to 3 mm, monitoring the viewing of examinations, and computer-aided diagnosis have been used to improve the diagnostic capability of spiral CT in the detection of pulmonary nodules (133–136).

Future large scale randomized studies have to confirm whether in fact spiral CT screening will lead to a reduction in lung cancer mortality. In a randomized study, the following questions arise: (a) what is the optimal high-risk group to study and what should be the control arm? (b) what should be the end points (goals) of the studies? The ultimate goal is to reduce the lung cancer mortality. However, although this is a long-term goal, intermediate end points from such studies should be evaluated. The change to more curable stages at diagnosis for the lung cancer patients is one such immediate goal; (c) what is the optimal workup and the morbidity of this program? (d) what is the cost of such a screening program? and (e) what is the false-positive rate of the screening findings? Incorporation of smoking cessation programs should be included in the future design of screening studies because it has been shown that screening with low-dose CT in participants who are still smoking provides substantial motivation for smoking cessation (137).

The studies with spiral CT-scan have demonstrated the superior diagnostic ability in the detection of small peripherally located tumors, most of the malignant ones of adenocarcinoma type of histology. The diagnostic sensitivity of spiral CT for more centrally located tumors (mostly squamous cell carcinoma) is significantly lower than for the peripherally located ones. Through these spiral CT studies, we will learn about the biology, pathology, and clinical course of these small tumors, which might be different from what we know about clinically more evident tumors detected routinely in previous studies.

Because lung cancer is so common, the introduction of any new screening technique in this area has to be underpinned by careful definition of the cost implications and must be justified by compelling evidence. The cost-effectiveness of the spiral CT approach should be assessed by evaluating the rate of over-diagnosing nonmalignant, relatively common abnormalities and comparing CT imaging to other diagnostic technologies.

PET with FDG has recently emerged as a practical and useful imaging modality in the preoperative staging of patients with lung cancer. However, whereas CT is most frequently used to provide additional anatomical and morphological information about lesions, the FDG PET imaging provides physiological and metabolic information that characterizes lesions that are indeterminate by CT. FDG PET imaging takes advantage of the increased accumulation of FDG in transformed cells and is sensitive (~95%) for the detection of cancer in patients who have indeterminate lesions on CT (138). The specificity (~85%) of PET imaging is slightly less than its sensitivity because some inflammatory processes avidly accumulate FDG. The high negative predictive value of PET suggests that lesions considered negative on the study are benign, biopsy is not needed, and radiographic follow-up is recommended. Several studies have documented the increased accuracy of PET compared with CT in the evaluation of the hilar and mediastinal lymph node status

in patients with lung cancer (138). However, the PET resolution is sufficient only for nodules ≥ 6 cm and will not be helpful in detecting the very small nodules. Compared with low-dose spiral CT, the FDG PET scan is more expensive and time consuming. The role of PET scan in early diagnosis of lung cancer in an asymptomatic high-risk population is not yet evaluated. However, future studies have to include PET evaluation to define its role in a population screening setting.

Conclusion

Recent advances in molecular biology and pathology have led to a better understanding and documentation of morphological changes in the bronchial epithelium before development of clinical evident lung carcinomas. Combined with technical developments in radiological and bronchoscopic techniques, these procedures offer great promise in diagnosing lung cancer far in advance of clinical presentation. Any of these individual procedures could be incorporated into the routine management of individuals at risk for developing primary or secondary lung cancer, and for several of these methods, clinical studies are under way. Preliminary reported data are very promising for the early detection of lung cancer. Future studies must incorporate the different methods in a multidisciplinary scientific setting to evaluate the role of the individual method in the overall management for individuals at high risk for developing lung cancer. Several of these tests might diagnose the disease at the stage of clonal expansion before invasive carcinoma has developed. A management and intervention strategy appropriate to that stage of disease have to be developed. Preliminary studies of chemoprevention agents are reported, and new agents based on other biological mechanisms are under development and ready for clinical trials. It is now time to plan clinical trials that evaluate both diagnostic and therapeutic approaches to access their impact on the incidence of clinical lung cancer.

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References

- Greenlee, R. T., Murray, T., Bolden, S., and Wingo, P. A. Cancer Statistics, 2000. *CA Cancer J. Clin.*, 50: 7–30, 2000.
- Mountain, C. T. Revision in the international system for staging of lung cancer. *Chest*, 111: 1710–1717, 1997.
- Idée, D. C. Chemotherapy of lung cancer. *N. Engl. J. Med.*, 327: 1434–1441, 1992.
- McLined, M. R., Flehinger, B. J., Zaman, M. B., Heelan, R. T., Patchick, W. A., and Martini, N. Screening for lung cancer: results of the Memorial Sloan-Kettering study in New York. *Chest*, 86: 44–53, 1984.
- Fontana, R. S., Sanderson, D. R., Woolner, L. B., Taylor, W. F., Miller, W. E., and Muhm, J. R. Lung Cancer Screening: The Mayo program. *J. Occup. Med.*, 28: 746–750, 1986.
- Tockman, M. S. Survival and mortality from lung cancer in a screened population: The Johns Hopkins Study. *Chest*, 89: 3245–3255, 1986.

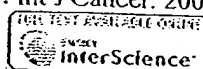
7. Kubik, A., Parkin, D. M., Khlát, M., Erban, J., Polak, J., and Adfamec, M. Lack of benefit from semi-annual screening for cancer of the lung: follow-up report of a randomized controlled trial on a population of high-risk males in Czechoslovakia. *Int. J. Cancer*, 45: 26-33, 1990.
8. Fontana, R. S., Sanderson, D. R., Woolner, L. B., Taylor, W. F., Miller, W. E., Muhm, J. R., Bernatz, P. E., Payne, W. S., Pairolero, P. C., and Bergstralh, E. J. Screening of lung cancer. A critique of the Mayo Lung Project. *Cancer (Phila.)*, 67: 1155-1164, 1991.
9. Strauss, G. M., Gleason, R. E., and Sugarbaker, D. J. Screening for lung cancer. Another look: a different view. *Chest*, 111: 754-768, 1997.
10. Hirsch, F. R., Brambilla, E., Gray, N., Gritz, E., Kelloff, G. J., Linnoila, I., Pastorino, U., and Mulshine, J. L. Prevention and early detection of lung cancer—clinical aspects. *Lung Cancer (Limerick)*, 17: 163-174, 1997.
11. Hong, W. K. Chemoprevention of Lung Cancer. *Oncology (Basel)*, 13 (Suppl. 5): 135-141, 1999.
12. Mulshine, J. L., and Henschke, C. I. Prospects for lung-cancer screening. *Lancet*, 355: 592-593, 2000.
13. Travis, W. D., Colby, T. V., Corrin, B., Shimosato, Y., and Brambilla, E. Histological typing of tumours of lung and pleura. In: L. H. Sobin (ed.), *World Health Organization International Classification of Tumours*, Ed. 3. New York: Springer-Verlag, 1999.
14. Franklin, W. A. Pathology of lung cancer. *J. Thorac. Imaging*, 15: 3-12, 2000.
15. Colby, T. V. Precursor lesions to pulmonary neoplasia. In: C. Brambilla and E. Brambilla (eds.), *Lung Tumors. Fundamental Biology and Clinical Management*, pp. 61-87. New York: Marcel Dekker Inc., 1999.
16. Peters, E. J., Morice, R., Benner, S. E., Lippman, S., Lukeman, J., Lee, J. S., Ro, J. L., and Hong, W. K. Squamous metaplasia of the bronchial mucosa and its relationship to smoking. *Chest*, 103: 1429-1432, 1993.
17. Auerbach, O., Gere, B., Forman, J. B., Petrick, T. G., Smolin, H. J., Muchsam, G. E., Kassouny, D. Y., and Stout, A. P. Changes in bronchial epithelium in relation to smoking and cancer of the lung. *N. Engl. J. Med.*, 256: 97-104, 1957.
18. Wistuba, I. J., Behrens, C., Milchgrub, S., Bryant, D., Hung, J., Minna, J. D., and Gazdar, A. F. Sequential molecular abnormalities are involved in the multistage development of squamous cell lung carcinoma. *Oncogene*, 18: 643-650, 1999.
19. Lam, S., LeRiche, J. C., Zheng, Y., Coldman, A., MacAulay, C., Hawk, E., Kelloff, G., and Gazdar, A. F. Sex-related differences in bronchial epithelial changes associated with tobacco smoking. *J. Natl. Cancer Inst.*, 91: 691-696, 1999.
20. Saccomanno, G., Archer, V. E., Auerbach, O., Saunders, R. P., and Brennan, L. M. Development of carcinoma of the lung as reflected in exfoliated cells. *Cancer (Phila.)*, 32: 256-270, 1974.
21. Frost, J. K., Ball, W. C., Jr., Levin, M. L., Tockman, M. S., Erozan, Y. S., Gupta, K., Eggleston, J. C., Pressman, N. J., Donithan, M. P., and Kimball, A. W. Sputum cytopathology: use and potential in monitoring the workplace environment by screening for biological effects of exposure. *J. Occup. Med.*, 28: 692-703, 1986.
22. Venman, B. J. W., van Boxem, T. J. M., Smit, E. F., Postmus, P. E., and Sutedja, T. G. Outcome of bronchial carcinoma *in situ*. *Chest*, 117: 1572-1576, 2000.
23. Slebos, R. J., Baas, I. O., Clement, M. J., Offerhaus, G. J., Askin, F. B., Hruban, R. H., and Westra, W. H. p53 alterations in atypical alveolar hyperplasia of the human lung. *Hum. Pathol.*, 29: 801-808, 1998.
24. Kitamura, H., Kameda, Y., Ito, T., and Hayashi, H. Atypical adenomatous hyperplasia of the lung. Implications for the pathogenesis of peripheral lung adenocarcinoma. *Am. J. Clin. Pathol.*, 111: 610-622, 1999.
25. Diedrich, S., Semik, M., Lentschig, M. G., Wintet, F., Scheld, H. H., Ross, N., and Bongartz, G. Helical CT of pulmonary nodules in patients with extrathoracic malignancy: CT-surgical correlation. *Am. J. Roentgenol.*, 172: 353-360, 1999.
26. Keith, R. L., Miller, Y. E., Gemmill, R. M., Drabkin, H. A., Dempsey, E. C., Kennedy, T. C., Prindiville, S., and Franklin, W. A. Angiogenic squamous dysplasia in bronchi of individuals at high risk for lung cancer. *Clin. Cancer Res.*, 6: 1616-1625, 2000.
27. Muller, K. M., and Muller, G. The ultrastructure of preneoplastic changes in bronchial mucosa. *Curr. Top. Pathol.*, 73: 233-263, 1983.
28. Hirsch, F. R., Gazdar, A. F., Gabrielson, E., Lam, S., and Franklin, W. A. Histopathologic evaluation of premalignant and early malignant bronchial lesions: an interactive program based on internet digital images to improve WHO criteria for early diagnosis of lung cancer and for monitoring chemoprevention studies—a SPOR collaborative project. *Lung Cancer (Limerick)*, 29 (Suppl. 2): 209, 2000.
29. Lengauer, C., Kinzler, K. W., and Vogelstein, B. Genetic instabilities in human cancers. *Nature (Lond.)*, 396: 643-649, 1998.
30. Fong, K. M., Sekido, Y., and Minna, J. D. Molecular pathogenesis of lung cancer. *J. Thorac. Cardiovasc. Surg.*, 118: 1136-1152, 1999.
31. Hirano, T., Franzen, B., Kato, H., Ebihara, Y., and Auer, G. Genesis of squamous cell lung carcinoma: sequential changes of proliferation, DNA ploidy, and p53 expression. *Am. J. Pathol.*, 144: 296-302, 1994.
32. Betticher, D. C., Heighway, J., Thatcher, N., and Hasleton, P. S. Abnormal expression of CCND1 and RB1 in resection margin epithelia of lung cancer patients. *Br. J. Cancer*, 75: 1761-1768, 1997.
33. Satoh, Y., Ishikawa, Y., Nakagawa, K., Hirano, T., and Tsuchiya, E. A follow-up study of progression from dysplasia to squamous cell carcinoma with immunohistochemical examination of p53 protein overexpression in the bronchi of ex-chuomate workers. *Br. J. Cancer*, 75: 678-683, 1997.
34. Li, Z. H., Zheng, J., Weiss, L. M., and Shibata D. c-k *ras*, and p53 mutations occur very early in adenocarcinoma of the lung. *Am. J. Pathol.*, 144: 303-309, 1994.
35. Brambilla, E., Gazzeri, S., Lantuejoul, S., Coll, J. L., Moro, D., Negoescu, A., and Brambilla, C. p53 mutant immunophenotype and deregulation of p53 transcription pathway (Bcl2, Bax, and Waf1) in precursor bronchial lesions of lung cancer. *Clin. Cancer Res.*, 4: 1609-1618, 1998.
36. Hung, J., Kishimoto, Y., Sugio, K., Virmani, A., McIntire, D. D., Minna, J. D., and Gazdar, A. F. Allele-specific chromosome 3p deletions occur at an early stage in the pathogenesis of lung carcinoma. *J. Am. Med. Assoc.*, 273: 558-563, 1995.
37. Kishimoto, Y., Sugio, K., Hung, J. Y., Virmani, A. K., McIntire, D. D., Minna, J. D., and Gazdar, A. F. Allele-specific loss in chromosome 9p loci in preneoplastic lesions accompanying non-small cell lung cancers. *J. Natl. Cancer Inst.*, 87: 1224-1229, 1995.
38. Sugio, K., Kishimoto, Y., Virmani, A. K., Hung, J. Y., and Gazdar, A. F. K-ras mutations are a relatively late event in the pathogenesis of lung carcinomas. *Cancer Res.*, 54: 5811-5815, 1994.
39. Wistuba, I. J., Behrens, C., Virmani, A. K., Milchgrub, S., Syed, S., Lam, S., Mackay, B., Minna, J. D., and Gazdar, A. F. Allelic losses at chromosome 8p21-23 are early and frequent events in the pathogenesis of lung cancer. *Cancer Res.*, 59: 1973-1979, 1999.
40. Varella-Garcia, M., Gemmill, R. M., Rabenhorst, S. H., Lott, A., Drabkin, H. A., Archer, P. A., and Franklin, W. A. Chromosomal duplication accompanies allelic loss in non-small cell lung carcinoma. *Cancer Res.*, 58: 4701-4707, 1998.
41. Westra, W. H., Baas, I. O., Hruban, R. H., Askin, F. B., Wilson, K., Offerhaus, G. J., Slebos, R. J. K-ras oncogene activation in atypical alveolar hyperplasias of the human lung. *Cancer Res.*, 56: 2224-2228, 1996.
42. Wistuba, I. J., Lam, S., Behrens, C., Virmani, A. K., Fong, K. M., LeRiche, J., Samet, J. M., Srivastava, S., Minna, J. D., and Gazdar, A. F. Molecular damage in the bronchial epithelium of current and former smokers. *J. Natl. Cancer Inst.*, 89: 1366-1377, 1997.
43. Mao, L., Lee, J. S., Kurie, J. M., Fan, Y. H., Lippman, S. M., Lee, J. J., Ro, J. Y., Broxson, A., Yu, R., Morice, R. C., Kemp, B. L., Khuri, F. R., Walsh, G. L., Hittelman, W. N., and Hong, W. K. Clonal genetic

- alterations in the lungs of current and former smokers. *J. Natl. Cancer Inst.*, 89: 857-862, 1997.
44. Smith, A. L., Hung, J., Walker, L., Rogers, T. E., Vuitch, F., Lee, E., and Gazdar, A. F. Extensive areas of aneuploidy are present in the respiratory epithelium of lung cancer patients. *Br. J. Cancer*, 73: 203-209, 1996.
 45. Franklin, W. A., Gazdar, A. F., Haney, J., Wisnuba, I. I., La Rosa, F. G., Kennedy, T., Ritchey, D. M., and Miller, Y. E. Widely dispersed p53 mutation in respiratory epithelium: a novel mechanism for field carcinogenesis. *J. Clin. Invest.*, 100: 2133-2137, 1997.
 46. Johnston, W. W., and Elson, C. E. Respiratory tract. In: M. Bibbo (ed.), *Comprehensive Cytopathology*, pp. 325-401. Philadelphia: Saunders, 1997.
 47. Travis, W. D., Travis, L. B., and Devesa, S. S. Lung cancer. *Cancer (Phila.)*, 75 (Suppl. 1): 191-202, 1995.
 48. Wynder, E. L., and Muscat, J. E. The changing epidemiology of smoking and lung cancer histology. *Environ. Health Perspect.*, 103 (Suppl. 8): 143-148, 1995.
 49. Thun, M. J., Lally, C. A., Flannery, J. T., Calle, E. E., Flanders, W. D., and Heath, C. W., Jr. Cigarette smoking and changes in the histopathology of lung cancer. *J. Natl. Cancer Inst.*, 89: 1580-1586, 1997.
 50. Lam, S., and Shibuya, H. Early diagnosis of lung cancer. *Clin. Chest Med.*, 20: 53-61, 1999.
 51. Kennedy, T. C., Proudfoot, S., Piantadosi, S., Wu, L., Saccomanno, G., Petty, T. L., and Tockman, M. S. Efficacy of two sputum collection techniques in patients with air flow obstruction. *Acta Cytol.*, 43: 630-636, 1999.
 52. Browman, G. P., Arnold, A., Levine, M. N., and D'Souza, T. Use of screening phase data to evaluate observer variation of sputum cytodiagnosis as an outcome measure in a chemoprevention trial. *Cancer Res.*, 50: 1216-1219, 1990.
 53. Cantaboni, A., Pezzotta, M. G., Sironi, M., and Porcellai, M. Quality assurance in pathology: cytologic and histologic correlation. *Acta Cytol.*, 36: 717-721, 1992.
 54. Holiday, D. B., McLary, J. W., Farley, M. L., Mabry, L. C., Cozens, D., Roby, T., Waldron, E., Underwood, R. D., Anderson, E., and Culbreth, W. Sputum cytology within and across laboratories. A reliability study. *Acta Cytol.*, 39: 195-206, 1995.
 55. Risse, E. K., Vooijs, G. P., and van't Hoff, M. A. Relationship between the cellular composition of sputum and the cytologic diagnosis of lung cancer. *Acta Cytol.*, 31: 170-176, 1987.
 56. MacDougall, B., and Weinerman, B. The value of sputum cytology. *J. Gen. Intern. Med.*, 7: 11-12, 1992.
 57. Tockman, M. S., Erozan, Y. S., Gupta, P., Piantadosi, S., Mulshine, J. L., and Ruckdeschel, J. C. The early detection of second primary lung cancers by sputum immunostaining. *Chest*, 106 (Suppl.): 385S-390S, 1994.
 58. Tockman, M. S., Gupta, P. K., Myers, J. D., Frost, J. K., Baylin, S. B., Gold, E. B., Chase, A. M., Wilkinson, P. H., and Mulshine, J. L. Sensitive and specific monoclonal antibody recognition of human lung cancer antigen on preserved sputum cells: a new approach to early lung cancer detection. *J. Clin. Oncol.*, 6: 1685-1693, 1988.
 59. Zhou, J., Mulshine, J. L., Unsworth, E. J., Scott, F. M., Avis, I. M., Vos, M. D., and Treston, A. M. Purification and characterization of a protein that permits early detection of lung cancer. *J. Biol. Chem.*, 271: 10760-10766, 1996.
 60. Qiao Y-L., Tockman, M. S., Li, L., Erozan, Y. S., Yao, S. X., Barrett, M. J., Zhou, W. H., Giffen, C. A., Luo, X. C., and Taylor, P. R. A case-cohort study of an early biomarker of lung cancer in a screening cohort of Yunnan tin miners in China. *Cancer Epidemiol Biomark. Prev.*, 6: 893-900, 1997.
 61. Grover, F. L., and Piantadosi, S. Recurrence and survival following resection of bronchioloalveolar carcinoma of the lung: the Lung Cancer Study Group experience. *Ann. Surg.*, 209: 779-790, 1989.
 62. Tockman, M. S. Advances in sputum analysis for screening and early detection of lung cancer. *Cancer Control*, 7: 19-24, 2000.
 63. Sueoka, E., Goto, Y., Sueoka, N., Kai, Y., Kozu, T., and Fujiki, H. Heterogeneous nuclear ribonucleoprotein B1 as a new marker of early detection for human lung cancers. *Cancer Res.*, 59: 1404-1407, 1999.
 64. Mulshine, J. L. Reducing lung cancer risk. Early detection. *Chest*, 116: 493S-496S, 1999.
 65. Mao, L., Hruban, R. H., Boyle, J. O., Tockman, M., and Sidransky, D. Detection of oncogene mutations in sputum precedes diagnosis of lung cancer. *Cancer Res.*, 54: 1634-1637, 1994.
 66. Ruppert, J. M., Tokino, K., and Sidransky, D. Evidence for two bladder cancer suppressor loci on chromosome 9. *Cancer Res.*, 53: 5093-5095, 1993.
 67. Nawroz, H., van der Riet, P., Hruban, R. H., Koch, W., Ruppert, J. M., and Sidransky, D. Allelotype of head and neck squamous cell carcinoma. *Cancer Res.*, 54: 1152-1155, 1994.
 68. Miozzo, M., Sozzi, G., Musso, K., Pilotti, S., Incabone, M., Pastorino, U., and Pierotti, M. A. Microsatellite alterations in bronchial and sputum specimens of lung cancer patients. *Cancer Res.*, 56: 2285-2288, 1996.
 69. Shapiro, G. I., Park, J. E., Edwards, C. D., Mao, L., Merlo, A., Sidransky, D., Ewen, M. E., and Rollins, B. J. Multiple mechanisms of p16^{INK4a} inactivation in non-small cell lung cancer cell lines. *Cancer Res.*, 55: 6200-6209, 1995.
 70. Hamada, K., Kohno, T., Kawanishi, M., Ohwada, S., and Yokota, J. Association of CDKN2A(p16)/CDKN2B(p15) alterations and homozygous chromosome arm 9p deletions in human lung carcinoma. *Genes Chromosomes Cancer*, 22: 232-240, 1998.
 71. Belinsky, S. A., Nikula, K. J., Palmisano, W. A., Michels, R., Saccomanno, G., Gabrielson, E., Baylin, S. B., and Herman, J. G. Aberrant methylation of p16^{INK4a} is an early event in lung cancer and a potential biomarker for early diagnosis. *Proc. Natl. Acad. Sci. USA*, 95: 11891-11896, 1998.
 72. Esteller, M., Sanchez-Cespedes, M., Rosell, R., Sidransky, D., Baylin, S. B., and Herman, J. G. Detection of aberrant promoter hypermethylation of tumor suppressor genes in serum DNA from non-small cell lung cancer patients. *Cancer Res.*, 59: 67-70, 1999.
 73. Nieburgs, H. E. Recent progress in the interpretation of malignancy associated changes (MAC). *Acta Cytol.*, 32: 445-453, 1988.
 74. Payne, P. W., Sebo, T. J., Doudkine, A., Garner, D., MacAulay, C., Lam, S., LeRiche, J. C., and Palcic, B. Sputum screening by quantitative microscopy: a reexamination of a portion of the National Cancer Institute Cooperative Early Lung Cancer Study. *Mayo Clin. Proc.*, 72: 697-704, 1997.
 75. Lam, S., Palcic, B., Garner, D., Beveridge, J., MacAulay, C., LeRiche, J., and Coldman, A. Lung Cancer Control Strategy in the New Millennium. *Lung Cancer*, 29 (Suppl. 2): 145, 2000.
 76. Ahrendt, S. A., Chow, J. T., Xu, L. H., Yang, S. C., Eisenberger, C. F., Esteller, M., Herman, J. G., Wu, L., Decker, P. A., Jen, J., and Sidransky, D. Molecular detection of tumor cells in bronchoalveolar lavage fluid from patients with early stage lung cancer. *J. Natl. Cancer Inst.*, 91: 332-339, 1999.
 77. Gazdar, A. F., and Minna, J. D. Molecular detection of early lung cancer. *J. Natl. Cancer Inst.*, 91: 299-301, 1999.
 78. Field, J. K., Liloglou, T., Xinarianos, G., Prime, W., Fielding, P., Walshaw, M. J., and Turnbull, L. Genetic alterations in bronchial lavage as a potential marker for individuals with a high risk of developing lung cancer. *Cancer Res.*, 59: 2690-2695, 1999.
 79. Liloglou, T., Maloney, P., Xinarianos, G., Fear, S., and Field, J. K. Sensitivity and limitations of high throughput fluorescent microsatellite analysis for the detection of allelic imbalance. Application in lung tumors. *Int. J. Oncol.*, 16: 5-14, 2000.
 80. Fielding, P., Turnbull, L., Prime, W., Walshaw, M., and Field, J. K. Heterogeneous nuclear ribonucleoprotein A2/B1 up-regulation in bronchial lavage specimens: a clinical marker of early lung cancer detection. *Clin. Cancer Res.*, 5: 404S-405S, 1999.
 81. Pantel, K., Schlimok, G., Angstwurm, M., Weckermann, D., Schmaus, W., Gath, H., Passlick, B., Izbicke, J. R., and Rietman, G. Methodological analysis of immunocytochemical screening for dissem-

- inated epithelial tumor cells in bone marrow. *J. Hematother.*, 3: 165-173, 1994.
82. Lambrechts, A. C., van't Veer, L. J., and Rodenhuis, S. The detection of minimal numbers of contaminating epithelial tumor cells in blood or bone marrow: use, limitations and future of RNA-based methods. *Ann. Oncol.*, 9: 1269-1276, 1998.
83. Steinman, C. R. Free DNA in serum and plasma from normal adults. *J. Clin. Investig.*, 66: 1391-1399, 1980.
84. Raptis, L., and Menard, H. A. Quantitation and characterization of plasma DNA in normals and patients with lupus erythematosus. *J. Clin. Investig.*, 66: 1391-1399, 1980.
85. Leon, S. A., Shapiro, B., Sklaroff, D. M., and Yaros, M. J. Free DNA in the serum of cancer patients and the effect of therapy. *Cancer Res.*, 37: 646-650, 1977.
86. Stroun, M., Anker, P., Maurice, P., Lyautey, J., Lederrey, C., and Beljanski, M. Neoplastic characteristics of the DNA found in the plasma of cancer patients. *Oncology*, 46: 318-322, 1989.
87. Shapiro, B., Chakrabaty, M., Cohn, E., and Leon, S. A. Determination of circulating DNA levels in patients with benign or malignant gastrointestinal disease. *Cancer (Phila.)*, 51: 2116-2120, 1983.
88. Chen, X. Q., Stroun, M., Magnenat, J.-L., Nicod, L. P., Kurt, A. M., Lyautey, J., Lederrey, C., and Anker, P. Microsatellite alterations in plasma DNA of small cell lung cancer patients. *Nat. Med.*, 2: 1033-1037, 1996.
89. Anker, P., Lefort, F., Vasioukhin, V., Lyautey, J., Lederrey, C., Chen, X. Q., Stroun, M., Mulcahy, H. E., and Farthing, M. J. K-ras mutations are found in DNA extracted from the plasma of colorectal cancer patients. *Gastroenterology*, 112: 1114-1129, 1997.
90. Kopreski, M. S., Benko, F. A., Kwee, C., Leitzel, K. E., Eskander, E., Lipton, A., and Gocke, C. D. Detection of mutant K-ras DNA in plasma or serum of patients with colorectal cancer. *Br. J. Cancer*, 76: 1293-1299, 1997.
91. Hibi, K., Robinson, R., Wu, L., Hamilton, S. R., Sidransky, D., and Jen, J. Molecular detection of genetic alterations in the serum of colorectal cancer patients. *Cancer Res.*, 58: 1405-1407, 1998.
92. Sorenson, G. D., Pribish, D. M., Valone, F. H., Memoli, V. A., Bizik, D. J., and Yao, S. L. Soluble normal and mutated DNA sequences from single copy genes in human blood. *Cancer Epidemiol. Biomark. Prev.*, 3: 67-71, 1994.
93. Mulcahy, H. E., Lyautey, J., Lederrey, C., Chen, X., Anker, P., Alstead, E. M., Ballinger, A., Farthing, M. J., and Stroun, M. A prospective study of K-ras mutations in the plasma of pancreatic cancer patients. *Clin. Cancer Res.*, 4: 271-275, 1998.
94. Vasioukhin, V., Anker, P., Maurice, P., Lyautey, J., Lederrey, C., and Stroun, M. Point mutations of the N-ras in the blood plasma DNA of patients with myelodysplastic syndrome or acute myelogenous leukaemia. *Br. J. Haematol.*, 86: 774-779, 1994.
95. Woolner, L. B. Pathology of cancer detected cytologically. In: *Atlas of Early Lung Cancer*. National Cancer Institute, NIH, United States Department of Health and Human Services, pp. 107-203. Tokyo: Igaku-Shoin, 1983.
96. Kato, H., and Cortese, D. A. Early detection of lung cancer by means of hematoporphyrin derivative fluorescence and laser photoradiation. *Clin. Chest Med.*, 6: 237-253, 1985.
97. Profio, A. E., Dohren, D. R., and King, E. G. L. Laser fluorescence bronchoscopy for localization of occult lung tumors. *Med. Phys. (NY)*, 6: 532-535, 1979.
98. Kinsey, J. H., and Cortese, D. A. Endoscopic system for simultaneous visual examination and electronic detection of fluorescence. *Rev. Sci. Instrum.*, 51: 1403-1406, 1980.
99. Profio, A. E., Dohren, D. R., and Sarnak, J. Fluorometer for endoscopic diagnosis of tumors. *Med. Phys. (NY)*, 11: 516-520, 1984.
100. Montan, S., Svanberg, K., and Svanberg, S. Multicolor imaging and contrast enhancement in cancer-tumor localization using laser-induced fluorescence in hematoporphyrin-derivative-bearing tissue. *Optics Lett.*, 10: 56-58, 1985.
101. Kato, H., Imaizumi, T., Aizawa, K., Iwabuchi, H., Yamamoto, H., Ikeda, N., Tsuchida, T., Tamachi, Y., Ito, T., and Hayata, Y. Photodynamic diagnosis in respiratory tract malignancy, using an excimer dye laser system. *J. Photochem. Photobiol. Biol.*, 6: 189-196, 1990.
102. Kato, H., and Ikeda, N. The role of fluorescence diagnosis in the early detection of high-risk bronchial lesions. *J. Bronchol.*, 5: 273-274, 1998.
103. Palcic, B., Lam, S., Hung, J., and MacAulay, C. Detection and localization of early lung cancer by imaging techniques. *Chest*, 99: 742-743, 1991.
104. Hung, J., Lam, S., LeRiche, J. C., and Palcic, B. Autofluorescence of normal and malignant bronchial tissue. *Lasers Surg. Med.*, 11: 99-105, 1991.
105. Lam, S., Kennedy, T., Unger, M., Miller, Y. E., Gelinhout, D., Rusch, V., Gipe, B., Howard, D., LeRiche, J. C., Goldman, A., and Gazdar, A. F. Localization of bronchial intraepithelial neoplastic lesions by fluorescence bronchoscopy. *Chest*, 113: 696-702, 1998.
106. Kurie, J. M., Lee, J. S., Morice, R. C., Walsh, G. L., Khuri, F. R., Broxson, A., Ro JY, Franklin, W. A., Yu, R., and Hong, W. K. Autofluorescence bronchoscopy in the detection of squamous metaplasia and dysplasia in current and former smokers. *J. Natl. Cancer Inst.*, 90: 991-995, 1998.
107. Venmans, B. J., van der Linden, J. C., van Boxem, A. J., Postmus, P. E., Smit, E. F., and Sutedja, G. Early detection of pre-invasive lesions in high risk patients. A comparison of conventional fiberoptic and fluorescence bronchoscopy. *J. Bronchol.*, 5: 280-283, 1998.
108. Vermylen, P., Pierard, P., Roufosse, C., Bosschaerts, T., Verhest, A., Sculier, J.-P., and Ninane, V. Detection of bronchial preneoplastic lesions and early lung cancer with fluorescence bronchoscopy: a study about its ambulatory feasibility under local anaesthesia. *Lung Cancer (Limerick)*, 25: 161-168, 1999.
109. Kennedy, T., Hirsch, F. R., Miller, Y., Prindiville, S., Bunn, P. A., Jr., and Franklin, W. A randomized study of fluorescence bronchoscopy versus white-light bronchoscopy for early detection of lung cancer in high risk patients. *Lung Cancer (Limerick)*, 29 (Suppl. 2): 244, 2000.
110. Gazdar, A. F., and Minna, J. D. Angiogenesis and the multistage development of lung cancers. *Clin. Cancer Res.*, 6: 1611-1612, 2000.
111. Henschke, C. I., Miettinen, O. S., Yankelevitz, D. F., Libby, D. M., and Smith, J. P. Radiographic screening for cancer: proposed paradigm for requisite research. *Clin. Imaging*, 18: 6-20, 1994.
112. Miettinen, O. S. Screening for lung cancer. *Radiol. Clin. N. Am.*, 38: 479-486, 2000.
113. Marcus, P. M., Bergstralh, E. J., Fagerstrom, M., Williams, D. E., Fontana, R., Taylor, W. F., and Prorok, P. C. Lung cancer mortality in the Mayo Lung Project: impact of extended follow-up. *J. Natl. Cancer Inst.*, 92: 1308-1316, 2000.
114. Black, W. C. Overdiagnosis. An underrecognized cause of confusion and harm in cancer screening. *J. Natl. Cancer Inst.*, 92: 1280-1282, 2000.
115. The Health, and Welfare Statistics Foundation. The current issue on the screening project for elderly people by the Ministry of Health and Welfare Japan. *J. Health Welfare*, 42: 37, 1995.
116. Sobue, T., Suzuki, T., and Naruke, T. The Japanese Lung Cancer Screening Research Group: a case-control study for evaluating lung cancer screening in Japan. *Int. J. Cancer*, 50: 230-237, 1992.
117. Okamoto, N., Suzuki, T., Hasegawa, H., Gotoh, T., Hagiwara, S., Sekimoto, M., and Kaneko, M. Evaluation of a clinic-based screening program for lung cancer with a case-control design in Kanagawa, Japan. *Lung Cancer (Limerick)*, 25: 77-85, 1999.
118. Muhl, J. R., Miller, W. F., Fontana, R. S., Sanderson, D. R., and Uhlenhuth, M. A. Lung cancer detected during a screening program using four-month chest radiographs. *Radiology*, 148: 609-615, 1983.
119. Hayabuchi, N., Russell, W. J., and Murakami, J. Problems in radiographic detection and diagnosis of lung cancer. *Acta Radiol. (CPII)*, 30: 163-167, 1989.
120. Woodring, J. H. Pitfalls in the radiologic diagnosis of lung cancer. *Am. J. Roentgenol.*, 154: 1165-1175, 1990.

121. Greene, R. E. Missed lung nodules: lost opportunities for cancer care. *Radiology*, 182: 8-9, 1992.
122. Austin, J. H. M., Romney, B. M., Goldsmith, L. S. Missed bronchogenic carcinoma: radiographic findings in 27 patients with a potentially resectable lesions evident in retrospect. *Radiology*, 182: 115-122, 1992.
123. Schaner, E. G., Chang, A. E., Doppan, J. L., Conkle, D. M., Flye, M. W., and Rosenberg, S. A. Comparison of computed and conventional whole lung tomography in detecting pulmonary nodules. *Am. J. Roentgenol.*, 131: 51-54, 1978.
124. Webb, W. R. Radiologic evaluation of solitary pulmonary nodule. *Am. J. Roentgenol.*, 154: 701-708, 1990.
125. Kalender, W. A., Seissler, W., Klotz, E., and Vock, P. Spiral volumetric CT with single-breath-hold technique, continuous transport, and continuous scanner rotation. *Radiology*, 176: 181-183, 1990.
126. Costello, P., Anderson, W. A., and Blume, D. Pulmonary nodule: evaluation with spiral volumetric CT. *Radiology*, 179: 875-876, 1991.
127. Remy-Jardin, M., Giraud, F., and Marquette, C. H. Pulmonary nodules: detection with thick section spiral CT versus conventional CT. *Radiology*, 187: 513-520, 1993.
128. Davis, S. CT evaluation for pulmonary metastases in patients with extrathoracic malignancy. *Radiology*, 180: 1-12, 1991.
129. Kaneko, M., Eguchi, K., Ohmatsu, H., Kakinuma, R., Naruke, T., Suemasu, K., and Moriyama, N. Peripheral lung cancer: screening and detection with low-dose spiral CT versus radiography. *Radiology*, 201: 798-802, 1996.
130. Sone, S., Takashima, S., Li, F., Yang, Z., Honda, T., Maruyama, Y., Hasegawa, M., Yamada, T., Kubo, K., Hanamura, K., and Asakura, K. Mass screening for lung cancer with mobile spiral computed tomography scanner. *Lancet*, 351: 1242-1245, 1998.
131. Henschke, C. I., McCauley, D. I., Yankelevitz, D. F., Naidich, D. P., McGuinness, G., Miettinen, O. S., Libby, D. M., Pasmantier, M. W., Koizumi, J., Altorki, N. K., and Smith, J. P. Early Lung Cancer Action Project: overall design and findings from baseline screening. *Lancet*, 354: 99-105, 1999.
132. Kakinuma, R., Ohmatsu, H., Kaneko, M., Eguchi, K., Naruke, T., Nagai, K., Nishiwaki, Y., Suzuki, A., and Moriyama, N. Detection failures in spiral CT screening for lung cancer: analysis of CT findings. *Radiology*, 212: 61-66, 1999.
133. Paranjpe, D. V., and Bergin, C. J. Spiral CT of the lungs: optimal technique and resolution compared with conventional CT. *Am. J. Roentgenol.*, 167: 561-567, 1994.
134. Seltzer, S. E., Judy, P. F., Adams, D. F., Jacobson, F. L., Stark, P., Kikinis, R., Swenson, R. G., Hooton, S., Head, B., and Feldman, U. Spiral CT of the chest: comparison of cine and film-based viewing. *Radiology*, 197: 73-78, 1995.
135. Croisille, P., Souto, M., Cova, M., Wood, S., Afework, Y., Kuhlman, J. E., and Zerhouni, E. A. Pulmonary nodules: improved detection with vascular segmentation and extraction with spiral CT. *Radiology*, 197: 397-401, 1995.
136. Kanazawa, K., Kawata, Y., Niki, N., Satoh, H., Ohmatsu, H., Kakinuma, R., and Kaneko, M. Computer-aided diagnosis for pulmonary nodules based on helical CT images. *Comput. Med. Imaging Graph.*, 22: 157-167, 1998.
137. Buckshar, N., Agnello, K., Yankelevitz, D. F., Mancuso, C., and Henschke, C. I. Smoking habits and overall satisfaction after early lung cancer screening using low-dose CT. *ALA/ATS International Conference*, 1999.
138. Coleman, R. E. PET in lung cancer. *J. Nucl. Med.*, 40: 814-820, 1999.
139. Maitra, A., Wisstuba, I. J., Virmani, A. K., Sakaguchi, M., Park, I., Stucky, A., Milchgrub, S., Gibbons, D., Minna, J. D., and Gazdar, A. F. Enrichment of epithelial cells for molecular studies. *Nat. Med.*, 5: 459-463, 1999.
140. Kopelovich, I., Henson, D. F., Gazdar, A. F., Dubb, B., Srivastava, S., Kelloff, G. J., and Greenwald, P. Surrogate anatomic/functional sites for evaluating cancer risk: an extension of the field effect. *Clin. Cancer Res.*, 5: 3899-3905, 1999.
141. Crowell, R. E., Gilliland, F. D., Temes, R. T., Harms, H. J., Neft, R. E., Heaphy, E., Auckley, D. H., Crooks, L. A., Jordan, S. W., Samet, J. M., Lechner, J. F., and Belinsky, S. A. Detection of trisomy 7 in nonmalignant bronchial epithelium from lung cancer patients and individuals at risk for lung cancer. *Cancer Epidemiol. Biomark. Prev.*, 5: 631-637, 1996.
142. Somers, V. A., van Henten, A. M., ten Velde, G. P., Arends, J. W., and Thunnissen, F. B. Additional value of K-ras point mutations in bronchial wash fluids for diagnosis of peripheral lung tumours. *Eur. Respir. J.*, 13: 1120-1124, 1999.
143. Yahata, N., Ohyashiki, K., Ohyashiki, J. H., Iwama, H., Hayashi, S., Ando, K., Hirano, T., Tsuchida, T., Katô, H., Shay, J. W., and Toyama, K. Telomerase activity in lung cancer cells obtained from bronchial washings. *J. Natl. Cancer Inst.*, 90: 684-690, 1998.
144. Wistuba, I. J., and Gazdar, A. F. Molecular abnormalities in the sequential development of lung carcinoma. In: S. Srivastava, D. E. Henson, and A. F. Gazdar (eds.), *Molecular Pathology of Early Cancer*, pp. 265-276. Amsterdam: IOS Press, 1999.
145. Yashima, K., Litzky, L. A., Kaiser, L., Rogers, T., Lam, S., Wistuba, I. J., Milchgrub, S., Srivastava, S., Piatyszek, M. A., Shay, J. W., and Gazdar, A. F. Telomerase expression in respiratory epithelium during the multistage pathogenesis of lung carcinomas. *Cancer Res.*, 57: 2373-2377, 1997.
146. Scott, F. M., Treston, A. M., Shaw, G. L., Avis, I., Sorenson, J., Kelly, K., Dempsey, E. C., Cantor, A. B., Tockman, M., and Mulshine, J. L. Peptide amidating activity in human bronchoalveolar lavage fluid. *Lung Cancer (Limerick)*, 14: 239-251, 1998.
147. Scott, F. M., Modali, R., Lehman, T. A., Seddon, M., Kelly, K., Dempsey, E. C., Wilson, V., Tockman, M. S., and Mulshine, J. L. High frequency of K-ras codon 12 mutations in bronchoalveolar lavage fluid of patients at high risk for second primary lung cancer. *Clin. Cancer Res.*, 3: 479-482, 1997.
148. Mills, N. E., Fishman, C. L., Rom, W. N., and Jacobson, D. R. Ras oncogene detection in bronchoalveolar lavage fluid from patients with lung cancer. *Lung Cancer (Limerick)*, 1 (Suppl.): 11, 1994.
149. Nawroz, H., Koch, W., Anker, P., Stroun, M., and Sidransky, D. Microsatellite alterations in serum DNA of head and neck cancer patients. *Nat. Med.*, 2: 1035-1037, 1996.
150. Brennan, J. A., Mao, L., Hruban, R. H., Boyle, J. O., Eby, Y. J., Koch, W. M., Goodman, S. N., and Sidransky, D. Molecular assessment of histopathological staging in squamous-cell carcinoma of the head and neck. *N. Engl. J. Med.*, 332: 429-435, 1995.
151. Ahrendt, S. A., Yang, S. C., Wu, L., Westra, W. H., Jen, J., Califano, J. A., and Sidransky, D. Comparison of oncogene mutation detection and telomerase activity for the molecular staging of non-small cell lung cancer. *Clin. Cancer Res.*, 3: 1207-1214, 1997.
152. Gazdar, A. F., and Minna, J. D. NCI series of cell lines: an historical perspective. *J. Cell. Biochem.*, 24 (Suppl.): 1-11, 1996.
153. Wistuba, I. J., Bryant, D., Behrens, C., Milchgrub, S., Virmani, A. K., Ashfaq, R., Minna, J. D., and Gazdar, A. F. Comparison of features of human lung cancer cell lines and their corresponding tumors. *Clin. Cancer Res.*, 5: 991-1000, 1999.
154. Amstad, P., Reddel, R. R., Pfeifer, A., Malan, S. L., Mark, G. D., and Harris, C. C. Neoplastic transformation of a human bronchial epithelial cell line by a recombinant retrovirus encoding viral Harvey ras. *Mol. Carcinog.*, 1: 151-160, 1988.
155. Franklin, W. A., Folkvord, J. M., Varela-Garcia, M., Kennedy, T., Proudfoot, S., Cook, R., Dempsey, E. C., Hehn, K., Bunn, P. A., and Miller, Y. E. Expansion of bronchial epithelial cell populations by in vitro culture of explants from dysplastic and histologically normal sites. *Am. J. Respir. Cell Mol. Biol.*, 15: 297-304, 1996.
156. Kelsey, K., Spitz, M., Zuo, A., and Wiencke, J. Deletion of glutathione S-transferase class μ and class θ genes interacts to enhance susceptibility to lung cancer in minority populations. *Cancer Causes Control*, 1997.

157. Wu, X., Zhao, Y., Hoon, S. E., Tomlinson, G. E., Minna, J. D., Hong, W. K., and Spitz, M. R. Benzo[a]pyrene diol epoxide-induced 3p21.3 aberrations and genetic predisposition to lung cancer. *Cancer Res.*, 58: 1605-1608, 1998.
158. Virmani, A. K., Fong, K. M., Kodagoda, D., McIntire, D., Hung, J., Tonk, V., Minna, J. D., and Gazdar, A. F. Allelotyping demonstrates common and distinct patterns of chromosomal loss in human lung cancer types. *Genes Chromosomes Cancer*, 21: 308-319, 1998.
159. Takahashi, T., Nau, M. M., Chiba, I., Birrer, M. J., Rosenberg, R. K., Vinocour, M., Levitt, M., Pass, H., Gazdar, A. F., and Minna, J. D. p53: a frequent target for genetic abnormalities in lung cancer. *Science (Washington DC)*, 246: 491-494, 1989.
160. Forgacs, E., Biesterveld, E. J., Sekido, Y., Fong, K. M., Muncer, S., Wistuba, I., Milchgrub, S., Brezinschek, R., Virmani, A., Gazdar, A. F., and Minna, J. D. Mutation analysis of the *PTEN/MMAC1* gene in lung cancer. *Oncogene*, 17: 1557-1565, 1998.
161. Mitsudomi, T., Steinberg, S., Oie, H. K., Mulshine, J. L., Phelps, R., Viallet, J., Pass, H., Minna, J. D., and Gazdar, A. F. *ras* gene mutations in non-small cell lung cancers are associated with shortened survival irrespective of treatment intent. *Cancer Res.*, 51: 4999-5002, 1991.
162. Sozzi, G., Veronese, M. L., Negrini, M., Baffa, R., Corticelli, M. G., Inoue, H., Tomielli, S., Pilotti, S., Ohta, M., Huebner, K., and Croce, C. M. The *FHIT* gene at 3p14.2 is abnormal in lung cancer. *Cell*, 85: 17-26, 1996.
163. Ambazhagan, R., Tihan, T., Bornman, D. M., Johnston, J. C., Saltz, J. H., Weigering, A., Piantadosi, S., and Gabrielson, E. Classification of small cell lung cancer and pulmonary carcinoid by gene expression profiles. *Cancer Res.*, 59: 5119-5122, 1999.
164. Hibi, K., Liu, Q., Beaudry, G. A., Madden, S. L., Westra, W. H., Wehage, S. L., Yang, S. C., Heilmiller, R. F., Bertelsen, A. H., Sidransky, D., and Jen, J. Serial analysis of gene expression in non-small cell lung cancer. *Cancer Res.*, 58: 5690-5694, 1998.
165. Sozzi, G., Pastorino, U., Moiraghi, L., Tagliabue, E., Pezzella, F., Ghirelli, C., Tomielli, S., Sard, L., Huebner, K., Pierotti, M. A., Croce, C. M., and Pilotti, S. Loss of *FHIT* function in lung cancer and preinvasive bronchial lesions. *Cancer Res.*, 58: 5032-5037, 1998.
166. Tockman, M. S., and Mulshine, J. L. Sputum screening by quantitative microscopy: a new dawn for detection of lung cancer? *Mayo Clin. Proc.*, 72: 788-790, 1997.
167. Jizzo, J., and Hittelman, W. N. Characterization of multistep tumorigenesis by *in situ* hybridization. In: M. Andreeff and D. Pinkel (eds.), *Introduction to FISH*. In press, 2001.
168. Neft, R. E., Crowell, R. E., Gilliland, F. D., Murphy, M. M., Lane, J. L., Harms, H., Coons, T., Heaphy, E., Belinsky, S. A., and Lechner, J. F. Frequency of trisomy 20 in nonmalignant bronchial epithelium from lung cancer patients and cancer-free former uranium miners and smokers. *Cancer Epidemiol. Biomark. Prev.*, 7: 1051-1054, 1998.
169. Heppell-Parton, A. C., Nacheva, E., Carter, N. P., and Rabbitts, P. H. A combined approach of conventional and molecular cytogenetics for detailed karyotypic analysis of the small cell lung carcinoma cell line U2020. *Cancer Genet. Cytogenet.*, 108: 110-119, 1999.
170. Lechner, J. F., Neft, R., Gilliland, F. D., Crowell, R. E., Auckely, D. H., Ternes, R. T., and Belinsky, S. A. Individuals at high risk for lung cancer have airway epithelial cells with chromosome aberrations frequently found in lung tumor cells. *In Vivo* 12: 23-26, 1998.
171. Walch, A. K., Zitzelsberger, H. F., Aubele, M. M., Mattis, A. E., Baehninger, M., Candidus, S., Prauer, H. W., Werner, M., and Hoffer, H. Typical and atypical carcinoid tumors of the lung are characterized by 11q deletions as detected by comparative genomic hybridization. *Am. J. Pathol.*, 153: 1089-1098, 1998.
172. Petersen, I., Bujard, M., Petersen, S., Wolf, G., Goeze, A., Schwendel, A., Langreck, H., Gellert, K., Reichel, M., Just, K., du Manoir, S., Cremer, T., Dietel, M., and Ried, T. Patterns of chromosomal imbalances in adenocarcinoma and squamous cell carcinoma of the lung. *Cancer Res.*, 57: 2331-2335, 1997.
173. Smith, A. L., Hung, J., Walker, L., Rogers, T. E., Vuitch, F., Lee, E., and Gazdar, A. F. Extensive areas of aneuploidy are present in the respiratory epithelium of lung cancer patients. *Br. J. Cancer*, 73: 203-209, 1996.
174. MacAulay, C. E., Lam, S., Kein-Parker, H., Gazdar, A., Guillaud, M., Payne, P., LeRiche, J., Dawe, C., Band, P., and Palci, B. Intermediate endpoint biomarkers for lung cancer chemoprevention. *SPICE*, 3260: 207-211, 1998.



Localization of tissue inhibitor of metalloproteinases 1 (TIMP-1) in human colorectal adenoma and adenocarcinoma.

Holten-Andersen MN, Hansen U, Brunner N, Nielsen HJ, Illemann M, Nielsen BS.

The Finsen Laboratory, Rigshospitalet, Denmark.

Tissue inhibitor of matrix metalloproteinases 1 (TIMP-1) inhibits the proteolytic activity of matrix metalloproteinases and hereby prevents cancer invasion. However, TIMP-1 also possesses other functions such as inhibition of apoptosis, induction of malignant transformation and stimulation of cell-growth. We have previously demonstrated that TIMP-1 is elevated in blood from colorectal cancer patients and that high TIMP-1 levels predict poor prognosis. To clarify the role of TIMP-1 in colorectal tumorigenesis, the expression pattern of TIMP-1 in benign and malignant colorectal tumors was studied. In all of 24 cases of colorectal adenocarcinoma TIMP-1 mRNA was detected by in situ hybridization. In all cases TIMP-1 expression was found in fibroblast-like cells located at the invasive front but was seen only sporadically in normal mucosa. No TIMP-1 mRNA was seen in any of the cases in benign or malignant epithelial cells, in vascular cells or smooth muscle cells. Comparison of sections processed for TIMP-1 in situ hybridization with sections immunohistochemically stained with antibodies against TIMP-1 showed good correlation between TIMP-1 mRNA and immunoreactivity. Combining TIMP-1 in situ hybridization with immunohistochemical staining for alpha-smooth muscle actin or CD68 showed TIMP-1 mRNA in myofibroblasts but not in macrophages. TIMP-1 mRNA was detected in 2 of 7 adenomatous polyps in the adenoma area: in both cases associated with focal stromal inflammation at the epithelial-stromal interface. In conclusion, TIMP-1 expression is a rare event in benign human colon tissue but is highly expressed by myofibroblasts in association with invading colon cancer cells.

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Localization of Tissue Inhibitor of Metalloproteinases 1 (TIMP-1) in Human Colorectal Adenoma and Adenocarcinoma

Mads N. Holten-Andersen^{1,2*}, Ulla Hansen³, Nils Brønner², Hans Jørgen Nielsen⁴, Martin Illemann¹ and Boye Schnack Nielsen¹

¹The Finsen-Laboratory, Rigshospitalet, Denmark

²The Institute for Pharmacology and Pathobiology, The Royal Veterinary and Agricultural University, Denmark

³Department of Pathology, University Hospital Hvidovre, Denmark

⁴Department of Surgical Gastroenterology, University Hospital Hvidovre, Denmark

Tissue inhibitor of matrix metalloproteinases 1 (TIMP-1) inhibits the proteolytic activity of matrix metalloproteinases and hereby prevents cancer invasion. However, TIMP-1 also possesses other functions such as inhibition of apoptosis, induction of malignant transformation and stimulation of cell growth. We have previously demonstrated that TIMP-1 is elevated in blood from colorectal cancer patients and that high TIMP-1 levels predict poor prognosis. To clarify the role of TIMP-1 in colorectal tumorigenesis, the expression pattern of TIMP-1 in benign and malignant colorectal tumors was studied. In all of 24 cases of colorectal adenocarcinoma TIMP-1 mRNA was detected by *in situ* hybridization. In all cases TIMP-1 expression was found in fibroblast-like cells located at the invasive front but was seen only sporadically in normal mucosa. No TIMP-1 mRNA was seen in any of the cases in benign or malignant epithelial cells, in vascular cells or smooth muscle cells. Comparison of sections processed for TIMP-1 *in situ* hybridization with sections immunohistochemically stained with antibodies against TIMP-1 showed good correlation between TIMP-1 mRNA and immunoreactivity. Combining TIMP-1 *in situ* hybridization with immunohistochemical staining for α -smooth muscle actin or CD68 showed TIMP-1 mRNA in myofibroblasts but not in macrophages. TIMP-1 mRNA was detected in 2 of 7 adenomatous polyps in the adenoma area; in both cases associated with focal stromal inflammation at the epithelial-stromal interface. In conclusion, TIMP-1 expression is a rare event in benign human colon tissue but is highly expressed by myofibroblasts in association with invading colon cancer cells.

Key words: TIMP-1; MMP; *in situ* hybridization; immunohistochemistry; myofibroblast

A prerequisite for cancer cell invasion and metastasis is the breakdown of tissue barriers mediated by proteolytic enzymes such as the matrix metalloproteinases (MMP).^{1,2} Under normal physiologic conditions, the tissue degrading activities of the MMPs are kept at bay by the presence of the naturally occurring inhibitors: tissue inhibitors of metalloproteinases (TIMP). TIMP-1, a 28 kDa glycoprotein demonstrated to be present in most bodily tissues and fluids, binds and inhibits MMPs in a 1:1 stoichiometric manner.^{3,4} Overexpression of TIMP-1 in various cancer models has shown a suppressive role in the malignant progression.⁵ However, as opposed to this anti-invasive role of TIMP-1, several recent studies have demonstrated quite different functions of this MMP-inhibitor including stimulation of cell growth, malignant transformation and inhibition of apoptosis, suggesting a possible tumor-promoting role of TIMP-1 in very early stages of tumorigenesis.^{6–10} Thus, it has been speculated that TIMP-1 may actually play a dual role in cancer progression and metastasis.¹¹

Several studies have demonstrated that tumor tissue levels of MMP mRNA and protein are significantly increased in various malignant diseases and that such MMP elevations are correlated with cancer cell invasion, metastasis and short patient survival.^{12,13} In addition, many reports have described similar overexpression of TIMP-1 mRNA and protein in several cancer types.^{14–20} Moreover, we and others have demonstrated that measurement of increased plasma levels of TIMP-1 by immunoassay serves as a strong marker for short survival and recurrence of disease in patients with colorectal cancer.^{21–25} Similarly, a strong correlation

between high protein levels and poor prognosis is known for the type-1 plasminogen activator inhibitor (PAI-1).^{24,25} Considering the protease inhibiting function of these inhibitors, these findings seemed controversial; however, alternative functions have been reported both for TIMP-1 as mentioned above as well as for PAI-1.²⁶

In order to better understand the role of TIMP-1 in colorectal cancer, histochemical analyses may provide some indications. A number of studies of the localization of TIMP-1 in colorectal cancer have been published; however, the results of these reports are somewhat contradictory. Newell and colleagues²⁷ reported that TIMP-1 mRNA was expressed both in invasive adenocarcinoma, carcinoma *in situ* and adenoma and that the expression was observed in both the stromal as well as the epithelial compartment of the tissues studied. In contrast, Zeng and colleagues^{12,28} reported that TIMP-1 mRNA was expressed only in the stromal compartment of colorectal adenocarcinomas in spindle-shaped cells surrounding the invasive cancer cells. The results of immunohistochemical studies of TIMP-1 in colon are also conflicting: Hewitt and colleagues¹⁹ reported that TIMP-1 was expressed in the connective tissue and basement membrane in both normal mucosa, adenomas and adenocarcinomas with only little staining of the neoplastic epithelium. On the other hand, Tomita and colleagues²⁹ reported that TIMP-1 was expressed in both stromal and epithelial cells in colonic polyps and adenomas, as well as in adenocarcinomas, in which the neoplastic cells were strongly immunoreactive.

In order to resolve these inconsistencies, we undertook our study and by *in situ* hybridization and immunohistochemistry demonstrated that TIMP-1 is expressed in myofibroblasts in the stroma at the invasive front of colorectal adenocarcinomas. Because TIMP-1 was virtually absent from normal colorectal epithelium, we evaluated the possibility of using TIMP-1 as a diagnostic tool to differentiate colorectal adenomas from Dukes' stage A colorectal adenocarcinomas.

Material and methods

Tissue samples

All tissue material included was obtained from University Hospital of Hvidovre (Copenhagen, Denmark) in accordance with a permission given by the local scientific ethical committee (KF 01-078/93). Fourteen archival samples (formalin fixed and paraffin embedded) collected from 1989 to 1993 included Dukes' stage A colorectal adenocarcinomas ($n = 8$) and colorectal adenomatous polyps ($n = 6$, 3 were pedunculated (1 with mild and 2 with

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*Correspondence to: Mads N. Holten-Andersen, The Institute for Pharmacology and Pathobiology, The Royal Veterinary and Agricultural University, Denmark.

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moderate dysplasia) and 3 were sessile (1 with moderate and 2 with focally severe dysplasia). Samples from 16 colorectal adenocarcinomas (1 Duke's stage A, 6 Duke's stage B, 8 Duke's stage C and 1 Duke's stage D), 1 villous adenoma and 1 malignant colon lymphoma were prospectively collected during 1999–2000. These prospectively collected tissue specimens were dissected so that samples contained both normal mucosa and tumor tissue and were obtained within 30 min following surgical bowel resection. The specimens were immediately fixed in 4% neutral buffered formalin for 20–24 hr and then paraffin embedded. The 14 archival samples had also been formalin fixed and paraffin embedded.

Generation of nonoverlapping TIMP-1 cDNA fragments by PCR

The full length TIMP-1 cDNA (GenBank NM_003254) cloned in pSP64 vector²⁹ was used as template to generate 2 nonoverlapping PCR fragments for *in vitro* transcription, and named f104 (bp 56–378) and f106 (bp 398–680). First, the whole insert (~780 bp) was cut out by digestion with *Hind* III and *Bam* HI and purified after agarose gel electrophoresis using the Qiaex II gel extraction kit (Qiagen, Crawley, United Kingdom). To generate nonoverlapping antisense probes and the corresponding sense probe, 2 PCR fragments were generated using upstream primers flanked by a linker sequence containing an *Eco*RI restriction enzyme site (underlined nucleotides) and a T3 polymerase binding sequence (boldface) 5'-(**gagaattc**attaacctcactaaaggaga)-3', and downstream primers flanked by a linker sequence containing a *Bam*HI restriction enzyme site and a T7 polymerase binding sequence 5'-(**ggatccta**atagcactatagggag)-3'. The TIMP-1 specific upstream primers were 5'-accacatggcccttg-3' for f104 and 5'-(linker)-gcaggatggactcttcaca-3' for f106, and the downstream primers were 5'-(linker)-actctcgtcgtcggttg-3' for f104 and 5'-(linker)-tatctggagccgaggact-3' for f106. PCR using the 2 f104 primers or the 2 f106 primers was done as previously described.³⁰

The PCR products were purified by column chromatography using S-200HR microspin columns (Amersham Pharmacia Biotech, Inc., Piscataway, NJ), and their size tested by agarose gel electrophoresis. Both migrated as ~300 bp fragments in accordance with the predicted size (322 and 282 bp, respectively). An ABI PRISM 310 genetic analyzer was employed for DNA sequencing analysis and was performed according to the manufacturer's instructions (Perkin Elmer, Applied Biosystems, Foster City, CA) using the primers specified above. The DNA sequences obtained were confirmed by comparison with the specific TIMP-1 cDNA nucleic acid sequence (GenBank NM_003254).

Plasmids containing human MMP-2 cDNA (pCol7201, bp 647–1284) and human MMP-9 cDNA (pCol9202, bp 1751–2326) have been described elsewhere.³¹

In vitro transcription

Antisense and sense riboprobes were labeled with ³⁵S UTP (NEN, Boston, MA) by *in vitro* transcription using T7 and T3 RNA polymerases (Roche, Basel, Switzerland). The DNA template was digested with DNase (Promega, Madison, WI). Nonincorporated ³⁵S UTP and DNA was removed by column chromatography using S-200HR microspin columns (Amersham Pharmacia Biotech, Inc., Piscataway, NJ). The ³⁵S activity was adjusted for every probe by dilution to 500,000 cpm/μl.

In situ hybridization

In situ hybridization was performed essentially as described previously.³² In brief, 3 μm paraffin sections were deparaffinized in xylene, hydrated with graded ethanol and boiled in a microwave oven for 10–12 min in 10 mM citrate buffer, pH 6.0. After additional 20 min at room temperature, the sections were dehydrated with graded ethanol and the ³⁵S labeled probes (2 × 10⁶ cpm in 20 μl hybridization mixture³¹ per slide) incubated overnight at 55°C in a humidified chamber. Sections were washed in Hellendahl chambers with SSC buffers containing 0.1% SDS and 10 mM DTT at 150 rpm at 55°C using a Bühler incubation shaker (Johanna Otto GmbH, Hechingen, Germany) for 10 min in 2 × SSC,

10 min in 0.5 × SSC, and 10 min in 0.2 × SSC. Sections were then RNase A treated for 10 min to remove nonspecifically bound riboprobe. Subsequent wash was performed in 0.2 × SSC as specified above. Sections were dehydrated and soaked into an autoradiographic emulsion (Ilford), exposed for 5–7 days if not otherwise stated and finally developed. Sections were counterstained with haematoxylin and eosin.

Immunoperoxidase staining

Immunohistochemistry was performed essentially as described previously.³² Five micrometer paraffin sections were deparaffinized with xylene and hydrated through ethanol/water dilutions. Tissue pretreatment was performed with protease-K (5 μg/ml) digestion for 20 min. Sections were blocked for endogenous peroxidase activity by treatment with 1% hydrogen peroxide for 15 min. The sections were washed in 50 mM Tris 150 mM NaCl, pH 7.6, containing 0.5% Triton X-100 (TBS-T). Incubation with antibodies was done overnight at 4°C. Sheep polyclonal antibodies (pAb) against TIMP-1 and nonimmune goat IgG were used at a final concentration of 4.0 μg/ml. Two monoclonal antibodies (MAb) against TIMP-1,³³ NM4 (clone rTIX6A, NeoMarkers, Fremont, CA) and CalB2 (clone 147-6D11, CalBiochem, Oncogene Res. Products, Cambridge, MA), and a MAb against trinitrophenyl (TNP)³⁴ were all incubated at 1.0 μg/ml (all 3 MABs are IgG1). CalB2 MAB recognizes both free TIMP-1 and TIMP-1 in complex with MMPs.³³ NM4 MAB only recognizes free TIMP-1.³³ According to the manufacturer's descriptions, both MABs are raised using recombinant human TIMP-1. The sheep polyclonal antibodies were raised by immunization with TIMP-1 purified from human dermal fibroblasts. The IgG was obtained by triple precipitation using ammonium-sulfate and characterized by immunodiffusion and rocket immunoelectrophoresis.³⁵ In addition, we have shown that the pAb recognize both free and MMP-complexed TIMP-1.³⁶ Furthermore, the specificity of the antibodies was analyzed by Western blotting analysis against recombinant human TIMP-1 expressed in NSO mouse myeloma cells. Here, the antibody preparation recognizes a band of approximately 28 kDa in accordance with the molecular weight of TIMP-1. To certify that the pAb recognize TIMP-1 in colon tumors, the antibodies were immobilized on a sepharose column. Total protein extracted from 3 colon adenocarcinomas was passed through the column 5 times and the bound and subsequently eluted protein analyzed in a Western blot using a TIMP-1 monoclonal antibody (MAC15). A single band of approximately 28 kDa was revealed in accordance with the molecular weight of TIMP-1 (results not shown). In immunohistochemistry, the sheep pAb were detected with biotinylated rabbit-anti-goat IgG, which cross-react with sheep IgG (1:100, code E466, DakoCytomation) followed by horseradish peroxidase in complex with streptavidin (code K377, DakoCytomation). The MABs were detected with the Envision-mouse reagent (EnVision reagent, K4003, DakoCytomation), followed by tyramine amplification, using biotinyl tyramine substrate as specified by the manufacturer (Nen, Boston, MA). Sections were developed with NovaRed substrate as specified by the manufacturer (Vector Laboratories, Burlingame, CA) for 15 min. Finally, sections were counterstained in Mayers haematoxylin, dehydrated in ethanol and mounted.

Combined in situ hybridization and immunohistochemistry

Double labeling by combining *in situ* hybridization and immunohistochemistry on paraffin sections has been described previously.³² In brief, using MAB against α-sm-actin (clone 1A4) diluted 1:1000, against cytokeratin (clone AE1/AE3) diluted 1:1000, or against CD68 (clone PGM1) diluted 1:200, sections were incubated for 2 hours at room temperature and then detected with anti-mouse-IgG/horse radish peroxidase-conjugated polymers (Envision-mouse reagent, DakoCytomation, Glostrup, Denmark). Sections were developed with diaminobenzidine (DAB) for 7–10 min, and immediately dehydrated for *in situ* hybridization, which was performed as described above using the antisense probes of f104. Sections were counterstained with haematoxylin.

Results

Analysis of TIMP-1 probes and antibodies for *in situ* hybridization and immunohistochemistry

Histopathological diagnosis of prospectively collected specimens from 18 colorectal lesions revealed 16 colorectal adenocarcinomas, 1 villous adenoma and 1 malignant lymphoma. ³⁵S-labeled antisense and sense RNA probes were generated by *in vitro* transcription from 2 nonoverlapping DNA sequences of the human TIMP-1 cDNA and tested by *in situ* hybridization on adjacent sections from 5 of the colorectal adenocarcinomas. The 2 antisense probes showed an identical hybridization pattern in all the 5 cases, located in the stromal compartment surrounding the invading cancer cells, while no specific signal was seen with the 2 sense probes (Fig. 1). To test whether the TIMP-1 mRNA was accompanied by TIMP-1 protein expression, immunohistochemistry was performed on 8 of the adenocarcinomas (including the 5 mentioned above) and the malignant lymphoma using sheep anti-human TIMP-1 polyclonal antibodies on sections adjacent to TIMP-1 *in situ* hybridized sections. The TIMP-1 mRNA and immunoreactivity was observed in the same cells in all of the 9 cases tested (Fig. 2A), including the malignant lymphoma. The anti-TIMP-1 polyclonal antibodies did not react with other cell populations in all of 8 adenocarcinomas and the malignant lymphoma apart from some normal and malignant epithelial cells that were weakly stained on the luminal apical surface. Two MABs against TIMP-1 (CalB2 and NM4) required strong signal amplification but showed a staining pattern similar to that of the polyclonal antibody preparation (Fig. 2B), with the only exception that neither of the 2 MABs stained the luminal apical surface of the normal and malignant epithelium. No signal was obtained with nonimmune goat serum or a MAB (of same subclass as CalB2 and NM4) directed against the synthetic hapten trinitrophenyl (TNP).

TIMP-1 mRNA expression patterns in colon cancer

Expression of TIMP-1 mRNA was then analyzed in the remaining 9 colorectal lesions by *in situ* hybridization. TIMP-1 mRNA

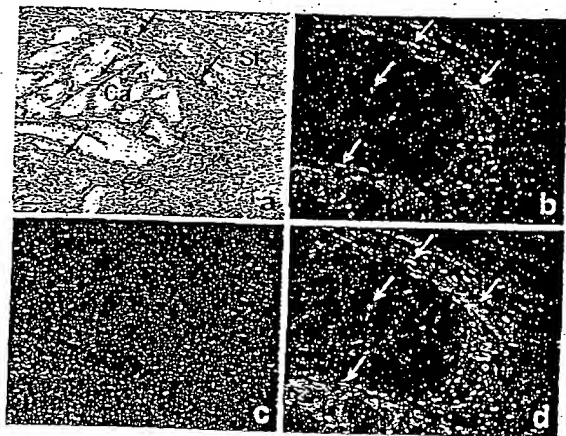


FIGURE 1—*In situ* hybridization with 2 nonoverlapping TIMP-1 specific probes in human colon cancer. Three adjacent sections from a colon adenocarcinoma were incubated with 2 nonoverlapping ³⁵S-labeled antisense probes for TIMP-1 mRNA (f106 (a,b) and f104 (d)) and a corresponding TIMP-1 sense probe, f106 (c). The *in situ* hybridization signal is identified as black silver grains and demonstrated in brightfield (a) and as a white pattern in darkfield illumination (b-d). The 2 antisense probes show the same hybridization pattern and the hybridization signal is seen in the same cells (arrows in a, b and d), whereas no specific signal is seen with the sense probe (c). Note that the TIMP-1 mRNA signal is located in the tumor stroma (indicated by St) surrounding the invasive cancer cells (Ca) that are devoid of TIMP-1 *in situ* hybridization signal. (a)-(d): Bar = 100 μ m.

expression was in all the cases of colon adenocarcinoma (including those mentioned above) highly expressed in stromal fibroblast-like cells located at the invasive front (Fig. 3a,d). TIMP-1 mRNA signal was also observed in fibroblast-like cells located in the tumor stroma towards the colonic lumen in 8 of 10 cases where this tissue structure was present (data not shown). No or little TIMP-1 mRNA was detected in the central part of the carcinomas. In 5 of the 16 colorectal adenocarcinomas, we observed TIMP-1 mRNA signal in some fibroblast-like cells located around the muscle layer of some arteries located in the submucosa distant from the cancer area. The normal colonic mucosa, including the lamina propria that was present in all samples tested, was generally negative (Fig. 3b,e). Only a relatively weak TIMP-1 mRNA signal was detected in stromal fibroblast-like cells surrounding one or a very few normal crypts (Fig. 3c,f) in 3 out of 6 cases tested with extended exposure time (10 days vs. usually 5 days). In the villous adenoma, we saw only a few TIMP-1 mRNA positive cells associated with focal inflammation (data not shown). In the malignant lymphoma of the colon, TIMP-1 mRNA expressing fibroblast-like cells were, different from the adenocarcinomas, located in a diffuse pattern throughout the whole tumor. No TIMP-1 mRNA signal was observed in any of the 18 cases in the cancer cells, smooth muscle cells or vascular cells.

Characterization of TIMP-1 mRNA expressing cells

To test whether the TIMP-1 mRNA positive fibroblast-like cells could be (myo)fibroblasts and/or macrophages, sections from 4 colorectal adenocarcinomas and the malignant lymphoma were first immunohistochemically stained with antibodies directed against α -sm-actin [for detection of myofibroblast/smooth muscle cells (SMC)] or CD68 (for detection of macrophages) and subsequently incubated with a TIMP-1 mRNA antisense probe. In normal colon tissue, α -sm-actin is expressed by vascular smooth muscle cells, smooth muscle cells of lamina muscularis mucosae and tunica muscularis as well as pericryptal myofibroblasts.³⁷ In colon tumors, α -sm-actin is expressed by tumor-associated fibroblast-like cells located throughout the tumor stroma, which are defined as myofibroblasts. No TIMP-1 mRNA was detected in any α -sm-actin positive smooth muscle cells, including those of the vessels, the lamina muscularis mucosae and the tunica muscularis. In addition, no TIMP-1 mRNA was detected in the α -sm-actin positive pericryptal myofibroblasts of the lamina propria in any of the 5 lesions. TIMP-1 mRNA signal was in contrast seen in α -sm-actin-positive tumor associated myofibroblasts located at the invasive front of the colon cancers. In 3 of the adenocarcinomas, more than 80% of TIMP-1 mRNA-positive cells located close to the invading cancer cells were α -sm-actin-positive (Fig. 4). TIMP-1 mRNA positive fibroblast-like cells located more distant from the invasive cancer cells, towards the submucosa, expressed little or no α -sm-actin. In 1 adenocarcinoma and in the malignant lymphoma approximately 50% of the TIMP-1 mRNA positive cells expressed α -sm-actin. Thus, the TIMP-1 mRNA expressing cells constitute a subpopulation of tumor-associated myofibroblasts located at the invasive front of the tumor. No TIMP-1 mRNA signal could be identified in any of the CD68-positive cells (Fig. 4).

Expression of TIMP-1 and MMP-2 and 9 in colon cancer

MMP-2 and MMP-9 are 2 type IV collagenases expressed in the invasive cancer tissue of colorectal adenocarcinomas. Thus MMP-2 has been reported to be expressed by fibroblast-like cells in the cancer stroma,^{12,31} and MMP-9 by macrophages at the leading edge of the invasive cancer.³⁸ To directly compare the expression patterns of MMP-2 and MMP-9 with that of TIMP-1, adjacent sections from 5 colorectal adenocarcinomas were hybridized with probes for TIMP-1, MMP-2 and MMP-9 mRNAs. We found that the expression of TIMP-1 mRNA was localized characteristically at the invasive front of the growing tumor, whereas the expression of MMP-2 mRNA was most intense in the central areas, showing decreased expression towards the invasive front (Fig. 5a). MMP-9 mRNA expressing cells were found at the

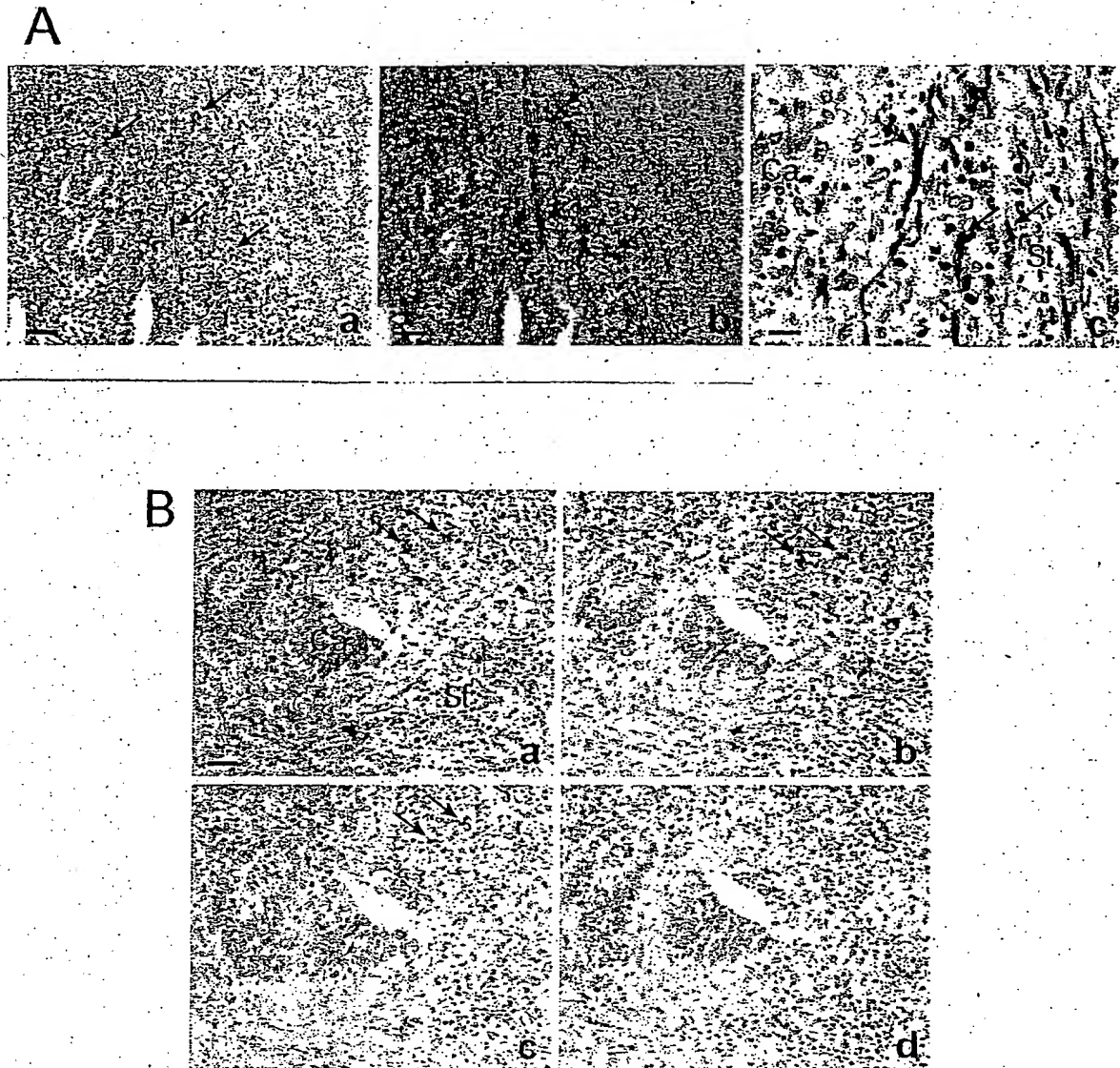


FIGURE 2—*In situ* hybridization and immunohistochemistry for TIMP-1 in human colon cancer. (A) Two adjacent sections from a human colon adenocarcinoma were incubated with polyclonal antibodies against TIMP-1 (a,c) and a TIMP-1 mRNA antisense probe (b). The TIMP-1 immunoreactivity (red-brown color, arrows in a and c) and the TIMP-1 mRNA (silver grains, arrows in b) are identified in the same cells (arrows in a,b). Immunoperoxidase staining with the TIMP-1 pAb reveals the TIMP-1-positive cells as fibroblast-like cells (arrows in c) located in the stroma (St). No TIMP-1 immunoreactivity is seen in cancer cells (Ca). a,b: bars = 50 μ m; c: bars = 13 μ m. (B) Four consecutive adjacent sections were incubated with CalB2 MAb anti-TIMP-1 (a), NM4 MAb (b), sheep anti TIMP-1 pAb (c) or mouse anti TNP (d). The 3 MABs were detected with Envision reagent followed by TS amplification and the sheep pAb with biotinylated rabbit anti-goat followed by HRP-conjugated streptavidin (see Material and methods). The 3 TIMP-1 antibodies react with the same cells (arrows). No immunoreactivity is seen when the sections are incubated with anti-TNP.

invasive front like those expressing TIMP-1 mRNA but with a distinctly different distribution. Foci with high expression of TIMP-1 mRNA were not accompanied by increased expression of MMP-9 mRNA and vice-versa (Fig. 5b). Thus, TIMP-1 mRNA expression is not coregulated with MMP-2 or MMP-9 mRNA expression.

TIMP-1 in adenomas and Dukes' stage A carcinomas

TIMP-1 antigen can readily be measured in blood and we have previously reported that levels of TIMP-1 in blood are significantly

elevated in colorectal cancer patients compared to healthy donors and that high plasma TIMP-1 levels are associated with short survival of colorectal cancer patients.^{21,56} TIMP-1 has therefore been suggested to be a novel marker for detection of early stage colorectal cancer and for prognostic stratification of colorectal cancer patients.^{21,39} These findings, together with the characteristic expression pattern of TIMP-1 at the invasive front of virtually all the colon cancers and the absence or minute TIMP-1 expression in normal and benign colon mucosa, prompted the evaluation of TIMP-1 expression as a marker for early invasive colon cancer.

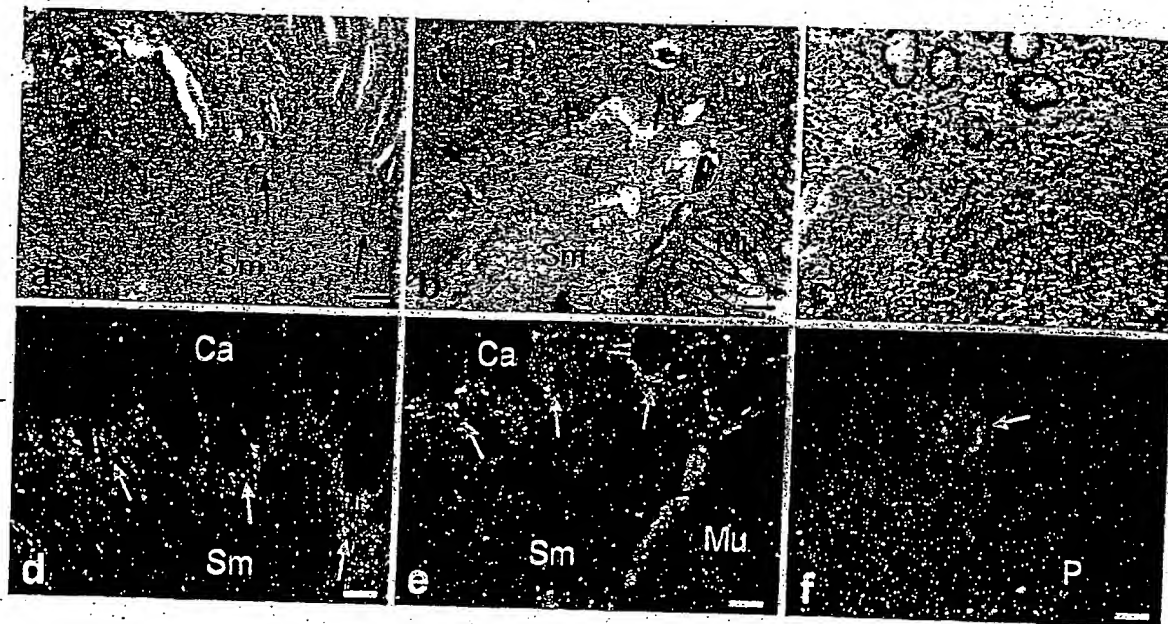


FIGURE 3—Expression patterns of TIMP-1 mRNA in human colon cancer. Sections were incubated with an antisense probe for TIMP-1 mRNA. The TIMP-1 mRNA signal is demonstrated in brightfield (a-c) and darkfield (d-f). The TIMP-1 mRNA is highly expressed in a subpopulation of stromal fibroblast-like cells at the invasive front of the cancer (a,d), whereas a low hybridization signal is seen within central parts of the cancer area (Ca) and in the submucosa (Sm). The normal mucosa (indicated by Mu in b and e) is negative. A low *in situ* hybridization signal was detected in a few stromal cells surrounding a few normal-looking glands (c,f), see also text. Exposure time: a,b,d,e, 5 days; d,f, 10 days. (a,b,d,f) bars = 200 μ m; (c,f) bars = 25 μ m.

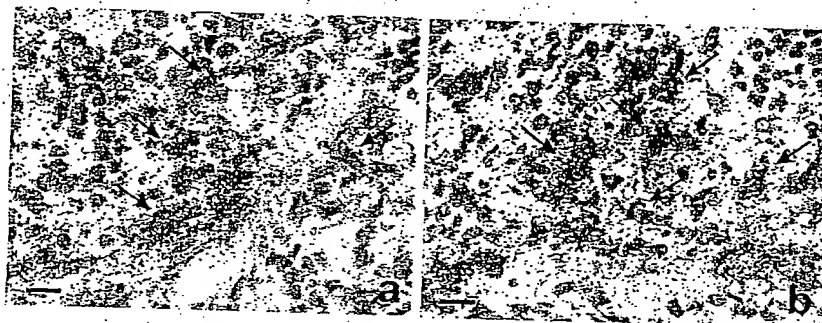


FIGURE 4—Double labeling for TIMP-1 mRNA and α -sm-actin or CD68 in human colon cancer. Sections were first processed for immunohistochemistry incubating antibodies against α -sm-actin (a) or CD68 (b) and subsequently by *in situ* hybridization using the 5104 TIMP-1 specific antisense probe. TIMP-1 mRNA signal is colocalized with α -sm-actin immunoreactivity in fibroblast-like cells that are considered as myofibroblasts (black arrows in a), whereas no CD68 immunoreactivity is seen in the TIMP-1 mRNA positive cells (black arrows in b) in an area with several CD68-positive macrophages (red arrows in b). Note that some of the α -sm-actin-positive myofibroblasts have little or no TIMP-1 mRNA signal (red arrows in a). Bars = 13 μ m.

Therefore, we compared TIMP-1 mRNA expression in an additional 6 colorectal adenomatous polyps with the expression in an additional 8 Duke's stage A colorectal adenocarcinomas. TIMP-1 mRNA signal was detected in 2 of the 6 adenomas, whereas all 8 Duke's stage A carcinomas showed TIMP-1 mRNA signal at the invasive front (Fig. 6). In 1 positive adenoma (pedunculated type), TIMP-1 mRNA expression was confined to a single focus in fibroblast-like cells associated with focal stromal inflammation (Fig. 6). However, histological analysis of additional sections from this sample clearly revealed disruption of the dysplastic epithelium in the same area. In the other TIMP-1 mRNA positive adenoma (sessile type), a few TIMP-1 mRNA expressing fibroblasts were located around small arteries not directly associated with the tumor area (data not shown).

Taken together, all of the 9 Duke's stage A carcinomas analyzed showed TIMP-1 mRNA expression in myofibroblasts located at the invasive front of the tumors, whereas expression was detected in only 3 of 7 adenomas, and in these was seen in fibroblast-like cells associated with focal inflammation at the epithelial-stromal interface in 2 of the cases and with arteries in the submucosa in 1 case.

Discussion

Our study was undertaken to clarify the expression and cellular localization of the MMP inhibitor TIMP-1 in human colon adenocarcinomas. Our studies were founded on the use of 2 specific antisense RNA probes derived from 2 nonoverlapping TIMP-1

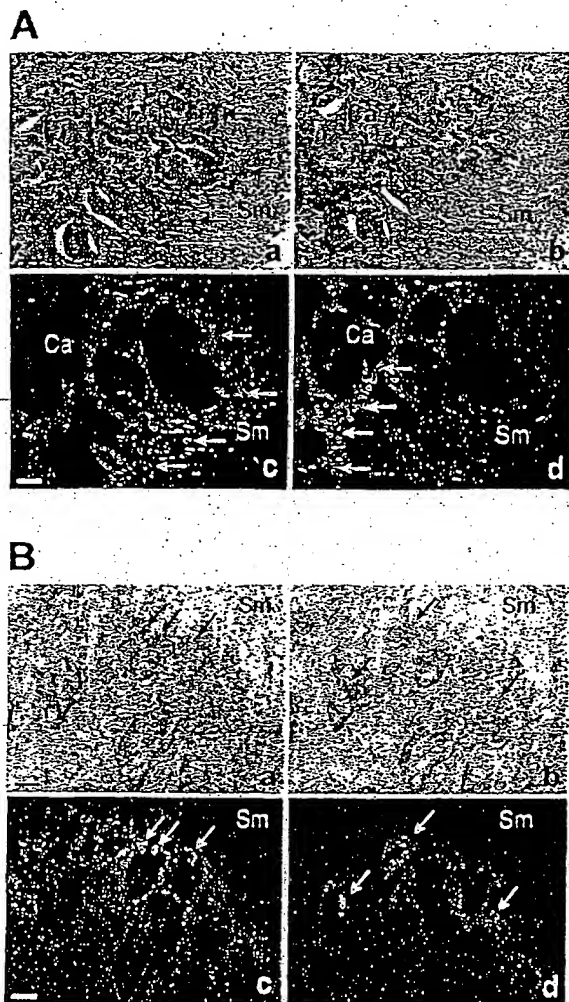


FIGURE 5—*In situ* hybridization for TIMP-1, MMP-2 and MMP-9 in human colon cancer. (A) Adjacent sections were incubated with probes specific for TIMP-1 mRNA (a,c) and MMP-2 mRNA (b,d), respectively, and is shown in brightfield (a,b) and darkfield illumination (c,d). The TIMP-1 mRNA signal increases towards the submucosa (Sm) whereas the MMP-2 mRNA signal decreases and is most intense in the central areas (Ca). (B) Adjacent sections were incubated with probes specific for TIMP-1 mRNA (a,c) and MMP-9 mRNA (b,d), respectively, and is here shown in brightfield (a,b) and darkfield illumination (c,d). Both the TIMP-1 mRNA signal and the MMP-9 mRNA signal are most intense at the invasive front towards the submucosa (Sm), but their expression patterns are quite different, with MMP-9 showing the most restricted expression. Bars = 100 μ m.

cDNA fragments and specific pAb and MAbs against human TIMP-1. The TIMP-1 mRNA signal in all colorectal adenocarcinomas investigated was seen in fibroblast-like cells located in the tumor periphery. An identical hybridization pattern was observed with the 2 antisense TIMP-1 probes and application of complementary sense probes on neighboring tissue sections as negative controls did not result in any hybridization signal; therefore, we conclude that the hybridization signal generated with the antisense probes represents the genuine TIMP-1 mRNA. TIMP-1 immunoreactivity was also distinctly located in fibroblast-like stromal cells in the tumor periphery, and these cells were identified to be the same cells as the TIMP-1 mRNA expressing cells. A preparation

of sheep pAb against human TIMP-1³⁵ and 2 well-characterized MAbs stained the very same cells in the tumor stroma. Weak staining of the apical surface of some normal and malignant epithelial cells was observed with the pAb in some of the samples. No staining was obtained when the anti-TIMP-1 antibodies were substituted with nonimmune goat serum or anti-TNP MAb incubated at the same concentrations. These immunohistochemical findings strongly suggest that the TIMP-1 antigen detected in the fibroblast-like cells represents the genuine TIMP-1 protein.

In our study, we found TIMP-1 mRNA expression in stromal fibroblast-like cells located in the tumor periphery in all colorectal adenocarcinomas tested, whereas no expression was detected in the cancer cells in any of the cases tested. This finding is in agreement with studies by Zeng and colleagues,^{12,20} but is partly in disagreement with findings by Newell and colleagues.²⁷ In addition to TIMP-1 mRNA signal in fibroblast-like cells in the tumor periphery, Newell and colleagues²⁷ detected a weak TIMP-1 mRNA signal in both benign and malignant epithelial cells.²⁷ This observation was, however, based on the use of probes from a single TIMP-1 cDNA subclone and no additional controls to verify the expression pattern. The difference between our results and those of Newell and colleagues may be explained by methodological differences, since the procedure employed by Newell and colleagues was in several steps different from the one used in the present study, e.g., Newell and colleagues used ³H-labeled probes, whereas we used ³⁵S-labeled probes. It is in this context noteworthy that in order to look for a low expression level of TIMP-1 mRNA, we performed *in situ* hybridization experiments with prolonged exposure time (10 days vs. usually 5 days) with both our TIMP-1 antisense probes and both TIMP-1 sense probes, but with this challenge we did not detect any TIMP-1 mRNA in any epithelial cells. It cannot be excluded though that the TIMP-1 mRNA is expressed in epithelial cells below the detection limit of our *in situ* hybridization procedure.

An interesting observation in our study was the characteristic intense TIMP-1 mRNA and protein expression in the tumor periphery of all colon adenocarcinomas, while little or no expression was seen in the center of the carcinomas. Only in the colorectal lymphoma did we find TIMP-1 mRNA and protein expression in fibroblast-like cells located throughout the tumor tissue. The TIMP-1 expression pattern in the colon adenocarcinomas is in contrast to the expression pattern reported by Hewitt and colleagues,¹⁹ who found that the TIMP-1 staining in most colorectal adenocarcinomas was equally intense in fibroblasts throughout the tumors and that some of the cases even showed decreased TIMP-1 signal intensity towards the tumor periphery. This difference may be explained by possible cross-reactivity of the polyclonal antibodies employed by Hewitt and colleagues or that Hewitt and colleagues employed cryostat sections, while we analyzed paraffin sections.

The TIMP-1 expressing cells had a fibroblast-like morphology and using combined *in situ* hybridization for TIMP-1 mRNA and immunohistochemistry for α -sm-actin, we found that many, generally more than 50%, of the TIMP-1 mRNA positive cells coexpressed α -sm-actin. According to the cellular morphology of the TIMP-1 expressing cells and their localization in the invasive front, we could conclude that the cells were myofibroblasts and not smooth muscle cells.

The myofibroblast is a cell type present in the normal colon mucosa, that originally was described as a pericryptal fibroblast^{37,40} and later was identified with antibodies against α -sm-actin.²⁷ In the lamina propria, the myofibroblasts form a continuous cell layer just below the intestinal epithelium. The pericryptal myofibroblasts are phenotypically different from the neighboring quiescent interstitial fibroblasts that do not express markers of smooth muscle cells.⁴¹ During early steps of colonic tumorigenesis the number of myofibroblasts is significantly increased.⁴¹ The TIMP-1 expressing myofibroblasts may be generated after activation of the pericryptal myofibroblasts and/or the quiescent inter-

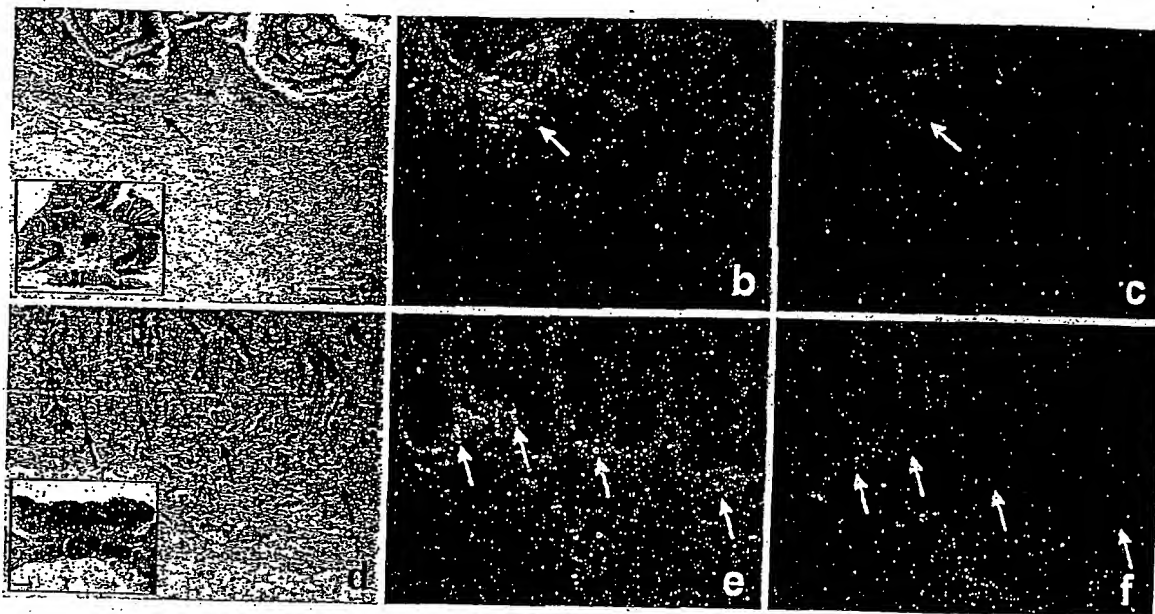


FIGURE 6—*In situ* hybridization for TIMP-1 in colon adenoma and Dukes' stage A adenocarcinoma. Sections from a moderate dysplastic sessile adenoma of the colon (a-c) and a Dukes' A colon carcinoma (d-f) were hybridized with TIMP-1 antisense (a,b and d,e) and sense (c,f) probes, shown in brightfield (a and d) and darkfield illumination (b,c and e,f). Inserts in a and d show virtually the whole tissue section with the arrow indicating the magnified area. TIMP-1 mRNA expression is confined to a single focus in the adenoma (arrows in a,b), whereas the TIMP-1 mRNA is detected along the invasive front in the Dukes' A colon carcinoma (arrows in d,e). (a)-(f): bar = 200 μ m. Bar in inserts = 1,250 μ m.

stitial fibroblasts. Adegboyega and colleagues⁴¹ hypothesized that the tumor-associated myofibroblasts originate from the quiescent interstitial fibroblasts of the lamina propria, rather than from pericryptal myofibroblast or smooth muscle cells, which may help to explain why we found some of the TIMP-1 mRNA expressing fibroblast-like cells α -sm-actin-positive and some α -sm-actin-negative.

Several MMPs including MMP-2, MMP-11 and MMP-14 are expressed by fibroblast-like cells in human colon cancer,^{27,51,42-44} some of which may indeed be myofibroblasts. The role of the (myo-) fibroblasts in colon cancer progression is not known. Since TIMP-1 in human colon cancer appears only to be expressed by fibroblast-like cells most of which are myofibroblasts, and since high TIMP-1 levels measured in blood or tumor extracts from colon cancer patients are strongly associated with a poor prognosis,^{20,21,27,28} it could be argued that the TIMP-1 expressing myofibroblasts play a tumor-promoting role. Immunohistochemical localization studies of proteins involved in the activation and regulation of the efficient serine protease plasminogen, including urokinase plasminogen activator (uPA) and its specific inhibitor PAI-1 show that both are mainly expressed by myofibroblasts in human breast cancer.^{45,46} High levels of uPA and PAI-1 are strongly correlated with poor prognosis in breast cancer,^{47,48} supporting the assumption that the myofibroblast express a promoting role in cancer invasion. We recently reported that the predominant PAI-1 expressing cell in human colorectal cancer also is the myofibroblast,⁴⁹ and earlier studies indicated that elevated levels of PAI-1 in colon cancer patients are associated with poor prognosis.²⁵ Together these findings indicate that myofibroblasts are strongly contributing to the expression of proteins involved in the regulation of extracellular matrix degrading proteases that facilitate cancer invasion and metastasis.

A particularly interesting finding of the present study was the absence of TIMP-1 mRNA in 4 of 7 adenomas, whereas in all of 9 Dukes' stage A carcinomas the TIMP-1 mRNA was expressed in fibroblast-like cells along the invasive front. In the 2 benign lesions, in which the TIMP-1 mRNA was seen in the adenoma area, the

TIMP-1 mRNA positive cells were confined to a single focus with locally increased inflammation related to the dysplastic epithelium. Evident disruption of the dysplastic epithelium was observed in the adenoma with most intense TIMP-1 mRNA signal. Intestinal inflammation may be caused by disruption of the mucous epithelium that leads to focal leakage of mucinous colon material into the lamina propria. Increased intestinal permeability is a common deficiency in Crohn's disease and interestingly TIMP-1 mRNA was found in the intestinal granulation tissue of Crohn's disease⁵⁰ and is expressed by myofibroblasts isolated from Crohn's disease.⁵¹ Induction of TIMP-1 in myofibroblasts in a benign or preinvasive tumor may also be a response to locally increased MMP activity or a response to the presence of a specific MMP in the local microenvironment. MMP-2 and MMP-9 mRNA expression, however, did not appear to be coregulated with TIMP-1 mRNA expression in the colorectal adenocarcinomas. Specific MMPs may indeed be involved in the transition of noninvasive to invasive disease; in studies of preinvasive lesions (ductal carcinomas *in situ*) of the human breast we recently reported that MMP-13 is specifically expressed in myofibroblasts associated with microinvasive events.³² Future studies may clarify whether TIMP-1 expression in colorectal adenomas is correlated with expression of specific MMPs, cytokines and/or growth factors, such as TGF- β 1 and TGF- β 251, and whether TIMP-1 can be used as a histopathological marker for malignancy in colorectal tumors.

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References

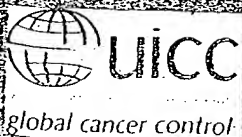
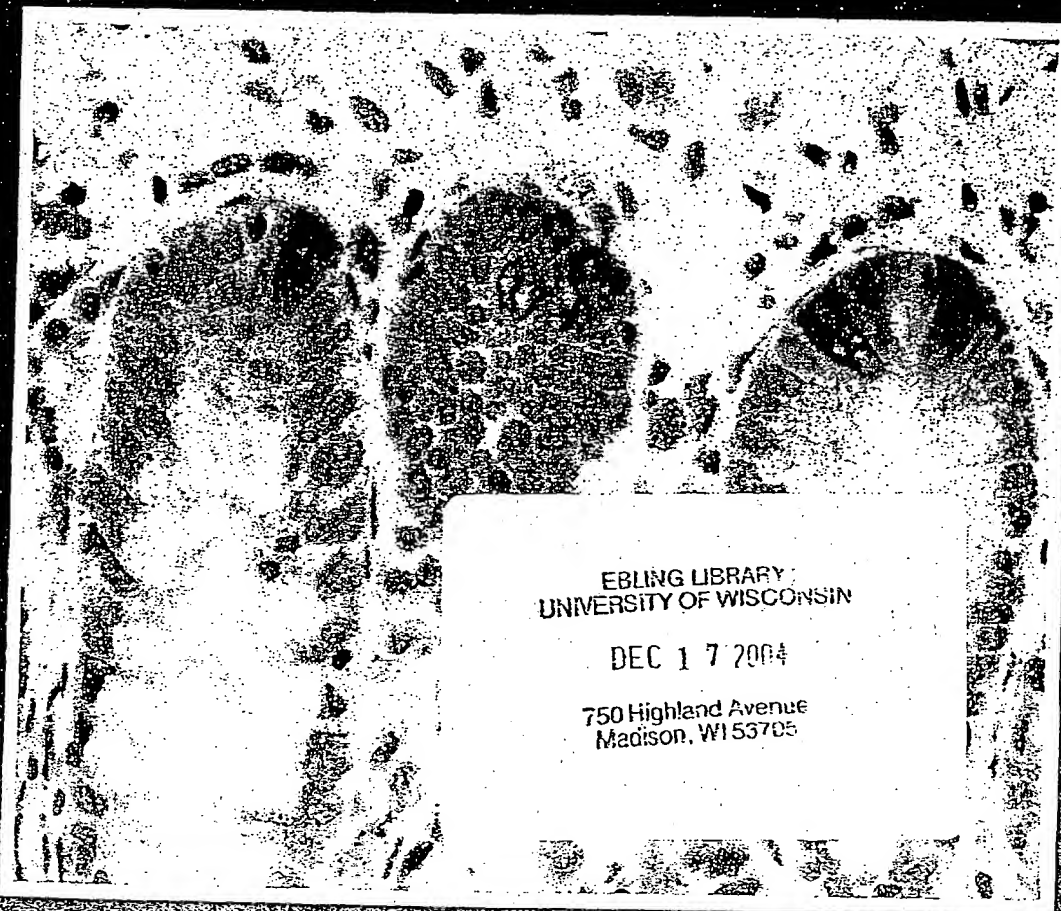
- Liotta LA, Steeg PS, Stetler-Stevenson WG. Cancer metastasis and angiogenesis: an imbalance of positive and negative regulation. *Cell* 1991;64:327-36.
- Stetler-Stevenson WG, Yn AE. Proteases in invasion: matrix metalloproteinases. *Semin Cancer Biol* 2001;11:143-52.
- Murphy G, Kokkiliis P, Carne AF. Dissociation of tissue inhibitor of metalloproteinases (TIMP) from enzyme complexes yields fully active inhibitor. *Biochem J* 1989;261:1031-4.
- Welgus HG, Stricklin GP. Human skin fibroblast collagenase inhibitor. Comparative studies in human connective tissues, serum, and amniotic fluid. *J Biol Chem* 1983;258:12259-64.
- Khokha R, Waterhouse P. The role of tissue inhibitor of metalloproteinase-1 in specific aspects of cancer progression and reproduction. *J Neurooncol* 1994;18:123-7.
- Docherty AJ, Lyons A, Smith BJ, Wright EM, Stephens PE, Harris TJ, Murphy G, Reynolds JJ. Sequence of human tissue inhibitor of metalloproteinases and its identity to erythroid-potentiating activity. *Nature* 1985;318:66-9.
- Guedez I, Courtemanche L, Stetler-Stevenson M. Tissue inhibitor of metalloproteinase (TIMP)-1 induces differentiation and an antiapoptotic phenotype in germinal center B cells. *Blood* 1998;92:1342-9.
- Hewitt RE, Brown KE, Corcoran M, Stetler-Stevenson WG. Increased expression of tissue inhibitor of metalloproteinases type 1 (TIMP-1) in a more tumorigenic colon cancer cell line. *J Pathol* 2000;192:455-9.
- Guedez I, Stetler-Stevenson WG, Wolff L, Wang J, Fukushima P, Mansoor A, Stetler-Stevenson M. In vitro suppression of programmed cell death of B cells by tissue inhibitor of metalloproteinases-1. *J Clin Invest* 1998;102:2002-10.
- Li G, Fridman R, Kim HR. Tissue inhibitor of metalloproteinase-1 inhibits apoptosis of human breast epithelial cells. *Cancer Res* 1999;59:6267-75.
- Jiang Y, Goldberg ID, Shi YE. Complex roles of tissue inhibitors of metalloproteinases in cancer. *Oncogene* 2002;21:2245-52.
- Zeng ZS, Guillem JG. Distinct pattern of matrix metalloproteinase 9 and tissue inhibitor of metalloproteinase 1 mRNA expression in human colorectal cancer and liver metastases. *Br J Cancer* 1995;72:575-82.
- Gomez DE, Alonso DF, Yoshiji H, Thorgerisson UP. Tissue inhibitors of metalloproteinases: structure, regulation and biological functions. *Eur J Cell Biol* 1997;74:111-22.
- Guillem JG, Levy MF, Hsieh LL, Johnson MD, LoGerfo P, Forde KA, Weinstein IB. Increased levels of p16, c-myc, and ornithine decarboxylase RNAs in human colon cancer. *Mol Carcinog* 1990;3:68-74.
- Lu XQ, Levy M, Weinstein IB, Santella RM. Immunological quantitation of levels of tissue inhibitor of metalloproteinase-1 in human colon cancer. *Cancer Res* 1991;51:6231-5.
- Mimori K, Mori M, Shiraiishi T, Fujie T, Baba K, Haraguchi M, Abe R, Ueo H, Akiyoshi T. Clinical significance of tissue inhibitor of metalloproteinase expression in gastric carcinoma. *Br J Cancer* 1997;76:531-6.
- Fong KM. TIMP1 and adverse prognosis in non-small cell lung cancer. *Clinical Cancer Research* 1996;2:1369-72.
- Ree AH, Florenes VA, Berg JP, Maclandsmo GM, Nesland JM, Fodstad O. High levels of messenger RNAs for tissue inhibitors of metalloproteinases (TIMP-1 and TIMP-2) in primary breast carcinomas are associated with development of distant metastases. *Clin Cancer Res* 1997;3:1623-8.
- Hewitt RE, Leach JH, Powe DG, Clark IM, Cawston TE, Turner DR. Distribution of collagenase and tissue inhibitor of metalloproteinases (TIMP) in colorectal tumours. *Int J Cancer* 1991;49:666-72.
- Zeng ZS, Cohen AM, Zhang ZF, Stetler-Stevenson W, Guillem JG. Elevated tissue inhibitor of metalloproteinase 1 RNA in colorectal cancer stroma correlates with lymph node and distant metastases. *Clin Cancer Res* 1995;1:899-906.
- Holten-Andersen MN, Stephens RW, Nielsen HJ, Murphy G, Christensen JJ, Stetler-Stevenson W, Brunner N. High preoperative plasma tissue inhibitor of metalloproteinase-1 levels are associated with short survival of patients with colorectal cancer. *Clin Cancer Res* 2000;6:4292-9.
- Oberg A, Hoyhtya M, Tavelin B, Stenling R, Lindmark G. Limited value of preoperative serum analyses of matrix metalloproteinases (MMP-2, MMP-9) and tissue inhibitors of matrix metalloproteinases (TIMP-1, TIMP-2) in colorectal cancer. *Anticancer Res* 2000;20:1085-91.
- Pellegrini P, Contasta I, Berghella AM, Gargano E, Mammarella C, Adorno D. Simultaneous measurement of soluble carcinoembryonic antigen and the tissue inhibitor of metalloproteinase TIMP1 serum levels for use as markers of pre-invasive to invasive colorectal cancer. *Cancer Immunol Immunother* 2000;49:388-94.
- Grondahl-Hansen J, Christensen JJ, Rosenquist C, Brunner N, Mouridsen HT, Dano K, Blichert-Toft M. High levels of urokinase-type plasminogen activator and its inhibitor PAI-1 in cytosolic extracts of breast carcinomas are associated with poor prognosis. *Cancer Res* 1993;53:2513-21.
- Nielsen HJ, Pappot H, Christensen JJ, Brunner N, Thordarson-Ussing O, Moesgaard F, Dano K, Grondahl-Hansen J. Association between plasma concentrations of plasminogen activator inhibitor-1 and survival in patients with colorectal cancer. *BMJ* 1998;316:829-30.
- Wind T, Hansen M, Jensen JK, Andreasen PA. The molecular basis for anti-proteolytic and non-proteolytic functions of plasminogen activator inhibitor type-1: roles of the reactive centre loop, the shutter region, the flexible joint region and the small serpin fragment. *Biol Chem* 2002;383:21-36.
- Newell KJ, Witty JP, Rodgers WH, Matrisian LM. Expression and localization of matrix-degrading metalloproteinases during colorectal tumorigenesis. *Mol Carcinog* 1994;10:199-206.
- Tomita T, Iwata K. Matrix metalloproteinases and tissue inhibitors of metalloproteinases in colonic adenomas-adenocarcinomas. *Dis Colon Rectum* 1996;39:1255-64.
- O'Shea M, Willenbrock F, Williamson RA, Cockeitt MI, Freedman RB, Reynolds JJ, Docherty AJ, Murphy G. Site-directed mutations that alter the inhibitory activity of the tissue inhibitor of metalloproteinases-1: importance of the N-terminal region between cysteine 3 and cysteine 13. *Biochemistry* 1992;31:10146-52.
- Engelholm LH, Nielsen BS, Netzel-Arnett S, Solberg H, Chen XD, Lopez Garcia JM, Lopez-Otin C, Young MF, Birkedal-Hansen H, Dano K, Lund LR, Behrendt N, et al. The urokinase plasminogen activator receptor-associated protein/endo180 is coexpressed with its interaction partners urokinase plasminogen activator receptor and matrix metalloproteinase-13 during osteogenesis. *Lab Invest* 2001;81:1403-14.
- Pyke C, Ralfkiaer E, Huhtala P, Hurskainen T, Dano K, Tryggevason K. Localization of messenger RNA for Mr 72,000 and 92,000 type IV collagenases in human skin cancers by *in situ* hybridization. *Cancer Res* 1992;52:1336-41.
- Nielsen BS, Rank F, Lopez JM, Balbin M, Vizoso F, Lund LR, Dano K, Lopez-Otin C. Collagenase-3 expression in breast myofibroblasts as a molecular marker of transition of ductal carcinoma *in situ* lesions to invasive ductal carcinomas. *Cancer Res* 2001;61:7091-100.
- Holten-Andersen MN, Brunner N, Maimonis P, Jensen V, Murphy G, Piironen T. Characterization of monoclonal antibodies to tissue inhibitor of metalloproteinases-1. *Journal of clinical ligand assay* 2002;25:87-90.
- Shulman M, Wilde CD, Kohler G. A better cell line for making hybridomas secreting specific antibodies. *Nature* 1978;276:269-70.
- Hemby RM, Murphy G, Reynolds JJ. Immunolocalization of tissue inhibitor of metalloproteinases (TIMP) in human cells. Characterization and use of a specific antiserum. *J Cell Sci* 1985;73:105-19.
- Holten-Andersen MN, Murphy G, Nielsen HJ, Pedersen AN, Christensen JJ, Hoyer-Hansen G, Brunner N, Stephens RW. Quantitation of TIMP-1 in plasma of healthy blood donors and patients with advanced cancer. *Br J Cancer* 1999;80:495-503.
- Sappino AP, Dietrich PY, Skalli O, Widgren S, Gabbiani G. Colonic pericryptal fibroblasts. Differentiation pattern in embryogenesis and phenotypic modulation in epithelial proliferative lesions. *Virchows Arch A Pathol Anat Histopathol* 1989;415:551-7.
- Nielsen BS, Timshel S, Kjeldsen L, Sehested M, Pyke C, Borregaard N, Dano K. 92 kDa type IV collagenase (MMP-9) is expressed in neutrophils and macrophages but not in malignant epithelial cells in human colon cancer. *Int J Cancer* 1996;65:57-62.
- Holten-Andersen MN, Christensen JJ, Nielsen HJ, Stephens RW, Jensen V, Nielsen OH, Sorensen S, Overgaard J, Lilja H, Harris A, Murphy G, Brunner N. Total levels of tissue inhibitor of metalloproteinases 1 in plasma yield high diagnostic sensitivity and specificity in patients with colon cancer. *Clin Cancer Res* 2002;8:156-64.
- Kaye GI, Lane N, Pascal RR. Colonic pericryptal fibroblast sheath: replication, migration, and cytodifferentiation of a mesenchymal cell system in adult tissue. II. Fine structural aspects of normal rabbit and human colon. *Gastroenterology* 1968;54:852-65.
- Adegboyega PA, Mifflin RC, DiMari JF, Saada JJ, Powell DW. Immunohistochemical study of myofibroblasts in normal colonic mucosa, hyperplastic polyps, and adenomatous colorectal polyps. *Arch Pathol Lab Med* 2002;126:829-36.
- Poulsom R, Pignatelli M, Stetler-Stevenson WG, Liotta LA, Wright PA, Jeffery RE, Longcroft JM, Rogers L, Stamp GW. Stromal expression of 72 kDa type IV collagenase (MMP-2) and TIMP-2 mRNAs in colorectal neoplasia. *Am J Pathol* 1992;141:389-96.
- Rouyer N, Wolf C, Chénard MP, Rio MC, Chambou P, Bellocq JP, Basset P. Stromelysin-3 gene expression in human cancer: an overview. *Invasion Metastasis* 1994;14:269-75.

44. Okada A, Bellocq JP, Ronyer N, Chapard MP, Rio MC, Chambon P, Basset P. Membrane-type matrix metalloproteinase (MT-MMP) gene is expressed in stromal cells of human colon, breast, and head and neck carcinomas. *Proc Natl Acad Sci U S A* 1995;92:2730-4.
45. Nielsen BS, Sehested M, Dunn S, Rank F, Timshel S, Rygaard J, Johnsen M, Dano K. Urokinase plasminogen activator is localized in stromal cells in ductal breast cancer. *Lab Invest* 2001;81:1485-501.
46. Offersen BV, Nielsen BS, Høyer-Hansen G, Rank F, Hamilton-Dutoit S, Overgaard J, Andreasen PA. The myofibroblast is the predominant plasminogen activator inhibitor-1 expressing cell type in human breast carcinomas. *Am J Pathol* 2003;163:1887-99.
47. Duffy MJ, Reilly D, O'Sullivan C, O'Higgins N, Fennelly JJ, Andreasen P. Urokinase-plasminogen activator, a new and independent prognostic marker in breast cancer. *Cancer Res* 1990;50:6827-9.
48. Grøndahl HJ, Christensen JJ, Rosenquist C, Brunner N, Mouridsen HT, Dano K, Blichert TM. High levels of urokinase-type plasminogen activator and its inhibitor PAI-1 in cytosolic extracts of breast carcinomas are associated with poor prognosis. *Cancer Res* 1993;53:2513-21.
49. Illemann M, Hansen U, Nielsen HJ, Andreasen PA, Høyer-Hansen G, Lund LR, Dano K, Nielsen BS. Leading edge myofibroblasts in human colon cancer express PAI-1. *Am J Clin Pathol* 2004;122:256-65.
50. Saarialho-Kere UK, Vaalamo M, Puolakkainen P, Airola K, Parks WC, Karjalainen-Lindsberg ML. Enhanced expression of matrilysin, collagenase, and stromelysin-1 in gastrointestinal ulcers. *Am J Pathol* 1996;148:519-526.
51. McKaig BC, McWilliams D, Watson SA, Mahida YR. Expression and regulation of tissue inhibitor of metalloproteinase-1 and matrix metalloproteinases by intestinal myofibroblasts in inflammatory bowel disease. *Am J Pathol* 2003;162:1355-1360.

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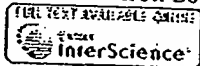
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Tissue plasminogen activator induced by dengue virus infection of human endothelial cells.

Huang YH, Lei HY, Liu HS, Lin YS, Chen SH, Liu CC, Yeh TM.

Department of Microbiology and Immunology, College of Medicine, National Cheng Kung University, Tainan, Taiwan, ROC.

Dengue hemorrhagic fever and dengue shock syndrome (DHF/DSS) are severe complications of dengue virus (DV) infection. However, the pathogenesis of hemorrhage induced by dengue virus infection is poorly understood. Since endothelial cells play a pivotal role in the regulation of hemostasis, we studied the effect of DV infection on the production of tissue plasminogen activator (tPA) and plasminogen activator inhibitor 1 (PAI-1) in vitro using both primary isolated endothelial cells, human umbilical cord veins cells, and a human microvascular endothelial cell line. DV infection significantly induced the secretion of tPA but not PAI-1 of human endothelial cells. In addition, tPA mRNA of endothelial cells was induced by DV as demonstrated by RT-PCR. Antibody against IL-6 but not control antibody inhibited DV-induced tPA production of endothelial cells. Furthermore, a good correlation between sera levels of IL-6 and tPA was found in DHF but not DF patients. These results suggest that IL-6 can regulate DV-induced tPA production of endothelial cells, which may play important roles in the pathogenic development of DHF/DSS. Copyright 2003 Wiley-Liss, Inc.

PMID: 12794725 [PubMed - indexed for MEDLINE]

Neu oncogene expression in ovarian tumors: a quantitative study.

Huettner PC, Carney WP, Naber SP, DeLellis RA, Membrino W, Wolfe HJ.

Department of Pathology, Tufts University School of Medicine, Massachusetts.

We studied neu mRNA expression by slot blot analysis and protein product expression by capture ELISA and immunohistochemistry in 57 primary and metastatic ovarian neoplasms, two paraovarian leiomyosarcomas, and eight normal ovaries. Some 61% of ovarian tumors but none of the paraovarian neoplasms or normal ovaries overexpressed neu mRNA. A total of 96% of the ovarian tumors that overexpressed neu were of epithelial type. Epithelial ovarian tumors had significantly higher amounts of the neu oncogene product as determined by capture ELISA than either germ cell and stromal tumors or normal ovaries (p less than 0.025). Different subtypes of ovarian carcinomas had significantly different amounts of neu oncogene product as measured by capture ELISA; endometrioid tumors had the highest, and poorly differentiated carcinomas not otherwise specified had the lowest (p less than 0.025). ELISA values, mRNA overexpression, and immunohistochemical staining intensity did not correlate with stage at diagnosis or architectural or nuclear grade in ovarian tumors. We conclude that capture ELISA is a simple, effective way to measure the neu oncogene protein product and that there is a good correlation between ELISA levels and immunohistochemical staining intensity. However, ELISA values did not correlate with stage or histologic prognostic factors in ovarian neoplasms.

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Real-time quantitative RT-PCR of cyclin D1 mRNA in mantle cell lymphoma: comparison with FISH and immunohistochemistry.

Hui P, Howe JG, Crouch J, Nimmakayalu M, Qumsiyeh MB, Tallini G, Flynn SD, Smith BR.

Department of Laboratory Medicine, Yale University School of Medicine, 333 Cedar Street, P.O. Box 208035, New Haven, CT 06520-8035, USA.

Presence of the balanced translocation t(11;14)(q13;q32) and the consequent overexpression of cyclin D1 found in mantle cell lymphoma (MCL) has been shown to be of important diagnostic value. Although many molecular and immunohistochemical approaches have been applied to analyze cyclin D1 status, correlative studies to compare different methods for the diagnosis of MCL are lacking. In this study, we examined 39 archived paraffin specimens from patients diagnosed with a variety of lymphoproliferative diseases including nine cases meeting morphologic and immunophenotypic criteria for MCL by: (1) real-time quantitative RT-PCR to evaluate cyclin D1 mRNA expression; (2) dual fluorescence in situ hybridization (FISH) to evaluate the t(11;14) translocation in interphase nuclei; and (3) tissue array immunohistochemistry to evaluate the cyclin D1 protein level. Among the nine cases of possible MCL, seven cases showed overexpression of cyclin D1 mRNA (cyclin D1 positive MCL) and two cases showed no cyclin D1 mRNA increase (cyclin D1 negative "MCL-like"). In six of seven cyclin D1 positive cases, the t(11;14) translocation was demonstrated by FISH analysis; in one case FISH was unsuccessful. Six of the seven cyclin D1 mRNA overexpressing cases showed increased cyclin D1 protein on tissue array immunohistochemistry; one was technically suboptimal. Among the two cyclin D1 negative MCL-like cases, FISH confirmed the absence of the t(11;14) translocation in both cases. All other lymphoproliferative diseases studied were found to have low or no cyclin D1 mRNA expression and were easily distinguishable from the cyclin D1 overexpressing MCLs by all three techniques. In addition, to confirming the need to assess cyclin D1 status, as well as, morphology and immunophenotyping to establish the diagnosis of MCL, this study demonstrates good correlation and comparability between measure of cyclin D1 mRNA, the 11;14 translocation and cyclin D1 protein.

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Elevation of topoisomerase I messenger RNA, protein, and catalytic activity in human tumors: demonstration of tumor-type specificity and implications for cancer chemotherapy.

Husain I, Mohler JL, Seigler HF, Besterman JM.

Department of Cell Biology, Glaxo Inc. Research Institute, Research Triangle Park, North Carolina 27709.

Topoisomerase I has been identified as an intracellular target of camptothecin, a plant alkaloid with anticancer activity. Various lines of evidence suggest that the sensitivity of cells to this drug is directly related to the topoisomerase I content. In humans, the levels of topoisomerase I have been shown to be elevated in colorectal tumors, compared to normal colon mucosa. The aim of our study was to determine whether (a) topoisomerase I levels are elevated in other solid tumors, (b) the elevated enzyme is catalytically active in these tumors, and (c) the increase in topoisomerase I levels in colorectal tumors is a result of increased transcription or translation. Topoisomerase I levels were quantitated in crude extracts from colorectal, prostate, and kidney tumors and their matched normal counterparts by Western blotting and by direct determination of catalytic activity, and mRNA levels were determined by Northern blotting. By Western blotting, colorectal tumors showed 5-35-fold increases in topoisomerase I levels, compared to their normal colon mucosa. In the case of prostate tumors, the increase was 2-10-fold, compared with benign hyperplastic prostate tissue from the same patients. However, no difference was observed in topoisomerase I levels in kidney tumors, compared to their normal counterparts. The catalytic activity of topoisomerase I was determined by a quantitative ³²P-transfer assay in crude homogenates, without isolating nuclei. Colorectal and prostate tumors exhibited 11-40- and 4-26-fold increases, respectively, in catalytic activity. However, kidney tumors did not show any alteration in catalytic activity, compared to their normal matched samples. Thus, for all three tumor types there was a good correlation between enzyme levels and catalytic activity. Finally, colorectal tumors were analyzed for steady state mRNA levels. A 2-33-fold increase in mRNA levels was found in colorectal tumors, compared to normal colon mucosa. These results suggest that alterations in topoisomerase I expression in humans are tumor type specific and that the increase in topoisomerase I levels results from either increased transcription of the topoisomerase I gene or increased mRNA stability.

PMID: 8275492 [PubMed - indexed for MEDLINE]

High-level mRNA quantification of proliferation marker pKi-67 is correlated with favorable prognosis in colorectal carcinoma.

Ihmann T, Liu J, Schwabe W, Hausler P, Behnke D, Bruch HP, Broll R, Windhovel U, Duchrow M.

St. Elisabeth Klinik, Klinik für Anesthesiologie, Schmerztherapie und Intensivmedizin, Saarlouis, Germany.

PURPOSE: The present study retrospectively examines the expression of pKi-67 mRNA and protein in colorectal carcinoma and their correlation to the outcome of patients. **METHODS:** Immunohistochemistry and quantitative RT-PCR were used to analyze the expression of pKi-67 in 43 archival specimens of patients with curatively resected primary colorectal carcinoma, who were not treated with neo-adjuvant therapy. **RESULTS:** We determined a median pKi-67 (MIB-1) labeling index of 31.3% (range 10.3-66.4%), and a mean mRNA level of 0.1769 (DeltaC(T): range 0.01-0.69); indices and levels did not correlate. High pKi-67 mRNA DeltaC(T) values were associated with a significantly favorable prognosis, while pKi-67 labeling indices were not correlated to prognostic outcome. A multivariate analysis of clinical and biological factors indicated that tumor stage (UICC) and pKi-67 mRNA expression level were independent prognostic factors. **CONCLUSION:** Quantitatively determined pKi-67 mRNA can be a good and new prognostic indicator for primary resected colorectal carcinoma.

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- Evaluation Studies

PMID: 15449182 [PubMed - indexed for MEDLINE]



Modulation of glucagon receptor expression and response in transfected human embryonic kidney cells.

Ikegami T, Cypess AM, Bouscarel B.

Department of Medicine, George Washington University Medical Center, Washington, District of Columbia 20037, USA.

The modulation of glucagon receptor (GR) expression and biological response was investigated in human embryonic kidney cell (HEK-293) clones permanently expressing the GR with different densities. The GR mRNA expression level in these clones was upregulated by cellular cAMP accumulation and presented a good correlation with both the protein expression level and the maximum number of glucagon binding sites. However, the determination of glucagon-induced cAMP accumulation in these cell lines revealed that the enhancement of receptor expression did not lead to a proportional increase in cAMP formation. Under these conditions, the maximum cAMP production induced by NaF and forskolin was not significantly different among selected clones, regardless of the receptor expression level. High receptor-expressing clones showed the greatest susceptibility for agonist-induced desensitization compared with clones with lower GR expression levels. The results of the present study suggest that the GR can recruit non-GR-specific desensitization mechanism(s). Furthermore, the partial inhibition or alteration of the overall cAMP synthesis pathway at the receptor level may be a necessary adaptive step for a cell in response to a massive increase in membrane receptor expression level.

PMID: 11546678 [PubMed - indexed for MEDLINE]

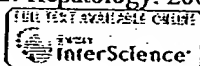
Developmental regulation of acidic fibroblast growth factor (aFGF) expression in bovine retina.

Jacquemin E, Jonet L, Oliver L, Bugra K, Laurent M, Courtois Y, Jeanny JC.

Unite de Recherches Gerontologiques, U. 118 INSERM, Paris, France.

Acidic fibroblast growth factor (aFGF) is a signalling molecule implicated in a wide variety of biological processes such as cell growth, differentiation and survival. It has been purified from bovine retina. The present study was carried out to detect which cells in the bovine retina expressed aFGF at the different stages of embryonic and post-natal development. The specific aFGF mRNA and protein were detected by in situ hybridization employing riboprobes and immunocytochemistry using affinity purified polyclonal human recombinant aFGF antibodies respectively. No signal was detected by either technique until 4-5 months and then there was progressive expression of aFGF with terminal morphogenesis of the retina. By 8-9 months of embryonic development, nuclei of the 3 neuronal layers (ganglion cell layer, inner and outer nuclear layers) were all uniformly and intensely labeled. A slight labeling of the pigmented epithelium of the retina was also visible throughout development and maturation. These results showed a good correlation between message and protein expression in these cell types. In contrast, glial cells in the nerve fiber layer and vascular endothelial cells displayed a nuclear immunostaining for the protein in the absence of message. These data suggest that aFGF plays a role in the late steps of retinal differentiation by autocrine and paracrine mechanisms.

PMID: 7507349 [PubMed - indexed for MEDLINE]



The p21(Cip1) protein, a cyclin inhibitor, regulates the levels and the intracellular localization of CDC25A in mice regenerating livers.

Jaime M, Pujol MJ, Serratos J, Pantoja C, Canela N, Casanovas O, Serrano M, Agell N, Bachs O.

Department of Cell Biology and Pathology, Faculty of Medicine, Institut d'Investigacions Biomediques August Pi Sunyer (IDIBAPS), University of Barcelona, Barcelona, Spain.

Liver cells from p21(Cip1^{-/-}) mice subjected to partial hepatectomy (PH) progress into DNA synthesis faster than those from wild-type mice. These cells also show a premature induction of cyclin E/cyclin-dependent kinase (CDK) 2 activity. We studied the mechanisms whereby cells lacking p21(Cip1) showed a premature induction of this activity. Whereas the levels of CDK2, cyclin E, and p27(Kip1) were similar in both wild-type and p21(Cip1^{-/-}) mice, those of the activator CDC25A were much higher in p21(Cip1^{-/-}) quiescent and regenerating livers than in wild-type animals. Moreover, p21(Cip1^{-/-}) cells also showed a premature translocation of CDC25A from cytoplasm into the nucleus. The ectopic expression of p21(Cip1) into mice embryo fibroblasts from p21(Cip1^{-/-}) mice decreased the levels of CDC25A and delayed its nuclear translocation. The levels of CDC25A messenger RNA in p21(Cip1^{-/-}) cells were higher than in wild-type cells, suggesting that this increase might be responsible, at least in part, for the high levels of CDC25A protein in these cells. Thus, the results reported here indicate that p21(Cip1) regulates the levels and the intracellular localization of CDC25A. We also found a good correlation between CDC25A nuclear translocation and cyclin E/CDK2 activation. In conclusion, premature translocation of CDC25A to the nucleus might be involved in the advanced induction of cyclin E/CDK2 activity and DNA replication in cells from animals lacking p21(Cip1).

PMID: 11981756 [PubMed - indexed for MEDLINE]



Alteration of frizzled expression in renal cell carcinoma.

Janssens N, Andries L, Janicot M, Perera T, Bakker A.

Department of Biochemistry, University of Antwerp, Wilrijk, Belgium.
njansse9@prdbe.juj.com

To evaluate the involvement of frizzled receptors (Fzds) in oncogenesis, we investigated mRNA expression levels of several human Fzds in more than 30 different human tumor samples and their corresponding (matched) normal tissue samples, using real-time quantitative PCR. We observed that the mRNA level of Fzd5 was markedly increased in 8 of 11 renal carcinoma samples whilst Fzd8 mRNA was increased in 7 of 11 renal carcinoma samples. Western blot analysis of crude membrane fractions revealed that Fzd5 protein expression in the matched tumor/normal kidney samples correlated with the observed mRNA level. Wnt/beta-catenin signaling pathway activation was confirmed by the increased expression of a set of target genes. Using a kidney tumor tissue array, Fzd5 protein expression was investigated in a broader panel of kidney tumor samples. Fzd5 membrane staining was detected in 30% of clear cell carcinomas, and there was a strong correlation with nuclear cyclin D1 staining in the samples. Our data suggested that altered expression of certain members of the Fzd family, and their downstream targets, could provide alternative mechanisms leading to activation of the Wnt signaling pathway in renal carcinogenesis. Fzd family members may have a role as a biomarker.

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Alteration of Frizzled Expression in Renal Cell Carcinoma

Nico Janssens^{a,c} Luc Andries^b Michel Janicot^c Tim Perera^c
Annette Bakker^c

^aDepartment of Biochemistry, University of Antwerp, Wilrijk, ^bHistoGeneX, Edegem, and ^cOncology Discovery Research, Johnson & Johnson Pharmaceutical Research and Development, Beerse, Belgium

Key Words

β -Catenin · Cyclin D1 · Frizzled receptor · Renal cell carcinoma · Wnt

Abstract

To evaluate the involvement of frizzled receptors (Fzds) in oncogenesis, we investigated mRNA expression levels of several human Fzds in more than 30 different human tumor samples and their corresponding (matched) normal tissue samples, using real-time quantitative PCR. We observed that the mRNA level of Fzd5 was markedly increased in 8 of 11 renal carcinoma samples whilst Fzd8 mRNA was increased in 7 of 11 renal carcinoma samples. Western blot analysis of crude membrane fractions revealed that Fzd5 protein expression in the matched tumor/normal kidney samples correlated with the observed mRNA level. Wnt/ β -catenin signaling pathway activation was confirmed by the increased expression of a set of target genes. Using a kidney tumor tissue array, Fzd5 protein expression was investigated in a broader panel of kidney tumor samples. Fzd5 membrane staining was detected in 30% of clear cell carcinomas, and there was a strong correlation with nuclear cyclin D1 staining in the samples. Our data suggested that altered expression of certain members of the

Fzd family, and their downstream targets, could provide alternative mechanisms leading to activation of the Wnt signaling pathway in renal carcinogenesis. Fzd family members may have a role as a biomarker.

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Introduction

The Wnt signaling pathway is evolutionary conserved and controls many events during embryonic development. Members of the Wnt gene family of secreted glycoproteins are involved in embryonic induction, generation of cell polarity, cell proliferation and the determination of cell fate [1, 2]. Recently, it has become evident that the Wnt pathway is also deregulated in a range of tumors [3].

The Wnt signaling pathway is activated when Wnt proteins bind to a cell surface receptor complex consisting of a member of the frizzled receptor (Fzd) family and either low-density-lipoprotein receptor-related protein (LRP)5 or LRP6 [4, 5]. A detailed characterization of the Fzds and the immediate downstream events after Wnt binding has been hampered by the lack of pure biologically active Wnts.

Downstream of the receptor complex, three pathways may be initiated, depending on the composition of the

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Nico Janssens
Oncology Discovery Research, JAJPRD
Turnhoutseweg 30
BE-2340 Beerse (Belgium)
Tel. +32 14 603831, Fax +32 14 605403, E-Mail njansse@prdbe.jnj.com

ligand and receptor complex. The 'Wnt/ β -catenin pathway', the 'Wnt/ Ca^{2+} pathway' or the 'Wnt polarity pathway' [6]. The Wnt/ β -catenin pathway has been linked to carcinogenesis. Genetic alterations in components of this pathway (adenomatous polyposis coli, APC, axin and β -catenin) can result in the accumulation of non-phosphorylated β -catenin [3, 7] and this can promote carcinogenesis. Conversely, neither the Wnt/ Ca^{2+} pathway nor the Wnt polarity pathway involves the activation of β -catenin [for review, see ref. 1, 6].

Mutations in one of the three regulatory genes (APC, β -catenin and axin), overexpression of Wnts and Fzds or the expression of a constitutively active Fzd have been linked to Wnt/ β -catenin pathway activation in various tumors [8, 9].

To evaluate the involvement of Fzds in oncogenesis, we investigated mRNA expression levels of several human Fzds (Fzd2, 3, 5, 6, 7, 8 and 9) in more than 30 different human tumor samples using real-time quantitative PCR. Each sample was compared with its corresponding (matched) normal tissue sample. The most striking observation was the dramatically increased Fzd5 and Fzd8 mRNA expression seen in the renal carcinoma samples. This was confirmed at the protein level using Western blotting. Kidney tumor tissue arrays confirmed Fzd5 membrane staining in 30% of clear cell carcinomas, with nuclear cyclin D1 showing a strong correlation with the Fzd5 membrane labeling. Fzd8 protein expression analysis was not performed due to the lack of suitable reagents. These data suggest that Fzd5 may have a role in renal cell carcinogenesis due to its frequent overexpression observed in these tumor samples. Potential future applications could include uses in tumor targeting or as a potential biomarker.

Materials and Methods

Tissue Samples

Frozen tumor tissue samples with corresponding normal tissue from the same patient were derived either from human biopsy or autopsy material (Department of Pathology, University of Antwerp, kindly provided by Prof. E. Van Marck). Tissue specimens were snap-frozen in liquid nitrogen and kept at -80°C until use. Frozen sections of kidney tumor and normal tissue samples were stained with hematoxylin-eosin to support the pathologist's observations and to confirm the type of kidney tumor. Paraffin-embedded tissue slides of renal carcinoma, lung carcinoma, breast and colon carcinoma were obtained, after encryption, from the Department of Pathology (Middelheim Hospital, Antwerp, Belgium). The CL1 human kidney cancer (SuperBioChips Laboratories) tissue array used in this study contained 59 tissue samples consisting of 9 normal kidney tissues.

30 clear cell renal carcinoma samples and another 20 renal cell tumor types (chromophil, chromophobe, papillary type, collecting duct carcinoma and samples with mixed types).

RNA Isolation and Reverse Transcription

Total RNA was extracted from tissue specimens using Ultraspec Reagent (Biotecx, USA) according to the manufacturer's instructions. All total RNA was routinely treated with DNase (DNA-free kit, Ambion, USA). 1 μg of total RNA was used to synthesize cDNA using oligo-dT primers (Superscript; Invitrogen, Merelbeke, Belgium). Reverse transcription was performed at 42°C for 60 min, followed by 70°C for 10 min.

Real-Time PCR

Real-time PCR was performed on either an ABI Prism 7700 or 7900 Sequence detection system (Perkin-Elmer Applied Biosystems, Foster City, Calif., USA) using the 5' nuclease assay (TaqmanTM). Primer and probe sequences were designed using Primer Express (PE Applied Biosystems) and are shown in table 1. Quantitative values were obtained from the threshold cycle number (Ct) at which the increase in the signal associated with exponential growth of PCR products is detected using PE Biosystems analysis software, according to the manufacturer's instructions.

We have used the $2^{-\Delta\Delta\text{Ct}}$ method to analyze the relative changes in gene expression of the different genes between tumor and corresponding normal tissue samples. We used the mitochondrial ATP synthase 6 (ATP6) as the endogenous RNA control [10; Janssens et al., in prep.], and each sample was normalized to its ATP6 content. The relative expression of the target gene was also normalized to the corresponding normal tissue sample (calibrator). Results, expressed as the amount of target sample relative to the ATP6 gene and the calibrator, were determined as follows: $N = 2^{-(\Delta\text{Ct}_{\text{sample}} - \Delta\text{Ct}_{\text{calibrator}})}$, where the ΔCt values of the sample and calibrator were determined by subtracting the average Ct value of the sample and the calibrator from the average Ct value of the ATP6 gene. Amplification was done essentially as described previously [10]. Briefly, 50 μl of reaction mixture containing 1 μl of cDNA template were amplified as follows: incubation at 50°C for 2 min, denaturation at 95°C for 10 min, and 50 cycles at 95°C for 15 s and 60°C for 1 min.

Membrane Preparation, Gel Electrophoresis and Immunoblotting

Tissue samples were weighed, suspended at a 40 times dilution [$= 40$ volumes/original wet weight of tissue (v/w)] in 50 mM Tris-HCl buffer, pH 7.4, and homogenized with an Ultra-Turrax homogenizer. After centrifugation for 10 min, 24,000 g at 4°C , the pellet was washed three times by resuspension in the Tris-HCl buffer followed by centrifugation. The final membrane pellets were stored at -80°C in the Tris-HCl buffer at a concentration of 0.5–1 mg/ml. The Bradford protein assay (Pierce, Aalst, Belgium) was used for protein determination. Proteins (50 μg) were separated by 8% SDS-PAGE and transferred to nitrocellulose membranes. After primary and secondary antibody incubation, the antigen-antibody-peroxidase complex was detected by chemiluminescence (Pierce, Aalst, Belgium) according to the manufacturer's instructions.

Immunohistochemistry

Immunohistochemistry was performed on 10- μm -thick cryosections of unfixed tumor tissue and on 6- μm -thick paraffin sections from renal tumor tissue fixed by formalin or by an alcohol-based fixative. Adjacent tissue blocks from renal tumors were processed with

Table 1. Real-time PCR primer and probe sequences

| Target cDNA | Primer/probe sequences ^a | Fragment position ^b | Accession No. ^c |
|---------------|--|--------------------------------|----------------------------|
| FZD2 | (a) 5'-atcccggtcccgcc-3' (b) 5'-gtattgacatgagacgtgaagtc-3' (c) 5'-FAM-tacacgccgcgcagtcgc-TAMRA-3' | 1,548-1,613 | AB017364 |
| FZD3 | (a) 5'-tcacgccagtcagtcgg-3' (b) 5'-ttgtcaccttcaattttatcatcg-3' (c) 5'-FAM-catecccggaactctaacatcatccttt-TAMRA-3' | 1,473-1,547 | AB039723 |
| FZD5 | (a) 5'-tgcacaggtcacttccgtt-3' (b) 5'-tctcgaagtcgctcg-3' (c) 5'-FAM-ccttcatgggtgtgtccccc-TAMRA-3' | 2,143-2,204 | HSU43318 |
| FZD6 | (a) 5'-ctagcaccctcaggttaagaga-3' (b) 5'-ctcagagagtcgtggaaggat-3' (c) 5'-FAM-tgtgtgaacctgctcgcag-TAMRA-3' | 2,094-2,170 | AF072873 |
| FZD7 | (a) 5'-ccttggaaaggcataactgtg-3' (b) 5'-aaccaacgggaacccaga-3' (c) 5'-FAM-aagcaactttataggcaagcagcgca-TAMRA-3' | 2,687-2,762 | AB017365 |
| FZD8 | (a) 5'-tgtgtcgggtgcttctgtt-3' (b) 5'-cgctccatgtcgataaggag-3' (c) 5'-FAM-ccaccttgcacccgtctcca-TAMRA-3' | 853-919 | AB043703 |
| FZD9 | (a) 5'-ccccggagctacggac-3' (b) 5'-tagtcatgtgcaagaccacgg-3' (c) 5'-FAM-tggcgcgcacttgcactataaggct-TAMRA-3' | 1,696-1,763 | HSU82169 |
| ATP5b | (a) 5'-gggtagggtggttgggt-3' (b) 5'-gggcgcagtgattataggctt-3' (c) 5'-FAM-aagtggtctagggtcatttttaatttagagcg-TAMRA-3' | 580-503 | AF368271 |
| c-myc | (a) 5'-accacagcagcgcctga-3' (b) 5'-tccagcagaaggatccagact-3' (c) 5'-FAM-accctttgcaggagcttgcctct-TAMRA-3' | 1,297-1,413 | HSMYC1 |
| Cyclin D1 | (a) 5'-gaacctggccgaatgac-3' (b) 5'-cgctcttggcatttggga-3' (c) 5'-FAM-cgcacgalltcattgaacatt-TAMRA-3' | 4,148-4,211 | AF511593 |
| PPAR δ | (a) 5'-agcatcttaccggcaaa-3' (b) 5'-gtctgatgtcgggaicaca-3' (c) 5'-FAM-ccagccacacggccct-TAMRA-3' | 932-990 | NM-006238 |

^a (a) = Sense primer; (b) = antisense primer; (c) = probe.

^b Fragment positions are given according to the EMBL/GenBank accession No. of cloned sequence.

^c EMBL/GenBank accession No. of cloned sequence.

formalin and with the alcohol-based fixative. Paraffin and cryosections were mounted on poly-L-lysine or 3-aminopropyltriethoxysilane-gelatin-coated slides. The 59 tissue samples on the CLI human renal cancer tissue array slides were all fixed with formalin and embedded in paraffin, and the sections were mounted on silane-coated slides (SuperBioChips Laboratories). In addition to renal carcinoma tissue, sections from 10 formalin-fixed paraffin-embedded lung carcinomas were stained for Fzd5. Colon and breast tumors

were used as positive controls for β -catenin and cyclin D1 immunostaining.

The following primary antibodies were used: Fzd5 (Upstate Biotechnology), β -catenin (Zymed), cyclin D1 (Zymed), E-cadherin (Novocastra) and cytokeratin 8 (Biogenex). Cryosections were fixed in 4% paraformaldehyde for 5 min; acetone for 5 min at -20°C and 70% ethanol for 5 min. Endogenous peroxidase activity was quenched using 3% H_2O_2 . Paraffin sections of formalin- and alcohol-

Table 2. Fzd mRNA expression in tumor samples

| Sample ^a | Tissue | Tumor type | x-fold expression increase ^b | | | | | | |
|---------------------|--------------|----------------------------|---|-------|-------|------|-------|-------|-------|
| | | | FZD2 | FZD3 | FZD5 | FZD6 | FZD7 | FZD8 | FZD9 |
| 133702 | kidney | adenocarcinoma | 0.17 | 0.5 | 3.72 | 2.13 | 0.06 | 1.32 | - |
| 137770 | kidney | renal cell carcinoma | 1.23 | 3.56 | 8.26 | 2.61 | 1.3 | 8.21 | 5.44 |
| 138844 | kidney | renal cell carcinoma | 0.31 | 0.11 | 6.84 | 1.18 | 0.23 | 3.18 | 2.8 |
| 137146 | kidney | renal cell carcinoma | 0.47 | 0.45 | 3.16 | 2.23 | 0.73 | 4.42 | 1.37 |
| 137564 | kidney | renal cell carcinoma | 3.43 | 2.95 | 9.6 | 1.57 | 7.64 | 3.52 | - |
| 133408 | kidney | renal cell carcinoma | 23.97 | 0.98 | 6.39 | 2.48 | 5.05 | 16.72 | - |
| 139188 | kidney | renal cell carcinoma | 3.7 | 0.56 | 0.66 | 1.33 | 0.37 | 4.41 | 1.53 |
| 135699 | kidney | renal cell carcinoma | 2.6 | 0.36 | 4.83 | 0.9 | 0.34 | 2.54 | 2.31 |
| 139064 | kidney | renal cell carcinoma | 5.16 | 1.82 | 1.25 | 2.36 | 1.8 | 2.19 | 2.85 |
| 134585 | kidney | renal cell carcinoma | 1.47 | 0.72 | 1.38 | 0.47 | 0.09 | 0.6 | - |
| 140279 | kidney | renal cell carcinoma | 7.33 | 18.17 | 3.93 | 6.05 | 6.41 | 4.65 | 7.91 |
| 137252 | ovary | carcinosarcoma | 0.4 | 3.39 | 0.49 | 0.44 | 0.92 | 0.54 | 0.53 |
| 138256 | ovary | papillary carcinoma | 0.7 | 5.17 | 1.22 | 2.19 | 0.4 | 0.11 | - |
| 146472 | ovary | serous papillary carcinoma | 0.39 | 3.29 | 2.56 | 1.34 | 3.94 | 0.67 | 0.59 |
| 145845 | colon | adenocarcinoma | 3.45 | 3.36 | 0.58 | 1.22 | 1.67 | 1.24 | 1.99 |
| 146145 | colon | adenocarcinoma | 5.46 | 6.74 | 4.42 | 6.57 | 3.36 | 6.08 | 0.15 |
| 146630 | colon | adenocarcinoma | 4.01 | 4.73 | 0.3 | 1.16 | 0.54 | 0.55 | 0.12 |
| 146633 | colon | adenocarcinoma | 1.87 | 1.07 | 1.62 | 1.47 | 0.51 | 2.07 | - |
| 147055 | colon | adenocarcinoma | 0.66 | 1.41 | 1.01 | 1.33 | 0.24 | 0.79 | 4.87 |
| 142253 | lung | adenocarcinoma | 0.67 | 5.17 | 0.43 | 1.87 | 0.33 | 0.59 | 0.41 |
| 143036 | lung | adenocarcinoma | 0.67 | 1.24 | 2.63 | 1.13 | 1.11 | 1.04 | 9.19 |
| 138938 | lung | adenocarcinoma | 0.93 | 0.88 | 1.07 | 1.3 | 1.73 | 0.38 | 0.76 |
| 133563 | lung | adenocarcinoma | 2.76 | 1.23 | 0.31 | 1.34 | 0.99 | 0.78 | 4.97 |
| 144387 | lung | adenocarcinoma | 12.85 | 0.43 | 0.49 | 0.25 | 2.73 | 0.54 | - |
| 137304 | lung | acinary adenocarcinoma | 0.54 | 9.15 | 1.65 | 1.25 | 0.63 | 3.41 | 2.64 |
| 144546 | lung | epithelial carcinoma | 0.1 | 0.67 | 0.21 | 8.11 | 0.27 | 0.97 | 1.32 |
| 137621 | lung | epithelial carcinoma | 1.52 | 2.19 | 0.37 | 2.26 | 0.52 | 0.45 | 1.06 |
| 145552 | lung | epithelial carcinoma | 1.09 | - | 0.16 | 0.91 | 1.47 | 0.13 | 0.5 |
| 143987 | testis | embryonal carcinoma | 0.24 | 0.6 | 19.43 | 1.33 | 23.12 | 2.43 | 0.33 |
| 137332 | stomach | leiomyoma | 19.32 | 3.91 | 0.45 | 1.53 | 18.98 | 32.82 | 16.7 |
| 139026 | stroma | gastrointestinal carcinoma | 66.1 | 3.51 | 0.03 | 6.67 | 8.44 | 2.28 | 20.82 |
| 136049 | rectum | adenocarcinoma | 1.71 | 0.52 | 0.4 | 0.54 | 0.29 | 0.88 | 0.28 |
| 140794 | gall bladder | adenosquamous carcinoma | 0.19 | - | 0.93 | 3.02 | 0.2 | - | - |

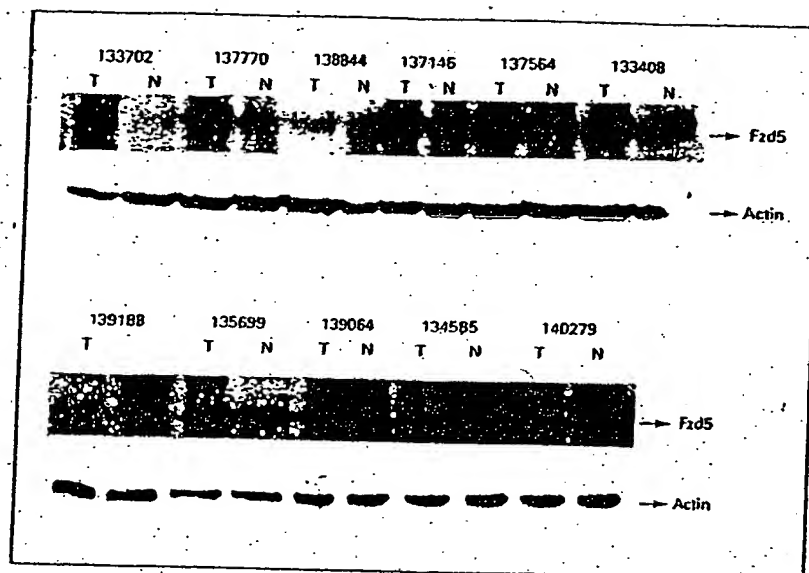
^a Sample identification numbers were given by the pathologist.

^b Results are expressed as x-fold increase of the gene in the tumor tissue sample compared to its matched normal tissue sample after normalizing both samples on the basis of their ATP5b content. A cutoff of 3-fold was used to define differential expression. Significant (> 3-fold) increases in the expression level of the Fzd receptor are shown in italics. - = Expression of the target gene undetectable in one or both samples (tumor and/or normal).

fixed tissue were processed with a trypsin-tritrate-microwave pretreatment or with an EDTA-microwave pretreatment to unmask epitopes, respectively. Sections were then sequentially processed with primary antibodies, biotinylated secondary antibodies and streptavidin-biotin-peroxidase (Fzd5, E-cadherin and cytokeratin 8). For β -catenin, polyclonal rabbit antibody with the EnVision detection system (DAKO) was used. The slides were further developed using 3-amino-9-ethylcarbazole, counterstained with hemalaun and mount-

ed with glycerin gelatin. Stained sections were observed with an Axioplan 2 microscope equipped with an Axiocam digital camera. Staining intensity for β -catenin was scored as no staining (value 0), weak and fragmentary staining of cell membranes (value 1), moderate membrane staining of less than 50% of the tumor cells (value 2), moderate membrane staining of more than 50% of tumor cells (value 3) and strong membrane staining of more than 75% of tumor cells (value 4). The cyclin D1 staining was quantified as a percentage of

Fig. 1. Fzd5 protein expression in matched tumor/normal kidney samples. T = Tumor sample; N = matched normal sample. Sample identification numbers are given by the pathologist.



cyclin D1-immunoreactive nuclei in tumor cells in three fields (area: 18,641 μm^2) of each tumor sample. The total number of tumor nuclei ranged from 51 to 164. The correlation between Fzd5 and β -catenin staining, and between Fzd5 and cyclin D1 staining was evaluated by the Mann-Whitney U test.

Results

Fzd mRNA Expression in Matched Human Tumor/Normal Tissue Samples

Fzd expression in tumor tissue was compared with Fzd expression in matched normal tissue samples and normalized to the expression of the housekeeping gene mitochondrial ATP5b (table 2). A 3-fold increase was considered significant.

In the kidney tumor samples, in which 10 of 11 samples were clear cell carcinomas, Fzd5 was upregulated in 8 of the 11 samples. A similar observation was made for Fzd8 and Fzd2, which were upregulated in 7 and 5 renal tumor samples, respectively. None of the other Fzds showed consistent upregulation.

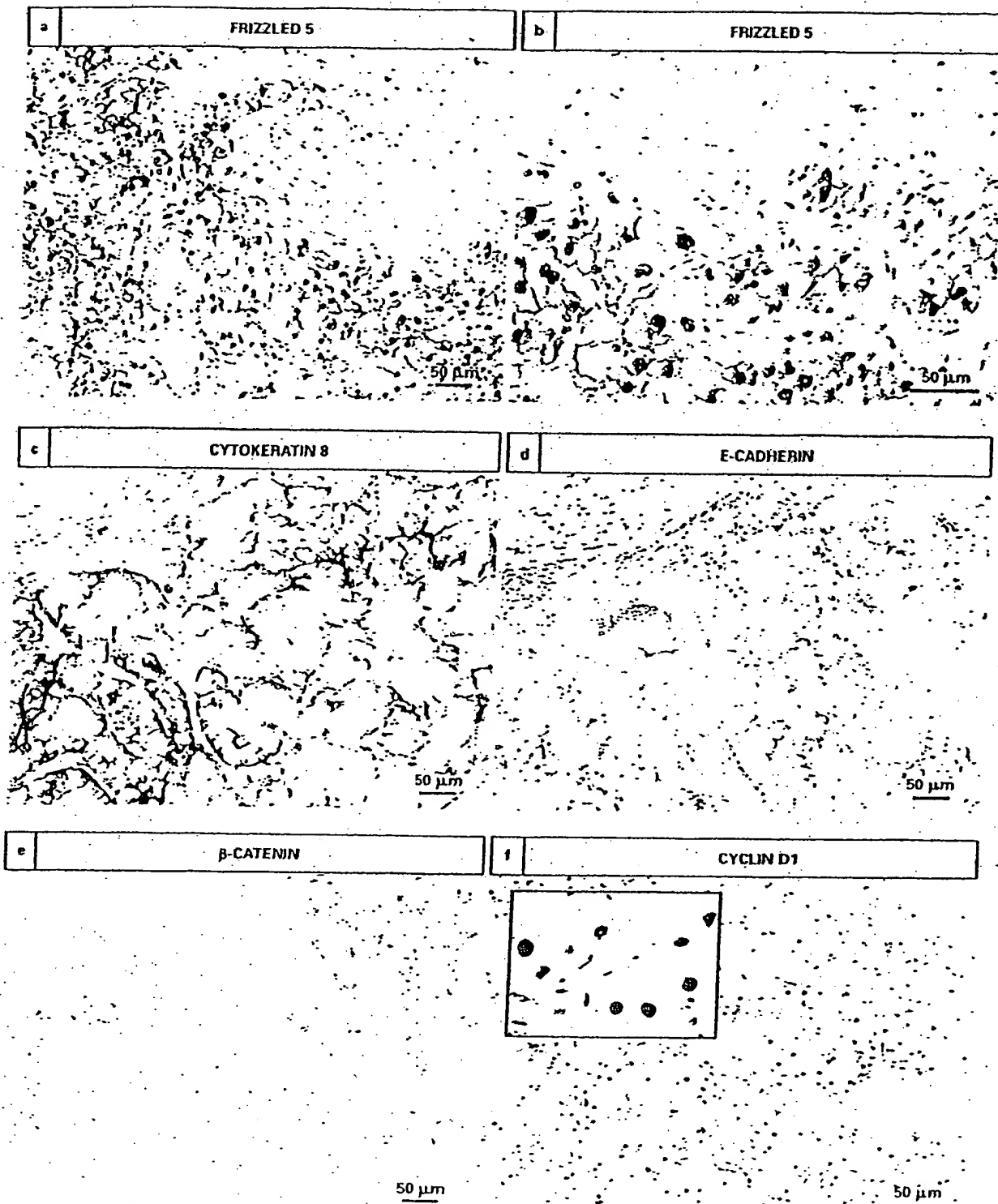
Both Fzd2 and Fzd3 were upregulated in 3 of 5 colon adenocarcinoma samples. No other Fzd expression was significantly different compared to the normal colon tissue sample. Fzd3 showed an increased expression in all 3 ovarian carcinoma samples. Fzd expression was not altered in any of the lung tumor samples. The Fzd expres-

sion level was observed to be relatively low in these lung tissues compared to the other tissues investigated.

Western Blot and Immunohistochemistry Analysis on Renal Carcinomas

Western blotting was used to evaluate Fzd5 protein expression in the renal tissue samples used for mRNA expression analysis. Membrane fractions of the renal carcinoma and corresponding normal tissue samples were prepared. As previously shown (table 2), Fzd5 mRNA upregulation was detected in 8 of the 11 matched tumor/normal samples. Increased expression of Fzd5 protein was seen in membrane fractions from 9 of 11 samples (fig. 1). In most cases, concomitant increases in Fzd5 mRNA and protein levels were observed.

Hematoxylin-eosin staining of the cryosectioned tumors confirmed the presence of clear cell carcinoma. Fzd5 immunostaining in clear cell carcinoma (fig. 2a, b) was observed to be localized to cell membranes and to nuclei. Cytokeratin 8 (fig. 2c) and E-cadherin (fig. 2d) were also detected. E-cadherin labeling of cell membranes in clear cell carcinoma was less intense and patchy compared to epithelial cells of normal renal tissue. β -Catenin staining was confined to the cell membrane. β -Catenin levels in the clear cell carcinoma membranes were highly variable. Nuclear β -catenin staining was not observed in any of the samples. Epithelial cells in normal renal tissue showed intense membrane staining and some cytoplasmic



staining. In addition, weak β -catenin staining of endothelial cells was observed. A high number of cyclin D1-immunoreactive nuclei was observed in clear cell carcinoma (fig. 2f).

On the CL1 human kidney cancer tissue array, 30% ($n = 9$) of the clear cell carcinoma tumor samples ($n = 30$) showed Fzd5 immunoreactivity (fig. 3a). Membrane-associated β -catenin staining was observed in 33% of the Fzd5-positive tumor samples and 57% of Fzd5-negative clear renal cell carcinoma samples (table 3; fig. 3c, d). Again, nuclear β -catenin staining was never observed. Statistical analysis did not reveal a difference in the expression of β -catenin between Fzd5-positive and Fzd5-negative tumor samples (fig. 4a).

Nuclear cyclin D1 was observed in 89% of the Fzd5-positive clear cell carcinoma samples (table 3; fig. 3e). Only 38% of the Fzd5-negative clear cell carcinoma samples contained nuclear cyclin D1. Statistical analysis showed a significantly higher cyclin D1 expression in Fzd5-positive compared to Fzd5-negative tumor samples (fig. 4b).

c-myc, Cyclin D1 and Peroxisome Proliferator-Activated Receptor δ Expression in Renal Carcinomas

Wnt/ β -catenin pathway activation in the kidney tissue samples was investigated looking at the expression of a number of target genes, which have previously been shown to be upregulated when the pathway is active. Gene expression of *c-myc*, cyclin D1 and peroxisome proliferator-activated receptor δ (PPAR δ) was analyzed. Increased expression of both *c-myc* and cyclin D1 genes have been implicated in cell proliferation, and carcinogenesis, and they represent two of the more important and closely studied target genes of the Wnt signaling pathway.

Fig. 2. Distribution of Fzd5 (a, b), cytokeratin 8 (c), E-cadherin (d) and β -catenin (e) immunoreactivity in paraffin sections from a renal tumor processed by an alcohol fixative. From the same tumor, a formalin-fixed block was used for cyclin D1 immunostaining (f). Fzd5 immunostaining shows distinct immunoreactivity in cell membranes and in nuclei of clear cell renal carcinoma. Clear cells are immunoreactive for cytokeratin 8. β -Catenin and E-cadherin staining of membranes is rather weak, and not uniform, in clear cell renal carcinoma. Nuclear β -catenin immunoreactivity was not observed. In clear cell renal carcinoma, many nuclei showed cyclin D1 immunoreactivity. The inset in f shows a detailed view of the cyclin D1 labeling of nuclei in clear cell renal carcinoma.

Expression of PPAR δ was investigated because it represents a direct target of the β -catenin pathway with T cell factor binding sites in its promoter. Expression of *c-myc* was found to be upregulated in 7 of 11, whilst cyclin D1 was upregulated in 10 of 11 kidney tumor samples (table 4). PPAR δ was upregulated in 9 cases. All three selected target genes showed a marked upregulation in the majority of renal tumors, which suggested that the Wnt/ β -catenin pathway was activated in these samples.

Discussion

Fzd family member overexpression has been postulated to play key roles in different tumor types such as esophageal carcinoma [11], gastric cancer [12] and head and neck squamous cell carcinoma [13]. The current study evaluated the potential implication of Fzds as tumor-associated antigens in different tumor types. We screened a number of matched normal/tumor tissue samples for the expression of a variety of Fzds using real-time quantitative PCR.

Results obtained revealed that both Fzd5 and Fzd8 mRNA were overexpressed in the majority of renal carcinoma samples when compared to the matched normal kidney samples. Fzd2 and Fzd3 were upregulated in 3 of 5 colon adenocarcinoma samples. Fzd3 was also upregulated in the ovarian tumor tissue samples compared to the matched normal tissue samples. None of the other Fzds evaluated showed a specific differential expression pattern in any of the samples studied. Fzd5 and Fzd8 show 69.1% similarity and belong to the same subgroup of Fzds [14]. The significantly higher expression of Fzd5 and Fzd8 in the renal tumor samples, as compared to the normal renal samples, suggests a higher probability that this subgroup may be implicated in the progression of renal cancer. Therefore, we decided to further examine the possible role of Fzd5 in renal carcinoma.

We observed, using Western blotting, that protein levels were mostly consistent with mRNA levels in the tumor samples. In order to be able to determine the Fzd5 expression in a broader range of kidney tissues, we utilized a tissue array. Fzd5 membrane staining was detected in 9 of 30 (30%) clear cell carcinomas, and importantly, membrane staining was not detected in the matched 9 normal kidney tissue samples.

Since the Wnt signaling pathway appears to play an important role in embryonic development, in particular embryonic kidney induction [15, 16], activation of this pathway in the adult kidney due to mutation or overex-

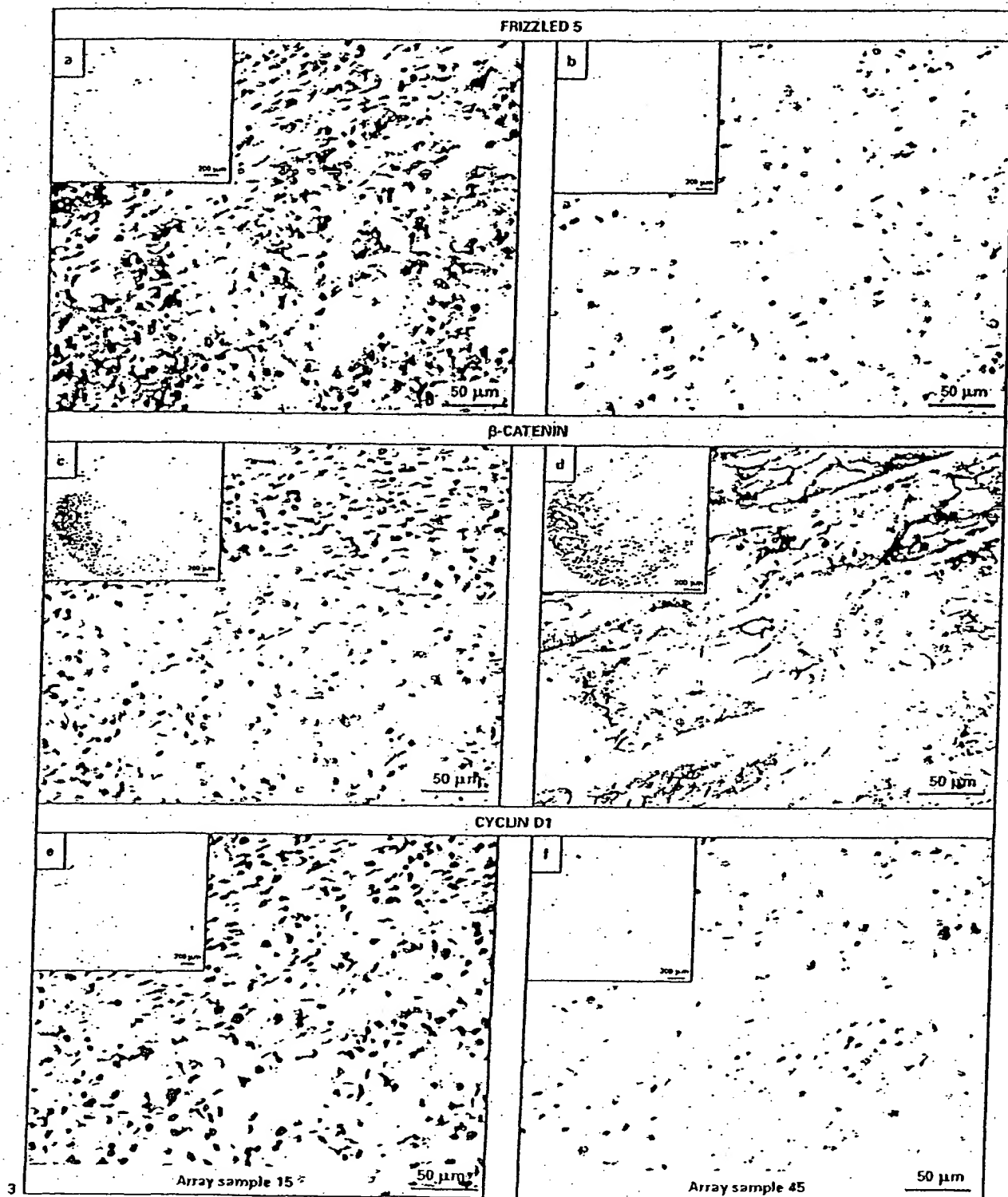


Table 3. Correlation between Fzd5 and β -catenin or cyclin D1 expression

| | Fzd5, % | |
|--------------------|---------|----|
| | + | - |
| β -Catenin + | 33 | 57 |
| β -Catenin - | 67 | 43 |
| Cyclin D1 + | 89 | 38 |
| Cyclin D1 - | 11 | 62 |

pression of one of the components of the pathway could be a determining factor in the development of renal cancers. Therefore, several studies have looked into the possible function the Wnt/ β -catenin pathway plays in renal carcinogenesis. APC gene mutations have been demonstrated not to be involved in renal carcinoma [17, 18]. In addition, β -catenin mutations are rare events in renal carcinoma [19, 20]. Nevertheless, cytoplasmic accumulation of β -catenin has been reported in a number of renal cell carcinomas [19], and thus the Wnt signaling pathway

Table 4. Wnt/ β -catenin target gene mRNA expression in tumor samples

| Sample ^a | Tissue | Tumor type | x-fold expression increase ^b | | |
|---------------------|--------|----------------------|---|-----------|---------------|
| | | | c-myc | cyclin D1 | PPAR δ |
| 133702 | kidney | adenocarcinoma | 0.54 | 4.52 | 0.52 |
| 137770 | kidney | renal cell carcinoma | 13.9 | 28.91 | 5.53 |
| 138844 | kidney | renal cell carcinoma | 2.39 | 31.49 | 7.48 |
| 137146 | kidney | renal cell carcinoma | 7.62 | 15.38 | 3.15 |
| 137564 | kidney | renal cell carcinoma | 33.82 | 19.65 | 8.65 |
| 133408 | kidney | renal cell carcinoma | 7.8 | 8.92 | 4.86 |
| 139188 | kidney | renal cell carcinoma | 2.22 | 9.92 | 11.67 |
| 135699 | kidney | renal cell carcinoma | 12.18 | 22.73 | 5.53 |
| 139064 | kidney | renal cell carcinoma | 22.11 | 5.04 | 6.33 |
| 134585 | kidney | renal cell carcinoma | 1.79 | 1.14 | 1.67 |
| 140279 | kidney | renal cell carcinoma | 61.68 | 54.95 | 14.62 |

^a Sample identification numbers were given by the pathologist.

^b Results are expressed as x-fold increase of the gene in the tumor tissue sample compared to its matched normal tissue sample after normalizing both samples on the basis of their ATP5b content. A cutoff of 3-fold was used to define differential expression. Significant (>3-fold) increases in the expression level of the Fzd receptors are in italics.

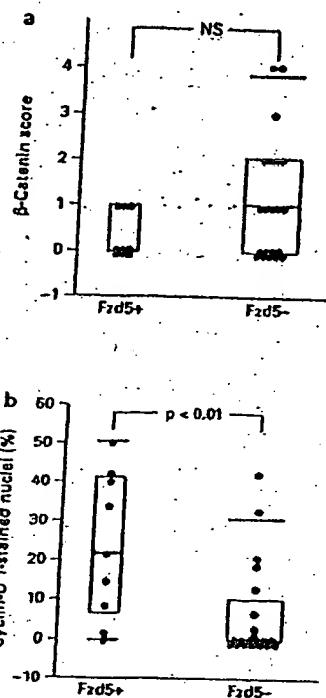


Fig. 3. Fzd5, β -catenin and cyclin D1 immunostaining of the CL1 renal carcinoma tissue arrays. The left column of images represents serial sections from tumor sample 15. Insets show an overview of each tumor section on the serial tissue arrays. The Fzd5-immunoreactive clear cell renal carcinoma (a) of this tumor sample does not express β -catenin (e). Immunostaining for cyclin D1 (e) detects distinct labeling of nuclei in clear cell renal carcinoma. The right column of images is taken from serial sections of tumor sample 45. Clear cell renal carcinoma from this tumor sample does not express Fzd5 (b) and cyclin D1 (f) but does show distinct membrane β -catenin staining (d).

Fig. 4. Box plot charts (thick black line = median) illustrating the relationship between Fzd5 immunostaining and β -catenin (a) and cyclin D1 (b) expression in clear cell renal carcinoma. No significant correlation was observed between the β -catenin scores of Fzd5-positive and -negative clear cell renal carcinoma. Nuclear cyclin D1 staining in clear cell renal carcinoma showed a significant difference between Fzd5-positive and Fzd5-negative tumor samples.

might act as an inducer of tumorigenesis in the kidney. This view is supported by the observation that aberrant activity of the Wnt signaling pathway has been reported in renal-cancer-derived cell lines. Zang et al. [21] observed a higher expression level of Wnt5a and Fzd5 mRNA in the renal cancer cell line GRC-1 than in the normal renal cell line HK-2. Expression of β -catenin was also higher in GRC-1 than in HK-2.

To determine the status of the canonical Wnt signaling pathway in our renal carcinoma samples, we have quantitated the mRNA levels of three important target genes of T cell factor/lymphoid enhancer factor activation by β -catenin. The mRNA levels of these three target genes (c-myc, cyclin D1 and PPAR δ) correlated largely with the expression of Fzd5 in these samples, suggesting that the canonical pathway is activated. On the kidney tissue array, cyclin D1 protein expression showed a highly significant correlation with the Fzd5 expression in the tumor samples (table 3). Cyclin D1 protein is frequently overexpressed in various tumors, but in only a proportion of the cases is it due to amplification of the cyclin D1 gene [22]. Therefore, other mechanisms such as upregulation of gene transcription may play a substantial role in the overexpression of cyclin D1 [23–26]. Our data, showing increased cyclin D1 expression in renal carcinoma samples, are consistent with the results of Stassar et al. [27]. They studied genes that are associated with human renal carcinoma by suppression subtractive hybridization and reported 14 differentially expressed genes, including cyclin D1. Although we would have expected an increased nuclear β -catenin staining, nuclear accumulation of β -cate-

nin was not observed in any of the tumors or on the tissue array. This result is consistent with the data presented for renal cell carcinomas by Kim et al. [19]. They did not detect nuclear β -catenin staining in the 52 renal cell carcinomas examined. The lack of nuclear β -catenin staining has also been reported by others in tumors that might have arisen from Wnt/ β -catenin pathway activation [28–31].

While expression of both Wnt5a and Fzd5 does induce duplication of the *Xenopus* head, exogenous expression of Fzd5 in a *Xenopus* model does not induce duplication of the head [32]. Fzd5 does not activate the β -catenin signaling pathway on its own, as the presence of its endogenous ligand is also required. Our results suggest that Fzd5 may have a role in renal cell carcinogenesis due to its frequent overexpression observed in these tumor samples, and we hypothesize that if Fzd5 is overexpressed, it has a rather limited effect on β -catenin signaling. However, in the presence of its endogenous still unknown ligand, it activates the canonical Wnt signaling pathway. The elucidation of this ligand and its binding characteristics is still under investigation. Ultimately, knowledge of the specific expression patterns of both Wnt and Fzd members could lead to directed tumor targeting or could be used as a tumor marker.

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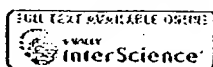
References

- Pandur P, Maurus D, Kuhl M: Increasingly complex: New players enter the Wnt signaling network. *Bioessays* 2002;24:881–884.
- Dale TC: Signal transduction by the Wnt family of ligands. *Biochem J* 1998;329:209–223.
- Oving JM, Clevers HC: Molecular causes of colon cancer. *Eur J Clin Invest* 2002;32:448–457.
- Pinson KJ, Brennan J, Monkley S, Avery BJ, Skarnes WC: An LDL-receptor-related protein mediates Wnt signalling in mice. *Nature* 2000;407:535–538.
- Tamai K, Semenov M, Kato Y, Spokony R, Liu CM, Katsuyama Y, Hess F, Saint-Jeannet JP, He X: LDL-receptor-related proteins in Wnt signal transduction. *Nature* 2000;407:530–535.
- Miller JR: The Wnts. *Genome Biol* 2002; vol 3.
- Sato S, Daigo Y, Furukawa Y, Kato T, Miwa N, Nishiwaki T, Kawasoe T, Ishiguro H, Fujita M, Tokino T, Sasaki Y, Imaoka S, Murata M, Shimano T, Yamaoka Y, Nakamura Y: AXIN1 mutations in hepatocellular carcinomas, and growth suppression in cancer cells by virus-mediated transfer of AXIN1. *Nat Genet* 2000;24:245–250.
- Vider BZ, Zimmer A, Chastre E, Prevot S, Gespach C, Estlin D, Wolkoch Y, Tronick SR, Gazit A, Yaniv A: Evidence for the involvement of the Wnt 2 gene in human colorectal cancer. *Oncogene* 1996;12:153–158.
- Iozzo RV, Eichstetter I, Danielson KG: Aberrant expression of the growth factor Wnt-5A in human malignancy. *Cancer Res* 1995;55:3495–3499.
- Gerard CJ, Andrejka LM, Macina RA: Mitochondrial ATP synthase 6 as an endogenous control in the quantitative RT-PCR analysis of clinical cancer samples. *Mol Diagn* 2000;5:39–46.
- Tanaka S, Akiyoshi T, Mori M, Wands JR, Sugimachi K: A novel frizzled gene identified in human esophageal carcinoma mediates APC/beta-catenin signals. *Proc Natl Acad Sci USA* 1998;95:10164–10169.
- To KF, Chan MW, Leung WK, Yu J, Tong JH, Lee TL, Chan FK, Sung JJ: Alterations of frizzled (FzE3) and secreted frizzled related protein (sFRP) expression in gastric cancer. *Life Sci* 2001;70:483–489.
- Rhee CS, Sen M, Lu D, Wu C, Leoni L, Rubin J, Corr M, Carson DA: Wnt and frizzled receptors as potential targets for immunotherapy in head and neck squamous cell carcinomas. *Oncogene* 2002;21:6598–6605.

- 14 Saitoh T, Hirai M, Katoh M: Molecular cloning and characterization of human frizzled-8 gene on chromosome 10p11.2. *Int J Oncol* 2001;18: 991-996.
- 15 Dressler GR: Tubulogenesis in the developing mammalian kidney. *Trends Cell Biol* 2003;12: 390-395.
- 16 Vainio SJ, Itaranta PV, Perasaari JP, Uusitalo MS: Wnts as kidney tubule inducing factors. *Int J Dev Biol* 1999;43:419-423.
- 17 Bohm M, Wicland I, Stinhofer C, Otto T, Rubben H: Detection of loss of heterozygosity in the APC tumor suppressor gene in nonpapillary renal cell carcinoma by microdissection and polymerase chain reaction. *Urol Res* 1997;25: 161-165.
- 18 Suzuki H, Ueda T, Komiya A, Okano T, Isaka S, Shimazaki J, Ito H: Mutational state of von Hippel-Lindau and adenomatous polyposis coli genes in renal tumors. *Oncology* 1997;54: 252-257.
- 19 Kim YS, Kang YK, Kim JB, Han SA, Kim KJ, Paik SR: Beta-catenin expression and mutational analysis in renal cell carcinomas. *Pathol Int* 2000;50:725-730.
- 20 Ueda M, Gemmill RM, West J, Winn R, Sugita M, Tanaka N, Ueki M, Drabkin HA: Mutations of the β - and γ -catenin genes are uncommon in human lung, breast, kidney, cervical and ovarian carcinomas. *Br J Cancer* 2001;85: 64-68.
- 21 Zang T, Zhuang L, Zhang Z, Xin D, Guo Y: Aberrant activity of WNT/frizzled signaling pathway in renal cancer cell lines. *Chin Sci Bull* 2000;45:1703-1707.
- 22 Ozturk M: Genetic aspects of hepatocellular carcinogenesis. *Semin Liver Dis* 1999;19:235-242.
- 23 Aktas H, Cai H, Cooper GM: Ras links growth factor signaling to the cell cycle machinery via regulation of cyclin D1 and the Cdk inhibitor p27KIP1. *Mol Cell Biol* 1997;17:3850-3857.
- 24 Weber JD, Raben DM, Phillips PJ, Baldassare JJ: Sustained activation of extracellular-signal-regulated kinase 1 (ERK1) is required for the continued expression of cyclin D1 in G1 phase. *Biochem J* 1997;326:61-68.
- 25 Lavoie JN, L'Allemain G, Brunet A, Muller R, Pouyssegur J: Cyclin D1 expression is regulated positively by the p42/p44MAPK and negatively by the p38/HOGMAPK pathway. *J Biol Chem* 1996;271:20608-20616.
- 26 Treinies I, Paterson HF, Hooper S, Wilson R, Marshall CJ: Activated MEK stimulates expression of AP-1 components independently of phosphatidylinositol 3-kinase (PI3-kinase) but requires a PI3-kinase signal to stimulate DNA synthesis. *Mol Cell Biol* 1999;19:321-329.
- 27 Stassar MJ, Devitt G, Brosius M, Rinnab L, Prang J, Schradin T, Simon J, Petersen S, Kopp-Schneider A, Zoller M: Identification of human renal cell carcinoma associated genes by suppression subtractive hybridization. *Br J Cancer* 2001;85:1372-1382.
- 28 Wong SC, Lo SF, Lee KC, Yam JW, Chan JK, Wendy Hsiao WL: Expression of frizzled-related protein and Wnt-signalling molecules in invasive human breast tumours. *J Pathol* 2002; 196:145-153.
- 29 Ueta T, Ikeguchi M, Hirooka Y, Kaibara N, Terada T: Beta-catenin and cyclin D1 expression in human hepatocellular carcinoma. *Oncol Rep* 2002;9:1197-1203.
- 30 Tanaka Y, Kato K, Notohara K, Nakatani Y, Miyake T, Ijiri R, Nishimata S, Ishida Y, Kigasawa H, Ohama Y, Tsukayama C, Kobayashi Y, Horie H: Significance of aberrant (cytoplasmic/nuclear) expression of beta-catenin in pancreaticoblastoma. *J Pathol* 2003;199: 185-190.
- 31 Qiao Q, Ramadani M, Gonsauge S, Gonsauge F, Leder G, Beger HG: Reduced membranous and ectopic cytoplasmic expression of beta-catenin correlate with cyclin D1 overexpression and poor prognosis in pancreatic cancer. *Int J Cancer* 2001;95:194-197.
- 32 He X, Saint-Jeannet JP, Wang Y, Nathans J, Dawid I, Varmus H: A member of the frizzled protein family mediating axis induction by Wnt-5A. *Science* 1997;275:1652-1654.

Erratum in:

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Immunohistochemical analysis of NY-ESO-1 antigen expression in normal and malignant human tissues.

Jungbluth AA, Chen YT, Stockert E, Busam KJ, Kolb D, Iversen K, Coplan K, Williamson B, Altorki N, Old LJ.

Ludwig Institute for Cancer Research, Memorial Sloan-Kettering Cancer Center, New York, NY, USA. jungblua@mskcc.org

NY-ESO-1, a member of the CT (cancer/testis) family of antigens, is expressed in normal testis and in a range of human tumor types. Knowledge of NY-ESO-1 expression has depended on RT-PCR detection of mRNA and there is a need for detecting NY-ESO-1 at the protein level. In the present study, a method for the immunochemical detection of NY-ESO-1 in paraffin-embedded tissues has been developed and used to define the expression pattern of NY-ESO-1 in normal tissues and in a panel of human tumors. No normal tissue other than testis showed NY-ESO-1 reactivity, and expression in testis was restricted to germ cells particularly spermatogonia. In human tumors, the frequency of NY-ESO-1 antigen expression corresponds with past analysis of NY-ESO-1 mRNA expression e.g., 20-30% of lung cancers, bladder cancers and melanoma, and no expression in colon and renal cancer. Co-typing of NY-ESO-1 antigen and mRNA expression in a large panel of lung cancers showed a good correlation. There is great variability in NY-ESO-1 expression in individual tumors, ranging from an infrequent homogeneous pattern of staining to highly heterogeneous antigen expression. Copyright 2001 Wiley-Liss, Inc.

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Multidrug resistance phosphoglycoprotein (ABCB1) in the mouse placenta: fetal protection.

Kalabis GM, Kostaki A, Andrews MH, Petropoulos S, Gibb W, Matthews SG.

Department of Physiology, University of Toronto, Ontario, Canada.

The multidrug resistance phosphoglycoprotein ATP-binding cassette subfamily B (ABCB1) actively extrudes a range of structurally and functionally diverse xenobiotics as well as glucocorticoids. ABCB1 is present in many cancer cell types as well as in normal tissues. Although it has been localized within the mouse placenta, virtually nothing is known about its regulation. In the mouse, two genes, *Abcb1a* and *Abcb1b*, encode ABCB1. We hypothesized that there are changes in placental *Abcb1a* and *Abcb1b* gene expression and ABCB1 protein levels during pregnancy. Using in situ hybridization, we demonstrated that *Abcb1b* mRNA is the predominant placental isoform and that there are profound gestational changes in the expression of both *Abcb1a* and *Abcb1b* mRNA. Placentas from pregnant mice were analyzed between Embryonic Days (E) 9.5 and 19 (term approximately 19.5d). *Abcb1b* mRNA was detected in invading trophoblast cells by E9.5, peaked within the placental labyrinth at E12.5, and then progressively decreased toward term ($P < 0.0001$). *Abcb1a* mRNA, although lower than that of *Abcb1b* at midgestation, paralleled changes in *Abcb1b* mRNA. Changes in *Abcb1* mRNA were reflected by a significant decrease in ABCB1 protein ($P < 0.05$). A strong correlation existed between placental *Abcb1b* mRNA and maternal progesterone concentrations, indicating a potential role of progesterone in regulation of placental *Abcb1b* mRNA. In conclusion, there are dramatic decreases in *Abcb1a* and *Abcb1b* mRNA and in ABCB1 at the maternal-fetal interface over the second half of gestation, suggesting that the fetus may become increasingly susceptible to the influences of xenobiotics and natural steroids in the maternal circulation.

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Expression of human telomerase reverse transcriptase gene and protein, and of estrogen and progesterone receptors, in breast tumors: Preliminary data from neo-adjuvant chemotherapy.

Kamori M, Izumiyama N, Hashimoto M, Nakamura K, Okano T, Kurabayashi R, Naoki H, Honma N, Ogawa T, Kaminishi M, Takubo K.

Division of Breast and Endocrine Surgery, Department of Surgery, Graduate School of Medicine, The University of Tokyo, Bunkyo-ku, Tokyo 113-8655, Japan. kamori-dis@umin.ac.jp.

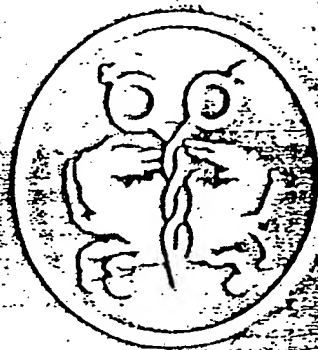
Human telomerase reverse transcriptase (hTERT), the catalytic subunit of telomerase, is very closely associated with telomerase activity. Telomerase has been implicated in cellular immortalization and carcinogenesis. In situ detection of hTERT will aid in determining the localization of telomerase-positive cells. The aim of this study was to detect expression of hTERT mRNA, hTERT protein, estrogen receptor (ER) and progesterone receptor (PR) in paraffin-embedded breast tissue samples and to investigate the relationship between hTERT expression and various clinicopathological parameters in breast tumorigenesis. We used in situ hybridization (ISH) to examine hTERT gene expression, and immunohistochemistry (IHC) to examine expression of hTERT protein, ER and PR, in breast tissues including 64 adenocarcinomas, 2 phyllode tumors and their adjacent normal breast tissues. hTERT gene expression was detected by ISH in 56 (88%) carcinomas, but in neither of the 2 phyllode tumors. hTERT protein expression was detected by IHC in 52 (81%) carcinomas, but in neither of the 2 phyllode tumors. Moreover, ER and PR were expressed in 42 (66%) and 42 (66%) carcinomas, respectively, and in neither of the 2 phyllode tumors. In 4 cases of breast carcinoma that strongly expressed hTERT gene and protein before treatment, neo-adjuvant chemotherapy led to disappearance of gene and protein expression in all cases. There was a strong correlation between detection of hTERT gene expression by ISH and of hTERT protein by IHC in tissue specimens from breast tumors. These results suggest that detection of hTERT protein by IHC can be used to distinguish breast cancers as a potential diagnostic and therapeutic marker.

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Expression of human telomerase reverse transcriptase gene and protein, and of estrogen and progesterone receptors, in breast tumors: Preliminary data from neo-adjuvant chemotherapy

MAKOTO KAMMORI^{1,2}, NAOTAKA IZUMIYAMA², MASANORI HASHIMOTO¹, KEN-ICHI NAKAMURA²,
TADAO OKANO³, RIE KURABAYASHI¹, HIKI NAOKI¹, NAOKO HONMA², TOSHIHISA OGAWA¹,
MICHIO KAMINISHI¹ and KAIYO TAKUBO²

¹Division of Breast and Endocrine Surgery, Department of Surgery, Graduate School of Medicine,
The University of Tokyo; ²Human Tissue Research Group, Tokyo Metropolitan Institute of Gerontology;

³Tokyo Metropolitan Tama Cancer Detection Center, Tokyo, Japan

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Abstract. Human telomerase reverse transcriptase (hTERT), the catalytic subunit of telomerase, is very closely associated with telomerase activity. Telomerase has been implicated in cellular immortalization and carcinogenesis. *In situ* detection of hTERT will aid in determining the localization of telomerase-positive cells. The aim of this study was to detect expression of hTERT mRNA, hTERT protein, estrogen receptor (ER) and progesterone receptor (PR) in paraffin-embedded breast tissue samples and to investigate the relationship between hTERT expression and various clinicopathological parameters in breast tumorigenesis. We used *in situ* hybridization (ISH) to examine hTERT gene expression, and immunohistochemistry (IHC) to examine expression of hTERT protein, ER and PR, in breast tissues including 64 adenocarcinomas, 2 phyllode tumors and their adjacent normal breast tissues. hTERT gene expression was detected by ISH in 56 (88%) carcinomas, but in neither of the 2 phyllode tumors. hTERT protein expression was detected by IHC in 52 (81%) carcinomas, but in neither of the 2 phyllode tumors. Moreover, ER and PR were expressed in 42 (66%) and 42 (66%) carcinomas, respectively, and in neither of the 2 phyllode tumors. In 4 cases of breast carcinoma that strongly expressed hTERT gene and protein before treatment, neo-adjuvant chemotherapy led to disappearance of gene and protein expression in all cases. There was a strong correlation between detection of hTERT gene expression by ISH and of hTERT protein by ICH in tissue specimens from breast tumors. These results suggest that detection of hTERT

protein by ICH can be used to distinguish breast cancers as a potential diagnostic and therapeutic marker.

Introduction

Breast cancer is the most frequent malignancy in women, affecting up to one in every eight females worldwide. The most important clinicopathological prognostic parameter so far identified is the absence or presence of lymph node metastasis, but the identification of further parameters for both lymph node-positive and -negative patients would facilitate an individually based risk-directed therapy (1). A promising emerging molecular marker is telomerase, a ribonucleoprotein enzyme complex, which when activated or upregulated allows tumor cells to escape from cellular senescence and to proliferate indefinitely (2). The human telomere is a simple repeat sequence of six bases (TTAGGG) that is located at the ends of each chromosome (3). Telomeres are believed to protect against degeneration, reconstruction, fusion, and loss (4) and to promote the homologous pairing of chromosomes (5). The end-to-end chromosome fusions observed in some tumors may result from the loss of telomeres and may be partly responsible for the genetic instability associated with tumorigenesis. Telomerase catalyzes the synthesis of telomere DNA and facilitates cell immortalization through the stabilization of chromosomal structure (6-8). Although the expression of the human RNA component of telomerase (hTERC) is widespread, the restricted expression pattern of the mRNA of hTERT, the human telomerase catalytic subunit gene, is correlated with telomerase activity (8-13). As telomerase activity seems to be the key player in tumor cell immortality, it has importance as a target molecule for anti-cancer therapy. Telomerase activity has been shown to correlate with poor clinical outcome in neuroblastomas and other tumors (14). For breast cancer, however, telomerase activity is a controversial prognostic marker: some studies suggest that telomerase activity, clinicopathological parameters and disease outcome are linked, whereas others do not find this association (14-23).

Correspondence to: Dr Makoto Kammori, Division of Breast and Endocrine Surgery, Department of Surgery, Graduate School of Medicine, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-8655, Japan
E-mail: kammori-dis@umin.ac.jp

Key words: human telomerase reverse transcriptase, telomerase, estrogen receptor, breast cancer, chemotherapy

We have succeeded in very clearly and sensitively demonstrating hTERT mRNA in thyroid, colorectal, parathyroid and lung tissues by use of an oligonucleotide probe (13,24-26). Strong correlation has been observed between hTERT mRNA and/or protein expression and telomerase in a variety of malignant tumors (13,14,24,25,27,28). In the present study, we used ISH to examine expression of the hTERT gene, and IHC to examine expression of hTERT protein, ER and PR, in 64 carcinomas and 2 phyllodes tumors of breast to determine whether hTERT protein can be used to differentiate breast cancers. We also analyzed hTERT mRNA and protein expression with special reference to clinical features and histological findings to investigate the potential role of hTERT mRNA expression analysis in predicting the biological characteristics of breast cancers. Since hTERT expression in breast tumors has not previously been analysed by ISH or IHC, our investigation also examined various clinicopathological parameters, including age, histopathological type, tumor size, lymph node status, relapses, and the expression of ER and PR.

Materials and methods

Tissue collection. Sixty-six samples were obtained during 66 mastectomies: 64 breast carcinomas, 2 phyllodes tumors and 66 specimens of the adjacent normal breast gland. In 4 cases, samples were obtained during core needle biopsies (CNB) before neo-adjuvant therapy, and again during mastectomies after neo-adjuvant therapy; for these cases, all measurements and examinations were performed both before and after the neo-adjuvant therapy. The patients ranged in age from 32 to 90 years. The patients with carcinomas ranged in age from 37 to 90, mean 56, years and were all women. The women with phyllode tumors were aged 32 and 38 years, respectively. The surgical and CNB tissue samples were frozen rapidly with liquid nitrogen and stored at -80°C until fixation. Then, they were fixed in 10% buffer formalin solution and embedded in paraffin. Surgical and CNB samples were collected from the patients after obtaining their informed consent, and the study protocol was approved by the Medical Department of the University of Tokyo Ethics Committee. The pathologic diagnoses were made by the surgical pathological specialists at our institute on the basis of examination of hematoxylin-eosin stained slides. A pathological review was performed for all breast tumors according to the BRE score. pT and pN staging were assigned according to the 1997 WHO classification (7th edition).

MCF-7 human breast cancer cells, kindly provided by the Cell Resource Center for Biomedical Research, Institute of Development, Aging and Cancer, Tohoku University, were used as positive controls. The cells were incubated in RPMI-1640 medium with 25 mM HEPES buffer, L-glutamine, and 10% fetal bovine serum (Gibco, Grand Island, NY) on a chamber-attached slideglass (Lab-Tek³ Chamber Slide™; Nalge Nunc International, Naperville, IL) in a humidified 5% CO₂ atmosphere at 37°C. The cells were then fixed with 10% buffered neutral formalin (Sigma Chemical Co., St. Louis, MO). The cultured MCF-7 cell line that was used as a positive control was tested for telomerase with a PCR-based standard TRAP assay (6,13). These cells were also used to prepare cell blocks. Briefly, the cells were fixed in 10% buffered neutral

formalin, resuspended in molten agarose and then embedded in paraffin. Sections from these cell blocks were used as positive procedural controls in ISH and IHC. The negative control in ISH was obtained by replacing the oligonucleotide probe with RNase. The negative control in IHC was obtained by replacing the primary antibody with Tris-buffered saline (TBS).

Oligonucleotide probe for ISH. The specificity of the oligonucleotide sequence was initially determined by a GenEMBL database search using the Genetics Computer Group Sequence Analysis Program (GCG, Madison, WI) based on the fastA algorithm (29); the sequence exhibited 100% homology with the hTERT gene sequence. A d(T)₂₀ oligonucleotide was used to verify the integrity and lack of degradation of the mRNA in each sample. All oligonucleotide probes were synthesized with a hapten-labeled nucleotide, such as digoxigenin-dUTP (Boehringer-Mannheim), at the 3' end via direct coupling by using standard phosphoramidite chemistry (Research Genetics, Huntsville, AL) (30). The probe used for detection of hTERT by ISH was generated from the original sequence for *Homo sapiens* telomerase reverse transcriptase (AF015950), 2766-2800: 5'-GCCTCGTCTTCTACAGGA AGTTCACCACTGTCTT-3' (13,24-26).

ISH. ISH was performed with the GenPoint nucleic acid hyper-detection system (Dako, Carpinteria, CA) (31). Formalin-fixed, paraffin-embedded tissue sections (5 µm thick) were deparaffinized in xylene and a graded alcohol series. Tissue sections and CNB samples were then pretreated with target retrieval solution (Dako, S1700) at 95°C and proteinase K (Dako, S3004) at room temperature. Next, the tissues and CNB samples were fixed in 0.3% hydrogen peroxide followed by a methyl alcohol series at room temperature. Digoxigenin-labeled anti-sense oligonucleotide in mRNA *in situ* hybridization solution (Dako, S3304) was placed over the tissues and CNB samples. After hybridization at 37°C overnight, the slides were washed in stringent wash solution (Dako, GenPoint System Kit) at 45°C. The tissues and CNB samples were exposed to avidin blocking solution (Dako, X0590) at room temperature followed by biotin blocking solution (Dako, X0590) at room temperature. The tissues and samples were then incubated at room temperature with a sheep monoclonal hapten-labeled anti-digoxigenin antibody (Dako, p5104), and the slides were then fixed with biotinyl tyramine (Dako, GenPoint System Kit) at room temperature. Finally, the slides were incubated with HRP-conjugated streptavidin (Dako, GenPoint System Kit) at room temperature. Since 3,3'-diaminobenzidine tetrahydrochloride (DAB) was used as the substrate, a positive reaction was visible as a brown color under a light microscope. The sections were weakly counter-stained with 0.1% hematoxylin.

IHC. IHC was performed by the avidin-biotin complex/horseradish peroxidase method. Tissue sections were stained for hTERT with commercially available monoclonal antibody (NCL-L-hTERT; Novocastra, Newcastle upon Tyne, UK). Sections were dewaxed in xylene. Antigen retrieval was done by incubating sections immersed in 0.01 M citrate buffer at pH 6.0 in a microwave oven at 99°C. The sections were allowed to cool down at room temperature. The sections

MCF-7 Lysis Case 6 Case 14 Case 23 Case 44 Case 59

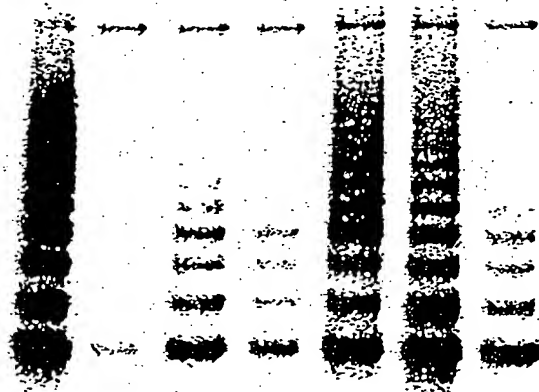


Figure 1. Representative results of the TRAP assay. If functional telomerase is present, the enzyme adds DNA to the substrate in 6-base-pair (bp) increments, resulting in a ladder-like distribution of products. The 6-bp ladder signals are apparent for MCF-7 and breast cancers (case nos. 6, 14, 23, 44 and 59) and are not apparent for lysis buffer as the negative control. An extract of MCF-7 was used as a positive control for the TRAP assay and as an Internal Telomerase Assay Standard (ITAS) positive control for PCR amplification, with lysis buffer as the negative control (Lysis).

were then immersed in 1% hydrogen peroxide (H_2O_2) in methanol to block endogenous peroxidase activity. Following that, the sections were washed in TBS (pH 7.6) before being incubated in normal rabbit serum for 20 min to block non-specific binding. After draining off the excess serum, the sections were incubated with the primary antibody at room temperature. The sections were washed in TBS before being incubated with the secondary antibody (biotinylated rabbit anti-mouse, Dako). The sections were washed again with TBS and incubated with avidin-biotin complex/horseradish peroxidase. After washing the sections with TBS, peroxidase activity was visualized under light microscopy by applying DAB chromogen (Dako). The sections were counterstained with hematoxylin, dehydrated in increasing grades of alcohol and finally mounted in dibutyl phthalate (DPX) mountant.

Homogeneous staining or a speckled/dotted pattern in the nucleus was considered positive staining, and absence of distinct nuclear staining was taken as negative staining. Grading of the percentage of stained cells (hTERT labeling index) was performed by previously published criteria (1) as follows: Grade 1, negative staining; Grade 2, 1-10% positive staining nuclei; Grade 3, 11-50% positive nuclei; and Grade 4, >50% positive nuclei. Immunostained slides for ER and PR were scored as previously described (32,33). In brief, each entire slide was evaluated by light microscopy. First, a proportion score was assigned, which represents the estimated proportion of positive-staining tumor cells (0, none; 1, <1/100; 2, 1/100 to 1/10; 3, 1/10 to 1/3; 4, 1/3 to 2/3; and 5, >2/3). Next, an intensity score was assigned, which represents the average intensity of positive tumor cells (0, none; 1, weak; 2, intermediate; and 3, strong). The proportion and intensity scores were then added to obtain a total score, which ranged from 0 to 8. Slides were scored by pathologists who did not have knowledge of ligand-binding results or patient outcome.

Table I. Relationships between mRNA status (negative/positive) by ISH and standard clinical, pathological, and biological factors in the 66 tumors.

| | Total population (%) | No. of patients (%) | | P-value ^a |
|--------------------------------|----------------------|---------------------|----------------|----------------------|
| | | hTERT negative | hTERT positive | |
| Total | 66 | 10 (15.2) | 56 (84.8) | |
| Age | | | | NS |
| ≤50 | 26 | 6 (23.1) | 20 (76.9) | |
| >50 | 40 | 6 (15.0) | 34 (85.0) | |
| Histopathological type | | | | NS |
| Scirrhous | 32 | 2 (6.4) | 30 (93.6) | |
| Papillotubular | 20 | 2 (10.0) | 18 (90.0) | |
| Solid tubular | 6 | 1 (16.7) | 5 (83.3) | |
| Mucinous | 2 | 1 (50.0) | 1 (50.0) | |
| Non-invasive | 4 | 2 (50.0) | 2 (50.0) | |
| Phyllodes | 2 | 2 (100) | 0 (0) | |
| Tumor size (cm) ^b | | | | NS |
| T1 (<2.0) | 20 | 0 (0) | 20 (100) | |
| T2 (2.0-5.0) | 34 | 6 (17.6) | 28 (82.4) | |
| T3 (>5.0) | 10 | 2 (20.0) | 8 (80.0) | |
| Lymph node status ^b | | | | NS |
| pN0 | 34 | 6 (17.6) | 28 (82.4) | |
| pN1 | 26 | 2 (7.7) | 24 (92.3) | |
| pN2+pNM | 4 | 0 (0) | 4 (100) | |
| Relapse | | | | NS |
| + | 10 | 0 (0) | 10 (100) | |
| - | 56 | 10 (17.6) | 46 (82.4) | |
| ER expression | | | | NS |
| + (≥2) | 42 | 6 (14.3) | 36 (85.7) | |
| - (<2) | 24 | 4 (16.7) | 20 (83.3) | |
| PR expression | | | | NS |
| + (≥2) | 42 | 4 (9.5) | 38 (90.5) | |
| - (<2) | 24 | 6 (25.0) | 18 (75.0) | |

^a χ^2 test. NS, not significant. ^bInformation available for 64 patients.

Statistical analysis. Differences in p-values were analyzed with the χ^2 test for independence, and Fisher's test was used for correlations. In all comparisons, $p < 0.05$ was considered significant.

Results

Representative results of the TRAP assay are shown in Fig. 1. The cultured cells, which were tested for telomerase activity

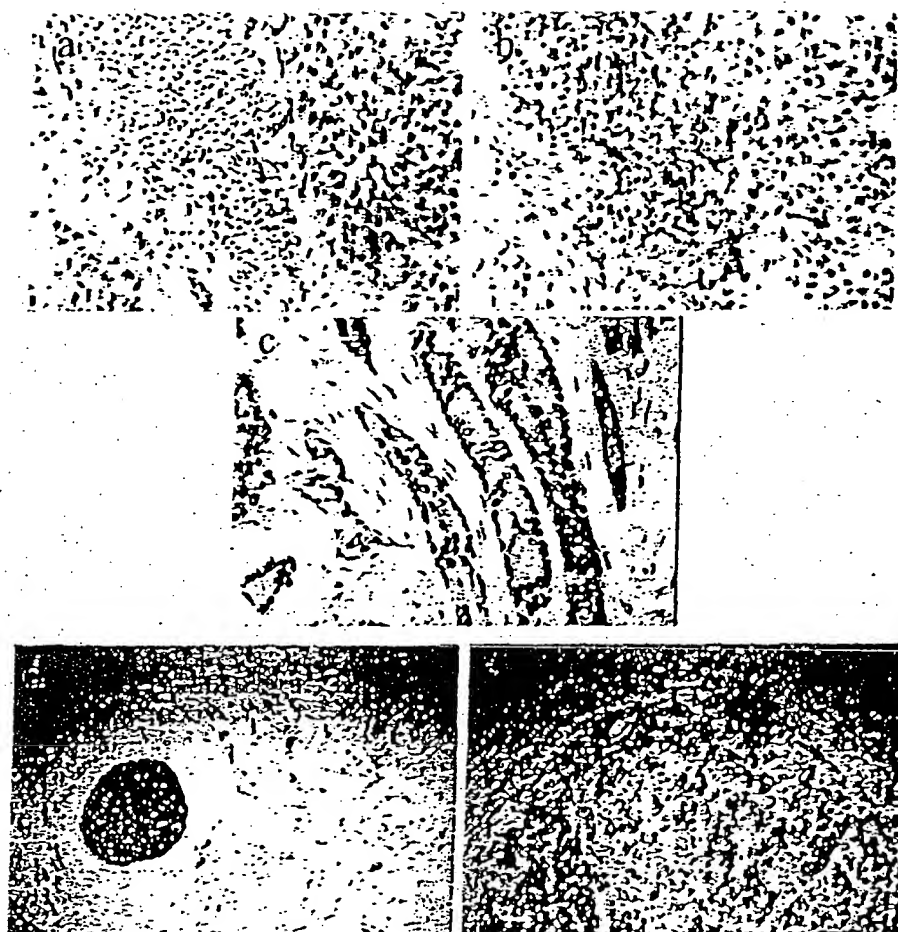


Figure 2. Correlation between histologic diagnosis, human telomerase reverse transcriptase (hTERT) mRNA by ISH, hTERT protein by IHC, estrogen receptor (ER) by IHC and progesterone receptor (PR) by IHC in breast cancers. (a, H&E); (b, hTERT mRNA); (c, hTERT protein); (d, ER) and (e, PR).

with the TRAP assay, gave positive results with all procedural controls (MCF-7 and 5 breast cancer samples) (Fig. 1).

ISH revealed that hTERT mRNA was strongly expressed in the nuclei and cytoplasm of almost all of the MCF-7 human cancer cells (data not shown). Expression of hTERT mRNA was detected in 56 (88%) of the 64 breast cancers and in none of the phyllodes tumors of the breast (Table I) with the anti-sense probe, whereas no expression was detected with the anti-sense probe treated with RNase (data not shown). The levels of expression were heterogeneous within the carcinomatous regions. Strong expression of hTERT mRNA was not confined to the carcinomatous regions but was also detected in infiltrating lymphocytes (Fig. 2a and b). Higher expression levels of both signals of hTERT mRNA were detected in some sections containing both carcinomas and lymphocytes, compared with the adjacent non-cancerous mammary gland, but no clear differences in signal intensity were observed between carcinomas and lymphocytes. The signals in both the normal and cancer tissues were mainly present in the lymphocytes, and the signal intensity was similar in both, although a precise quantitative comparison of the *in situ* signals was impossible.

IHC revealed that hTERT protein was strongly expressed in the nuclei, nuclear membrane and cytoplasm of almost all of the MCF-7 human cancer cells (data not shown). Expression of hTERT protein was detected in 52 (81%) of the 64 breast cancers and in none of the phyllode tumors of the breast (Table II). The levels of expression were heterogeneous within the carcinomatous regions. As shown in Fig. 2c, strong expression of hTERT protein was observed in nuclei, nuclear membrane and cytoplasm, similar to the pattern in MCF-7 human cancer cells. Normal mammary gland and stromal cells generally showed negative immunoreactivity against hTERT protein antibody.

A nuclear signal for the ER (Fig. 2d), as assessed by IHC, was observed in 36 (56%) of the 64 breast cancers and in none of the phyllode tumors of the breast, with positive scores ranging from 2 to 8 (Tables I and II). A nuclear signal for the PR (Fig. 2e), as assessed by IHC, was observed in 38 (59%) of the 64 breast cancers and in none of the phyllode tumors of the breast, with positive scores ranging from 2 to 8 (Tables I and II).

We used ISH and IHC to examine hTERT expression in 4 cases of breast cancer before and after neo-adjuvant

Table II. Relationships between mRNA status (negative/positive) by IHC and standard clinical, pathological, and biological factors in the 66 tumors.

| | Total | No. of patients (%) | | P-value ^a |
|--------------------------------|-------|---------------------|----------------|----------------------|
| | | hTERT negative | hTERT positive | |
| Total | 66 | 14 (21.2) | 52 (78.8) | |
| Age | | | | NS |
| ≤50 | 26 | 10 (38.5) | 16 (61.5) | |
| >50 | 40 | 4 (10.0) | 36 (90.0) | |
| Histopathological type | | | | NS |
| Scirrhous | 32 | 2 (6.4) | 30 (93.6) | |
| Papillotubular | 20 | 4 (20.0) | 16 (80.0) | |
| Solid tubular | 6 | 4 (66.7) | 2 (33.3) | |
| Mucinous | 2 | 0 (0) | 2 (100) | |
| Non-invasive | 4 | 2 (50.0) | 2 (50.0) | |
| Phyllodes | 2 | 2 (100) | 0 (0) | |
| Tumor size (cm) ^b | | | | NS |
| T1 (<2.0) | 20 | 4 (20.0) | 16 (80.0) | |
| T2 (2.0-5.0) | 34 | 6 (17.6) | 28 (82.4) | |
| T3 (>5.0) | 10 | 2 (20.0) | 8 (80.0) | |
| Lymph node status ^b | | | | NS |
| pN0 | 34 | 6 (17.6) | 28 (82.4) | |
| pN1 | 26 | 6 (23.1) | 20 (76.9) | |
| pN2+pNM | 4 | 0 (0) | 4 (100) | |
| Relapse | | | | NS |
| + | 10 | 1 (10.0) | 9 (90.0) | |
| - | 56 | 13 (23.2) | 43 (79.6) | |
| ER expression | | | | NS |
| + (≥2) | 42 | 6 (14.3) | 36 (85.7) | |
| - (<2) | 24 | 8 (33.3) | 16 (66.7) | |
| PR expression | | | | NS |
| + (≥2) | 42 | 6 (14.3) | 36 (85.7) | |
| - (<2) | 24 | 8 (33.3) | 16 (66.7) | |

^a χ^2 test. NS, not significant. ^bInformation available for 64 patients.

chemotherapy. Before chemotherapy, all 4 of the breast carcinomas strongly expressed hTERT by both ISH and IHC. After chemotherapy, hTERT expression completely disappeared in all 4 cases (Table III). hTERT expression by lymphocytes was detectable by ISH and IHC both before and after chemotherapy in all 4 cases, and the level of expression did not appear to be altered by treatment.

No correlation was observed between hTERT mRNA expression and any of the clinicopathological parameters age, histopathological type, tumor size, lymph node status,

Table III. Relationship of hTERT mRNA and protein expression before and after neoadjuvant chemotherapy.

| Case | Age | Neoadjuvant | hTERT mRNA | | hTERT protein | |
|------|-----|------------------|------------|-------|---------------|-------|
| | | | Before | After | Before | After |
| 1 | 80 | Anastrozole | + | - | + | - |
| 2 | 78 | Anastrozole | + | - | + | - |
| 3 | 35 | FEC ^a | + | - | + | - |
| 4 | 37 | AC ^b | + | - | + | - |

^aFEC, 5FU (500 mg/m²), Epirubicin (70 mg/m²), Cyclophosphamide (500 mg/m²). ^bAC, Doxorubicin (60 mg/m²), Cyclophosphamide (500 mg/m²). Before, before neoadjuvant chemotherapy. After, after neoadjuvant chemotherapy. +, positive; -, negative.

relapses, and the expression of ER and PR. Similarly, there was no correlation between hTERT protein expression and any of these clinicopathological parameters. There was a correlation between hTERT mRNA expression and hTERT protein expression in breast cancers ($p < 0.005$).

Discussion

This study reports a comparison of hTERT mRNA expression by ISH and hTERT protein expression by IHC in tissue sections from breast tumors. hTERT mRNA was detected by ISH in 56 of the 64 breast cancers and in MCF-7 human breast cancer cells. Breast cancer cell nuclei stained strongly positive with the specific anti-sense probe but not with the anti-sense probe treated with RNase (data not shown). Tissue lymphocytes also stained positively with the anti-sense probe, but the stromal cells did not. Expression of hTERT protein was observed by IHC in 52 of the 64 breast cancers. hTERT mRNA and protein expressions were highly correlated in breast cancers ($p < 0.005$). Detection of the hTERT protein by IHC has permitted further analysis of carcinogenesis and cancer diagnosis (34).

In recent years, there has been disagreement over the suitability of telomerase activity as a prognostic biologic marker in breast cancer that may help to differentiate patients for individually based risk-related therapy. Hiyama *et al* (15), in a study of 140 breast cancer specimens with the TRAP assay, found a strong association between telomerase activity and stage classification and observed telomerase activity in 68% of stage I tumors and 95% of stage IV tumors. Potemba *et al* (1), using tissue microarrays, found a statistically significant correlation between tumor-specific survival (overall survival) and hTERT expression in breast cancer. However, some problems in interpretation may affect this apparent consensus. First, some samples of breast cancer tissue may be extensively contaminated by infiltrating lymphocytes during operative manipulations, especially in advanced disease, causing overestimation of telomerase activity and/or hTERT expression. In our previous reports, higher expression levels

of signals for both hTERT mRNA and protein were detected in some sections containing both carcinoma and lymphocytes in thyroid and colorectal cancers (13,24). Secondly, Poremba *et al* (1) used polyclonal antibodies against hTERT protein as a signal for expression. In our hands, polyclonal antibodies against hTERT protein give rise to strong background signals and are not clearly specific for measuring expression in cancer tissues. We have carefully compared the reactivity against hTERT protein of the monoclonal antibody used in the present study with that of some polyclonal antibodies. Use of the monoclonal antibody in IHC allowed clear demonstration of hTERT protein expression, with results similar to those of ISH for hTERT mRNA expression. Furthermore, IHC is technically much easier to perform than ISH, since contamination of samples by RNase is not an issue in IHC.

To the best of our knowledge, this report is the first on the study of hTERT expression in breast cancer as a function of neo-adjuvant treatment. We examined hTERT expression in 4 cases of breast cancer before and after chemotherapy. Before chemotherapy, hTERT was strongly expressed in all 4 carcinomas, but after chemotherapy hTERT expression had completely disappeared in all 4 cases. hTERT expression by lymphocytes was detectable by ISH and IHC both before and after chemotherapy in all 4 cases, and the level of expression did not appear to be altered by treatment.

In conclusion, determination of hTERT mRNA expression by ISH and hTERT protein expression by IHC can be used to obtain information contributing to a histopathological diagnosis during screening of breast cancers. By use of a monoclonal antibody, we could very clearly and sensitively demonstrate hTERT protein expression in breast cancer tissues but not in non-cancerous tissues. We also demonstrated that 4 carcinomas with originally positive immunoreactivity against hTERT protein became negative after neo-adjuvant chemotherapy. These results suggest that determination of hTERT protein by ICH can be used as a potential diagnostic and therapeutic marker to distinguish breast cancers.

Acknowledgments

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References

- Poremba C, Heine B, Diallo R, Heinecke A, Wai D, Schaefer KL, Braun Y, Schuck A, Lanvers C, Bankfalvi A, Kneif S, Thorhorst J, Zuber M, Kochli OR, Mross F, Dieterich H, Sauter G, Stein H, Fogt F and Boecker W: Telomerase as a prognostic marker in breast cancer: high-throughput tissue microarray analysis of hTERT and hTR. *J Pathol* 198: 181-189, 2002.
- Healy KC: Telomere dynamics and telomerase activation in tumor progression: prospects for prognosis and therapy. *Oncol Res* 7: 121-130, 1995.
- Shampay J and Blackburn EH: Tetrahymena micronuclear sequences that function as telomeres in yeast. *Nucleic Acids Res* 17: 3247-3260, 1989.
- Levy MZ, Allsopp RC, Futcher AB, Greider CW and Harley CB: Telomere end-replication problem and cell aging. *J Mol Biol* 225: 951-960, 1992.
- Moyzis RK, Buckingham JM, Cram LS, Dani M, Deaven LL, Jones MD, Meyne J, Ratliff RL and Wu JR: A highly conserved repetitive DNA sequence, (TTAGGG)_n, present at the telomeres of human chromosomes. *Proc Natl Acad Sci USA* 85: 6622-6626, 1988.
- Kim NW, Piatyszek MA, Prowse KR, Harley CB, West MD, Ho PL, Coviello GM, Wright WE, Weinrich SL and Shay JW: Specific association of human telomerase activity with immortal cells and cancer. *Science* 266: 2011-2015, 1994.
- Blackburn EH: Telomerase. *Annu Rev Biochem* 61: 113-129, 1992.
- Shay JW and Bacchetti S: A survey of telomerase activity in human cancer. *Eur J Cancer* 33: 787-791, 1997.
- Meyerson M, Counter CM, Eaton EN, Ellison LW, Steiner P, Caddle SD, Ziaugra L, Beijersbergen RL, Davidoff MJ, Liu Q, Bacchetti S, Haber DA and Weinberg RA: hEST2, the putative human telomerase catalytic subunit gene, is up-regulated in tumor cells and during immortalization. *Cell* 90: 785-795, 1997.
- Nakamura TM, Morin GB, Chapman KB, Weinrich SL, Andrews WH, Lingner J, Harley CB and Cech TR: Telomerase catalytic subunit homologs from fission yeast and human. *Science* 277: 955-959, 1997.
- Kilian A, Bowtell DD, Abud HE, Hime GR, Venter DJ, Keese PK, Duncan EL, Reddel RR and Jefferson RA: Isolation of a candidate human telomerase catalytic subunit gene, which reveals complex splicing patterns in different cell types. *Hum Mol Genet* 6: 2011-2019, 1997.
- Kamatori M, Nakamura KI, Kanauchi H, Obara T, Kawahara M, Mijima Y, Kaminishi M and Takubo K: Consistent decrease in telomere length in parathyroid tumors but alteration in telomerase activity limited to malignancies: a preliminary report. *World J Surg* 26: 1083-1087, 2002.
- Kamatori M, Nakamura K, Hashimoto M, Ogawa T, Kaminishi M and Takubo K: Clinical application of human telomerase reverse transcriptase gene expression in thyroid follicular tumors by fine-needle aspirations using *in situ* hybridization. *Int J Oncol* 22: 985-991, 2003.
- Poremba C, Scheel C, Hero B, Christiansen H, Schaefer KL, Nakayama J, Berthold F, Juergens H, Boecker W and Dockhorn-Dworniczak B: Telomerase activity and telomerase subunits gene expression patterns in neuroblastoma: a molecular and immunohistochemical study establishing prognostic tools for fresh-frozen and paraffin-embedded tissues. *J Clin Oncol* 18: 2582-2592, 2000.
- Hiyama E, Gollahon L, Kataoka T, Kuroi K, Yokoyama T, Gazdar AF, Hiyama K, Piatyszek MA and Shay JW: Telomerase activity in human breast tumors. *J Natl Cancer Inst* 88: 116-122, 1996.
- Suzino T, Yoshida K, Bolodczuk J, Tahara H, Buley J, Marek S, Wells C, Goodison S, Ide T, Suzuki T, Tahara E and Tarin D: Telomerase activity in human breast cancer and benign breast lesions: diagnostic applications in clinical specimens, including fine needle aspirates. *Int J Cancer* 69: 301-306, 1996.
- Nawaz S, Hashizumi TL, Markham NE, Shroyer AL and Shroyer KR: Telomerase expression in human breast cancer with and without lymph node metastases. *Am J Clin Pathol* 107: 542-547, 1997.
- Clark GM, Osborne CK, Levitt D, Wu F and Kim NW: Telomerase activity and survival of patients with node-positive breast cancer. *J Natl Cancer Inst* 89: 1874-1881, 1997.
- Poremba C, Boecker W, Willenbring H, Schaefer KL, Onerbach F, Burger H, Diallo R and Dockhorn-Dworniczak B: Telomerase activity in human proliferative breast lesions. *Int J Oncol* 12: 641-648, 1998.
- Hoos A, Hepp HH, Kaul S, Ahlert T, Basten G and Wallwiener D: Telomerase activity correlates with tumor aggressiveness and reflects therapy effect in breast cancer. *Int J Cancer* 79: 8-12, 1998.
- Roos G, Nilsson P, Cajander S, Nielsen NH, Arnerlov C and Landberg G: Telomerase activity in relation to p53 status and clinico-pathological parameters in breast cancer. *Int J Cancer* 79: 343-348, 1998.
- Mokbel K, Parris CN, Ghilchik M, Williams G and Newbold RF: The association between telomerase, histopathological parameters, and KI-67 expression in breast cancer. *Am J Surg* 178: 69-72, 1999.
- Umbrecht CB, Sherman ME, Dome J, Carey LA, Marks J, Kim N and Sukumar S: Telomerase activity in ductal carcinoma *in situ* and invasive breast cancer. *Oncogene* 18: 3407-3414, 1999.
- Kamatori M, Kanauchi H, Nakamura KI, Kawahara M, Weber TK, Mafune KI, Kaminishi M and Takubo K: Demonstration of human telomerase reverse transcriptase (hTERT) in human colorectal carcinomas by *in situ* hybridization. *Int J Oncol* 20: 15-21, 2002.

25. Kamitori M, Nakamura KI, Ogawa T, Mafune KI, Tatutomi Y, Obara T, Onoda N, Fujiwara M, Izumiya-Shimomura N, Mori M, Kaminishi M and Takubo K: Demonstration of human telomerase reverse transcriptase (hTERT) in human parathyroid tumors by *in situ* hybridization with a new oligonucleotide probe. *Clin Endocrinol* 58: 43-48, 2003.
26. Fukushima M, Shimomura N, Nakamura KI, Kamitori M, Koizumi K, Shimizu K and Takubo K: Demonstration of human telomerase reverse transcriptase by *in situ* hybridization in lung carcinoma. *Oncol Rep* 12: 1227-1232, 2004.
27. Frost M, Bobak JB, Gianani R, Kim N, Weinrich S, Spalding DC, Cass LG, Thompson LC, Enomoto T, Uribe-Lopez D and Shroyer KR: Localization of telomerase hTERT protein and hTR in benign mucosa, dysplasia, and squamous cell carcinoma of the cervix. *Am J Clin Pathol* 114: 726-734, 2000.
28. Kawakami Y, Kitamoto M, Nakanishi T, Yasui W, Tahara E, Nakayama J, Ishikawa F, Tahara H, Ide T and Kajiyama G: Immuno-histochemical detection of human telomerase reverse transcriptase in human liver tissues. *Oncogene* 19: 3888-3893, 2000.
29. Pearson WR and Lipman DJ: Improved tools for biological sequence comparison. *Proc Natl Acad Sci USA* 85: 2444-2448, 1988.
30. Caruthers MH, Beaucage SL, Efcavitch JW, Fisher EF, Goldman RA, De Haseth PL, Mandecki W, Matteucci MD, Rosendahl MS and Szabinsky Y: Chemical synthesis and biological studies on mutated gene-control regions. *Cold Spring Harbor Symp Quant Biol* 47: 411-418, 1982.
31. Tani Y: PCR *in situ* amplification and catalyzed signal amplification; approaches of higher sensitive, non-radioactive *in situ* hybridization. *Acta Histochem Cytochem* 32: 261-270, 1999.
32. Harvey JM, Clark GM, Osborne CK and Allred DC: Estrogen receptor status by immunohistochemistry is superior to the ligand-binding assay for predicting response to adjuvant endocrine therapy in breast cancer. *J Clin Oncol* 17: 1474-1481, 1999.
33. Allred DC, Harvey JM, Berardo M and Clark GM: Prognostic and predictive factors in breast cancer by immunohistochemical analysis. *Mod Pathol* 11: 155-168, 1998.
34. Tahara H, Yasui W, Tahara E, Fujimoto J, Ito K, Tamai K, Nakayama J, Ishikawa F, Tahara E and Ide T: Immunohistochemical detection of human telomerase catalytic component, hTERT, in human colorectal tumor and non-tumor tissue sections. *Oncogene* 18: 1561-1567, 1999.



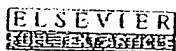
Expression of the ubiquitin-proteasome pathway and muscle loss in experimental cancer cachexia.

Khal J, Wyke SM, Russell ST, Hine AV, Tisdale MJ.

Pharmaceutical Sciences Research Institute, Aston University, Birmingham, UK.

Muscle protein degradation is thought to play a major role in muscle atrophy in cancer cachexia. To investigate the importance of the ubiquitin-proteasome pathway, which has been suggested to be the main degradative pathway mediating progressive protein loss in cachexia, the expression of mRNA for proteasome subunits C2 and C5 as well as the ubiquitin-conjugating enzyme, E2(14k), has been determined in gastrocnemius and pectoral muscles of mice bearing the MAC16 adenocarcinoma, using competitive quantitative reverse transcriptase polymerase chain reaction. Protein levels of proteasome subunits and E2(14k) were determined by immunoblotting, to ensure changes in mRNA were reflected in changes in protein expression. Muscle weights correlated linearly with weight loss during the course of the study. There was a good correlation between expression of C2 and E2(14k) mRNA and protein levels in gastrocnemius muscle with increases of 6-8-fold for C2 and two-fold for E2(14k) between 12 and 20% weight loss, followed by a decrease in expression at weight losses of 25-27%, although loss of muscle protein continued. In contrast, expression of C5 mRNA only increased two-fold and was elevated similarly at all weight losses between 7.5 and 27%. Both proteasome functional activity, and proteasome-specific tyrosine release as a measure of total protein degradation was also maximal at 18-20% weight loss and decreased at higher weight loss. Proteasome expression in pectoral muscle followed a different pattern with increases in C2 and C5 and E2(14k) mRNA only being seen at weight losses above 17%, although muscle loss increased progressively with increasing weight loss. These results suggest that activation of the ubiquitin-proteasome pathway plays a major role in protein loss in gastrocnemius muscle, up to 20% weight loss, but that other factors such as depression in protein synthesis may play a more important role at higher weight loss.

PMID: 16160695 [PubMed - in process]



Increased expression of proteasome subunits in skeletal muscle of cancer patients with weight loss.

Khal J, Hine AV, Fearon KC, Dejong CH, Tisdale MJ.

Pharmaceutical Sciences Research Institute, Aston University, Birmingham B4 7ET, UK.

Atrophy of skeletal muscle is common in patients with cancer and results in increased morbidity and mortality. In order to design effective therapy the mechanism by which this occurs needs to be elucidated. Most studies suggest that the ubiquitin-proteasome proteolytic pathway is most important in intracellular proteolysis, although there have been no reports on the activity of this pathway in patients with different extents of weight loss. In this report the expression of the ubiquitin-proteasome pathway in rectus abdominis muscle has been determined in cancer patients with weight loss of 0-34% using a competitive reverse transcriptase polymerase chain reaction to measure expression of mRNA for proteasome subunits C2 and C5, while protein expression has been determined by western blotting. Overall, both C2 and C5 gene expression was increased by about three-fold in skeletal muscle of cachectic cancer patients (average weight loss 14.5 \pm 2.5%), compared with that in patients without weight loss, with or without cancer. The level of gene expression was dependent on the amount of weight loss, increasing maximally for both proteasome subunits in patients with weight loss of 12-19%. Further increases in weight loss reduced expression of mRNA for both proteasome subunits, although it was still elevated in comparison with patients with no weight loss. There was no evidence for an increase in expression at weight losses less than 10%. There was a good correlation between expression of proteasome 20S α subunits, detected by western blotting, and C2 and C5 mRNA, showing that increased gene expression resulted in increased protein synthesis. Expression of the ubiquitin conjugating enzyme, E2(14k), with weight loss followed a similar pattern to that of proteasome subunits. These results suggest variations in the expression of key components of the ubiquitin-proteasome pathway with weight loss of cancer patients, and suggest that another mechanism of protein degradation must be operative for patients with weight loss less than 10%.

PMID: 16125116 [PubMed - in process]



Cell type-specific occurrence of caveolin-1alpha and -1beta in the lung caused by expression of distinct mRNAs.

Kogo H, Aiba T, Fujimoto T.

Department of Anatomy and Molecular Cell Biology, Nagoya University Graduate School of Medicine, Showa-ku, Nagoya 466-8550, Japan. hkogo@fujita-hu.ac.jp

Two isoforms of caveolin-1, alpha and beta, had been thought to be generated by alternative translation initiation of an mRNA (FL mRNA), but we showed previously that a variant mRNA (5'V mRNA) encodes the beta isoform specifically. In the present study, we demonstrated strong correlation between the expression of the caveolin-1 protein isoforms and mRNA variants in culture cells and the developing mouse lung. The alpha isoform protein and FL mRNA were expressed constantly during the lung development, whereas expression of the beta isoform protein and 5'V mRNA was negligible in the fetal lung before 17.5 days post coitum, and markedly increased simultaneously at 18.5 days post coitum, when the alveolar type I cells started to differentiate. Immunohistochemical analysis revealed the cell type-specific expression of the two isoforms; the alveolar type I cell expresses the beta isoform predominantly, while the endothelium harbors the alpha isoform chiefly. The mutually exclusive expression of caveolin-1 isoforms was verified by Western blotting of the selective plasma membrane preparation obtained from the endothelial and alveolar epithelial cells. The present result indicates that the two caveolin-1 isoforms are generated from distinct mRNAs in vivo and that their production is regulated independently at the transcriptional level. The result also suggests that the alpha and beta isoforms of caveolin-1 may have unique physiological functions.

PMID: 15067006 [PubMed - indexed for MEDLINE]

Oncogene and growth factor expression in ovarian cancer.

Kommoss F, Bauknecht T, Birmelin G, Kohler M, Tesch H, Pfeleiderer A.

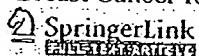
Department of Gynaecology, Albert-Ludwig University, Freiburg, Germany.

The varying tumor-biological behavior of ovarian carcinomas probably influences both their operability and response to chemotherapy, which are the most relevant prognostic factors. The phenotype of different ovarian carcinomas is obviously associated with an activation of the EGF/TGF-alpha signal pathway, including c-myc and c-jun expression. Analysis of EGF-R, TGF-alpha, c-myc and c-jun expression in 33 stage III/IV, and 2 stage I/II ovarian carcinomas with biochemical, molecular-chemical and immunohistochemical methods showed a correlation between the mRNA and protein levels of EGF-R and TGF-alpha for tumors with low or high expressing rates. However, the concentration of measurable free EGF-Rs seems to depend on the amount of TGF-alpha expression by the tumors. The EGF-R binding ligand TGF-alpha is produced by epithelial tumor cells; stromal cells are usually TGF-alpha-negative, as shown by immunohistochemistry. High expression rates of EGF-R, TGF-alpha and c-myc were detected in 6, 7, and 10 out of 35 ovarian carcinomas, respectively. C-jun mRNA was detected in 18/19 cases studied. Non-malignant tissues originating from myometrium or ovary expressed no (or only small amounts of) EGF-R or TGF-alpha mRNA, whereas a high c-myc expression was found in 1/7 normal myometria, and in 2/5 normal ovaries. There was no strong correlation between EGF-R/TGF-alpha and c-myc/c-jun expression. (ABSTRACT TRUNCATED AT 250 WORDS)

PMID: 1502888 [PubMed - indexed for MEDLINE]

15: Breast Cancer Res Treat. 2005 Jul;92(2):175-86.

Related Articles, Links



Somatostatin receptors in primary human breast cancer: quantitative analysis of mRNA for subtypes 1--5 and correlation with receptor protein expression and tumor pathology.

Kumar U, Grigorakis SI, Watt HL, Sasi R, Snell L, Watson P, Chaudhari S.

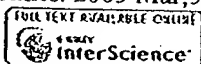
Fraser Laboratories For Diabetes Research, Department of Medicine, McGill University, Royal Victoria Hospital, 687 Pine Avenue West, H3A 1A1 Montreal, Quebec, Canada. ujendra.kumar@mhmc.mcgill.ca

Somatostatin receptors (SSTRs) have been identified in most hormone-producing tumors as well as in breast cancer. In the present study, we determined SSTR1-5 expression in primary ductal NOS breast tumors through semi-quantitative RT-PCR and immunocytochemistry. The results from the analysis of 98 samples were correlated with several key histological markers and receptor expression. All five SSTR subtypes are variably expressed at the mRNA level in breast tumors with 91% of samples showing SSTR1, 98% SSTR2, 96% SSTR3, 76% SSTR4, and 54% SSTR5. SSTR1-5 are localized to both tumor cells and the surrounding peritumoral regions as detected by immunocytochemistry. Levels of SSTR mRNA, when corrected for beta-actin levels, were highest for SSTR3 followed by SSTR1, SSTR2, SSTR5, and SSTR4. Furthermore, there was good correlation between mRNA and protein expression with 84% for SSTR1, 79% for SSTR2, 89% for SSTR3, 68% for SSTR4, 68% for SSTR5, and 78% for all five receptors. SSTR1, 2 and 4 were correlated with ER levels whereas SSTR2 showed an additional correlation with PR levels. These correlations were independent of patient age and histological grade. Moreover, using immunocytochemistry, blood vessels exhibited receptor-specific localization for SSTR2 and SSTR5. Our results indicate significant correlations between mRNA and protein expression along with receptor-specific distribution of SSTR subtypes in tumors and receptor-specific expression in vascular structures may be considered as a novel diagnosis for breast tumors with receptor subtype agonists.

Publication Types:

- Evaluation Studies

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A transcriptomic and proteomic analysis of the effect of CpG-ODN on human THP-1 monocytic leukemia cells.

Kuo CC, Kuo CW, Liang CM, Liang SM.

Institute of BioAgricultural Sciences, Academia Sinica, Taipei, Taiwan.

The CpG motif of bacterial DNA (CpG-DNA) is a potent immunostimulating agent whose mechanism of action is not yet clear. Here, we used both DNA microarray and proteomic approaches to investigate the effects of oligodeoxynucleotides containing the CpG motif (CpG-ODN) on gene transcription and protein expression profiles of CpG-ODN responsive THP-1 cells. Microarray analysis revealed that 2 h stimulation with CpG-ODN up-regulated 50 genes and down-regulated five genes. These genes were identified as being associated with inflammation, antimicrobial defense, transcriptional regulation, signal transduction, tumor progression, cell differentiation, proteolysis and metabolism. Longer stimulation (8 h) with CpG-ODN enhanced transcriptional expression of 58 genes. Among these 58 genes, none except one, namely WNT1 inducible signaling pathway protein 2, was the same as those induced after 2 h stimulation. Proteomic analysis by two-dimensional gel electrophoresis, followed by mass spectrometry identified several proteins up-regulated by CpG-ODN. These proteins included heat shock proteins, modulators of inflammation, metabolic proteins and energy pathway proteins. Comparison of microarray and proteomic expression profiles showed poor correlation. Use of more reliable and sensitive analyses, such as reverse transcriptase polymerase chain reaction, Western blotting and functional assays, on several genes and proteins, nonetheless, confirmed that there is indeed good correlation between mRNA and protein expression after CpG-ODN treatment. This study also revealed that several anti-apoptotic and neuroprotective related proteins, not previously reported, are activated by CpG-DNA. These findings have extended our knowledge on the activation of cells by CpG-DNA and may contribute to further understanding of mechanisms that link innate immunity with acquired immune response(s).

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REGULAR ARTICLE

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A transcriptomic and proteomic analysis of the effect of CpG-ODN on human THP-1 monocytic leukemia cells

Cheng-Chin Kuo¹, Chu-Wei Kuo², Chi-Ming Liang³ and Shu-Mei Liang¹

¹ Institute of BioAgricultural Sciences, Academia Sinica

² Graduate Institutes of Life Sciences, National Defense Medical Center

³ Institute of Biological Chemistry, Academia Sinica, Taipei, Taiwan, Republic of China

The CpG motif of bacterial DNA (CpG-DNA) is a potent immunostimulating agent whose mechanism of action is not yet clear. Here, we used both DNA microarray and proteomic approaches to investigate the effects of oligodeoxynucleotides containing the CpG motif (CpG-ODN) on gene transcription and protein expression profiles of CpG-ODN responsive THP-1 cells. Microarray analysis revealed that 2 h stimulation with CpG-ODN up-regulated 50 genes and down-regulated five genes. These genes were identified as being associated with inflammation, antimicrobial defense, transcriptional regulation, signal transduction, tumor progression, cell differentiation, proteolysis and metabolism. Longer stimulation (8 h) with CpG-ODN enhanced transcriptional expression of 58 genes. Among these 58 genes, none except one, namely WNT1 inducible signaling pathway protein 2, was the same as those induced after 2 h stimulation. Proteomic analysis by two-dimensional gel electrophoresis, followed by mass spectrometry identified several proteins up-regulated by CpG-ODN. These proteins included heat shock proteins, modulators of inflammation, metabolic proteins and energy pathway proteins. Comparison of microarray and proteomic expression profiles showed poor correlation. Use of more reliable and sensitive analyses, such as reverse transcriptase polymerase chain reaction, Western blotting and functional assays, on several genes and proteins, nonetheless, confirmed that there is indeed good correlation between mRNA and protein expression after CpG-ODN treatment. This study also revealed that several anti-apoptotic and neuroprotective related proteins, not previously reported, are activated by CpG-DNA. These findings have extended our knowledge on the activation of cells by CpG-DNA and may contribute to further understanding of mechanisms that link innate immunity with acquired immune response(s).

Keywords:

ADP-ribosylation factor 3 / CpG-ODN / Heat shock protein / Microarray

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Correspondence: Dr. Shu-Mei Liang, Institute of BioAgricultural Sciences, Academia Sinica, Taipei 115, Taiwan, Republic of China
E-mail: smyang@gate.sinica.edu.tw
Fax: +886-2-26515120

Abbreviations: ARF3, ADP-ribosylation factor 3; FPR1, formyl peptide receptor-like 1; HEK293, human embryonic kidney 293 cells; HSP, heat shock protein; IL, interleukin; JNK, c-Jun NH₂-terminal kinase; LPS, lipopolysaccharide; LXR, nuclear receptor subfamily 1; MyD88, myeloid differentiation factor 88; PKC, protein kinase C gamma; PGK, phosphoglycerate kinase; TLR9, toll-like receptor 9; WISP-2, WNT1 inducible signaling pathway protein

1 Introduction

Mammals protect themselves against pathogen infection primarily via innate and adaptive immunity [1]. The innate immune system relies on a set of pattern recognition receptors (e.g., Toll-like receptors) to recognize foreign molecular structures such as lipopolysaccharide (LPS) and bacterial DNA [2, 3]. Innate immune cells recognize these molecular structures and initiate not only innate but also adaptive immunity by producing immunomodulatory cytokines and activating T and B immune cells [1]. Bacterial DNA can directly activate B cells to

proliferate and secrete immunoglobulins in a T cell-independent manner [4–6]. It also induces B cells and monocytes to activate transcription factor NF- κ B and secrete cytokines, including interleukin (IL) 12, tumor necrosis factor α (TNF- α), and interferon α/β [7–10]. The immunostimulatory activity of bacterial DNA has been assigned to unmethylated CpG motifs (GACGTT for murine, GTCGTT for human) [11]. Recent evidence shows that synthetic oligodeoxynucleotides containing a CpG motif (CpG-ODN), like bacterial DNA with the CpG moiety (CpG-DNA), induce potent Th1-like immune responses that are protective against several infectious agents and immune disorders in animal models [12, 13]. Biologically active CpG-ODN, like bacteria DNA, activates macrophages and immature dendritic cells to increase expression of MHC class II and costimulatory molecules, thereby transcribing cytokine mRNAs, and producing pro-inflammatory cytokines including TNF α , IL-1, IL-6 and IL-12 [9, 14–16]. CpG-ODN can therefore serve as an adjuvant and immunomodulator in vaccines against a wide variety of targets, including infectious agents, cancer antigens and allergens [17].

It has been suggested that unmethylated CpG-DNA-mediated immune activation functions through a toll-like receptor 9 (TLR9) signaling pathway [18]. Endocytosis and sequentially endosomal maturation as well as binding of heat shock protein (HSP) 90 to CpG-DNA are essential for induction of TLR9 signal transduction [19, 20]. It has also been shown that recognition of CpG-DNA causes TLR to form a dimer, which recruits the adaptor molecule, myeloid differentiation factor 88 (MyD88), through interaction between their C-terminal Toll/IL-1R domains. This recruitment of MyD88 to the Toll/IL-1R domain of TLR9 initiates a signaling pathway that sequentially involves IL-1R-associated kinase 1 and TNF- α receptor-associated factor 6 [18, 21, 22]. Studies using gene-deficient mice and RAW264.7 cells transiently transfected with dominant-negative forms of these molecules have indicated that the MyD88-mediated signaling pathway is essential for CpG-DNA-induced activation of NF- κ B and c-Jun N-terminal kinase (JNK), as well as subsequent production of cytokines in monocytic cells [18, 21, 22]. The precise mechanism of action of CpG-DNA and CpG-ODN, nonetheless, is still not thoroughly understood. To further elucidate the molecular events after binding of CpG-ODN to TLR9, in this study, we treated CpG-ODN responsive THP-1 cells with CpG-ODN and evaluated changes by using DNA microarray and proteomic approaches. We have discovered up-regulation of more than 50 distinguished genes/proteins and identified induction of several anti-apoptotic and neuroprotecting genes by CpG-ODN treatment.

2 Materials and methods

2.1 Reagents

Phosphorothioate-modified CpG-ODN and GpC-ODN were synthesized by MDBio (Taipei, Taiwan). Human specific ODN sequences are: CpG-ODN, 5'-TCG TCG TTT TGT CGT

TTT GTC GTT-3'; GpC-ODN, 5'-TGC TGC TTT TGT GCT TTT GTG CTT-3'. The mouse specific CpG-ODN sequence is 5'-TCC ATG ACG TTC CTG ATG CT-3'. CHCA was from Sigma (St. Louis, MO, USA).

2.2 Cell culture

Cell lines were obtained from the American Type Culture Collection (Rockville, MD). Mouse RAW264.7 macrophage and human embryonic kidney 293 cells (HEK293) were cultured in DMEM supplemented with 10% heat inactivated fetal bovine serum, 100 U/mL penicillin, 100 μ g/mL streptomycin sulfate, 200 mmol/L L-glutamine, and 50 μ M β -mercaptoethanol in a humidified atmosphere of 5% CO₂ at 37°C. The medium was changed every 2 days for all experiments. Human THP-1 monocytic leukemia cells, which have been shown to express TLR9 and respond to CpG-DNA stimulation [23, 24], were cultured in RPMI1640 with the same supplements as for RAW264.7 cell cultures.

2.3 Human cDNA microarray

Total RNAs extracted from cultured THP-1 cells were isolated with TRIzol (Invitrogen, Leek, The Netherlands) and submitted to Genasia Biotechnology (Taipei, Taiwan) for further processing. In brief, 4 μ g of total RNA from CpG-ODN stimulated, or normal THP-1 cells was labeled with a fluorescence marker (U-vision, (Taipei, Taiwan)). Different colored fluorescence dyes (Cy5 and Cy3) were used to distinguish total RNA from normal and ODN stimulated cells. The labeled RNA was used for hybridization with the Human 1 cDNA microchip from Agilent Technologies (Palo Alto, CA, USA). The chips were scanned and the expression pattern was analyzed using genechip software. Genes showing up-regulation or down-regulation of RNA levels were analyzed and identified on a genomic database as suggested by the manufacturer of the microchip.

2.4 Protein preparation

THP-1 cells were seeded in a 175 cm² tissue culture flask at a density of 10⁶ cells per milliliter in culture medium. The cells were stimulated with or without 1.5 μ M CpG-ODN at defined times and harvested by centrifugation at 4°C, 1000 \times g for 15 min. Cell pellets were washed twice with ice-cold PBS, resuspended and sonicated in extraction buffer containing 25 mM Tris-HCl (pH 7.5), 2 mM β -mercaptoethanol and protease inhibitor cocktail. After centrifugation at 10 000 \times g for 20 min, ammonium sulfate was added to the supernatant until the final concentration reached 50% saturation w/v. The solution was stirred at 4°C for 30 min and centrifuged at 10 000 \times g for 30 min at 4°C. The supernatant fraction was then transferred into a fresh tube, and the precipitated protein pellet solubilized in extraction buffer. To remove salts and other contaminants, the extracts were treated with a pre-cooled (–20°C) solution of 10% TCA in acetone with 0.07% β -mercaptoethanol. Proteins were allowed to precipitate overnight at –20°C.

After centrifugation, the pellet was washed with ice-cold acetone, containing 0.07% β -mercaptoethanol. The supernatant was discarded and the pellet dried in a SpeedVac system (Model AES1010; Savant, Holbrook, NY, USA).

2.5 2-DE

2-DE was performed using an IPGphor IEF and a Hofer DALT vertical unit (Amersham Biosciences, Piscataway, NJ, USA). One milligram of dried protein sample was dissolved in 350 μ L of rehydration buffer solution, containing 7 M urea, 2 M thiourea, 4% w/v CHAPS, 5 mM tributyl phosphine, and 2% IPG and loaded onto an immobilized pH 3–10 linear gradient strip (18 cm), followed by rehydration for 16 h. IEF was then performed in the following manner: 100 V for 30 min, 250 V for 30 min, 500 V for 30 min, 1000 V for 30 min, 4000 V for 30 min, 6000 V for 55 000 Vh. At the end of IEF, the IPG strips were equilibrated for 15 min in buffer containing 6 M urea, 2% w/v SDS, 30% v/v glycerol, and 50 mM Tris, pH 6.8, then reduced with 65 mM dithioerythritol (DTE) and subsequently alkylated with 135 mM iodoacetamide for another 15 min. After equilibration, the IPG strips were immediately placed on top of a 12% SDS-PAGE (1.5 mm, 20 \times 24 cm). The second dimension gels were then overlaid with molten 0.8% agarose solution in SDS electrophoresis buffer. Electrophoresis was performed at 16°C, starting at 10 mA per gel for 1 h, followed by 45 mA per gel until the dye front reached the bottom of the gels.

2.6 Staining and image acquisition

Immediately after electrophoresis, gels were stained with SYPRO Ruby (Molecular Probes, Eugene, OR, USA). In brief, gels were fixed for 30 min in 10% methanol, 7% acetic acid, and then stained overnight in SYPRO Ruby stain. The staining solution was removed and gels were washed in 10% methanol and 7% acetic acid for 3 h. After staining, image acquisition was carried out on a Typhoon 9200 (Amersham Biosciences). To identify a protein, spot detection, quantification and matching of 2-D results were analyzed using ImageMaster software (Amersham Biosciences). The M_r of the proteins were calibrated according to the LMW-SDS Marker Kit (Amersham Biosciences), and their pI values were estimated from the position of the protein spots on the 2-D gel and confirmed with the information supplied by the manufacturer. Since most of the pI values for the truncated proteins had not been reported previously, the pI values of the truncated proteins were estimated from the position of the observed spots. To omit the variation due to the use of separate gels, after background subtraction, the intensity levels of protein spots on each gel were normalized as a proportion of one reference spot, and protein quantities were calculated by integrating the density over the spot area. Protein spots that showed reproducible modulation exceeding ~80% after CpG-ODN treatment in three experiments were further analyzed by MS.

2.7 In-gel digestion with trypsin and extraction of peptides

Protein spots were excised from stained gels and cut into pieces. In brief, gel spots were dehydrated with ACN for 10 min and dried in a vacuum centrifuge. Gel pieces were reswelled with 55 mM DTE in 25 mM ammonium bicarbonate (pH 8.5) at 37°C for 1 h. The solution was then exchanged with alkylation solution, which contained 100 mM iodoacetamide in 25 mM ammonium bicarbonate (pH 8.5), at room temperature for 1 h. After alkylation, the gel pieces were washed twice with 50% ACN in 25 mM ammonium bicarbonate (pH 8.5) for 15 min. The wash solution was discarded and the pieces of gel were dehydrated with ACN for 10 min and dried in a vacuum centrifuge. Tryptic digestion was initiated by reswelling the gel in 25 mM ammonium bicarbonate solution with 25 ng of trypsin (Promega, Madison, WI, USA). After incubation at 37°C for 16 h, tryptic peptides were extracted twice with 50% ACN containing 5% formic acid for 15 min with moderate sonication. The extracted solutions were pooled and evaporated to dryness in a vacuum centrifuge. The dried peptide mixture was dissolved in 0.1% formic acid and used for MS.

2.8 MALDI-Q-TOF MS and protein identification

Tryptic peptides analyses were performed using a Micromass Q-TOF Ultima MALDI (Micromass, Wythenshawe, U.K.) equipped with a 337 nm nitrogen laser and operated in reflection positive ion mode. Peptide mixtures (1 μ L) were premixed with 1 μ L of the matrix (5 mg CHCA in 50% ACN with 0.1% TFA) then spotted onto the MALDI target plate. Mass spectra were acquired for the mass range of 900–3500 Da and the individual spectra from MALDI MS or MS/MS were processed using the Micromass MassLynx 4.0 software. The generated peak list files were used to query the Swiss-Prot database using the MASCOT program (<http://www.matrixscience.com>) with the following parameters: peptide mass tolerance, 50 ppm; MS/MS ion mass tolerance, 0.25 Da; allowance of missed cleavage, 1; and consideration for variable modifications such as oxidation of methionine and carboxyamidomethylation of cysteines. Only significant hits as defined by MASCOT probability analysis were considered initially. In addition, when the PMF matches were between 5 and 9, at least one peptide sequence was manually checked by MALDI MS/MS analysis.

2.9 RT-PCR analysis

cDNA from THP-1 cells was produced with Superscript II reverse transcriptase (Invitrogen) using a oligo(dT)₁₅ primer for 1 h at 42°C. PCR of cDNA was performed using specific primers for the gene of interest and control β -actin. All PCR products were electrophoresed on a 1.5% agarose gel, and DNA bands were visualized by staining the gel with ethidium bromide.

2.10 Immunoblotting

Human THP-1 or mouse macrophage RAW264.7 cells ($5 \times 3 \times 10^6$ /well) were cultured in a six-well culture plate and treated with or without $1.5 \mu\text{M}$ CpG-ODN for the designated times. After stimulation, cells were harvested by centrifugation at $1000 \times g$ for 15 min in a refrigerated centrifuge and washed twice with cold PBS buffer. The cells were lysed on ice for 15 min with 300 μL lysis buffer (Pierce, Rockford, USA), supplemented with protease inhibitor cocktail (Sigma). The lysates were centrifuged at $12000 \times g$ for 15 min at 4°C , and protein concentrations of supernatant were determined using the Bio-Rad Protein Assay (Hercules, CA, USA). The lysates (50 μg of protein/lane) were subjected to 12% SDS-PAGE and transferred to NC membranes (Amersham Biosciences). The membranes were blocked in PBS-0.1% Tween 20 (PBST) containing 5% non-fat skim milk at room temperature for 1 h, followed by staining with anti-ADP-ribosylation factor 3 (ARF-3) monoclonal antibody (0.1 $\mu\text{g}/\text{mL}$; Sigma). The membranes were then incubated with horseradish peroxidase-conjugated secondary antibody (dilution, 1:3000) for 1 h. After washing three times with PBST, specific bands were detected by chemiluminescence according to the manufacturer's protocol (Amersham Biosciences).

2.11 Cell transfection and luciferase assay

HEK293 cells ($5 \times 3 \times 10^6$ /well) were transfected using FuGENE 6 (Roche Molecular Biochemicals, Indianapolis, IN, USA) plus 0.1 μg p5xNF- κB -luc (Stratagene, La Jolla, CA, USA), 0.05 μg pCDNA3.1- β -galactosidase, and pCDNA3.1-hTLR9 overnight. The cells were incubated with or without $1.5 \mu\text{M}$ CpG-ODN for 8 h and then lysed. NF- κB luciferase activity assays were performed as recommended by the manufacturer (Promega). β -galactosidase activity was used to normalize the data.

2.12 Enzyme activity assay

Pyruvate kinase activity was assayed in a solution (1 mL) containing 100 mM Tris-HCl (pH 8.0), 100 mM KCl, 10 mM MgCl_2 , 0.2 mM NADH, 10 mM PEP, 1.5 mM ADP, 1 unit of lactate dehydrogenase, and an appropriate amount of cell lysate from CpG-ODN untreated or treated THP-1 cells. The reaction was monitored at 30°C for a period of time by measuring the decrease in absorbance at 340 nm. PGK activity was assayed in a coupled reaction with glyceraldehyde 3-phosphate dehydrogenase (GAPDH) as described by Lee [25]. In brief, the assay was performed at 30°C in a total volume of 1 mL containing 100 mM Tris-HCl (pH 7.9), 10 mM MgCl_2 , 0.15 mM NADH, 2 mM ATP, 6 mM 3-phosphoglycerate, 0.1 mg/mL BSA, 50 mg of GAPDH, and an appropriate amount of cell lysate. NADH consumption was monitored at 340 nm.

3 Results

3.1 Effect of CpG-ODN on gene expression profiles of human THP-1 cells

To elucidate the effect of CpG-ODN on gene expression, THP-1, a cell line known to express TLR9 and respond to CpG-DNA [23, 24] was cultured with or without CpG-ODN. Since preliminary experiments showed that $1.5 \mu\text{M}$ CpG-ODN caused more contrasting results between normal and CpG-ODN treated cells, $1.5 \mu\text{M}$ CpG-ODN was used throughout the experiments. To distinguish CpG-ODN treated samples from the control, total cellular RNA of normal and CpG-ODN treated cells was isolated and labeled with the fluorescence dyes, Cy5 and Cy3, respectively. The labeled RNA was then used for hybridization with a Human 1 cDNA microchip from Agilent Technologies. Of the 13000 human genes represented on the gene array, a total of 55 genes changed expression significantly after 2 h CpG-ODN treatment. Among these, 50 genes were up-regulated while five genes were down-regulated by a factor ≥ 2 . These genes were sorted by functions and are listed in Tables 1 and 2. They included notably, IL-18 receptor accessory protein, MGA beta gene, thioredoxin, pro-pol-dUTPase polypeptide, Sp140, connexin 59 gene, Grb2-like 2, enoyl-coenzyme A hydratase, propionyl coenzyme A carboxylase, cytochrome P450, and WNT1 inducible signaling pathway protein 2 (WISP-2) etc. The function of these genes are known to be related to inflammatory responses, antimicrobial defense, transcriptional regulation, intracellular signal transduction, tumor progression, cell differentiation, proteolysis etc.

Table 1. Genes up-regulated (≥ 2 fold) in human THP-1 cells after 2 h of CpG-ODN treatment

| Gene name and description (changed fold ≥ 2) | Genebank number |
|---|--------------------|
| Inflammation and receptor | |
| IL-18 receptor accessory protein | AF077346 |
| T cell receptor V beta gene | X58806 |
| MGA, beta gene | U03019 |
| Antigen gene (PA) | M21896 |
| TIED | NM_004791 |
| Platelet activating receptor | AF002986 |
| Antimicrobial defence | |
| Thioredoxin | NM_003329 |
| Pro-Pol-dUTPase polypeptide | AC004748 |
| Nuclear body protein Sp140 | U63420 |
| Transcriptional regulation | |
| Putative transcription factor LUZP | A1986271 |
| General transcription factor II, I, pseudogene 1 | A1700706 |
| Connexin 59 gene | L29277 |
| Basic helix-loop-helix protein class B 1 (BHLHB1) | AF221520 |

Table 1. Continued

| Gene name and description (changed fold ≥ 2) | Genebank number |
|---|--------------------|
| Regulatory protein | |
| Advillin | AF041449 |
| Channel and transport | |
| Small GTP binding protein Rab9 | U44103 |
| FXD domain-containing ion transport regulator 1 | AI125364 |
| Signal transduction | |
| SH3-domain Grb2-like 2 | AF036268 |
| Titin | X90568 |
| Titin associated protein (165 kD protein) | X69089 |
| KIAA1451 protein | AB040884 |
| Vasoactive intestinal peptide receptor | U11087 |
| Enzyme and protease | |
| Enoyl-Coenzyme A hydratase | AI800553 |
| Nephrin, B-type metallopeptidase | U65090 |
| Propionyl Coenzyme A carboxylase | AB011145 |
| Acyloxyacyl hydrolase (neutrophil) | M62840 |
| Cytochrome P450 | U79716 |
| Intestinal alkaline phosphatase | M31008 |
| Tumor progression and cell differentiation | |
| Retinoblastoma 1 | L11910 |
| Human genomic DNA of 9q32 anti-oncogene of flat Epithelium cancer, segment 6/10 | AB036268 |
| WNT1 inducible signaling pathway protein 2 (WISP-2) | AF100780 |
| Structure protein | |
| Collagen, type IV, alpha 6 | D21337 |
| Beta Myosin heavy chain | M58018 |
| Other | |
| Homo sapiens Cri-du-chat region mRNA, clone NIBB11 | U52827 |
| Human mRNA for laminin alpha 5 chain, partial cds. | AB010099 |
| NIK like and Thyroxine-binding globulin precursor | Z83850 |
| Hypothetical protein DKFZp434M0331 | AL137720 |
| Hypothetical protein FLJ11021 similar to splicing factor | AK023985 |
| Hypothetical protein | AL049851 |
| Chromosome 18 open reading frame 1 | NM_004338 |
| Arlapin 1 | AW408785 |
| Zinc finger protein 8 (ZFP8) | M29581 |
| Zinc finger protein 137 (clone pHZ-30) | U09414 |
| Olfactomedin related ER localized protein | AI738468 |
| Cyclin-dependent kinase 8 | BE467537 |
| Integrin, alpha 1 | D87462 |
| KIAA0421 protein | AB007881 |
| KIAA1233 protein | AB033059 |
| Unnamed protein product | AK026362 |
| NBL4 | X75535 |
| BC331191_1 | AAD39268 |

Table 2. Genes down-regulated (≥ 2 fold) in human THP-1 cells after 2 h of CpG-ODN treatment

| Gene name and description (changed fold ≥ 2) | Genebank number |
|---|--------------------|
| Zinc-finger homeodomain protein 4 | BAB03600 |
| Human protein kinase MEKK2b mRNA, complete cds. | AF239798 |
| Glypican 5 | U66033 |
| Human genomic DNA, chromosome 22q11.2, clone N75A12. | AP000362 |
| Collagen, type I, alpha 1 | Z74615 |

Longer stimulation of THP-1 cells with CpG-ODN (8 h) resulted in the up-regulation of 58 genes. These genes included notably IL-10 receptor beta, formyl peptide receptor-like 1 (FPR1), vitamin D receptor, nuclear receptor subfamily 1 (LxR), early B-cell factor, protein kinase C gamma (PKC), Nck, Ash, phospholipase C binding protein (NAP4), phosphoriboxyl pyrophosphate amidotransferase, disheveled 3, WISP-2 *etc.* Analysis of the functions of the 58 up-regulated genes showed that they are associated with anti-inflammation, transcriptional regulation, intracellular signal transduction, tumor progression, cell differentiation, proteolysis, neurodegeneration, neuroprotection *etc.* (Table 3). We also found that the stimulation of THP-1 cells with CpG-ODN for different periods of times resulted in different profiles. Several defense related genes such as IL-18 receptor accessory protein, Pro-Pol-dUTPase polypeptide, Sp140 and connexin 59 were transiently up-regulated at 2 h short stim-

Table 3. Genes up-regulated (≥ 2 fold) in human THP-1 cells after 8 h of CpG-ODN treatment

| Gene name and description (changed fold ≥ 2) | Genebank number |
|---|--------------------|
| Inflammation and receptor | |
| IL-10 receptor (beta) | U08988 |
| Formyl peptide receptor-like 1 (FPR1) | AF081535 |
| Vitamin D receptor | J03258 |
| NMDAR1 | Z32774 |
| CD44 antigen | AW028346 |
| Nuclear receptor subfamily 1 (LxR) | NM_005693 |
| Neuromedin B receptor | M73482 |
| Transcriptional regulation | |
| Early B-cell factor (ebf) | AF208502 |
| Neurogenic differentiation 1 (Neuro D) | AB018693 |
| MAX dimerization protein (NESH protein) | AB037886 |
| Ribosomal protein S6 kinase | AF090421 |
| ASH2L | AB022785 |
| Regulatory protein | |
| Hypothetical protein DKFZp434H0820 | AL137555 |
| Peroxisomal farnesylated protein | X75535 |
| LTBP4 | AF051344 |
| Neuronal pentraxin II | U29195 |

Table 3. Continued

| Gene name and description (changed fold ≥ 2) | Genebank number |
|--|--------------------|
| Channel and transport | |
| Gamma-aminobutyric [61] A receptor | NM_004961 |
| ATP synthase subunit F6 | M37104 |
| Transient receptor potential channel 1 | Z73903 |
| Choroideremia (Rab escort protein 1) | X57637 |
| Signal transduction | |
| Protein kinase C, gamma | Z15114 |
| Regulator of G-protein signaling 5 | A1674877 |
| Nck, Ash and phospholipase C binding protein (NAP4) | AB005216 |
| Highly similar to adenylyl kinase gene | AB016886 |
| Enzyme and protease | |
| Phenylalanine hydroxylase | AA203389 |
| Carboxypeptidase A1 | X67318 |
| Xylulokinase | AK001205 |
| Pancreatic lipase | J05125 |
| Ubiquitin specific protease 12 | AF022789 |
| Transmembrane protease, serine2 | U75329 |
| Aspartate beta-hydroxylase | U03109 |
| Phosphoribosyl pyrophosphate amidotransferase | D13757 |
| Tumor progression and cell differentiation | |
| CDC23 | AF053977 |
| WISP-2 | AF100780 |
| Microseminoprotein, beta | M34376 |
| Dishevelled 3 | NM_004423 |
| Structure protein | |
| Trichohyalin | L09190 |
| Kertain | AF061809 |
| Other | |
| Human transferrin pseudogene | M22376 |
| TIMP-2 | U44383 |
| Collagen-like protein | U67921 |
| Human genomic DNA, chromosome 21q, section 60/105 | AP001716 |
| Human genomic DNA, chromosome 21q, section 64/105 | AP001720 |
| KIAA0136 | D50926 |
| KIAA0379 | AB002377 |
| KIAA0489 | AB007958 |
| KIAA1114 | AL049732 |
| KIAA1451 | AB040884 |
| KIAA0756 | AB018299 |
| Zinc finger protein 267 | AF220492 |
| Hypothetical protein FLJ10633 | AK001495 |
| Hypothetical protein EU01IMAGE 1955967 | AK026108 |
| Myb1 homolog like 1 | AK001893 |
| Antizyme inhibitor | D88674 |
| Disintegrin-like and metalloprotease (reprolysin type) with Thrombospondin type 1 motif, 3 | AB002364 |
| ADP-ribosylation factor 3 (ARF-3) | M74491 |
| Testis specific protein, Y-linked | M98525 |
| Unnamed protein | AK026042 |

Table 4. List of antimicrobial and anti-inflammatory genes modulated by CpG-ODN treatment of THP-1 cells

| Gene name | Genebank number | Expression fold | |
|-------------------------------------|--------------------|-----------------|-----------------|
| | | 2 h | 8 h |
| Connexin 59 gene | L29277 | 2.12 \pm 0.05 | 1.74 \pm 0.08 |
| IL-18 receptor accessory protein | X58806 | 2.32 \pm 0.21 | 1.19 \pm 0.13 |
| Integrin, α 1 | X68742 | 2.01 \pm 0.03 | 1.05 \pm 0.10 |
| Nuclear body protein Sp140 | U63420 | 2.22 \pm 0.11 | 1.39 \pm 0.04 |
| Pro-Pol-dUTPase polyprotein | AC004748 | 2.33 \pm 0.18 | 0.95 \pm 0.03 |
| Thioredoxin | NM_003329 | 2.20 \pm 0.08 | 1.07 \pm 0.01 |
| FPRL1 | AF081535 | 0.95 \pm 0.06 | 2.13 \pm 0.11 |
| IL-10 receptor | U08988 | 1.22 \pm 0.18 | 2.21 \pm 0.07 |
| LxR | NM_005693 | 0.90 \pm 0.03 | 2.37 \pm 0.31 |
| Vitamin D receptor | J03258 | 1.37 \pm 0.23 | 2.39 \pm 0.11 |

Expression fold is designated as the ratio of CpG-ODN treated over control

ulation but were down-regulated thereafter, while anti-inflammatory associated genes such as FPRL1, IL10 receptor, vitamin D receptor and LxR were up-regulated after 8 h stimulation (Tables 1, 3 and 4).

3.2 Verification of the microarray results with RT-PCR or Western blotting

To verify the results from the microarray analysis, we also performed RT-PCR on the up-regulated genes (Fig. 1). Consistent with results obtained in the microarray gene expression analysis, RT-PCR studies showed that the mRNA levels of some selected genes, including ubiquitin specific protease 12, regulator of G-protein signaling 5, NAP4 and ASH2L, were increased in response to CpG-ODN (Fig. 1, Table 3). In addition, the protein expression level of ARF-3

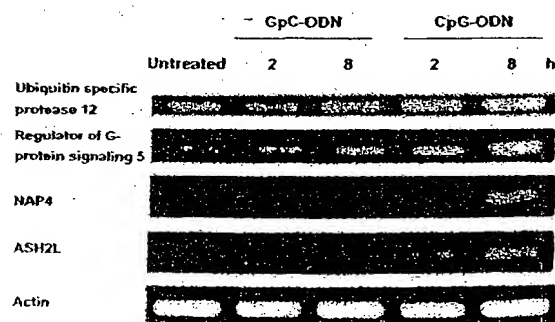


Figure 1. Induction of various genes by CpG-ODN. THP1 cells were stimulated with medium alone, 1.5 μ M GpC-ODN (as the negative control) or CpG-ODN for the indicated times. RT-PCR was then performed to analyze gene expression levels. β -actin was used as an internal control. The experiment was repeated three times with similar results.

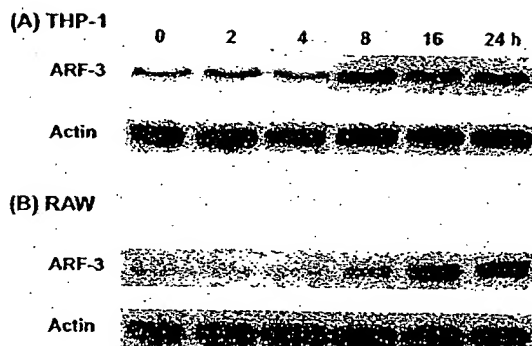


Figure 2. CpG-ODN induced ARF-3 protein expression in human THP-1 or mouse RAW264.7 cells. THP-1 (A) or mouse RAW264.7 (B) cells were incubated with 1.5 μ M CpG-ODN for the indicated time points. The protein expression level of ARF-3 was determined by Western blotting of cell extracts using anti-ARF-3 antibody. The experiment was repeated three times with similar results.

was shown to increase in Western blotting analysis in cell lysates from THP-1 cells treated with CpG-ODN for 8–24 h (Fig. 2A). Similar studies showed that the ARF-3 protein was also induced by mouse specific CpG-ODN in other TLR9 expression cell lines such as the mouse macrophage RAW264.7 cell line (Fig 2B).

3.3 Proteins regulated in CpG-ODN stimulated THP-1 cells

To further assess whether there was any correlation between regulation of gene expression and expression of cellular proteins, a proteomic approach was adopted to identify protein expression profiles. THP-1 cells were treated with CpG-ODN for defined times (from 8 to 40 h), and their cytoplasmic proteins were extracted for 2-DE analysis. Although the use of high concentrations of urea might give us a broader view of all the proteins affected by CpG-ODN, preliminary results from 2-D gels showed that the resolution of the protein mixtures were not satisfactory. To improve and get the best resolution from 2-DE, total proteins were roughly separated into supernatant and precipitated fractions using 50% saturated ammonium sulfate solution. To remove salts and other contaminants, both protein fractions were precipitated with TCA solution and then subjected to 2-DE. By protein spot determination analysis, about 500 and 450 well-resolved spots were observed on each pH 3.0–10.0 gel for precipitated or supernatant fractions, respectively. Comparative analysis of 2-DE between treatments and control showed that the intensities of the protein spots from the ammonium sulfate precipitated fraction did not change, while several protein spots were up-regulated by at least ~80% in the supernatant fraction of 8 h CpG-ODN stimulated THP-1 cells (Fig. 3).

The protein spots were individually excised from gels for further identification. After trypsin digestion, several protein spots were identified without ambiguity by MS MALDI-

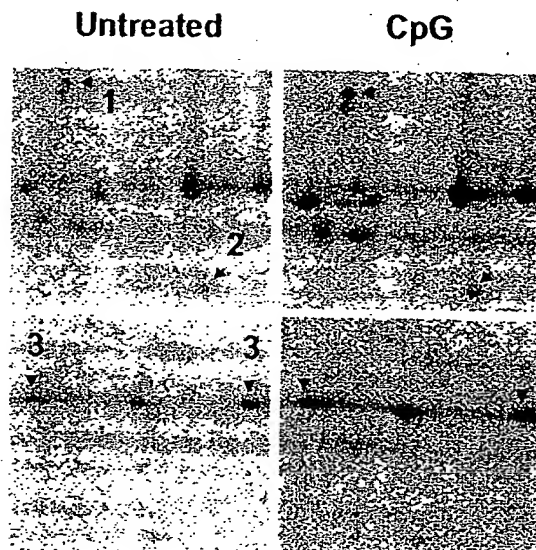


Figure 3. The effect of CpG-ODN on the 2-DE profile of THP-1 cells. THP-1 cells were treated with or without 1.5 μ M CpG-ODN for 8 h. Total proteins were extracted and roughly separated into two fractions by 50% saturation with ammonium sulfate. The supernatant fraction was then separated by 2-DE. Protein spots were visualized by SYPRO Ruby staining. Comparison of CpG-ODN treated THP-1 cells to untreated cells showed that these proteins changed in intensity by over 80%. Protein spots were identified by trypsin digestion and MS. Localization of protein spots 1 (enol-coenzyme A hydratase), 2 (proteasome α) and 3 (cyclophilin A; two isoforms) are shown. The experiment was repeated three times with similar results.

TOF. These proteins included HSP60, HSP90, cyclophilin A, enol-coenzyme A hydratase, eukaryotic translation elongation factor, proteasome α and β chain and ATP synthase beta chain (Table 5). Similar experiments on cells treated for a longer period of time with CpG-ODN stimulation (25 h) revealed that 27 protein spots were changed in intensity by at least ~80%. These protein spots contained members of HSPs (HSP27, hsc70, grp78 and grp94), metabolic enzymes (phosphoglycerate kinase (PGK) and pyruvate kinase (PYK)), macrophage capping protein and cyclophilin A (Table 6). Among these proteins, macrophage capping protein, PGK, PYK, cyclophilin A and HSP27 (Figs. 4 and 5A) were found to be up-regulated. Interestingly, we found that a truncated form of grp78 with an expected mass of 25 kDa and pI of 5.3 was up-regulated while grp78 itself was down-regulated. A similar situation was also found for grp94 and hsc70 and their truncated derivatives (Table 7 and Fig. 5). In addition, we also observed six down-regulated protein spots on 2-D gels in samples after 25 h CpG-ODN treatment. Among these six proteins, we have successfully identified three as 40s ribosomal protein SA, grp78 and hsc70, respectively (Table 6), while the other three, due to their relative low abundance, have not been identified yet.

Table 5. List of proteins modulated by 8 h CpG-ODN treatment

| Protein name | Accession no. | M_r (theor.) | pI (theor.) | Matched no. | Coverage% | Score | Expression fold |
|--|-------------------------|----------------|---------------|-------------|-----------|-------|-----------------|
| ATP synthase beta-chain | gi114549 | 56 525 | 5.26 | 16 | 58 | 171 | 2.33 ± 0.06 |
| Cyclophilin A | P05092 | 17 870 | 7.82 | 5 | 35 | 62 | 2.85 ± 0.13 |
| Enoyl-Coenzyme A hydratase | gi4503447 ^{a)} | 35 971 | 6.61 | 11 | 44 | 62 | 2.52 ± 0.05 |
| Eukaryotic translation elongation factor | gi4503481 | 50 087 | 6.25 | 6 | 25 | 68 | 3.41 ± 0.21 |
| HSP60 | P10809 ^{b)} | 57 963 | 5.24 | 13 | 27 | 76 | 2.78 ± 0.03 |
| HSP90-beta | P08238 | 83 133 | 4.97 | 10 | 18 | 65 | 2.36 ± 0.10 |
| Proteasome α chain | gi4506181 | 25 882 | 6.92 | 11 | 59 | 80 | 2.52 ± 0.11 |
| Proteasome β chain | gi4506193 | 26 472 | 8.27 | 9 | 46 | 84 | 3.85 ± 0.17 |

Expression fold is designated as the ratio of CpG-ODN treated over control

a) NCBI accession number

b) Swiss-Prot accession number

Table 6. List of proteins modulated by 16 and 25 h CpG-ODN treatment

| Protein name | Swiss-Prot no. | M_r (theor.) | pI (theor.) | Matched no. | Coverage% | Score | Expression fold | |
|---|----------------|----------------|---------------|-------------|-----------|-------|-----------------|-----------------|
| | | | | | | | 16 h | 25 h |
| Cyclophilin A | P05092 | 17 870 | 7.82 | 5 | 35 | 62 | 2.53 ± 0.02 | 2.48 ± 0.15 |
| 78 kDa glucose regulated protein (grp78) | P11021 | 72 288 | 5.07 | 13 | 30 | 148 | 0.61 ± 0.04 | 0.29 ± 0.06 |
| HSP27 | P04792 | 22 768 | 5.98 | 12 | 59 | 124 | 1.00 ± 0.01 | 2.61 ± 0.12 |
| Heat shock cognate 70 kDa protein (hsc70) | P11142 | 70 854 | 5.37 | 16 | 34 | 114 | 0.64 ± 0.01 | 0.31 ± 0.03 |
| Macrophage capping protein | P40121 | 38 494 | 5.88 | 9 | 30 | 58 | 1.31 ± 0.01 | 2.58 ± 0.03 |
| Phosphoglycerate kinase | P00558 | 44 284 | 7.052 | 11 | 33 | 71 | 2.32 ± 0.11 | 4.23 ± 0.19 |
| Pyruvate kinase | P14618 | 57 710 | 7.95 | 17 | 32 | 114 | 1.65 ± 0.07 | 2.70 ± 0.12 |
| 40s ribosomal protein SA (RSP40) | P08865 | 32 833 | 4.79 | 5 | 23 | 61 | 0.33 ± 0.02 | 0.35 ± 0.05 |

Table 7. List of truncated proteins detected in THP-1 cells after 25 h CpG-ODN treatment

| Protein name | Swiss-Prot no. | M_r (obs.) | pI (obs.) | Matched no. | Coverage% | Score | Expression fold |
|--|----------------|--------------|-------------|-------------|-----------|-------|-------------------|
| 94 kDa glucose-regulated protein (grp94) | P14625 | ~59 700 | ~5.00 | 12 | 14 | 104 | New ^{a)} |
| Truncated form of grp78 | P11021 | ~25 000 | ~5.30 | 12 | 22 | 75 | New |
| Truncated form of hsc70 | P11142 | ~22 000 | ~5.80 | 12 | 20 | 96 | New |
| Truncated form of hsc70 | P11142 | ~19 000 | ~6.10 | 11 | 18 | 113 | New |

a) New designated proteins detected in the CpG-ODN treated gel but not in the corresponding control gel

3.4 Comparison of microarray and proteomic results

Table 8 shows the expression of six genes and their corresponding proteins that were modulated by 8 h treatment of THP-1 cells with CpG-ODN. Besides enoyl-coenzyme A hydratase, there was poor correlation between the expression

of genes and their corresponding proteins (Table 8), suggesting that more in-depth studies were needed. To further evaluate whether changes observed in protein expression correlated with changes in mRNA levels, we randomly chose two proteins (PGK and PYK) that were induced after 16 h CpG-ODN treatment and determined their mRNA levels by

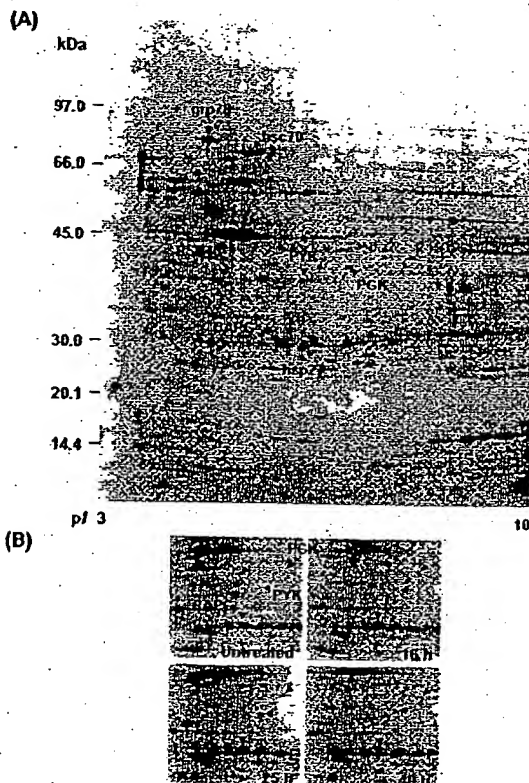


Figure 4. 2-D gel electrophoretic analysis of CpG-ODN-treated THP-1 cells. (A) Total cell protein from unstimulated THP-1 cells was subjected to 2-DE. (B) THP-1 cells were treated with or without 1.5 μ M CpG-ODN for defined times. Cellular proteins were extracted and separated by 2-DE. Several up-regulated proteins are shown in the SYPRO Ruby staining gel. Comparison of CpG-ODN treated THP-1 cells to untreated cells showed that these proteins changed in intensity by over 80%. The experiment was repeated three times with similar results.

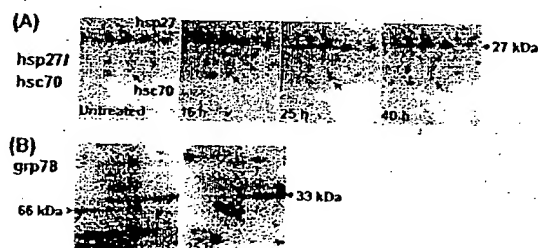


Figure 5. 2-D gel electrophoretic analysis of CpG ODN-treated THP-1 cells. THP-1 cells were treated with or without 1.5 μ M CpG-ODN for defined times. Cellular proteins were extracted and separated by 2-DE. Protein spots were detected by SYPRO Ruby staining. (A) Expression of HSP27 was induced by increasing the period of CpG-ODN stimulation. A truncated form of hsc70 was detected on the gel. (B) The native form of grp78 was detected in untreated cells, while the truncated form of grp78 was observed after 25 h stimulation. The experiment was repeated three times with similar results.

Table 8. Comparison of gene and protein expression levels in THP-1 cells after 8 h CpG-ODN treatment

| Protein name | Gene expression fold from microarray | Protein expression fold from 2-D gel |
|--|--------------------------------------|--------------------------------------|
| Enoyl-Coenzyme A hydratase | 1.72 \pm 0.31 | 2.52 \pm 0.05 |
| Eukaryotic translation elongation factor | 0.95 \pm 0.01 | 3.41 \pm 0.21 |
| HSP60 | 0.94 \pm 0.07 | 2.78 \pm 0.03 |
| HSP90-beta | 1.58 \pm 0.11 | 2.36 \pm 0.10 |
| Proteasome α chain | 0.98 \pm 0.03 | 2.52 \pm 0.11 |
| Proteasome β chain | 1.09 \pm 0.11 | 3.85 \pm 0.17 |

RT-PCR. Our results showed that mRNA levels of PYK increased after 16 h CpG-ODN treatment, while mRNA levels of PGK were dramatically increased after 24 h stimulation (Fig. 6). In addition, we also performed enzyme activity analysis and found that the activity of PGK and PYK were

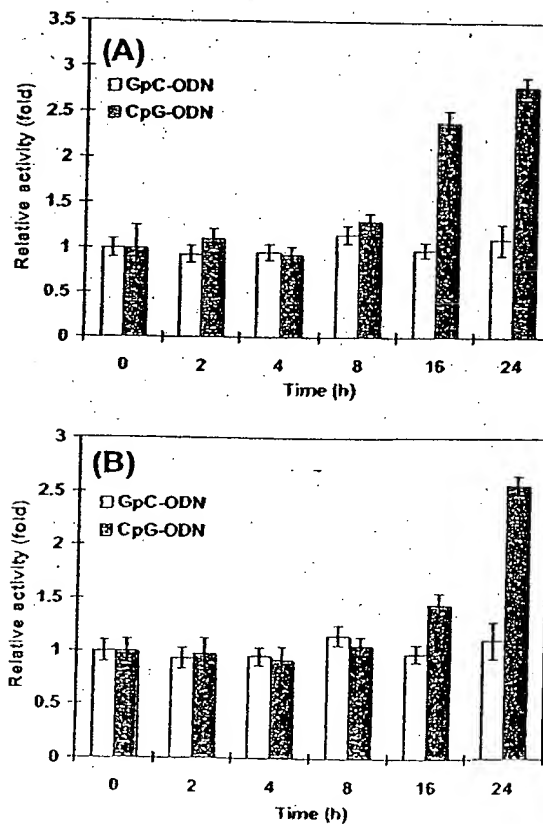


Figure 6. Activities of PYK and PGK induced by CpG-ODN. THP1 cells were stimulated with medium alone, 1.5 μ M GpC-ODN (as the negative control) or CpG-ODN for the indicated times. Cell lysates were extracted and assayed for (A) PYK and (B) PGK activities. Data represent mean \pm SEM. ($n = 3$).

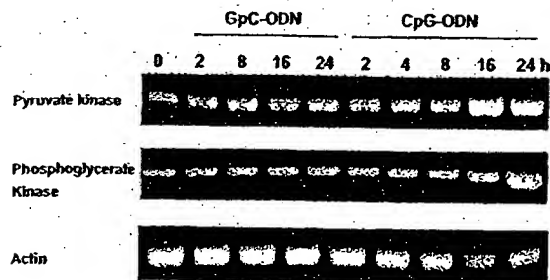


Figure 7. Induction of PYK and PGK transcripts by CpG-ODN. THP1 cells were stimulated with medium alone, 1.5 μ M GpC-ODN or CpG-ODN for the indicated times. RT-PCR was then performed to analyze gene expression. β -actin was used as an internal control. The experiment was repeated three times with similar results.

indeed increased by a factor ~ 2.5 after CpG-ODN stimulation (Fig. 7). To confirm that mRNA induced by CpG-ODN would also be accompanied by an increase in protein expression even though it was not detected in 2-D gel analysis, we used more sensitive and specific Western blotting analysis. As shown in Table 3 and Fig. 2, ARF-3 was identified in the microarray gene profile but not in the proteomic expression profile. Nevertheless, we observed enhanced protein expression of ARF-3 after CpG-ODN stimulation by Western blotting. Moreover, to investigate whether the up-regulation of ARF-3 by CpG-ODN is mediated through the TLR9 pathway, TLR9-deficient HEK293 cells were transiently cotransfected with hTLR9 and luciferase-reporter gene driven by a NF- κ B-dependent promoter. Our data showed that NF- κ B activity of untransfected HEK293 cells were not responsive to CpG-ODN stimulation, while in transfected HEK293 cells expressing hTLR9, NF- κ B luciferase activity was up-regulated 12-fold after 8 h CpG-ODN stimulation. The activation of NF- κ B induced by CpG-ODN was blocked by pretreatment of the transfected cells with an ARF-3 inhibitor, such as brefeldin A (Fig. 8), suggesting CpG-ODN induces ARF-3 and activates NF- κ B after the interaction of CpG-ODN with TLR9.

4 Discussion

In this study both microarray and proteomic approaches were used to evaluate the effect of CpG-ODN on gene/protein expression profiles of THP-1 cells at several time points. Comparison of the gene expression profiles showed that stimulation of the cells with CpG-ODN for different periods of time resulted in different profiles (Tables 1–4). The differences in mRNA expression between the cells with short and long stimulation could be attributable to the low reproducibility. However, to avoid experimental variations, we not only used the same batch of microarrays from the same manufacturer but also applied the samples of short and long term stimulation at the same time. In this way, we found that the

changes in expression fold of mRNA after CpG-ODN treatment were quite reproducible as shown by their mean \pm SEM (Table 4). A more likely explanation for the difference in the expression level of mRNA after different periods of stimulation with CpG-ODN is that the transient increase or decrease in these mRNA by CpG-ODN plays a significant role in modulating biological functions. For example, we found that the IL-18 receptor accessory protein from THP-1 cells was up-regulated after 2 h of CpG-ODN stimulation. The IL-18/IL18R system is known to activate Th1-mediated immune responses that play a critical role in host defense against infection [26]. Together with IL-18/IL18 R, several genes for antimicrobial defense were also increased, including thioredoxin, Pro-Pol-dUTPase polyprotein and Sp140. After 8 h of CpG-ODN stimulation, however, none of these genes was activated any more (Table 4). Since sustained or excessive production of these antimicrobial molecules might lead to inflammation and cellular damage [27], a plausible explanation is that THP-1 cells fight against the invasion of pathogens by up-regulating antimicrobial defense-associated genes at an early stage of stimulation and then shut them down to avoid over-activation. Whether this explanation is true remains to be verified.

It is noteworthy that our data also identified the up-regulation of several anti-inflammatory associated genes after 8 h of CpG-ODN stimulation. These genes included FPRL1, IL-10 receptor, vitamin D receptor, and LxR (Table 3). FPR and FPRL1 have been defined as chemotactic factors involved in host defense against bacterial infection and in the clearance of damaged cells. Additional studies have indicated that FPRL1 interacts with a menagerie of structurally diverse pro- and anti-inflammatory ligands associated with diseases, including amyloidosis, Alzheimer's diseases, prion disease and HIV [28, 29]. Therefore, FPRL1 may play an important role in regulating and/or balancing the production of pro- and anti-inflammatory molecules in CpG stimulated THP-1 cells. Additionally, a recent study has demonstrated that LxRs and their ligands act as negative regulators of macrophage inflammatory gene expression and inhibit the expression of inflammatory mediators such as inducible nitric oxide synthase, cyclooxygenase and IL-6 in response to bacterial infection or LPS stimulation [30]. Of interest, we found that a transcription factor gene connexin 59, a regulator of IL-6 expression, was up-regulated after 2 h of CpG-ODN stimulation. It is thus likely that CpG-ODN stimulation of THP-1 cells for 2 h may induce the expression of the pro-inflammatory cytokine IL-6 through the up-regulation of the connexin 59 gene, while 8 h of CpG-ODN treatment may counterbalance the initial inflammatory response by inducing LxR to inhibit IL-6 production.

Signal transduction molecules play an important role in cellular activation. Intracellular signal transduction systems employing various intermolecular interactions through docking elements, including SH2 and SH3 domains, have been reported [31–33]. Here we found that THP-1 cells treated with CpG-ODN for 2 h up-regulated gene expression

of Grb2-like protein (which contains an SH3 domain), while 8 h of stimulation induced Nck, Ash and phospholipase C binding protein (NAP4 which contains an SH2 domain). It is thus possible that Grb2-like protein and NAP4 may play important roles in CpG-ODN mediated signaling pathways. Furthermore, recent studies have also revealed that binding of CpG-DNA to TLR9 results in activation of JNK [34]. Since JNK is activated by Nck adaptor protein and Nck interacting kinase [35, 36], it is possible that CpG-ODN may activate JNK via up-regulation of NAP4. Although a recent publication described the gene expression profiles of a cultured mouse macrophage cell line after CpG-DNA stimulation [34], their microarray results were only conducted at one time-point (6 h stimulation). Moreover, they did not report the measurement of protein expression profiles in response to CpG-ODN stimulation.

Comparison of the gene and protein expression profiles showed that there was discordance between mRNA and protein levels (Table 8). Similar discordance between the expression pattern of genes and proteins was also reported in other system using different stimuli [37–41]. The discordance between mRNA and protein levels could be due to screening capability such as detection sensitivity, choice of cut-off point, quantitativity of microarray and 2-D gels, as well as time discrepancy between gene and protein expression [39, 40, 42, 43]. Alternatively, it could also be explained by post-transcriptional events, such as alternative splicing or PTM [39, 40, 42, 43]. Another possible explanation is that most of the spots observed in the 2-D gels are isoforms of some proteins. The intensity of each spots does not necessarily represent total amount of a certain protein and thus does not correlate with its mRNA level. Our finding that microarray results correlated better with Western blotting results (e.g., ARF-3 in Fig. 2), an approach more suitable than 2-D gels for determining the total amount rather than isoforms of a given protein, seems to suggest that formation of isoforms should be carefully taken into consideration when one tries to correlate mRNA and protein expression data.

Using a proteomics approach, we found that CpG-ODN treatment up-regulated the expression of many proteins including HSPs, metabolic enzymes, structural proteins, as well as macrophage capping protein, cyclophilin and proteasome α and β chain *etc.* HSPs are the most abundant and ubiquitous soluble intracellular proteins. They are up-regulated by various stressors including temperature, glucose deprivation, microbial infection and cancer [44]. They function as molecular chaperones to prevent protein aggregation and contribute to the folding of nascent and altered proteins. In addition, they are able to regulate immune responses, including production of inflammatory cytokines and chemokines and activation or maturation of immune cells [45, 46]. Beside HSPs, cyclophilin as well as proteasome α and β chain have also been reported to be involved in the immune response [39, 47]; proteasome β chain is consistently up-regulated in human neutrophils following LPS exposure [39]. Our finding that the protein levels of HSPs, cyclophilin, and

proteasome α and β chain were increased after CpG-ODN treatment suggests that these molecules might play a role in the immunostimulating effect of CpG-ODN. To what extent these proteins contribute to the immune responses of the cells to CpG-ODN is currently under study. Proteomic analysis also showed that truncated forms of grp78, grp94 and hsc70 were induced, a phenomenon similar to calreticulin observed by Richards and his coworkers [48]. The expression of full length hsc70 and grp78 were decreased while the levels of their truncated derivatives was increased after CpG-ODN treatment. These results suggest that the degradation of these proteins has been enhanced. We also found that proteasome α and β chains as well as ubiquitin specific protease 12 were increased by CpG-ODN. Whether these enzymes or other enzymes were responsible for the generation of truncated hsc70 and grp78 remains to be elucidated.

Cells rely on multiple signaling pathways to determine their fates of survival, proliferation or apoptosis [49]. In fact, apoptosis plays an important role in regulating pathogen infection. To be able to grow and replicate in the target cells, pathogens may have to block apoptosis. Results from several laboratories have made it clear that HSP70 and HSP27 protect cells not only from heat, but also from most apoptotic stimuli [48, 50] by binding to Akt and subsequently mediating anti-apoptotic activity through activation of Akt [51–53]. Since our data revealed that CpG-ODN induced the expression of HSP90 and HSP27, it is possible that CpG-ODN might prevent apoptosis by up-regulation of HSPs.

Interestingly, our microarray data also showed that CpG-ODN mediated the induction of a set of genes associated with tumor progression and cell proliferation. Among these, one gene, WISP-2, was up-regulated by CpG-ODN after both 2 and 8 h stimulation. WISP genes were first identified as downstream targets of the Wnt-1 β -catenin signaling pathway. They belong to the CCN family of growth factors that have been receiving increasing attention lately due to some of the family members having been reported to be involved in angiogenesis and tumorigenesis [54]. It would be interesting to evaluate whether CpG-ODN plays a role in angiogenesis and tumorigenesis by regulating WISP-2. In addition, we found that some genes associated with neurodegeneration or neuroprotection, such as FPR1, NMDAR (NMDA) receptor, PKC and dishevelled 3 were up-regulated. To our knowledge, this is the first report to suggest an association between these genes and CpG-ODN stimulation. As mentioned above, FPR1 plays a crucial role in proinflammatory aspects of systemic amyloidosis and neurodegenerative disease such as Alzheimer's disease and prion disease [28]. NMDAR, PKC and dishevelled are involved in modulating amyloid precursor protein metabolism, which is central to the pathogenesis of Alzheimer's disease [55–57]. Most notably, recent studies have shown that the TLR4-dependent pathway is involved in neurodegeneration of the central nervous system [58]. Whether CpG-ODN moieties of pathogens play any role in neurodegenerative diseases such as Alzheimer's remains to be elucidated.

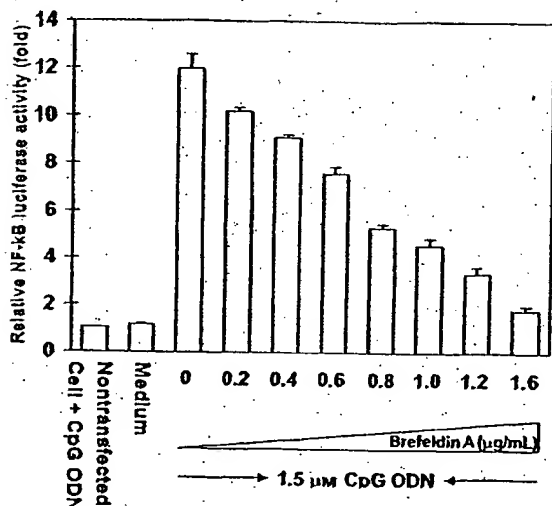


Figure 8. ARF-3 participates in the CpG-ODN-TLR9-NF-κB pathway. HEK293 cells were cotransfected with p5xNF-κB and human TLR9. After overnight transfection, the cells were incubated with or without 1.5 μM CpG-ODN for 8 h in the presence or absence of increasing concentrations of the ARF-3 inhibitor brefeldin A. After incubation, cells were lysed and NF-κB luciferase activity was measured. Data represent mean ± SEM. ($n = 3$).

Exposure of cells to LPS or microbial infection has been known to induce several genes encoding metabolic enzyme [34, 39]. Our microarray data also revealed that a large number of genes encoding proteins involved in energy synthesis and fatty acid oxidation, such as enoyl-coenzyme A hydratase, propionyl coenzyme A carboxylase and cytochrome p450 were activated by CpG-ODN treatment. In addition, we found that other proteins such as ARF-3 were up-regulated (Table 3, Fig. 8). ARFs are 20 kDa GTPases of the ras superfamily that are critical to vesicular trafficking, including exocytic protein transport and endocytosis [59, 60]. This study demonstrates for the first time that ARF-3 is involved in the activation of NF-κB induced by CpG-ODN (as shown in Fig. 8).

CpG-DNA/ODN has been shown to elicit primarily responses via the TLR9/MyD88 dependent pathway [18, 21, 22]. Chromosome location analysis showed that instead of localizing on one or two chromosome, the genes/proteins modulated by CpG-ODN stimulation are scattered on all chromosomes except chromosomes 23 and 24. These results seem to suggest that CpG-ODN either affects multiple chromosomes simultaneously or subsequently via cascades of cellular messengers. More studies are needed to elucidate its mechanism of actions.

5 Concluding remarks

In summary, by using microarray and proteomic approaches to evaluate the effect of CpG-ODN at different time points, we have found that genes/proteins regulated by CpG-ODN

are related to inflammatory responses, antimicrobial defense, transcriptional regulation, intracellular signal transduction, tumor progression, cell differentiation, proteolysis, anti-apoptosis as well as neurodegeneration and neuroprotection. Our results may help delineate the CpG-ODN mediated pathway and contribute to further understanding of mechanisms that link innate immunity with acquired immune response(s).

We thank Mr. Yen-Chieh Huang and Ms. V. R. Kavitha for technical assistance in RT-PCR analysis. We also thank the Core Facilities for Proteomics Research at the Academia Sinica, Taiwan for mass spectrometry analyses. This work was supported by the National Science Council (Grant NSC 91-3112-P001-002-Y) and Academia Sinica (Grant AS 91IBC3PP), Republic of China.

6 References

- [1] Akira, S., Hemmi, H., *Immunol. Lett.* 2003, **85**, 85–95.
- [2] Janeway, C. A. Jr., Medzhitov, R., *Semin. Immunol.* 1998, **10**, 349–350.
- [3] Wagner, H., *Curr. Opin. Microbiol.* 2002, **5**, 62–69.
- [4] Halpern, M. D., Kurlander, R. J., Pisetsky, D. S., *Cell. Immunol.* 1996, **167**, 72–78.
- [5] Messina, J. P., Gilkeson, G. S., Pisetsky, D. S., *J. Immunol.* 1991, **147**, 1759–1764.
- [6] Yi, A. K., Krieg, A. M., *J. Immunol.* 1998, **160**, 1240–1245.
- [7] Tokunaga, T., Yamamoto, H., Shimada, S., Abe, H. et al., *J. Natl. Cancer Inst.* 1984, **72**, 955–962.
- [8] Yamamoto, S., Kuramoto, E., Shimada, S., Tokunaga, T., *Jpn. J. Cancer Res.* 1988, **79**, 866–873.
- [9] Stacey, K. J., Sweet, M. J., Hume, D. A., *J. Immunol.* 1996, **157**, 2116–2122.
- [10] Yi, A. K., Tuetken, R., Redford, T., Waldschmidt, M. et al., *J. Immunol.* 1998, **160**, 4755–4761.
- [11] Krieg, A. M., Yi, A. K., Matson, S., Waldschmidt, T. J. et al., *Nature* 1995, **374**, 546–549.
- [12] Kline, J. N., Waldschmidt, T. J., Businga, T. R., Lemish, J. E. et al., *J. Immunol.* 1998, **160**, 2555–2559.
- [13] Shiota, H., Sano, K., Hirasawa, N., Terui, T. et al., *J. Immunol.* 2001, **167**, 66–74.
- [14] Lipford, G. B., Sparwasser, T., Bauer, M., Zimmermann, S. et al., *Eur. J. Immunol.* 1997, **27**, 3420–3426.
- [15] Sparwasser, T., Miethke, T., Lipford, G., Borschert, K. et al., *Nature* 1997, **386**, 336–337.
- [16] Sparwasser, T., Miethke, T., Lipford, G., Erdmann, A. et al., *Eur. J. Immunol.* 1997, **27**, 1671–1679.
- [17] Takeda, K., Akira, S., *Genes Cells* 2001, **6**, 733–742.
- [18] Hemmi, H., Takeuchi, O., Kawai, T., Kaisho, T. et al., *Nature* 2000, **408**, 740–745.
- [19] Hacker, H., Mischak, H., Miethke, T., Liptay, S. et al., *EMBO J.* 1998, **17**, 6230–6240.
- [20] Bandholtz, L., Guo, Y., Palmberg, C., Mattsson, K. et al., *Cell. Mol. Life Sci.* 2003, **60**, 422–429.

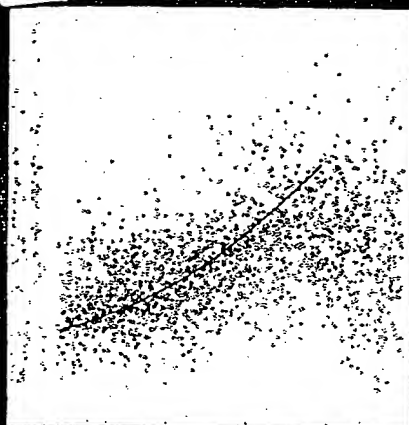
- [21] Bauer, S., Kirschning, C. J., Hacker, H., Redecke, V. et al., *Proc. Natl. Acad. Sci. USA* 2001, 98, 9237–9242.
- [22] Hacker, H., Vabulas, R. M., Takeuchi, O., Hoshino, K. et al., *J. Exp. Med.* 2000, 192, 595–600.
- [23] Takeshita, F., Leifer, C. A., Gursel, I., Ishii, K. J. et al., *J. Immunol.* 2001, 167, 3555–3558.
- [24] Akhtar, M., Watson, J. L., Nazli, A., McKay, D. M., *FASEB J.* 2003, 17, 1319–1321.
- [25] Lee, C. Y., *Methods Enzymol.* 1982, 90 Pt E, 121–126.
- [26] Kawakami, K., *J. Immunother.* 2002, 25 Suppl. 1, S12–19.
- [27] Nakanishi, K., Yoshimoto, T., Tsutsui, H., Okamura, H., *Annu. Rev. Immunol.* 2001, 19, 423–474.
- [28] Le, Y., Oppenheim, J. J., Wang, J. M., *Cytokine Growth Factor Rev.* 2001, 12, 91–105.
- [29] Le, Y., Murphy, P. M., Wang, J. M., *Trends Immunol.* 2002, 23, 541–548.
- [30] Joseph, S. B., Castrillo, A., Laffitte, B. A., Mangelsdorf, D. J., Tontonoz, P., *Nat. Med.* 2003, 9, 213–219.
- [31] Cohen, G. B., Ren, R., Baltimore, D., *Cell* 1995, 80, 237–248.
- [32] Pawson, T., *Nature* 1995, 373, 573–580.
- [33] Birge, R. B., Knudsen, B. S., Besser, D., Hanafusa, H., *Genes Cells* 1996, 1, 595–613.
- [34] Gao, J. J., Diest, V., Wittmann, T., Morrison, D. C. et al., *J. Leukocyte Biol.* 2002, 72, 1234–1245.
- [35] Minden, A., Lin, A., Claret, F. X., Abo, A., Karin, M., *Cell* 1995, 81, 1147–1157.
- [36] Becker, E., Huynh-Do, U., Holland, S., Pawson, T. et al., *Mol. Cell Biol.* 2000, 20, 1537–1545.
- [37] Anderson, L., Seilhamer, J., *Electrophoresis* 1997, 18, 533–537.
- [38] Gygi, S. P., Rochon, Y., Franz, B. R., Aebersold, R., *Mol. Cell Biol.* 1999, 19, 1720–1730.
- [39] Fessler, M. B., Malcolm, K. C., Duncan, M. W., Worthen, G. S., *J. Biol. Chem.* 2002, 277, 31291–31302.
- [40] Kim, C. H., Kim do, K., Choi, S. J., Choi, K. H. et al., *Proteomics* 2003, 3, 2454–2471.
- [41] Scheurer, S. B., Raybak, J. N., Rosli, C., Neri, D., Elia, G., *Proteomics* 2004, 4, 1737–1760.
- [42] Corthals, G. L., Wasinger, V. C., Hochstrasser, D. F., Sanchez, J. C., *Electrophoresis* 2000, 21, 1104–1115.
- [43] Hegde, P. S., White, I. R., Debouck, C., *Curr. Opin. Biotechnol.* 2003, 14, 647–651.
- [44] Robert, J., *Dev. Comp. Immunol.* 2003, 27, 449–464.
- [45] Basu, S., Srivastava, P. K., *Cell Stress Chaperones* 2000, 5, 443–451.
- [46] Berwin, B., Nicchitta, C. V., *Traffic* 2001, 2, 690–697.
- [47] Fluckiger, S., Fijten, H., Whitley, P., Blaser, K., Cramer, R., *Eur. J. Immunol.* 2002, 32, 10–17.
- [48] Richards, J., Le Naour, F., Hanash, S., Beretta, L., *Ann. N. Y. Acad. Sci.* 2002, 975, 91–100.
- [49] Underhill, D. M., Ozinsky, A., *Annu. Rev. Immunol.* 2002, 20, 825–852.
- [50] Somersan, S., Larsson, M., Fonteneau, J. F., Basu, S. et al., *J. Immunol.* 2001, 167, 4844–4852.
- [51] Rane, M. J., Pan, Y., Singh, S., Powell, D. W. et al., *J. Biol. Chem.* 2003, 278, 27828–27835.
- [52] Solit, D. B., Basso, A. D., Olshen, A. B., Scher, H. I., Rosen, N., *Cancer Res.* 2003, 63, 2139–2144.
- [53] Sato, S., Fujita, N., Tsuruo, T., *Proc. Natl. Acad. Sci. USA* 2000, 97, 10832–10837.
- [54] Pennica, D., Swanson, T. A., Welsh, J. W., Roy, M. A. et al., *Proc. Natl. Acad. Sci. USA* 1998, 95, 14717–14722.
- [55] Lipton, S. A., *Cell Death Differ.* 1999, 6, 943–951.
- [56] Du, J., Zhou, S., Coggeshall, R. E., Carlton, S. M., *Neuroscience* 2003, 118, 547–562.
- [57] Mudher, A., Chapman, S., Richardson, J., Asuni, A. et al., *J. Neurosci.* 2001, 21, 4987–4995.
- [58] Lehnardt, S., Massillon, L., Follett, P., Jensen, F. E. et al., *Proc. Natl. Acad. Sci. USA* 2003.
- [59] Lenhard, J. M., Kahn, R. A., Stahl, P. D., *J. Biol. Chem.* 1992, 267, 13047–13052.
- [60] Morinaga, N., Adamik, R., Moss, J., Vaughan, M., *J. Biol. Chem.* 1999, 274, 17417–17423.
- [61] Balog, R. P., de Souza, Y. E., Tang, H. M., DeMasellis, G. M. et al., *Anal. Biochem.* 2002, 309, 301–310.

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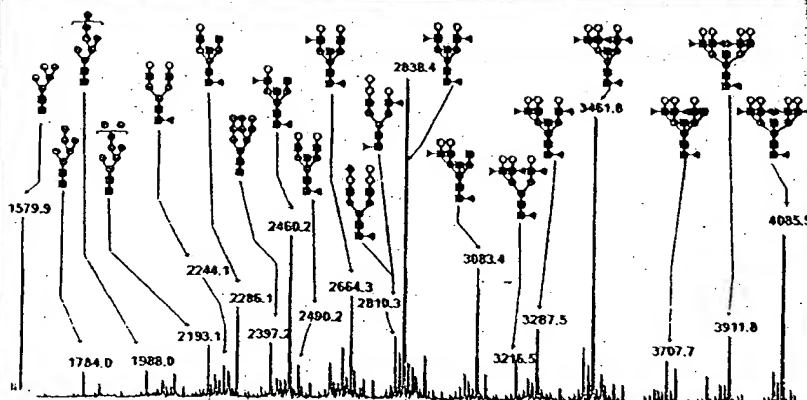
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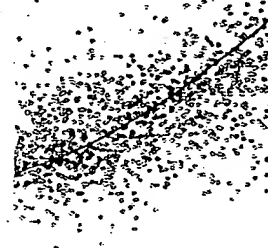
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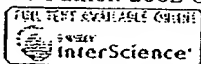
Cellular location and age-dependent changes of the regulatory subunits of cAMP-dependent protein kinase in rat testis.

Landmark BF, Oyen O, Skälhegg BS, Fauske B, Jahnsen T, Hansson V.

Institute of Medical Biochemistry, University of Oslo, Norway.

This study was undertaken to examine the expression and cellular location of the various cAMP-dependent protein kinase (PKA) subunits in different testicular cell types, using cDNA probes, isoenzyme-specific antibodies and activity measurements. Amounts of mRNA and protein were examined in cultured Sertoli cells, cultured peritubular cells, germ cells (pachytene spermatocytes, round spermatids), Leydig cell tumours as well as whole testes from rats of various ages. In Sertoli cells, there was a good correlation between the amount of mRNA and the respective immunoreactive proteins. In other types of cell, such as germ cells and Leydig tumour cells, this was not always the case. Large amounts of RII beta mRNA were found in Leydig tumour cells, whereas the amount of immunoreactive protein was low. Furthermore, large amounts of small-sized, germ cell-specific mRNAs for RI alpha (1.7 kb) and RII alpha (2.2 kb) were also found in the developing rat testis after 30 to 40 days of age, but the large amounts of mRNA were only partially reflected at the protein level. Pachytene spermatocytes and round spermatids were practically devoid of both RII alpha and RII beta protein. During spermatid differentiation, there was a decrease in RI alpha and an increase in RII alpha protein. Cell specific distribution of the various PKA subunits in testicular cell types is described. In some types of cell, discrepancies between mRNA and protein were demonstrated, which clearly suggest cell specific differences in translational efficiencies for some of these mRNAs, particularly the small-sized mRNAs for RI alpha and RII alpha in meiotic and post-meiotic germ cells.

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Quantification of CK20 gene and protein expression in colorectal cancer by RT-PCR and immunohistochemistry reveals inter- and intratumour heterogeneity.

Lassmann S, Bauer M, Soong R, Schreglmann J, Tabiti K, Nahrig J, Ruger R, Hofer H, Werner M.

Pathologisches Institut, Universitätsklinikum Freiburg, Albertstrasse 19, 79104 Freiburg, Germany. lassmann@ukl.uni-freiburg.de

Cytokeratin 20 (CK20) is an epithelial protein expressed almost exclusively in the gastrointestinal (GI) tract and is widely used as immunohistochemical marker for routine diagnosis. In contrast, CK20 gene expression is not an established marker for the classification of tumours and the detection of disseminated cancer cells in colorectal cancer. Recently, real-time reverse transcriptase polymerase chain reaction (RT-PCR) has provided the means for reproducible and quantitative investigation of molecular markers. This report directly compares CK20 mRNA and protein expression in serial sections of archival, formalin-fixed, paraffin-embedded (FFPE) colorectal adenocarcinomas. CK20 expression was detected by immunohistochemistry (IHC) in 60/63 (95.2%) cases, by conventional RT-PCR in 58/60 (96.7%) and by quantitative RT-PCR using the LightCycler (LightCycler is a trademark of a Member of the Roche Group) System in 29/32 (90.6%) microdissected cases, one case yielding variable results. Despite the high detection rate of all three techniques, marked heterogeneity of CK20 expression was seen between different cases and also within individual cases. CK20 expression profiles were not related to particular histopathological features of the tumours. A good correlation ($r = 0.8964$) was found between CK20 mRNA and protein expression by comparing quantitative RT-PCR with IHC in 32 cases. This was also true for selected heterogeneous tumour cells within individual cases. Both RT-PCR and IHC are therefore valuable tools for CK20 detection in colorectal adenocarcinoma, with real-time RT-PCR providing supplementary quantitative information. This suggests a promising supportive role for quantitative RT-PCR in molecular pathology. Copyright 2002 John Wiley & Sons, Ltd.

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- Evaluation Studies

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Expression of the multidrug resistance-associated protein (MRP) mRNA and protein in normal peripheral blood and bone marrow haemopoietic cells.

Legrand O, Perrot JY, Tang R, Simonin G, Gurbuxani S, Zittoun R, Marie JP.

Laboratoire de Cinetique et de Cultures Cellulaires, Hotel Dieu, Paris, France.

We studied the expression of multidrug resistance-associated protein (MRP) in normal haemopoietic cells from peripheral blood and bone marrow. The MRP mRNA levels were estimated by RT/PCR and in situ hybridization (ISH) assay, and the protein levels by flow cytometry. 21 samples of peripheral blood and 21 samples of bone marrow (11 normal bone marrow donors, 10 patients in complete remission after chemotherapy for large cell lymphoma or acute myeloid leukaemia) were analysed. In peripheral blood the mean MRP mRNA level in CD3+ cells was statistically higher than in the other cells (3-fold by the methods used). The levels of MRP in CD3+ varied from one individual to another (4.5-34.8 units by RT/PCR and 5-23 grains/cell by ISH); however, this was proportional to the variation in all the cell lineages of same individual ($r = 0.84$). In bone marrow the mean MRP levels of the various cell lineages (including CD34+) were similar to the basal level in HL60 cells. Individual expression levels were again variable; however, there was no difference between untreated normal bone marrow and post chemotherapy normal bone marrow. MRP protein expression was determined by flow cytometry with the monoclonal antibody MRPM6. The CD4+ lymphocytes exhibited a higher MRP protein expression than the other cell lineages, including CD8+ cells. There was a good correlation between the three methods used (RT/PCR and ISH, $P = 0.0001$, $r = 0.87$; RT/PCR and flow cytometry, $P = 0.0001$, $r = 0.85$; ISH and flow cytometry, $P = 0.002$, $r = 0.67$).

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Vascular endothelial growth factor enhances cardiac allograft arteriosclerosis.

Lemstrom KB, Krebs R, Nykanen AI, Tikkanen JM, Silvola RK, Aaltola EM, Hayry PJ, Wood J, Alitalo K, Yla-Herttuala S, Koskinen PK.

Cardiopulmonary Research Group, Transplantation Laboratory, University of Helsinki and Helsinki University Central Hospital, Helsinki, Finland. Karl.Lemstrom@helsinki.fi

BACKGROUND: Cardiac allograft arteriosclerosis is a complex process of alloimmune response, chronic inflammation, and smooth muscle cell proliferation that includes cross talk between cytokines and growth factors. **METHODS AND RESULTS:** Our results in rat cardiac allografts established alloimmune response as an alternative stimulus capable of inducing vascular endothelial growth factor (VEGF) mRNA and protein expression in cardiomyocytes and graft-infiltrating mononuclear inflammatory cells, which suggests that these cells may function as a source of VEGF to the cells of coronary arteries. Linear regression analysis of these allografts with different stages of arteriosclerotic lesions revealed a strong correlation between intragraft VEGF protein expression and the development of intimal thickening, whereas blockade of signaling downstream of VEGF receptor significantly reduced arteriosclerotic lesions. In addition, in cholesterol-fed rabbits, intracoronary perfusion of cardiac allografts with a clinical-grade adenoviral vector that encoded mouse VEGF(164) enhanced the formation of arteriosclerotic lesions, possibly secondary to increased intragraft influx of macrophages and neovascularization in the intimal lesions. **CONCLUSIONS:** Our findings suggest a positive regulatory role between VEGF and coronary arteriosclerotic lesion formation in the allograft cytokine microenvironment.

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GENIES

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CHAPTER 29

Regulation of transcription

The phenotypic differences that distinguish the various kinds of cells in a higher eukaryote are largely due to differences in the expression of genes that code for proteins, that is, those transcribed by RNA polymerase II. In principle, the expression of these genes might be regulated at any one of several stages. The concept of the "level of control" implies that gene expression is not necessarily an automatic process once it has begun. It could be regulated in a gene-specific way at any one of several sequential steps. We can distinguish (at least) five potential control points, forming the series:

Activation of gene structure
↓
Initiation of transcription
↓
Processing the transcript
↓
Transport to cytoplasm
↓
Translation of mRNA

The existence of the first step is implied by the discovery that genes may exist in either of two structural conditions. Relative to the state of most of the genome, genes are found in an "active" state in the cells in which they are expressed (see Chapter 27). The change of structure is distinct from the act of transcription, and indicates that the gene is "transcribable." This suggests that acquisition of the "active" structure must be the first step in gene expression.

Transcription of a gene in the active state is

controlled at the stage of initiation, that is, by the interaction of RNA polymerase with its promoter. This is now becoming susceptible to analysis in the *in vitro* systems (see Chapter 28). For most genes, this is a major control point; probably it is the most common level of regulation.

There is at present no evidence for control at subsequent stages of transcription in eukaryotic cells, for example, via antitermination mechanisms.

The primary transcript is modified by capping at the 5' end, and usually also by polyadenylation at the 3' end. Introns must be spliced out from the transcripts of interrupted genes. The mature RNA must be exported from the nucleus to the cytoplasm. Regulation of gene expression by selection of sequences at the level of nuclear RNA might involve any or all of these stages, but the one for which we have most evidence concerns changes in splicing; some genes are expressed by means of alternative splicing patterns whose regulation controls the type of protein product (see Chapter 30).

Finally, the translation of an mRNA in the cytoplasm can be specifically controlled. There is little evidence for the employment of this mechanism in adult somatic cells, but it does occur in some embryonic situations, as described in Chapter 7. The mechanism is presumed to involve the blocking of initiation of translation of some mRNAs by specific protein factors.

But having acknowledged that control of gene expression can occur at multiple stages, and that production of RNA cannot inevitably be equated with production of protein, it is clear

that the overwhelming majority of regulatory events occur at the initiation of transcription. Regulation of tissue-specific gene transcription lies at the heart of eukaryotic differentiation; indeed, we see examples in Chapter 38 in which proteins that regulate embryonic development prove to be transcription factors. A regulatory transcription factor serves to provide

common control of a large number of target genes, and we seek to answer two questions about this mode of regulation: what identifies the common target genes to the transcription factor; and how is the activity of the transcription factor itself regulated in response to intrinsic or extrinsic signals?

Response elements identify genes under common regulation

The principle that emerges from characterizing groups of genes under common control is that *they share a promoter element that is recognized by a regulatory transcription factor*. An element that causes a gene to respond to such a factor is called a response element; examples are the HSE (heat shock response element), GRE (glucocorticoid response element), SRE (serum response element).

The properties of some inducible transcription factors and the elements that they recognize are summarized in Table 29.1. Response elements have the same general characteristics as upstream elements of promoters or enhancers. They contain short consensus sequences, and copies of the response elements found in different genes are closely related, but not necessarily identical. The region bound by the factor extends for a short distance on either side of

the consensus sequence. In promoters, the elements are not present at fixed distances from the startpoint, but are usually <200 bp upstream of it. The presence of a single element usually is sufficient to confer the regulatory response, but sometimes there are multiple copies.

Response elements may be located in promoters or in enhancers. Some types of elements are typically found in one rather than the other: usually an HSE is found in a promoter, while a GRE is found in an enhancer. We assume that all response elements function by the same general principle. *A gene is regulated by a sequence at the promoter or enhancer that is recognized by a specific protein. The protein functions as a transcription factor needed for RNA polymerase to initiate. Active protein is available only under conditions when the gene is to be expressed; its absence means that the promoter is not activated by this particular circuit.*

An example of a situation in which many genes are controlled by a single factor is provided by the heat shock response. This is common to a wide range of prokaryotes and eukaryotes and involves multiple controls of gene expression: an increase in temperature turns off transcription of some genes, turns on transcription of the heat shock genes, and causes changes in the translation of mRNAs. The control of the heat shock genes illustrates the differences between prokaryotic and eukaryotic modes of control. In bacteria, a new sigma factor is synthesized that directs RNA polymerase holoenzyme to recognize an alter-

Table 29.1 Inducible transcription factors bind to response elements that identify groups of promoters or enhancers subject to coordinate control.

| Regulatory Agent | Module | Consensus | Factor |
|------------------|--------|-----------------|----------|
| Heat shock | HSE | CNNGAANNTCGNG | HSTF |
| Glucocorticoid | GRE | TGGTACAAATGTTCT | Receptor |
| Phorbol ester | TRE | TGACTCA | AP1 |
| Serum | SRE | CCATATTAGG | SRE |



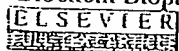
Retinal preconditioning and the induction of heat-shock protein 27.

Li Y, Roth S, Laser M, Ma JX, Crosson CE.

Department of Ophthalmology, Medical University of South Carolina, Charleston, South Carolina 29425, USA.

PURPOSE: Brief periods of ischemia have been shown to protect the retina from potentially damaging periods of ischemia. This phenomenon has been termed ischemic preconditioning or ischemic tolerance. In the present study the cellular changes in levels of heat shock protein (Hsp)27, -70, and -90 mRNA and expression of Hsp in the rat retina associated with ischemic preconditioning were evaluated. **METHODS:** Unilateral retinal ischemia was created in Long-Evans and Sprague-Dawley rats for 5 minutes. Rats were then left for 1 hour to 7 days, to allow the retina to reperfuse. Retinas were dissected, the mRNA and protein isolated, and Northern and Western blot analyses conducted to detect changes in expression of Hsp27, -70, and -90. Immunohistochemical studies were used to identify retinal regions where Hsp changes occurred. Selected animals were subjected to a second ischemic event, 60 minutes in duration, to correlate the changes in expression of Hsp with functional protection of the retina from ischemic injury. **RESULTS:** In control and sham-treated animals retinal Hsp27, -70, and -90 mRNAs were detectable. Five hours after retinal preconditioning, levels of Hsp27 mRNA were elevated above control levels, and 24 hours later, mRNA levels increased 200% over basal levels. Hsp27 expression remained elevated for up to 72 hours and then began to return to control levels. Hsp27 protein levels were increased by 200% over basal levels 24 hours after retinal preconditioning, remained at this level for 72 hours, and then returned to control levels. In contrast, no consistent change in Hsp70 or -90 mRNA or protein levels was observed during the course of the study. Immunohistochemical studies demonstrated that the increase in expression of Hsp27 was localized to neuronal and non-neuronal cells in the inner layers of the retina. Electroretinography studies demonstrated a strong correlation between the protection of retinal function from ischemic injury and the expression of Hsp27. **CONCLUSIONS:** These results provide evidence that the induction of Hsp27 is a gene-specific event associated with ischemic preconditioning in the retina. This increase in expression of Hsp27 occurs in both neuronal and non-neuronal retinal cells, and appears to be one component of the neuroprotective events induced by ischemic preconditioning in the retina.

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Enhanced expressions of arachidonic acid-sensitive tandem-pore domain potassium channels in rat experimental acute cerebral ischemia.

Li ZB, Zhang HX, Li LL, Wang XL.

Institute of Materia Medica, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing 100050, China.

To further explore the pathophysiological significance of arachidonic acid-sensitive potassium channels, RT-PCR and Western blot analysis were used to investigate the expression changes of TREK channels in cortex and hippocampus in rat experimental acute cerebral ischemia in this study. Results showed that TREK-1 and TRAAK mRNA in cortex, TREK-1 and TREK-2 mRNA in hippocampus showed significant increases 2 h after middle cerebral artery occlusion (MCAO). While the mRNA expression levels of the all three channel subtypes increased significantly 24 h after MCAO in cortex and hippocampus. At the same time, the protein expressions of all the three channel proteins showed significant increase 24 h after MCAO in cortex and hippocampus, but only TREK-1 showed increased expression 2 h after MCAO in cortex and hippocampus. Immunohistochemical experiments verified that all the three channel proteins had higher expression levels in cortical and hippocampal neurons 24 h after MCAO. These results suggested a strong correlation between TREK channels and acute cerebral ischemia. TREK channels might provide a neuroprotective mechanism in the pathological process.

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Increasing expression of tissue plasminogen activator and plasminogen activator inhibitor type 2 in dog gingival tissues with progressive inflammation.

Lindberg P, Kinnby B, Lecander I, Lang NP, Matsson L.

Center for Oral Health Sciences, Malmo University, S-214 21 Malmo, Sweden.
pia.lindberg@od.mah.se

Urokinase and tissue-type plasminogen activators (u-PA and t-PA) are serine proteases that convert plasminogen into plasmin, which degrades matrix proteins and activates metalloproteinases. The PAs are balanced by specific inhibitors (PAI-1 and PAI-2). Local production of t-PA and PAI-2 was recently demonstrated in human gingival tissues. The aim now was to investigate the production and localization of t-PA and PAI-2 in gingival tissues from dogs in three well-defined periodontal conditions; clinically healthy gingiva, chronic gingivitis and an initial stage of ligature-induced loss of attachment. At the start of the experiment the gingiva showed clear signs of inflammation. Clinically healthy gingiva were obtained after 21 days period of intense oral hygiene. Attachment loss was induced by placing rubber ligatures around the neck of some teeth. Biopsies were taken from areas representing the different conditions and prepared for in situ hybridization and immunohistochemistry. In clinically healthy gingiva both t-PA mRNA and antigen were expressed in a thin outer layer of the sulcular and junctional epithelia. No t-PA signals or staining were seen in connective tissue. Both mRNA signaling and immunostaining for t-PA were stronger in chronic gingivitis. In areas with loss of attachment, t-PA mRNA as well as antigen were found in the sulcular and junctional epithelia to a similar degree as in gingivitis. Occasionally the connective tissue was involved, especially in connection with vessels. PAI-2 mRNA was seen in a thin outer layer of the sulcular and junctional epithelia in clinically healthy gingiva, but no signals were seen in connective tissue. PAI-2 antigen was found primarily in the outer layer of the sulcular and junctional epithelia. Some cells in the connective tissue were stained. In gingivitis, PAI-2 signals were mainly found in the same locations, but more intense and extending towards the connective tissue. Immunostaining was seen in the outer half of the sulcular and junctional epithelia as well as in the upper part of the connective tissue, close to the sulcular epithelium. In sites with loss of attachment, PAI-2 mRNA was found throughout the sulcular and junctional epithelia, as was the antigen, which stained intensely. No PAI-2 mRNA was seen in connective tissue; the antigen was found scattered, especially near vessels. This study shows that the expression of both t-PA and PAI-2 increases with experimental gingival inflammation in the dog, and furthermore, the two techniques demonstrate a strong correlation between the topographical distribution of the site of protein synthesis and the tissue location of the antigens for both t-PA and PAI-2. The distribution correlates well with previous findings in humans.

Effect of duration of fixation on quantitative reverse transcription polymerase chain reaction analyses.

Macabeo-Ong M, Ginzinger DC, Deldker N, McMillan A, Regezi JA, Wong DT, Jordan RC.

Oral Pathology, Department of Stomatology, University of California San Francisco, California 94143-0424, USA.

Increasingly, there is the need to analyze gene expression in tumor tissues and correlate these findings with clinical outcome. Because there are few tissue banks containing enough frozen material suitable for large-scale genetic analyses, methods to isolate and quantify messenger RNA (mRNA) from formalin-fixed, paraffin-embedded tissue sections are needed. Recovery of RNA from routinely processed biopsies and quantification by the polymerase chain reaction (PCR) has been reported; however, the effects of formalin fixation have not been well studied. We used a proteinase K-salt precipitation RNA isolation protocol followed by TaqMan quantitative PCR to compare the effect of formalin fixation for 24, 48, and 72 hours and for 1 week in normal (2), oral epithelial dysplasia (3), and oral squamous cell carcinoma (4) specimens yielding 9 fresh and 36 formalin-fixed samples. We also compared mRNA and protein expression levels using immunohistochemistry for epidermal growth factor receptor (EGFR), matrix metalloproteinase (MMP)-1, p21, and vascular endothelial growth factor (VEGF) in 15 randomly selected and routinely processed oral carcinomas. We were able to extract RNA suitable for quantitative reverse transcription (RT) from all fresh (9/9) and formalin-fixed (36/36) specimens fixed for differing lengths of time and from all (15/15) randomly selected oral squamous cell carcinoma. We found that prolonged formalin fixation (>48 h) had a detrimental effect on quantitative RT polymerase chain reaction results that was most marked for MMP-1 and VEGF but less evident for p21 and EGFR. Comparisons of quantitative RT polymerase chain reaction and immunohistochemistry showed that for all markers, except p21, there was good correlation between mRNA and protein levels. p21 mRNA was overexpressed in only one case, but protein levels were elevated in all but one tumor, consistent with the established translational regulation of p21. These results show that RNA can be reliably isolated from formalin-fixed, paraffin-embedded tissue sections and can produce reliable quantitative RT-PCR data. However, results for some markers are adversely affected by prolonged formalin fixation times.

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Id-1 and Id-2 are overexpressed in pancreatic cancer and in dysplastic lesions in chronic pancreatitis.

Maruyama H, Kleeff J, Wildt S, Friess H, Buchler MW, Israel MA, Kore M.

Division of Endocrinology, Department of Medicine, University of California, Irvine, USA.

Id proteins antagonize basic helix-loop-helix proteins, inhibit differentiation, and enhance cell proliferation. In this study we compared the expression of Id-1, Id-2, and Id-3 in the normal pancreas, in pancreatic cancer, and in chronic pancreatitis (CP). Northern blot analysis demonstrated that all three Id mRNA species were expressed at high levels in pancreatic cancer samples by comparison with normal or CP samples. Pancreatic cancer cell lines frequently coexpressed all three Ids, exhibiting a good correlation between Id mRNA and protein levels, as determined by immunoblotting with highly specific anti-Id antibodies. Immunohistochemistry using these antibodies demonstrated the presence of faint Id-1 and Id-2 immunostaining in pancreatic ductal cells in the normal pancreas, whereas Id-3 immunoreactivity ranged from weak to strong. In the cancer tissues, many of the cancer cells exhibited abundant Id-1, Id-2, and Id-3 immunoreactivity. Scoring on the basis of percentage of positive cells and intensity of immunostaining indicated that Id-1 and Id-2 were increased significantly in the cancer cells by comparison with the respective controls. Mild to moderate Id immunoreactivity was also seen in the ductal cells in the CP-like areas adjacent to these cells and in the ductal cells of small and interlobular ducts in CP. In contrast, in dysplastic and atypical papillary ducts in CP, Id-1 and Id-2 immunoreactivity was as significantly elevated as in the cancer cells. These findings suggest that increased Id expression may be associated with enhanced proliferative potential of pancreatic cancer cells and of proliferating or dysplastic ductal cells in CP.

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Id-1 and Id-2 Are Overexpressed in Pancreatic Cancer and in Dysplastic Lesions in Chronic Pancreatitis

Haruhisa Maruyama,* Jörg Kleeff,* Stefan Wildi,*
Helmut Friess,[†] Markus W. Büchler,[‡]
Mark A. Israel,[‡] and Murray Korc*

From the Division of Endocrinology, Diabetes, and Metabolism,*
Departments of Medicine, Biological Chemistry and
Pharmacology, University of California, Irvine, California; the
Department of Visceral and Transplantation Surgery,[†] University
of Bern, Bern, Switzerland; and the Preuss Laboratory,[‡]
Department of Neurological Surgery, University of California,
San Francisco, California

Id proteins antagonize basic helix-loop-helix proteins, inhibit differentiation, and enhance cell proliferation. In this study we compared the expression of Id-1, Id-2, and Id-3 in the normal pancreas, in pancreatic cancer, and in chronic pancreatitis (CP). Northern blot analysis demonstrated that all three Id mRNA species were expressed at high levels in pancreatic cancer samples by comparison with normal or CP samples. Pancreatic cancer cell lines frequently coexpressed all three Ids, exhibiting a good correlation between Id mRNA and protein levels, as determined by immunoblotting with highly specific anti-Id antibodies. Immunohistochemistry using these antibodies demonstrated the presence of faint Id-1 and Id-2 immunostaining in pancreatic ductal cells in the normal pancreas, whereas Id-3 immunoreactivity ranged from weak to strong. In the cancer tissues, many of the cancer cells exhibited abundant Id-1, Id-2, and Id-3 immunoreactivity. Scoring on the basis of percentage of positive cells and intensity of immunostaining indicated that Id-1 and Id-2 were increased significantly in the cancer cells by comparison with the respective controls. Mild to moderate Id immunoreactivity was also seen in the ductal cells in the CP-like areas adjacent to these cells and in the ductal cells of small and interlobular ducts in CP. In contrast, in dysplastic and atypical papillary ducts in CP, Id-1 and Id-2 immunoreactivity was as significantly elevated as in the cancer cells. These findings suggest that increased Id expression may be associated with enhanced proliferative potential of pancreatic cancer cells and of proliferating or dysplastic ductal cells in CP. (*Am J Pathol* 1999, 155:815-822)

Basic helix-loop-helix (bHLH) proteins play an important role as transcription factors in cellular development, proliferation, and differentiation.^{1,2} The basic domain of the bHLHs is required for binding to an E-box DNA sequence, thus promoting transcription of specific target genes. The HLH domain promotes dimer formation with various members of the bHLH protein family.^{1,2} Homodimers of the class B family of bHLH proteins, including MyoD, NeuroD, and numerous other proteins, are known to activate tissue-specific genes.³⁻⁵ These tissue-specific bHLHs typically form heterodimers with widely expressed class A bHLHs, which include proteins encoded by E2A, E2-2, HEB, and other genes (also termed E-proteins).⁶⁻⁹ These heterodimers activate transcription of genes that are associated with differentiation.

Id genes encode a family of four HLH proteins that lack the basic DNA binding domain.^{1,10} They act as dominant-negative HLH proteins by forming high affinity heterodimers with other bHLH proteins, thereby preventing them from binding to DNA and inhibiting transcription of differentiation-associated genes.¹⁰⁻¹² Id gene expression is down-regulated on differentiation in many cell types *in vitro* and *in vivo*.¹³⁻¹⁸ In addition, Id proteins seem to be required for cell cycle progression through G₁/S phase in certain cell types, and interaction between Id-2 and pRB is associated with enhanced proliferation in some cell lines *in vitro*.¹⁹⁻²³

Pancreatic cancer is the fifth leading cause of cancer death in the United States, with a mortality rate that virtually equals its incidence rate.²⁴ This malignancy is often associated with the overexpression of a variety of mitogenic growth factors and their receptors, and by oncogenic mutations of K-ras and inactivation of the p53 tumor suppressor gene.²⁵ We have recently reported that pancreatic cancers overexpress the HLH protein Id-2, and that enhanced expression of this protein is evident in the cytoplasm of the cancer cells within the pancreatic tumor mass.²⁶ It is not known, however, whether the expression of other Id proteins is altered in this malignancy, or whether their expression is altered in chronic pancreatitis

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Address reprint requests to Dr. Murray Korc, Division of Endocrinology, Diabetes and Metabolism, Medical Sciences I, C240, University of California, Irvine, CA 92697. E-mail: mkorc@uci.edu.

(CP), an inflammatory disease that is characterized by dysplastic ducts, foci of proliferating ductal cells, acinar cell degeneration, and fibrosis.²⁷ We now report that there is a five- to sixfold increase in Id-1 and Id-2 mRNA levels and a twofold increase in Id-3 mRNA levels in pancreatic cancer by comparison with the normal pancreas. In contrast, overall Id mRNA levels are not increased in CP.

Patients and Methods

Normal human pancreatic tissue samples from 7 male and 5 female donors (median age 41.8 years, range 14–68 years), CP tissues from 13 males and 1 female (median age 42.1 years; range 30–56 years), and pancreatic cancer tissues from 10 male and 6 female donors (median age 62.6 years; range 53–83 years) were obtained through an organ donor program and from surgical specimens from patients with severe symptomatic chronic pancreatitis or pancreatic cancer. A partial duodenopancreatectomy (Whipple/pylorus-preserving Whipple; $n = 13$), a left resection of the pancreas ($n = 2$), or a total pancreatectomy ($n = 1$) were carried out in the pancreatic cancer patients. According to the TNM classification of the Union Internationale Contre le Cancer (UICC) 6 tumors were stage 1, 1 was stage 2, and 9 were stage 3 ductal cell adenocarcinoma. Freshly removed tissue samples were fixed in 10% formaldehyde solution for 12 to 24 hours and paraffin-embedded for histological analysis. In addition, tissue samples were frozen in liquid nitrogen immediately on surgical removal and maintained in -80°C until use for RNA extraction. All studies were approved by the Ethics Committee of the University of Bern, Bern, Switzerland, and by the Human Subjects Committee at the University of California, Irvine, California.

Northern Blot Analysis

Northern blot analysis was carried out as described previously.^{26,28} Briefly, total RNA was extracted by the single step acid guanidinium thiocyanate phenol chloroform method. RNA was size-fractionated on 1.2% agarose/1.8 mol/L formaldehyde gels; electrotransferred onto nylon membranes, and cross-linked by UV irradiation. Blots were prehybridized and hybridized with cDNA probes and washed under high stringency conditions. The following cDNA probes were used: a 979-bp human Id-1 cDNA probe, a 440-bp human Id-2 cDNA probe, and a 450-bp human Id-3 cDNA probe, covering the entire coding regions of Id-1, Id-2, and Id-3, respectively. A *Bam*HI 190-bp fragment of mouse 7S cDNA that hybridizes with human cytoplasmic RNA was used to confirm equal RNA loading and transfer. Blots were then exposed at -80°C to Kodak BioMax-MS films and the resulting autoradiographs were scanned to quantify the intensity of the radiographic bands.^{26,28} For each sample the ratio of Id mRNA expression to 7S expression was calculated. To compare the relative increase in expression of the respective Id mRNA species in the cancer and CP samples, the same normal samples were used for normal/

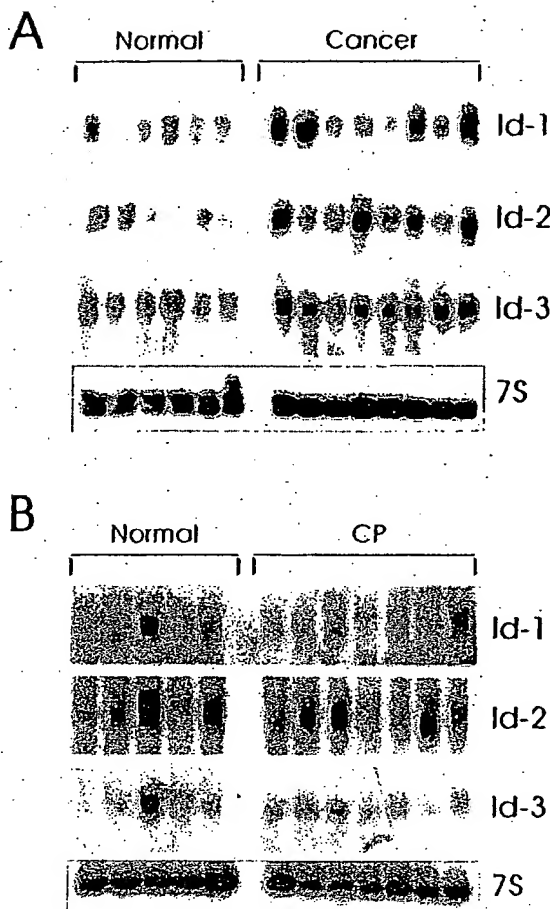


Figure 1. mRNA expression of Id-1, Id-2, and Id-3 in pancreatic cancer and chronic pancreatitis. Total RNA (20 $\mu\text{g}/\text{lane}$) from six normal, eight cancerous, and seven chronic pancreatitis tissue samples were subjected to Northern blot analysis using ^{32}P -labeled cDNA probes (500,000 cpm/ml) specific for Id-1, Id-2, and Id-3, respectively. A 7S cDNA probe (50,000 cpm/ml) was used as a loading and transfer control. Exposure times of the normal/cancer blots were 1 day for all Id probes, and 2 days for the normal/CP blots. Exposure time was 4 hours for mouse 7S cDNA. By comparison with the normal samples, Id-1 and Id-3 mRNA levels were elevated in 8 and 9 cancer samples, respectively, whereas Id-2 was elevated in 6 cancer samples.

cancer and normal/CP membranes. The median score for Id-1, Id-2, and Id-3 mRNA levels in these normal samples was set to 100. Statistical analysis was performed with SigmaStat software (Jandel Scientific, San Raphael, CA). The rank sum test was used, and $P < 0.05$ was taken as the level of significance.

Cell Culture and Western Blot Analysis

PANC-1, MIA-PaCa-2, ASPC-1, and CAPAN-1 human pancreatic cell lines were obtained from ATCC (Manassas, VA). COLO-357 human pancreatic cells were a gift from Dr. R. S. Metzger (Durham, NC). Cells were routinely grown in DMEM (COLO-357, MIA-PaCa-2, PANC-1) or RPMI (ASPC-1, CAPAN-1) supplemented with 10% fetal bovine serum, 100 U/ml penicillin, and 100 $\mu\text{g}/\text{ml}$ streptomycin. For immunoblot analysis, exponentially growing

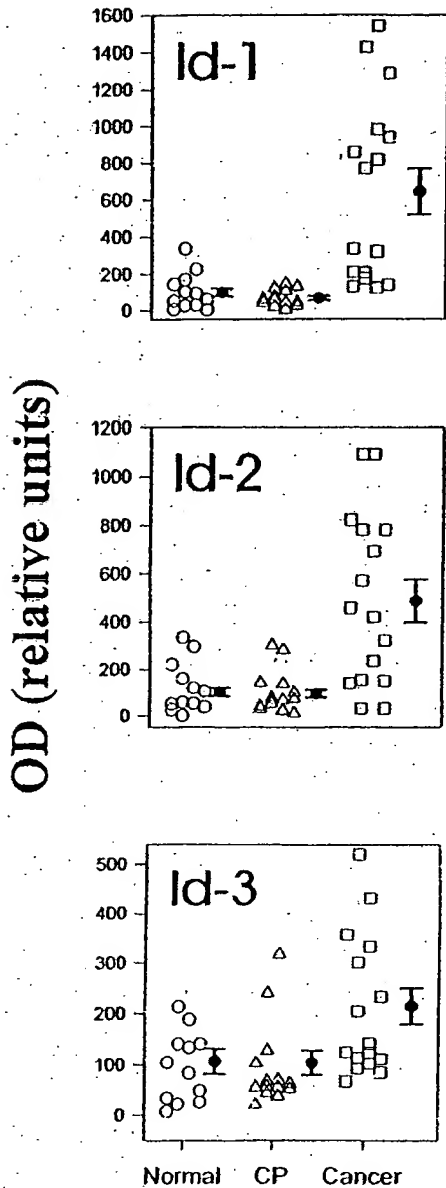


Figure 2. Densitometric analysis of Northern blots. Autoradiographs of Northern blots from 12 normal, 14 CP, and 16 pancreatic cancers were analyzed by densitometry. mRNA levels were determined by calculating the ratio of the optical density for the respective Id mRNA species in relation to the optical density of mouse 7S cDNA. To compare the relative increase in expression of the respective Id mRNA species in the cancer and CP samples, the same normal samples were used for normal/cancer and normal/CP membranes. Normal pancreatic tissues are indicated by circles, CP tissues by triangles, and cancer tissues by squares. Data are expressed as median scores \pm SD. By comparison with the normal samples, only the cancer samples exhibited significant increases: 6.5-fold ($P < 0.01$) for Id-1, fivefold ($P < 0.01$) for Id-2, and twofold ($P = 0.027$) for Id-3.

cells (60–70% confluent) were solubilized in lysis buffer containing 50 mmol/L Tris-HCl, pH 7.4, 150 mmol/L NaCl, 1 mmol/L EDTA, 1 μ g/ml pepstatin A, 1 mmol/L phenylmethylsulfonyl fluoride (PMSF), and 1% Triton X-100. Proteins were subjected to sodium dodecyl sulfate polyacryl-

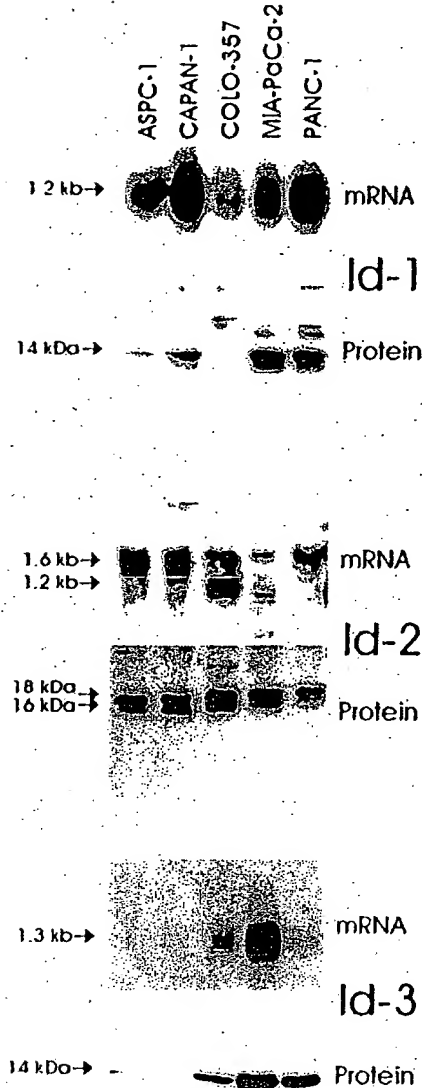


Figure 3. Id mRNA and protein expression in pancreatic cancer cell lines. Upper panels: Total RNA (20 μ g/lane) from 5 pancreatic cancer cell lines were subjected to Northern blot analysis using 32 P-labeled cDNA probes (500,000 cpm/ml) specific for Id-1, Id-2, and Id-3, respectively. Exposure times were 1 day for all Id probes. Lower panels: Immunoblotting. Cell lysates (30 μ g/lane) were subjected to SDS-PAGE. Membranes were probed with specific Id-1, Id-2, and Id-3 antibodies. Visualization was performed by enhanced chemiluminescence.

amide gel electrophoresis (SDS-PAGE), transferred to Immobilon P membranes, and incubated for 90 minutes with the indicated antibodies and for 60 minutes with secondary antibodies against rabbit IgG. Visualization was performed by enhanced chemiluminescence.

Immunohistochemistry

Specific rabbit anti-human Id-1 (C-20), Id-2 (C-20), and Id-3 (C-20; all from Santa Cruz Biotechnology, Santa

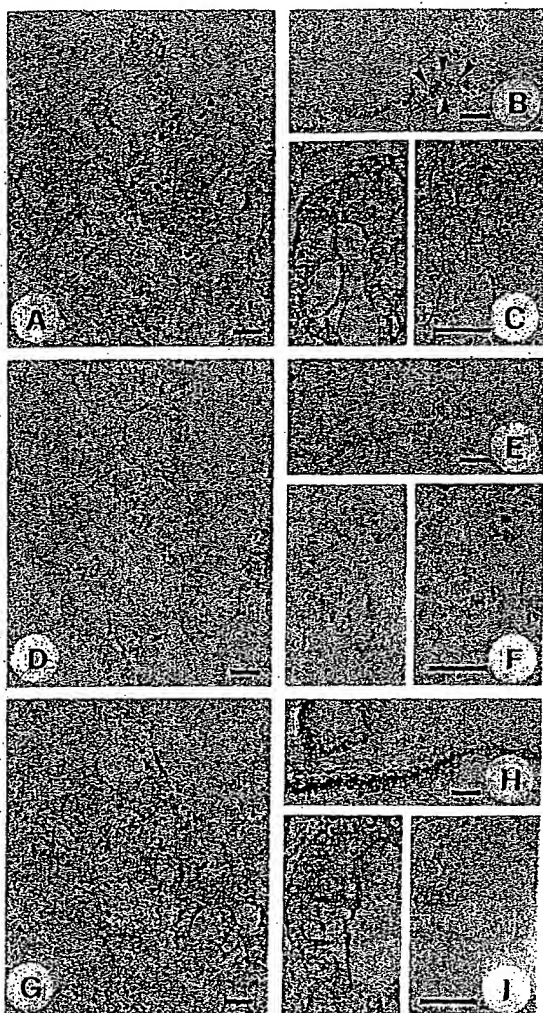


Figure 4. Normal and cancerous pancreatic tissues were subjected to immunostaining using highly specific anti-Id-1 (A-C), anti-Id-2 (D-F), and anti-Id-3 (G-I) antibodies as described in the Methods section. Moderate to strong Id-1 immunoreactivity was present in the cytoplasm of duct-like cancer cells (A and C, left panel). In the normal pancreas there was weak Id-1 immunoreactivity in the ductal cells (B). Preabsorption with the Id-1-specific blocking peptide abolished the Id-1 immunoreactivity (C, right panel). Strong Id-2 immunoreactivity was observed in the cytoplasm of the cancer cells that exhibited duct-like structures (D and F, left panel), whereas in the normal pancreas, there was only weak Id-2 immunoreactivity in the ductal cells (E). Preabsorption with the Id-2-specific blocking peptide abolished the Id-2 immunoreactivity (F, right panel). Moderate to strong Id-3 immunoreactivity was present in the duct-like cancer cells (G and I, left panel). Moderate to strong Id-3 immunoreactivity was also present in the ductal cells of normal pancreatic tissue samples (H). Id-3 immunoreactivity was completely abolished by preabsorption with the Id-3 specific blocking peptide (I, right panel). A, D, and G constitute serial sections of a pancreatic cancer sample, revealing coexpression of the three Id proteins. Scale bars, 25 μ m.

Cruz, CA) polyclonal antibodies were used for immunohistochemistry. These affinity-purified rabbit polyclonal antibodies specifically react with Id-1, Id-2, and Id-3, respectively, of human origin, as determined by Western blotting. Paraffin-embedded sections (4 μ m) were subjected to immunostaining using the streptavidin-peroxidase technique. Where indicated, immunostaining for all three Id proteins was performed on serial sections. En-

dogenous peroxidase activity was blocked by incubation for 30 minutes with 0.3% hydrogen peroxide in methanol. Tissue sections were incubated for 15 minutes (23°C) with 10% normal goat serum and then incubated for 16 hours at 4°C with the indicated antibodies in PBS containing 1% bovine serum albumin. Bound antibodies were detected with biotinylated goat anti-rabbit IgG secondary antibodies and streptavidin-peroxidase complex, using diaminobenzidine tetrahydrochloride as the substrate. Sections were counterstained with Mayer's hematoxylin. Preabsorption with Id-1-, Id-2-, or Id-3-specific blocking peptides completely abolished immunoreactivity of the respective primary antibody. The immunohistochemical results were semiquantitatively analyzed as described previously.^{29,30} The percentage of positive cancer cells was stratified into four groups: 0, no cancer cells exhibiting immunoreactivity; 1, <33% of the cancer cells exhibiting immunoreactivity; 2, 33 to 67% of the cancer cells exhibiting immunoreactivity; 3 >67% of the cancer cells exhibiting immunoreactivity. The intensity of the immunohistochemical signal was also stratified into four groups: 0, no immunoreactivity; 1, weak immunoreactivity; 2, moderate immunoreactivity; 3, strong immunoreactivity. Finally, the sum of the results of the cell score and the intensity score was calculated. Statistical analysis was performed with SigmaStat software. The rank sum test was used, and $P < 0.05$ was taken as the level of significance.

Results

Northern blot analysis of total RNA isolated from 12 normal pancreatic tissues and 16 pancreatic cancers revealed the presence of the 1.2-kb Id-1 transcript and the 1.6-kb Id-2 mRNA transcript in 11 of the 12 normal pancreatic samples, and the 1.3-kb Id-3 mRNA transcript in all normal pancreatic samples (Figure 1A, 2). In the cancer tissues, Id-1 mRNA levels were elevated in 8 of 16 samples, Id-2 mRNA levels were elevated in 9 of these samples, and Id-3 mRNA levels were elevated in 6 of these samples (Figure 1A, 2). Concomitant overexpression of all three Id species was observed in 6 of the cancer samples (38%). In contrast, none of the Id mRNA species were overexpressed in CP by comparison with normal controls (Figure 1B, 2). Densitometric analysis of all of the autoradiograms indicated that there was a 6.5-fold increase ($P < 0.01$) in Id-1 mRNA levels, a fivefold increase ($P < 0.01$) in Id-2 mRNA levels, and a twofold increase ($P = 0.027$) in Id-3 mRNA levels in the pancreatic cancer samples in comparison to normal controls (Figure 2). In contrast, there was no statistically significant difference in the expression levels of Id-1, Id-2, and Id-3, in CP tissues in comparison to the corresponding levels in the normal pancreas (Figure 2).

Next, we assessed the expression of the three Id genes in 5 human pancreatic cancer cell lines by Northern and Western blot analyses. Id-1 mRNA was present at varying levels in all 5 cell lines (Figure 3). ASPC-1, CAPAN-1, MIA-PaCa-2, and PANC-1 expressed moderate to high levels of Id-1 mRNA, whereas COLO-357 cells

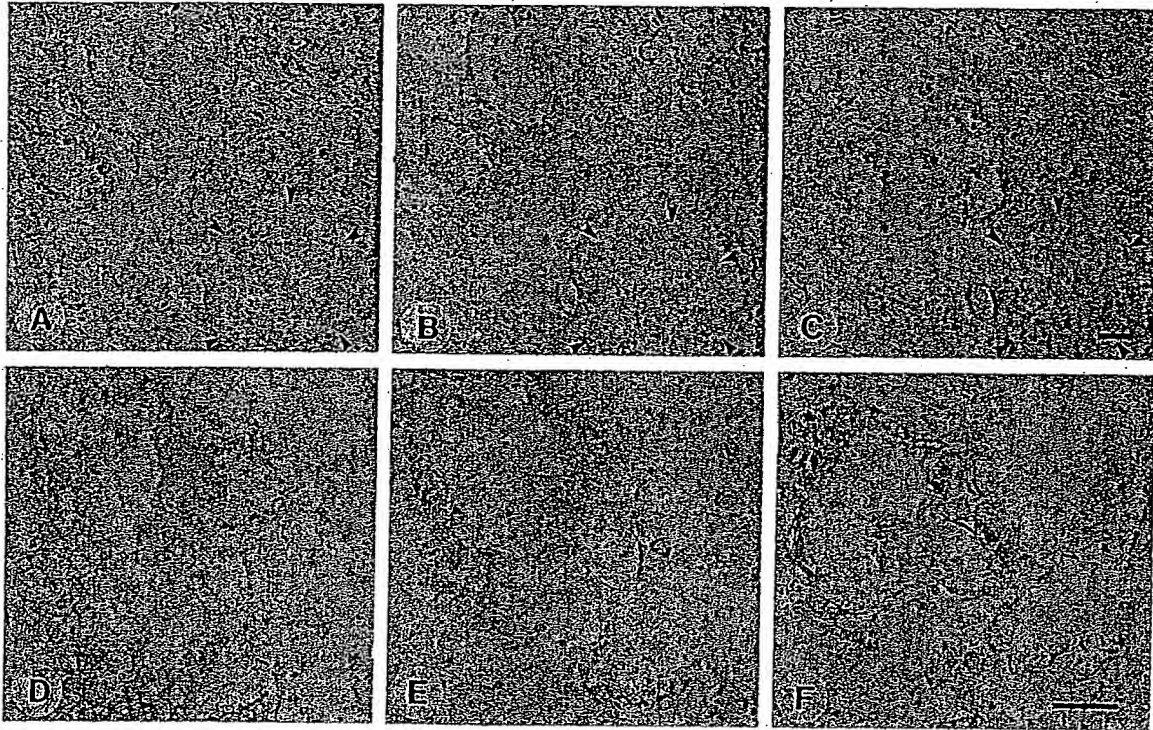


Figure 5. Immunohistochemistry of pancreatic cancer and dysplastic ducts in CP tissues. In the pancreatic cancer tissues (A-C) there was moderate to strong Id-1 (A), Id-2 (B), and Id-3 (C) immunoreactivity in the ductal cells in the areas adjacent to the cancer cells that exhibited CP-like alterations. Islet cells did not exhibit Id immunoreactivity (outlined by solid arrowheads). In the CP samples, moderate to strong Id-1 (D), Id-2 (E), and Id-3 (F) immunoreactivity was present in the cytoplasm of epithelial cells forming large dysplastic ducts. Scale bar, 25 μ m.

expressed relatively low levels of this mRNA moiety. Western blotting with a highly specific anti-Id-1 antibody confirmed the presence of the approximately 14-kd Id-1 protein in the 4 cell lines that expressed high levels of Id-1 mRNA (Figure 3). Furthermore, the three cell lines with the highest Id-1 mRNA expression (CAPAN-1, MIA-PaCa-2, and PANC-1) also exhibited the highest Id-1 protein expression. Variable levels of the 1.6-kb Id-2 mRNA transcript were present in all 5 cell lines. In addition, a minor band of approximately 1.2 kb was visible in COLO-357 and MIA-PaCa-2 cells. Immunoblot analysis with a highly specific anti-Id-2 antibody revealed two bands of approximately 16 and 18 kd at relatively high levels in all of the cell lines with exception of PANC-1 cells, in which the 16-kd band was relatively faint (Figure 3). With the exception of MIA-PaCa-2 cells, there was a good correlation between Id-2 mRNA and protein levels (Figure 3). Id-3 mRNA was present at high levels in MIA-PaCa-2 cells, at moderate levels in COLO-357 cells, and at low levels in PANC-1 cells. Id-3 mRNA was not detectable in ASPC-1 and CAPAN-1 cells (Figure 3). Immunoblot analysis with a highly specific anti-Id-3 antibody revealed an approximately 14-kd band that was most abundant in MIA-PaCa-2 cells, and was also readily apparent in COLO-357 and PANC-1 cells. In contrast, only a faint Id-3 band was seen in ASPC-1 and CAPAN-1 cells. Thus, with the exception of PANC-1 cells, there was a good correlation between Id-3 mRNA and protein levels.

To determine the localization of Id-1, Id-2, and Id-3, immunostaining was carried out using the same highly specific anti-Id antibodies. In the pancreatic cancers, moderate to strong Id-1 immunoreactivity was present in the cancer cells in 9 of 10 randomly selected cancer samples. An example of moderate Id-1 immunoreactivity is shown in Figure 4A, and of strong immunoreactivity in Figure 4C (left panel). In contrast, in the normal pancreas, faint Id-1 immunoreactivity was present only in the ductal cells of pancreatic ducts (Figure 4B, arrowheads). Preabsorption with the Id-1-specific blocking peptide completely abolished the Id-1 immunoreactivity (Figure 4C, right panel). The cancer cells also exhibited strong Id-2 (Figure 4, D and F, left panel) and moderate to strong Id-3 immunoreactivity. An example of moderate Id-3 immunoreactivity is shown in Figure 4G, and of strong immunoreactivity in Figure 4I (left panel). In contrast, only faint Id-2 immunoreactivity was present in the ductal cells in the normal pancreas (Figure 4E), whereas Id-3 immunoreactivity in these cells was more variable and ranged from moderate to occasionally strong (Figure 4H). Islet cells and acinar cells were always devoid of Id immunoreactivity. Preabsorption of the respective antibody with the blocking peptides specific for Id-2 (Figure 4F, right panel) and Id-3 (Figure 4I, right panel) completely abolished immunoreactivity. Analysis of serial pancreatic cancer sections revealed that there was often colocalization of the

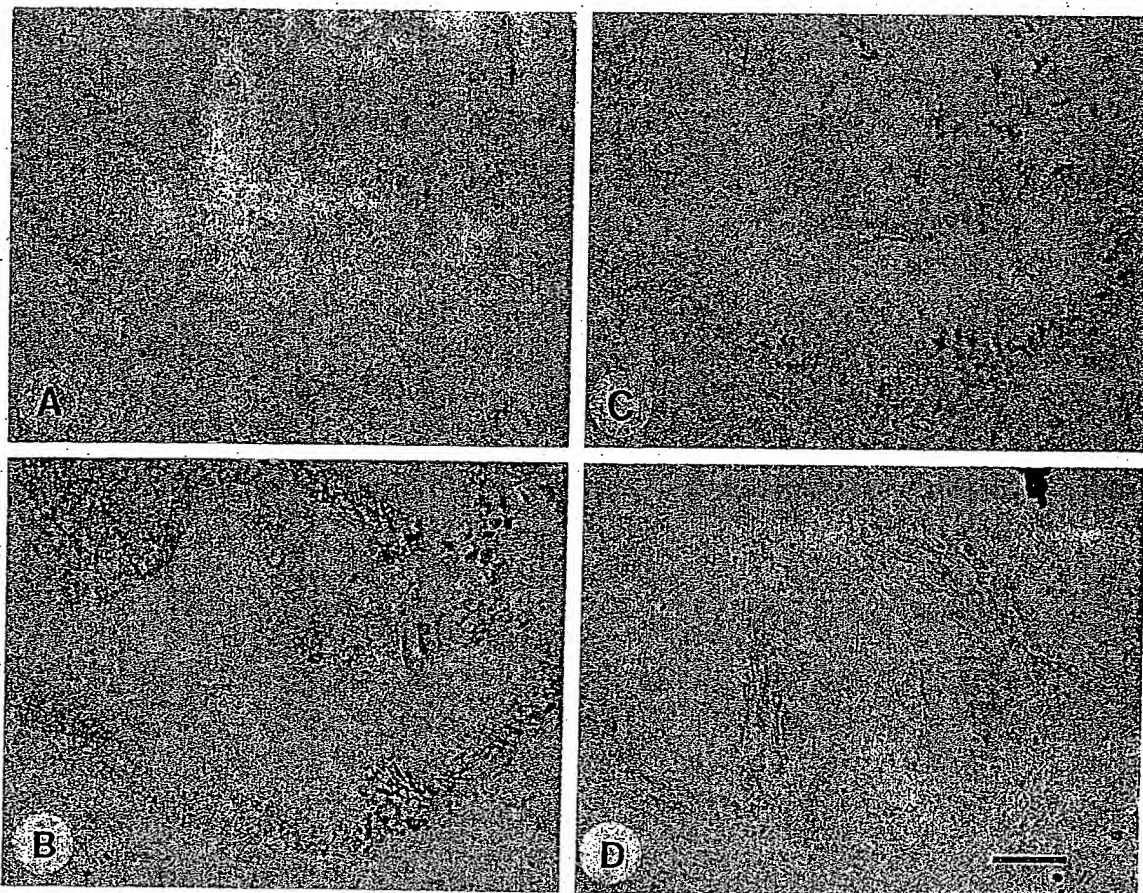


Figure 6. Immunohistochemistry of atypical papillary epithelium in CP tissues. Serial section analysis of some CP samples revealed the presence of large duct-like structures with atypical papillary epithelium. Mild to moderate Id-1 (A) and Id-2 (B) immunoreactivity and weak Id-3 (C) immunoreactivity was present in the cytoplasm of the cells forming these large ducts with papillary structures. Some CP samples also exhibited moderate Id-3 immunoreactivity in these cells (D). Scale bar, 25 μ m.

three Id proteins. An example of serial sections from a pancreatic cancer tissue is shown in Figure 4, A, D, and G.

Id-1, Id-2, and Id-3 immunoreactivity was also present at moderate levels in the cytoplasm of ductal cells within CP-like areas adjacent to the cancer cells (Figure 5, A-C). As in the normal pancreas, islet cells (outlined by arrowheads) did not exhibit Id immunoreactivity. In 4 of 9 CP samples, there were foci of ductal cell dysplasia of relatively large interlobular ducts, all of which exhibited moderate to strong Id-1, Id-2, and Id-3 immunoreactivity (Figure 5, D-F). Five of 9 CP samples also contained foci of large ducts exhibiting atypical papillary epithelium. Serial section analysis of one of those CP samples revealed mild to moderate Id-1 and Id-2 immunoreactivity and weak Id-3 immunoreactivity in the cells of these atypical papillary ducts (Figure 6, A-C). In contrast, in some of these CP samples, moderate to strong Id-3 immunoreactivity was also observed (Figure 6D). However, most of the ductal cells forming the typical ductular structures of CP, such as large interlobular ducts and small proliferating ducts, exhibited generally only weak to occasionally moderate Id immunoreactivity (data not shown).

The immunohistochemical data for Id-1, Id-2, and Id-3 are summarized in Table 1. In the case of Id-1 and Id-2, the cancer cells as well as the dysplastic and atypical papillary ducts in CP exhibited a significantly higher score than the ductal cells in the normal pancreas. In contrast, due to the marked variability in Id-3 immunostaining in the normal pancreas, the differences between normal and cancer cells and normal and dysplastic cells did not achieve statistical significance.

Discussion

Id proteins constitute a family of HLH transcription factors that are important regulators of cellular differentiation and proliferation.^{1,2} To date, four members of the human Id family have been identified.^{1,10-12} Their expression is enhanced during cellular proliferation and in response to mitogenic stimuli,^{19,31} and overexpression of Id genes inhibits differentiation and/or enhances proliferation in several different cell types.^{15,32-34} The forced expression of Id-1 in mouse small intestinal epithelium results in

Table 1. Histological Scoring

| | | Id-1 | Id-2 | Id-3 |
|-----------------|----------------------------------|------------------------|------------------------|-----------|
| Normal (n = 6) | Ductal cells | 2.0 ± 0.4 | 2.3 ± 0.2 | 2.5 ± 0.9 |
| Cancer (n = 10) | Cancer cells | 4.5* ± 0.5 | 5.2 [§] ± 0.3 | 4.5 ± 0.6 |
| CP (n = 9) | Typical CP lesions (n = 9) | 2.7 ± 0.5 | 3.1 ± 0.6 | 3.4 ± 0.7 |
| | Dysplastic ducts (n = 4) | 5.3 [†] ± 0.2 | 5.8 [†] ± 0.2 | 5.3 ± 0.4 |
| | Atypical papillary ducts (n = 5) | 4.4 [‡] ± 0.2 | 5.2 [‡] ± 0.2 | 5.0 ± 0.4 |

Scoring of the histological specimens was performed as described in the Patients and Methods section. Values are the means ± SD of the number of samples indicated in parenthesis. *P* values are based on comparisons with the respective controls in the normal samples.

* *P* < 0.02; [†]*P* < 0.01; [‡]*P* = 0.004; [§]*P* = 0.001.

adenoma formation in these animals.³⁵ The growth-promoting effects of Id genes are thought to occur through several mechanisms. For example, Id-2 can bind to members of the pRB tumor suppressor family, thus blocking their growth-suppressing activity,^{20,21} and Id-1 and Id-2 can antagonize the bHLH-mediated activation of known inhibitors of cell cycle progression such as the cyclin-dependent kinase inhibitor p21.²³

In the present study, we determined by Northern blot analysis that a significant percentage of human pancreatic cancers expressed increased Id-1, Id-2, and Id-3 mRNA levels. Increased expression was most evident for Id-1 (6.5-fold) and Id-2 (fivefold). In contrast, Id-3 mRNA levels were only twofold increased in the cancer samples, partly because this mRNA was present at relatively high levels in the normal pancreas. Immunohistochemical analysis confirmed the presence of Id-1, Id-2, and Id-3 in the cancer cells within the tumor mass, whereas in the normal pancreas faint Id-1 and Id-2 immunoreactivity and moderate to occasionally strong Id-3 immunoreactivity was present in some ductal cells. Pancreatic acinar and islet cells in the normal pancreas were devoid of Id-1, Id-2, and Id-3 immunoreactivity. In the cancer samples, all three Id proteins often colocalized in the cancer cells. Coexpression of all three Id genes was also observed in cultured pancreatic cancer cell lines, which often exhibited a close correlation between Id mRNA and protein expression. However, in MIA-PaCa-2 there was a divergence of Id-2 mRNA and protein levels, and in PANC-1 cells, Id-3 mRNA levels did not correlate well with Id-3 protein expression. These observations suggest that in these cells, the half-life of either Id mRNA or Id protein may be altered by comparison with the other cell lines. Interestingly, Id-2 immunoblotting revealed two closely spaced bands of approximately 16 and 18 kd in 4 of 5 cell lines. In view of the fact that two possible initiation codons have been reported for the Id-2 gene,³⁶ our observation raises the possibility that the two Id-2-immunoreactive bands may represent separate translation products of the Id-2 gene.

Pancreatic cancers often harbor p53 tumor suppressor gene mutations³⁷ and exhibit alterations in apoptosis pathways. Thus, these cancers often exhibit increased expression of anti-apoptotic proteins such as Bcl-2³⁸ and abnormal resistance to Fas-ligand-mediated apoptosis.³⁹ It has been shown recently that forced constitutive expression of Id genes together with the expression of anti-apoptotic genes such as Bcl-2 or Bcl_x can result in

malignant transformation of human fibroblasts,¹¹ raising the possibility that the enhanced Id expression in pancreatic cancers together with increased expression of anti-apoptotic genes may contribute to the malignant potential of pancreatic cancer cells *in vivo*.

In the CP tissues there was no significant increase in Id-1, Id-2, and Id-3 mRNA levels in comparison to the normal pancreas. Immunohistochemical analysis of pancreatic cancer samples revealed colocalization of weak to moderate Id-1, Id-2, and Id-3 immunoreactivity in proliferating ductal cells in the CP-like regions adjacent to the cancer cells, indicating that Id expression was not restricted to the cancer cells. Similarly, analysis of CP samples indicated weak Id-1, Id-2, and Id-3 immunoreactivity in the cells of small proliferating ducts and large ducts without dysplastic changes. In general, there was a correlation between weak immunoreactivity and low Id mRNA levels. However, in samples that harbored large ducts with papillary structures there was moderate Id immunoreactivity, and in the cells forming dysplastic ducts there was moderate to strong Id immunoreactivity. In these CP samples, Id mRNA levels were relatively higher than in the CP samples that were devoid of these histological changes. Overall, however, increased Id expression, most notably of Id-1 and Id-2, distinguished a subgroup of pancreatic cancers from CP (Table 1).

Epidemiological studies have shown that the risk of developing pancreatic cancer is increased up to 16-fold in patients with pre-existing CP in comparison to the general population.⁴⁰ The mechanisms that contribute to neoplastic transformation in CP are not known. Although there is no established tumor progression model for pancreatic cancer, such as the adenoma-carcinoma sequence of colorectal carcinoma,⁴¹ it is generally accepted that K-ras and p16 mutations occur relatively early in pancreatic carcinogenesis, whereas p53 mutations occur late in this process.^{37,41-43} Increased Id expression may contribute to malignant transformation of cultured cell lines *in vitro*¹¹ and has been linked to cell invasion in a murine mammary epithelial cell line.⁴⁴ In view of the current findings that Id-1, Id-2, and Id-3 are overexpressed in pancreatic cancer and in dysplastic/metaplastic ducts in CP, these observations raise the possibility that elevated levels of Id-1, Id-2, and, to a lesser extent, Id-3 may represent relatively early markers of pancreatic malignant transformation and may contribute to the pathobiology of pancreatic cancer.

References

- Jan YN, Jan LY: HLH proteins, fly neurogenesis, and vertebrate myogenesis. *Cell* 1993, 75:827-830
- Olson EN, Klein WH: bHLH factors in muscle development: dead lines and commitments, what to leave in and what to leave out. *Genes Dev* 1994, 8:1-8
- Begley CG, Aplan PD, Denning SM, Haynes BF, Waldmann TA, Kirsch IR: The gene SCL is expressed during early hematopoiesis and encodes a differentiation-related DNA-binding motif. *Proc Natl Acad Sci USA* 1989, 86:10128-10132
- Johnson JE, Birren SJ, Anderson DJ: Two rat homologues of *Drosophila achaete-scute* specifically expressed in neuronal precursors. *Nature* 1990, 346:858-861
- Weintraub H: The MyoD family and myogenesis: redundancy, networks, and thresholds. *Cell* 1993, 75:1241-1244
- Hu JS, Olson EN, Kingston RE: HEB, a helix-loop-helix protein related to E2A, and ITF2 that can modulate the DNA-binding ability of myogenic regulatory factors. *Mol Cell Biol* 1992, 12:1031-1042
- Langlands K, Yin X, Anand G, Prochownik EV: Differential interactions of Id proteins with basic-helix-loop-helix transcription factors. *J Biol Chem* 1997, 272:19785-19793
- Murre C, Bain G, van Dijk MA, Engel I, Furnari BA, Massari ME, Matthews JR, Quong MW, Rivera RR, Stuver MH: Structure and function of helix-loop-helix proteins. *Biochim Biophys Acta* 1994, 1218:129-135
- Murre C, McCaw PS, Vaessin H, Caudy M, Jan LY, Jan YN, Cabrera CV, Buskin JN, Hauschka SD, Lassar AB, Baltimore D: Interactions between heterologous helix-loop-helix proteins generate complexes that bind specifically to a common DNA sequence. *Cell* 1989, 58:537-544
- Benezra R, Davis RL, Lockshon D, Turner DL, Weintraub H: The protein Id: a negative regulator of helix-loop-helix DNA binding proteins. *Cell* 1990, 61:49-59
- Norton JD, Atherton GT: Coupling of cell growth control and apoptosis functions of Id proteins. *Mol Cell Biol* 1998, 18:2371-2381
- Norton JD, Deed RW, Craggs G, Sablitzky F: Id helix-loop-helix proteins in cell growth and differentiation. *Trends Cell Biol* 1998, 8:58-65
- Christy BA, Sanders LK, Lau LF, Copeland NG, Jenkins NA, Nathans D: An Id-related helix-loop-helix protein encoded by a growth factor-inducible gene. *Proc Natl Acad Sci USA* 1991, 88:1815-1819
- Kawaguchi N, DeLuca HF, Noda M: Id gene expression and its suppression by 1,25-dihydroxyvitamin D3 in rat osteoblastic osteosarcoma cells. *Proc Natl Acad Sci USA* 1992, 89:4569-4572
- Kreider BL, Benezra R, Rovera G, Kadesch T: Inhibition of myeloid differentiation by the helix-loop-helix protein Id. *Science* 1992, 255:1700-1702
- Le Jossic C, Ilyin GP, Loyer P, Glaise D, Cariou S, Guguen-Guillouzo C: Expression of helix-loop-helix factor Id-1 is dependent on the hepatocyte proliferation and differentiation status in rat liver and in primary culture. *Cancer Res* 1994, 54:6065-6068
- Sun XH, Copeland NG, Jenkins NA, Baltimore D: Id proteins Id1 and Id2 selectively inhibit DNA binding by one class of helix-loop-helix proteins. *Mol Cell Biol* 1991, 11:5603-5611
- Wilson RB, Kiledjian M, Shen CP, Benezra R, Zwollo P, Dymecki SM, Desiderio SV, Kadesch T: Repression of immunoglobulin enhancers by the helix-loop-helix protein Id: implications for B-lymphoid-cell development. *Mol Cell Biol* 1991, 11:6185-6191
- Hara E, Yamaguchi T, Nojima H, Ide T, Campisi J, Okayama H, Oda K: Id-related genes encoding helix-loop-helix proteins are required for G1 progression and are repressed in senescent human fibroblasts. *J Biol Chem* 1994, 269:2139-2145
- Iavarone A, Garg P, Lasorella A, Hsu J, Israel MA: The helix-loop-helix protein Id-2 enhances cell proliferation and binds to the retinoblastoma protein. *Genes Dev* 1994, 8:1270-1284
- Lasorella A, Iavarone A, Israel MA: Id2 specifically alters regulation of the cell cycle by tumor suppressor proteins. *Mol Cell Biol* 1996, 16:2570-2578
- Peverali FA, Rånqvist T, Saffrich R, Pepperkok R, Barone MV, Philipson L: Regulation of G1 progression by E2A and Id helix-loop-helix proteins. *EMBO J* 1994, 13:4291-4301
- Prabhu S, Ignatova A, Park ST, Sun XH: Regulation of the expression of cyclin-dependent kinase inhibitor p21 by E2A and Id proteins. *Mol Cell Biol* 1997, 17:5888-5896
- Warshaw AL, Fernandez-del Castillo C: Pancreatic carcinoma. *N Engl J Med* 1992, 326:455-465
- Korc M: Role of growth factors in pancreatic cancer. *Surg Oncol Clin North Am* 1998, 7:25-41
- Kleeff J, Ishiwata T, Friess H, Büchler MW, Israel MA, Korc M: The helix-loop-helix protein Id2 is overexpressed in human pancreatic cancer. *Cancer Res* 1998, 58:3769-3772
- Oertel JE, Heffes CS, Oertel YC: *Pancreas. Diagnostic Surgical Pathology*. Edited by SS Sternberg. New York, Raven Press, 1989, pp. 1057-1093
- Korc M, Chandrasekar B, Yamanaka Y, Friess H, Büchler MW, Beger HG: Overexpression of the epidermal growth factor receptor in human pancreatic cancer is associated with concomitant increase in the levels of epidermal growth factor and transforming growth factor α . *J Clin Invest* 1992, 90:1352-1360
- Saeki T, Stromberg K, Qi CF, Gullick WJ, Tahara E, Normanno N, Ciardiello F, Kenney N, Johnson GR, Salomon DS: Differential immunohistochemical detection of amphiregulin and crip1 in human normal colon and colorectal tumors. *Cancer Res* 1992, 52:3467-3473
- Cantero D, Friess H, Defforin J, Zimmermann A, Bröndler MA, Riesle E, Korc M, Büchler MW: Enhanced expression of urokinase plasminogen activator and its receptor in pancreatic carcinoma. *Br J Cancer* 1997, 75:388-395
- Desprez PY, Hara E, Bissell MJ, Campisi J: Suppression of mammary epithelial cell differentiation by the helix-loop-helix protein Id-1. *Mol Cell Biol* 1995, 15:3398-3404
- Shoji W, Yamamoto T, Obinata M: The helix-loop-helix protein Id inhibits differentiation of murine erythroleukemia cells. *J Biol Chem* 1994, 269:5078-5084
- Cross JC, Flannery ML, Blonar MA, Steingrimsson E, Jenkins NA, Copeland NG, Rutter WJ, Werb Z: Hxt encodes a basic helix-loop-helix transcription factor that regulates trophoblast cell development. *Development* 1995, 121:2513-2523
- Sun XH: Constitutive expression of the Id1 gene impairs mouse B cell development. *Cell* 1994, 79:893-900
- Wice BM, Gordon JI: Forced expression of Id-1 in the adult mouse small intestinal epithelium is associated with development of adenomas. *J Biol Chem* 1998, 273:25310-25319
- Barone MV, Pepperkok R, Peverali FA, Philipson L: Id proteins control growth induction in mammalian cells. *Proc Natl Acad Sci USA* 1994, 91:4985-4988
- Barton CM, Staddon SL, Hughes CM, Hall PA, O'Sullivan C, Kloppel G, Theis B, Russell RC, Neoptolmos J, Williamson RCN, Lane DP, Lemoine NR: Abnormalities of the p53 tumour suppressor gene in human pancreatic cancer. *Br J Cancer* 1991, 64:1076-1082
- Ohshio G, Suwa H, Imamura T, Yamaki K, Tanaka T, Hashimoto Y, Imamura M: An immunohistochemical study of bcl-2 and p53 protein expression in pancreatic carcinomas. *Scand J Gastroenterol* 1998, 33:535-539
- Ungeloren H, Voss M, Jansen M, Roeder C, Henne-Bruns D, Kremer B, Kalthoff H: Human pancreatic adenocarcinomas express Fas and Fas ligand yet are resistant to Fas-mediated apoptosis. *Cancer Res* 1998, 58:1741-1749
- Niedermaier C, Niedermaier MC, Heintges T, Lüthen R: Epidemiology: relation between chronic pancreatitis and pancreatic carcinoma. *Cancer of the Pancreas*. Edited by HG Beger, MW Büchler, MH Schoenberg. Ulm, Germany: Universitätsverlag Ulm GmbH, 1996, pp 6-9
- Moskaluk CA, Kern SE: Molecular genetics of pancreatic carcinoma. *Pancreatic Cancer: Pathogenesis, Diagnosis, and Treatment*. Edited by HA Reber. Totowa, NJ, Humana Press, 1998, pp 3-20
- Moskaluk CA, Huban RH, Kern SE: p16 and K-ras gene mutations in the intraductal precursors of human pancreatic adenocarcinoma. *Cancer Res* 1997, 57:2140-2143
- Tada M, Ohashi M, Shiratori Y, Okudaira T, Komatsu Y, Kawabe T, Yoshida H, Machinami R, Kishi K, Omata M: Analysis of K-ras gene mutation in hyperplastic duct cells of the pancreas without pancreatic disease. *Gastroenterology* 1996, 110:227-231
- Desprez PY, Lin CO, Thomasset N, Sympon CJ, Bissell MJ, Campisi J: A novel pathway for mammary epithelial cell invasion induced by the helix-loop-helix protein Id-1. *Mol Cell Biol* 1998, 18:4577-4588



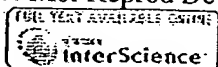
Tightly regulated and inducible expression of a yoked hormone-receptor complex in HEK 293 cells.

Meehan TP, Puett D, Narayan P.

Department of Biochemistry and Molecular Biology, University of Georgia, Athens, Georgia 30602, USA.

We have previously reported the construction of a constitutively active luteinizing hormone receptor by covalently linking a fused heterodimeric hormone to the extracellular domain of the G protein-coupled receptor. This yoked hormone-receptor complex (YHR) was found to produce high levels of cAMP in the absence of exogenous hormone. Stable lines expressing YHR were generated in HEK 293 cells to obtain lines with different expression levels; however, in a relatively short time of continued passage, it was found that YHR expression was greatly reduced. Herein, we describe the development of clonal lines of HEK 293 cells in which the expression of YHR is under the control of a tetracycline-regulated system. Characterization of clonal lines revealed tight control of YHR expression both by dose and time of incubation with doxycycline. These experiments demonstrated a good correlation between expression levels of the receptor and basal cAMP production. Moreover, the reduction in receptor expression following doxycycline removal revealed that YHR mRNA and protein decayed at similar rates, again suggesting a strong linkage between mRNA and protein levels. The controlled expression of YHR in this cell system will allow for a more detailed analysis of the signaling properties associated with constitutive receptor activation and may prove to be advantageous in developmental studies with transgenic animals.

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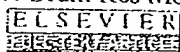
c-fos and estrogen receptor gene expression pattern in the rat uterine epithelium during the estrous cycle.

Mendoza-Rodriguez CA, Merchant-Larios H, Segura-Valdez ML, Moreno-Mendoza N, Cruz ME, Arteaga-Lopez P, Camacho-Arroyo I, Dominguez R, Cerbon M.

Facultad de Quimica, Universidad Nacional Autonoma de Mexico, Ciudad Universitaria, Coyoacan 04510, Mexico, D.F., Mexico.

Different studies in ovariectomized estrogen treated animals support the idea that c-fos plays a role in the proliferation of uterine epithelial cells. However, these studies invite us to reassess the role played by c-fos in epithelial cell types of the endometrium during the estrous cycle. The present study was undertaken to determine the c-fos and estrogen receptor (ER) gene expression pattern in the rat uterine epithelium during the estrous cycle in which natural and cyclic changes of steroid hormones occur, and correlate these changes with the proliferation status of this cellular types. Proliferation was assessed during the estrous cycle using bromodeoxyuridine incorporation to DNA. ERalpha and beta proteins were assessed by immunohistochemistry. The regulation of c-fos gene expression in the uterus of intact animals during the estrous cycle was evaluated using both in situ hybridization and immunohistochemistry. Estradiol (E(2)) and progesterone (P(4)) plasma levels were assessed by radioimmunoassay. The results indicated that luminal (LE) and glandular epithelia (GE) presented maximal proliferation during the metestrus (M) and the diestrus (D) days. However, during the proestrus (P) day only LE presented proliferation, and during the estrus (E) day only the stromal cells proliferated. A marked immunostaining for ERalpha was detected in both LE and GE cells during the early phases of the cycle but diminished on the P and the E day. In contrast, ERbeta was undetectable in both epithelia during all stages of the cycle. The highest c-fos mRNA level was detected in both epithelia on the M day, followed by a significant reduction during the other days of the cycle. The highest protein content was observed on the M and D days, and the minimal value was detected on the E day. The c-Fos protein level in LE was increased during M and D days, presenting a high correlation with the cellular proliferation pattern of this cell type. In conclusion, the overall results indicate that c-Fos protein presented a good correlation with uterine epithelial cell proliferation of LE. In the case of GE, the same tendency was observed, although no significant correlation was found. Both in LE and GE, c-fos mRNA did not strictly correlate with its protein levels. c-fos seems to have a posttranscriptional regulation in uterine epithelial cells during the rat's estrous cycle. Copyright 2003 Wiley-Liss, Inc.

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[3H]MK-801 binding and the mRNA for the NMDAR1 subunit of the NMDA receptor are differentially distributed in human and rat forebrain.

Meoni P, Mugnaini M, Bunnemann BH, Trist DG, Bowery NG.

Department of Pharmacology, Medical School, University of Birmingham, UK.
meonip@novel15.bham.ac.uk

The distributions of [3H]MK-801 binding and the NMDA NR1 subunit mRNA were studied using receptor autoradiography and in-situ hybridization in rat and human brain whole-hemisphere coronal sections. Receptor protein detected by radioligand autoradiography and the mRNA for the key subunit of the receptor presented similar distributions in the forebrain, with a few areas showing an imbalance between the levels of mRNA and receptor protein. Human frontal cortex showed a relative abundance of NMDAR1 mRNA as compared to [3H]MK-801 binding. The same area in rat brain did not show any difference in the two distributions. In comparison, the rat claustrum presented a relative excess of NMDAR1 mRNA which was not detected in human sections. Human caudate nucleus exhibited relatively high levels of [3H]MK-801 binding that were unmatched in rat caudate. The hippocampi of either species presented similar levels of [3H]MK-801 binding and NMDAR1 mRNA, but when the two signals were measured in specific subfields of the hippocampal formation, the differential distribution of the two signals reflected the anatomy of hippocampal connections assuming a preferential dendritic distribution for MK-801 binding. Interestingly, rat and human hippocampi also showed some important species-dependent difference in the relative distribution of the receptor protein and mRNA. The data presented show an overall good correlation between the mRNA for the key subunit of the NMDA receptor and the functional receptor detected with radioligand binding and highlight the presence of local differences in their ratio. This may reflect different splicing of the mRNA for the NMDAR1 subunit in specific brain areas of rat and human. The species-dependent differences in the relative distribution of the mRNA for the key subunit of the NMDA receptor and that of a marker of functional receptors also highlights important differences in the NMDA function in rat and human brain.

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Review

Translation Initiation in Cancer: A Novel Target for Therapy¹

Funda Meric² and Kelly K. Hunt

Department of Surgical Oncology, The University of Texas M. D. Anderson Cancer Center, Houston, Texas 77030

Abstract

Translation initiation is regulated in response to nutrient availability and mitogenic stimulation and is coupled with cell cycle progression and cell growth. Several alterations in translational control occur in cancer. Variant mRNA sequences can alter the translational efficiency of individual mRNA molecules, which in turn play a role in cancer biology. Changes in the expression or availability of components of the translational machinery and in the activation of translation through signal transduction pathways can lead to more global changes, such as an increase in the overall rate of protein synthesis and translational activation of the mRNA molecules involved in cell growth and proliferation. We review the basic principles of translational control, the alterations encountered in cancer, and selected therapies targeting translation initiation to help elucidate new therapeutic avenues.

Introduction

The fundamental principle of molecular therapeutics in cancer is to exploit the differences in gene expression between cancer cells and normal cells. With the advent of cDNA array technology, most efforts have concentrated on identifying differences in gene expression at the level of mRNA, which can be attributable either to DNA amplification or to differences in transcription. Gene expression is quite complicated, however, and is also regulated at the level of mRNA stability, mRNA translation, and protein stability.

The power of translational regulation has been best recognized among developmental biologists, because transcription does not occur in early embryogenesis in eukaryotes. For example, in *Xenopus*, the period of transcriptional quiescence continues until the embryo reaches midblastula transition, the 4000-cell stage. Therefore, all necessary mRNA molecules are transcribed during oogenesis and stockpiled in a translationally inactive, masked form. The mRNA are translationally activated at appropriate times during oocyte maturation, fertilization, and

early embryogenesis and thus, are under strict translational control.

Translation has an established role in cell growth. Basically, an increase in protein synthesis occurs as a consequence of mitogenesis. Until recently, however, little was known about the alterations in mRNA translation in cancer, and much is yet to be discovered about their role in the development and progression of cancer. Here we review the basic principles of translational control, the alterations encountered in cancer, and selected therapies targeting translation initiation to elucidate potential new therapeutic avenues.

Basic Principles of Translational Control

Mechanism of Translation Initiation

Translation initiation is the main step in translational regulation. Translation initiation is a complex process in which the initiator tRNA and the 40S and 60S ribosomal subunits are recruited to the 5' end of a mRNA molecule and assembled by eukaryotic translation initiation factors into an 80S ribosome at the start codon of the mRNA (Fig. 1). The 5' end of eukaryotic mRNA is capped, i.e., contains the cap structure m⁷GpppN (7-methyl-guanosine-triphospho-5'-ribonucleoside). Most translation in eukaryotes occurs in a cap-dependent fashion, i.e., the cap is specifically recognized by the eIF4E,³ which binds the 5' cap. The eIF4F translation initiation complex is then formed by the assembly of eIF4E, the RNA helicase eIF4A, and eIF4G, a scaffolding protein that mediates the binding of the 40S ribosomal subunit to the mRNA molecule through interaction with the eIF3 protein present on the 40S ribosome. eIF4A and eIF4B participate in melting the secondary structure of the 5' UTR of the mRNA. The 43S initiation complex (40S/eIF2/Met-tRNA^{GTP} complex) scans the mRNA in a 5'→3' direction until it encounters an AUG start codon. This start codon is then base-paired to the anticodon of initiator tRNA, forming the 48S initiation complex. The initiation factors are then displaced from the 48S complex, and the 60S ribosome joins to form the 80S ribosome.

Unlike most eukaryotic translation, translation initiation of certain mRNAs, such as the picornavirus RNA, is cap independent and occurs by internal ribosome entry. This mechanism does not require eIF4E. Either the 43S complex can bind the initiation codon directly through interaction with the IRES in the 5' UTR such as in the encephalomyocarditis virus, or it can

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² To whom requests for reprints should be addressed, at Department of Surgical Oncology, Box 444, The University of Texas M. D. Anderson Cancer Center, 1515 Holcombe Boulevard, Houston, TX 77030. Phone: (713) 745-4453; Fax: (713) 745-4926; E-mail: fmeric@mdanderson.org.

³ The abbreviations used are: eIF4E, eukaryotic initiation factor 4E; UTR, untranslated region; IRES, internal ribosome entry site; 4E-BP1, eukaryotic initiation factor 4E-binding protein 1; S6K, ribosomal p70 S6 kinase; mTOR, mammalian target of rapamycin; ATM, ataxia telangiectasia mutated; PI3K, phosphatidylinositol 3-kinase; PTEN, phosphatase and tensin homolog deleted from chromosome 10; PP2A, protein phosphatase 2A; TGF- β 3, transforming growth factor- β 3; PAP, poly(A) polymerase; EPA, eicosapentaenoic acid; mda-7, melanoma differentiation-associated gene 7.

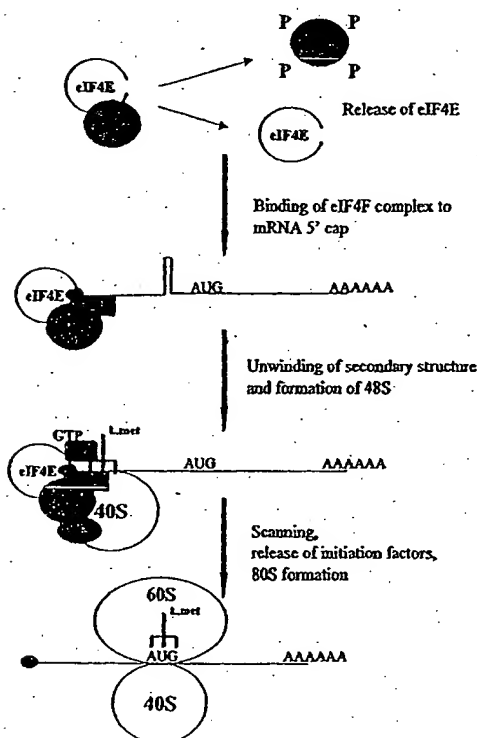


Fig. 1. Translation initiation in eukaryotes. The 4E-BPs are hyperphosphorylated to release eIF4E so that it can interact with the 5' cap, and the eIF4F initiation complex is assembled. The interaction of poly(A) binding protein with the initiation complex and circularization of the mRNA is not depicted in the diagram. The secondary structure of the 5' UTR is melted, the 40S ribosomal subunit is bound to eIF3, and the ternary complex consisting of eIF2, GTP, and the Met-tRNA are recruited to the mRNA. The ribosome scans the mRNA in a 5'→3' direction until an AUG start codon is found in the appropriate sequence context. The initiation factors are released, and the large ribosomal subunit is recruited.

initially attach to the IRES and then reach the initiation codon by scanning or transfer, as is the case with the poliovirus (1).

Regulation of Translation Initiation

Translation initiation can be regulated by alterations in the expression or phosphorylation status of the various factors involved. Key components in translational regulation that may provide potential therapeutic targets follow.

eIF4E. eIF4E plays a central role in translation regulation. It is the least abundant of the initiation factors and is considered the rate-limiting component for initiation of cap-dependent translation. eIF4E may also be involved in mRNA splicing, mRNA 3' processing, and mRNA nucleocytoplasmic transport (2). eIF4E expression can be increased at the transcriptional level in response to serum or growth factors (3). eIF4E overexpression may cause preferential translation of mRNAs containing excessive secondary structure in their 5' UTR that are normally discriminated against by the trans-

lational machinery and thus are inefficiently translated (4–7). As examples of this, overexpression of eIF4E promotes increased translation of vascular endothelial growth factor, fibroblast growth factor-2, and cyclin D1 (2, 8, 9).

Another mechanism of control is the regulation of eIF4E phosphorylation. eIF4E phosphorylation is mediated by the mitogen-activated protein kinase-interacting kinase 1, which is activated by the mitogen-activated pathway, activating extracellular signal-related kinases and the stress-activated pathway acting through p38 mitogen-activated protein kinase (10–13). Several mitogens, such as serum, platelet-derived growth factor, epidermal growth factor, insulin, angiotensin II, src kinase overexpression, and ras overexpression, lead to eIF4E phosphorylation (14). The phosphorylation status of eIF4E is usually correlated with the translational rate and growth status of the cell; however, eIF4E phosphorylation has also been observed in response to some cellular stresses when translational rates actually decrease (15). Thus, further study is needed to understand the effects of eIF4E phosphorylation on eIF4E activity.

Another mechanism of regulation is the alteration of eIF4E availability by the binding of eIF4E to the eIF4E-binding proteins (4E-BP, also known as PHAS-I). 4E-BPs compete with eIF4G for a binding site in eIF4E. The binding of eIF4E to the best characterized eIF4E-binding protein, 4E-BP1, is regulated by 4E-BP1 phosphorylation. Hypophosphorylated 4E-BP1 binds to eIF4E, whereas 4E-BP1 hyperphosphorylation decreases this binding. Insulin, angiotensin, epidermal growth factor, platelet-derived growth factor, hepatocyte growth factor, nerve growth factor, insulin-like growth factors I and II, interleukin 3, granulocyte-macrophage colony-stimulating factor + steel factor, gastrin, and the adenovirus have all been reported to induce phosphorylation of 4E-BP1 and to decrease the ability of 4E-BP1 to bind eIF4E (15, 16). Conversely, deprivation of nutrients or growth factors results in 4E-BP1 dephosphorylation, an increase in eIF4E binding, and a decrease in cap-dependent translation.

p70 S6 Kinase. Phosphorylation of ribosomal 40S protein S6 by S6K is thought to play an important role in translational regulation. S6K $-/-$ mouse embryonic cells proliferate more slowly than do parental cells, demonstrating that S6K has a positive influence on cell proliferation (17). S6K regulates the translation of a group of mRNAs possessing a 5' terminal oligopyrimidine tract (5' TOP) found at the 5' UTR of ribosomal protein mRNAs and other mRNAs coding for components of the translational machinery. Phosphorylation of S6K is regulated in part based on the availability of nutrients (18, 19) and is stimulated by several growth factors, such as platelet-derived growth factor and insulin-like growth factor I (20).

eIF2 α Phosphorylation. The binding of the initiator tRNA to the small ribosomal unit is mediated by translation initiation factor eIF2. Phosphorylation of the α -subunit of eIF2 prevents formation of the eIF2/GTP/Met-tRNA complex and inhibits global protein synthesis (21, 22). eIF2 α is phosphorylated under a variety of conditions, such as viral infection, nutrient deprivation, heme deprivation, and apoptosis (22). eIF2 α is phosphorylated by heme-regulated inhibitor, nutrient-regulated protein kinase, and the IFN-induced, double-stranded RNA-activated protein kinase (PKR; Ref. 23).

The mTOR Signaling Pathway. The macrolide antibiotic rapamycin (Siralimus; Wyeth-Ayerst Research, Collegeville, PA) has been the subject of intensive study because it inhibits signal transduction pathways involved in T-cell activation. The rapamycin-sensitive component of these pathways is mTOR (also called FRAP or RAFT1). mTOR is the mammalian homologue of the yeast TOR proteins that regulate G₁ progression and translation in response to nutrient availability (24). mTOR is a serine-threonine kinase that modulates translation initiation by altering the phosphorylation status of 4E-BP1 and S6K (Fig. 2; Ref. 25).

4E-BP1 is phosphorylated on multiple residues. mTOR phosphorylates the Thr-37 and Thr-46 residues of 4E-BP1 *in vitro* (26); however, phosphorylation at these sites is not associated with a loss of eIF4E binding. Phosphorylation of Thr-37 and Thr-46 is required for subsequent phosphorylation at several COOH-terminal, serum-sensitive sites; a combination of these phosphorylation events appears to be needed to inhibit the binding of 4E-BP1 to eIF4E (25). The product of the *ATM* gene, p38/MSK1 pathway, and protein kinase C α also play a role in 4E-BP1 phosphorylation (27-29).

S6K and 4E-BP1 are also regulated, in part, by PI3K and its downstream protein kinase Akt. PTEN is a phosphatase that negatively regulates PI3K signaling. PTEN null cells have constitutively active Akt, with increased S6K activity and S6 phosphorylation (30). S6K activity is inhibited both by PI3K inhibitors wortmannin and LY294002 and by mTOR inhibitor rapamycin (24). Akt phosphorylates Ser-2448 in mTOR *in vitro*, and this site is phosphorylated upon Akt activation *in vivo* (31-33). Thus, mTOR is regulated by the PI3K/Akt pathway; however, this does not appear to be the only mode of regulation of mTOR activity. Whether the PI3K pathway also regulates S6K and 4E-BP1 phosphorylation independent of mTOR is controversial.

Interestingly, mTOR autophosphorylation is blocked by wortmannin but not by rapamycin (34). This seeming inconsistency suggests that mTOR-responsive regulation of 4E-BP1 and S6K activity occurs through a mechanism other than intrinsic mTOR kinase activity. An alternate pathway for 4E-BP1 and S6K phosphorylation by mTOR activity is by the inhibition of a phosphatase. Treatment with calyculin A, an inhibitor of phosphatases 1 and 2A, reduces rapamycin-induced dephosphorylation of 4E-BP1 and S6K by rapamycin (35). PP2A interacts with full-length S6K but not with a S6K mutant that is resistant to dephosphorylation resulting from rapamycin. mTOR phosphorylates PP2A *in vitro*; however, how this process alters PP2A activity is not known. These results are consistent with the model that phosphorylation of a phosphatase by mTOR prevents dephosphorylation of 4E-BP1 and S6K, and conversely, that nutrient deprivation and rapamycin block inhibition of the phosphatase by mTOR.

Polyadenylation. The poly(A) tail in eukaryotic mRNA is important in enhancing translation initiation and mRNA stability. Polyadenylation plays a key role in regulating gene expression during oogenesis and early embryogenesis. Some mRNA that are translationally inactive in the oocyte are polyadenylated concomitantly with translational activation in oocyte maturation, whereas other mRNAs that are translationally active during oogenesis are deadenylated and trans-

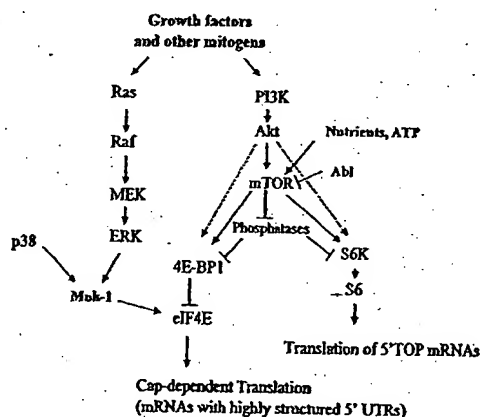


Fig. 2. Regulation of translation initiation by signal transduction pathways. Signaling via p38, extracellular signal-related kinase, PI3K, and mTOR can all activate translation initiation.

lationally silenced (36-38). Thus, control of poly(A) tail synthesis is an important regulatory step in gene expression. The 5' cap and poly(A) tail are thought to function synergistically to regulate mRNA translational efficiency (39, 40).

RNA Packaging. Most RNA-binding proteins are assembled on a transcript at the time of transcription, thus determining the translational fate of the transcript (41). A highly conserved family of Y-box proteins is found in cytoplasmic messenger ribonucleoprotein particles, where the proteins are thought to play a role in restricting the recruitment of mRNA to the translational machinery (41-43). The major mRNA-associated protein, YB-1, destabilizes the interaction of eIF4E and the 5' mRNA cap *in vitro*, and overexpression of YB-1 results in translational repression *in vivo* (44). Thus, alterations in RNA packaging can also play an important role in translational regulation.

Translation Alterations Encountered in Cancer

Three main alterations at the translational level occur in cancer: variations in mRNA sequences that increase or decrease translational efficiency, changes in the expression or availability of components of the translational machinery, and activation of translation through aberrantly activated signal transduction pathways. The first alteration affects the translation of an individual mRNA that may play a role in carcinogenesis. The second and third alterations can lead to more global changes, such as an increase in the overall rate of protein synthesis, and the translational activation of several mRNA species.

Variations in mRNA Sequence

Variations in mRNA sequence affect the translational efficiency of the transcript. A brief description of these variations and examples of each mechanism follow.

Mutations. Mutations in the mRNA sequence, especially in the 5' UTR, can alter its translational efficiency, as seen in the following examples.

c-myc. Saito *et al.* proposed that translation of full-length *c-myc* is repressed, whereas in several Burkitt lymphomas that have deletions of the mRNA 5' UTR, translation of *c-myc* is more efficient (45). More recently, it was reported that the 5' UTR of *c-myc* contains an IRES, and thus *c-myc* translation can be initiated by a cap-independent as well as a cap-dependent mechanism (46, 47). In patients with multiple myeloma, a C→T mutation in the *c-myc* IRES was identified (48) and found to cause an enhanced initiation of translation via internal ribosomal entry (49).

BRCA1. A somatic point mutation (117 G→C) in position -3 with respect to the start codon of the *BRCA1* gene was identified in a highly aggressive sporadic breast cancer (50). Chimeric constructs consisting of the wild-type or mutated *BRCA1* 5' UTR and a downstream luciferase reporter demonstrated a decrease in the translational efficiency with the 5' UTR mutation.

Cyclin-dependent Kinase Inhibitor 2A. Some inherited melanoma kindreds have a G→T transversion at base -34 of cyclin-dependent kinase inhibitor-2A, which encodes a cyclin-dependent kinase 4/cyclin-dependent kinase 6 kinase inhibitor important in G₁ checkpoint regulation (51). This mutation gives rise to a novel AUG translation initiation codon, creating an upstream open reading frame that competes for scanning ribosomes and decreases translation from the wild-type AUG.

Alternate Splicing and Alternate Transcription Start Sites. Alterations in splicing and alternate transcription sites can lead to variations in 5' UTR sequence, length, and secondary structure, ultimately impacting translational efficiency.

ATM. The *ATM* gene has four noncoding exons in its 5' UTR that undergo extensive alternative splicing (52). The contents of 12 different 5' UTRs that show considerable diversity in length and sequence have been identified. These divergent 5' leader sequences play an important role in the translational regulation of the *ATM* gene.

mdm. In a subset of tumors, overexpression of the oncoprotein *mdm2* results in enhanced translation of the *mdm2* mRNA. Use of different promoters leads to two *mdm2* transcripts that differ only in their 5' leaders (53). The longer 5' UTR contains two upstream open reading frames, and this mRNA is loaded with ribosomes inefficiently compared with the short 5' UTR.

BRCA1. In a normal mammary gland, *BRCA1* mRNA is expressed with a shorter leader sequence (5' UTRa), whereas in sporadic breast cancer tissue, *BRCA1* mRNA is expressed with a longer leader sequence (5' UTRb); the translational efficiency of transcripts containing 5' UTRb is 10 times lower than that of transcripts containing 5' UTRa (54).

TGF-β3. *TGF-β3* mRNA includes a 1.1-kb 5' UTR, which exerts an inhibitory effect on translation. Many human breast cancer cell lines contain a novel *TGF-β3* transcript with a 5' UTR that is 870 nucleotides shorter and has a 7-fold greater translational efficiency than the normal *TGF-β3* mRNA (55).

Alternate Polyadenylation Sites. Multiple polyadenylation signals leading to the generation of several transcripts with differing 3' UTR have been described for several mRNA species, such as the *RET* proto-oncogene (56), *ATM* gene (52), tissue inhibitor of metalloproteinases-3 (57), *RHOA*

proto-oncogene (58), and calmodulin-I (59). Although the effect of these alternate 3' UTRs on translation is not yet known, they may be important in RNA-protein interactions that affect translational recruitment. The role of these alterations in cancer development and progression is unknown.

Alterations in the Components of the Translation Machinery

Alterations in the components of translation machinery can take many forms.

Overexpression of eIF4E. Overexpression of eIF4E causes malignant transformation in rodent cells (60) and the deregulation of HeLa cell growth (61). Polunovsky *et al.* (62) found that eIF4E overexpression substitutes for serum and individual growth factors in preserving viability of fibroblasts, which suggests that eIF4E can mediate both proliferative and survival signaling.

Elevated levels of eIF4E mRNA have been found in a broad spectrum of transformed cell lines (63). eIF4E levels are elevated in all ductal carcinoma *in situ* specimens and invasive ductal carcinomas, compared with benign breast specimens evaluated with Western blot analysis (64, 65). Preliminary studies suggest that this overexpression is attributable to gene amplification (66).

There are accumulating data suggesting that eIF4E overexpression can be valuable as a prognostic marker. eIF4E overexpression was found in a retrospective study to be a marker of poor prognosis in stages I to III breast carcinoma (67). Verification of the prognostic value of eIF4E in breast cancer is now under way in a prospective trial (67). However, in a different study, eIF4E expression was correlated with the aggressive behavior of non-Hodgkin's lymphomas (68). In a prospective analysis of patients with head and neck cancer, elevated levels of eIF4E in histologically tumor-free surgical margins predicted a significantly increased risk of local-regional recurrence (9). These results all suggest that eIF4E overexpression can be used to select patients who might benefit from more aggressive systemic therapy. Furthermore, the head and neck cancer data suggest that eIF4E overexpression is a field defect and can be used to guide local therapy.

Alterations in Other Initiation Factors. Alterations in a number of other initiation factors have been associated with cancer. Overproduction of eIF4G, similar to eIF4E, leads to malignant transformation *in vitro* (69). eIF-2α is found in increased levels in bronchioloalveolar carcinomas of the lung (3). Initiation factor eIF-4A1 is overexpressed in melanoma (70) and hepatocellular carcinoma (71). The p40 subunit of translation initiation factor 3 is amplified and overexpressed in breast and prostate cancer (72), and the eIF3-p110 subunit is overexpressed in testicular seminoma (73). The role that overexpression of these initiation factors plays on the development and progression of cancer, if any, is not known.

Overexpression of S6K. S6K is amplified and highly overexpressed in the MCF7 breast cancer cell line, compared with normal mammary epithelium (74). In a study by Bartlund *et al.* (74), S6K was amplified in 59 of 668 primary breast tumors, and a statistically significant association was observed between amplification and poor prognosis.

Overexpression of PAP. PAP catalyzes 3' poly(A) synthesis. PAP is overexpressed in human cancer cells compared with normal and virally transformed cells (75). PAP enzymatic activity in breast tumors has been correlated with PAP protein levels (76) and, in mammary tumor cytosols, was found to be an independent factor for predicting survival (76). Little is known, however, about how PAP expression or activity affects the translational profile.

Alterations in RNA-binding Proteins. Even less is known about alterations in RNA packaging in cancer. Increased expression and nuclear localization of the RNA-binding protein YB-1 are indicators of a poor prognosis for breast cancer (77), non-small cell lung cancer (78), and ovarian cancer (79). However, this effect may be mediated at least in part at the level of transcription, because YB-1 increases chemoresistance by enhancing the transcription of a multidrug resistance gene (80).

Activation of Signal Transduction Pathways

Activation of signal transduction pathways by loss of tumor suppressor genes or overexpression of certain tyrosine kinases can contribute to the growth and aggressiveness of tumors. An important mutant in human cancers is the tumor suppressor gene *PTEN*, which leads to the activation of the PI3K/Akt pathway. Activation of PI3K and Akt induces the oncogenic transformation of chicken embryo fibroblasts. The transformed cells show constitutive phosphorylation of S6K and of 4E-BP1 (81). A mutant Akt that retains kinase activity but does not phosphorylate S6K or 4E-BP1 does not transform fibroblasts, which suggests a correlation between the oncogenicity of PI3K and Akt and the phosphorylation of S6K and 4E-BP1 (81).

Several tyrosine kinases such as platelet-derived growth factor, insulin-like growth factor, HER2/neu, and epidermal growth factor receptor are overexpressed in cancer. Because these kinases activate downstream signal transduction pathways known to alter translation initiation, activation of translation is likely to contribute to the growth and aggressiveness of these tumors. Furthermore, the mRNA for many of these kinases themselves are under translational control. For example, HER2/neu mRNA is translationally controlled both by a short upstream open reading frame that represses HER2/neu translation in a cell type-independent manner and by a distinct cell type-dependent mechanism that increases translational efficiency (82). HER2/neu translation is different in transformed and normal cells. Thus, it is possible that alterations at the translational level can, in part account for the discrepancy between *HER2/neu* gene amplification detected by fluorescence *in situ* hybridization and protein levels detected by immunohistochemical assays.

Translation Targets of Selected Cancer Therapy

Components of the translation machinery and signal pathways involved in the activation of translation initiation represent good targets for cancer therapy.

Targeting the mTOR Signaling Pathway: Rapamycin and Tumstatin

Rapamycin inhibits the proliferation of lymphocytes. It was initially developed as an immunosuppressive drug for organ

transplantation. Rapamycin with FKBP 12 (FK506-binding protein, M_r 12,000) binds to mTOR to inhibit its function.

Rapamycin causes a small but significant reduction in the initiation rate of protein synthesis (83). It blocks cell growth in part by blocking S6 phosphorylation and selectively suppressing the translation of 5' TOP mRNAs, such as ribosomal proteins, and elongation factors (83–85). Rapamycin also blocks 4E-BP1 phosphorylation and inhibits cap-dependent but not cap-independent translation (17, 86).

The rapamycin-sensitive signal transduction pathway, activated during malignant transformation and cancer progression, is now being studied as a target for cancer therapy (87). Prostate, breast, small cell lung, glioblastoma, melanoma, and T-cell leukemia are among the cancer lines most sensitive to the rapamycin analogue CCI-779 (Wyeth-Ayerst Research; Ref. 87). In rhabdomyosarcoma cell lines, rapamycin is either cytostatic or cytotoxic, depending on the p53 status of the cell; p53 wild-type cells treated with rapamycin arrest in the G₁ phase and maintain their viability, whereas p53 mutant cells accumulate in G₁ and undergo apoptosis (88, 89). In a recently reported study using human primitive neuroectodermal tumor and medulloblastoma models, rapamycin exhibited more cytotoxicity in combination with cisplatin and camptothecin than as a single agent. *In vivo*, CCI-779 delayed growth of xenografts by 160% after 1 week of therapy and 240% after 2 weeks. A single high-dose administration caused a 37% decrease in tumor volume. Growth inhibition *in vivo* was 1.3 times greater, with cisplatin in combination with CCI-779 than with cisplatin alone (90). Thus, preclinical studies suggest that rapamycin analogues are useful as single agents and in combination with chemotherapy.

Rapamycin analogues CCI-779 and RAD001 (Novartis, Basel, Switzerland) are now in clinical trials. Because of the known effect of rapamycin on lymphocyte proliferation, a potential problem with rapamycin analogues is immunosuppression. However, although prolonged immunosuppression can result from rapamycin and CCI-779 administered on continuous-dose schedules, the immunosuppressive effects of rapamycin analogues resolve in ~24 h after therapy (91). The principal toxicities of CCI-779 have included dermatological toxicity, myelosuppression, infection, mucositis, diarrhea, reversible elevations in liver function tests, hyperglycemia, hypokalemia, hypocalcemia, and depression (87, 92–94). Phase II trials of CCI-779 have been conducted in advanced renal cell carcinoma and in stage III/IV breast carcinoma patients who failed with prior chemotherapy. In the results reported in abstract form, although there were no complete responses, partial responses were documented in both renal cell carcinoma and in breast carcinoma (94, 95). Thus, CCI-779 has documented preliminary clinical activity in a previously treated, unselected patient population.

Active investigation is under way into patient selection for mTOR inhibitors. Several studies have found an enhanced efficacy of CCI-779 in *PTEN*-null tumors (30, 96). Another study found that six of eight breast cancer cell lines were responsive to CCI-779, although only two of these lines lacked *PTEN* (97). There was, however, a positive correlation between Akt activation and CCI-779 sensitivity (97). This correlation suggests that activation of the PI3K-Akt pathway,

regardless of whether it is attributable to a PTEN mutation or to overexpression of receptor tyrosine kinases, makes cancer cell amenable to mTOR-directed therapy. In contrast, lower levels of the target of mTOR, 4E-BP1, are associated with rapamycin resistance; thus, a lower 4E-BP1/eIF4E ratio may predict rapamycin resistance (98).

Another mode of activity for rapamycin and its analogues appears to be through inhibition of angiogenesis. This activity may be both through direct inhibition of endothelial cell proliferation as a result of mTOR inhibition in these cells or by inhibition of translation of such proangiogenic factors as vascular endothelial growth factor in tumor cells (99, 100).

The angiogenesis inhibitor tnmstatin, another anticancer drug currently under study, was also found recently to inhibit translation in endothelial cells (101). Through a requisite interaction with integrin, tnmstatin inhibits activation of the PI3K/Akt pathway and mTOR in endothelial cells and prevents dissociation of eIF4E from 4E-BP1, thereby inhibiting cap-dependent translation. These findings suggest that endothelial cells are especially sensitive to therapies targeting the mTOR-signaling pathway.

Targeting eIF2 α EPA, Clotrimazole, mda-7, and Flavonoids

EPA is an n-3 polyunsaturated fatty acid found in the fish-based diets of populations having a low incidence of cancer (102). EPA inhibits the proliferation of cancer cells (103), as well as in animal models (104, 105). It blocks cell division by inhibiting translation initiation (105). EPA releases Ca²⁺ from intracellular stores while inhibiting their refilling, thereby activating PKR. PKR, in turn phosphorylates and inhibits eIF2 α , resulting in the inhibition of protein synthesis at the level of translation initiation. Similarly, clotrimazole, a potent antiproliferative agent *in vitro* and *in vivo*, inhibits cell growth through depletion of Ca²⁺ stores, activation of PKR, and phosphorylation of eIF2 α (106). Consequently, clotrimazole preferentially decreases the expression of cyclins A, E, and D1, resulting in blockage of the cell cycle in G₁.

mda-7 is a novel tumor suppressor gene being developed as a gene therapy agent. Adenoviral transfer of mda-7 (Ad-mda7) induces apoptosis in many cancer cells including breast, colorectal, and lung cancer (107–109). Ad-mda7 also induces and activates PKR, which leads to phosphorylation of eIF2 α and induction of apoptosis (110).

Flavonoids such as genistein and quercetin suppress tumor cell growth. All three mammalian eIF2 α kinases, PKR, heme-regulated inhibitor, and PERK/PEK, are activated by flavonoids, with phosphorylation of eIF2 α and inhibition of protein synthesis (111).

Targeting eIF4A and eIF4E: Antisense RNA and Peptides

Antisense expression of eIF4A decreases the proliferation rate of melanoma cells (112). Sequestration of eIF4E by overexpression of 4E-BP1 is proapoptotic and decreases tumorigenicity (113, 114). Reduction of eIF4E with antisense RNA decreases soft agar growth, increases tumor latency, and increases the rates of tumor doubling times (7). Antisense eIF4E RNA treat-

ment also reduces the expression of angiogenic factors (115) and has been proposed as a potential adjuvant therapy for head and neck cancers, particularly when elevated eIF4E is found in surgical margins. Small molecule inhibitors that bind the eIF4G/4E-BP1-binding domain of eIF4E are proapoptotic (116) and are also being actively pursued.

Exploiting Selective Translation for Gene Therapy

A different therapeutic approach that takes advantage of the enhanced cap-dependent translation in cancer cells is the use of gene therapy vectors encoding suicide genes with highly structured 5' UTR. These mRNA would thus be at a competitive disadvantage in normal cells and not translate well, whereas in cancer cells, they would translate more efficiently. For example, the introduction of the 5' UTR of fibroblast growth factor-2'5' to the coding sequence of herpes simplex virus type-1 thymidine kinase gene, allows for selective translation of herpes simplex virus type-1 thymidine kinase gene in breast cancer cell lines compared with normal mammary cell lines and results in selective sensitivity to ganciclovir (117).

Toward the Future

Translation is a crucial process in every cell. However, several alterations in translational control occur in cancer. Cancer cells appear to need an aberrantly activated translational state for survival, thus allowing the targeting of translation initiation with surprisingly low toxicity. Components of the translational machinery, such as eIF4E, and signal transduction pathways involved in translation initiation, such as mTOR, represent promising targets for cancer therapy. Inhibitors of the mTOR have already shown some preliminary activity in clinical trials. It is possible that with the development of better predictive markers and better patient selection, response rates to single-agent therapy can be improved. Similar to other cytostatic agents, however, mTOR inhibitors are most likely to achieve clinical utility in combination therapy. In the interim, our increasing understanding of translation initiation and signal transduction pathways promise to lead to the identification of new therapeutic targets in the near future.

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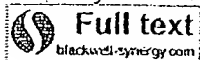
References

1. Pestova, T. V., Kolupaeva, V. G., Lomakin, I. B., Pilipenko, E. V., Shatsky, I. N., Agol, V. I., and Hellen, C. U. Molecular mechanisms of translation initiation in eukaryotes. *Proc. Natl. Acad. Sci. USA*, 98: 7029–7036, 2001.
2. Rosenwald, I. B., Kaspar, R., Rousseau, D., Gehlke, L., Leboulch, P., Chen, J. J., Schmidt, E. V., Sonenberg, N., and London, I. M. Eukaryotic translation initiation factor 4E regulates expression of cyclin D1 at transcriptional and post-transcriptional levels. *J. Biol. Chem.*, 270: 21176–21180, 1995.
3. Rosenwald, I. B., Hutzler, M. J., Wang, S., Savas, L., and Fraire, A. E. Expression of eukaryotic translation initiation factors 4E and 2 α is increased frequently in bronchioloalveolar but not in squamous cell carcinomas of the lung. *Cancer (Phila.)*, 92: 2164–2171, 2001.

4. Darveau, A., Pelletier, J., and Sonenberg, N. Differential efficiencies of *in vitro* translation of mouse c-myc transcripts differing in the 5' untranslated region. *Proc. Natl. Acad. Sci. USA*, 82: 2315-2319, 1985.
5. Kozak, M. Influences of mRNA secondary structure on initiation by eukaryotic ribosomes. *Proc. Natl. Acad. Sci. USA*, 83: 2850-2854, 1986.
6. Koromilas, A. E., Lazaris-Karatzas, A., and Sonenberg, N. mRNAs containing extensive secondary structure in their 5' non-coding region translate efficiently in cells overexpressing initiation factor eIF-4E. *EMBO J.*, 11: 4153-4158, 1992.
7. Finkler-Schaeffer, C. W., Graff, J. R., De Benedetti, A., Zimmer, S. G., and Rhoads, R. E. Decreasing the level of translation initiation factor 4E with antisense RNA causes reversal of ras-mediated transformation and tumorigenesis of cloned rat embryo fibroblasts. *Int. J. Cancer*, 55: 841-847, 1993.
8. Kevil, C. G., De Benedetti, A., Payne, D. K., Coe, L. L., Laroux, F. S., and Alexander, J. S. Translational regulation of vascular permeability factor by eukaryotic initiation factor 4E: implications for tumor angiogenesis. *Int. J. Cancer*, 65: 785-790, 1996.
9. Nathan, C. A., Franklin, S., Abreo, F. W., Nassar, R., De Benedetti, A., and Glass, J. Analysis of surgical margins with the molecular marker eIF4E: a prognostic factor in patients with head and neck cancer. *J. Clin. Oncol.*, 17: 2909-2914, 1999.
10. Fukunaga, R., and Hunter, T. MNK1, a new MAP kinase-activated protein kinase, isolated by a novel expression screening method for identifying protein kinase substrates. *EMBO J.*, 16: 1921-1933, 1997.
11. Waskiewicz, A. J., Flynn, A., Proud, C. G., and Cooper, J. A. Mitogen-activated protein kinases activate the serine/threonine kinases Mnk1 and Mnk2. *EMBO J.*, 16: 1909-1920, 1997.
12. Wang, X., Flynn, A., Waskiewicz, A. J., Webb, B. L., Vries, R. G., Baines, I. A., Cooper, J. A., and Proud, C. G. The phosphorylation of eukaryotic initiation factor eIF4E in response to phorbol esters, cell stresses, and cytokines is mediated by distinct MAP kinase pathways. *J. Biol. Chem.*, 273: 9373-9377, 1998.
13. Pyronnet, S., Inatoka, H., Gingras, A. C., Fukunaga, R., Hunter, T., and Sonenberg, N. Human eukaryotic translation initiation factor 4G (eIF4G) recruits Mnk1 to phosphorylate eIF4E. *EMBO J.*, 18: 270-279, 1999.
14. Kleijn, M., Scheper, G. C., Voorma, H. O., and Thomas, A. A. Regulation of translation initiation factors by signal transduction. *Eur. J. Biochem.*, 253: 531-544, 1998.
15. Raught, B., and Gingras, A. C. eIF4E activity is regulated at multiple levels. *Int. J. Biochem. Cell Biol.*, 31: 43-57, 1999.
16. Takeuchi, K., Shibamoto, S., Nagamine, K., Shigemori, I., Omura, S., Kitamura, N., and Ito, F. Signaling pathways leading to transcription and translation cooperatively regulate the transient increase in expression of c-Fos protein. *J. Biol. Chem.*, 276: 26077-26083, 2001.
17. Kawasome, H., Popst, P., Webb, S., Keller, G. M., Johnson, G. L., Gelfand, E. W., and Terada, N. Targeted disruption of p70(s6k) defines its role in protein synthesis and rapamycin sensitivity. *Proc. Natl. Acad. Sci. USA*, 95: 5033-5038, 1998.
18. Christie, G. R., Hajdich, E., Hundal, H. S., Proud, C. G., and Taylor, P. M. Intracellular sensing of amino acids in *Xenopus laevis* oocytes stimulates p70 S6 kinase in a target of rapamycin-dependent manner. *J. Biol. Chem.*, 277: 9952-9957, 2002.
19. Hara, K., Yonezawa, K., Weng, Q. P., Kozłowski, M. T., Betham, C., and Avruch, J. Amino acid sufficiency and mTOR regulate p70 S6 kinase and eIF-4E BP1 through a common effector mechanism. *J. Biol. Chem.*, 273: 14484-14494, 1998.
20. Graves, L. M., Bornfeldt, K. E., Argast, G. M., Krebs, E. G., Kong, X., Lin, T. A., and Lawrence, J. C., Jr. cAMP- and rapamycin-sensitive regulation of the association of eukaryotic initiation factor 4E and the translational regulator PHAS-I in aortic smooth muscle cells. *Proc. Natl. Acad. Sci. USA*, 92: 7222-7226, 1995.
21. Merrick, W. C., and Hershey, J. W. B. The pathway and mechanism of eukaryotic protein synthesis. In: J. W. B. Hershey and M. B. Mathews (eds.), *Translational Control*, pp. 31-69. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory, 1996.
22. Kimball, S. R. Eukaryotic initiation factor eIF2. *Int. J. Biochem. Cell Biol.*, 31: 25-29, 1999.
23. Jagus, R., Joshi, B., and Barber, G. N. PKR, apoptosis and cancer. *Int. J. Biochem. Cell Biol.*, 31: 123-138, 1999.
24. Thomas, G., and Hall, M. N. TOR signalling and control of cell growth. *Curr. Opin. Cell Biol.*, 9: 782-787, 1997.
25. Gingras, A. C., Raught, B., and Sonenberg, N. Regulation of translation initiation by FRAP/mTOR. *Genes Dev.*, 15: 807-826, 2001.
26. Gingras, A. C., Gygi, S. P., Raught, B., Polakiewicz, R. D., Abraham, R. T., Hoekstra, M. F., Aebersold, R., and Sonenberg, N. Regulation of 4E-BP1 phosphorylation: a novel two-step mechanism. *Genes Dev.*, 13: 1422-1437, 1999.
27. Kumar, V., Pandey, P., Sabatini, D., Kumar, M., Majumder, P. K., Bhardi, A., Carmichael, G., Kufe, D., and Karthanda, S. Functional interaction between RAFT1/FRAP/mTOR and protein kinase C δ in the regulation of cap-dependent initiation of translation. *EMBO J.*, 19: 1087-1097, 2000.
28. Yang, D. Q., and Kastan, M. B. Participation of ATM in insulin signaling through phosphorylation of eIF-4E-binding protein 1. *Nat. Cell Biol.*, 2: 893-898, 2000.
29. Liu, G., Zhang, Y., Bode, A. M., Ma, W. Y., and Dong, Z. Phosphorylation of 4E-BP1 is mediated by the p38/Msk1 pathway in response to UVB irradiation. *J. Biol. Chem.*, 277: 8810-8816, 2002.
30. Neshat, M. S., Mellingerhoff, I. K., Tran, C., Siles, B., Thomas, G., Petersen, R., Frost, P., Gibbons, J. J., Wu, H., and Sawyers, C. L. Enhanced sensitivity of PTEN-deficient tumors to inhibition of FRAP/mTOR. *Proc. Natl. Acad. Sci. USA*, 98: 10314-10319, 2001.
31. Sekulic, A., Hudson, C. C., Homme, J. L., Yin, P., Ottemess, D. M., Karnitz, L. M., and Abraham, R. T. A direct linkage between the phosphoinositide 3-kinase-AKT signaling pathway and the mammalian target of rapamycin in mitogen-stimulated and transformed cells. *Cancer Res.*, 60: 3504-3513, 2000.
32. Scott, P. H., and Lawrence, J. C., Jr. Attenuation of mammalian target of rapamycin activity by increased cAMP in 3T3-L1 adipocytes. *J. Biol. Chem.*, 273: 34496-34501, 1998.
33. Reynolds, I. T., Bodine, S. C., and Lawrence, J. C., Jr. Control of Ser2448 phosphorylation in the mammalian target of rapamycin by insulin and skeletal muscle load. *J. Biol. Chem.*, 277: 17657-17662, 2002.
34. Peterson, R. T., Beal, P. A., Comb, M. J., and Schreiber, S. L. FKBP12-rapamycin-associated protein (FRAP) autophosphorylates at serine 2481 under translationally repressive conditions. *J. Biol. Chem.*, 275: 7416-7423, 2000.
35. Peterson, R. T., Desai, B. N., Hardwick, J. S., and Schreiber, S. L. Protein phosphatase 2A interacts with the 70-kDa S6 kinase and is activated by inhibition of FKBP12-rapamycin-associated protein. *Proc. Natl. Acad. Sci. USA*, 96: 4438-4442, 1999.
36. McGrew, L. L., Dworkin-Rastl, E., Dworkin, M. B., and Richter, J. D. Poly(A) elongation during *Xenopus* oocyte maturation is required for translational recruitment and is mediated by a short sequence element. *Genes Dev.*, 3: 803-815, 1989.
37. Sheets, M. D., Wu, M., and Wickens, M. Polyadenylation of c-mos mRNA as a control point in *Xenopus* meiotic maturation. *Nature (Lond.)*, 374: 511-516, 1995.
38. Yamum, S. M., and Wormington, W. M. Deadenylation of maternal mRNAs during *Xenopus* oocyte maturation does not require specific cis-sequences: a default mechanism for translational control. *Genes Dev.*, 4: 2278-2286, 1990.
39. Gallie, D. R. The cap and poly(A) tail function synergistically to regulate mRNA translational efficiency. *Genes Dev.*, 5: 2108-2116, 1991.
40. Sachs, A. B., and Varani, G. Eukaryotic translation initiation: there are (at least) two sides to every story. *Nat. Struct. Biol.*, 7: 356-361, 2000.
41. Wolffe, A. P., and Meric, F. Coupling transcription to translation: a novel site for the regulation of eukaryotic gene expression. *Int. J. Biochem. Cell Biol.*, 28: 247-257, 1996.
42. Evdokimova, V. M., Wei, C. L., Sitkov, A. S., Simonenko, P. N., Lazarev, O. A., Vasilenko, K. S., Ustinov, V. A., Hershey, J. W., and Ovchinnikov, L. P. The major protein of messenger ribonucleoprotein particles in somatic cells is a member of the Y-box binding transcription factor family. *J. Biol. Chem.*, 270: 3186-3192, 1995.
43. Matsumoto, K., Meric, F., and Wolffe, A. P. Translational repression dependent on the interaction of the *Xenopus* Y-box protein FRGY2 with mRNA: Role of the cold shock domain, tail domain, and selective RNA sequence recognition. *J. Biol. Chem.*, 271: 22706-22712, 1996.

44. Evdokimova, V., Ruzanov, P., Imataka, H., Raught, B., Svitkin, Y., Ovchinnikov, L. P., and Sonenberg, N. The major mRNA-associated protein YB-1 is a potent 5' cap-dependent mRNA stabilizer. *EMBO J.*, 20: 5491-5502, 2001.
45. Saito, H., Hayday, A. C., Wiman, K., Hayward, W. S., and Tonegawa, S. Activation of the c-myc gene by translocation: a model for translational control. *Proc. Natl. Acad. Sci. USA*, 80: 7476-7480, 1983.
46. Nambu, C., Lafon, I., Audigier, S., Gensac, M. C., Vagner, S., Huez, G., and Prats, A. C. Alternative translation of the proto-oncogene c-myc by an internal ribosome entry site. *J. Biol. Chem.*, 272: 32061-32066, 1997.
47. Stoneley, M., Paulin, F. E., Le Quesne, J. P., Chappell, S. A., and Willis, A. E. c-Myc 5' untranslated region contains an internal ribosome entry segment. *Oncogene*, 16: 423-428, 1998.
48. Paulin, F. E., West, M. J., Sullivan, N. F., Whitney, R. L., Lyne, L., and Willis, A. E. Aberrant translational control of the c-myc gene in multiple myeloma. *Oncogene*, 13: 505-513, 1996.
49. Chappell, S. A., LeQuesne, J. P., Paulin, F. E., de Schoolmeester, M. L., Stoneley, M., Soutar, R. L., Rakston, S. H., Hellrich, M. H., and Willis, A. E. A mutation in the c-myc-IRES leads to enhanced internal ribosome entry in multiple myeloma: a novel mechanism of oncogene de-regulation. *Oncogene*, 19: 4437-4440, 2000.
50. Signori, E., Bagni, C., Papa, S., Primerano, B., Rinaldi, M., Amaldi, F., and Fazio, V. M. A somatic mutation in the 5'UTR of *BRCA1* gene in sporadic breast cancer causes down-modulation of translation efficiency. *Oncogene*, 20: 4596-4600, 2001.
51. Liu, L., Dilworth, D., Gao, L., Monzon, J., Summers, A., Lassar, N., and Hogg, D. Mutation of the CDK2A 5' UTR creates an aberrant initiation codon and predisposes to melanoma. *Nat. Genet.*, 21: 128-132, 1999.
52. Savitsky, K., Platzer, M., Uziel, T., Gilad, S., Sartiel, A., Rosenthal, A., Elroy-Stein, O., Shiloh, Y., and Rotman, G. Ataxia-telangiectasia: structural diversity of untranslated sequences suggests complex post-transcriptional regulation of *ATM* gene expression. *Nucleic Acids Res.*, 25: 1678-1684, 1997.
53. Brown, C. Y., Mize, G. J., Pineda, M., George, D. L., and Morris, D. R. Role of two upstream open reading frames in the translational control of oncogene *mdm2*. *Oncogene*, 18: 5631-5637, 1999.
54. Sobczak, K., and Krzyzosiak, W. J. Structural determinants of *BRCA1* translational regulation. *J. Biol. Chem.*, 277: 17349-17358, 2002.
55. Arrick, B. A., Grendell, R. L., and Griffin, L. A. Enhanced translational efficiency of a novel transforming growth factor $\beta 3$ mRNA in human breast cancer cells. *Mol. Cell. Biol.*, 14: 619-628, 1994.
56. Myers, S. M., Eng, C., Ponder, B. A., and Mulligan, L. M. Characterization of *RET* proto-oncogene 3' splicing variants and polyadenylation sites: a novel C-terminus for *RET*. *Oncogene*, 11: 2039-2045, 1995.
57. Byrne, J. A., Tomasello, C., Rouyer, N., Bellocq, J. P., Fio, M. C., and Basset, P. The tissue inhibitor of metalloproteinases-3 gene in breast carcinoma: identification of multiple polyadenylation sites and a stromal pattern of expression. *Mol. Med.*, 1: 418-427, 1995.
58. Moscow, J. A., He, R., Gudas, J. M., and Cowan, K. H. Utilization of multiple polyadenylation signals in the human *RHOA* protooncogene. *Gene (Amst.)*, 144: 229-236, 1994.
59. Senterre-Lesentants, S., Alag, A. S., and Sobel, M. E. Multiple mRNA species are generated by alternate polyadenylation from the human *calmodulin-1* gene. *J. Cell. Biochem.*, 58: 445-454, 1995.
60. Lazaris-Karatzas, A., Montine, K. S., and Sonenberg, N. Malignant transformation by a eukaryotic initiation factor subunit that binds to mRNA 5' cap. *Nature (Lond.)*, 345: 544-547, 1990.
61. De Benedetti, A., and Rhoads, R. E. Overexpression of eukaryotic protein synthesis initiation factor 4E in HeLa cells results in aberrant growth and morphology. *Proc. Natl. Acad. Sci. USA*, 87: 8212-8216, 1990.
62. Polunovsky, V. A., Rosenwald, I. B., Tan, A. T., White, J., Chiang, L., Sonenberg, N., and Bitterman, P. B. Translational control of programmed cell death: eukaryotic translation initiation factor 4E blocks apoptosis in growth-factor-restricted fibroblasts with physiologically expressed or de-regulated *Myc*. *Mol. Cell. Biol.*, 16: 6573-6581, 1996.
63. Miyagi, Y., Sugiyama, A., Asai, A., Okazaki, T., Kuchino, Y., and Ken, S. J. Elevated levels of eukaryotic translation initiation factor eIF-4E mRNA in a broad spectrum of transformed cell lines. *Cancer Lett.*, 91: 247-252, 1995.
64. Kerkatte, V., Smiley, K., Hu, B., Smith, A., Gelder, F., and De Benedetti, A. The proto-oncogene/translation factor *eIF4E*: a survey of its expression in breast carcinomas. *Int. J. Cancer*, 64: 27-31, 1995.
65. Li, B. D., Liu, L., Dawson, M., and De Benedetti, A. Overexpression of eukaryotic initiation factor 4E (eIF4E) in breast carcinoma. *Cancer (Phila.)*, 79: 2385-2390, 1997.
66. Sorrelis, D. L., Black, D. R., Meschonat, C., Rhoads, R., De Benedetti, A., Gao, M., Williams, B. J., and Li, B. D. Detection of *eIF4E* gene amplification in breast cancer by competitive PCR. *Ann. Surg. Oncol.*, 5: 232-237, 1998.
67. Li, B. D., McDonald, J. C., Nassar, R., and De Benedetti, A. Clinical outcome in stage I to III breast carcinoma and *eIF4E* overexpression. *Ann. Surg.*, 227: 756-761; discussion, 761-763, 1998.
68. Wang, S., Rosenwald, I. B., Hutzler, M. J., Pham, G. A., Savas, L., Chen, J. J., and Woda, B. A. Expression of the eukaryotic translation initiation factors 4E and 2a in non-Hodgkin's lymphomas. *Am. J. Pathol.*, 155: 247-255, 1999.
69. Fukuchi-Shimogori, T., Ishii, L., Kashiwagi, K., Mashiba, H., Ekimoto, H., and Igarashi, K. Malignant transformation by overproduction of translation initiation factor eIF4G. *Cancer Res.*, 57: 5041-5044, 1997.
70. Eberle, J., Krasagakis, K., and Orlan, C. E. Translation initiation factor eIF-4A1 mRNA is consistently overexpressed in human melanoma cells *in vitro*. *Int. J. Cancer*, 71: 396-401, 1997.
71. Shida, M., Kondoh, N., Tanaka, K., Ryo, A., Wakatsuki, T., Hada, A., Goseki, N., Igari, T., Matsue, K., Aihara, T., Horiuchi, S., Shichita, M., Yamamoto, N., and Yamamoto, M. Enhanced expression of translation factor mRNAs in hepatocellular carcinoma. *Anticancer Res.*, 20: 2489-2494, 2000.
72. Nupponen, N. N., Porlita, K., Kakkola, L., Tanner, M., Persson, K., Borg, A., Isola, J., and Visakorpi, T. Amplification and overexpression of p40 subunit of eukaryotic translation initiation factor 3 in breast and prostate cancer. *Am. J. Pathol.*, 154: 1777-1783, 1999.
73. Rothe, M., Ko, Y., Albers, P., and Wernert, N. Eukaryotic initiation factor 3 p110 mRNA is overexpressed in testicular seminomas. *Am. J. Pathol.*, 157: 1597-1604, 2000.
74. Bartlund, M., Forozan, F., Kononen, J., Bubendorf, L., Chen, Y., Bittner, M. L., Thorst, J., Haas, P., Bucher, C., Sauter, G., Kallioniemi, O. P., and Kallioniemi, A. Detecting activation of ribosomal protein S6 kinase by complementary DNA and tissue microarray analysis. *J. Natl. Cancer Inst.* (Bethesda), 92: 1252-1259, 2000.
75. Topalian, S. L., Kaneko, S., Gonzales, M. I., Bond, G. L., Ward, Y., and Manley, J. L. Identification and functional characterization of neo-poly(A) polymerase, an RNA processing enzyme overexpressed in human tumors. *Mol. Cell. Biol.*, 21: 5614-5623, 2001.
76. Scorilas, A., Taleri, N., Ardavanis, A., Courtis, N., Dimitriadis, E., Yofis, J., Tsiapalis, C. M., and Triangas, T. Polyadenylate polymerase enzymatic activity in mammary tumor cytosols: a new independent prognostic marker in primary breast cancer. *Cancer Res.*, 60: 5427-5433, 2000.
77. Janz, M., Harbeck, N., Dettmar, P., Berger, U., Schmidt, A., Jurchott, K., Schmitt, M., and Royer, H. D. Y-box factor YB-1 predicts drug resistance and patient outcome in breast cancer independent of clinically relevant tumor biologic factors *HER2*, *uPA* and *PAI-1*. *Int. J. Cancer*, 97: 278-282, 2002.
78. Shibahara, K., Sugio, K., Osaki, T., Uchiumi, T., Maehara, Y., Kohno, K., Yasumoto, K., Sugimachi, K., and Kuwano, M. Nuclear expression of the Y-box binding protein, YB-1, as a novel marker of disease progression in non-small cell lung cancer. *Clin. Cancer Res.*, 7: 3151-3155, 2001.
79. Kamura, T., Yahata, H., Amada, S., Ogawa, S., Sonoda, T., Kobayashi, H., Mitsumoto, M., Kohno, K., Kuwano, M., and Nakano, H. Is nuclear expression of Y box-binding protein-1 a new prognostic factor in ovarian serous adenocarcinoma? *Cancer (Phila.)*, 85: 2450-2454, 1999.
80. Bargou, R. C., Jurchott, K., Wagener, C., Bergmann, S., Metzner, S., Bommer, K., Mager, M. Y., Winer, K. J., Dietel, M., Dorken, B., and Royer, H. D. Nuclear localization and increased levels of transcription factor YB-1 in primary human breast cancers are associated with intrinsic *MDR1* gene expression. *Nat. Med.*, 3: 447-450, 1997.
81. Aoki, M., Blazek, E., and Vogt, P. K. A role of the kinase mTOR in cellular transformation induced by the oncoproteins *P3k* and *Akt*. *Proc. Natl. Acad. Sci. USA*, 98: 136-141, 2001.
82. Child, S. J., Miller, M. K., and Geballe, A. P. Cell type-dependent and -independent control of *HER-2/neu* translation. *Int. J. Biochem. Cell Biol.*, 31: 201-213, 1999.

83. Jefferies, H. B., Reinhard, C., Kozma, S. C., and Thomas, G. Rapamycin selectively represses translation of the "polypyrimidine tract" mRNA family. *Proc. Natl. Acad. Sci. USA*, 91: 4441-4445, 1994.
84. Terada, N., Patel, H. R., Takase, K., Kohno, K., Naim, A. C., and Gelfand, E. W. Rapamycin selectively inhibits translation of mRNAs encoding elongation factors and ribosomal proteins. *Proc. Natl. Acad. Sci. USA*, 91: 11477-11481, 1994.
85. Jefferies, H. B., Fumagalli, S., Dennis, P. B., Reinhard, C., Pearson, R. B., and Thomas, G. Rapamycin suppresses 5'TOP mRNA translation through inhibition of p70S6k. *EMBO J.*, 16: 3693-3704, 1997.
86. Beretta, L., Gingras, A. C., Svitkin, Y. V., Hall, M. N., and Sonenberg, N. Rapamycin blocks the phosphorylation of 4E-BP1 and inhibits cap-dependent initiation of translation. *EMBO J.*, 15: 658-664, 1996.
87. Hidalgo, M., and Rowinsky, E. K. The rapamycin-sensitive signal transduction pathway as a target for cancer therapy. *Oncogene*, 19: 6680-6686, 2000.
88. Hosoi, H., Dilling, M. B., Shikata, T., Liu, L. N., Shu, L., Ashmun, R. A., Germain, G. S., Abraham, R. T., and Houghton, P. J. Rapamycin causes poorly reversible inhibition of mTOR and induces p53-independent apoptosis in human rhabdomyosarcoma cells. *Cancer Res.*, 59: 886-894, 1999.
89. Huang, S., and Houghton, P. J. Resistance to rapamycin: a novel anticancer drug. *Cancer Metastasis Rev.*, 20: 69-78, 2001.
90. Georger, B., Kerr, K., Tang, C. B., Fung, K. M., Powell, B., Sutton, L. N., Phillips, P. C., and Janss, A. J. Antitumor activity of the rapamycin analog CCI-779 in human primitive neuroectodermal tumor/medulloblastoma models as single agent and in combination chemotherapy. *Cancer Res.*, 61: 1527-1532, 2001.
91. Gibbons, J. J., Discalcani, C., Peterson, R., Hernandez, R., Skotnick, J., and Frost, P. The effect of CCI-779, a novel macrolide anti-tumor agent, on the growth of human tumor cells *in vitro* and in nude mouse xenografts *in vivo*. *Proc. Am. Assoc. Cancer Res.*, 40: 301, 1999.
92. Hidalgo, M., Rowinsky, E., Erlichman, C., Marshall, B., Marks, R., Edwards, T., and Buckner, J. J. A Phase I and pharmacological study of CCI-779 cycle inhibitor. *Ann. Oncol.*, 11 (Suppl. 4): 133, 2001.
93. Alexandre, J., Raymond, E., Depenbrock, H., Melchard, S., Angevin, E., Pallet, C., Hanauke, A., Frisch, J., Feussner, A., and Armand, J. P. CCI-779, a new rapamycin analog, has antitumor activity at doses inducing only mild cutaneous effects and mucositis: early results of an ongoing Phase I study. *Proceedings of the 1999 AACR-NCI-EORTC International Conference, Clin. Cancer Res.*, 5 (Suppl.): 3730s, 1999.
94. Chan, S., Johnston, S., Scheulen, M. E., Moss, K., Morant, A., Lahr, A., Feussner, A., Berger, M., and Kirsch, T. First report: a Phase 2 study of the safety and activity of CCI-779 for patients with locally advanced or metastatic breast cancer failing prior chemotherapy. *Proc. Am. Soc. Clin. Oncol.*, 21: 44a, 2002.
95. Atkins, M. B., Hidalgo, M., Stadler, W., Logan, T., Dutcher, J. P., Hudes, G., Park, Y., Marshall, B., Boni, J., and Dukat, G. A randomized double-blind Phase 2 study of intravenous CCI-779 administered weekly to patients with advanced renal cell carcinoma. *Proc. Am. Soc. Clin. Oncol.*, 21: 10a, 2002.
96. Smith, S. G., Trinh, C. M., Inge, L. J., Thomas, G., Cloughsey, T. F., Sawyers, C. L., and Mischel, P. S. PTEN expression status predicts glioblastoma cell sensitivity to CCI-779. *Proc. Am. Assoc. Cancer Res.*, 43: 335, 2002.
97. Yu, K., Toral-Barza, L., Discalcani, C., Zhang, W. G., Skotnick, J., Frost, P., and Gibbons, J. J. mTOR, a novel target in breast cancer: the effect of CCI-779, an mTOR inhibitor, in preclinical models of breast cancer. *Endocr. Relat. Cancer*, 8: 249-258, 2001.
98. Dilling, M. B., Germain, G. S., Dudkin, L., Jayaraman, A. L., Zhang, X., Hanwood, F. C., and Houghton, P. J. 4E-binding proteins, the suppressors of eukaryotic initiation factor 4E, are downregulated in cells with acquired or intrinsic resistance to rapamycin. *J. Biol. Chem.*, 277: 13907-13917, 2002.
99. Guba, M., von Breitenbuch, P., Steinhilber, M., Koehl, G., Fiegel, S., Hornung, M., Bruns, C. J., Zuelke, C., Farkas, S., Anhuber, M., Jauch, K. W., and Geissler, E. K. Rapamycin inhibits primary and metastatic tumor growth by antiangiogenesis: involvement of vascular endothelial growth factor. *Nat. Med.*, 8: 128-135, 2002.
100. Lane, H. A., Schell, C., Theuer, A., O'Reilly, T., and Wood, J. Anti-angiogenic activity of RAD001, an orally active anticancer agent. *Proc. Am. Assoc. Cancer Res.*, 43: 184, 2002.
101. Maeshima, Y., Suchak, A., Lively, J. C., Ueki, K., Kharbada, S., Kahn, C. R., Sonenberg, N., Hynes, R. O., and Kalluri, R. Tumstatin, an endothelial cell-specific inhibitor of protein synthesis. *Science (Wash. DC)*, 295: 140-143, 2002.
102. Caygill, C. P., Charlett, A., and Hill, M. J. Fat, fish, fish oil and cancer. *Br. J. Cancer*, 74: 159-164, 1996.
103. Falconer, J. S., Ross, J. A., Fearon, K. C., Hawkins, R. A., O'Riordan, M. G., and Carter, D. C. Effect of eicosapentaenoic acid and other fatty acids on the growth *in vitro* of human pancreatic cancer cell lines. *Br. J. Cancer*, 69: 826-832, 1994.
104. Noguchi, M., Minami, M., Yagasaki, R., Kinoshita, K., Earashi, M., Kitagawa, H., Taniya, T., and Miyazaki, I. Chemoprevention of DMBA-induced mammary carcinogenesis in rats by low-dose EPA and DHA. *Br. J. Cancer*, 75: 348-353, 1997.
105. Palakurthi, S. S., Fluckiger, R., Aktas, H., Changolkar, A. K., Shahsafari, A., Hameit, S., Klic, E., and Halperin, J. A. Inhibition of translation initiation mediates the anticancer effect of the n-3 polyunsaturated fatty acid eicosapentaenoic acid. *Cancer Res.*, 60: 2919-2925, 2000.
106. Aktas, H., Fluckiger, R., Acosta, J. A., Savage, J. M., Palakurthi, S. S., and Halperin, J. A. Depletion of intracellular Ca^{2+} stores, phosphorylation of eIF2 α , and sustained inhibition of translation initiation mediate the anticancer effects of clotrimazole. *Proc. Natl. Acad. Sci. USA*, 95: 8260-8265, 1998.
107. Mhashikar, A. M., Schrock, R. D., Hindi, M., Liao, J., Siegel, K., Kourouma, F., Zou-Yang, X. H., Onishi, E., Tak, O., Vedvick, T. S., Fanger, G., Stewart, L., Watson, G. J., Snary, D., Fisher, P. B., Saeid, T., Roth, J. A., Ramesh, R., and Chada, S. Melanoma differentiation associated gene-7 (*mda-7*): a novel anti-tumor gene for cancer gene therapy. *Mol. Med.*, 7: 271-282, 2001.
108. Su, Z. Z., Madireddi, M. T., Lin, J. J., Young, C. S., Kilada, S., Reed, J. C., Goldstein, N. L., and Fisher, P. B. The cancer growth suppressor gene *mda-7* selectively induces apoptosis in human breast cancer cells and inhibits tumor growth in nude mice. *Proc. Natl. Acad. Sci. USA*, 95: 14400-14405, 1998.
109. Saeid, T., Mhashikar, A., Chada, S., Branch, C., Roth, J. A., and Ramesh, R. Tumor-suppressive effects by adenovirus-mediated *mda-7* gene transfer in non-small cell lung cancer cell *in vitro*. *Gene Ther.*, 7: 2051-2057, 2000.
110. Palaer, A., Vorburger, S. A., Barber, G. N., Chada, S., Mhashikar, A. M., Zou-Yang, H., Stewart, A. L., Balachandran, S., Roth, J. A., Hunt, K. K., and Swisher, S. G. Adenoviral transfer of the melanoma differentiation-associated gene 7 (*mda7*) induces apoptosis of lung cancer cells via up-regulation of the double-stranded RNA-dependent protein kinase (PKR). *Cancer Res.*, 62: 2239-2243, 2002.
111. Ito, T., Wamken, S. P., and May, W. S. Protein synthesis inhibition by flavonoids: roles of eukaryotic initiation factor 2 α kinases. *Biochem. Biophys. Res. Commun.*, 265: 589-594, 1999.
112. Eberle, J., Fecker, L. F., Bittner, J. U., Orfanos, C. E., and Geilen, C. C. Decreased proliferation of human melanoma cell lines caused by antisense RNA against translation factor eIF-4A1. *Br. J. Cancer*, 86: 1957-1962, 2002.
113. Polunovsky, V. A., Gingras, A. C., Sonenberg, N., Peterson, M., Tan, A., Rubins, J. B., Marivel, J. C., and Bitterman, P. B. Translational control of the antiapoptotic function of Ras. *J. Biol. Chem.*, 275: 24776-24780, 2000.
114. D'Cunha, J., Kratzke, M. G., Alter, M. D., Polunovsky, V. A., Bitterman, P. B., and Kratzke, R. A. Over-expression of the translational repressor 4E-BP1 inhibits NSCLC tumorigenicity *in vivo*. *Proc. Am. Assoc. Cancer Res.*, 43: 816-817, 2002.
115. DeFatta, R. J., Nathan, C. A., and De Benedetti, A. Antisense RNA to eIF4E suppresses oncogenic properties of a head and neck squamous cell carcinoma cell line. *Laryngoscope*, 110: 928-933, 2000.
116. Herbert, T. P., Fahraeus, R., Prescott, A., Lane, D. P., and Proud, C. G. Rapid induction of apoptosis mediated by peptides that bind initiation factor eIF4E. *Curr. Biol.*, 10: 793-796, 2000.
117. DeFatta, R. J., Li, Y., and De Benedetti, A. Selective killing of cancer cells based on translational control of a suicide gene. *Cancer Gene Ther.*, 9: 573-578, 2002.



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Mezzano SA, Droguett MA, Burgös ME, Ardiles LG, Aros CA, Caorsi I, Egido J.

Division of Nephrology, School of Medicine, Universidad Austral, Valdivia, Chile.
smezzano@uach.cl

Overexpression of chemokines, fibrogenic cytokines, and myofibroblasts in human membranous nephropathy. **BACKGROUND:** Proteinuria plays a central role in the progression of glomerular disease, and there is growing evidence suggesting that it may determine tubular cell activation with release of chemokines and fibrogenic factors, leading to interstitial inflammatory reaction. However, most studies on this subject have been performed in experimental models, and the experience in human kidney biopsies has been scarce. We analyzed the tissue sections of patients with idiopathic membranous nephropathy (IMN), a noninflammatory glomerular disease that may follow a progressive disease with heavy persistent proteinuria, interstitial cell infiltration, and decline of renal function. **METHODS:** Paraffin-embedded biopsy specimens from 25 patients with IMN (13 progressive and 12 nonprogressive) were retrospectively studied by immunohistochemistry [monocyte chemoattractant protein-1 (MCP-1), regulated on activation normal T-cell expressed and secreted chemokine (RANTES), osteopontin (OPN), platelet-derived growth factor-BB (PD-GF-BB)] and in situ hybridization [MCP-1, RANTES, PDGF-BB, transforming growth factor-beta1 (TGF-beta1)]. Moreover, we studied the presence of myofibroblasts, which were identified by the expression of alpha-smooth muscle actin (alpha-SMA), the monocytes/macrophages (CD68-positive cells), and T-cell infiltration (CD4+ and CD8+ cells). All of the patients were nephrotic and without treatment at time of the biopsy. **RESULTS:** A strong up-regulation of MCP-1, RANTES, and OPN expression was observed, mainly in tubular epithelial cells, with a significant major intensity in the progressive IMN patients. A strong correlation between the mRNA expression and the corresponding protein was noted. The presence of these chemokines and OPN was associated with interstitial cell infiltration. TGF-beta and PDGF were also up-regulated, mainly in tubular epithelial cells, with a stronger expression in the progressive IMN, and an association with the presence of myofibroblasts was found. **CONCLUSIONS:** Patients with severe proteinuria and progressive IMN have an overexpression in tubular epithelial cells of the chemokines MCP-1, RANTES, and OPN and the profibrogenic cytokines PDGF-BB and TGF-beta. Because this up-regulation was associated with an interstitial accumulation of mononuclear cells and an increase in myofibroblastic activity, it is suggested that those mediators are potential predictors of progression in IMN. Finally, based on experimental data and the findings of this article, we speculate that severe proteinuria is the main factor responsible for the up-regulation of these factors in tubular epithelial cells.



Decreased uncoupling protein expression and intramyocytic triglyceride depletion in formerly obese subjects.

Mingrone G, Rosa G, Greco AV, Manco M, Vega N, Hesselink MK, Castagneto M, Schrauwen P, Vidal H.

Istituto di Clinica Medica and. Clinica Chirurgica and Centro CNR Fisiopatologia Shock, Università Cattolica S Cuore, Rome, Italy. gmingrone@rm.unicatt.it

OBJECTIVE: To examine the muscular uncoupling protein expression 2 (UCP2) and UCP3 gene expression in morbid obese subjects before and after bariatric surgery [bilio-pancreatic diversion (BPD)]. **RESEARCH METHODS AND PROCEDURES:** Eleven obese subjects (BMI = 49 ± 2 kg/m²) were studied before BPD and 24 months after BPD. Skeletal muscle UCP2 and UCP3 mRNA was measured using reverse transcriptase-competitive polymerase chain reaction and UCP3 protein by Western blotting. Intramyocytic triglycerides were quantified by high-performance liquid chromatography. Twenty-four-hour energy expenditure and respiratory quotient (RQ) were measured in a respiratory chamber. **RESULTS:** After BPD, the average weight loss was approximately 38%. Nonprotein RQ was increased in the postobese subjects (0.73 ± 0.00 vs. 0.83 ± 0.02 , $p < 0.001$). The intramyocytic triglyceride level dropped (3.66 ± 0.16 to 1.60 ± 0.29 mg/100 mg of fresh tissue, $p < 0.0001$) after BPD. Expression of UCP2 and UCP3 mRNA was significantly reduced (from $35.9 \pm 6.1\%$ to $18.6 \pm 4.5\%$ of cyclophilin, $p = 0.02$; from $60.2 \pm 14.0\%$ to $33.4 \pm 8.5\%$, $p = 0.03$; respectively). UCP3 protein content was also significantly reduced (272.19 ± 84.13 vs. 175.78 ± 60.31 , AU, $p = 0.04$). A multiple regression analysis ($R(2) = 0.90$) showed that IMTG levels ($p = 0.007$) represented the most powerful independent variable for predicting UCP3 variation. **DISCUSSION:** The strong correlation of UCP expression and decrease in IMTG levels suggests that triglyceride content plays an even more important role in the regulation of UCP gene expression than the circulating levels of free fatty acids or the achieved degree of weight loss.

PMID: 12740453 [PubMed - indexed for MEDLINE]

Differential expression of the short and long forms of the gamma 2 subunit of the GABAA/benzodiazepine receptors.

Miralles CP, Gutierrez A, Khan ZU, Vitorica J, De Blas AL.

Division of Molecular Biology and Biochemistry, School of Biological Sciences,
University of Missouri-Kansas City 64110-2499.

The distribution of the mRNAs encoding the gamma 2S and gamma 2L subunits of the GABAA receptor in the rat brain has been revealed by in situ hybridization, northern blot and dot blot analysis using specific antisense oligonucleotides. In addition, the quantitative distribution of the gamma 2S and gamma 2L subunit peptides participating in the fully assembled GABAA receptors/benzodiazepine receptors has been mapped by immunoprecipitation with specific anti-gamma 2S and anti-gamma 2L antibodies. Several neuronal types and brain regions are enriched in gamma 2L such as neurons of the layer II of striate cortex and cerebellar Purkinje cells as well as the inferior colliculus, superior colliculus, deep cerebellar nuclei, medulla and pons. Other neuronal types and regions are enriched in gamma 2S such as the mitral cells of the olfactory bulb, pyramidal neurons of the pyriform cortex, layer VI of the neocortex, granule cells of the dentate gyrus and pyramidal cells of the hippocampus. Other cortical areas and cerebellar granule cells express both gamma 2S and gamma 2L in comparable amounts. There is a good correlation between the relative expression of gamma 2S and gamma 2L mRNAs and the relative presence of these protein subunits in fully assembled and mature receptors in the studied brain regions. The differential distribution of gamma 2S and gamma 2L might result in differential ethanol sensitivity of the neurons expressing these GABAA receptor subunits.

PMID: 7968350 [PubMed ; indexed for MEDLINE]

Follicle-Stimulating Hormone Receptor and Its Messenger Ribonucleic Acid Are Present in the Bovine Cervix and Can Regulate Cervical Prostanoid Synthesis¹

D. Mizrachi and M. Shemesh²

Department of Hormone Research, Kimron Veterinary Institute, Bet Dagan, Israel 50250

ABSTRACT

The hypothesis that FSH regulates the bovine cervical prostaglandin E₂ (PGE₂) synthesis that is known to be associated with cervical relaxation and opening at the time of estrus was investigated. Cervical tissue from pre-estrous/estrous, luteal, and postovulatory cows were examined for 1) the presence of bovine (b) FSH receptor (R) and its corresponding mRNA and 2) the effect of FSH on the PGE₂ regulatory pathway in vitro. The presence of bFSHR mRNA in the cervix (maximal during pre-estrous/estrous) was demonstrated by the expression of a reverse transcription (RT) polymerase chain reaction (PCR) product (384 base pairs) specific for bFSHR mRNA and sequencing. Northern blotting revealed three transcripts (2.5, 3.3, and 3.8 kilobases [kb]) in cervix from pre-estrous/estrous cows. The level of FSHR (75 kDa) was significantly higher ($p < 0.01$) in Western blots of pre-estrous/estrous cervix than in other cervical tissues. There was a good correlation between the 75-kDa protein expression and its corresponding transcript of 2.55 kb throughout the estrous cycle as described by Northern blot analysis as well as RT-PCR. Incubation of FSH (10 ng/ml) with pre-estrous/estrous cervix resulted in a 3-fold increase in the expression of FSHR and a 2-fold increase in both G protein (α) and cyclooxygenase II. FSH (5–20 ng/ml) significantly increased ($p < 0.01$) cAMP, inositol phosphate ($p < 0.01$), and PGE₂ ($p < 0.01$) production by pre-estrous/estrous cervix but not by cervix at the other stages. We conclude that bovine cervix at the time of the peripheral plasma FSH peak (pre-estrous/estrous) contains high levels of FSHR and responds to FSH by increasing the PGE₂ production responsible for cervical relaxation at estrus.

INTRODUCTION

FSH and LH are the two known pituitary hormones that regulate gonadal functions. FSH is essential for female fertility, specifically for folliculogenesis, and human ovarian failure has been related to mutations both in the FSH receptor (FSHR) and in the FSHB gene [1]. However, unlike LH, for which extra-gonadal binding has been documented for the uterus in the cow [2], pig [3], and human [4], no extra-gonadal effects of FSH have been reported. The uterine LH receptors have been shown to be physiologically active in the cow [2, 5]. The concentration of uterine LH receptors is related to the phase of the cycle, and binding of LH to uterine receptors increases uterine prostaglandin synthesis [2, 5]. Preliminary observations in our laboratory indicated that both FSH and LH receptors are also present in the cervix. We therefore investigated whether FSHRs are associated with the regulation of cervical prostaglandin, particularly around the time of estrus, when both cervical

PGE₂ production [6] and peripheral FSH concentrations are maximal.

The present investigation was therefore conducted to determine the presence of FSHR and its mRNA, and its physiological activity, in the bovine reproductive tract. Since the cervix is a prostaglandin E₁ (PGE₁)- and PGE₂-sensitive tissue and responds to hormonal stimulation, we also determined whether FSH could increase cervical cyclooxygenase expression and production of PGE₂ in vitro, particularly at the time near estrus.

MATERIALS AND METHODS

Animals

Cervical and ovarian tissues were collected from Holstein cows at a local abattoir. The stage of the cycle was determined according to signs of ovulation and the status of corpora lutea (CL; weight, color). These parameters allowed the classification of the cervix into three groups: 1) pre-estrous/estrous (follicular, 18–20 days postovulation; regressed CL < 1.0 g; presence of cervical mucus; $n = 54$); 2) postovulatory (1–4 days postovulation; presence of corpora hemorrhagica; signs of follicular rupture; $n = 35$), and 3) luteal (12–16 days postovulation; CL 4–6 g; $n = 44$). Bovine granulosa cells were collected from pre-estrous/estrous follicles and used as a positive control for FSHR expression [7].

The cervix external os segment was taken as the initial 3 cm of the cervix (total length of cervix is about 9 cm). The cervical os segments, primarily luminally oriented muscularis with epithelium, were sliced into horizontal strips (1 cm in length), pooled, and minced finely with a scalpel. The Kimron Veterinary Institute Animal Care and Use Committee approved all procedures.

FSHR Gene Expression

RNA isolation. Bovine cervical or ovarian tissues were rapidly dissected and frozen in liquid nitrogen (within 20 min after slaughter). Total RNA was extracted using the acid phenol, guanidine thiocyanate technique [8], using TriReagent (Molecular Research Center, Inc., Cincinnati, OH) according to the manufacturer's instructions.

Reverse transcription (RT)-polymerase chain reaction (PCR). Bovine FSHR mRNA was detected as previously described for the LH receptor mRNA [5]. Oligonucleotides corresponding to the published sequence of the bovine FSHR [7] were synthesized and used as primers in a PCR reaction. Bovine ovarian and cervical tissue cDNA obtained by RT of 2 μ g RNA was used as a template in the PCR reaction using a pair of primers corresponding to the transmembrane segments of the FSHR. The primers, 20-mer each, were selected using the OLIGO Program (Oligo R1 primer analysis software; National Biosciences, Plymouth, MN). The forward primer corresponded to position 1399–1419 (5'CGGCTTTTTCACCTGTCTTTG3') on the

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²Correspondence: M. Shemesh, Department of Hormone Research, Kimron Veterinary Institute, Bet Dagan, POB 12, Israel 50250. FAX: 972 3 9681753; e-mail: mshem@vs.moag.gov.il

bovine FSHR mRNA [7]. The reverse primer was a 20-mer oligonucleotide complementary to position 1763–1783 (5'-CGCTTGGCTATCTTGGTGTC3'). The predicted size of the RT-PCR product was 384 basepairs (bp). Four microliters of the RT reaction were used as a template for PCR; the reaction was allowed to proceed 35 cycles using 2 U *Termus Aquaticus* DNA polymerase (recombinant) provided by MBI Fermentas, Vilnius, Lithuania, and 200 pmol of each primer. The cycling parameters of the PCR were 98°C for 20 sec, 56°C for 45 sec, and 72°C for 45 sec. After amplification the samples were separated on a 1% agarose gel, stained with ethidium bromide, and photographed under UV light. The 384-bp fragment was extracted using the Wizard PCR Preps Kit (Promega, Madison, WI) and sequenced with the upper primer using an automatic sequencer (Applied Biosystems, Foster City, CA). For internal control, a bovine β -actin fragment of 890 bp was produced from an upper primer (5'-ACCA-ACTGGGACGACATGGAG3'; 21 mer) and a lower primer (5'-GCATTTGCGGTGGACAATGGA3'; 21 mer) as previously described [5]. Each PCR amplification was standardized using ovarian granulosa cDNA to produce a specific band of the predicted size. Standardization was based on temperature, cycle number, Mg level, and pH.

Western Blots

Cervical, bovine seminal vesicle, or muscle minces were sonicated for 45 sec, lysed in 500 μ l lysing buffer (1% Nonidet-40 [Sigma, St. Louis, MO], 2 mM EGTA, 5 mM $MgCl_2$, 1 mM PMSF in PBS) and incubated for 2 h at 4°C. After centrifugation, aliquots were taken for protein determination using gamma globulin (Sigma) as reference standard [9] and a protein dye binding method (Bio-Rad Laboratories, Richmond, CA). Sixty micrograms of protein of the soluble tissue extract was then separated by electrophoresis on 5–9%-gradient SDS-PAGE and electroblotted onto nitrocellulose paper (Amersham, Little Chalfont, Bucks, UK) as described previously [9]. The nitrocellulose membrane was washed with PBS containing 0.05% TWEEN-20 and blocked with 10% horse serum (Kimron Veterinary Institute) in washing solution. The nitrocellulose membrane was then treated with either 1) an anti-peptide antibody (code name 179) raised against human (h) FSHR peptide 265–295, diluted 1:500 (donation of Dr. J.A. Dias, Wadsworth Center, New York State Department of Health); 2) rabbit antiserum for bovine G protein (α_s) diluted 1:1000; UBI, Lake Placid, NY); or 3) rabbit anti-bovine cyclooxygenase polyclonal antiserum (diluted 1:200; Kimron Veterinary Institute) [9]. Different dilutions were used for each antibody, and the final dilution used was 75% of the dilution, which gave a maximal signal as previously described [9]. The nitrocellulose paper was then incubated with horseradish peroxidase-conjugated goat anti-rabbit IgG (Sigma Israel, Rehovot, Israel; diluted 1:2000 in washing buffer) for 1 h at room temperature. The presence of FSHR, G protein (α_s), or cyclooxygenase was then visualized by means of a color reaction as follows. The nitrocellulose paper was incubated in a substrate solution containing 3,3'-diaminobenzidine (0.5 mg/ml; Sigma) in a mixture of PBS containing 0.5% $CaCl_2$ and 6% H_2O_2 . The antibody to hFSHR recognized the 75-kDa protein of the FSHR. The antibody to G (α_s) recognized both the 42-kDa and 87-kDa forms of this protein, and the antibody to cyclooxygenase recognized the 72-kDa form (cyclooxygenase II). Extract of bovine seminal vesicles that are known to have a high

content of cyclooxygenase was used as positive control as previously described [9]. The densitometric scans were obtained using a bio-imaging system (B.I.S. 2020; Rhenium Dingo, Jerusalem, Israel) and processed with Tina 2.0 software (Fuji, Japan). Linearity of detection [9] was determined for densitometry for both Western and Northern blots. Each Western blot was evaluated in the absence of the first antibody, and no signal was detected.

Northern Blot Analysis

For Northern blots, 10 μ g of total RNA was denatured in 20 μ l 50% formamide/2.2 M formaldehyde in single-strength 3-(*n*-morpholino) propanesulfonic acid (MOPS) buffer (0.04 M MOPS, 10 mM sodium acetate, 1 mM EDTA) at 65°C for 10 min. Samples were placed on ice, and 5 μ l of loading buffer (0.5% SDS, 0.25% bromophenol blue, 25% glycerol, 25 mM EDTA) was added. Samples were subjected to electrophoresis through a 1.2% agarose formaldehyde gel and transferred by capillary blotting to nylon membranes (Nytran; Schleicher and Schuell, Keene, NH). RNA was subsequently UV cross-linked to membranes. The RT-PCR product fragments of 384 bp for FSHR and 890 bp for β -actin were used to generate biotinylated probes using random primer biotin labeling of DNA for chemiluminescence (NEBlot phototype kit; Biolabs, Beverly, MA) according to the manufacturer's instructions. The blotted membrane was prehybridized at 65°C for 1 h in prehybridization solution (6-strength SSC [3 M NaCl, 0.3 M sodium citrate], 5-strength Denhardt's reagent [1% ficoll, 1% polyvinylpyrrolidone, 1% BSA], 0.5% SDS, and 100 μ g/ml denatured salmon sperm DNAs) and hybridized overnight in the same solution with the denatured biotinylated probe to the target RNA. The membrane was then washed in double-strength SSC, 0.1% SDS at room temperature for 10 min and washed again in 0.1-strength SSC, 0.1% SDS at 68°C for 15 min followed by subsequent chemiluminescence, development, and detection on x-ray film (RX; Fuji Film, Tokyo, Japan). To obtain quantitative data for the specific mRNA, we used the densitometric value for β -actin mRNA to normalize each specific mRNA value. To remove FSHR probe, membranes were stripped using water for 15 min, then incubated in 0.4 M NaOH, 0.1% SDS at 80°C for 30 min. The membrane was rinsed again in 0.2 M Tris-HCl, 0.1-strength SSC for 30 min at 25°C. After stripping, prehybridization and hybridization with β -actin probe were performed as described for FSHR mRNA.

Cyclic AMP Determinations

Cervical tissue minces (50 mg/ml) were incubated for 10 min in wells of 1.5 ml containing 1.0 ml of Tissue Culture Medium-199 (TCM-199 without serum (Biological Industries, Beit Haemek, Israel) in the absence or presence of physiological (5, 10 ng/ml) and pharmacological doses (20, 40 ng/ml) of FSH (USDA-bFSH-8-1; no further increase in cAMP production was seen at 50 or 100 ng/ml) or forskolin, a stimulator of adenylate cyclase (10 μ M; Sigma). The dose of forskolin was selected to give a 3-fold increase in cAMP. At the end of the incubation, tissues were removed, blotted on filter paper to remove mucus, and incubated overnight at 4°C with 400 μ l of 3% $HClO_4$. The solution was then neutralized with 150 μ l of $KHCO_3$ and centrifuged, and 50- μ l aliquots were taken for the radioreceptor assay as described by Brown et al. [10] and modified for endometrium by Miyazaki et al. [11]. Standards (0–1000 pg of cAMP) or samples were incubated at room tem-

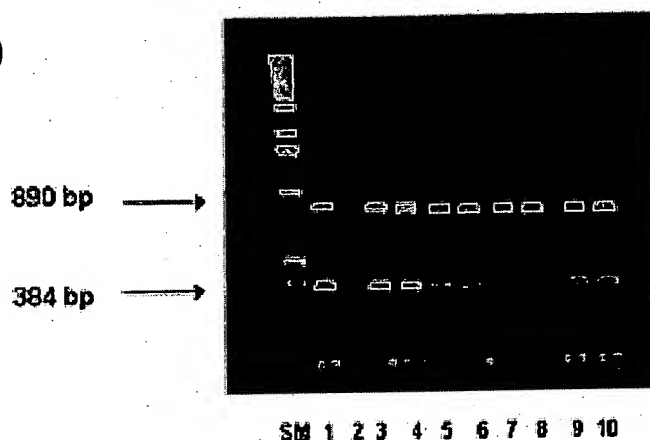


FIG. 1. RT-PCR amplification of bovine cervix FSHR and β -actin mRNA. RT-PCR was performed as described in *Materials and Methods* for FSHR and β -actin. Separate RT-PCR amplification products of FSHR (estimated 384 bp) and β -actin (estimated 890 bp) were combined and separated on agarose gel electrophoresis and stained with ethidium bromide. RT-PCR was performed using 2 μ g RNA of bovine granulosa cells as a positive control (lane 1), cervical tissue at the pre-estrous/estrous (lanes 3–4), post-ovulatory (lanes 5–6), and luteal phases (lanes 7–8). Lanes 9–10 represent reamplification, using as template the PCR product, cDNA, from the luteal-phase amplification. Lane 2 represents negative control in the absence of RNA. The tissue was taken from 4 cows at each stage of the estrous cycle, and two representative samples from each stage are shown. SM, size markers.

perature with 100 μ l of cAMP binding protein prepared from bovine adrenal extracts as described [10]. The buffer used for the assay was added to form a final reaction volume of 400 μ l. After 2-h incubation in a cold room, the reaction was stopped by adding 300 μ l of a charcoal-dextran solution; centrifugation followed, and the supernatant was removed for counting in a scintillation counter. The sensitivity of the assay was 60 pg/tube. Quadruplicate assays were made for each value determined. The within-assay and between-assay coefficients of variance were 8% and 10%, respectively.

1401
 BTFSH GCTTTTTCAC TGCTTTGCC AGTGAGCTCT CAGTCTACAC TCTGACCGCC
 Cervix GCTTTTTCAC TGCTTTTACT AGTGAGCTCT CAGTCTACAC TCTGACCGCC
 BTFSH ATCAGCTGG AAAGATGGCA TACCATCACC CATGCCATGC AGCTCGAATG
 Cervix ATCAGCTGG AAAGATGGCA TCCCATCACC CGTGCCATGC AGCTCGAATG
 BTFSH CAAAGTCGAC CTCCGCCATG CTGCCAGCAT CATGCTGGTG GGCTGGATCT
 Cervix CAAAGTCGAC CTCCGCCATG CTGCCAGCAT CATGCTGGTG GGCTGGATCT
 BTFSH TTGCTTTTGC AGTTGCCCTT TTTCCCATCT TTGGCATCAG CAGCTACATG
 Cervix TTGCTTTTGC AGTTGCCCTT TTTCCCATCT TTGGCATCAG CAGCTACATG
 BTFSH AAGGTGAGCA TCTGCTGCC CATGGACATT GACAGCCCTT GTGCACAAT
 Cervix AAGGTGAGCA TCTGCTGCC CATGGACATT GACAGCCCTT GTGCACAAT
 BTFSH CTATGTCATG TCCCTCCTTG TGCTCAATGT CCTGGCCTTT GTGGTCATCT
 Cervix CTATGTCATG TCCCTCCTTG TGCTCAATGT CCTGGCCTTT GTGGTCATCT
 BTFSH GTGGCTGCTA CACTCACATC TACCTCACGG TGAGGAACCC CAACATCACA
 Cervix GTGGCTGCTA CACTCACATC TACCTCACGG TGAGGAACCC CAACATCACA
 1761
 BTFSH TCCTCTTCTA GTG
 Cervix TCCTCTTCTA GTG

FIG. 2. Automated nucleotide sequencing and homology between *Bos taurus* mRNA FSHR (GenBank accession number L22319) from nucleotides 1401 to 1763 and the PCR products obtained using the upper primer as described in *Materials and Methods*. A homology of 97.5% was found between *Bos taurus* and our amplified cDNA obtained by RT-PCR from cervical mRNA at pre-estrous/estrous, suggesting that they are complementary.

Measurement of Phospholipase C (PLC) Activity

Cervical minces (30 mg) were incubated in 1 ml TCM-199 containing 5 mCi [3 H]myo-inositol for 90 min. Slices were rinsed with 1 ml TCM-199 and incubated for another 60 min to remove unincorporated [3 H]myo-inositol. At the end of the preincubations, slices were incubated in 1 ml TCM-199 containing 10 mM LiCl to inhibit inositol phosphate (IP) hydrolysis, allowing [3 H]-IP, [3 H]-IP₂, and [3 H]-IP₃ to accumulate in the tissue after stimulation of PLC, in absence or presence of 0–20 ng/ml of FSH or 0–20 ng/ml LH (USDA bLH-I-1). Incubations were terminated by adding 2 ml ice-cold chloroform/methanol/hydrochloric acid (5:10:0.1, v:v:v) and then 1 ml chloroform and 1 ml EDTA (5 mM) and extracted as described by Kisielewska et al. [12]. [3 H]-IP, [3 H]-IP₂, and [3 H]-IP₃ were separated from the labeled inositol compounds by using Dowex AG 1-X8 (formate form). [3 H]-Labeled IP was eluted from the column by sequential elution into 10 fractions. Aliquots of each fraction were added to scintillation vials containing 5 ml scintillation fluid, and radioactivity was determined using a scintillation counter. Total activity of PLC is expressed as cpm of total IPs/30 mg/30 min.

RIA for PGE₂ and PGF_{2 α}

Aliquots of 100 μ l were taken at the end of the incubation period for specific RIA of PGE₂ and PGF_{2 α} , which were performed without chromatographic separation. The antisera for PGF_{2 α} (Sigma) reacts preferentially with PGF_{2 α} but cross-reacts with PGF_{1 α} (60%) and to a negligible extent (< 0.1%) with prostaglandins of the A, B, and E series. The antisera to PGE₂ (Sigma Israel) reacts preferentially with PGE₂ but cross-reacts with PGE₁ (20%), PGA₁, PGA₂, PGF_{1 α} , and PGF_{2 α} (< 10%) and to a negligible extent with PGB₁ and PGB₂ (< 0.1%). The intraassay coefficients of variation were 9% and 11%, and the interassay coefficients of variance were 12.3% and 13% for PGF_{2 α} and PGE₂.

Statistical Analysis

ANOVA was performed with a significance level of $p < 0.05$. Data were further analyzed using Tukey's procedure ($p < 0.05$ or $p < 0.01$) to assess significance between treatments. Student's t -test was used where appropriate. Values are expressed as means \pm SEM.

RESULTS

FSHR mRNA Expression in the Cervix

RT-PCR demonstrated that the FSHR gene is expressed in bovine cervix as indicated by the presence of the 384-bp fragment corresponding to bFSHR mRNA. The mRNA was maximally expressed in the cervix during the pre-estrous/estrous phase. In contrast, the expression of the gene was reduced in postovulatory cervix and not detectable in luteal-phase cervix (Fig. 1). However, reamplification of the PCR product, cDNA, from the luteal phase produced a detectable signal (Fig. 1).

To demonstrate that the absence of FSHR mRNA in luteal-phase cervix was specific, bovine β -actin mRNA was used as internal control. PCR demonstrated that the specific 890 bp of the β -actin cDNA band was produced by the cervical tissues of all the stages. The band for β -actin was

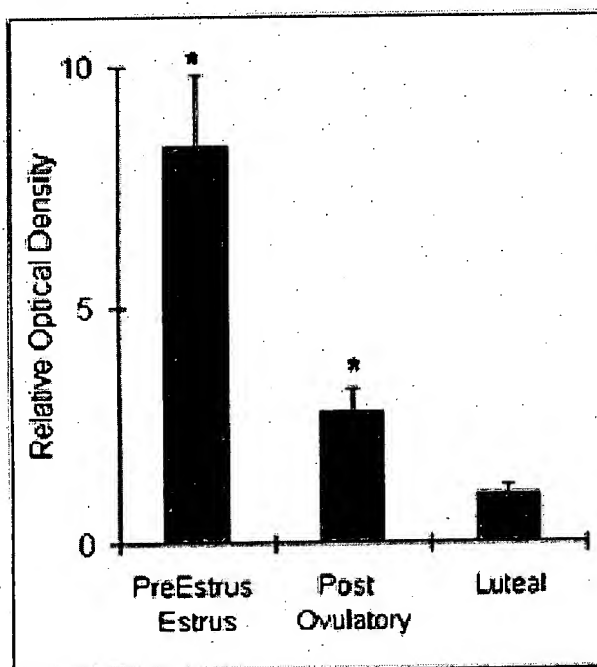
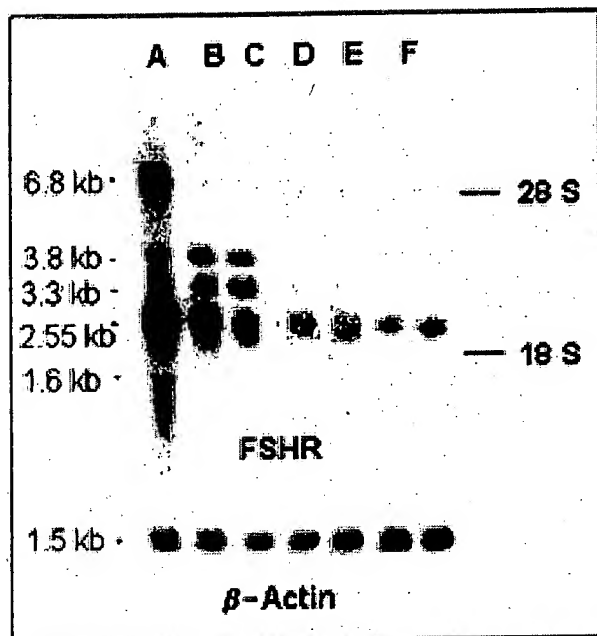


FIG. 3. Northern blot for analysis of total RNA (10 μ g) extracted from bovine cervical tissues collected from pre-estrous/estrous, postovulatory, and luteal-phase cows, and bovine granulosa cells as a positive control. Relative migration of 28S and 18S ribosomal RNA is indicated. To localize mRNA for FSHR and β -actin, our RT-PCR products (384-bp and 890-bp fragments, respectively) were used to generate biotinylated probes using biotin random primer. Figure shows results for FSHR mRNA in bovine granulosa cells as a positive control (lane A); and in cervical tissues at pre-estrous/estrous (lanes B–C), postovulation, (lanes D–E) and the luteal phase (lanes F–G). The corresponding β -actin band was obtained by re-probing of the membranes previously tested for FSHR mRNA as described in *Materials and Methods*. Columns show mean \pm SEM of the optical density of the ratio FSHR: β -actin, in which the luteal phase is given an arbitrary value of 1.0. *Significantly different from luteal-phase value ($p < 0.01$); $n = 5$ for each stage.

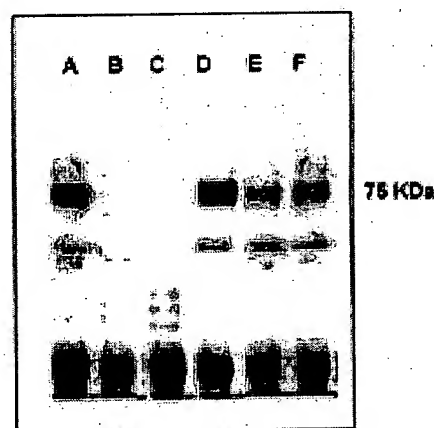
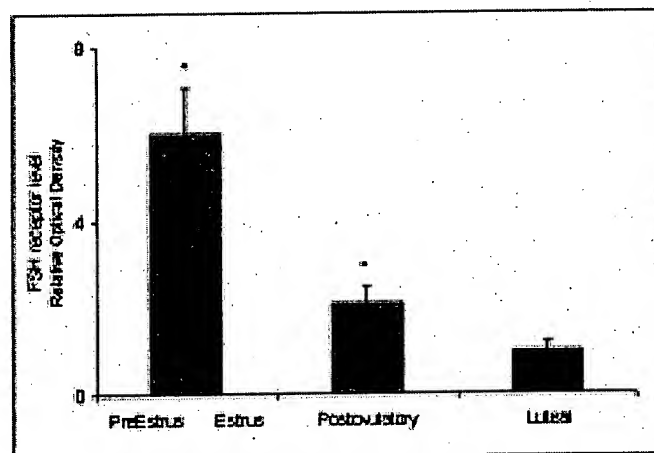


FIG. 4. Presence of FSHR protein in cervical tissues as determined by Western blot. Soluble cell extracts (60 μ g protein) were used to determine the amount of FSHR as described in *Materials and Methods*. Western blot proteins were transferred to nitrocellulose paper and probed with anti-serum to peptide hFSHR, 265–296. Western blot displays a representative result for bovine granulosa cells, used as positive control (lane A); bovine seminal vesicles (lane B) and muscle, used as a negative control (lane C); and cervical tissues from three stages of the estrous cycle: pre-estrous/estrous (lane D), postovulatory (lane E), and luteal (lane F). Histogram summarizes the results for the three stages of the estrous cycle. Columns show mean \pm SEM of the relative optical density in which the luteal phase is given an arbitrary value of 1.0. *Significantly different from luteal-phase value ($p < 0.01$); $n = 7$ for each stage.

present even though the 384-bp band corresponding to the FSHR was absent (Fig. 1).

The nucleotide sequence (Fig. 2) of the 384-bp fragment obtained from pre-estrous/estrous cervix was compared with that for the *Bos taurus* FSHR reported by Houde et al. [7]. It was found that there was a 97.5% homology between the 384-bp fragment and the nucleotide range 1410–1763 found in the *Bos taurus* FSHR.

The Northern blot contained the expected three transcripts of 2.55, 3.3, and 3.8 kilobases (kb) present in the bovine granulosa. However, the high (6.8-kb) and low (1.6-kb) transcripts found in the ovary were not found in the cervix. The three transcripts were found in pre-estrous/estrous cervix, but only one transcript (2.55 kb) was seen in luteal cervix (1/8 of the pre-estrus/estrus level; $n = 5$) or postovulatory cervix (1/3 of the pre-estrus/estrus level; $n = 5$) (Fig. 3).

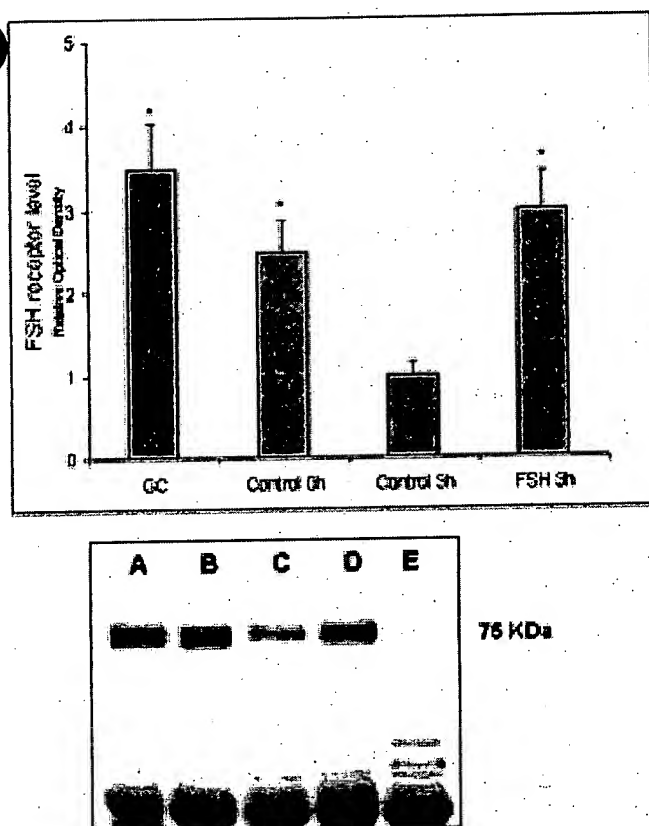


FIG. 5. Effect of incubation of cervical tissues with FSH on the expression of the FSHR protein. Cervical tissue from 5 pre-estrous/estrous cows was incubated in the absence or presence of FSH (0 or 10 ng/ml). Western blot is representative for FSH protein concentration after 3 h of incubation, with granulosa cell extract serving as positive control. Lane A, Granulosa cells (GC); lane B, cervical tissue at Time 0; lane C, cervical tissue after 3 h of incubation in TCM-199; lane D, cervical tissue after 3 h of incubation with 10 ng/ml of FSH. Lane E represents muscle tissue incubated with FSH (10 ng/ml) for 3 h as a negative control. Histogram summarizes the results of six experiments. Columns are means \pm SEM. *Significantly different ($p < 0.01$) from the column representing the control at 3 h (a given value of 1).

FSHR Protein in the Cervix

The presence of FSHR protein in the cervix was demonstrated as determined by Western blot analysis using a specific antipeptide antibody for hFSHR. The antibody recognized a 75-kDa protein in both bovine granulosa and cervical preparations identical with the predicted molecular size of FSHR. The signal for the 75-kDa protein was strongest in pre-estrous/estrous cervix when compared with postovulation- (3-fold) or luteal-phase (6-fold) cervix (Fig. 4). With the use of 60 μ g protein in each lane, this protein was observed to be expressed throughout the estrous cycle.

In Vitro Effect of FSH on Induction of FSHRs

A time-course (0.75, 1.5, and 3 h) and dose-response (0, 10, and 20 ng/ml) study was carried out to determine in vitro effect of FSH on its own receptor. Cervical minces (100 mg) from cows in the pre-estrous/estrous, postovulatory, and luteal phases were used. It was found, as determined by Western blot, that FSH induced its own receptor in cervical tissues at pre-estrus/estrus, with a 3-fold increase ($p < 0.01$) observed at 10 ng/ml after 3 h of incubation (Fig. 5). However, no effect of FSH was seen at earlier

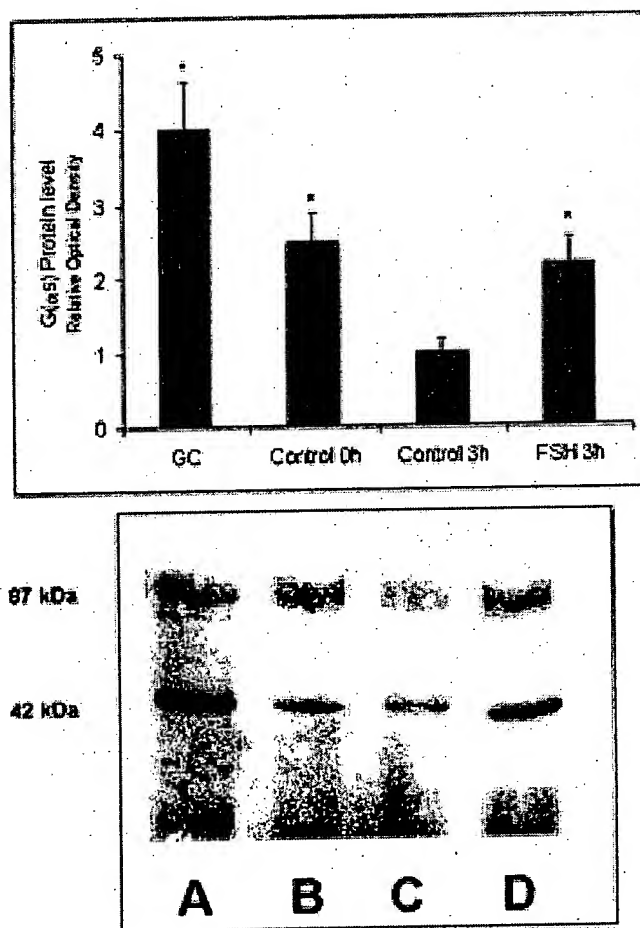


FIG. 6. Effect of incubation of cervical tissues with FSH on the synthesis of G protein. Cervical tissues from 5 pre-estrous/estrous cows were incubated in the absence or presence of FSH (10 ng/ml). Both the 87-kDa G protein (α , β , γ complex) and 42-kDa protein (α_s) are elevated at 3 h of incubation. Granulosa cell extract was used as a positive control. Lane A, Granulosa cells (GC); lane B, cervical tissue at Time 0; lane C, cervical tissue after 3 h of incubation in TCM-199; lane D, cervical tissue after 3 h of incubation with 10 ng/ml of FSH. Columns are means \pm SEM of 42-kDa protein. *Significantly different ($p < 0.05$) from the column representing the control at 3 h (with an arbitrary value of 1.0).

times at the 10 ng/ml level. When 20 ng/ml was used, a stimulatory effect (2-fold) was seen after 1.5 h. No significant response was observed when tissues of the postovulatory or luteal phase were used at any dose or time tested (data not shown).

Effect of FSH Treatment on G Protein (α_s)

A time-course and dose-response study was carried out to determine whether cervical FSHR was coupled to G protein. Cervical minces (100 mg) from cows in the pre-estrous/estrous, postovulatory, and luteal phases were incubated in the presence of FSH (0–20 ng/ml) for 0.75, 1.5, and 3 h. It was found in pre-estrous/estrous cervical tissue that, in the presence of FSH (10 ng/ml), there was a significantly ($p < 0.05$) higher level of G protein (α_s) at the level of both the 87-kDa (α , β , γ complex) and the 42-kDa (α_s) proteins after 3 h of incubation (Fig. 6). FSH, at either 10 or 20 ng/ml, did not have a stabilizing effect on the level of G protein (α_s) at shorter times of incubation (0.75 or 1.5 h). No significant response was observed when tis-

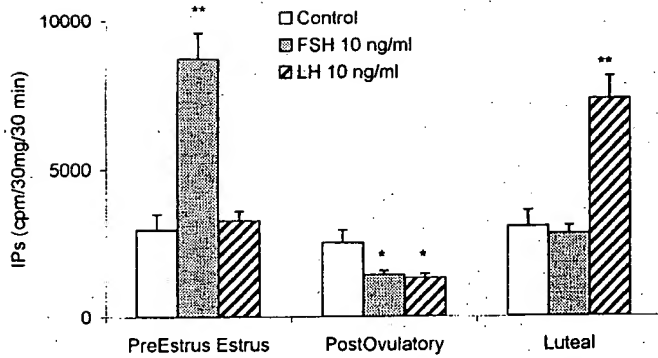


FIG. 7. Effect of FSH on IP production in cervical tissue. Cervical tissues from six cows at each of three stages of the cycle were incubated for 30 min in the absence or presence of FSH (10 ng/ml) or LH (10 ng/ml). IPs produced were determined in four replicates by amount of labeled compound (cpm) produced. Columns are means \pm SEM (* p < 0.05; ** p < 0.01; significantly different from control value for each stage of the cycle).

sues of the postovulatory or luteal phase were used at any dose or time tested (data not shown).

IP Pathway Activation by FSH

For IP measurement, cervical tissue minces (30 mg) from pre-estrous/estrous, postovulatory, and luteal-phase cows were incubated for 30 min in the absence or presence of FSH (10 ng/ml) or LH (10 ng/ml). FSH significantly (p < 0.01) increased the level of IP, IP_2 , and IP_3 at pre-estrous/estrous. However, FSH significantly (p < 0.05) inhibited IP level in the postovulatory stage. In contrast, LH, but not FSH, stimulated IPs significantly (p < 0.01) at the luteal phase (Fig. 7).

Adenylate Cyclase Activation by FSH

A dose-response analysis was used to determine whether the cervical FSHR was coupled to adenylate cyclase. In these experiments, cervical tissue minces (50 mg) obtained from pre-estrous/estrous, postovulatory, and luteal-phase cows were incubated with FSH (0–40 ng/ml). FSH increased cAMP accumulation in a dose-dependent manner when incubated with pre-estrous/estrous cervical tissues, with maximal stimulation 2.5 times (p < 0.01) that of control in the presence of 10 ng/ml (Fig. 8A). In contrast, cervical tissue from luteal or postovulation did not respond to FSH even though tissues from all phases of the cycle responded to forskolin with a significant (p < 0.01) increase in cAMP (Fig. 8B).

Effect of FSH on PGE Production by Cervical Tissues

A dose-response analysis was carried out to determine whether FSHR was associated with cervical prostaglandin production. PGE_2 and $PGF_{2\alpha}$ production by cervical minces (100 mg/ml) in the presence of FSH (0–20 ng/ml) was determined by RIA. FSH stimulated (p < 0.01) basal PGE_2 production in a dose-dependent manner and caused a 3-fold increase in PGE_2 production at 10 ng/ml in pre-estrous/estrous cervical tissues (n = 8, Fig. 9). FSH had no effect on the small amount of basal $PGF_{2\alpha}$ (< 1 ng/100 mg) produced by cervical tissue (data not shown). FSH did not elevate PGE_2 in cervical tissues from the luteal phase; furthermore, a small but significant (p < 0.05) inhibition was observed at postovulation (Fig. 9).

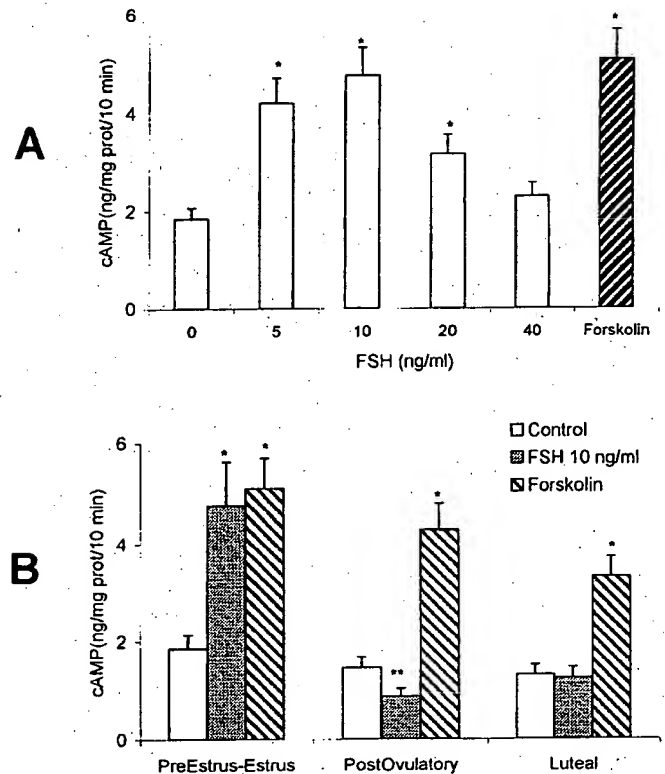


FIG. 8. Effect of FSH on adenylate cyclase production in cervical tissue. A) Cervical tissues from six cows at pre-estrous/estrous were incubated (4 replicates) for 10 min in the absence or presence of FSH (0, 5, 10, 20, or 40 ng/ml) or forskolin (10 μ mol/L), and the amount of cAMP produced was measured by a protein binding assay. B) Cervical tissues from six cows at each of the three stages of the estrous cycle were incubated (4 replicates) for 10 min in the absence or presence of FSH (10 ng/ml) or forskolin (10 μ M). The amount of cAMP produced was measured by a protein binding assay. Columns with asterisks were statistically different from their own control (* p < 0.01; ** p < 0.05).

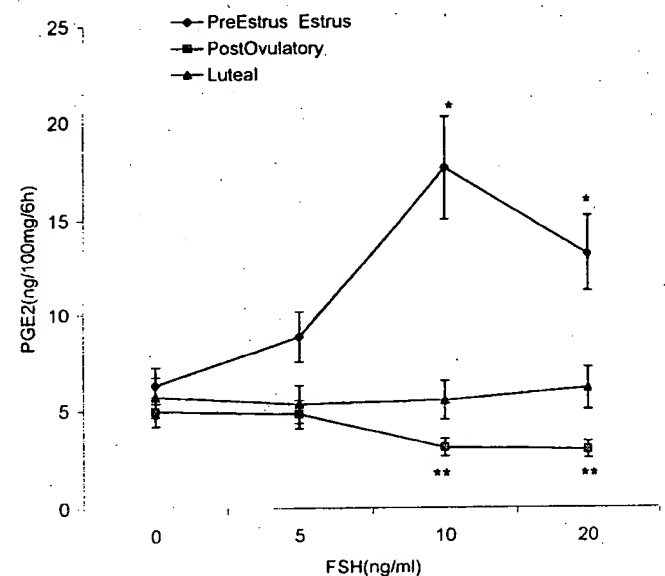


FIG. 9. FSH dose response on production of PGE_2 by cervical tissues. Cervical tissues from 8 pre-estrous/estrous, postovulatory, or luteal-phase cows were incubated in the absence or presence of FSH (5, 10, or 20 ng/ml) for 6 h, and the effect on the production of PGE_2 was measured by RIA. * p < 0.01; ** p < 0.05.

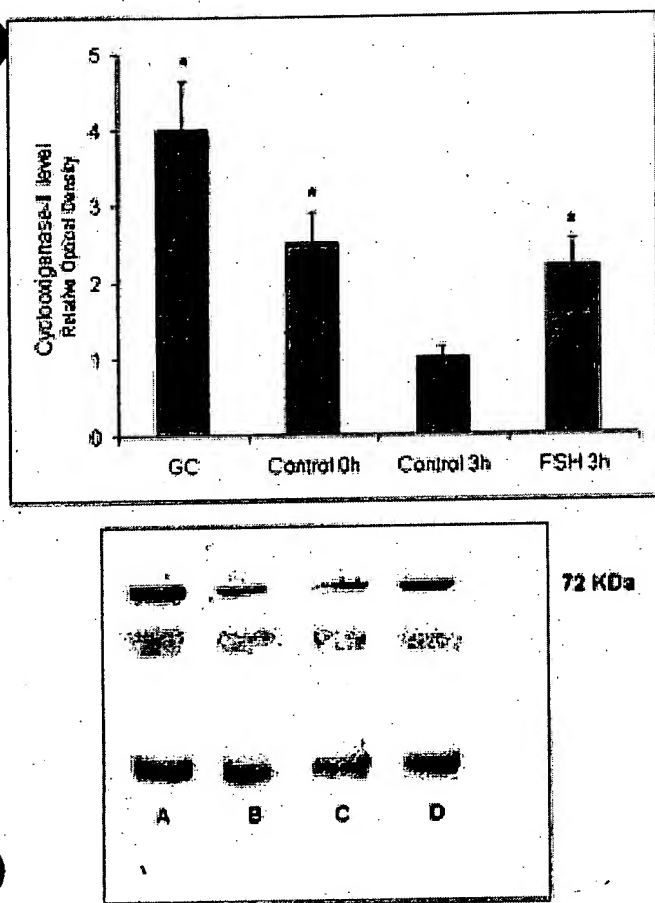


FIG. 10. Effect of incubation of cervical tissues with FSH on the expression of cyclooxygenase. Cervical tissues from 5 pre-estrous/estrous cows were incubated in the absence or presence of FSH (10 ng/ml). The 72-kDa cyclooxygenase II protein was elevated at 3 h of incubation. Lane A, bovine granulosa cells (GC); lane B, cervical tissue at Time 0; lane C, cervical tissue after 3 h of incubation in TCM-199; lane D, cervical tissue after 3 h of incubation with 10 ng/ml of FSH. Columns are means \pm SEM of 72-kDa protein. *Significantly different ($p < 0.01$) from the column representing the control at 3 h (with an arbitrary value of 1.0).

Cyclooxygenase Activation by FSH

To determine whether the presence of FSHR was associated with the expression of cyclooxygenase, cervical minces were incubated for 3 h in the presence or absence of FSH (10 ng/ml). Tissues from pre-estrous/estrous, post-ovulatory, and luteal-phase cows were extracted, separated on SDS-PAGE, and tested for the cyclooxygenase II (72 kDa) using a specific antibody. It was found that in cervical tissues from six pre-estrous/estrous cows, FSH induced a 200% increase in the expression of cyclooxygenase after 3 h of incubation (Fig. 10). In contrast, no effect on cyclooxygenase expression was seen in cervical tissues from luteal-phase cows, and an insignificant ($p > 0.05$) elevation at the postovulatory phase was observed (data not shown).

DISCUSSION

Results of this study demonstrate by both PCR amplification and Northern blot analysis that FSHR mRNA is present in the bovine cervix. The bands corresponding to the receptor (75 kDa) and its mRNA were maximally expressed during the pre-estrous/estrous phase. Incubation of cervical tissues with FSH increased the expression of cyclooxygenase

ase and the production of PGE_2 (and possibly PGE_1) at the pre-estrous/estrous phase. The stimulation of cervical cyclooxygenase was associated with activation of both the PLC and adenylate cyclase second messenger G-protein related pathways.

Northern blot analysis of bovine cervical tissues from pre-estrous/estrous cows revealed multiple transcripts for bovine cervical FSHR. The sizes of the major transcripts were 2.5 kb, 3.3 kb, and 3.8 kb, which were similar to those obtained in bovine granulosa cells [7], rat testis and ovary [13, 14], and human ovary [15] and myometrium [16]. Other minor transcripts have been reported in all of these species. The RT-PCR product yielded a single band of 384 bp, which was the expected molecular size, so the different isoforms of the FSHR are probably the result of differential splicing of the same transcript. Isoforms of FSHR have been characterized in human [17] and primate [18] ovary and in ovine testis [19]. Since only a single 2.5-kb isomer was present in luteal-phase cervix (which did weakly produce FSHR), it would appear that there are different isoforms present in the bovine cervix as well.

The nucleotide sequence homology between our RT-PCR 384-bp product was 97.3% identical to the comparable region of the bovine FSHR, showing that our amplified cDNA was complementary to the *Bos taurus* mRNA FSHR. The PCR product of 384-bp cDNA was found primarily in the pre-estrous/estrous cervix.

Using specific antibody raised in rabbits against hFSHR (amino acid sequence 265–296) for Western blot analysis resulted in a major signal for a 75-kDa protein. A protein of similar molecular mass has been reported in rat and human ovary [20]. A strong signal for this protein was seen in the pre-estrous/estrous cervix compared with cervix from the other stages of the cycle. This cervical receptor was regulated by FSH itself, similar to the regulation of the FSHR in the rat ovary [14].

FSH increased cAMP production by cervical tissue from the pre-estrous/estrous phase but did not elevate cAMP in the postovulatory and luteal phases. FSH elevation of IPs displayed the same pattern. It would therefore appear that the cervical FSHR is associated with signal transduction pathways in a way similar to the accepted mechanism of LH action in the ovary and testes, i.e., activation of the second messenger pathways—adenylate cyclase [21] and phosphatidylinositol (PLC) [22, 23]. The inhibitory effect of FSH on IPs and cAMP at postovulation occur when FSHR is low. Since FSH activity is biphasic, i.e., is stimulated by a low dose and inhibited by a high dose, an inhibitory effect at low concentrations could be the result of the change in the ligand-receptor ratio.

FSH induced about a 200% increase in the expression of cyclooxygenase at pre-estrus/estrus but had no effect on the two other stages of the cycle. This was similar to the response of cyclooxygenase to LH that was observed for the bovine endometrium [2] and uterine vein [6]. However, in the endometrium, LH increased cyclooxygenase during both the luteal and postovulatory phases but not at estrus [2]. Furthermore, the induction of cyclooxygenase by gonadotropin in the endometrium was associated with an increased level of $\text{PGF}_{2\alpha}$, while in the cervix PGE_2 was the major product.

The bFSH was of immunological grade and devoid of residual biological contamination of LH, thyroid-stimulating hormone, growth hormone, prolactin, or ACTH. The effects on increasing cAMP, PGE_2 , and cyclooxygenase as well as induction of FSHR were therefore specific. Fur-

thermore, it was found that highly purified hFSH (10–20 ng/ml) (donation of J.A. Dias) had the same effects as bFSH when incubated with cervical tissue under the same conditions (2.5-fold increase in cAMP; 3-fold increase in PGE₂; 2-fold increase in cyclooxygenase).

The relaxation and opening of the gravid cervix is due to active biochemical and structural changes in the cervical connective tissue that are mediated in part by prostaglandins [24]. It is thought that stretching of the cervix is a factor in causing the release of these prostaglandins [25, 26]. The effect of PGE in causing cervical softening in the ewe is also well documented [27–29]. In both nonpregnant and pregnant cows, PGE₂ causes an increase in cervical opening within 3 h of treatment [30]. PGE₁ administered intra-cervically in a jelly was shown to decrease cervical resistance within 24 h [31].

Recently, we reported [6] that oxytocin caused a significant stimulation of PGE₂ production in vitro in cervical tissues from pre-estrous/estrous cows but had no effect on PGE₂ production in cervical tissue from other stages of the estrous cycle of the cow. Similarly, in an initial report [31], it was shown that administration of oxytocin to pre-estrous cows increased the concentration of PGE₂ in the cervical exudate. However, peripheral oxytocin concentrations during estrus are lower than during the luteal phase [32, 33], and elevations of both peripheral oxytocin and cervical oxytocin receptor concentration are necessary for oxytocin to cause cervical softening towards parturition [24]. Furthermore, progesterone in vitro induced a dose-dependent inhibition of PGE₂ release by cervical tissues from pre-estrous/estrous cows, and this was associated with a decrease in both basal and oxytocin-stimulated PGE₂ production [6]. It would therefore seem that hormones other than oxytocin are responsible for the increase in cervical PGE₂ in the pre-estrous/estrous cow. The present report indicates that FSH, which has its peak peripheral concentration at the time of estrus, could be the hormone that increases cervical PGE₂, as the FSHR expression is maximal at this time and FSH in vitro increases PGE₂ production by the cervix.

The bovine cervix at pre-estrus/estrus has high levels of FSHR protein and its corresponding mRNA. Activation of the receptor by FSH is associated with the G-protein-coupled receptor family that mediates the cAMP and IP signalling pathways. These signalling pathways then increase the expression of cyclooxygenase and production of PGE₂. The expression of the FSHR was maximal at the time of the FSH peak in the blood and suggests a physiological role for FSH in the relaxation and opening of the cervix at estrus.

ACKNOWLEDGMENTS

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REFERENCES

1. Dias JA, Lindau-Shepard B, Hauer C, Auger I. Human follicle-stimulating hormone structure-activity relationships. *Biol Reprod* 1998; 58:1331–1336.
2. Friedman S, Gurevich M, Shemesh M. Bovine cyclic endometrium contains affinity luteinizing hormone/human chorionic gonadotropin. *Biol Reprod* 1995; 52:1020–1026.
3. Ziecik AJ, Stanchev PD, Tilton JE. Evidence for the presence of luteinizing hormone/human chorionic gonadotropin-binding sites in the porcine uterus. *Endocrinology* 1986; 119:1159–1163.
4. Reshef E, Lei ZM, Rao ChV, Pridham DD, Chegini N, Luborsky JL.

- The presence of gonadotropin receptors in nonpregnant human uterus, human placenta, fetal membranes, and decidua. *J Clin Endocrinol Metab* 1990; 70:421–430.
5. Shemesh M, Gurevich M, Mizrahi D, Dombrowski L, Stram Y, Fields MJ, Shore LS. Expression of functional luteinizing hormone receptor and its mRNA in bovine uterine veins; luteinizing hormone induction of cyclooxygenase and augmentation of prostaglandin production in bovine uterine veins. *Endocrinology* 1997; 138:4844–4851.
6. Shemesh M, Dombrowski L, Gurevich M, Friedman S, Shore LS, Fuchs AR, Fields MF. Regulation of bovine cervical secretion of prostaglandins and synthesis of cyclooxygenase by oxytocin. *Reprod Fertil Dev* 1997; 9:525–530.
7. Houde A, Lambert A, Saumande J, Silversides DW, Lussier JG. Structure of bovine follicle-stimulating hormone receptor complementary DNA and expression in bovine tissues. *Mol Reprod Dev* 1994; 39:127–135.
8. Chomczynski P, Sacchi N. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal Biochem* 1987; 162:156–159.
9. Izhar M, Pasmanik M, Marcus S, Shemesh M. Dexamethasone inhibition of cyclooxygenase expression in bovine term placenta. *Prostaglandins* 1992; 43:239–254.
10. Brown BL, Albano JDM, Ekins RP, Sgherzi AM, Tampion W. A simple and sensitive saturation assay method for measurement of adenosine 3':5'-cyclic monophosphate. *Biochem J* 1971; 121:561–562.
11. Miyazaki K, Tanaka N, Kawakami S, Okamura H. Adenylyl cyclase activity in human decidua and myometrium during pregnancy and labour. *J Reprod Fertil* 1995; 105:141–145.
12. Kisielska J, Flint APF, Ziecik AJ. Phospholipase C and adenylyl cyclase signalling systems in the action of hCG on porcine myometrial smooth muscle cells. *J Endocrinol* 1996; 148:175–180.
13. Heckert LL, Griswold MD. Expression of follicle-stimulating hormone receptor mRNA in rat testes and sertoli cells. *Mol Endocrinol* 1991; 5:670–677.
14. LaPolt PS, Tilly JL, Aihara T, Nishimori K, Hsueh AJW. Gonadotropin-induced up-and-down-regulation of ovarian follicle-stimulating hormone (FSH) receptor gene expression in immature rats: effects of pregnant mare's gonadotropin, human chorionic gonadotropin, and recombinant FSH. *Endocrinology* 1992; 130:1289–1295.
15. Tilly JL, Aihara T, Nishimori K, Jia X-C, Billig H, Kowalski KI, Perlas EA, Hsueh AJW. Expression of recombinant human follicle-stimulating hormone receptor: species-specific ligand binding, signal transduction, and identification of multiple ovarian messenger ribonucleic acid transcripts. *Endocrinology* 1992; 131:799–806.
16. Kornyei JL, Li X, Lei ZM, Rao ChV. Restoration of human chorionic gonadotropin response in human myometrial smooth muscle cells by treatment with follicle-stimulating hormone (FSH): evidence for the presence of FSH receptors in human myometrium. *Eur J Endocrinol* 1996; 134:225–231.
17. Gromell J, Gutermann T, Nieschlag E. Molecular cloning of truncated isoform of the human follicle stimulating hormone receptor. *Biochem Biophys Res Commun* 1992; 188:1077–1083.
18. Gromell J, Dankbar B, Sharma RS, Nieschlag E. Molecular cloning of the testicular follicle stimulating hormone receptor of the nonhuman primate *Macaca fascicularis* and identification of multiple transcripts in the testes. *Biochem Biophys Res Commun* 1993; 19:1066–1072.
19. Khan H, Yarney TA, Sairam MR. Cloning of alternately spliced mRNA transcripts coding for variants of ovine testicular follitropin receptor lacking the G protein coupling domains. *Biochem Biophys Res Commun* 1993; 190:888–894.
20. Liu X, DePasquale JA, Griswold MD, Dias JA. Accessibility of rat and human follitropin receptor primary sequence (R265-S296) in situ. *Endocrinology* 1994; 135:682–691.
21. Marsh JM. The role of cyclic AMP in gonadal function. In: Greengard P, Robinson GA (eds.), *Cyclic Nucleotide Research*. Vol. 6. New York: Raven Press; 1975: 137–199.
22. Davis JS. Stimulation of intracellular free Ca²⁺ by luteinizing hormone in isolated bovine luteal cells. *Adv Exp Med Biol* 1987; 219:671–675.
23. Davis RJ. The mitogen-activated protein kinase signal transduction pathway. *J Biol Chem* 1993; 268:14553–14556.
24. Fuchs A-R, Goeschen J, Rasmussen AB, Rehnström JV. Cervical ripening by endocervical and extra-amniotic PGE₂. *Prostaglandins* 1984; 28:217–227.
25. Ellwood DA, Mitchell MD, Anderson ABM, Turnbull AC. Specific changes in the in vitro production of prostanoids by the ovine cervix at parturition. *Prostaglandins* 1980; 39:675–684.

26. Hiller K, Coad N. Synthesis of prostaglandins by the human uterine cervix in vitro during passive mechanical stretch. *J Pharm Pharmacol* 1982; 34:262-263.
27. Stys SJ, Dresser BL, Ott TE, Clark KE. Effects of prostaglandin E₂ on cervical compliance in pregnant ewes. *Am J Obstet Gynecol* 1981; 140:415-419.
28. Ledger WL, Ellwood DA, Taylor MJ. Cervical softening in late pregnant sheep by infusion of prostaglandin E₂ into cervical artery. *J Reprod Fertil* 1983; 69:511-515.
29. Khalifa RME, Sayre BL, Lewis GS. Exogenous oxytocin dilates the cervix in the ewes. *J Anim Sci* 1992; 70:38-42.
30. Duchens M, Fredriksson G, Kindahl H, Aiumlamai S. Effect of intra-cervical administration of a prostaglandin E₂ gel in pregnant and non-pregnant heifers. *Vet Rec* 1993; 133:546-549.
31. Graddy L, Fields M, Kowalski A, Chang S-M, Fuchs A-R. Effects of oxytocin and PGE₁ on bovine cervix at estrus. *Biol Reprod* 1998; 58(suppl 1):174 (abstract 327).
32. Schams D. Oxytocin determination by radioimmunoassay. III. Improvement to subpicogram sensitivity and application to blood levels in cyclic cattle. *Acta Endocrinol* 1983; 103:180-183.
33. Ohtani M, Kobayashi S-I, Miyamoto A, Hayashi K, Yutaka F. Real-time relationships between intraluteal and plasma concentrations of endothelin, oxytocin and progesterone during prostaglandin F_{2α}-induced luteolysis in the cow. *Biol Reprod* 1998; 58:103-108.

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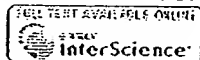
The alpha(v)beta6 integrin receptor for Foot-and-mouth disease virus is expressed constitutively on the epithelial cells targeted in cattle.

Monaghan P, Gold S, Simpson J, Zhang Z, Weinreb PH, Violette SM, Alexandersen S, Jackson T.

Institute for Animal Health, Pirbright Laboratory, Ash Road, Pirbright, Surrey GU24 0NF, UK.

Field strains of Foot-and-mouth disease virus (FMDV) use a number of alpha(v)-integrins as receptors to initiate infection on cultured cells, and integrins are believed to be the receptors used to target epithelial cells in animals. In this study, immunofluorescence confocal microscopy and real-time RT-PCR were used to investigate expression of two of the integrin receptors of FMDV, alpha(v)beta6 and alpha(v)beta3, within various epithelia targeted by this virus in cattle. These studies show that alpha(v)beta6 is expressed constitutively on the surfaces of epithelial cells at sites where infectious lesions occur during a natural infection, but not at sites where lesions are not normally formed. Expression of alpha(v)beta6 protein at these sites showed a good correlation with the relative abundance of beta6 mRNA. In contrast, alpha(v)beta3 protein was only detected at low levels on the vasculature and not on the epithelial cells of any of the tissues investigated. Together, these data suggest that in cattle, alpha(v)beta6, rather than alpha(v)beta3, serves as the major receptor that determines the tropism of FMDV for the epithelia normally targeted by this virus.

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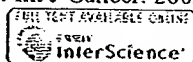
Urokinase-mediated posttranscriptional regulation of urokinase-receptor expression in non small cell lung carcinoma.

Montuori N, Mattiello A, Mancini A, Tagliatela P, Caputi M, Rossi G, Rago P.

Istituto di Endocrinologia ed Oncologia Sperimentale, Consiglio Nazionale delle Ricerche, Naples, Italy.

The urokinase-type plasminogen activator (uPA) and its cellular receptor (uPAR) are involved in the proteolytic cascade required for tumor cell dissemination and metastasis, and are highly expressed in many human tumors. We have recently reported that uPA, independently of its enzymatic activity, is able to increase the expression of its own receptor in uPAR-transfected kidney cells at a posttranscriptional level. In fact, uPA, upon binding uPAR, modulates the activity and/or the level of a mRNA-stabilizing factor that binds the coding region of uPAR-mRNA. We now investigate the relevance of uPA-mediated posttranscriptional regulation of uPAR expression in non small cell lung carcinoma (NSCLC), in which the up-regulation of uPAR expression is a prognostic marker. We show that uPA is able to increase uPAR expression, both at protein and mRNA levels, in primary cell cultures obtained from tumor and adjacent normal lung tissues of patients affected by NSCLC, thus suggesting that the enzyme can exert its effect in lung cells. We investigated the relationship among the levels of uPA, uPAR and uPAR-mRNA binding protein(s) in NSCLC. Lung tissue analysis of 35 NSCLC patients shows an increase of both uPA and uPAR in tumor tissues; as compared to adjacent normal tissues, in 27 patients (77%); 19 of these 27 patients also show a parallel increase of the level and/or binding activity of a cellular protein capable of binding the coding region of uPAR-mRNA. Therefore, in tumor tissues, a strong correlation is observed among these 3 parameters; uPA, uPAR and the level and/or the activity of a uPAR-mRNA binding protein. We then suggest that uPA regulates uPAR expression in NSCLC at a posttranscriptional level by increasing uPAR-stability through a cellular factor that binds the coding region of uPAR-mRNA. Copyright 2003 Wiley-Liss, Inc.

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Vascular endothelial growth factor expression correlates with matrix metalloproteinases MT1-MMP, MMP-2 and MMP-9 in human glioblastomas.

Munaut C, Noel A, Hougrand O, Foidart JM, Boniver J, Deprez M.

Laboratory of Tumour and Development Biology, University of Liege, Liege, Belgium.

Vascular endothelial growth factor (VEGF) is the major endothelial mitogen in central nervous system neoplasms and it is expressed in 64-95% of glioblastomas (GBMs). Tumour cells are the main source of VEGF in GBMs whereas VEGF receptors (VEGFR-1, its soluble form sVEGFR-1, VEGFR-2 and neuropilin-1) are expressed predominantly by endothelial cells. Infiltrating tumour cells and newly-formed capillaries progress through the extracellular matrix by local proteolysis involving matrix metalloproteinases (MMPs). Recent studies have shown that VEGF expression and bioavailability can be modulated by MMPs. We reported previously that the expression of MT1-MMP in human breast cancer cells was associated with an enhanced VEGF expression. We used quantitative RT-PCR, Western blot, gelatin zymography and immunohistochemistry to study the expression of VEGF, VEGFR-1, VEGFR-2, sVEGFR-1, neuropilin-1, MT1-MMP, MMP-2, MMP-9 and TIMP-2 in 20 human GBMs and 5 normal brains. The expression of these MMPs was markedly increased in most GBMs with excellent correlation between mRNA and protein levels; activated forms of MMP-2 and MMP-9 were present in 8/18 and 7/18 of GBMs. A majority of GBMs (17/20) also expressed high levels of VEGF, as previously reported, with strong correlation between VEGF and MT1-MMP gene expression levels, and double immunostaining showed that VEGF and MT1-MMP peptides co-localize in tumour and endothelial cells. Our results suggest that the interplay between metalloproteinases and VEGF previously described in experimental tumours may also be operative in human GBMs. Because of its dual ability to activate MMP-2 and to up-regulate VEGF, MT1-MMP might be of central importance in the growth of GBMs and represent an interesting target for anti-cancer treatments. Copyright 2003 Wiley-Liss, Inc.

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VASCULAR ENDOTHELIAL GROWTH FACTOR EXPRESSION CORRELATES WITH MATRIX METALLOPROTEINASES MT1-MMP, MMP-2 AND MMP-9 IN HUMAN GLIOBLASTOMAS

Carine MUNAUT¹, Agnès NOËL¹, Olivier HOUGRAND², Jean-Michel FOIDART¹, Jacques BONIVER² and Manuel DEPREZ^{2*}

¹Laboratory of Tumour and Development Biology, University of Liège, Liège, Belgium

²Laboratory of Neuropathology, University of Liège, Liège, Belgium

Vascular endothelial growth factor (VEGF) is the major endothelial mitogen in central nervous system neoplasms and it is expressed in 64–95% of glioblastomas (GBMs). Tumour cells are the main source of VEGF in GBMs whereas VEGF receptors (VEGFR-1, its soluble form sVEGFR-1, VEGFR-2 and neuropilin-1) are expressed predominantly by endothelial cells. Infiltrating tumour cells and newly-formed capillaries progress through the extracellular matrix by local proteolysis involving matrix metalloproteinases (MMPs). Recent studies have shown that VEGF expression and bioavailability can be modulated by MMPs. We reported previously that the expression of MT1-MMP in human breast cancer cells was associated with an enhanced VEGF expression. We used quantitative RT-PCR, Western blot, gelatin zymography and immunohistochemistry to study the expression of VEGF, VEGFR-1, VEGFR-2, sVEGFR-1, neuropilin-1, MT1-MMP, MMP-2, MMP-9 and TIMP-2 in 20 human GBMs and 5 normal brains. The expression of these MMPs was markedly increased in most GBMs with excellent correlation between mRNA and protein levels; activated forms of MMP-2 and MMP-9 were present in 8/18 and 7/18 of GBMs. A majority of GBMs (17/20) also expressed high levels of VEGF, as previously reported, with strong correlation between VEGF and MT1-MMP gene expression levels, and double immunostaining showed that VEGF and MT1-MMP peptides co-localize in tumour and endothelial cells. Our results suggest that the interplay between metalloproteinases and VEGF previously described in experimental tumours may also be operative in human GBMs. Because of its dual ability to activate MMP-2 and to up-regulate VEGF, MT1-MMP might be of central importance in the growth of GBMs and represent an interesting target for anti-cancer treatments.

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Key words: VEGF; MMPs; glioblastomas; brain; tumor

Angiogenesis is critical for the development of normal tissue and solid tumours. This process includes the degradation of the extracellular matrix (ECM) and the proliferation, migration and differentiation of endothelial cells, and is finely regulated by inhibitory and promoting factors.^{1,2} Among positive factors, vascular endothelial growth factor (VEGF) has been proposed as the major endothelial mitogen in central nervous system (CNS) neoplasms.³ Strong VEGF expression has been detected by immunohistochemistry in 64–95% of glioblastomas (GBMs).^{4–6} GBMs are the most common malignant primitive tumours of the CNS in adults.⁷ Microvascular proliferation is characteristic of these tumours and is an essential WHO diagnostic criteria.⁷ Tumour cells are the main source of VEGF in GBMs whereas VEGF receptors are predominantly expressed by endothelial cells.^{8,9} These receptors differ both in terms of affinity and transduction signaling.^{10–12} VEGFR-1 (flt-1) and VEGFR-2 (KDR) belong to the Class II tyrosine-kinase receptor family. VEGFR-1 has a soluble isoform (sVEGFR-1) that modulates VEGF availability.^{8,13} Neuropilin-1 (NRP1) is a co-receptor for VEGF that increases by 10-fold the affinity of the VEGF₁₆₅ isoform for VEGFR-2.¹⁴ NRP1 is thought to modulate VEGF-mediated tumour angiogenesis in human malignant astrocytomas.¹⁰ The coordinated up-regulation of VEGF and its receptors appears as a critical event in the control of angiogenesis.^{8,12}

Infiltrating tumour cells and newly-formed capillaries progress through the ECM by local proteolysis involving matrix metalloproteinases (MMPs).^{15,16} MMPs are proteolytic enzymes that are synthesized as inactive zymogens. Their activation requires the removal of a propeptide by proteinase cleavage and can be inhibited by various tissue inhibitors of MMPs (TIMPs). Most MMPs are secreted as soluble enzymes but a subset of them are inserted in the cell membrane by a transmembrane domain or by a glycosylphosphatidyl-inositol anchor and are classified as membrane-type MMPs (MT-MMPs).^{17,18} In cultured tumour cells, MT-MMPs tend to accumulate on the cytoplasmic membrane of invadopodia where they selectively mediate local pericellular proteolysis. GBMs express high levels of MT1-MMP, MMP-2 and MMP-9.^{19,20} Among these MMPs, MT1-MMP might play a central role in the remodeling of the ECM as this membrane-bound protease is able to activate MMP-2 and MMP-13.^{21,22} Moreover, MT1-MMP has been shown to promote cell migration in various carcinoma cell lines by its ability to cleave laminin-5, a major constituent of basement membrane,^{23,24} and through the processing of CD44H (the major receptor for hyaluronan)²⁵ and of $\alpha_5\beta_1$ integrin.²⁶

MT1-MMP is involved in both developmental and tumour angiogenesis.²⁷ MT1-MMP overexpression in human melanoma cells has been associated with enhanced *in vitro* invasion and increased *in vivo* tumour growth and vascularization.²⁸ We have shown previously that in MCF-7 breast cancer cells, VEGF transcription is upregulated when MT1-MMP is overexpressed.²⁹ Up-regulation of VEGF by MT1-MMP has also been reported in a model of human glioma xenograft by Deryugina *et al.*³⁰ These experimental data suggest a link between MT1-MMP and the VEGF network. We have tested for the presence of such a link in human glioblastomas. We compared the expression of VEGF and its receptors (VEGFR-1, sVEGFR-1, VEGFR-2, NRP1) with

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*Correspondence to: Laboratory of Neuropathology, Tower of Pathology B23/1, B-4000 Sart Tilman (Liège), Belgium. Fax: +32-4-3662919. E-mail: Manuel.Deprez@ulg.ac.be

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MT1-MMP, MMP-2, MMP-9 and TIMP-2 in a series of 20 GBMs and 5 normal brains. Using quantitative RT-PCR, gelatin zymography, Western blot and immunohistochemistry, we showed a strong correlation between the expression of VEGF, MT1-MMP, MMP-2 and MMP-9 in GBMs. These results are in accordance with previous *in vitro* studies and add to the evidence of an interplay between VEGF and MMPs in the progression of human GBMs.

MATERIAL AND METHODS

Patients

We studied 20 GBMs diagnosed at the Laboratory of Neuropathology-CHU Liège between 1997 and 2001. The series included 17 primary GBMs (*i.e.*, no previous history of lower grade diffuse astrocytoma) and 3 secondary GBMs (*i.e.*, previous history of lower grade diffuse astrocytoma). Clinical information on these 20 cases have been reported previously as part of a larger series.³¹ The gender ratio was 1/1, and the age at time of diagnosis ranged from 41–79 years (mean 56 years). Normal brain cortex and white matter were obtained from 5 patients with intractable epilepsy treated by partial temporal lobectomy. Histological examination of these specimens showed severe hippocampal sclerosis; frozen tissue was sampled from microscopically normal inferior temporal gyri. Our study was approved by the Ethical Committee of the Faculty of Medicine of the University of Liège.

RNA extraction and cDNA synthesis

Total RNA was extracted from cryosections with RNeasy Mini Kit (QIAGEN GmbH, Hilden, Germany) according to the manufacturer's protocol. Total RNA (1 µg) was reverse transcribed with a ThermoScript reverse transcriptase (ThermoScript RT-PCR System, Invitrogen, Carlsbad, CA) and random hexamers as primers.

Primers

Primers pairs used in our study are described in Table 1. Primers for the VEGF gene were chosen to distinguish between VEGF₁₈₉, VEGF₁₆₅, VEGF₁₄₅ and VEGF₁₂₁ mRNA isoforms. Intron-spanning primers and probes for the TaqMan system (primers for VEGFR-1 (Flt-1), sVEGFR-1, VEGFR-2 (KDR/Flk-1) and NRP1) were designed to meet specific criteria by using Primer Express software (Perkin Elmer, Foster City, CA). All primers were synthesized by Eurogentec (Liège, Belgium). The 5'- and 3'-end nucleotides of the probe were labeled with a reporter (FAM = 6-carboxy-fluorescein) and a quencher dye (TAMRA = 6-carboxy-tetramethylrhodamine). We conducted BLASTn (National Center for Biotechnology Information, Bethesda) searches against dbEST and the non redundant set of GenBank, EMBL, and DDBJ database sequences to confirm the total gene specificity of the nucleotide sequences chosen for the primers. The specificity of the amplified PCR products was confirmed either by restriction digest or by sequencing. The 18S ribosomal RNA was measured using the Pre-Developed TaqMan Assay Reagents Endogenous control kit from Applied Biosystems (Foster City, CA).

TABLE 1—SEQUENCE OF PRIMERS AND TaqMan PROBES USED FOR RT-PCR STUDIES

| Gene and accession number | Position | Sequence | Size | Cycles |
|---------------------------|----------|--|--------------------------------------|--------|
| MMP-2 FP | 1740F | 5'-AGATCTTCTCTCAAGGACCGGT-3' | 225 bp | 33 |
| MMP-2-RP NM_004530 | 1964R | 5'-GGCTGGTCACTGGCTTGGGGTA-3' | | |
| MMP-9-FP | 1592F | 5'-GCGGAGATTGGGAACCACTGTA-3' | 208 bp | 37 |
| MMP-9-RP J05070 | 1800R | 5'-GACCGCGCTGTGTACACCCACA-3' | | |
| MMP-14-FP | 1288F | 5'-GGATACCCAATGCCCATTTGGCCA-3' | 221 bp | 32 |
| MMP-14-RP NM_004995 | 1508R | 5'-CCATTGGGCATCCAGAAAGAGAGC-3' | | |
| TIMP1-FP | 78F | 5'-CATCCTGTGTGTGCTGTGGCTGAT-3' | 168 bp | 33 |
| TIMP1-RP M12670 | 245R | 5'-GTCATCTTGATCTCATAACGCTGG-3' | | |
| TIMP-2-FP | 78F | 5'-CTCGCTGGACGTTGGAGGAAAGAA-3' | 155 bp | 30 |
| TIMP-2-RP NM_003255 | 245R | 5'-AGCCCATCTGGTACCTGTGGTTCA-3' | | |
| VEGF-FP | 1208F | 5'-CCTGGTGGACATCTCCAGGAGTA-3' | 479 bp 407 bp 347 bp 275 bp | 33 |
| VEGF-RP AH001553 | 1687R | 5'-CTCACCGCTCGGCTTGTCACA-3' | | |
| 28S rRNA-RP | 12403F | 5'-GTTCACCCACTAATAGGGAACGTGA-3' | | |
| 28S rRNA-RP U13369 | 12614R | 5'-GATTCTGACTTAGAGGCGTTCACT-3' | | |
| VEGFR1-FP | 2438F | 5'-TCCCTTATGATGCCAGCAAGT-3' | 79 bp | 40 |
| VEGFR1-RP | 2516R | 5'-CCAAAAGCCCTCTTCCAA-3' | | |
| VEGFR1 Probe AF063657 | 2469 | 5'-CCGGGAGAGACTTAACTGGGCAAAATCA-3' | | |
| sVEGFR1-FP | 2209F | 5'-ACAATCAGAGGTGAGCACTGCAA-3' | 180 bp | 40 |
| sVEGFR1-RP | 2388R | 5'-TCCGAGCCTGAAAGTTAGCAA-3' | | |
| sVEGFR1 Probe U01134 | 2257 | 5'-TCCAAATTTAAAGCACAAGGAATGATTGTACCAC-3' | | |
| VEGFR2-FP | 791F | 5'-CTTCGAAGCATCAGCATAAAGAACT-3' | 156 bp | 40 |
| VEGFR2-RP | 946R | 5'-TGGTCATCAGCCCACTGGAT-3' | | |
| VEGFR2 Probe AF063658 | 820 | 5'-AACCGAGACCTAAAAACCCAGTCTGGGAGT-3' | | |
| NRP1-FP | 1831F | 5'-CACAGTGGAAACAGGTGATGACTTC-3' | 112 bp | 40 |
| NRP1-RP | 1942R | 5'-AACCATATGTTGGAACCTCTGATTGT-3' | | |
| NRP1 Probe XM_034725 | 1883 | 5'-CCACAGAAAAGCCACGGTCATAGACA-3' | | |

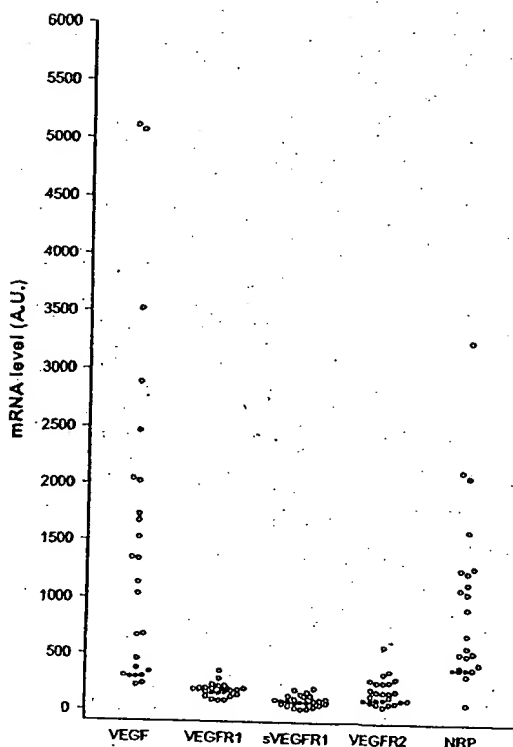


FIGURE 1 – VEGF and VEGF receptors mRNA quantification: scatter of the distribution. Normal brain (black spots) and GBMs (white spots) mRNA levels are expressed as normalized values (as described in Material and Methods: A.U. = arbitrary units). Each point represents the mean of 3 separate experiments.

End point quantitative PCR for MT1-MMP, MMP-2, MMP-9, TIMP-2 mRNA and VEGF mRNA isoforms

MT1-MMP, MMP-2, MMP-9, TIMP-2 and VEGF mRNA isoforms (VEGF₁₈₉, VEGF₁₆₅, VEGF₁₄₅ and VEGF₁₂₁) were measured in 10 ng aliquots of cDNA using Taq polymerase (Takara, Shiga, Japan) and 5 pmol of each primers (Table I). The thermal cycling conditions included 2 min at 95°C for denaturation and then amplification 15 sec at 94°C, 20 sec at 66°C and 20 sec at 72°C (30 sec for VEGF isoforms) with a final incubation 2 min at 72°C. PCR products were resolved on 2% Nusieve 3:1 agarose gels (BioWhittaker, Rockland, MD) and analyzed using a Fluor-S MultiImager (Bio-Rad, Hercules, CA) after ethidium bromide staining. Specific mRNA levels were expressed as the ratio of specific transcripts/28S transcripts. Experiments were repeated at least 3 times in duplicate.

Real-time quantitative PCR for VEGFR-1, sVEGFR-1, VEGFR-2 and NRP1 mRNA

Real-time quantitative RT-PCR analyses for VEGFR-1, sVEGFR-1, VEGFR-2, NRP1 mRNAs and 18S rRNA were carried out using the ABI PRISM 7700 Sequence Detection System instrument and software (PE Applied Biosystems). The sequences of the PCR primer pairs and fluorogenic probes that were used for each gene are shown in Table I. A standard curve was generated by 5-fold serial dilution of placenta cDNA to cover the range of 50,000–80 ng and was run in duplicate during every experiment. For each experimental sample, the amount of target gene was determined from this standard curve. The relative expression level of the target gene was normalized against 18S rRNA to compen-

sate for variation in the quality of RNA and the amount of input cDNA (as described by the manufacturer PE Applied Biosystems in User Bulletin 2). PCR was carried out with the TaqMan Universal PCR Master Mix (Applied Biosystems) using 5 µl of diluted cDNA (equivalent to 10 ng total RNA), 200 nM of the probe, and 400 nM primers in a 25 µl final reaction mixture. After a 2 min incubation at 50°C to allow for UNG cleavage, AmpliTaq Gold was activated by an incubation for 10 min at 95°C. Each of the 40 PCR cycles consisted of 15 sec of denaturation at 95°C and hybridization of probe and primers for 1 min at 60°C.

To confirm amplification specificity, the PCR products were also examined by subsequent 2% agarose gel electrophoresis. Experiments were repeated at least 3 times in duplicate.

Immunohistochemistry for VEGF and MT1-MMP

Sections (4 µm thick) were cut from formalin-fixed, paraffin embedded tumour tissue. They were hydrated through graded alcohols and incubated in H₂O₂ (0.3% 15 min). Sections were autoclaved for 11 min at 126°C in citrate buffer pH6 for antigen retrieval (Dako, Glostrup, Denmark). For double immunostaining sections were incubated in primary monoclonal Ab anti-MT1-MMP (Ab-4) 1:100 (Oncogene Research Products, San Diego, CA) followed by peroxidase-conjugated EnVision (Dako). Immunoreactivity was visualized with 3,3'-diaminobenzidine (DAB+, Dako). Sections were then incubated with polyclonal Ab anti-VEGF 1:150 (Santa Cruz, Santa Cruz, CA) for 1 hr at room temperature, followed by alkaline phosphatase-conjugated EnVision (Dako). Immunoreactivity for VEGF was visualized with Fast Red chromogenic substrate (Dako). Single immunostaining was also carried out on serial sections using each primary antibody alone with the corresponding enzyme-chromogene combination. Negative controls were obtained by omitting the primary antibodies.

Gelatin zymography assay

MMP-2 and MMP-9 activities were quantified by gelatin zymography on 2 normal brains and 18 GBMs. Ten cryosections (10 µm) were homogenized in buffer (0.1 M Tris-HCl pH 8.1, 0.4% Triton X-100) and centrifuged for 20 min at 5,000g. The pellets were discarded. 25 µg of total protein from homogenate supernatants were mixed with non reducing sample buffer (62.5 mM Tris-HCl, pH 6.8; 2% SDS; 10% glycerol; 0.1% bromophenol blue) and electrophoresed directly on 10% SDS-polyacrylamide gels (SDS-PAGE) containing 0.1% gelatin (w/v).³² After electrophoresis, gels were washed for 1 hr at room temperature in a 2% (v/v) Triton X-100 solution to remove SDS, transferred to a buffer (50 mM Tris-HCl, pH 7.6, containing 10 mM CaCl₂) and incubated for 18 hr at 37°C. Gels were stained for 30 min with 0.1% (w/v) Coomassie brilliant blue G250 in 45% (v/v) methanol/10% (v/v) acetic acid and destained in 10% (v/v) acetic acid/20% (v/v) methanol. Gels were analyzed with Quantity One software (version 4.2.2, Bio-Rad Laboratories, Hercules, CA) after densitometric scanning of the gels using a Fluor-S MultiImager (BioRad).

Western blot

MT1-MMP protein levels were analyzed in 2 normal brains and 15 GBMs. Brain extracts (25 µg) were mixed with 1/2 sample buffer [0.25 M Tris (pH6.8), 10% SDS (w/v), 4% sucrose (v/v), 5% β-mercaptoethanol (v/v) and 0.125% bromophenol blue (w/v)] and boiled for 5 min. They were separated on 10% SDS-PAGE gels and transferred to a PVDF filter (NEN, Boston, MA). After blocking with 5% milk (w/v), 0.1% tween 20 (w/v) in PBS for 2 hr at room temperature, membranes were exposed to the primary antibody (10 µg/ml, clone 113-5B7, Ab-4, Oncogene Research Products, San Diego, CA) at 4°C overnight followed by incubation with a horseradish peroxidase-conjugated rabbit anti-mouse antibody (1.3 µg/ml, Dako, Glostrup, Denmark). Signals were detected with an enhanced chemoluminescence (ECL) kit (NEN, Boston, MA). The relative intensities of the immunoreactive bands were analyzed with Quantity One software (version 4.2.2, Bio-Rad).

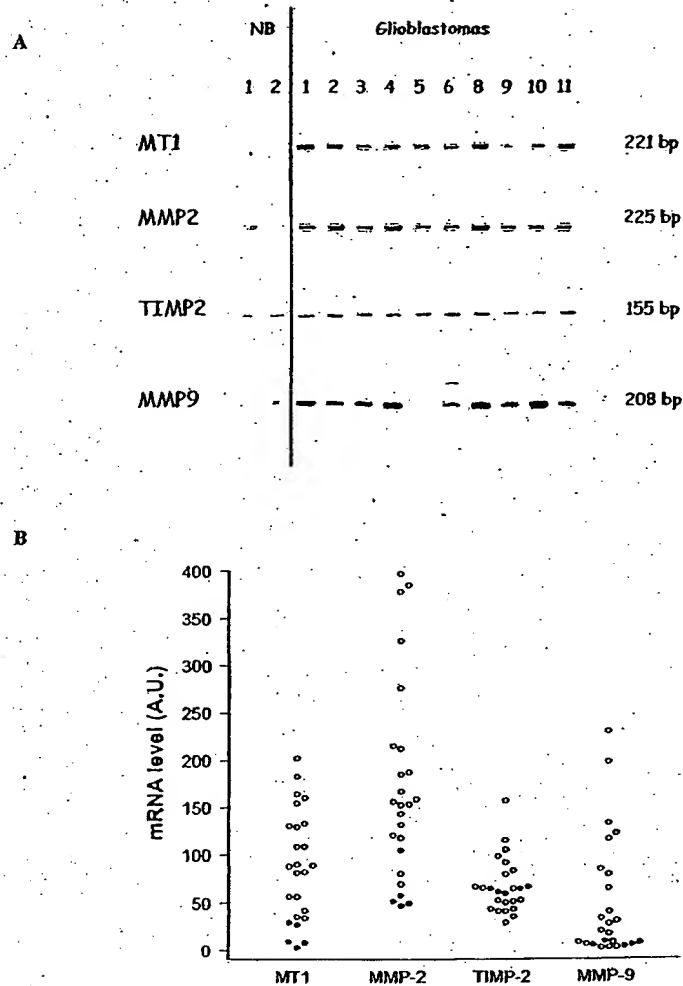


FIGURE 2—MMPs and TIMP-2 mRNA quantification. (a) Representative 2% agarose gels of RT-PCR products for MT1-MMP, MMP-2, TIMP-2, and MMP-9 in 2 normal brains (NB) and 10 GBMs. (b) Scatter plots (as described in Fig. 1). Experiment was repeated at least 3 times in duplicate.

Laboratories) after densitometric scanning of the X-ray films using a Fluor-S Multimager (Bio-Rad).

Statistics

VEGF, VEGFRs, MMPs and TIMP-2 expression values in GBMs were correlated using Spearman's test. Correlation was considered significant for 2-tailed p -value < 0.05 . Statistical analysis was carried out using the Prism 3.0 software (GraphPad, San Diego, CA).

RESULTS

Expression of VEGF and VEGF receptors

VEGF mRNA was present in normal brains (295–375, arbitrary units; mean = 322) and in all GBM samples (217–5,112; mean = 1,774) as reported previously (Fig. 1).³¹ In most GBMs, VEGF mRNA levels were raised 2–15-fold above normal brain values. The most abundant isoform in all cases was VEGF₁₆₅, followed by VEGF₁₂₁, VEGF₁₈₉ and VEGF₁₄₅ (data not shown). VEGFR-1 expression was found at similar levels in GBMs (89–357; mean = 182) and normal controls (154–198; mean = 181). There was no correlation between VEGFR-1 and VEGF mRNA levels ($p = 0.35$) in GBMs. VEGFR-2 was expressed in all GBMs (48–582;

mean = 210) and in 8/20 cases at least twice normal values (87–111; mean = 103). VEGF and VEGFR-2 expressions were correlated significantly ($p = 0.0035$) in GBMs. NRP1 expression varied broadly between GBMs (75–3,260; mean = 1,061) contrasting with a constant baseline expression in normal controls (383–397; mean = 390). In tumours, NRP1 correlated with VEGFR-2 ($p = 0.0119$) but not with VEGF ($p = 0.084$), nor VEGFR-1 ($p = 0.066$). sVEGFR-1 was expressed at low levels both in normal brains (84–92; mean = 87) and GBMs (25–208; mean = 101). sVEGFR-1, however, was found to correlate with VEGFR-1 ($p = 0.0289$), VEGFR-2 ($p = 0.0029$), and NRP1 ($p = 0.0027$) but not with VEGF ($p = 0.053$).

Expression of MMPs and TIMP-2

MT1-MMP, MMP-2 and MMP-9 were expressed in both normal brains and GBMs but at much higher levels in the latter (Fig. 2a). MT1-MMP mRNA levels were constantly higher in GBMs (34–202; mean = 106) than in normal controls (3–29; mean = 15). MMP-2 and MMP-9 mRNA levels were higher than controls in 18/20 and 14/20 cases respectively (Fig. 2b) and correlated with each other ($p = 0.0187$). MT1-MMP mRNA levels correlated with MMP-2 ($p = 0.0008$) and MMP-9 ($p = 0.005$). TIMP-2 had a non-discriminative distribution in relation to the controls. TIMP-2

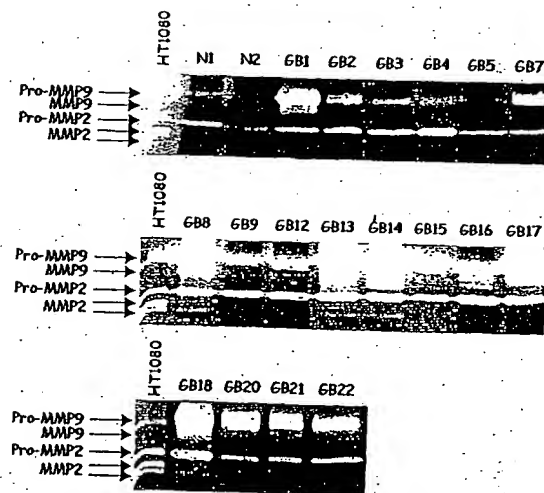


FIGURE 3—Zymographic analysis of MMP-2 and MMP-9 in tissue extracts either from normal brain (N) or GBMs (GB). Medium conditioned by human HT1080 cells was included as positive control. Positions of pro-MMP-9, MMP-9, pro-MMP-2 and MMP-2 are indicated by arrows.

TABLE II—ZYMOGRAMS AND WESTERN BLOT QUANTIFICATION OF MMPs IN NORMAL BRAIN AND GBMs¹

| | Gelatin zymography | | | | Western blot MT1-MMP |
|------|--------------------|-------|----------|-------|-------------------------|
| | proMMP-9 | MMP-9 | proMMP-2 | MMP-2 | |
| N1 | 0.41 | 0.00 | 1.52 | 0.00 | 0.44 |
| N2 | 0.28 | 0.00 | 0.83 | 0.00 | 0.47 |
| GB1 | 9.00 | 1.25 | 2.45 | 0.35 | 1.58 |
| GB2 | 4.28 | 0.00 | 2.88 | 0.09 | 0.47 |
| GB3 | 1.67 | 0.00 | 2.86 | 0.00 | 0 |
| GB4 | 1.40 | 0.00 | 4.33 | 0.03 | 0 |
| GB5 | 0.47 | 0.00 | 1.15 | 0.00 | ND |
| GB7 | 5.29 | 0.00 | 1.12 | 0.00 | 0 |
| GB8 | 5.75 | 3.78 | 4.63 | 0.65 | 0.55 |
| GB9 | 5.02 | 0.00 | 4.09 | 0.00 | 0.45 |
| GB12 | 5.08 | 0.00 | 4.79 | 0.20 | 3.18 |
| GB13 | 6.86 | 4.58 | 4.93 | 0.48 | 1.55 |
| GB14 | 5.59 | 6.75 | 5.53 | 0.76 | 0.94 |
| GB15 | 8.00 | 0.00 | 5.13 | 0.00 | ND |
| GB16 | 5.55 | 1.10 | 5.66 | 0.00 | 2.50 |
| GB17 | 6.61 | 2.04 | 6.00 | 0.00 | 0 |
| GB18 | 14.79 | 4.34 | 8.20 | 0.40 | 1.97 |
| GB20 | 18.85 | 0.00 | 2.12 | 0.00 | 0 |
| GB21 | 16.52 | 0.00 | 2.37 | 0.00 | ND |
| GB22 | 20.66 | 0.00 | 5.11 | 0.00 | 0.86 |

¹Extracts expressed as arbitrary units. N, normal brain; GB, glioblastoma; ND, not determined.

was correlated with MT1-MMP ($p = 0.0019$) and MMP-2 ($p = 0.0002$) but not with MMP-9 ($p = 0.1408$).

Correlation between MT1-MMP protein and activated MMP-2 and -MMP-9

By gelatin zymography, pro-MMP-2 and pro-MMP-9 were detected in the 18 GBMs and 2 controls examined (Fig. 3, Table II). In most GBMs, levels of these inactive forms were higher than in normal brains and were correlated with their respective mRNA levels (MMP-2: $p < 0.0001$; MMP-9: $p = 0.01$). Activated forms of MMP-2 and MMP-9 were not found in normal brain. By contrast, they were present in 8/18 (MMP-2) and 7/18 (MMP-9) GBMs. MT1-MMP protein levels were quantified by Western blot in 15 GBMs and 2 normal brains (Fig. 4, Table II). They were signifi-

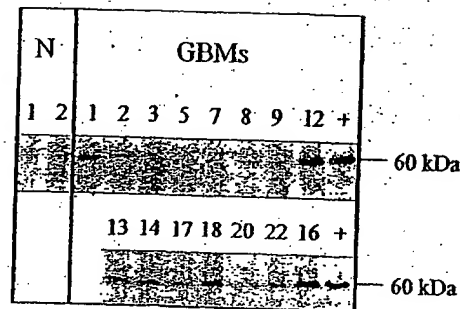


FIGURE 4—Western blot analysis using the ab4 antibody (clone 113-5B7) raised against the catalytic domain of MT1-MMP. Protein extracts from MT1-MMP transfected A2058 cells (clone SL5²⁸) were used as a positive control (+). MT1-MMP protein is detected in normal brain (N) and GBMs.

TABLE III—CORRELATION BETWEEN VEGF, VEGF RECEPTORS, MMPs AND TIMP-2 EXPRESSED AS p VALUES DERIVED FROM SPEARMAN'S TEST

| | MT1-MMP | MMP-2 | MMP-9 | TIMP-2 |
|----------|---------|--------|---------|--------|
| VEGF | 0.0250 | 0.0245 | 0.0053 | 0.0094 |
| VEGFR-1 | 0.0073 | 0.0710 | <0.0001 | 0.4542 |
| VEGFR-2 | <0.0001 | 0.0168 | 0.0004 | 0.0153 |
| NRP1 | 0.0053 | 0.1334 | 0.2457 | 0.5480 |
| sVEGFR-1 | 0.0313 | 0.0469 | 0.0194 | 0.2563 |

cantly correlated with zymogram-derived activated MMP-2 levels ($p = 0.0226$) but not with activated MMP-9 levels ($p = 0.06$). Interestingly, MT1-MMP protein and mRNA levels were correlated significantly ($p = 0.089$), arguing for a predominantly transcriptional regulation in GBMs.

Correlation between VEGF network and MMPs

mRNA levels of VEGF and VEGF receptors were compared to MT1-MMP, MMP-2, MMP-9 and TIMP-2 (Table III). There was a significant correlation between VEGF expression and MT1-MMP, MMP-2 and MMP-9. A similar correlation was also observed between VEGFR-2 and MMPs. Interestingly, TIMP-2 expression was correlated with VEGF and VEGFR-2 but not with other VEGF receptors.

Immunohistochemistry for VEGF and MT1-MMP

VEGF immunoreactivity was shown in both tumour and endothelial cells, as previously reported (Fig. 5a,b).³¹ By single immunostaining, MT1-MMP was detected in glioblastoma cells as a diffuse cytoplasmic staining (Fig. 5c,d). MT1-MMP positivity was also seen in endothelial cells and perivascular cells (Fig. 5c). By double immunostaining, we observed the co-localization of VEGF and MT1-MMP in the cytoplasm of numerous tumour cells (Fig. 5e,f).

DISCUSSION

GBMs are highly malignant tumours with poor prognosis. They show major microvascular proliferation and express high levels of VEGF.⁴⁻⁹ VEGF is a strong mitogen for endothelial cells thereby promoting angiogenesis. Previous reports^{32,34} have suggested that VEGF also stimulates tumour cell invasion, migration and survival in malignant epithelial cells through an autocrine loop by which overexpression of MMPs induces VEGF secretion and leads to subsequent amplification of cell proliferation and protection against apoptosis. We and others reported previously that in human melanoma and breast carcinoma cells, MT1-MMP upregulates VEGF expression whereas TIMP-2 reduces it.^{28,29,35} Therefore the

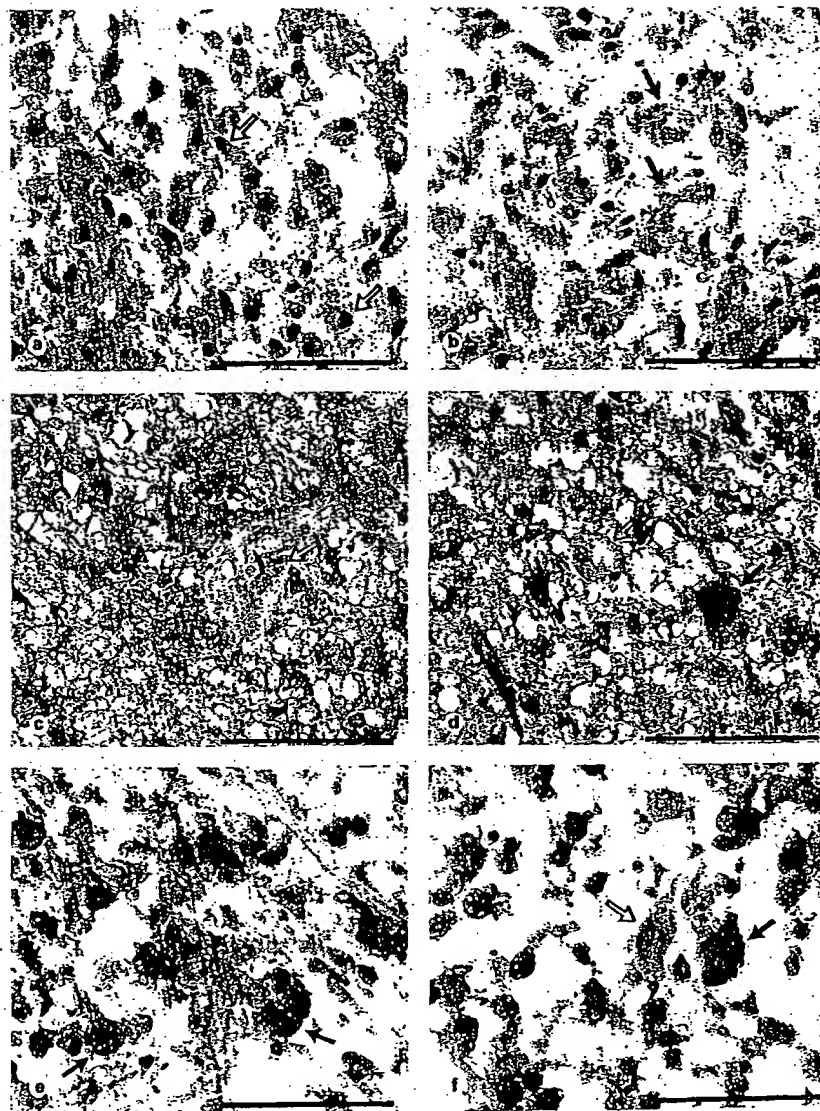


FIGURE 5—Immunohistochemistry (scale bar = 50 µm). (a,b) VEGF positive tumour and endothelial cells (plain arrows) show granular red staining of the cytoplasm. Negative cells (empty arrow) are seen in their close vicinity. (c) MT1-MMP positive cells show a strong brown cytoplasmic staining. They include tumour cells (plain arrow) and endothelial cells (empty arrow). (d) MT1-MMP positive tumour cells (plain arrow) are mixed with negative cells (empty arrow). (e,f) Double staining with VEGF (red) and MT1-MMP (brown). Double positive tumour cells (plain arrows) contrast with negative or single weakly positive cell (empty arrow).

pericellular proteolysis mediated by MT1-MMP in GBMs could also induce an autocrine loop resulting in enhanced VEGF expression. In turn, VEGF could act as a paracrine factor on endothelial cells to stimulate angiogenesis or possibly as an autocrine factor promoting glioblastoma cells survival-migration and invasion as demonstrated recently in the various tumour cell culture models.

We compared the expression of VEGF and its receptors with MT1-MMP, MMP-2 and MMP-9 in 20 GBMs and 5 normal brains. The expression of these MMPs was markedly increased in most GBMs with excellent correlation between mRNA and protein levels. MT1-MMP expression has been shown previously to correlate with glioma aggressiveness and its transfection in different tumour cell lines triggers an angiogenic phenotype and promotes tumour growth.^{20,28–30,36–38} A majority of GBMs (17/20) also expressed high levels of VEGF, as previously reported, with a strong correlation between VEGF and MT1-MMP gene expression levels. Double immunostaining studies showed co-expression of VEGF and MT1-MMP by the same tumour cells. These data suggest that the transcrip-

tional control of VEGF by MT1-MMP could be operative not only *in vitro* but also *in vivo* in human GBMs.

MT1-MMP could also promote the growth of GBMs by its ability to activate MMP-2 in the presence of low concentration of TIMP-2.⁴⁰ Pro-MMP-2 activation occurs after the formation of a ternary complex that contains pro-MMP-2 linked to cell surface MT1-MMP via a TIMP-2 bridge. In accordance with this hypothesis, we found that MMP-2 activation occurred in 8/18 of our GBMs^{20,41} among which 7/7 tested for MT1-MMP showed high contents of this protease.

Activated MMP-9 was also found in 7/18 of our GBMs. This is an interesting finding as active MMP-9 is able to mobilize VEGF from its ECM reservoir.³⁹ Therefore, MMPs could promote VEGF-mediated angiogenesis in GBMs by both transcriptional (MT1-MMP) and post translational (MMP-9) mechanisms.

VEGF binding to VEGFR-2 triggers the proliferation and migration of endothelial cells whereas its binding to VEGFR-1 has

opposite effects on glioblastoma cell lines.^{12,42} In our study, VEGF mRNA levels were correlated with VEGFR-2 but not VEGFR-1, NRP1 and sVEGFR-1. Collectively our data suggest that GBMs display a specific and complex pattern of VEGF receptors, transducing VEGF signaling toward cell proliferation and migration.

In conclusion, our study adds to the evidence for an interplay between metalloproteinases and VEGF in human GBMs as previously documented in experimental tumours. Because of its dual ability to activate MMP-2 and to up-regulate VEGF, MT1-MMP might be of central importance in the growth of human glioblastomas and represent an interesting target for anti-cancer treatments.

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REFERENCES

- Folkman J, Klagsbrun M. Angiogenic factors. *Science* 1987;235:442-7.
- Carmeliet P, Jain RK. Angiogenesis in cancer and other diseases. *Nature* 2000;407:249-57.
- Plate KH, Breier G, Weich HA, Risau W. Vascular endothelial growth factor is a potential tumour angiogenesis factor in human gliomas in vivo. *Nature* 1992;359:845-8.
- Oehring RD, Miletic M, Valter MM, Pietsch T, Neumann J, Fimmers R, Schlegel U. Vascular endothelial growth factor (VEGF) in astrocytic gliomas—a prognostic factor? *J Neurooncol* 1999;45:117-25.
- Pietsch T, Valter MM, Wolf HK, von Deimling A, Huang HJ, Cavenee WK, Wiestler OD. Expression and distribution of vascular endothelial growth factor protein in human brain tumors. *Acta Neuropathol (Berl)* 1997;93:109-17.
- Nishikawa R, Cheng SY, Nagashima R, Huang HJ, Cavenee WK, Matsutani M. Expression of vascular endothelial growth factor in human brain tumors. *Acta Neuropathol (Berl)* 1998;96:453-62.
- Kleihues P, Cavenee WK. Pathology and genetics of tumours of the nervous system. Lyon: IARC Press, 2000. 29-39.
- Plate KH, Breier G, Weich HA, Mennel HD, Risau W. Vascular endothelial growth factor and glioma angiogenesis: coordinate induction of VEGF receptors, distribution of VEGF protein and possible in vivo regulatory mechanisms. *Int J Cancer* 1994;59:520-9.
- Machein MR, Plate KH. VEGF in brain tumors. *J Neurooncol* 2000;50:109-20.
- Ding H, Wu X, Roicari L, Lau N, Shannon P, Nagy A, Guha A. Expression and regulation of neuropilin-1 in human astrocytomas. *Int J Cancer* 2000;88:584-92.
- Kunkel P, Ulbricht U, Bohlen P, Brockmann MA, Fillbrandt R, Stavrou D, Westphal M, Lamszus K. Inhibition of glioma angiogenesis and growth in vivo by systemic treatment with a monoclonal antibody against vascular endothelial growth factor receptor-2. *Cancer Res* 2001;61:6624-8.
- Herold-Mende C, Steiner HH, Andl T, Riede D, Buttler A, Reisser C, Fussenig NE, Mueller MM. Expression and functional significance of vascular endothelial growth factor receptors in human tumor cells. *Lab Invest* 1999;79:1573-82.
- Hiratsuka S, Minowa O, Kuno J, Noda T, Shibuya M. Flt-1 lacking the tyrosine kinase domain is sufficient for normal development and angiogenesis in mice. *Proc Natl Acad Sci USA* 1998;95:9349-54.
- Soker S, Takashima S, Miao HQ, Neufeld G, Klagsbrun M. Neuropilin-1 is expressed by endothelial and tumor cells as an isoform-specific receptor for vascular endothelial growth factor. *Cell* 1998;92:735-45.
- Egeblad M, Werb Z. New functions for the matrix metalloproteinases in cancer progression. *Nat Rev Cancer* 2002;2:161-74.
- Pepper MS. Role of the matrix metalloproteinase and plasminogen activator-plasmin systems in angiogenesis. *Arterioscler Thromb Vasc Biol* 2001;21:1104-17.
- Sternlicht MD, Werb Z. How matrix metalloproteinases regulate cell behavior. *Annu Rev Cell Dev Biol* 2001;17:463-516.
- Sounni NE, Janssen M, Foidart JM, Noel A. Membrane Type-1 Matrix Metalloproteinase and TIMP2 in tumor angiogenesis. *Matrix Biol* 2003;22:55-61.
- Behen AT, Paganelli PA, Schwab ME. Membrane-type 1 matrix metalloproteinase (MT1-MMP) enables invasive migration of glioma cells in central nervous system white matter. *J Cell Biol* 1999;144:373-84.
- Lampert K, Machein U, Machein MR, Conca W, Peter HH, Volk B. Expression of matrix metalloproteinases and their tissue inhibitors in human brain tumors. *Am J Pathol* 1998;153:429-37.
- Sato H, Takino T, Okada Y, Cao J, Shinagawa A, Yamamoto E, Seiki M. A matrix metalloproteinase expressed on the surface of invasive tumour cells. *Nature* 1994;370:61-5.
- Knauper V, Will H, Lopez-Otin C, Smith B, Atkinson SJ, Stanton H, Hemby RM, Murphy G. Cellular mechanisms for human procollagenase-3 (MMP-13) activation. Evidence that MT1-MMP (MMP-14) and gelatinase a (MMP-2) are able to generate active enzyme. *J Biol Chem* 1996;271:17124-31.
- Koshikawa N, Giannelli G, Cirulli V, Miyazaki K, Quaranta V. Role of cell surface metalloproteinase MT1-MMP in epithelial cell migration over laminin-5. *J Cell Biol* 2000;148:615-24.
- Gilles C, Polette M, Coraux C, Tournier JM, Meneguzzi G, Munaut C, Volders L, Rousselle P, Birembaut P, Foidart JM. Contribution of MT1-MMP and of human laminin-5 gamma2 chain degradation to mammary epithelial cell migration. *J Cell Sci* 2001;114:2967-76.
- Kajita M, Itoh Y, Chiba T, Mori H, Okada A, Kinoh H, Seiki M. Membrane-type 1 matrix metalloproteinase cleaves CD44 and promotes cell migration. *J Cell Biol* 2001;153:893-904.
- Deryugina EI, Ratnikov B, Monosov E, Postnova TI, DiScipio R, Smith JW, Strongin AY. MT1-MMP initiates activation of pro-MMP-2 and integrin $\alpha v \beta 3$ promotes maturation of MMP-2 in breast carcinoma cells. *Exp Cell Res* 2001;263:209-23.
- Zhou Z, Apte SS, Soiminen R, Cao R, Baaklini GY, Rausser RW, Wang J, Cao Y, Tryggvason K. Impaired endochondral ossification and angiogenesis in mice deficient in membrane-type matrix metalloproteinase 1. *Proc Natl Acad Sci USA* 2000;97:4052-7.
- Sounni NE, Baramova EN, Munaut C, Maquoi E, Frankenne F, Foidart JM, Noel A. Expression of membrane type 1 matrix metalloproteinase (MT1-MMP) in A2058 melanoma cells is associated with MMP-2 activation and increased tumor growth and vascularization. *Int J Cancer* 2002;98:23-8.
- Sounni NE, Devy L, Hajitou A, Frankenne F, Munaut C, Gilles C, Deroanne C, Thompson EW, Foidart JM, Noel A. MT1-MMP expression promotes tumor growth and angiogenesis through an upregulation of vascular endothelial growth factor expression. *FASEB J* 2002;16:555-64.
- Deryugina EI, Sorocanu L, Strongin AY. Upregulation of vascular endothelial growth factor by membrane-type 1 matrix metalloproteinase stimulates human glioma xenograft growth and angiogenesis. *Cancer Res* 2002;62:580-8.
- Munaut C, Boniver J, Foidart JM, Deprez M. Macrophage migration inhibitory factor (MIF) expression in human glioblastomas correlates with VEGF expression. *Neuropathol Appl Neurobiol* 2002;28:452-60.
- Munaut C, Noel A, Weidle UH, Krell HW, Foidart JM. Modulation of the expression of interstitial and type-IV collagenases in coculture of HT1080 fibrosarcoma cells and fibroblasts. *Invasion Metastasis* 1995;15:169-78.
- Bachelder RE, Crago A, Chung J, Wendt MA, Shaw LM, Robinson G, Mercurio AM. Vascular endothelial growth factor is an autocrine survival factor for neuropilin-expressing breast carcinoma cells. *Cancer Res* 2001;61:5736-40.
- Chung J, Bachelder RE, Lipscomb EA, Shaw LM, Mercurio AM. Integrin ($\alpha 6 \beta 4$) regulation of eIF-4E activity and VEGF translation: a survival mechanism for carcinoma cells. *J Cell Biol* 2002;158:165-74.
- Hajitou A, Sounni NE, Devy L, Grignet-Debrus C, Lewalle JM, Li H, Deroanne CF, Lu H, Colige A, Nussens BV, Frankenne F, Maron A.

- et al. Downregulation of vascular endothelial growth factor by tissue inhibitor of metalloproteinase-2: effect on in vivo mammary tumor growth and angiogenesis. *Cancer Res* 2001;61:3450-7.
36. Forsyth PA, Laing TD, Gibson AW, Rewcastle NB, Brasher P, Sutherland G, Johnston RN, Edwards DR. High levels of gelatinase-B and active gelatinase-A in metastatic glioblastoma. *J Neurooncol* 1998;36:21-9.
37. Nakada M, Nakamura H, Ikeda E, Fujimoto N, Yamashita J, Sato H, Seiki M, Okada Y. Expression and tissue localization of membrane-type 1, 2, and 3 matrix metalloproteinases in human astrocytic tumors. *Am J Pathol* 1999;154:417-28.
38. Kachra Z, Beaulieu E, Delbecchi L, Mousseau N, Berthelet F, Moun-djian R, Del Maestro R, Beliveau R. Expression of matrix metallo-proteinases and their inhibitors in human brain tumors. *Clin Exp Metastasis* 1999;17:555-66.
39. Bergers G, Brekken R, McMahon G, Vu TH, Itoh T, Tamaki K, Tanzawa K, Thorpe P, Itohara S, Werb Z, Hanahan D. Matrix metalloproteinase-9 triggers the angiogenic switch during carcinogenesis. *Nat Cell Biol* 2000;2:737-44.
40. Strongin AY, Collier I, Bamikov G, Mariner BL, Grant GA, Goldberg GL. Mechanism of cell surface activation of 72-kDa type IV collagenase. Isolation of the activated form of the membrane metalloproteinase. *J Biol Chem* 1995;270:5331-8.
41. Nakada M, Kita D, Futami K, Yamashita J, Fujimoto N, Sato H, Okada Y. Roles of membrane type 1 matrix metalloproteinase and tissue inhibitor of metalloproteinases 2 in invasion and dissemination of human malignant glioma. *J Neurosurg* 2001;94:464-73.
42. Millauer B, Witzmann-Voos S, Schumacher H, Martinez R, Moller NP, Risau W, Ullrich A. High affinity VEGF binding and developmental expression suggest Flk-1 as a major regulator of vasculogenesis and angiogenesis. *Cell* 1993;72:835-46.

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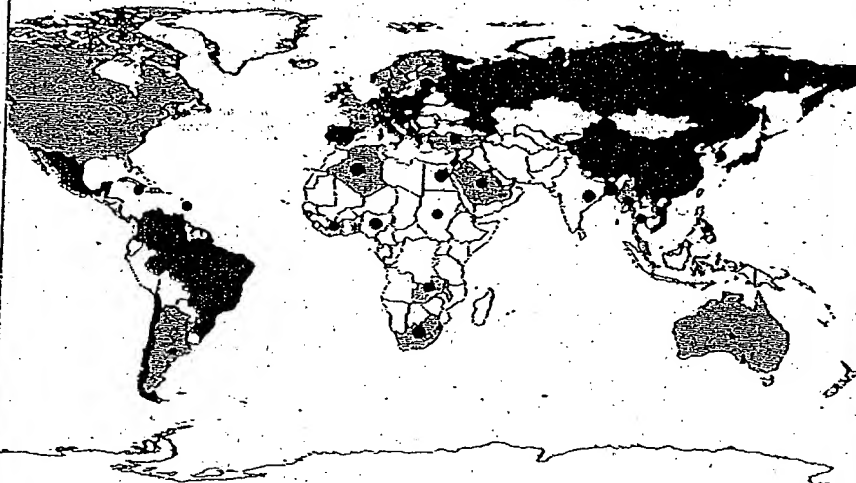
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Helicobacter pylori and gastric cancer: facing the enigmas



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Prevalence of *H. pylori*
(%)

- 1st quartile (19-39.9)
- 2nd quartile (39.9-67.5)
- 3rd quartile (67.5-81.2)
- 4th quartile (81.2-96.3)

Gastric cancer incidence*
(100 000)

- ≤ 5
- ▨ 5-10
- ▩ 10-15
- 15-20
- > 20

* Age and sex-standardized



International Union Against Cancer

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Founded in 1933, the UICC is a non-governmental, independent association of more than 290 member organizations in over 80 countries. Members are voluntary cancer leagues and societies, cancer research and/or treatment centers, and in some countries ministries of health.

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The UICC organizes an International Cancer Congress every 4 years as well as annual symposia, workshops, and training courses. It publishes the *International Journal of Cancer* (30 issues per year), *UICC News* (quarterly), the *International Calendar of Meetings on Cancer* (bi-annually), and a number of technical reports, textbooks and manuals.

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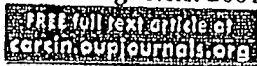
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DNA hypermethylation is a mechanism for loss of expression of the HLA class I genes in human esophageal squamous cell carcinomas.

Nie Y, Yang G, Song Y, Zhao X, So C, Liao J, Wang LD, Yang CS.

Laboratory for Cancer Research, College of Pharmacy, Rutgers-The State University of New Jersey, 164 Frelinghuysen Road, Piscataway, NJ 08854-8020, USA.

The three human leukocyte antigen (HLA) class I antigens, HLA-A, HLA-B and HLA-C, play important roles in the elimination of transformed cells by cytotoxic T cells. Frequent loss of expression of these antigens at the cell surface has been observed in many human cancers. Various mechanisms for post-transcriptional regulation have been proposed and tested but the molecular mechanisms for transcriptional regulation are not clear. We show by immunohistochemistry that the HLA class I antigens are absent in 26 of 29 (89%) samples of human esophageal squamous cell carcinomas (ESCC). Eleven of the 26 ESCC samples lost mRNA expression for at least one of the HLA genes, as shown by RT-PCR. DNA from the 29 pairs of ESCC and neighboring normal epithelium were examined for CpG island hypermethylation, homozygous deletion, microsatellite instability (MSI) and loss of heterozygosity (LOH). DNA from normal epithelial tissues had no detectable methylation of the CpG islands of any of these gene loci. Thirteen of 29 ESCC samples (45%) exhibited methylation of one or more of the three HLA loci and six samples (21%) exhibited methylation of all three loci. The HLA-B gene locus was most frequently methylated (38%). HLA-B mRNA expression in an ESCC cell line, where HLA-B was hypermethylated and did not express mRNA, was activated after treatment with 5-aza-2'-deoxycytidine. Homozygous deletion of these three gene loci was not observed. Relatively low rates of LOH and MSI were observed for the microsatellite markers D6S306, D6S258, D6S273 and D6S1666, close to the HLA-A, -B and -C loci, although a high ratio of LOH was observed at a nearby locus (represented by the markers D6S1051 and D6S1560), where the tumor suppressor gene p21(Waf1) resides. A strong correlation between genetic alterations and mRNA inactivation was observed in the ESCC samples. Our results indicate that HLA class I gene expression was frequently down-regulated in ESCC at both the protein and mRNA levels and that hypermethylation of the promoter regions of the HLA-A, -B and -C genes is a major mechanism of transcriptional inactivation.

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Comment in:

- [Hum Pathol. 2003 Jul;34\(7\):635-8.](#)

Human Pathology

Molecular and immunohistochemical analysis of HER2/neu oncogene in synovial sarcoma.

Nuciforo PG, Pellegrini C, Fasani R, Maggioni M, Coggi G, Parafioriti A, Bosari S.

Department of Medicine, Surgery and Dental Sciences, University of Milan, A.O.S. Paolo and IRCCS Ospedale Maggiore, Italy.

Amplification and/or overexpression of HER2/neu have been documented in many types of epithelial tumor and recently has been reported in sarcomas, particularly in osteosarcomas. But the role of HER2/neu alterations in soft tissue tumors remains poorly understood. Thus the present study investigates the expression of HER2/neu in 13 patients with synovial sarcoma (SS). In this study, HER2/neu mRNA levels were measured in frozen tissue samples using a real-time reverse transcription-polymerase chain reaction assay; protein expression was assessed by immunohistochemistry using an anti-HER2/neu polyclonal antibody. Six normal skeletal muscle specimens were used to establish basal levels of HER2/neu mRNA. HER2/neu transcripts were detected in all normal tissues and SSs. Four of 13 sarcomas (31%) demonstrated HER2/neu mRNA levels above the mean value, whereas 3 tumors (23%) displayed HER2/neu protein overexpression. Both membranous and cytoplasmic patterns of immunostaining were observed, and a strong correlation was found between protein expression and mRNA level ($P = 0.01$). Increased HER2/neu mRNA levels were significantly associated with a lower risk of developing recurrences ($P = 0.02$). Moreover, none of the patients with HER2/neu overexpression developed metastasis. Our data demonstrate that HER2/neu is expressed in SSs and that both membrane and cytoplasmic HER2/neu expression correlate with mRNA levels. Our results show that the presence of increased levels of HER2/neu in SSs is associated with a more favorable clinical course. Further studies are needed to assess the role of this oncogene in SSs and to evaluate the application of inhibitory humanized monoclonal antibodies in the treatment regimens for this malignancy.

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Original Contributions

Molecular and Immunohistochemical Analysis of HER2/neu Oncogene in Synovial Sarcoma

PAOLO GIOVANNI NUCIFORO, MD, CATERINA PELLEGRINI, PhD,
ROBERTA FASANI, MD, MARCO MAGGIONI, MD,
GUIDO COGGI, MD, ANTONINA PARAFIORITI, MD,
AND SILVANO BOSARI, MD

Amplification and/or overexpression of HER2/neu have been documented in many types of epithelial tumor and recently has been reported in sarcomas, particularly in osteosarcomas. But the role of HER2/neu alterations in soft tissue tumors remains poorly understood. Thus the present study investigates the expression of HER2/neu in 13 patients with synovial sarcoma (SS). In this study, HER2/neu mRNA levels were measured in frozen tissue samples using a real-time reverse transcription-polymerase chain reaction assay; protein expression was assessed by immunohistochemistry using an anti-HER2/neu polyclonal antibody. Six normal skeletal muscle specimens were used to establish basal levels of HER2/neu mRNA. HER2/neu transcripts were detected in all normal tissues and SSs. Four of 13 sarcomas (31%) demonstrated HER2/neu mRNA levels above the mean value, whereas 3 tumors (23%) displayed HER2/neu protein overexpression. Both membranous and cytoplasmic patterns of immunostaining were observed, and a strong correlation was

found between protein expression and mRNA level ($P = 0.01$). Increased HER2/neu mRNA levels were significantly associated with a lower risk of developing recurrences ($P = 0.02$). Moreover, none of the patients with HER2/neu overexpression developed metastasis. Our data demonstrate that HER2/neu is expressed in SSs and that both membrane and cytoplasmic HER2/neu expression correlate with mRNA levels. Our results show that the presence of increased levels of HER2/neu in SSs is associated with a more favorable clinical course. Further studies are needed to assess the role of this oncogene in SSs and to evaluate the application of inhibitory humanized monoclonal antibodies in the treatment regimens for this malignancy. *Hum Pathol* 34:639-645. © 2003 Elsevier Inc. All rights reserved.

Key Words: HER2/neu, synovial sarcoma, real-time RT-PCR, immunohistochemistry.

Abbreviations: FISH, fluorescence in situ hybridization; RT-PCR, reverse transcription-polymerase chain reaction; SS, synovial sarcoma.

Synovial sarcoma (SS) is an aggressive soft tissue tumor that accounts for up to 10% of sarcomas, with a peak incidence in adolescents and young adults. This tumor occurs in 2 major forms, biphasic and monophasic, and it is cytogenetically characterized by the t(X;18)(p11;q11) translocation, found in >95% of cases. Although traditionally considered to be a high-grade neoplasm, recent investigations have suggested that different factors influence prognosis, including morphological and cytogenetic features, treatment strategies, the ploidy status, and the apoptotic index.¹

The development of new therapeutic advancements, such as the specific targeting of molecular alterations present in human malignancies, has brought to light the

need to identify not only prognostic factors, but also tumor features that are predictive of response to therapy.

One of the most extensively studied molecular targets for therapy is the HER-2/neu proto-oncogene. The HER-2/neu oncogene (also known as c-erbB-2), located on chromosome 17q21, is a member of the tyrosine kinase receptor family and encodes for a 185-kilodalton protein that shows 50% homology with the epidermal growth factor receptor.^{2,3} This gene is amplified and/or overexpressed in 20% to 30% of breast carcinomas^{4,5} and in various other tumors,⁶ and usually is associated with tumor aggressiveness and poor prognosis.^{7,8} Several studies have supported the value of HER-2/neu to predict the response to chemotherapy in breast cancer, and the use of recombinant humanized antibodies to HER-2/neu protein (Trastuzumab) in the care of patients with advanced, metastatic breast tumors has been approved.⁹

The role of HER-2/neu activation in soft tissue tumors remains poorly understood, and scarce molecular data backing immunohistochemical studies have been reported. HER-2/neu protein expression was immunohistochemically studied in 204 sarcomas, including 6 SSs, and overexpression was absent in all these malignant mesenchymal neoplasms.¹⁰

Recently, HER-2/neu alterations have been described in osteosarcoma, with a high incidence of pro-

From the Department of Medicine, Surgery and Dental Sciences, University of Milan, A.O.S. Paolo and IRCCS Ospedale Maggiore, Milan, Italy; Department of Pathology, Orthopedic Institute Gaetano Pini, Milan, Italy; and Interuniversity Center of Cancer Research, Milan, Italy. Accepted for publication March 5, 2003.

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Address correspondence and reprint requests to Paolo Giovanni Nuciforo, MD, Division of Anatomic Pathology, A.O.S. Paolo, Via A. Di Rudinì 8, 20142, Milano, Italy.

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TABLE 1. Clinicopathologic Features of 13 Patients With SS

| No. | Age(y)/G | Site | Subtype | Surgery | Chemo/Radio | Rec | Mets | HER2 | FU(mo) |
|-----|----------|------|---------|---------|-------------|-----|------|------|--------|
| 1 | 68/F | KI | MF | RR | No | Yes | Yes | L | 108 |
| 2 | 71/F | KE | BF | WA | No | No | No | L | 12 |
| 3 | 27/M | KI | BF | ME | Yes | Yes | Yes | L | 24 |
| 4 | 39/M | T | MF | WE | Yes | No | No | H | 48 |
| 5 | 15/M | N | MF,PD | WE | Yes | No | No | H | 48 |
| 6 | 41/F | F | MF,PD | WA | Yes | No | No | H | 48 |
| 7 | 57/F | KI | MF | WA | No | N/A | N/A | L | N/A |
| 8 | 48/M | F | BF,PD | ME | Yes | Yes | No | L | 36 |
| 9 | 29/F | T | PD | WE | Yes | Yes | Yes | L | 36 |
| 10 | 27/M | KE | BF | WE | No | No | No | H | 24 |
| 11 | 62/F | A | MF | WE | No | Yes | No | L | 24 |
| 12 | 62/F | F | MF | WA | No | No | No | L | 2 |
| 13 | 34/F | L | MF | RR | No | No | No | L | 2 |

Abbreviations: Age, age at diagnosis; G, gender; M, male; F, female; Site, anatomic location; KI, knee, intra-articular; KE, knee, extra-articular; T, thigh; N, neck; F, foot; A, arm; L, leg; Subtype, histological subtype; MF, monophasic fibrous; BF, biphasic; MF-PD, monophasic fibrous with poorly differentiated areas; BF-PD, biphasic with poorly differentiated areas; PD, poorly differentiated; Surgery, primary surgical therapy; RR, radical en bloc resection; WA, wide through-bone amputation; ME, marginal en bloc excision; WE, wide en bloc excision; Chemo/Radio, adjuvant postoperative chemotherapy and/or radiotherapy; Rec, recurrence; Mets, presence of metastasis; HER2, HER2 mRNA expression; L, low expression; H, high expression; FU, follow-up status; N/A, not available.

tein expression, ranging from 42% to 61%.¹¹⁻¹⁵ Indeed, despite limited information on Her2/neu in this type of malignancy, based mostly on immunohistochemical findings, 2 clinical trials of Trastuzumab have been initiated for recurrent and metastatic osteosarcoma patients (http://www.cancer.gov/clinical_trials: MSKCC-99097/NCI-T98-0083 and COG-AOST0121).

Therefore, we evaluated the mRNA expression and the gene product of HER-2/neu in 13 SS patients using real-time reverse transcription-polymerase chain reaction (RT-PCR) and immunohistochemistry. To the best of our knowledge, this is the first report documenting increased levels of HER-2/neu mRNA and protein in SS.

MATERIALS AND METHODS

Patient Population

Thirteen patients with primary SSs, obtained from the files of the Department of Pathology, Gaetano Pini Orthopedic Institute, were included in this study. Cases were chosen based on the availability of frozen primary tumor. Patient age ranged from 15 to 71 years (mean, 44.6 years). Anatomic sites included the knee (5 patients, 3 intra-articular), thigh (2 patients), foot (3 patients), arm (1 patient), neck (1 patient), and lower leg (1 patient). The histological subtypes were 3 biphasic (BF), 6 monophasic fibrous (MF), 2 monophasic fibrous with poorly differentiated areas (MF-PD), 1 biphasic with poorly differentiated areas (BF-PD), and 1 poorly differentiated (PD). Clinical staging was IIB for all the patients. Local surgical excision was performed in 9 patients; amputation, in 4 patients. Recurrence was observed in 5 cases with subsequent amputation (3 patients) and local excision (2 patients). Metastasis occurred to lung (2 patients) and inguinal lymph nodes (1 patient). Five patients received adjuvant postoperative chemotherapy, and 1 patient (case 4) was treated with chemotherapy associated with radiotherapy. Follow-up ranged from 2 to 108 months (mean, 34.3 months). Clinicopathologic data are summarized in Table 1.

Pathologic Studies and Frozen Tissue Selection

In all cases, the primary tumor was available for study. Surgically resected tumor tissues were partly snap-frozen in liquid nitrogen and stored at -80°C for RNA extraction, and partly fixed in buffered formalin and embedded in paraffin blocks. Hematoxylin and eosin-stained sections were re-evaluated and graded according to FNCLCC grading system.¹⁶ Frozen tissue blocks were handled as follows: 4-μ-thick frozen sections were cut and stained with hematoxylin and eosin to determine the percentage of tumor cells present in the specimen. We used tissue blocks with tumor cells comprising more than 80% of the specimen. About 10 20-μ-thick sections were collected into Eppendorf tubes. Another 4-μ-thick frozen section was cut after the serial sections and examined by light microscopy to guarantee the percentage of tumor cells collected. Trizol (Life Technologies; Gibco BRL, Gaithersburg, MD) was used for RNA extraction, according to the manufacturer's protocol. RNA was quantified spectrophotometrically.

cdNA Synthesis

Total RNA (200 ng) was reverse-transcribed in a total volume of 50 μL containing 1 × TaqMan buffer, 5.5 mmol MgCl₂, 1 mmol deoxynucleotides, 2.5 μmol random hexamers, 20 U RNase inhibitor, and 62.5 U MuL_v reverse transcriptase. The samples were incubated at 25°C for 10 minutes, 48°C for 30 minutes and 95°C for 5 minutes.

PCR Amplification

Amplification reactions were performed with the Universal TaqMan 2 × PCR mastermix in a volume of 25 μL containing 300 nmol of each primer, 100 nmol of probe, and 5 μL of cDNA. Both β-actin and HER2/neu amplification were done in duplicate for each sample.

The thermal cycling conditions included 2 minutes at 50°C and 10 minutes at 95°C, followed by 40 cycles of 95°C for 15 seconds and 60°C for 1 minute. All reagents used for

RT-PCR were purchased from Applied Biosystems (Foster City, CA).

Primers and Probes

Primers and probes for β -actin and HER2/neu mRNA were chosen using the computer program Primer Express (Applied Biosystems). Sequences of the forward primer for HER2/neu mRNA (GenBank accession number X03363) were 5'-TCC TGT GTG CAC CTG GAT CAC-3' and the reverse primer 5'-CCA AAG ACC ACC CCC AAG A-3'; the sequence of the TaqMan probe was 5'(FAM)-AGC ACA ATG CCA ACC ACC CCA CA-(TAMRA)-3'. Sequences of the forward primer for β -actin mRNA (GenBank accession number X00351) were 5'-TCC TTC CTG CCC ATG CAG-3' and the reverse primer 5'-AGC AGG ACC AAT GAT CTT GAT CTT-3'; the sequence of the TaqMan probe was 5'(FAM)-CCT GTG GCA TCC AGC AAA CTA CCT TC-(TAMRA)-3'. Probes were purchased from Applied Biosystems.

Real-Time RT-PCR

To measure HER2/neu expression in these tumors we used a real-time quantitative RT-PCR based on TaqMan methodology, as previously described,¹⁷ with minor modifications. Briefly, this technique allows, by means of fluorescence emission, to find the cycling point when PCR product is detectable (Ct value or threshold cycle). As previously reported, the Ct value correlates to the starting quantity of the target mRNA.¹⁸ To normalize the amount of total RNA present in each reaction, we amplified the housekeeping gene β -actin, which is assumed to be constant in both normal samples and tumor tissues.

Our results are expressed as relative levels of HER2/neu mRNA, referred to a sample, called a "calibrator," chosen to represent 1X expression of this gene. The calibrator was a breast cancer cellular line (MCF-7)¹⁹ that was analyzed on every assay plate with the unknown samples. All of the analyzed tumors expressed n-fold HER2/neu mRNA relative to the calibrator.

The amount of target, normalized to an endogenous reference (β -actin) and relative to the calibrator, was defined by the $\Delta\Delta Ct$ method as described by Livak K (Sequence Detector User Bulletin 2; Applied Biosystems). Specifically, the formula is applied as follows:

$$\text{target amount} = 2^{-\Delta\Delta Ct}$$

where $\Delta\Delta Ct = [Ct(\text{HER2/neu sample}) - Ct(\beta\text{-actin sample})] - [Ct(\text{HER2/neu calibrator}) - Ct(\beta\text{-actin calibrator})]$.

Immunohistochemistry

Formalin-fixed, paraffin-embedded tissue sections were deparaffinized, rehydrated, and exposed to the primary antibody using the EnVision+ system (Dako, Carpinteria, CA). Primary anti-HER2/neu antibody (rabbit polyclonal antibody, catalog number A0485; Dako) was applied in a dilution of 1:2000 for 60 minutes at room temperature. Before exposure to the primary antibody, sections were microwave-pretreated in EDTA, pH 8.0, to retrieve antigenicity, and incubated with endogenous peroxidase-blocking solution for 10 minutes at room temperature. Positive control, constituted by a breast carcinoma showing more than 80% positive staining for HER2/neu, as well as negative control, in which the primary antibody was omitted, were stained in parallel.

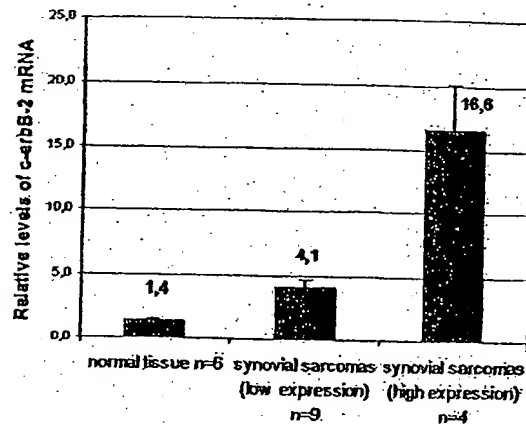


FIGURE 1. Distribution of HER2/neu mRNA levels in normal muscle tissues, and in low- and high-expression sarcomas. Data are expressed as mean and standard error of the mean for each group.

All cases were examined for both cytoplasmic and membrane immunoreactivity. Cytoplasmic staining was evaluated on a semiquantitative scale, according to Kilpatrick et al with minor modifications,²⁰ and reported as 0 (no staining or staining in <10% of cells), 1+ (weak staining in >10% of cells), 2+ (moderate staining in >10% of cells), or 3+ (strong staining in >10% of cells). The presence of a membranous pattern of staining was recorded separately and scored as absent (no staining or weak staining in <10% of cells) or present (complete and/or incomplete staining in >10% of cells). Tumors with a cytoplasmic score of 3+ were considered to have high HER2/neu protein expression.

Statistical Analysis

Statistical differences were calculated by Fisher's exact test. The t-test method was used to evaluate the differences between groups. Differences were considered statistically significant when P was <0.05.

RESULTS

HER2/neu mRNA Evaluation

All of the tissues analyzed contained detectable levels of HER2/neu mRNA. Six normal tissue samples (skeletal muscle) were used to establish basal level of HER2/neu mRNA. All the normal samples expressed very low levels of HER2/neu mRNA, ranging from 0.9 to 1.9 n (mean, 1.4 n). Among the 13 tumor samples, HER2/neu levels varied greatly, ranging from 2.1 to 24 n. Setting a cutoff level at 7.9 n (a value that represents the mean value of expression distribution of the SSs), 9 cases (69%) had low HER2/neu expression and 4 cases (31%) had high HER2/neu expression (Fig 1; Table 1). The difference between the 2 groups (low and high HER2/neu tumors) was statistically significant ($P = 0.0004$).

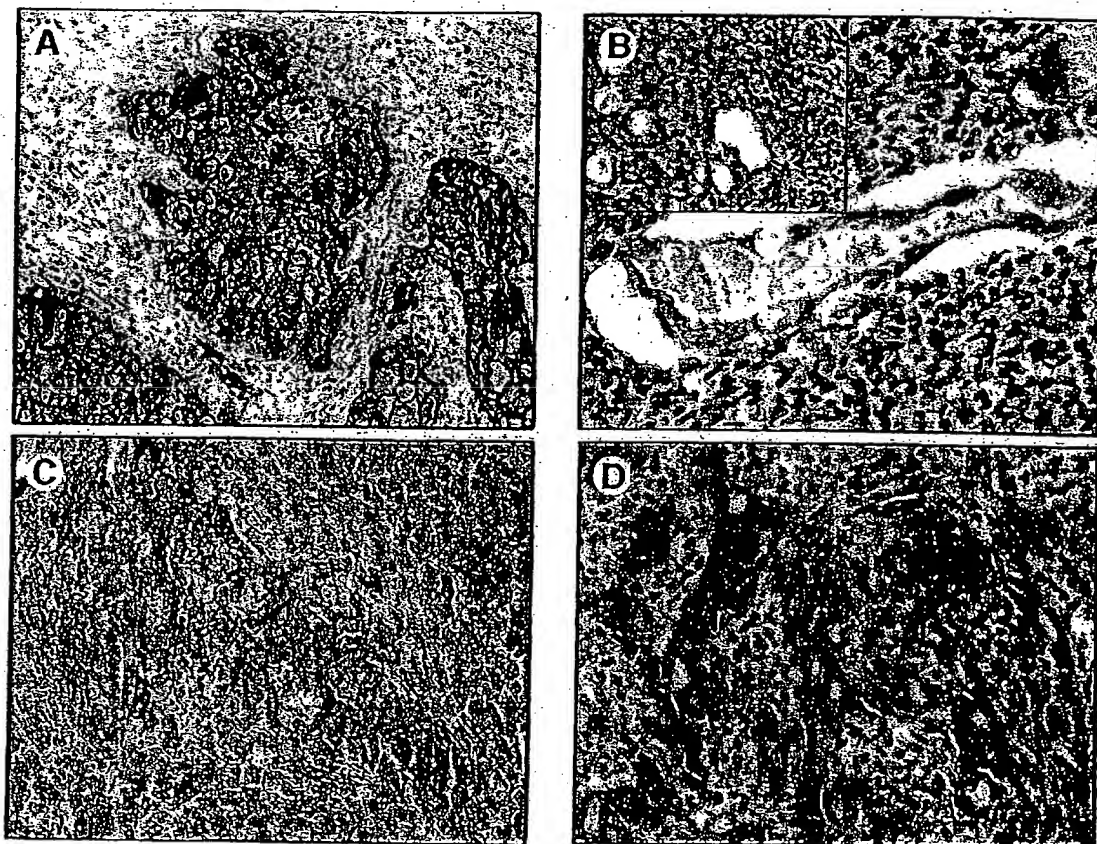


FIGURE 2. Immunohistochemical localization of HER2/neu in SS. (A) Positive control (breast cancer) showing typical strong membrane pattern of positivity. (B) Case 2, a biphasic SS, displaying very focal membrane staining limited to a gland. Inset: focal weak cytoplasmic positivity in the epithelial component of the same case. (C) Case 10, the epithelial component of this biphasic SS, displaying strong membrane pattern of staining. (D) Case 4, an epithelioid area in a monophasic fibrous SS, showing strong cytoplasmic positivity.

HER2/neu Protein Expression

Staining with HER2/neu antibody revealed a variable cytoplasmic and membrane staining pattern. Three tumors (23%) showed strong staining involving both the cell membrane and the cytoplasm (cases 4, 5, and 10); weak to moderate, exclusively cytoplasmic staining was observed in 7 cases (cases 1, 3, 6, 7, 8, 9, and 11). No staining was detected in 2 tumors (cases 12 and 13). In 1 case (case 2), a cluster of glandular structures representing <5% of the tumor showed weak cytoplasmic and very focal membrane staining, the latter limited to a single gland. The epithelial/epithelioid components exhibited stronger cytoplasmic staining compared with the spindle-cell component of the tumors. Membrane staining was predominantly incomplete and limited to the epithelial/epithelioid areas. All 3 cases with high immunohistochemical expression of HER2/neu (cases 4, 5, and 10) were grade III sarcomas, including 1 MF, 1 BF, and 1 MF-PD SSs.

Examples of HER2/neu cytoplasmic and membrane staining are depicted in Figure 2.

Correlation of Molecular and Immunohistochemical Results

A strong, statistically significant association was present between protein expression, for both membrane and cytoplasmic staining, and HER2/neu mRNA levels ($P = 0.01$), although 1 case (case 6) displayed discordant results. Interestingly, this neoplasm showed high HER2/neu mRNA levels, whereas only weak staining, limited to the cytoplasm of a minority of tumor cells, was detected by immunohistochemical analysis.

HER2/neu Expression and Clinicopathologic Parameters

Both HER2/neu protein expression and mRNA levels were evaluated to establish the relationships to

TABLE 2. Correlation Between Clinicopathologic Features and HER2/neu Expression as Detected by IHC and RT-PCR

| Variable | HER2/neu | | | | | |
|---------------------|----------|---|---------|-----|---|---------|
| | IHC | | | PCR | | |
| | L | H | P value | L | H | P value |
| Age (years) | | | | | | |
| <40 | 3 | 3 | | 3 | 3 | |
| >40 | 7 | 0 | NS | 6 | 1 | NS |
| Sex | | | | | | |
| Female | 8 | 0 | | 7 | 1 | |
| Male | 2 | 3 | 0.03 | 2 | 3 | NS |
| Tumor size (cm) | | | | | | |
| <5 | 3 | 2 | | 3 | 2 | |
| >5 | 7 | 1 | NS | 6 | 2 | NS |
| Histological grade | | | | | | |
| II | 3 | 0 | | 3 | 0 | |
| III | 7 | 3 | NS | 6 | 4 | NS |
| Histological type | | | | | | |
| MF | 5 | 1 | | 5 | 1 | |
| BF | 2 | 1 | | 2 | 1 | |
| PD | 3 | 1 | NS | 2 | 2 | NS |
| Chemo/Radiotherapy* | | | | | | |
| Yes | 4 | 2 | | 3 | 3 | |
| No | 5 | 1 | NS | 5 | 1 | NS |
| Recurrence† | | | | | | |
| Yes | 5 | 0 | | 5 | 0 | |
| No | 2 | 3 | NS | 1 | 4 | 0.02 |
| Metastasis† | | | | | | |
| Yes | 3 | 0 | | 3 | 0 | |
| No | 4 | 3 | NS | 3 | 4 | NS |

Abbreviations: L, low expression; H, high expression; NS, not significant; MF, monophasic fibrous; BF, biphasic; PD, poorly differentiated (including MF and BF with poorly differentiated areas).

*Information not available for case 7.

†Cases 7, 12, and 13 were excluded from the analysis.

clinicopathologic features, including local recurrence and metastatic disease. Two cases (cases 12 and 13) with follow-up less than 12 months and 1 case (case 7) for which clinical information was not available were excluded from the analysis of recurrences and metastatic behavior.

No correlation was observed between HER2/neu mRNA expression and age, sex, tumor size, tumor grade, histotype, and metastasis. A correlation between sex of the patients and HER2/neu protein expression was found. In fact, none of the female patients showed high HER2/neu protein expression ($P = 0.03$). Patients with high Her2/neu mRNA levels had a lower risk of recurrence than those with low Her2/neu mRNA levels ($P = 0.02$). None of the cases with high HER2/neu mRNA levels developed metastatic foci, although the small number of observations precluded reaching statistical significance ($P = 0.1$). Results are detailed in Table 2.

DISCUSSION

The present work provides the first combined molecular by real-time RT-PCR and immunohistochemical evidence that HER2/neu overexpression occurs in SSs.

Our results indicate that this parameter may provide prognostic information and suggest that a specific therapy with humanized monoclonal antibodies against HER2/neu may be considered in a significant number of SSs.

The HER2/neu oncogene has been extensively investigated as a prognostic factor and more recently as a predictor of response to therapy. It has been demonstrated in breast cancer, where HER2/neu overexpression is usually associated with gene amplification,²¹ and in other epithelial tumors, including ovarian, gastric, lung, and urinary bladder carcinomas.

HER2/neu amplification/overexpression appears to be an early event in oncogenic transformation by interacting with other members of the HER family.³ In breast cancer, it is involved in cell cycle and apoptotic pathways through the antiapoptotic effects mediated by p53 and p21 deregulation.^{22,23}

Whether HER2/neu overexpression plays an important role in mesenchymal neoplasms remains controversial. An immunohistochemical study of sarcomas, using a monoclonal antibody, reported no evidence of immunoreactivity for HER-2/neu in 6 SSs as well as in other 197 mesenchymal tumors, with cytoplasmic reactivity observed only in 1 case of peripheral neuroepithelioma.¹⁰ A recent investigation reported gene expression profiles of 41 soft tissue tumors with cDNA microarray analysis. Among these sarcomas, 6 monophasic SSs were characterized by a unique expression pattern of a cluster of 104 genes, including the epidermal growth factor receptor, which shows 50% homology with the HER2/neu gene.²⁴ These data also suggest that the erb-B receptor family plays a significant role in SS. It has been demonstrated that a variable number of osteosarcomas overexpress HER2/neu.¹¹⁻¹⁵ However, more recent studies^{20,25,26} were unable to detect any HER2/neu gene amplification and/or overexpression using fluorescence in situ hybridization (FISH), RT-PCR, and immunohistochemistry.

Differences in the techniques used may play an important role and explain (at least in part) these discrepancies. HER2/neu alterations can be evaluated using different techniques including immunohistochemistry, FISH, Southern hybridization, Northern blot, and competitive, differential, or real-time PCR.²⁷ Immunohistochemistry is the most common method for detection of HER2/neu overexpression, but it is significantly affected by the sensitivity and specificity of the antibodies used, the type of tissue (frozen versus formalin-fixed), and the various interpretative criteria and scoring systems used to evaluate cases. Indeed, most studies of HER2/neu expression in osteosarcoma used immunohistochemical techniques, with different monoclonal or polyclonal antibodies. The discrepancy in results may stem from the use of different antibodies, as well as a lack of standardized evaluation.

For these reasons, to evaluate HER2/neu immunoreactivity in our study, we used a polyclonal antibody (Dako, Carpinteria, CA), arguably the most diffuse and thoroughly tested antibody for HER2/neu assessment. Furthermore, we investigated HER2/neu mRNA ex-

pression with real-time RT-PCR, because it has been demonstrated that mRNA levels correlates tightly with protein expression.²⁸ At present, real-time RT-PCR probably represents the most powerful tool for quantitative analysis, because it allows better internal control and reduction of sample contamination, and provides more objective results.¹⁸

We analyzed HER2/neu gene expression at the mRNA and protein level in 13 cases of SS. HER2/neu expression was found in all of the cases investigated, and mRNA content in the tumors varied from 2.1 to 24 n.

The variability of mRNA levels in SSs is reflected on heterogeneity of protein expression pattern as detected by immunohistochemistry. We found that HER2/neu immunoreactivity correlates strongly with mRNA levels. A convincing cytoplasmic immunoreactivity was documented with the polyclonal antibody in 10 of 13 sarcomas. Distinct membranous staining was observed in 3 cases, although it was never comparable to the positive breast control. It was predominantly incomplete and identified in the epithelial/epithelioid component of SS. Interestingly, all of the cases with strong cytoplasmic staining also exhibited a membrane-staining pattern.

In breast cancers, a membranous pattern of staining is thought to be specific for HER2/neu protein expression and correlated with gene amplification, whereas cytoplasmic staining is usually considered non-specific.²⁹ However, cytoplasmic positivity for HER2/neu has been reported to be prognostically significant in other tumor types, including bladder, colon, pancreas, thyroid, and nasopharyngeal carcinomas, and even in breast cancer.³⁰⁻³⁵

Patients with high HER2/neu mRNA expression had a significantly lower risk of recurrence. Similarly, all of the cases with high HER2/neu expression did not metastasize, although this correlation did not reach statistical significance due to the small number of cases studied. These data suggest that HER2/neu plays a role in the biology of SS and that HER2/neu overexpression may be linked to a less-aggressive clinical behavior. Indeed, unlike many cancers where HER2/neu overexpression has been shown to correlate with poor prognosis, HER2/neu levels are linked to a more favorable clinical course in other malignant tumors, such as thyroid carcinoma and osteosarcoma.^{15,34}

The molecular mechanisms responsible for the action of HER2/neu in SSs are unknown. A possible interaction between HER2/neu and the other members of HER family could be important in tumorigenesis. Derangements of other oncogenes, tumor suppressors, and apoptosis regulators have been described in SSs. For instance, many SSs have been shown to be diffusely positive for bcl-2 family proteins (bcl-2, bax, bcl-x, and bak). These members of the bcl-2 family are involved in the regulation of apoptosis in SS.³⁶ This raises the hypothesis that complex alterations in apoptosis-controlling mechanisms are present in these neoplasms, with HER2/neu interacting with Bcl-2 family

members. Further studies are needed to clarify the mechanisms of apoptosis in SS.

Depending on the size and location, the therapy of choice for SS is radical local excision or amputation. Whenever radical surgery cannot be performed, radiotherapy in concert with local excision is suggested in an attempt to avoid the need for amputation. Only recently has a study evaluated the possible role of chemotherapy in the treatment of SS.³⁷

The role of molecular markers in predicting treatment responsiveness is currently the focus of extensive investigation. Breast cancer patients with high HER2/neu expression appeared to benefit from high-dose CAF (cyclophosphamide, adriamycin, and 5-fluorouracil) therapy.³⁸ In our study, 3 of 4 patients with high HER2/neu expression received adjuvant chemotherapy with ifosfamide; these patients had a favorable clinical outcome. These data raise the possibility that HER2/neu may have value in predicting which patients are likely to respond to specific adjuvant chemotherapy regimens. Whether the favorable significance of HER2/neu expression depends on predicting clinical recurrence, response to chemotherapy, or both remains to be fully elucidated in SS patients.

To the best of our knowledge, this is the first report that shows expression of HER2/neu in primary SS by real-time RT-PCR. Elevated levels of HER2/neu mRNA and protein are found in a significant group of SS patients, and these levels appear to correlate with features of good prognosis. Furthermore, our results suggest that this mechanism of disease in SS may be the target of specific inhibitory therapies based on humanized monoclonal antibodies. Considering the small number of patients examined, further investigation is needed to confirm these preliminary findings.

REFERENCES

1. Weiss SW, Goldblum JR: Synovial sarcoma, in Weiss SW, Goldblum JR (eds): *Enzinger and Weiss's Soft Tissue Tumors*, 4th ed. St Louis, MO, Mosby-Year Book, 2001, pp 1483-1509
2. Schechter AL, Stern DF, Vaidyanathan L, et al: The neu oncogene: An erb-B-related gene encoding an 185,000 Mr tumor antigen. *Nature* 312:513-516, 1984
3. Bargmann CI, Hung MC, Weinberg RA: The neu oncogene encodes an epidermal growth factor receptor-related protein. *Nature* 319:226-230, 1986
4. Ali IU, Campbell G, Lidereau R, et al: Lack of evidence for the prognostic significance of c-erbB-2 amplification in human breast carcinoma. *Oncogene Res* 3:139-146, 1988
5. Berger MS, Locher GW, Sauer S, et al: Correlation of c-erbB-2 gene amplification and protein expression in human breast carcinoma with nodal status and nuclear grading. *Cancer Res* 48:1238-1243, 1988
6. Scholl S, Beuzec-Boc P, Pouillart P: Targeting HER2 in other tumor types. *Ann Oncol* 12(Suppl1):S81-S87, 2001
7. Slamon DJ, Clark GM, Wong SG, et al: Human breast cancer: Correlation to relapse and survival with amplification of the HER-2/neu oncogene. *Science* 235:177-182, 1987
8. Wright C, Angus B, Nicholson S, et al: Expression of c-erbB-2 oncoprotein: A prognostic indicator in human breast cancer. *Cancer Res* 49:2087-2090, 1989
9. Slamon D, Leyland-Jones B, Shak S, et al: Use of chemotherapy plus a monoclonal antibody against Her2/neu for metastatic

HER2/neu ONCOGENE IN SYNOVIAL SARCOMA (Nuciforo et al)

- breast cancer that overexpress Her2/neu. *N Engl J Med* 344:783-792, 2001
10. George E, Niehans GA, Swanson PE, et al: Overexpression of the c-erbB-2 oncogene in sarcomas and small round cell tumors of childhood. An immunohistochemical investigation. *Arch Pathol Lab Med* 116:1033-1035, 1992
11. Onda M, Matsuda S, Higaki S, et al: ErbB-2 expression is correlated with poor prognosis for patients with osteosarcoma. *Cancer* 77:71-78, 1996
12. Gorlick R, Huvois AG, Heller G, et al: Expression of HER2/erbB-2 correlates with survival in osteosarcoma. *J Clin Oncol* 17:2781-2788, 1999
13. Akatsuka T, Wada T, Kokai Y, et al: Loss of ErbB2 expression in pulmonary metastatic lesions in osteosarcoma. *Oncology* 60:361-366, 2001
14. Morris CD, Gorlick R, Huvois G, et al: Human epidermal growth factor receptor 2 as a prognostic indicator in osteogenic sarcoma. *Clin Orthop* 382:59-65, 2001
15. Akatsuka T, Wada T, Kokai Y, et al: ErbB2 expression is correlated with increased survival of patients with osteosarcoma. *Cancer* 94:1397-1404, 2002
16. Coindre JM, Trojani M, Contesso G, et al: Reproducibility of a histopathologic grading system for adult soft tissue sarcoma. *Cancer* 58:306-309, 1986
17. Marchetti A, Pellegrini C, Buttitta F, et al: Prediction of survival in stage I lung carcinoma patients by telomerase function evaluation. *Lab Invest* 82:729-736, 2002
18. Heid CA, Stevens J, Livak KJ, et al: Real time quantitative PCR. *Genome Res* 6:986-994, 1996
19. Kraus MH, Popescu NC, Amsbaugh SC, et al: Overexpression of the EGF receptor-related proto-oncogene erbB-2 in human mammary tumor cell lines by different molecular mechanisms. *EMBO J* 6:605-610, 1987
20. Kilpatrick SE, Geisinger KR, King TS, et al: Clinicopathologic analysis of HER-2/neu immunoreexpression among various histologic subtypes and grades of osteosarcoma. *Mod Pathol* 14:1277-1283, 2001
21. Slamon DJ, Godolphin W, Jones LA, et al: Studies of the HER-2/neu proto-oncogene in human breast and ovarian cancer. *Science* 244:707-712, 1989
22. Harari D, Yarden Y: Molecular mechanisms underlying ErbB2/HER2 action in breast cancer. *Oncogene* 19:6102-6114, 2000
23. Zhou BP, Liao Y, Xia W, et al: HER-2/neu induces p53 ubiquitination via Akt-mediated MDM2 phosphorylation. *Nat Cell Biol* 3:973-982, 2001
24. Nielsen OT, West RB, Linn SC, et al: Molecular characterization of soft tissue tumours: A gene expression study. *Lancet* 359:1301-1307, 2002
25. Thomas DG, Giordano TJ, Sanders D, et al: Absence of HER2/neu gene expression in osteosarcoma and skeletal Ewing's sarcoma. *Clin Cancer Res* 8:788-793, 2002
26. Maitra A, Wanzer D, Weinberg AG, et al: Amplification of the Her-2/neu oncogene is uncommon in pediatric osteosarcomas. *Cancer* 92:677-683, 2001
27. Hanna W, Kahn HJ, Trudeau M: Evaluation of HER-2/neu (erbB-2) status in breast cancer: From bench to bedside. *Mod Pathol* 12:827-834, 1999
28. Naber SP, Tsutsumi Y, Yin S, et al: Strategies for the analysis of oncogene overexpression. Studies of the new oncogene in breast carcinoma. *Am J Clin Pathol* 94:125-136, 1990
29. Tetu B, Brisson J: Prognostic significance of HER-2/neu oncoprotein expression in node-positive breast cancer. The influence of the pattern of immunostaining and adjuvant therapy. *Cancer* 73:2359-2365, 1994
30. Coombs LM, Pigott DA, Sweeney E, et al: Amplification and over-expression of c-erbB-2 in transitional cell carcinoma of the urinary bladder. *Br J Cancer* 63:601-608, 1991
31. Kay EW, Mulcahy H, Walsh CB, et al: Cytoplasmic c-erbB-2 protein expression correlates with survival in Dukes' B colorectal carcinoma. *Histopathology* 25:455-461, 1994
32. Roychowdhury DF, Tseng A Jr, Fu KK, et al: New prognostic factors in nasopharyngeal carcinoma. Tumor angiogenesis and c-erbB-2 expression. *Cancer* 77:1419-1426, 1996
33. Hall PA, Hughes CM, Staddon SL, et al: The c-erb B-2 proto-oncogene in human pancreatic cancer. *J Pathol* 161:195-200, 1990
34. Sugg SL, Ezzat S, Zheng L, et al: Cytoplasmic staining of erbB-2 but not mRNA levels correlates with differentiation in human thyroid neoplasia. *Clin Endocrinol (Oxf)* 49:629-637, 1998
35. Keshgegian AA, Cnaan A: erbB-2 oncoprotein expression in breast carcinoma. Poor prognosis associated with high degree of cytoplasmic positivity using CB-11 antibody. *Am J Clin Pathol* 108:456-463, 1997
36. Kawauchi S, Fukuda T, Oda Y, et al: Prognostic significance of apoptosis in synovial sarcoma: Correlation with clinicopathologic parameters, cell proliferative activity, and expression of apoptosis-related proteins. *Mod Pathol* 13:755-765, 2000
37. Rosen G, Forscher C, Lowenbraun S, et al: Synovial sarcoma: Uniform response of metastasis to high dose ifosfamide. *Cancer* 73:2506-2511, 1994
38. Muss HB, Thor AD, Berry DA, et al: c-erbB-2 expression and response to adjuvant therapy in women with node-positive early breast cancer. *N Engl J Med* 330:1260-1266, 1994

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Differential expression of heat shock protein 70 in well healing and chronic human wound tissue.

Oberringer M, Baum HP, Jung V, Welter C, Frank J, Kuhlmann M, Mutschler W, Hanselmann RG.

Department of Traumatology, University of Saarland, Germany.

Heat shock protein 70 (hsp 70) is an important member of the heat shock protein family, which is induced by different forms of stress. We attempted to find out if hsp 70 is also involved in wound healing, which likewise resembles a stress situation for cells too. Therefore we collected tissue samples from well healing and chronic human wound tissue. We used Northern- and Western-blot analysis to study the expression of hsp 70. At the protein level we found a strong correlation between well healing wounds and high expression of hsp 70, whereas chronic wounds showed no or weak expression. Interestingly hsp 70 mRNA did not show this significant correlation, displaying a variant expression pattern in the same kind of wound tissue, possibly due to unknown posttranscriptional regulating step, which has to be investigated in further studies. To localize hsp 70 mRNA and protein was used insitu hybridization and immunohistochemistry. Both displayed an overexpression in endothelial cells of capillary vessels.

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Orntoft TF, Thykjaer T, Waldman FM, Wolf H, Celis JE.

Department of Clinical Biochemistry, Molecular Diagnostic Laboratory, Aarhus University Hospital, Skejby, DK-8200 Aarhus N, Denmark. orntoft@kba.sks.au.dk

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Genome-wide Study of Gene Copy Numbers, Transcripts, and Protein Levels in Pairs of Non-invasive and Invasive Human Transitional Cell Carcinomas*

Torben F. Ørntoft†§, Thomas Thykjaer¶, Frederic M. Waldman||, Hans Wolf**, and Julio E. Celis††

Gain and loss of chromosomal material is characteristic of bladder cancer, as well as malignant transformation in general. The consequences of these changes at both the transcription and translation levels is at present unknown partly because of technical limitations. Here we have attempted to address this question in pairs of non-invasive and invasive human bladder tumors using a combination of technology that included comparative genomic hybridization, high density oligonucleotide array-based monitoring of transcript levels (5600 genes), and high resolution two-dimensional gel electrophoresis. The results showed that there is a gene dosage effect that in some cases superimposes on other regulatory mechanisms. This effect depended ($p < 0.015$) on the magnitude of the comparative genomic hybridization change. In general (18 of 23 cases), chromosomal areas with more than 2-fold gain of DNA showed a corresponding increase in mRNA transcripts. Areas with loss of DNA, on the other hand, showed either reduced or unaltered transcript levels. Because most proteins resolved by two-dimensional gels are unknown it was only possible to compare mRNA and protein alterations in relatively few cases of well focused abundant proteins. With few exceptions we found a good correlation ($p < 0.005$) between transcript alterations and protein levels. The implications, as well as limitations, of the approach are discussed. *Molecular & Cellular Proteomics* 1:37–45, 2002.

Aneuploidy is a common feature of most human cancers (1), but little is known about the genome-wide effect of this

phenomenon at both the transcription and translation levels. High throughput array studies of the breast cancer cell line BT474 has suggested that there is a correlation between DNA copy numbers and gene expression in highly amplified areas (2), and studies of individual genes in solid tumors have revealed a good correlation between gene dose and mRNA or protein levels in the case of c-erb-B2, cyclin D1, *ems1*, and N-myc (3–5). However, a high cyclin D1 protein expression has been observed without simultaneous amplification (4), and a low level of c-myc copy number increase was observed without concomitant c-myc protein overexpression (6).

In human bladder tumors, karyotyping, fluorescent *in situ* hybridization, and comparative genomic hybridization (CGH)¹ have revealed chromosomal aberrations that seem to be characteristic of certain stages of disease progression. In the case of non-invasive pTa transitional cell carcinomas (TCCs), this includes loss of chromosome 9 or parts of it, as well as loss of Y in males. In minimally invasive pT1 TCCs, the following alterations have been reported: 2q–, 11p–, 1q+, 11q13+, 17q+, and 20q+ (7–12). It has been suggested that these regions harbor tumor suppressor genes and oncogenes; however, the large chromosomal areas involved often contain many genes, making meaningful predictions of the functional consequences of losses and gains very difficult.

In this investigation we have combined genome-wide technology for detecting genomic gains and losses (CGH) with gene expression profiling techniques (microarrays and proteomics) to determine the effect of gene copy number on transcript and protein levels in pairs of non-invasive and invasive human bladder TCCs.

EXPERIMENTAL PROCEDURES

Material—Bladder tumor biopsies were sampled after informed consent was obtained and after removal of tissue for routine pathology examination. By light microscopy tumors 335 and 532 were staged by an experienced pathologist as pTa (superficial papillary),

From the †Department of Clinical Biochemistry, Molecular Diagnostic Laboratory and **Department of Urology, Aarhus University Hospital, Skejby, DK-8200 Aarhus N, Denmark, ‡AROS Applied Biotechnology ApS, Gustav Wiedsvej 10, DK-8000 Aarhus C, Denmark, ¶UCSF Cancer Center and Department of Laboratory Medicine, University of California, San Francisco, CA 94143-0808, and ††Institute of Medical Biochemistry and Danish Centre for Human Genome Research, Ole Worms Allé 170, Aarhus University, DK-8000 Aarhus C, Denmark

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¹ The abbreviations used are: CGH, comparative genomic hybridization; TCC, transitional cell carcinoma; LOH, loss of heterozygosity; PA-FABP, psoriasis-associated fatty acid-binding protein; 2D, two-dimensional.

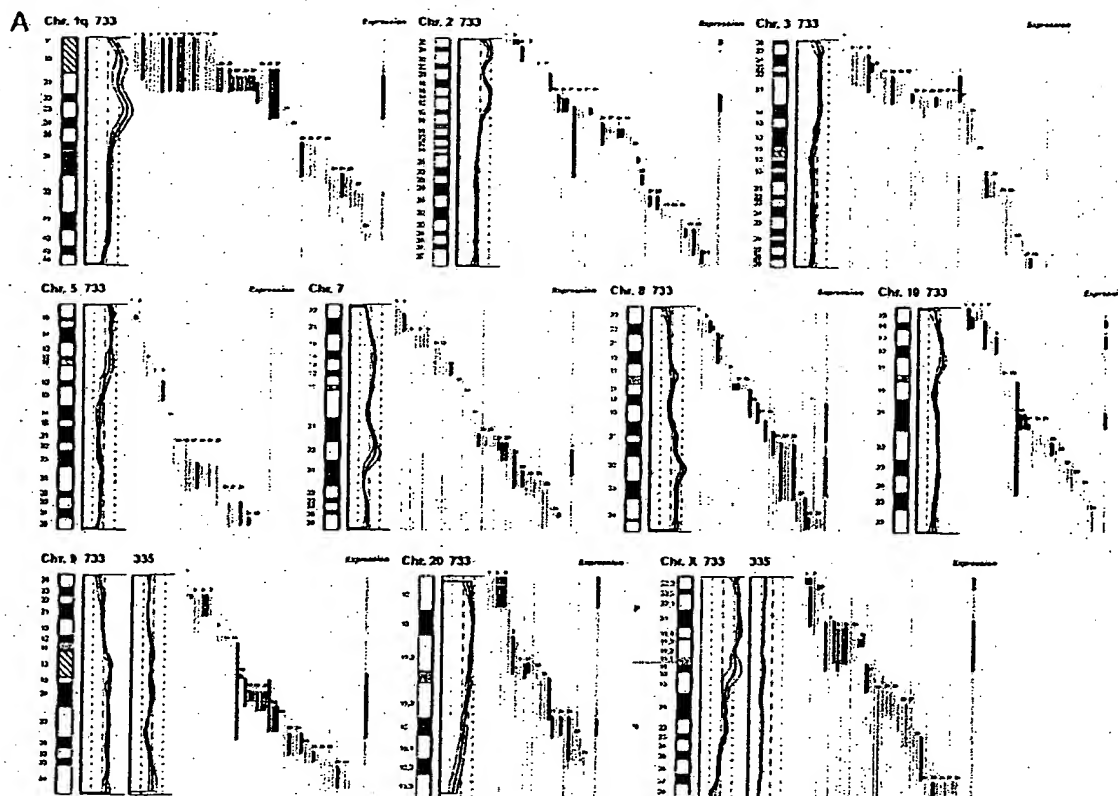


Fig. 1. DNA copy number and mRNA expression level. Shown from left to right are chromosome (Chr.), CGH profiles, gene location and expression level of specific genes, and overall expression level along the chromosome. **A**, expression of mRNA in invasive tumor 733 as compared with the non-invasive counterpart tumor 335. **B**, expression of mRNA in invasive tumor 827 compared with the non-invasive counterpart tumor 532. The average fluorescent signal ratio between tumor DNA and normal DNA is shown along the length of the chromosome (left). The bold curve in the ratio profile represents a mean of four chromosomes and is surrounded by thin curves indicating one standard deviation. The central vertical line (broken) indicates a ratio value of 1 (no change), and the vertical lines next to it (dotted) indicate a ratio of 0.5 (left) and 2.0 (right). In chromosomes where the non-invasive tumor 335 used for comparison showed alterations in DNA content, the ratio profile of that chromosome is shown to the right of the invasive tumor profile. The colored bars represent one gene each, identified by the running numbers above the bars (the name of the gene can be seen at www.MDL.DK/sdata.html). The bars indicate the purported location of the gene, and the colors indicate the expression level of the gene in the invasive tumor compared with the non-invasive counterpart; >2-fold increase (black), >2-fold decrease (blue), no significant change (orange). The bar to the far right, entitled *Expression* shows the resulting change in expression along the chromosome; the colors indicate that at least half of the genes were up-regulated (black), at least half of the genes down-regulated (blue), or more than half of the genes are unchanged (orange). If a gene was absent in one of the samples and present in another, it was regarded as more than a 2-fold change. A 2-fold level was chosen as this corresponded to one standard deviation in a double determination of ~1800 genes. Centromeres and heterochromatic regions were excluded from data analysis.

grade I and II, respectively, tumors 733 and 827 were staged as pT1 (invasive into submucosa), 733 was staged as solid, and 827 was staged as papillary, both grade III.

mRNA Preparation—Tissue biopsies, obtained fresh from surgery, were embedded immediately in a sodium-guanidinium thiocyanate solution and stored at -80°C . Total RNA was isolated using the RNeasy B RNA isolation method (WAK-Chemie Medical GmbH). poly(A)⁺ RNA was isolated by an oligo(dT) selection step (Oligotex mRNA kit; Qiagen).

cRNA Preparation—1 μg of mRNA was used as starting material. The first and second strand cDNA synthesis was performed using the SuperScript[®] choice system (Invitrogen) according to the manufacturer's instructions but using an oligo(dT) primer containing a T7 RNA polymerase binding site. Labeled cRNA was prepared using the ME-GAAscrip[®] *in vitro* transcription kit (Ambion). Biotin-labeled CTP and

UTP (Enzo) was used, together with unlabeled NTPs in the reaction. Following the *in vitro* transcription reaction, the unincorporated nucleotides were removed using RNeasy columns (Qiagen).

Array Hybridization and Scanning—Array hybridization and scanning was modified from a previous method (13). 10 μg of cRNA was fragmented at 94°C for 35 min in buffer containing 40 mM Tris acetate, pH 8.1, 100 mM KOAc, 30 mM MgOAc. Prior to hybridization, the fragmented cRNA in a 6 \times SSPE-T hybridization buffer (1 M NaCl, 10 mM Tris, pH 7.6, 0.005% Triton), was heated to 95°C for 5 min, subsequently cooled to 40°C , and loaded onto the Affymetrix probe array cartridge. The probe array was then incubated for 16 h at 40°C at constant rotation (60 rpm). The probe array was exposed to 10 washes in 6 \times SSPE-T at 25°C followed by 4 washes in 0.5 \times SSPE-T at 50°C . The biotinylated cRNA was stained with a streptavidin-phycoerythrin conjugate, 10 $\mu\text{g}/\text{ml}$ (Molecular Probes) in 6 \times SSPE-T

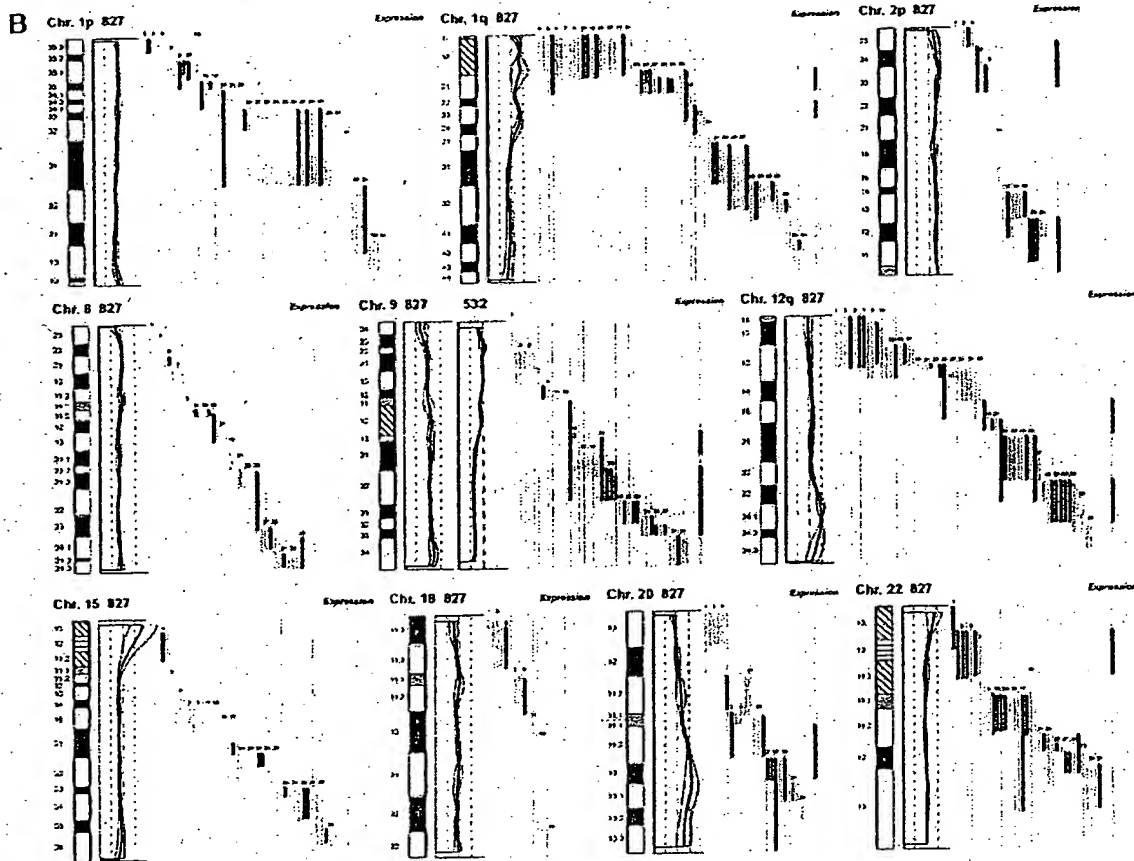


FIG. 1—continued

for 30 min at 25 °C followed by 10 washes in 6× SSPE-T at 25 °C. The probe arrays were scanned at 560 nm using a confocal laser scanning microscope (made for Affymetrix by Hewlett-Packard). The readings from the quantitative scanning were analyzed by Affymetrix gene expression analysis software.

Microsatellite Analysis—Microsatellite Analysis was performed as described previously (14). Microsatellites were selected by use of www.ncbi.nlm.nih.gov/genemap98, and primer sequences were obtained from the genome data base at www.gdb.org. DNA was extracted from tumor and blood and amplified by PCR in a volume of 20 μ l for 35 cycles. The amplicons were denatured and electrophoresed for 3 h in an ABI Prism 377. Data were collected in the Gene Scan program for fragment analysis. Loss of heterozygosity was defined as less than 33% of one allele detected in tumor amplicons compared with blood.

Proteomic Analysis—TCCs were minced into small pieces and homogenized in a small glass homogenizer in 0.5 ml of lysis solution. Samples were stored at -20 °C until use. The procedure for 2D gel electrophoresis has been described in detail elsewhere (15, 16). Gels were stained with silver nitrate and/or Coomassie Brilliant Blue. Proteins were identified by a combination of procedures that included microsequencing, mass spectrometry, two-dimensional gel Western immunoblotting, and comparison with the master two-dimensional gel image of human keratinocyte proteins; see biobase.dk/cgi-bin/celis.

CGH—Hybridization of differentially labeled tumor and normal DNA to normal metaphase chromosomes was performed as described previously (10). Fluorescein-labeled tumor DNA (200 ng), Texas Red-

labeled reference DNA (200 ng), and human Cot-1 DNA (20 μ g) were denatured at 37 °C for 5 min and applied to denatured normal metaphase slides. Hybridization was at 37 °C for 2 days. After washing, the slides were counterstained with 0.15 μ g/ml 4,6-diamidino-2-phenylindole in an anti-fade solution. A second hybridization was performed for all tumor samples using fluorescein-labeled reference DNA and Texas Red-labeled tumor DNA (inverse labeling) to confirm the aberrations detected during the initial hybridization. Each CGH experiment also included a normal control hybridization using fluorescein- and Texas Red-labeled normal DNA. Digital image analysis was used to identify chromosomal regions with abnormal fluorescence ratios, indicating regions of DNA gains and losses. The average green:red fluorescence intensity ratio profiles were calculated using four images of each chromosome (eight chromosomes total) with normalization of the green:red fluorescence intensity ratio for the entire metaphase and background correction. Chromosome identification was performed based on 4,6-diamidino-2-phenylindole banding patterns. Only images showing uniform high intensity fluorescence with minimal background staining were analyzed. All centromeres, p arms of acrocentric chromosomes, and heterochromatic regions were excluded from the analysis.

RESULTS

Comparative Genomic Hybridization—The CGH analysis identified a number of chromosomal gains and losses in the

Gene Copy Numbers, Transcripts, and Protein Levels

TABLE I
Correlation between alterations detected by CGH and by expression monitoring

Top, CGH used as independent variable (if CGH alteration – what expression ratio was found); bottom, altered expression used as independent variable (if expression alteration – what CGH deviation was found).

| CGH alterations | Tumor 733 vs. 335 | | Concordance | CGH alterations | Tumor 827 vs. 532 | |
|----------------------------|--|-----|-------------|----------------------------|---|-----|
| | Expression change clusters | | | | Expression change clusters | |
| 13 Gain | 10 Up-regulation 0 Down-regulation 3 No change | 77% | | 10 Gain | 8 Up-regulation 0 Down-regulation 2 No change | 80% |
| 10 Loss | 1 Up-regulation 5 Down-regulation 4 No change | 50% | | 12 Loss | 3 Up-regulation 2 Down regulation 7 No change | 17% |
| Expression change clusters | Tumor 733 vs. 335 | | Concordance | Expression change clusters | Tumor 827 vs. 532 | |
| | CGH alterations | | | | CGH alterations | |
| 16 Up-regulation | 11 Gain 2 Loss 3 No change | 69% | | 17 Up-regulation | 10 Gain 5 Loss 2 No change | 59% |
| 21 Down-regulation | 1 Gain 8 Loss 12 No change | 38% | | 9 Down-regulation | 0 Gain 3 Loss 6 No change | 33% |
| 15 No change | 3 Gain 3 Loss 9 No change | 60% | | 21 No change | 1 Gain 3 Loss 17 No change | 81% |

two invasive tumors (stage pT1, TCCs 733 and 827), whereas the two non-invasive papillomas (stage pTa, TCCs 335 and 532) showed only 9p-, 9q22-q33-, and X-, and 7+, 9q-, and Y-, respectively. Both invasive tumors showed changes (1q22-24+, 2q14.1-qter-, 3q12-q13.3-, 6q12-q22-, 9q34+, 11q12-q13+, 17+, and 20q11.2-q12+) that are typical for their disease stage, as well as additional alterations, some of which are shown in Fig. 1. Areas with gains and losses deviated from the normal copy number to some extent, and the average numerical deviation from normal was 0.4-fold in the case of TCC 733 and 0.3-fold for TCC 827. The largest changes, amounting to at least a doubling of chromosomal content, were observed at 1q23 in TCC 733 (Fig. 1A) and 20q12 in TCC 827 (Fig. 1B).

mRNA Expression in Relation to DNA Copy Number—The mRNA levels from the two invasive tumors (TCCs 827 and 733) were compared with the two non-invasive counterparts (TCCs 532 and 335). This was done in two separate experiments in which we compared TCCs 733 to 335 and 827 to 532, respectively, using two different scaling settings for the arrays to rule out scaling as a confounding parameter. Approximately 1,800 genes that yielded a signal on the arrays were searched in the Unigene and Genemap data bases for chromosomal location, and those with a known location (1096) were plotted as bars covering their purported locus. In that way it was possible to construct a graphic presentation of DNA copy number and relative mRNA levels along the individual chromosomes (Fig. 1).

For each mRNA a ratio was calculated between the level in the invasive versus the non-invasive counterpart. Bars, which represent chromosomal location of a gene, were color-coded according to the expression ratio, and only differences larger

than 2-fold were regarded as informative (Fig. 1). The density of genes along the chromosomes varied, and areas containing only one gene were excluded from the calculations. The resolution of the CGH method is very low, and some of the outlier data may be because of the fact that the boundaries of the chromosomal aberrations are not known at high resolution.

Two sets of calculations were made from the data. For the first set we used CGH alterations as the independent variable and estimated the frequency of expression alterations in these chromosomal areas. In general, areas with a strong gain of chromosomal material contained a cluster of genes having increased mRNA expression. For example, both chromosomes 1q21-q25, 2p and 9q, showed a relative gain of more than 100% in DNA copy number that was accompanied by increased mRNA expression levels in the two tumor pairs (Fig. 1). In most cases, chromosomal gains detected by CGH were accompanied by an increased level of transcripts in both TCCs 733 (77%) and 827 (80%) (Table I, top). Chromosomal losses, on the other hand, were not accompanied by decreased expression in several cases, and were often registered as having unaltered RNA levels (Table I, top). The inability to detect RNA expression changes in these cases was not because of fewer genes mapping to the lost regions (data not shown).

In the second set of calculations we selected expression alterations above 2-fold as the independent variable and estimated the frequency of CGH alterations in these areas. As above, we found that increased transcript expression correlated with gain of chromosomal material (TCC 733, 69% and TCC 827, 59%), whereas reduced expression was often detected in areas with unaltered CGH ratios (Table I, bottom). Furthermore, as a control we looked at areas with no alter-

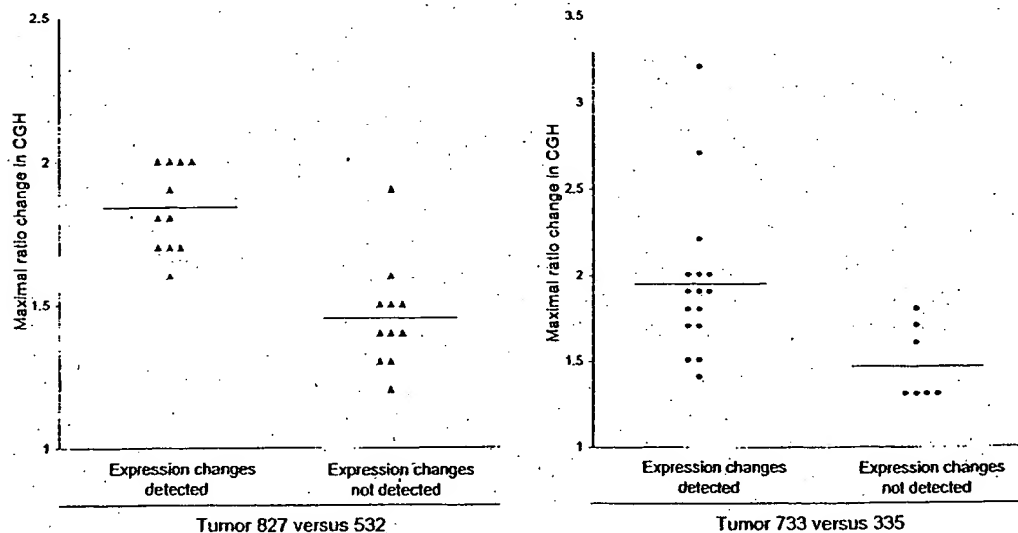


FIG. 2. Correlation between maximum CGH aberration and the ability to detect expression change by oligonucleotide array monitoring. The aberration is shown as a numerical -fold change in ratio between invasive tumors 827 (▲) and 733 (◆) and their non-invasive counterparts 532 and 335. The expression change was taken from the Expression line to the right in Fig. 1, which depicts the resulting expression change for a given chromosomal region. At least half of the mRNAs from a given region have to be either up- or down-regulated to be scored as an expression change. All chromosomal arms in which the CGH ratio plus or minus one standard deviation was outside the ratio value of one were included.

ation in expression. No alteration was detected by CGH in most of these areas (TCC 733, 60% and TCC 827, 81%; see Table I, bottom). Because the ability to observe reduced or increased mRNA expression clustering to a certain chromosomal area clearly reflected the extent of copy number changes, we plotted the maximum CGH aberrations in the regions showing CGH changes against the ability to detect a change in mRNA expression as monitored by the oligonucleotide arrays (Fig. 2). For both tumors TCC 733 ($p < 0.015$) and TCC 827 ($p < 0.00003$) a highly significant correlation was observed between the level of CGH ratio change (reflecting the DNA copy number) and alterations detected by the array based technology (Fig. 2). Similar data were obtained when areas with altered expression were used as independent variables. These areas correlated best with CGH when the CGH ratio deviated 1.6- to 2.0-fold (Table I, bottom) but mostly did not at lower CGH deviations. These data probably reflect that loss of an allele may only lead to a 50% reduction in expression level, which is at the cut-off point for detection of expression alterations. Gain of chromosomal material can occur to a much larger extent.

Microsatellite-based Detection of Minor Areas of Losses—In TCC 733, several chromosomal areas exhibiting DNA amplification were preceded or followed by areas with a normal CGH but reduced mRNA expression (see Fig. 1, TCC 733 chromosome 1q32, 2p21, and 7q21 and q32, 9q34, and 10q22). To determine whether these results were because of undetected loss of chromosomal material in these regions or

because of other non-structural mechanisms regulating transcription, we examined two microsatellites positioned at chromosome 1q25–32 and two at chromosome 2p22. Loss of heterozygosity (LOH) was found at both 1q25 and at 2p22 indicating that minor deleted areas were not detected with the resolution of CGH (Fig. 3). Additionally, chromosome 2p in TCC 733 showed a CGH pattern of gain/no change/gain of DNA that correlated with transcript increase/decrease/increase. Thus, for the areas showing increased expression there was a correlation with the DNA copy number alterations (Fig. 1A). As indicated above, the mRNA decrease observed in the middle of the chromosomal gain was because of LOH, implying that one of the mechanisms for mRNA down-regulation may be regions that have undergone smaller losses of chromosomal material. However, this cannot be detected with the resolution of the CGH method.

In both TCC 733 and TCC 827, the telomeric end of chromosome 11p showed a normal ratio in the CGH analysis; however, clusters of five and three genes, respectively, lost their expression. Two microsatellites (D11S1760, D11S922) positioned close to MUC2, IGF2, and cathepsin D indicated LOH as the most likely mechanism behind the loss of expression (data not shown).

A reduced expression of mRNA observed in TCC 733 at chromosomes 3q24, 11p11, 12p12.2, 12q21.1, and 16q24 and in TCC 827 at chromosome 11p15.5, 12p11, 15q11.2, and 18q12 was also examined for chromosomal losses using microsatellites positioned as close as possible to the gene loci

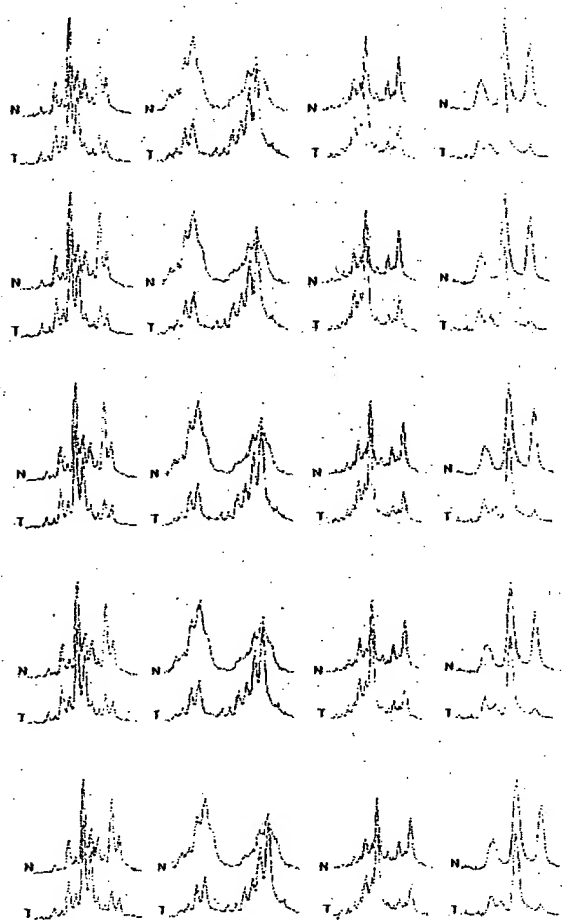


FIG. 3. Microsatellite analysis of loss of heterozygosity. Tumor 733 showing loss of heterozygosity at chromosome 1q25, detected (a) by D1S215 close to Hu class I histocompatibility antigen (gene number 38 in Fig. 1), (b) by D1S2735 close to cathepsin E (gene number 41 in Fig. 1), and (c) at chromosome 2p23 by D2S2251 close to general β -spectrin (gene number 11 on Fig. 1) and of (d) tumor 827 showing loss of heterozygosity at chromosome 18q12 by S18S1118 close to mitochondrial 3-oxoacyl-coenzyme A thiolase (gene number 12 in Fig. 1). The upper curves show the electropherogram obtained from normal DNA from leukocytes (N), and the lower curves show the electropherogram from tumor DNA (T). In all cases one allele is partially lost in the tumor amplicon.

showing reduced mRNA transcripts. Only the microsatellite positioned at 18q12 showed LOH (Fig. 3), suggesting that transcriptional down-regulation of genes in the other regions may be controlled by other mechanisms.

Relation between Changes in mRNA and Protein Levels—2D-PAGE analysis, in combination with Coomassie Brilliant Blue and/or silver staining, was carried out on all four tumors using fresh biopsy material. 40 well resolved abundant known proteins migrating in areas away from the edges of the pH

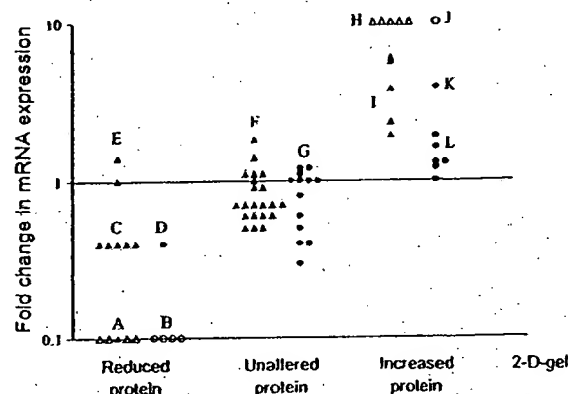


FIG. 4. Correlation between protein levels as judged by 2D-PAGE and transcript ratio. For comparison proteins were divided in three groups, unaltered in level or up- or down-regulated (horizontal axis). The mRNA ratio as determined by oligonucleotide arrays was plotted for each gene (vertical axis). Δ , mRNAs that were scored as present in both tumors used for the ratio calculation; Δ , mRNAs that were scored as absent in the invasive tumors (along horizontal axis) or as absent in non-invasive reference (top of figure). Two different scalings were used to exclude scaling as a confounder, TCCs 827 and 532 ($\Delta\Delta$) were scaled with background suppression, and TCCs 733 and 335 ($\bullet\bullet$) were scaled without suppression. Both comparisons showed highly significant ($p < 0.005$) differences in mRNA ratios between the groups. Proteins shown were as follows: Group A (from left), phosphoglucomutase 1, glutathione transferase class μ number 4, fatty acid-binding protein homologue, cytochrome 15, and cytochrome 13; B (from left), fatty acid-binding protein homologue, 28-kDa heat shock protein, cytochrome 13, and calnexin; C (from left), α -enolase, hnRNP B1, 28-kDa heat shock protein, 14-3-3- ϵ , and pre-mRNA splicing factor; D, mesothelial keratin K7 (type II); E (from top), glutathione S-transferase- π and mesothelial keratin K7 (type II); F (from top and left), adenylate cyclase-associated protein, E-cadherin, keratin 19, calgizzarin, phosphoglycerate mutase, annexin IV, cytoskeletal γ -actin, hnRNP A1, integral membrane protein calnexin (IP90), hnRNP H, brain-type clathrin light chain-a, hnRNP F, 70-kDa heat shock protein, heterogeneous nuclear ribonucleoprotein A/B, translationally controlled tumor protein, liver glyceraldehyde-3-phosphate dehydrogenase, keratin 8, aldehyde reductase, and Na,K-ATPase β -1 subunit; G, (from top and left), TCP20, calgizzarin, 70-kDa heat shock protein, calnexin, hnRNP H, cytochrome 15, ATP synthase, keratin 19, triosephosphate isomerase, hnRNP F, liver glyceraldehyde-3-phosphate dehydrogenase, glutathione S-transferase- π , and keratin 8; H (from left), plasma gelsolin, autoantigen calreticulin, thioredoxin, and NAD $^{+}$ -dependent 15-hydroxyprostaglandin dehydrogenase; I (from top), prolidyl 4-hydroxylase β -subunit, cytochrome 20, cytochrome 17, prothionin, and fructose 1,6-bisphosphatase; J, annexin II; K, annexin IV; L (from top and left), 90-kDa heat shock protein, prolidyl 4-hydroxylase β -subunit, α -enolase, GRP 78, cyclophilin, and cofillin.

gradient, and having a known chromosomal location, were selected for analysis in the TCC pair 827/532. Proteins were identified by a combination of methods (see "Experimental Procedures"). In general there was a highly significant correlation ($p < 0.005$) between mRNA and protein alterations (Fig. 4). Only one gene showed disagreement between transcript alteration and protein alteration. Except for a group of cyto-

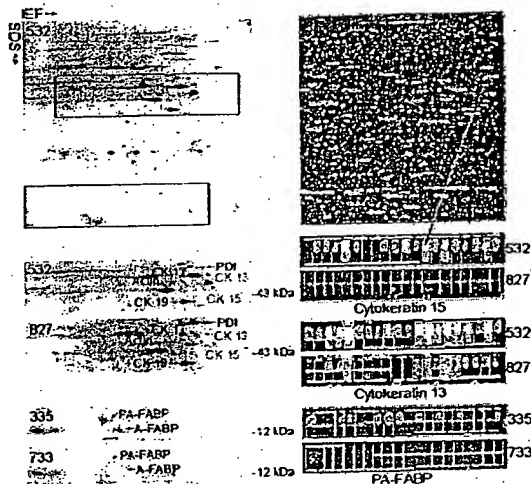


FIG. 5. Comparison of protein and transcript levels in invasive and non-invasive TCCs. The upper part of the figure shows a 2D gel (left) and the oligonucleotide array (right) of TCC 532. The red rectangles on the upper gel highlight the areas that are compared below. Identical areas of 2D gels of TCCs 532 and 827 are shown below. Clearly, cytokeratins 13 and 15 are strongly down-regulated in TCC 827 (red annotation). The tile on the array containing probes for cytokeratin 15 is enlarged below the array (red arrow) from TCC 532 and is compared with TCC 827. The upper row of squares in each tile corresponds to perfect match probes; the lower row corresponds to mismatch probes containing a mutation (used for correction for unspecific binding). Absence of signal is depicted as black, and the higher the signal the lighter the color. A high transcript level was detected in TCC 532 (6151 units) whereas a much lower level was detected in TCC 827 (absence of signals). For cytokeratin 13, a high transcript level was also present in TCC 532 (15659 units), and a much lower level was present in TCC 827 (623 units). The 2D gels at the bottom of the figure (left) show levels of PA-FABP and adipocyte-FABP in TCCs 335 and 733 (invasive); respectively. Both proteins are down-regulated in the invasive tumor. To the right we show the array tiles for the PA-FABP transcript. A medium transcript level was detected in the case of TCC 335 (1277 units) whereas very low levels were detected in TCC 733 (166 units). IEF, isoelectric focusing.

keratins encoded by genes on chromosome 17 (Fig. 5) the analyzed proteins did not belong to a particular family. 26 well focused proteins whose genes had a known chromosomal location were detected in TCCs 733 and 335, and of these 19 correlated ($p < 0.005$) with the mRNA changes detected using the arrays (Fig. 4). For example, PA-FABP was highly expressed in the non-invasive TCC 335 but lost in the invasive counterpart (TCC 733; see Fig. 5). The smaller number of proteins detected in both 733 and 335 was because of the smaller size of the biopsies that were available.

11 chromosomal regions where CGH showed aberrations that corresponded to the changes in transcript levels also showed corresponding changes in the protein level (Table II). These regions included genes that encode proteins that are found to be frequently altered in bladder cancer, namely cytokeratins 17 and 20, annexins II and IV, and the fatty acid-binding proteins PA-FABP and FBP1. Four of these proteins were encoded by genes in chromosome 17q, a frequently amplified chromosomal area in invasive bladder cancers.

DISCUSSION

Most human cancers have abnormal DNA content, having lost some chromosomal parts and gained others. The present study provides some evidence as to the effect of these gains and losses on gene expression in two pairs of non-invasive and invasive TCCs using high throughput expression arrays and proteomics, in combination with CGH. In general, the results showed that there is a clear individual regulation of the mRNA expression of single genes, which in some cases was superimposed by a DNA copy number effect. In most cases, genes located in chromosomal areas with gains often exhibited increased mRNA expression, whereas areas showing losses showed either no change or a reduced mRNA expression. The latter might be because of the fact that losses most often are restricted to loss of one allele, and the cut-off point for detection of expression alterations was a 2-fold change, thus being at the border of detection. In several cases, how-

TABLE II
Proteins whose expression level correlates with both mRNA and gene dose changes

| Protein | Chromosomal location | Tumor TCC | CGH alteration | Transcript alteration ^a | Protein alteration |
|-----------------------|----------------------|-----------|----------------|------------------------------------|--------------------|
| Annexin II | 1q21 | 733 | Gain | Abs to Pres ^a | Increase |
| Annexin IV | 2p13 | 733 | Gain | 3.9-Fold up | Increase |
| Cytokeratin 17 | 17q12-q21 | 827 | Gain | 3.8-Fold up | Increase |
| Cytokeratin 20 | 17q21.1 | 827 | Gain | 5.6-Fold up | Increase |
| (PA-)FABP | 8q21.2 | 827 | Loss | 10-Fold down | Decrease |
| FBP1 | 9q22 | 827 | Gain | 2.3-Fold up | Increase |
| Plasma gelsolin | 9q31 | 827 | Gain | Abs to Pres | Increase |
| Heat shock protein 28 | 15q12-q13 | 827 | Loss | 2.5-Fold up | Decrease |
| Prohibitin | 17q21 | 827/733 | Gain | 3.7-/2.5-Fold up ^b | Increase |
| Prolyl-4-hydroxyl | 17q25 | 827/733 | Gain | 5.7-/1.6-Fold up | Increase |
| hnRNBP1 | 7p15 | 827 | Loss | 2.5-Fold down | Decrease |

^a Abs, absent; Pres, present.

^b In cases where the corresponding alterations were found in both TCCs 827 and 733 these are shown as 827/733.

ever, an increase or decrease in DNA copy number was associated with *de novo* occurrence or complete loss of transcript, respectively. Some of these transcripts could not be detected in the non-invasive tumor but were present at relatively high levels in areas with DNA amplifications in the invasive tumors (e.g. in TCC 733 transcript from cellular ligand of annexin II gene (chromosome 1q21) from absent to 2670 arbitrary units; in TCC 827 transcript from small proline-rich protein 1 gene (chromosome 1q12-q21.1) from absent to 1326 arbitrary units). It may be anticipated from these data that significant clustering of genes with an increased expression to a certain chromosomal area indicates an increased likelihood of gain of chromosomal material in this area.

Considering the many possible regulatory mechanisms acting at the level of transcription, it seems striking that the gene dose effects were so clearly detectable in gained areas. One hypothetical explanation may lie in the loss of controlled methylation in tumor cells (17-19). Thus, it may be possible that in chromosomes with increased DNA copy numbers two or more alleles could be demethylated simultaneously leading to a higher transcription level, whereas in chromosomes with losses the remaining allele could be partly methylated, turning off the process (20, 21). A recent report has documented a ploidy regulation of gene expression in yeast, but in this case all the genes were present in the same ratio (22), a situation that is not analogous to that of cancer cells, which show marked chromosomal aberrations, as well as gene dosage effects.

Several CGH studies of bladder cancer have shown that some chromosomal aberrations are common at certain stages of disease progression, often occurring in more than 1 of 3 tumors. In pTa tumors, these include 9p-, 9q-, 1q+, Y- (2, 6), and in pT1 tumors, 2q-, 11p-, 11q-, 1q+, 5p+, 8q+, 17q+, and 20q+ (2-4, 6, 7). The pTa tumors studied here showed similar aberrations such as 9p- and 9q22-q33- and 9q- and Y-, respectively. Likewise, the two minimal invasive pT1 tumors showed aberrations that are commonly seen at that stage, and TCC 827 had a remarkable resemblance to the commonly seen pattern of losses and gains, such as 1q22-24 amplification (seen in both tumors), 11q14-q22 loss, the latter often linked to 17 q+ (both tumors), and 1q+ and 9p-, often linked to 20q+ and 11 q13+ (both tumors) (7-9). These observations indicate that the pairs of tumors used in this study exhibit chromosomal changes observed in many tumors, and therefore the findings could be of general importance for bladder cancer.

Considering that the mapping resolution of CGH is of about 20 megabases it is only possible to get a crude picture of chromosomal instability using this technique. Occasionally, we observed reduced transcript levels close to or inside regions with increased copy numbers. Analysis of these regions by positioning heterozygous microsatellites as close as possible to the locus showing reduced gene expression revealed loss of heterozygosity in several cases. It seems likely that multiple and different events occur along each chromosomal

arm and that the use of cDNA microarrays for analysis of DNA copy number changes will reach a resolution that can resolve these changes, as has recently been proposed (2). The outlier data were not more frequent at the boundaries of the CGH aberrations. At present we do not know the mechanism behind chromosomal aneuploidy and cannot predict whether chromosomal gains will be transcribed to a larger extent than the two native alleles. A mechanism as genetic imprinting has an impact on the expression level in normal cells and is often reduced in tumors. However, the relation between imprinting and gain of chromosomal material is not known.

We regard it as a strength of this investigation that we were able to compare invasive tumors to benign tumors rather than to normal urothelium, as the tumors studied were biologically very close and probably may represent successive steps in the progression of bladder cancer. Despite the limited amount of fresh tissue available it was possible to apply three different state of the art methods. The observed correlation between DNA copy number and mRNA expression is remarkable when one considers that different pieces of the tumor biopsies were used for the different sets of experiments. This indicate that bladder tumors are relatively homogenous, a notion recently supported by CGH and LOH data that showed a remarkable similarity even between tumors and distant metastasis (10, 23).

In the few cases analyzed, mRNA and protein levels showed a striking correspondence although in some cases we found discrepancies that may be attributed to translational regulation, post-translational processing, protein degradation, or a combination of these. Some transcripts belong to undertranslated mRNA pools, which are associated with few translationally inactive ribosomes; these pools, however, seem to be rare (24). Protein degradation, for example, may be very important in the case of polypeptides with a short half-life (e.g. signaling proteins). A poor correlation between mRNA and protein levels was found in liver cells as determined by arrays and 2D-PAGE (25), and a moderate correlation was recently reported by Ideker *et al.* (26) in yeast.

Interestingly, our study revealed a much better correlation between gained chromosomal areas and increased mRNA levels than between loss of chromosomal areas and reduced mRNA levels. In general, the level of CGH change determined the ability to detect a change in transcript. One possible explanation could be that by losing one allele the change in mRNA level is not so dramatic as compared with gain of material, which can be rather unlimited and may lead to a severalfold increase in gene copy number resulting in a much higher impact on transcript level. The latter would be much easier to detect on the expression arrays as the cut-off point was placed at a 2-fold level so as not to be biased by noise on the array. Construction of arrays with a better signal to noise ratio may in the future allow detection of lesser than 2-fold alterations in transcript levels, a feature that may facilitate the analysis of the effect of loss of chromosomal areas on transcript levels.

In eleven cases we found a significant correlation between DNA copy number, mRNA expression, and protein level. Four of these proteins were encoded by genes located at a frequently amplified area in chromosome 17q. Whether DNA copy number is one of the mechanisms behind alteration of these eleven proteins is at present unknown and will have to be proved by other methods using a larger number of samples. One factor making such studies complicated is the large extent of protein modification that occurs after translation, requiring immunoidentification and/or mass spectrometry to correctly identify the proteins in the gels.

In conclusion, the results presented in this study exemplify the large body of knowledge that may be possible to gather in the future by combining state of the art techniques that follow the pathway from DNA to protein (26). Here, we used a traditional chromosomal CGH method, but in the future high resolution CGH based on microarrays with many thousand radiation hybrid-mapped genes will increase the resolution and information derived from these types of experiments (2). Combined with expression arrays analyzing transcripts derived from genes with known locations, and 2D gel analysis to obtain information at the post-translational level, a clearer and more developed understanding of the tumor genome will be forthcoming.

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§ To whom correspondence should be addressed: Dept. of Clinical Biochemistry, Molecular Diagnostic Laboratory, Aarhus University Hospital, Skejby, DK-8200 Aarhus N, Denmark. Tel.: 45-89495100/45-86156201 (private); Fax: 45-89496018; E-mail: orntoft@kba.sks.au.dk.

REFERENCES

- Lengauer, C., Kinzler, K. W., and Vogelstein, B. (1998) Genetic instabilities in human cancers. *Nature* 396, 643-649.
- Pollack, J. R., Perou, C. M., Alizadeh, A. A., Eisen, M. B., Pergamenschikov, A., Williams, C. F., Jeffrey, S. S., Botstein, D., and Brown, P. O. (1999) Genome-wide analysis of DNA copy-number changes using cDNA microarrays. *Nat. Genet.* 23, 41-46.
- de Cremoux, P., Martin, E. C., Vincent-Salomon, A., Dieras, V., Barbaroux, C., Liva, S., Pouillart, P., Sastre-Garau, X., and Magdelenat, H. (1999) Quantitative PCR analysis of c-erb B-2 (HER2/neu) gene amplification and comparison with p185(HER2/neu) protein expression in breast cancer drill biopsies. *Int. J. Cancer* 83, 157-161.
- Brugier, P. P., Tamimi, Y., Shuring, E., and Schalken, J. (1996) Expression of cyclin D1 and EMS1 in bladder tumors; relationship with chromosome 11q13 amplifications. *Oncogene* 12, 1747-1753.
- Slavc, I., Ellenbogen, R., Jung, W. H., Vawter, G. F., Kretschmar, C., Grier, H., and Korl, B. R. (1990) *myc* gene amplification and expression in primary human neuroblastoma. *Cancer Res.* 50, 1459-1463.
- Sauter, G., Carroll, P., Moch, H., Kallioniemi, A., Kerschmann, R., Narayan, P., Mihatsch, M. J., and Waldman, F. M. (1995) *c-myc* copy number gains in bladder cancer detected by fluorescence *in situ* hybridization. *Am. J. Pathol.* 146, 1131-1139.
- Richter, J., Jiang, F., Gorog, J. P., Sartorius, G., Egenter, C., Gasser, T. C., Moch, H., Mihatsch, M. J., and Sauter, G. (1997) Marked genetic differences between stage pTa and stage pT1 papillary bladder cancer detected by comparative genomic hybridization. *Cancer Res.* 57, 2860-2864.
- Richter, J., Belfa, L., Wagner, U., Schraml, P., Gasser, T. C., Moch, H., Mihatsch, M. J., and Sauter, G. (1998) Patterns of chromosomal imbalances in advanced urinary bladder cancer detected by comparative genomic hybridization. *Am. J. Pathol.* 153, 1615-1621.
- Bruch, J., Wöhr, G., Hautmann, R., Mattfeldt, T., Bruderlein, S., Moller, P., Sauter, S., Hameister, H., Vogel, W., and Poiss, T. (1998) Chromosomal changes during progression of transitional cell carcinoma of the bladder and delineation of the amplified interval on chromosome arm 8q. *Genes Chromosomes Cancer* 23, 167-174.
- Hovey, R. M., Chu, L., Balazs, M., De Vries, S., Moore, D., Sauter, G., Carroll, P. R., and Waldman, F. M. (1998) Genetic alterations in primary bladder cancers and their metastases. *Cancer Res.* 58, 3555-3560.
- Simon, R., Burger, H., Brinkschmidt, C., Bocker, W., Hertle, L., and Törpe, H. J. (1998) Chromosomal aberrations associated with invasion in papillary superficial bladder cancer. *J. Pathol.* 185, 345-351.
- Koo, S. H., Kwon, K. C., Ihm, C. H., Jeon, Y. M., Park, J. W., and Sul, C. K. (1999) Detection of genetic alterations in bladder tumors by comparative genomic hybridization and cytogenetic analysis. *Cancer Genet. Cytogenet.* 110, 87-93.
- Wodicka, L., Dong, H., Mittmann, M., Ho, M. H., and Lockhart, D. J. (1997) Genome-wide expression monitoring in *Saccharomyces cerevisiae*. *Nat. Biotechnol.* 15, 1359-1367.
- Christensen, M., Sunde, L., Bolund, L., and Ørntoft, T. F. (1999) Comparison of three methods of microsatellite detection. *Scand. J. Clin. Lab. Invest.* 59, 167-177.
- Celis, J. E., Østergaard, M., Basse, B., Celis, A., Lauridsen, J. B., Ratz, G. P., Andersen, I., Hein, B., Wolf, H., Ørntoft, T. F., and Rasmussen, H. H. (1996) Loss of adipocyte-type fatty acid binding protein and other protein biomarkers is associated with progression of human bladder transitional cell carcinomas. *Cancer Res.* 56, 4782-4790.
- Celis, J. E., Ratz, G., Basse, B., Lauridsen, J. B., and Celis, A. (1994) In *Cell Biology: A Laboratory Handbook* (Celis, J. E., ed) Vol. 3, pp. 222-230. Academic Press, Orlando, FL.
- Ohlsson, R., Tycko, B., and Sapienza, C. (1998) Monoallelic expression: 'there can only be one'. *Trends Genet.* 14, 435-438.
- Hollander, G. A., Zuklys, S., Morel, C., Mizoguchi, E., Mobisson, K., Simpson, S., Terhorst, C., Wishart, W., Golan, D. E., Bhan, A. K., and Burakoff, S. J. (1998) Monoallelic expression of the interleukin-2 locus. *Science* 279, 2118-2121.
- Brannan, C. I., and Bartolomei, M. S. (1999) Mechanisms of genomic imprinting. *Curr. Opin. Genet. Dev.* 9, 164-170.
- Ohlsson, R., Cui, H., He, L., Pfeifer, S., Malmikumpu, H., Jiang, S., Feinberg, A. P., and Hedborg, F. (1999) Mosaic allelic insulin-like growth factor 2 expression patterns reveal a link between Wilms' tumorigenesis and epigenetic heterogeneity. *Cancer Res.* 59, 3889-3892.
- Cui, H., Hedborg, F., He, L., Nordenskjöld, A., Sandstedt, B., Pfeifer, Ohlsson, S., and Ohlsson, R. (1997) Inactivation of H19, an imprinted and putative tumor repressor gene, is a preneoplastic event during Wilms' tumorigenesis. *Cancer Res.* 57, 4469-4473.
- Galitski, T., Saldanha, A. J., Styles, C. A., Lander, E. S., and Fink, G. R. (1999) Ploidy regulation of gene expression. *Science* 285, 251-254.
- Tsao, J., Yatabe, Y., Markl, I. D., Hajyan, K., Jones, P. A., and Shibata, D. (2000) Bladder cancer genotype stability during clinical progression. *Genes Chromosomes Cancer* 29, 26-32.
- Zong, Q., Schummer, M., Hood, L., and Morris, D. R. (1999) Messenger RNA translation state: the second dimension of high-throughput expression screening. *Proc. Natl. Acad. Sci. U. S. A.* 96, 10632-10636.
- Anderson, L., and Seltham, J. (1997) Comparison of selected mRNA and protein abundances in human liver. *Electrophoresis* 18, 533-537.
- Ideker, T., Thorsson, V., Raniish, J. A., Christmas, R., Buhler, J., Eng, J. K., Bumgarner, R., Goodlett, D. R., Aebersold, R., and Hood, L. (2001) Integrated genomic and proteomic analyses of a systematically perturbed metabolic network. *Science* 292, 929-934.



Expression of bcr-abl mRNA in individual chronic myelogenous leukaemia cells as determined by in situ amplification.

Pachmann K, Zhao S, Schenk T, Kantarjian H, El-Naggar AK, Siciliano MJ, Guo JQ, Arlinghaus RB, Andreeff M.

The University of Texas M.D. Anderson Cancer Center, Department of Molecular Haematology and Therapy, 1515 Holcombe Boulevard, Houston, TX 77030, USA.

We present the results of a novel method developed for evaluation of in situ amplification, a molecular genetic method at the cellular level. Reverse transcription polymerase chain reaction (RT-PCR) was used to study bcr-abl transcript levels in individual cells from patients with chronic myelogenous leukaemia (CML). After hybridizing a fluorochrome-labelled probe to the cell-bound RT-PCR product, bcr-abl mRNA-positive cells were determined using image analysis. A dilution series of bcr-abl-positive BV173 into normal cells showed a good correlation between expected and actual values. In 25 CML samples, the percentage of in situ PCR-positive cells showed an excellent correlation with cytogenetic results ($r = 0.94$, $P < 0.0001$), interphase fluorescence in situ hybridization (FISH) ($r = 0.95$, $P = 0.001$) and hypermetaphase FISH ($r = 0.81$, $P < 0.001$). The fluorescence intensity was higher in residual CML cells after interferon (IFN) treatment than in newly diagnosed patients ($P = 0.004$), and was highest in late-stage CML resistant to IFN therapy and lowest in CML blast crisis ($P = 0.001$). Mean fluorescence values correlated with bcr-abl protein levels, as determined by Western blot analysis ($r = 0.62$). Laser scanning cytometry allowing automated analysis of large numbers of cells confirmed the results. Thus, fluorescence in situ PCR provides a novel and quantitative approach for monitoring tumour load and bcr-abl transcript levels in CML.

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Cell localization and regulation of expression of cytochrome P450 1A1 and 2B1 in rat lung after induction with 3-methylcholanthrene using mRNA hybridization and immunohistochemistry.

Pairon JC, Trabelsi N, Buard A, Fleury-Feith J, Bachelet CM, Poron F, Beaune P, Brochard P, Laurent P.

INSERM Unite 139, Hopital Henri Mondor, Creteil, France.

In order to characterize the response of various pulmonary cell types to polycyclic aromatic hydrocarbons, the expression of cytochrome P450 (CYP) 1A1 and 2B1 mRNA in the lung of rats, with or without induction by 3-methylcholanthrene (3MC), was analyzed by in situ hybridization using appropriate 35S-labeled riboprobes. The expression of the corresponding proteins was investigated immunohistochemically. Following induction with 3MC, the kinetics of mRNA expression differed considerably between Clara cells and type II pneumocytes and venous endothelial cells. In Clara cells, mRNA expression was detected as early as 1 h after induction, peaked between 2 and 4 h, and was completely undetectable at 14 h. In contrast, venous endothelial cells and type II pneumocytes exhibited permanent mRNA expression of CYP 1A1 in 3MC-pretreated rats. These kinetic results explain the striking absence of correlation between mRNA and protein expression observed in Clara cells 24 h after the end of the induction protocol, as these cells exhibited intense protein expression with no mRNA. In contrast, a good correlation was observed for mRNA and protein expression of CYP 2B1, with similar expressions for Clara cells and type II pneumocytes, but no expression in endothelial cells. This study clearly distinguished the regulation of CYP 1A1 expression in the rat lung from that described in the liver. The differences observed in the various lung cell types, whatever the post-transcriptional mechanisms involved, emphasize that studies must be performed at the cellular level in order to understand the specific response to xenobiotics, not only of this organ as a whole but also of its various anatomic structures.

PMID: 7917307 [PubMed - indexed for MEDLINE]



Correlative immunohistochemical and reverse transcriptase polymerase chain reaction analysis of somatostatin receptor type 2 in neuroendocrine tumors of the lung.

Papotti M, Croce S, Macri L, Funaro A, Pecchioni C, Schindler M, Bussolati G.

Department of Biomedical Sciences and Oncology, University of Turin, Italy.

Somatostatin receptors type 2 (sst2) have been frequently detected in neuroendocrine tumors and bind somatostatin analogues, such as octreotide, with high affinity. Receptor autoradiography, specific mRNA detection and, more recently, antisst2 polyclonal antibodies are currently employed to reveal sst2. The aim of the present study was to investigate by three different techniques the presence of sst2 in a series of 26 neuroendocrine tumors of the lung in which fresh frozen tissue and paraffin sections were available. It was possible, therefore, to compare, in individual cases, RNA analysis studied by reverse transcriptase polymerase chain reaction (RT-PCR), in situ hybridization (ISH), and immunohistochemistry. A series of 20 nonneuroendocrine lung carcinoma samples served as controls. RT-PCR was positive for sst2 in 22 of 26 samples, including 15 of 15 typical carcinoids, 5 of 6 atypical carcinoids, and 2 of 5 small-cell carcinomas. The sst2 mRNA signal obtained by RT-PCR was strong in the majority (87%) of typical carcinoids and of variable intensity in atypical carcinoids and small-cell carcinomas. A weakly positive signal was observed in 5 of 20 control samples. In immunohistochemistry, two different antibodies (anti-sst2) were employed, including a monoclonal antibody, generated in the Department of Pathology, University of Turin. In the majority of samples a good correlation between sst2 mRNA (as detected by RT-PCR) and sst2 protein expression (as detected by immunohistochemistry) was observed. However, one atypical carcinoid and one small-cell carcinoma had focal immunostaining but no RT-PCR signal. ISH performed in selected samples paralleled the results obtained with the other techniques. A low sst2 expression was associated with high grade neuroendocrine tumors and with aggressive behavior. It is concluded that 1) neuroendocrine tumors of the lung express sst2, and there is a correlation between the mRNA amount and the degree of differentiation; 2) immunohistochemistry and ISH are reliable tools to demonstrate sst2 in these tumors; and 3) sst2 identification in tissue sections may provide information on the diagnostic or therapeutic usefulness of somatostatin analogues in individual patients with neuroendocrine tumors.

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Correlative Immunohistochemical and Reverse Transcriptase Polymerase Chain Reaction Analysis of Somatostatin Receptor Type 2 in Neuroendocrine Tumors of the Lung

Mauro Papotti, M.D., Sabrina Croce, M.D., Luigia Macri, M.D.,
Aola Funaro, Ph.D., Carla Pecchioni, Marcus Schindler, M.D., and
Gianni Bussolati, M.D., F.R.C.Path.

Somatostatin receptors type 2 (sst2) have been frequently detected in neuroendocrine tumors and bind somatostatin analogues, such as octreotide, with high affinity. Receptor autoradiography, specific mRNA detection and, more recently, anti-sst2 polyclonal antibodies are currently employed to reveal sst2. The aim of the present study was to investigate by three different techniques the presence of sst2 in a series of 26 neuroendocrine tumors of the lung in which fresh frozen tissue and paraffin sections were available. It was possible, therefore, to compare, in individual cases, RNA analysis studied by reverse transcriptase polymerase chain reaction (RT-PCR), *in situ* hybridization (ISH), and immunohistochemistry. A series of 20 nonneuroendocrine lung carcinoma samples served as controls. RT-PCR was positive for sst2 in 22 of 26 samples, including 15 of 15 typical carcinoids, 5 of 6 atypical carcinoids, and 2 of 5 small-cell carcinomas. The sst2 mRNA signal obtained by RT-PCR was strong in the majority (87%) of typical carcinoids and of variable intensity in atypical carcinoids and small-cell carcinomas. A weakly positive signal was observed in 5 of 20 control samples. In immunohistochemistry, two different antibodies (anti-sst2) were employed, including a monoclonal antibody, generated in the Department of Pathology, University of Turin. In the majority of samples a good correlation between sst2 mRNA (as detected by RT-PCR) and sst2 protein expression (as detected by immunohistochemistry) was observed. However, one atypical carcinoid and one small-cell carcinoma had focal immunostaining but no RT-PCR signal. ISH performed in selected samples paralleled the results obtained with the other techniques. A low sst2 expression was associated with

high grade neuroendocrine tumors and with aggressive behavior. It is concluded that 1) neuroendocrine tumors of the lung express sst2, and there is a correlation between the mRNA amount and the degree of differentiation; 2) immunohistochemistry and ISH are reliable tools to demonstrate sst2 in these tumors; and 3) sst2 identification in tissue sections may provide information on the diagnostic or therapeutic usefulness of somatostatin analogues in individual patients with neuroendocrine tumors.

Key Words: Neuroendocrine—Lung—Tumors—Somatostatin receptors—Immunohistochemistry—Small cell carcinoma—Reverse transcriptase polymerase chain reaction.

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The somatostatin receptor family (sst) includes at least five isoforms that have been recently identified and characterized (18,32,41). The ssts are widely distributed in normal human tissues and in human tumors. Sst type 2 is more commonly detected in neuroendocrine tumors (32,37) and binds the somatostatin analogue octreotide with high affinity.

Sst localization had originally been demonstrated by means of binding assays of radiolabeled somatostatin analogues (20,25,31). Subsequently, specific sst messenger RNA (mRNA) detection was obtained by means of *in situ* hybridization (ISH) and reverse transcriptase polymerase chain reaction (RT-PCR) (14,32,37). Recently, polyclonal antibodies specific for different isoforms of sst were produced and used in immunohistochemistry (10,12,15,18,30,35,36). Given the well-known heterogeneity of neoplastic populations, *in situ* methods (immunohistochemistry and ISH) allow a more definite mapping of the distribution of the receptor in such tissues.

From the Department of Biomedical Sciences and Oncology (M.P., S.C., L.M., A.F., C.P., G.B.), University of Turin, Italy; and Glaxo Institute of Applied Pharmacology (M.S.), University of Cambridge, United Kingdom.

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A.F. is the recipient of a Research Contract by the University of Turin.

Address correspondence and reprint requests to Prof. M. Papotti, Department of Pathology, University of Turin, Via Santena 7, I-10126 Torino.

This is potentially useful for predicting the responsiveness of a given neoplastic cell population to medical treatment with somatostatin analogues, which are used in the clinical setting for both diagnostic and therapeutic purposes with special reference to neuroendocrine tumors.

The spectrum of neuroendocrine tumors of the lung includes well-differentiated neoplasms (so-called typical carcinoids) and poorly differentiated small-cell carcinomas (SCCs). Intermediate forms sharing features of both the aforementioned types also belong to this spectrum (so-called atypical carcinoids or well-differentiated neuroendocrine carcinomas). Finally, large-cell neuroendocrine carcinoma has been identified and included in this tumor group (4,40). The tissue distribution of *sst2* in neuroendocrine tumors of the lung has not been thoroughly characterized, although individual samples of bronchial carcinoids were found to express *sst2* (30). SCCs (but not non-small-cell types) were also shown to be *sst2* positive by receptor binding assay (33). Moreover, *sst2* has been detected in *in vitro* cell cultures of human SSC of the lung (39,42). No study on a series of neuroendocrine tumors of the lung including all neuroendocrine lung tumor types has been reported to date.

The aim of this study was therefore to investigate the presence of *sst2* mRNA and protein in a series of 26 neuroendocrine tumors of the lung, employing different technical approaches, such as RT-PCR, ISH, and immunohistochemistry. To this purpose a monoclonal antibody to *sst2* (N-terminal) was generated in the Department of Pathology, University of Turin. The results were then compared and related to the tumor grade and to other clinicopathologic parameters.

MATERIALS AND METHODS

Case Series and RNA Extraction

Twenty-six samples of neuroendocrine tumors of the lung, in which fresh frozen tissue was available, were retrieved from the surgical pathology file of the University of Turin, Italy. All samples were reviewed applying currently accepted criteria of classification (4,40), and the neuroendocrine nature was confirmed by positive immunostaining for chromogranin A (CgA) (with or without antigen retrieval) or synaptophysin, and by positive RT-PCR for CgA mRNA. According to the classifications described here, these included 15 well-differentiated neuroendocrine tumors (typical carcinoids), 6 well-differentiated neuroendocrine carcinomas (atypical carcinoids), and 5 SCCs.

A series of 20 non-small-cell lung carcinomas (10 squamous, 9 adenocarcinomas, and 1 large-cell anaplastic) lacking neuroendocrine differentiation, as demonstrated by negative immunohistochemistry and RT-PCR for CgA (1), served as a control group. Clinicopathologic data and follow-up information were obtained for all patients.

For hybridization analysis, total RNA was extracted using the guanidine thiocyanate-cesium chloride method (5). The concentration of RNA was estimated by spectrophotometry, and RNA degradation was assessed by agarose gel electrophoresis, as previously reported (37).

Reverse Transcriptase Polymerase Chain Reaction for *sst2* and Chromogranin A

Total RNA (2 μ g) was first digested, with 10 units of RNase-free DNase (Boehringer, Mannheim, Germany) in a 10- μ L solution containing 20 mmol/L $MgCl_2$, to avoid DNA contamination. The solution was kept at room temperature for 10 minutes, then heated for 5 minutes at 70°C to inactivate the DNase molecules; 40 pmol/L of oligodeoxythymidine primers (oligo-dT16) were added and the solution was heated again at 70°C for 10 minutes, then chilled on ice to allow the primer hybridization. The resulting solution was reverse transcribed using 100 units of reverse transcriptase (Gibco BRL, Gaithersburg, MD). Complementary DNA (cDNA) was generated in a 50- μ L final reaction volume containing 50 mmol/L Tris-HCl pH 8.3, 75 mmol/L KCl, 3 mmol/L $MgCl_2$, 10 mmol/L dithiothreitol, 1 mmol/L deoxynucleotide triphosphates (dNTPs), and 20 units of RNasin (Promega, Madison, WI). The solution was heated at 37°C for 90 minutes. Finally, the enzymes were inactivated by heating to 70°C for 10 minutes.

The efficiency of the reverse transcription was determined by performing a PCR reaction having the β_2 -microglobulin "housekeeping gene" as a target. PCR was carried out in a 10- μ L final reaction volume containing 1 μ L of cDNA template, 10 pmol of sense and antisense oligonucleotide primers, 67 mmol/L Tris-HCl pH 8.8, 16 mmol/L $(NH_4)_2SO_4$, 0.01% polysorbate 20, 2 mmol/L dNTPs, 1 mmol/L $MgCl_2$, and 0.5 units of Taq polymerase. β_2 -Microglobulin, *sst2*, and CgA PCR reactions were performed using the same protocol at the following PCR conditions: 35 cycles, each cycle consisting of denaturation at 94°C for 2 minutes, annealing at 55°C for 1 minute for β_2 -microglobulin, at 61°C for *sst2*, and at 68°C for CgA; extension was performed at 72°C for 1 minute. The primers used for RT-PCR (9,11,23,37) are reported in Table 1.

The amplified fragments were run in a 1% agarose gel, containing ethidium bromide. Strict precautions against contamination were undertaken (19) and negative controls (a no-template control and a no-reverse transcriptase control and distilled water to replace the RNA) were included. The RNA extracted from an H716 neuroendocrine colon carcinoma cell line and from a neuroblastoma (37) served as positive controls for CgA and *sst2*, respectively.

Antibodies

Two different antibodies specific for *sst2* were employed. The first one was a monoclonal antibody raised

TABLE 1. Sequences of primers used for reverse transcriptase polymerase chain reaction

| | Size of PCR product (bp) | Position | Study |
|---|--------------------------|----------|---------------------|
| 1) β_2 -microglobulin sense: 5' ACC CCC ACT GAA AAA GAT GA 3' | 120 | 286-305 | Gussow et al. (9) |
| 2) β_2 -microglobulin antisense: 5' ATC TTC AAA CCT CCA TGA TG 3' | | 389-408 | |
| 3) SSTR2 sense: 5' CAG TCA TGA GCA TCG ACC GA 3' | 284 | 402-421 | Sestini et al. (37) |
| 4) SSTR2 antisense: 5' GCA AAG ACA GAT GAT GGT GA 3' | | 665-684 | |
| 5) CgA sense: 5' GCT CCA AGA CCT CGC TCT CC 3' | 583 | 316-335 | Helman et al. (11) |
| 6) CgA antisense: 5' GAC CGA CTC TCG CCT TTC CG 3' | | 878-897 | |

PCR, polymerase chain reaction.

in the Department of Pathology (University of Turin) specific for an N-terminal sequence of the sst2 (shared by both A and B receptor isoforms). The octapeptide EPYYDLTS, corresponding to amino acids 35 to 42 of the human receptor (and differing by one amino acid from the mouse sequence), was synthesized, having a lysin added to the N-terminal. This sequence was similar to that used by other groups to produce polyclonal antibodies (17,18,27). This sequence was rather short but made it possible to avoid extensive homology with sst1. In addition, according to a genbank search using FASTA (28), this protein sequence is unique to human sst2 and has a partial homology only with rat and human nuclear receptor retinoid orphan nuclear receptor-beta (a protein having nuclear localization). Three Balb/c mice were immunized with the peptide conjugated to keyhole limpets hemocyanin (KLH) (Sigma, St. Louis, MO) following the standard procedure. After the first intrasplenic injection (100 μ g of protein) at time 0, the mice were intraperitoneally injected six times with the peptide-KLH conjugate (150 μ g) in the presence of Freund adjuvant. The reactivity of the sera from each animal was evaluated using an enzyme-linked immunosorbent assay, using the peptide coated onto the plastic. The hybridomas were produced by somatic fusion of immunized splenocytes with the mouse myeloma cell line Ag8.X63.653, following the standard technique (21). The monoclonal antibodies of interest were selected on the basis of the reactivity with the target peptide and with appropriate tissue sections. The latter included formalin-fixed and paraffin-embedded sections of pituitary gland and pancreatic islets and were analyzed by means of immunoperoxidase staining. Parallel control experiments were also performed by staining serial sections of these tissues, omitting the primary antibody or with the preimmune serum or with the antibody preadsorbed with high concentrations (1 mg/mL) of the antigenic peptide. In addition, the selected monoclonal antibodies (coded 10C6 and 10G4), both of IgM isotype, were further characterized by Western blotting. Membranes were prepared from stable transfected Chinese hamster ovary (CHO)-K1 cells, individually expressing recombinant human somatostatin receptors (sst1 to sst5). Western blotting was performed as previously described (36). The monoclonal

antibody was used as culture supernatant at 1:3 dilution for 2 hours at room temperature in Tris-buffered saline (TBS), supplemented with 0.1% polysorbate 20. Blots were washed in TTBS and incubated with peroxidase-conjugated goat antimouse IgM, diluted 1:1,500 for 90 minutes at room temperature. Then, blots were washed in TTBS and immunocomplexes were visualized using ECL following manufacturer's instructions (Amersham, Bucks, UK).

A second polyclonal antibody was produced that had been characterized previously (35,36). This antibody (coded K230) was raised in sheep and was specific for a sequence of the C-terminal portion of the sst2A (KSRL-NETTETQRTLLNEDLQ, amino acids 347 to 366).

Immunohistochemistry

Sections 4 or 5 μ thick, adjacent to those used for conventional histopathologic examination and immunostaining for neuroendocrine markers, were collected onto poly-L-lysine-coated slides. The proliferative activity of the tumors was assessed by means of Ki67 immunostaining (clone MIB1, Immunotech, Marseille, France), diluted 1:10 after microwave-based antigen retrieval in citrate buffer). The ascitic fluid of monoclonal antibody 10G4 was used in this study and was applied to tissue sections with prior antigen retrieval (three 3-minute passages in a microwave oven at 800 W in citrate buffer pH 6.0), at the dilution of 1:10,000 or 1:12,000 for 30 minutes at room temperature. The antiserum coded K230 was applied overnight at a dilution of 1:300 with no prior antigen retrieval. The immune reactions were then revealed with the immunoperoxidase technique (13) using the streptavidin-peroxidase kit and diaminobenzidine as chromogen. A weak nuclear counterstain or no counterstain was used in parallel sections. Control stainings for both antibodies included immunoperoxidase of serial sections using preimmune serum or antibody preadsorbed with the antigen or buffer instead of the primary antibody.

In Situ Hybridization

Selected tumors (12 samples) were also analyzed for sst2 mRNA expression by means of a nonradioactive, tyramide deposition-based ISH technique. The proce-

cedure of amplification was modified from procedures reported by Kerstens et al. (16), Speel et al. (38), and the GenPoint (biotinyl-tyramide) manufacturer (Dako, Glostrup, Denmark). Briefly, 5- μ m-thick paraffin sections were collected onto silane-coated slides and deparaffinized through xylene and graded alcohols to phosphate buffer saline (PBS). The slides were then incubated for 5 minutes in a microwave oven at 800 W in citrate buffer pH 6.0. After washing in PBS, they were digested with proteinase K (1 μ g/mL) for 10 minutes at 23°C. Endogenous peroxidase activity was blocked with 3% hydrogen peroxide and endogenous biotin was blocked using avidin-blocking reagent for 15 minutes followed by washing in PBS and biotin-blocking reagent for 15 minutes (3). Sections were then prehybridized for 1 hour at room temperature in a mixture composed of 4 \times SSC, 50% formamide, Denhardt's 1 \times , dextrane sulfate 5 \times , 500 μ g/mL salmon sperm DNA, and 250 μ g/mL tRNA. Hybridization took place overnight at 42°C in a solution containing the specific probe at a concentration of 1 pmol/mL. The probe was a digoxigenin-labeled 48-base oligonucleotide (32), complementary to positions 91 to 139 of the human *ss2* gene (41). After hybridization, excess hybridization buffer and coverslips were removed by a rapid wash in 4 \times SSC followed by stringent washing in 0.1 \times SSC for 10 minutes at 42°C. The hybrids were revealed by the following incubation steps: peroxidase-labeled antidigoxigenin (diluted 1:100 in PBS) for 30 minutes at room temperature, biotinylated tyramide (diluted 1:5 in PBS) for 15 minutes at room temperature, and peroxidase-labeled streptavidin for 15 minutes at room temperature. Diaminobenzidine was used as chromogen. Controls for ISH included staining of serial sections with sense probe, an unrelated probe (EBER-1 of the Epstein-Barr virus), and omission of the probe in the hybridization mixture, with all other experimental conditions identical to the procedure described here.

RESULTS

Reverse Transcriptase Polymerase Chain Reaction

All neuroendocrine tumors, but no nonneuroendocrine lung carcinomas, were positive for CgA mRNA (Fig. 1). *Sst2* mRNA was amplified in 22 of 26 samples of neuroendocrine tumor. The signals had variable intensities (Fig. 2) and were weak in moderately or poorly differentiated tumors (mostly in SCCs). No amplification was obtained in no-template or no-reverse transcriptase experiments. Control samples (nonneuroendocrine lung carcinomas proven by negative CgA RT-PCR) were weakly positive for *ss2* in 5 of 20 samples only (including 3 adenocarcinomas, 1 squamous, and the large-cell anaplastic carcinoma) (Fig. 3). These differences were statistically significant ($P < 0.01$) by χ^2 test.

Characterization of Monoclonal Antibodies to *ss2*.

Several clones were identified having a positive binding by enzyme-linked immunosorbent assay and a parallel immunoreactivity on formalin-fixed paraffin-embedded human endocrine tissues (pituitary and pancreatic islets). In Western blotting experiments, two clones (coded 10C6 and 10G4) specifically developed a band at approximately 70 kD. When the antibodies were used against CHO-transfected cells expressing recombinant somatostatin receptors 1 through 5, a specific band corresponding to *ss2* (at approximately 70 kD) was revealed by the monoclonal antibody 10G4. Monoclonal antibody 10C6 developed a strong band with *ss2* but displayed a weaker reactivity also with *ss1*, 3, and 5, at least in the present experimental conditions (Fig. 4 A,B). The same antibodies were also tested by means of immunoperoxidase staining on formalin-fixed, paraffin-embedded samples of normal human pituitary gland and pancreas. Monoclonal antibody 10G4 gave good results in immunohistochemistry and was used at increasing di-

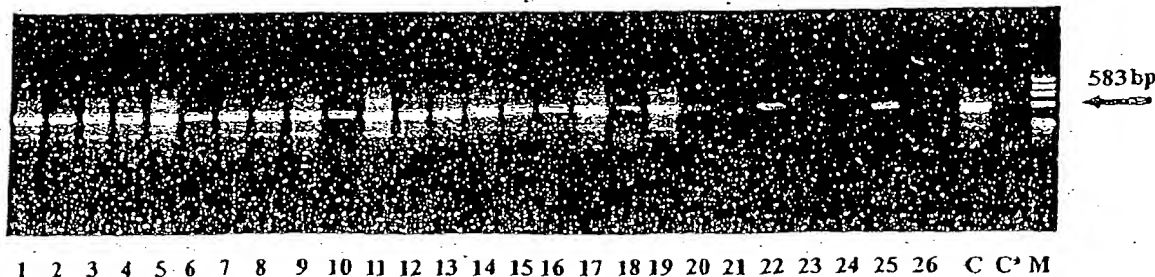


FIG. 1. Reverse transcriptase polymerase chain reaction for chromogranin A (CgA) mRNA in 26 samples of neuroendocrine tumor of the lung. Numbers in each lane correspond to sample numbers in Table 2. CgA mRNA is amplified at 429 bp. C and C* stand for positive (neuroendocrine colon carcinoma cell line, H716) and negative (distilled water) controls, respectively. The last column to the right represents the molecular weight marker. All samples are positive with a variable intensity of the amplification band.

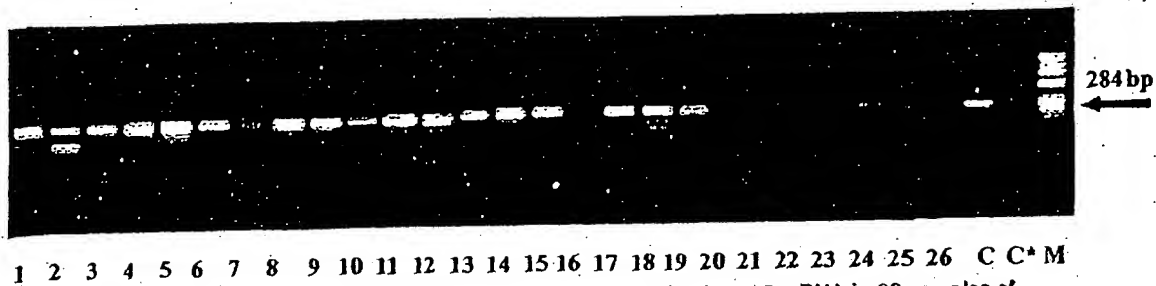


FIG. 2. Reverse transcriptase polymerase chain reaction for *sst2* mRNA in 26 samples of neuroendocrine tumor of the lung. Numbers in each lane correspond to sample numbers in Table 2. *sst2* mRNA is amplified at 284 bp. C and C* stand for positive (a neuroblastoma) and negative (distilled water) controls, respectively. The last column to the right represents the molecular weight marker. Twenty-two of 26 samples are positive with a variable intensity of the amplification band.

lutions (up to 1:15,000) with specific staining. Using thin sections (approximately 4 μ m), a strong membrane-bound and peripheral cytoplasmic immunoreactivity was found in an adenohypophyseal cell population (corresponding to growth hormone-secreting cells, as confirmed by double immunohistochemical analyses) and in pancreatic islets (Fig. 4 C,D). In the latter, the staining was apparently not restricted to a specific hormone-producing cell type and had a peripheral cytoplasmic or membrane distribution. Exocrine pancreatic cells (both acinar and ductal) were only occasionally immunostained. Immunohistochemistry performed on serial control sections, either omitting the primary antibody or using the preimmune serum or antibodies preabsorbed with the synthetic peptide, was negative in both tissues. Monoclonal antibody 10C6 had a relatively higher background staining at similar dilutions.

Immunohistochemistry

The antibodies to *sst2* (monoclonal antibody 10G4 and polyclonal K230) gave slightly different immunoreactions in 25 samples, and staining was not done in 1 sample because of lack of residual paraffin blocks. The monoclonal antibody 10G4 stained 21 of 25 samples, the

negative samples being 1 atypical carcinoid and 3 SCCs (Fig. 5). The tumors had 5% to 25% of the neoplastic cells immunoreactive. The staining was at the periphery of the cytoplasm, and omitting the counterstain its membrane-bound distribution was better outlined in most samples (Fig. 6). One sample of atypical carcinoid (no. 21) was focally immunoreactive for *sst2*, despite negative RT-PCR findings. Conversely, sample no. 26 was immunohistochemistry negative and RT-PCR positive. The antiserum anti-*sst2A* (code K230) gave positive signal in 19 of 25 samples, in 5% to 60% of the neoplastic cell population (Fig. 7). The location of the staining was at the membrane level associated with a weak cytoplasmic reactivity. The same pattern was seen in positive controls, e.g., pancreatic islets (Fig. 7, inset). Two samples (nos. 19 and 26) were negative in spite of a positive RT-PCR signal. Two other tumors (nos. 21 and 22), apparently devoid of *sst2* mRNA, showed a small percentage of immunoreactive cells. Incidentally, one of these latter samples (no. 21) was also immunoreactive with monoclonal antibody 10G4 (Table 2).

The five control samples positive by RT-PCR were also reactive with the antibodies. The type of immunocytochemical location of *sst2* receptors was similar to that described here, being a peripheral cytoplasmic stain-

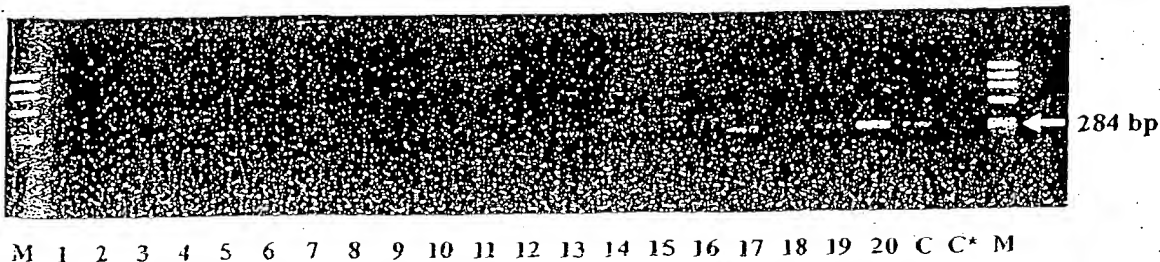


FIG. 3. Reverse transcriptase polymerase chain reaction for *sst2* mRNA in 20 control samples of nonneuroendocrine lung carcinoma. Five of 20 samples show a weak band at 284 bp corresponding to *sst2* mRNA. Control columns (C and C*) are identical to those in Fig. 2.

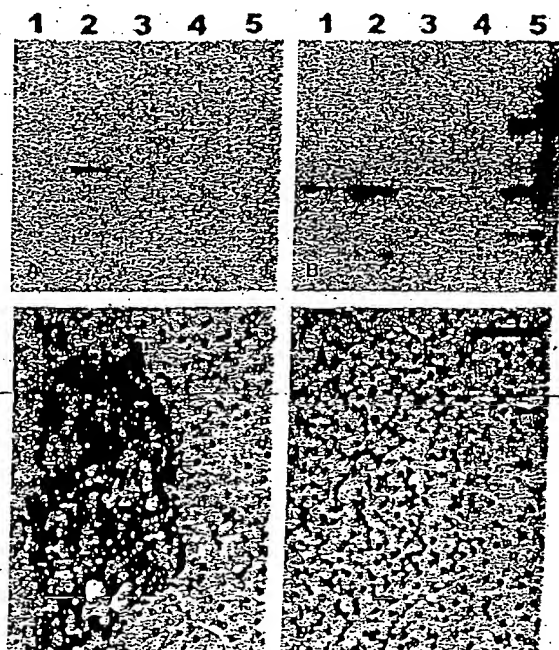


FIG. 4. Western blot analysis of monoclonal antibody clones 10G4 and 10C6 against sst2 in Chinese hamster ovary cells transfected with recombinant sst 1 through 5 (numbers of each column correspond to receptor type). Monoclonal antibody 10G4 shows a specific band at approximately 70 kD for sst2 only (A) as opposed to monoclonal antibody 10C6, which strongly reacts with sst2 but also has some degrees of cross-reactivity with sst 1, 3, and 5 (B). The lower figures show control formalin-fixed paraffin-embedded pancreatic islets immunostained with monoclonal antibody 10G4 without (C) and with (D) preadsorption with the peptide antigen, respectively. The majority of endocrine cells show a membrane-bound immunoreactivity (C) (immunoperoxidase). Bar: 90 μ m.

ing present in 40% to 70% of neoplastic cells. A weak and focal staining was also observed in five of the remaining RT-PCR-negative samples, when the antibody K230 was used (but not when the monoclonal was employed).

Several cells in peritumoral tissues were occasionally stained. Ciliated cells of bronchial mucosa had a peripheral staining at the cilia border. Mucous glands were negative. Rare chondrocytes had a membrane staining. The wall of peritumoral as well as of occasional distant vessels was stained at the endothelium level and in occasional smooth muscle cells.

The reactivity of both antibodies was abolished in serial sections when the reagents were preabsorbed with the respective synthetic peptides, but not when an unrelated peptide was used. The peritumoral bronchial mucosa had a focal staining of ciliated cells with both antibodies. This reactivity disappeared when the preabsorbed antibody was applied.

In Situ Hybridization

Eight of 12 samples stained by ISH were positive for sst2 mRNA. The mRNA was present in a percentage of cells (ranging from 10% to 40%) and gave a weak signal (Fig. 8), despite the amplification provided by the tyramide-based procedure. The background level was minimal using diluted biotinylated tyramide. Control sections stained with sense probe or an unrelated probe, or omitting the probe, were consistently negative.

Clinical Data

Clinicopathologic data are summarized in Table 2. At follow-up, the majority of patients with typical carcinoids are free from disease 1 to 11 years after surgery. Two patients are alive with stable metastatic disease. Patients affected by atypical carcinoids had disease progression in one third of samples. Finally, patients with SCC had fatal outcomes within 1 year from diagnosis (except the recent sample). Eight patients had preoperative octreotide scintigraphy performed at the time of diagnosis. All patients had positive octreoscan findings, and, in these patients, also the tumor was positive by RT-PCR and immu-



FIG. 5. sample no. 25 (small cell carcinoma). Absence of immunoreactivity for sst2 with the monoclonal 10G4. This sample was also negative by reverse transcriptase polymerase chain reaction and in situ hybridization. (Immunoperoxidase in a formalin-fixed paraffin-embedded sample. Nuclei slightly counterstained with hemalum.) Bar: 45 μ m.

nohistochemistry or ISH. In addition, three of these patients received octreotide therapy administered at the time of tumor recurrence or metastatic spread. Stable disease is recorded at follow-up more than 5 years after diagnosis.

Correlations

Overall, complete overlapping (i.e., RT-PCR, ISH, and immunohistochemistry with two antibodies) between *sst2* gene and protein expression was obtained in 21 of 25 samples (84%) and between RT-PCR results and immunohistochemical findings with at least one of the antibodies in 24 of 25 samples (96%). The monoclonal antibody 10G4 looked highly sensitive, being able to stain all but one sample (no. 26) (95%) positive for *sst2* mRNA by RT-PCR. *Sst2* expression, at mRNA as well as at protein levels, was reduced in high grade tumors, with SCCs being weakly positive in only two of five samples. Decreasing expression of *sst2* appears to cor-



FIG. 6. sample no. 16 (typical carcinoid). Immunohistochemical detection of *sst2* by means of monoclonal antibody 10G4. The neoplastic cells have a peripheral cytoplasmic staining and membrane positivity in some cells, whereas the peribronchial gland adjacent to the tumor is unreactive. (Immunoperoxidase in a formalin-fixed paraffin-embedded sample. Nuclei slightly counterstained with hemalum.) Bar: 45 μ m. The membrane-bound distribution of the immunostaining is better outlined in a parallel section stained for monoclonal antibody 10G4 omitting nuclear counterstain (inset).



FIG. 7. Same sample as in Fig. 6. Immunohistochemical detection of *sst2* by means of the polyclonal antibody K230. The immunostaining is more intense at the cell border (arrows), as observed with the monoclonal antibody. In the inset, a pancreatic islet, used as positive control, shows a predominant membrane-bound immunostaining of many neuroendocrine cells. (Immunoperoxidase in a formalin-fixed paraffin-embedded sample. Nuclei slightly counterstained with hemalum.) Bar: 45 μ m.

relate with high tumor grade and elevated proliferative activity, but not with other parameters such sex, age, or tumor size.

DISCUSSION

In this study, the presence of *sst2* mRNA has been demonstrated in a series of resected neuroendocrine tumors of the lung by means of RT-PCR and confirmed by a sensitive nonradioactive tyramide-based ISH procedure and by immunohistochemistry with anti-*sst2* antibodies. Samples of both carcinoid tumors and SCCs were *sst2* positive, although a reduced or absent signal was observed in poorly differentiated (small-cell) carcinomas. This is the first study of *sst2* expression in a relatively large series of neuroendocrine tumors of the lung. Single samples of human carcinoids and SCCs (including cell lines of the latter) had previously been analyzed and found to express *sst2* (7,15,30,32,33,39,42). Several methods have been used to detect these receptors and partially overlapping results were obtained.

In the present study, the expression of high amounts of

TABLE 2. Clinicopathologic data and somatostatin receptor type 2 (sst2) expression in 26 cases of neuroendocrine lung tumors

| Patient no. | Diagnosis | Sex/age | Size (cm) | Follow-up (mo) | CgA IHC | CgA RT-PCR | SYP IHC | Ki67 IHC* | sst2 RT-PCR | sst2 IHC Mab(10G4) | sst2 IHC (K230 Ab) |
|-------------|-----------|---------|-----------|----------------|---------|------------|---------|-----------|-------------|--------------------|--------------------|
| 1 | WD NET | F/35 | 3.5 | NED 90 | + | +++ | + F | 1.5 | +++ | + | + |
| 2 | WD NET | F/29 | 4 | NED 45 | + | ++ | + F | 3 | ++ | + | + |
| 3 | WD NET | F/41 | 2.5 | NED 70 | + | ++ | + | 0.1 | +++ | + | + F |
| 4 | WD NET† | F/25 | 4 | NED 21 | + | ++ | + | 2.6 | +++ | + F | + F |
| 5 | WD NET† | M/58 | 3.8 | NED 23 | + | +++ | + | 13 | +++ | + F | + |
| 6 | WD NET | M/52 | 2.5 | NED 42 | + | ++ | + | 4.5 | +++ | + F | + F |
| 7 | WD NET | F/69 | 3.5 | NED 47 | + | ++ | + | 1 | + | + | + |
| 8 | WD NET | M/29 | 3 | NED 70 | + | +++ | + | 1.5 | +++ | + | + |
| 9 | WD NET | M/27 | 4 | NED 108 | + | +++ | + | NT | +++ | + | + |
| 10 | WD NE Ca† | M/66 | 8 | AWD 55 | + | ++ | + F | 1.1 | ++ | + F | + |
| 11 | WD NET† | F/29 | 2 | AWD 56 | + | +++ | + | 2.5 | +++ | + | + |
| 12 | WD NET* | F/32 | 3 | NED 26 | + | ++ | + | 4 | +++ | + | + |
| 13 | WD NE Ca | M/60 | 3 | NED 133 | + | +++ | + | 1 | +++ | NT | NT |
| 14 | WD NET | M/28 | 4 | NED 130 | + | +++ | + | 1 | +++ | + | + |
| 15 | WD NET† | M/41 | 1.3 | AWD 53 | + | +++ | + | 1.5 | +++ | + | + |
| 16 | WD NET† | F/31 | 1 | NED 13 | + | ++ | ++ | 2.6 | + | + | + |
| 17 | WD NET | F/53 | 4 | NED 24 | + | +++ | + F | 4 | +++ | + F | + |
| 18 | WD NE Ca | M/62 | 3 | NED 6 | - | ++ | - | 13 | +++ | + | + F |
| 19 | WD NE Ca | F/73 | 5 | DOD 20 | + | +++ | + | 3 | ++ | + | - |
| 20 | SCC | M/57 | 6 | DOD 12 | + | ++ | + | 45 | + | + F | + F |
| 21 | SCC | M/51 | 4.5 | DOD 5 | + | + | + | 35 | - | + F | + F |
| 22 | WD NE Ca | M/60 | 6 | NED 51 | + | ++ | + | 1.5 | - | - | + |
| 23 | SCC | F/56 | 6 | DOD 11 | + | + | + | 50 | - | - | - |
| 24 | WD NE Ca | M/77 | 2.5 | NED 21 | + | + | + | 24 | + | + F | + F |
| 25 | SCC | M/57 | 5 | DOD 10 | + | ++ | ++ | 80 | - | - | - |
| 26 | SCC† | M/68 | 11 | recent case | + | + | ++ | 71 | + | - | - |

AWD, alive with disease; CgA, chromogranin A; DOD, died of disease; + F, focal: positive in <5% of cells; IHC, Immunohistochemistry; Mab, monoclonal antibody; NECa, Neuroendocrine carcinoma; NED, no evidence of disease; NET, neuroendocrine tumor; NT, not tested; RT-PCR, reverse transcriptase polymerase chain reaction; SCC, Small-cell lung carcinoma; SYP, synaptophysin; WD, well differentiated.

* Ki67 IHC: values correspond to percentage of positive nuclei of neoplastic cells.

† Patients who had preoperative octreoscan performed.

‡ Patients who had octreoscan performed and octreotide treatment.

sst2 mRNA was confirmed in well to moderately differentiated neuroendocrine tumors, in agreement with the results obtained by Reubi et al. (32) by means of radioactive ISH. The presence of sst2 mRNA in SCC had never been reported in human specimens, except for two samples included in Reubi et al.'s series (32). Although the data on cell lines support the observation that SCCs contain sst2 (42), slightly discrepant results were found in some of samples described here. Unfortunately, SCCs are rarely operated on, and therefore it is difficult to collect a large number of surgical specimens. The five samples studied in the current series by means of RT-PCR had a low amount (two samples) or absent (three samples) sst2 mRNA. This could be the result of the extensive necrosis commonly present in such tumor types. However, because care was taken to freeze fragments that were macroscopically devoid of necrotic areas, a more likely hypothesis is that sst2 expression is reduced in poorly differentiated tumors. Recently, Reisinger et al. (29) showed that the uptake of somatostatin analogues in patients with SCC undergoing chemotherapy is significantly lower, and therapeutic external factors may affect the receptor status of individual tumors. In addition, the uptake of somatostatin analogues

in metastatic deposits of SCC has been shown to be low or absent (29,2). The present findings suggest that the sst2 mRNA content is related to the degree of tumor differentiation. These data must be confirmed in larger series of nonneuroendocrine tumors to ascertain whether the observed loss or decrease of sst2 expression in neuroendocrine tumors is a common event linked to neoplastic dedifferentiation. In addition, further studies are needed to assess the functionality of such receptors, by comparing the profile of sst2 expression in tumor tissues with binding assays employing labeled somatostatin and with the clinical response to diagnostic and therapeutic administration of somatostatin analogues.

To this purpose, several investigators have demonstrated a correlation between clinical imaging or response to somatostatin analogue treatment and sst2 mRNA content in single samples of carcinoid tumors (15,22). Northern blotting and ISH were the techniques used for sst2 mRNA identification. This kind of correlation is useful for selecting patients for somatostatin analogue treatment, although the demonstration of receptor mRNA in a cell does not imply per se that the receptor is fully functional.

The present study relied on a highly sensitive tech-

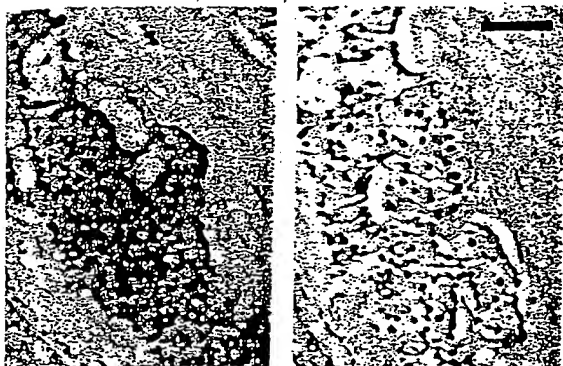


FIG. 8. sample no. 14 (typical carcinoid). In-situ hybridization (ISH) for sst2 mRNA shows a weak cytoplasmic staining (A) in most tumor cells. An ISH performed with an unrelated probe was negative in a serial section of the same tumor (B). This sample was strongly positive by reverse transcriptase polymerase chain reaction for sst2 mRNA and by immunohistochemistry. (Nonradioactive ISH revealed by peroxidase and diaminobenzidine, as substrate. Nuclei counterstained with hemalum.) Bar: 75 μ m.

nique, RT-PCR, to identify all samples bearing even small amounts of sst2 mRNA. Indeed, in a previous study, single samples exhibiting octreotide-binding sites had no demonstrable sst2 mRNA by means of ISH, possibly due to the low sensitivity of the ISH procedure (34). The RT-PCR has shown sst2 mRNA transcripts in the majority of samples here studied. Only four samples were negative, all belonging to poorly differentiated high grade tumors, which usually follow an aggressive course. A decrease of sst2 mRNA expression in association with neuroendocrine tumor dedifferentiation had also been reported in neuroblastomas (37). In the above report, as well as in the current study, samples having an unfavorable prognosis were found to contain a relatively low amount of sst2 mRNA, as compared with well-differentiated tumors.

In the current sample series, eight samples were investigated before surgery with radiolabeled octreotide. Despite the low figures, all the samples positive at the diagnostic procedure had a strong RT-PCR signal for sst2 mRNA. Three of eight patients were also responsive to octreotide treatment administered at the time of relapse or metastatic spread. More extensive correlative clinicopathologic studies on the sst status are needed to better define the tissue distribution of somatostatin binding sites and their potential clinical role in the treatment of patients.

Sst2 evaluation by means of ISH (14,32) or RT-PCR (26,37) is a highly sensitive and reliable procedure. Unfortunately, these techniques have limitations because frozen tissue is needed for some of them, and radioactive material or costly and time-consuming methods are nec-

essary for others. Immunohistochemical analysis of sst2 by means of specific antibodies represents an ideal, cheap, and rapid alternative, easily applicable to archival material. For these reasons, several investigators have raised polyclonal antibodies specific for sst (8,10,15,17, 18). In the current study, tested tumor fragments adjacent to those snap frozen for RT-PCR analysis were tested with a polyclonal antibody against a C-terminal portion of the sst2A splice variant (35,36). In addition, a monoclonal antibody was produced in the Department of Pathology (University of Turin) against an N-terminal sequence of the human sst2. This antibody was the first monoclonal developed against sst2 and was shown to be highly specific for sst2 in Western blot and immunohistochemical analysis. Both the monoclonal and the polyclonal antibodies specifically reacted with all samples also positive by RT-PCR (with minor discrepancies in two samples, likely due to tumor heterogeneity). The observed correlation between RT-PCR and immunohistochemistry indicates that the latter may be a reliable diagnostic tool and may allow immunohistochemical investigation for sst2 even in small biopsy samples. This in turn may enable a rapid screening of sst2-positive tumors for medical treatment with somatostatin analogues.

Having confirmed in a relatively large series that the vast majority of neuroendocrine tumors of the lung contain variable amounts of sst2 mRNA, a final comment is deserved for sst2 expression in nonneuroendocrine lung carcinomas. No data have been reported thus far in the literature concerning normal human lung, although in the present study some bronchial cells of peritumoral parenchyma were positive for sst2 when immunohistochemical analysis was performed with either antibody. The staining was specific because it was abolished using preabsorbed antibodies. Therefore, it is likely that normal human lung tissue contains sst2. This might be confirmed by alternative techniques (e.g., Western blot, RT-PCR). However, in situ morphologic procedures, such as those employed here, have definite advantages. In fact, the lung is rich in vessels, and in several tissues (either in tumoral or in inflammatory-reactive conditions) the vessels were recently shown to contain sst (6).

A low expression of sst2 was found in 25% of lung carcinomas of nonneuroendocrine type investigated in the present study by means of RT-PCR. Therefore, sst type 2, at least, does not appear to be extensively expressed in nonneuroendocrine carcinomas of the lung. However, because two tumors in the control group (a squamous carcinoma and an adenocarcinoma, respectively) had positive octreotide scintigraphy, but no sst2 mRNA, it is plausible that a heterogeneous distribution of sst occurs in nonneuroendocrine lung tumors. Other receptor types may be expressed in these tumors and may be responsible for the positive results in diagnostic testing. Because sst5 is also known to bind somatostatin

analogues, such as octreotide, with high affinity (24), the expression of this receptor type will be investigated in future studies. □

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REFERENCES

1. Abbondanza GC, Papotti M, Viberi L, Macri L, Siella A, Bussolati G. Chromogranin A gene expression in non-small cell lung carcinomas. *J Pathol* 1998;186:1-6.
2. Berenger N, Moretti JL, Boaziz C, Vigneron N, Morete JF, Breaux JL. Somatostatin receptor imaging in small cell lung cancer. *Eur J Cancer* 1996;32:1429-31.
3. Bussolati G, Gugliotta P, Volante M, Pace M, Papotti M. Retrieved endogenous biotin: a novel marker and potential pitfall in diagnostic immunohistochemistry. *Histopathology* 1997;31:400-7.
4. Capella C, Heitz PU, Hofler H, Solcia E, Kloppel G. Revised classification of neuroendocrine tumors of the lung, pancreas and gut. *Virchows Arch* 1995;425:547-60.
5. Chirgwin JM, Przybyla AE, MacDonald RJ, Rutter W. Isolation of biologically active RNA from sources enriched in ribonuclease. *Biochemistry* 1979;18:5294-7.
6. Denzler B, Reubi JC. Expression of somatostatin receptors in peritumoral veins of human tumors. *Cancer* 1999;85:189-98.
7. Fujita T, Yamaji Y, Sato M, Murao K, Takahara J. Gene expression of somatostatin receptor subtypes, SSTR1 and SSTR2, in human lung cancer cell lines. *Life Sci* 1994;55:1797-806.
8. Gu WZ, Schonbrunn A. Coupling specificity between somatostatin receptor sst2A and G proteins: isolation of the receptor-G protein complex with a receptor antibody. *Mol Endocrinol* 1997;11:527-37.
9. Gussow D, Rein R, Ginjzar I, Hochstenbach F, Seemann G, Kottman A, Ploegh HL. The human beta 2-microglobulin gene: primary structure and definition of the transcriptional unit. *J Immunol* 1987;139:3132-8.
10. Helboe L, Moller M, Norregaard L, Schiodt M, Sudsen CE. Development of selective antibodies against the human somatostatin receptor subtypes sst1-sst5. *Brain Res Mol Brain Res* 1997;49:82-8.
11. Helman JJ, Ahn TG, Levine MA, et al. Molecular cloning and primary structure of human chromogranin A (secretory protein I) cDNA. *J Biol Chem* 1988;263:11559-63.
12. Hofland LJ, Liu Q, Van Koetsveld FM, et al. Immunohistochemical detection of somatostatin receptor subtypes sst1 and sst2A in human somatostatin receptor positive tumors. *J Clin Endocrinol Metab* 1999;84:775-80.
13. Hsu SM, Raine L, Fanger H. Use of avidin- peroxidase complex (ABC) in immunoperoxidase techniques: comparison between ABC and unlabelled antibody (PAP) procedures. *J Histochem Cytochem* 1981;29:577-80.
14. Janson ET, Gobl A, Kalkne KM, Oberg K. A comparison between the efficacy of somatostatin receptor scintigraphy and that of *in situ* hybridization for somatostatin receptor subtype 2 messenger RNA to predict therapeutic outcome in carcinoid patients. *Cancer Res* 1996;56:2561-5.
15. Janson ET, Sridsberg M, Gobl A, Westlin JE, Oberg K. Determination of somatostatin receptors subtype 2 in carcinoid tumors by immunohistochemical investigation with somatostatin receptor subtype 2 antibodies. *Cancer Res* 1998;58:2375-8.
16. Kerstens HJM, Poddighe PJ, Hanselaar AGJM. A novel *in situ* hybridization signal amplification method based on the deposition of biotinylated tyramine. *J Histochem Cytochem* 1995;43:347-52.
17. Krisch B, Feindt J, Mentlein R. Immunoelectronmicroscopic analysis of the ligand-induced internalization of the somatostatin receptor subtype 2 in cultured human glioma cells. *J Histochem Cytochem* 1998;46:1233-42.
18. Kumar U, Laird D, Srikanth CB, Escher E, Patel YC. Expression of the five somatostatin receptors (SSTR1-5) subtypes in rat pituitary somatotrophs: quantitative analysis by double-label immunofluorescence confocal microscopy. *Endocrinology* 1997;138:4473-6.
19. Kwok S, Higuchi R. Avoiding false positives with PCR. *Nature* 1989;339:237-8.
20. Lamberts SWJ, Hofland LJ, Koetsveld FM, Reubi JC, Bruining HA, Bakker WH, Krenning EP. Parallel *in vivo* and *in vitro* detection of functional somatostatin receptors in human endocrine pancreatic tumors: consequences with regard to diagnosis, localization and therapy. *J Clin Endocrinol Metab* 1990;71:566-74.
21. Malavasi F, Funaro A, Bellone G, et al. Functional and molecular characterization by the CB04 monoclonal antibody of a cell surface structure exerting C3-complement receptor activity. *J Clin Immunol* 1985;5:412-20.
22. Nilsson O, Kolby L, Wangberg B, et al. Comparative studies on the expression of somatostatin receptor subtypes, outcome of octreotide scintigraphy and response to octreotide treatment in patients with carcinoid tumours. *Br J Cancer* 1998;77:632-7.
23. Pagani A, Forni M, Tonini GP, Papotti M, Bussolati G. Expression of members of the chromogranin family in primary neuroblastomas. *Diagn Mol Pathol* 1992;1:16-24.
24. Panetta R, Greenwood MT, Warszynska A, et al. Molecular cloning, functional characterization and chromosomal localization of a human somatostatin receptor (somatostatin receptor type 5) with preferential affinity for somatostatin-28. *Mol Pharmacol* 1994;45:417-27.
25. Papotti M, Macri L, Bussolati G, Reubi JC. Correlative study on neuroendocrine differentiation and presence of somatostatin receptors in breast carcinomas. *Int J Cancer* 1989;43:365-9.
26. Papotti M, Macri L, Pagani A, Aloisi F, Bussolati G. Quantitation of somatostatin receptor type 2 in neuroendocrine (Merkel cell) carcinoma of the skin by competitive RT-PCR. *Endocr Pathol* 1999;10:1-10.
27. Patel YC, Panetta R, Escher E, Greenwood M, Srikanth CB. Expression of multiple somatostatin receptor genes in AIT-20 cells: evidence for a novel somatostatin-28 selective receptor subtype. *J Biol Chem* 1994;269:1506-9.
28. Pearson WR, Lipman DJ. Improved tools for biological sequence comparison. *Proc Natl Acad Sci U S A* 1988;85:2444-8.
29. Reisinger I, Bohuslavzki KH, Brenner W, et al. Somatostatin receptor scintigraphy in small-cell lung cancer: results of a multicenter study. *J Nucl Med* 1998;39:224-7.
30. Reubi JC, Kappeler A, Waser B, Laissue J, Hipkin RW, Schonbrunn A. Immunohistochemical localization of somatostatin receptors sst2A in human tumors. *Am J Pathol* 1998;153:233-45.
31. Reubi JC, Maurer R, von Werder K, Torhorst J, Klijn JGM, Lamberts SWJ. Somatostatin receptors in human endocrine tumors. *Cancer Res* 1987;47:551-8.
32. Reubi JC, Schaefer JC, Waser B, Mengod G. Expression and localization of somatostatin receptors SSTR1, SSTR2, SSTR3 messenger RNA in primary human tumors using *in situ* hybridisation. *Cancer Res* 1994;54:3455-9.
33. Reubi JC, Waser B, Sheppard M, Macaulay V. Somatostatin receptors are present in small-cell but not in non-small-cell primary lung carcinomas: relationship to EGF receptors. *Int J Cancer* 1990;45:269-74.
34. Schaefer JC, Waser B, Mengod G, Reubi JC. Somatostatin receptor subtypes sst1, sst2, sst3 and sst5 expression in human pituitary, gastroentero-pancreatic and mammary tumors: comparison of mRNA analysis with receptor autoradiography. *Int J Cancer* 1997;70:530-7.
35. Schindler M, Holloway S, Humphrey PPA, Waldvogel H, Faull RLM, Berger W, Emson PC. Localization of the somatostatin sst2a receptor in human cerebral cortex, hippocampus and cerebellum. *Neuroreport* 1998;9:521-5.
36. Schindler M, Sellers LA, Humphrey PPA, Emson PC. Immunohistochemical localization of the somatostatin sst2(A) receptor in the rat brain and spinal cord. *Neuroscience* 1997;76:225-40.
37. Sestini R, Orlando C, Peri A, et al. Quantification of somatostatin

- receptor type-2 gene expression in neuroblastoma cell lines and primary tumors using competitive reverse transcription-polymerase chain reaction. *Clin Cancer Res* 1996;2:1757-65.
38. Speel EJM, Saremaslani P, Roth J, Hopman AHN, Komminoth P. Improved mRNA in situ hybridization on formaldehyde-fixed and paraffin-embedded tissue using signal amplification with different haptenized tyramides. *Histochem Cell Biol* 1998;420:1-7.
39. Taylor JE, Theveniau MA, Bashirzadeh R, Reisinc T, Eden PA. Detection of somatostatin receptor subtype 2 (SSTR2) in established tumors and tumor cell lines: evidence for sst2 heterogeneity. *Peptides* 1994;15:1229-36.
40. Travis WD, Gal AA, Colby TV, Klimstra DS, Falk R, Koss MN. Reproducibility of neuroendocrine lung tumor classification. *Hum Pathol* 1998;29:272-9.
41. Yamada Y, Post SR, Wang K, Tager H, Bell GI, Scino S. Cloning and functional characterization of a family of human and mouse somatostatin receptors expressed in brain, gastrointestinal tract and kidney. *Biochemistry* 1992;89:251-5.
42. Zhang CY, Yokogoshi Y, Yoshimoto K, Fujinaka Y, Matsumoto K, Saito S. Point mutation of the somatostatin receptor 2 gene in the human small cell lung cancer cell line COR-L103. *Biochem Biophys Res Commun* 1995;210:805-15.

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Expression of somatostatin receptor types 1-5 in 81 cases of gastrointestinal and pancreatic endocrine tumors. A correlative immunohistochemical and reverse-transcriptase polymerase chain reaction analysis.

Papotti M, Bongiovanni M, Volante M, Allia E, Landolfi S, Helboe L, Schindler M, Cole SL, Bussolati G.

Department of Biomedical Sciences and Oncology, University of Turin, Via Santena 7, 10126 Turin, Italy. mauro.papotti@unito.it

Somatostatin receptors (SSTRs) have been extensively mapped in human tumors by means of autoradiography, reverse-transcriptase polymerase chain reaction (RT-PCR), in situ hybridization (ISH) and immunohistochemistry (IHC). We analyzed the SSTR type 1-5 expression by means of RT-PCR and/or IHC in a series of 81 functioning and non-functioning gastroenteropancreatic (GEP) endocrine tumors and related normal tissues. Moreover, we compared the results with clinical, pathological and hormonal features. Forty-six cases (13 intestinal and 33 pancreatic) were studied for SSTR 1-5 expression using RT-PCR, IHC with antibodies to SSTR types 2, 3, 5 and ISH for SSTR2 mRNA. The vast majority of tumors expressed SSTR types 1, 2, 3 and 5, while SSTR4 was detected in a small minority. Due to the good correlation between RT-PCR and IHC data on SSTR types 2, 3, and 5, thirty-five additional GEP endocrine tumors were studied with IHC alone. Pancreatic insulinomas had an heterogeneous SSTR expression, while 100% of somatostatinomas expressed SSTR5 and 100% gastrinomas and glucagonomas expressed SSTR2. Pre-operative biopsy material showed an overlapping immunoreactivity with that of surgical specimens, suggesting that the SSTR status can be detected in the diagnostic work-up. It is concluded that SSTRs 1-5 are heterogeneously expressed in GEP endocrine tumors and that IHC is a reliable tool to detect SSTR types 2, 3 and 5 in surgical and biopsy specimens.

PMID: 12021920 [PubMed - indexed for MEDLINE]

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P-cadherin overexpression is an indicator of clinical outcome in invasive breast carcinomas and is associated with CDH3 promoter hypomethylation.

Paredes J, Albergaria A, Oliveira JT, Jeronimo C, Milanezi F, Schmitt FC.

Institute of Pathology and Molecular Immunology of Porto University (IPATIMUP), Braga, Portugal. jparedes@ipatimup.pt

PURPOSE: P-cadherin overexpression has been reported in breast carcinomas, where it was associated with proliferative high-grade histological tumors. This study aimed to analyze P-cadherin expression in invasive breast cancer and to correlate it with tumor markers, pathologic features, and patient survival. Another purpose was to evaluate the P-cadherin promoter methylation pattern as the molecular mechanism underlying this gene regulation. **EXPERIMENTAL DESIGN:** Using a series of invasive breast carcinomas, P-cadherin expression was evaluated and correlated with histologic grade, estrogen receptor, MIB-1, and p53 and c-erbB-2 expression. In order to assess whether P-cadherin expression was associated with changes in CDH3 promoter methylation, we studied the methylation status of a gene 5'-flanking region in these same carcinomas. This analysis was also done for normal tissue and for a breast cancer cell line treated with a demethylating agent. **RESULTS:** P-cadherin expression showed a strong correlation with high histologic grade, increased proliferation, c-erbB-2 and p53 expression, lack of estrogen receptor, and poor patient survival. This overexpression can be regulated by gene promoter methylation because the 5-Aza-2'-deoxycytidine treatment of MCF-7/AZ cells increased P-cadherin mRNA and protein levels. Additionally, we found that 71% of P-cadherin-negative cases showed promoter methylation, whereas 65% of positive ones were unmethylated ($P = 0.005$). The normal P-cadherin-negative breast epithelial cells showed consistent CDH3 promoter methylation. **CONCLUSIONS:** P-cadherin expression was strongly associated with tumor aggressiveness, being a good indicator of clinical outcome. Moreover, the aberrant expression of P-cadherin in breast cancer might be regulated by gene promoter hypomethylation.

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Mammary-derived growth inhibitor protein and messenger ribonucleic acid concentrations in different physiological states of the gland.

Politis I, Gorewit RC, Muller T, Grosse R.

Department of Animal Science, Cornell University, Ithaca 14853.

Expression of mammary-derived growth inhibitor in tissue from lactating and involuting bovine mammary glands was investigated. Seventeen lactating, pregnant (220 to 272 d in gestation) cows were divided in two groups of 8 and 9 cows each. Cows of the first group were slaughtered while in lactation. Cows of the second group (9 involuting cows) were slaughtered at 13 to 52 d following sudden cessation of milking. High concentrations of mammary-derived growth inhibitor (.63% of the total protein) were detected in mammary tissue of lactating cows. Mammary-derived growth inhibitor (less than .10% of the total protein) was dramatically reduced during most of the involution period (13 to 45 d following cessation of milking). Mammary-derived growth inhibitor was again detected (.28% of the total protein) during the last stage of the involution (46 to 53 d after cessation of milking), which coincided with colostrum formation. When steady state concentrations of mammary-derived growth inhibitor mRNA were examined, the results obtained mirrored those obtained at the protein concentration. These data suggest that regulation of mammary-derived growth inhibitor occurs via modulation of the steady state concentration of its mRNA. Furthermore, there is a strong correlation between mammary-derived growth inhibitor expression and lactation in dairy cows.

PMID: 1500548 [PubMed - indexed for MEDLINE]

Comment in:

- J Invest Dermatol. 1994 Nov;103(5):742-4.

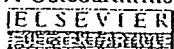
T-cell receptor V beta-family usage in primary cutaneous and primary nodal T-cell non-Hodgkin's lymphomas.

Preesman AH, Hu HZ, Tilanus MG, de Geus B, Schuurman HJ, Reitsma R, van Wichen DF, van Vloten WA, de Weger RA.

Department of Pathology, University Hospital Utrecht, The Netherlands.

To evaluate whether the expression of T-cell receptor (TCR) V beta families in eight cases of malignant T-cell lymphomas took place in a preferential manner, we analyzed four cases of mycosis fungoides (MF), the most common form of primary cutaneous T-cell non-Hodgkin's lymphomas (NHL), and four cases of primary nodal T-cell NHL. The usage of V beta families in T-cell populations was investigated on mRNA that was transcribed to cDNA using a C beta primer and reverse transcriptase. Subsequently, the specific usage of the families was analyzed by polymerase chain reaction (PCR) using combinations of the selected C beta-oligonucleotide primer and one of the family-specific V beta primers. Peripheral blood lymphocytes from four healthy volunteers and 1 "reactive" lymph node served as a control and expressed all 20 V beta families tested for. In T-cell lines, with restricted V beta expression, and in three patients with advanced MF, only one or two V beta families were expressed at the mRNA level. In an early MF lesion this monoclonal expression was absent: several V beta families were expressed with a weak intensity. This may indicate either a polyclonal origin of MF, or that too few monoclonal neoplastic cells were present in the tissue specimen. In the four nodal T-cell NHL, only one family could be clearly distinguished, whereas some of the other V beta families showed only a weak expression. These latter families represent the reactive T-cell component in the nodal T-cell NHL. Both in nodal T-cell NHL and in MF there was no preferential expression of a particular V beta family. There was a good correlation between PCR data and the expression of V beta-family protein products observed by immunohistochemistry on tissue sections of the T-cell lymphomas. All T-cell lines, three cases of MF, and three cases of nodal T-cell NHL showed a rearrangement of the TCR beta chain on DNA level.

PMID: 1331246 [PubMed - indexed for MEDLINE]



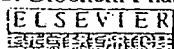
Matrilin-3 in human articular cartilage: increased expression in osteoarthritis.

Pullig O, Weseloh G, Klatt AR, Wagener R, Swoboda B.

Division of Orthopaedic Rheumatology, Department of Orthopaedics, University of Erlangen-Nuremberg, Rathsberger Str. 57, D-91054 Erlangen, Germany.
oliver.pullig@med.uni-erlangen.de

OBJECTIVE: Matrilin-3 is a member of the recently described matrilin family of extracellular matrix proteins containing von Willebrand factor A-like domains. The matrilin-3 subunit can form homo-tetramers as well as hetero-oligomers together with subunits of matrilin-1 (cartilage matrix protein). It has a restricted tissue distribution and is strongly expressed in growing skeletal tissues. Detailed information on expression and distribution of extracellular matrix proteins is important to understand cartilage function in health and in disease like osteoarthritis (OA). **METHODS:** Normal and osteoarthritic cartilage were systematically analysed for matrilin-3 expression, using immunohistochemistry, Western blot analysis, in situ hybridization, and quantitative PCR. **RESULTS:** Our results indicate that matrilin-3 is a mandatory component of mature articular cartilage with its expression being restricted to chondrocytes from the tangential zone and the upper middle cartilage zone. Osteoarthritic cartilage samples with only moderate morphological osteoarthritic degenerations have elevated levels of matrilin-3 mRNA. In parallel, we found an increased deposition of matrilin-3 protein in the cartilage matrix. Matrilin-3 staining was diffusely distributed in the cartilage matrix, with no cellular staining being detectable. In cartilage samples with minor osteoarthritic lesions, matrilin-3 deposition was restricted to the middle zone and to the upper deep zone. A strong correlation was found between enhanced matrilin-3 gene and protein expression and the extent of tissue damage. Sections with severe osteoarthritic degeneration showed the highest amount of matrilin-3 mRNA, strong signals in in situ hybridization, and prominent protein deposition in the middle and deep cartilage zone. **CONCLUSION:** We conclude that matrilin-3 is an integral component of human articular cartilage matrix and that the enhanced expression of matrilin-3 in OA may be a cellular response to the modified microenvironment in the disease. Copyright 2002 OsteoArthritis Research Society International.

PMID: 11950247 [PubMed - indexed for MEDLINE]



Up-regulation of mitochondrial peripheral benzodiazepine receptor expression by tumor necrosis factor alpha in testicular leydig cells. Possible involvement in cell survival.

Rey C, Mauduit C, Naureils O, Benahmed M, Louisot P, Gasnier F.

INSERM U. 189, Faculte de Medecine Lyon-Sud, BP12, 69921 cedex, Oullins, France.

Porcine Leydig cells in primary cultures are resistant to tumor necrosis factor alpha (TNFalpha) cytotoxicity. Here we report that these cells can be rendered sensitive to TNFalpha killing by treatment with the translational inhibitor cycloheximide, suggesting the existence of proteins that can suppress the death stimulus induced by the cytokine. In search of these cytoprotective proteins, we focused on the constituents of the mitochondrial permeability transition pore (PT pore), whose opening has been shown to play a critical role in the TNFalpha-mediated death pathway. We found that TNFalpha up-regulated mRNA and protein expression of the mitochondrial peripheral benzodiazepine receptor (PBR), an outer membrane-derived constituent of the pore. A strong correlation was established between the resistance of the cells to TNFalpha killing and the density of PBR-binding sites. Concomitantly, TNFalpha down-regulated Bcl-2 mRNA and protein expression. As Bcl-2 has been shown to be an endogenous inhibitor of the PT pore, we hypothesize that the TNFalpha-induced up-regulation of PBR expression may compensate for the decrease in Bcl-2 levels to prevent the opening of the PT pore.

PMID: 11077046 [PubMed - indexed for MEDLINE]

MetaPress

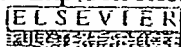
GLUT1 messenger RNA and protein induction relates to the malignant transformation of cervical cancer.

Rudlowski C, Becker AJ, Schroder W, Rath W, Buttner R, Moser M.

Dept of Gynecology and Obstetrics, University Hospital Heidelberg, Vossstr 7-9, D-69115 Heidelberg, Germany.

We studied whether induction of glucose transporters (GLUTs) 1 to 4 correlates with human papillomavirus (HPV)-dependent malignant transformation of cervical epithelium. Tissue samples of cervical intraepithelial neoplasia (CIN; grades 1 to 3), invasive carcinomas, and lymph node metastasis were examined. HPV typing was performed. Tissue sections were immunostained with GLUT1 to GLUT4 antibodies. Messenger RNA (mRNA) in situ hybridization confirmed GLUT1 protein expression. Weak expression of GLUT1 was found in nondysplastic HPV-positive and HPV-negative epithelium; significant expression was observed in preneoplastic lesions, correlating with the degree of dysplasia. In CIN 3 high-risk HPV lesions, cervical cancer, and metastasis, GLUT1 was expressed at highest levels with a strong correlation of GLUT1 mRNA and protein expression. Immunostains for GLUT2 to GLUT4 were negative. Cervical tumor cells respond to enhanced glucose utilization by up-regulation of GLUT1. The strong induction of GLUT1 mRNA and protein in HPV-positive CIN 3 lesions suggests GLUT1 overexpression as an early event in cervical neoplasia. GLUT1 is potentially relevant as a diagnostic tool and glucose metabolism as a therapeutic target in cervical cancer.

PMID: 14608894 [PubMed - indexed for MEDLINE]



Expression and distribution of laminin alpha1 and alpha2 chains in embryonic and adult mouse tissues: an immunochemical approach.

Sasaki T, Giltay R, Talts U, Timpl R, Talts JF.

Max-Planck-Institute for Biochemistry, Martinsried, D-82152, Germany.

Protein levels, mRNA expression, and localization of laminin alpha1 and alpha2 chains in development and in adult mice were examined. Recombinant fragments were used to obtain high-titer-specific polyclonal antibodies for establishing quantitative radioimmuno-inhibition assays. This often demonstrated an abundance of alpha2 chain, but also distinct amounts of alpha1 chain for adult tissues. The highest amounts of alpha1 were found in placenta, kidney, testis, and liver and exceeded those of alpha2. All other tissue extracts showed a higher content of alpha2, which was particularly high in heart and muscle when compared to alpha1. Content of gamma1 chain, shared by most laminins, was also analyzed. This demonstrated gamma1 chain levels being equal to or moderately exceeding the sum of alpha1 and alpha2 chains, indicating that these isoforms represent the major known laminin isoforms in most adult mouse tissues so far examined. Moreover, we found good correlation between radioimmuno-inhibition data and mRNA levels of adult tissues as measured by quantitative real-time reverse transcriptase-PCR. Embryonic tissues were also analyzed by radioimmuno-inhibition assays. This demonstrated for day 11 embryos comparable amounts of alpha1 and gamma1 and a more than 25-fold lower content of alpha2. This content increased to about 10% of alpha1 in day 13 embryos. The day 18 embryo showed in heart, kidney, and liver, but not yet in brain and lung, alpha1/alpha2 chain ratios comparable to those in adult tissues. Immunostaining demonstrated alpha1 in Reichert's membrane (day 7.5), while alpha2 could not be detected before day 11.5. These data were compared with immunohistochemical localization results on several more embryonic and adult tissue sections. Our results regarding localization are consistent with those of earlier work with some notable exceptions. This was in part due to epitope masking for monoclonal antibodies commonly used in previous studies in esophagus, intestine, stomach, liver, kidney, and spleen.

PMID: 11969289 [PubMed - indexed for MEDLINE]



Discordant regulation of granzyme H and granzyme B expression in human lymphocytes.

Sedelies KA, Sayers TJ, Edwards KM, Chen W, Pellicci DG, Godfrey DI, Trapani JA.

Cancer Immunology Laboratory, Peter MacCallum Cancer Centre, Locked Bag 1, A'Beckett Street, East Melbourne, 8006, Australia.

We analyzed the expression of granzyme H in human blood leukocytes, using a novel monoclonal antibody raised against recombinant granzyme H. 33-kDa granzyme H was easily detected in unfractionated peripheral blood mononuclear cells, due to its high constitutive expression in CD3(-)CD56(+) natural killer (NK) cells, whereas granzyme B was less abundant. The NK lymphoma cell lines, YT and Lopez, also expressed high granzyme H levels. Unstimulated CD4(+) and particularly CD8(+) T cells expressed far lower levels of granzyme H than NK cells, and various agents that classically induce T cell activation, proliferation, and enhanced granzyme B expression failed to induce granzyme H expression in T cells. Also, granzyme H was not detected in NK T cells, monocytes, or neutrophils. There was a good correlation between mRNA and protein expression in cells that synthesize both granzymes B and H, suggesting that gzmH gene transcription is regulated similarly to gzmB. Overall, our data indicate that although the gzmB and gzmH genes are tightly linked, expression of the proteins is quite discordant in T and NK cells. The finding that granzyme H is frequently more abundant than granzyme B in NK cells is consistent with a role for granzyme H in complementing the pro-apoptotic function of granzyme B in human NK cells.

PMID: 15069086 [PubMed - indexed for MEDLINE]

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BCL2 protein expression parallels its mRNA level in normal and malignant B cells.

Shen Y, Iqbal J, Huang JZ, Zhou G, Chan WC.

Department of Pathology and Microbiology, University of Nebraska Medical Center, Omaha, USA.

The regulation of B-cell lymphoma 2 (BCL2) protein expression in germinal center (GC) B cells has been controversial. Previous reports have indicated posttranscriptional regulation plays a dominant role. However, a number of recent studies contradicted these reports. Using real-time polymerase chain reaction (PCR) and Standardized Reverse Transcriptase-PCR (StaRT-PCR), we measured the level of mRNA expression in GC, mantle zone (MNZ), and marginal zone (MGZ) cells from laser capture microdissection. Both quantitative RT-PCR measurements of microdissected GC cells from tonsils showed that GC cells had low expression of BCL2 transcripts commensurate with the low protein expression level. These results are in agreement with microarray studies on fluorescence-activated cell sorter (FACS)-sorted cells and microdissected GC cells. We also examined BCL2 mRNA and protein expression on a series of 30 cases of diffuse large B-cell lymphoma (DLBCL) and found, in general, a good correlation. The results suggested that BCL2 protein expression is regulated at the transcriptional level in normal B cells and in the neoplastic cells in most B-cell lymphoproliferative disorders.

PMID: 15242877 [PubMed - indexed for MEDLINE]

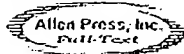
Quantitative determinations of the steady state transcript levels of hexokinase isozymes and glucose transporter isoforms in normal rat tissues and the malignant tumor cell line AH130.

Shinohara Y, Yamamoto K, Inoo K, Yamazaki N, Terada H.

Faculty of Pharmaceutical Sciences, University of Tokushima, Japan.
yasuo@ph.tokushima-u.ac.jp

The steady state transcript levels of the four hexokinase (HK) isozymes and four glucose transporter (GLUT) isoforms were determined quantitatively by Northern analysis of RNA samples from rat tissues using synthetic fragments of the RNAs encoding the HK isozymes and GLUT isoforms. Results showed that the levels of HK isozyme transcripts were low in rat tissues, the level of that most highly expressed, the type I isozyme (HKI), in the brain being 0.025% of the total poly(A)+ RNA. A good correlation was found between the reported HK activities and the total amounts of transcripts encoding all HK isozymes in various tissues, showing that the HK activities in tissues can be estimated from the total amount of transcripts encoding HK isozymes. The proposed associated expressions of HK isozymes and GLUT isoforms in particular tissues were confirmed at their transcript levels. The steady state transcript levels of type II HK and the type I GLUT isoform in the malignant tumor cell line AH130 were also determined quantitatively.

PMID: 9459591 [PubMed - indexed for MEDLINE]



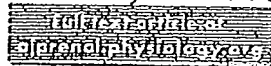
UVA irradiation-induced activation of activator protein-1 is correlated with induced expression of AP-1 family members in the human keratinocyte cell line HaCaT.

Silvers AL, Bowden GT.

Department of Radiation Oncology, Arizona Cancer Center, The University of Arizona, Tucson 85724, USA.

To determine whether the transcription factor activator protein-1 (AP-1) could be modulated by ultraviolet A (UVA) exposure, we examined AP-1 DNA-binding activity and transactivation after exposure to UVA in the human immortalized keratinocyte cell line HaCaT. Maximal AP-1 transactivation was observed with 250 kJ/m² UVA between 3 and 4 h after irradiation. DNA binding of AP-1 to the target 12-O-tetradecanoylphorbol-13-acetate response element sequence was maximally induced 1-3 h after irradiation. Both de novo transcription and translation contributed to the UVA-induced AP-1 DNA binding. c-Fos was implicated as a primary component of the AP-1 DNA-binding complex. Other components of the complex included Fra-2, c-Jun, JunB and JunD. UVA irradiation induced protein expression of c-Fos, c-Jun, Fra-1 and Fra-2. Phosphorylated forms of these induced proteins were determined at specific time points. A strong correlation existed between UVA-induced AP-1 activity and accumulation of c-Fos, c-Jun and Fra-1 proteins. UVA irradiation also induced c-fos and c-jun mRNA expression and transcriptional activation of the c-fos gene promoter. These results demonstrate that UVA irradiation activates AP-1 and that c-fos induction may play a critical role in the response of these human keratinocytes to UVA irradiation.

PMID: 11950097 [PubMed - indexed for MEDLINE]



Rat kidney glutamyl aminopeptidase (aminopeptidase A): molecular identity and cellular localization.

Song L, Ye M, Troyanovskaya M, Wilk E, Wilk S, Healy DP.

Department of Pharmacology, Mount Sinai School of Medicine, City University of New York, New York 10029.

Glutamyl aminopeptidase [aminopeptidase A (EAP), EC 3.4.11.7] is an ectoenzyme that selectively hydrolyzes acidic amino acid residues from the amino terminus of oligopeptides. EAP activity is highest within the kidney and small intestine. The murine pre-B cell BP-1/6C3 and the human kidney glycoprotein gp160 differentiation antigens have been reported to have biochemical properties indistinguishable from EAP. It is not known, however, if rat kidney EAP is a homologue of these antigens or molecularly distinct. Using the reverse transcription-polymerase chain reaction method with oligonucleotide primers based on the BP-1/6C3 nucleotide sequence, we isolated a 450-bp partial cDNA from rat kidney poly(A)⁺ RNA. The partial cDNA encoded a predicted protein that was 92% and 86% identical to the murine BP-1/6C3 and human gp160 antigens, respectively; the amino acid sequence within the zinc-binding domain was completely conserved. Purification of EAP from rat kidney and microsequence analysis of a tryptic digest peptide fragment (18-mer) indicated that the fragment was highly similar to a region within the BP-1/6C3 and gp160 proteins. Northern blot hybridization and immunoblot analyses were also consistent with labeling of products the same size as reported for the BP-1/6C3 and gp160 antigens. There was a good correlation between the cellular distribution of EAP mRNA and EAP immunoreactivity, with proximal tubules and glomerular mesangial cells having the highest densities. These results indicate that rat kidney EAP is a species homologue of the murine BP-1/6C3 and human gp160 antigens. Furthermore, on the basis of its cellular localization, rat kidney EAP is likely to be involved in degradation of oligopeptides within the glomerulus and the glomerular filtrate. Since cells that express EAP also express receptors for angiotensin II, an intrarenal vasoactive hormone that is a substrate for EAP, these results further suggest that EAP may play a role in modulating the activity of intrarenal angiotensin II.

PMID: 7943354 [PubMed - indexed for MEDLINE]

Tumor necrosis factor-alpha upregulates the prostaglandin E2 EP1 receptor subtype and the cyclooxygenase-2 isoform in cultured amnion WISH cells.

Spaziani EP, Benoit RR, Tsibris JC, Gould SE, O'Brien WF.

University of South Florida Health Science Center, Department of Obstetrics & Gynecology, Tampa 33612, USA. espazian@com1.med.usf.edu

Recent studies have demonstrated a strong correlation between infection and preterm labor. Preterm delivery is also associated with high levels of cytokines and prostaglandins in amniotic fluid. The purpose of this study was to investigate the effect of tumor necrosis factor-alpha (TNF-alpha) on the levels of cyclooxygenase, prostaglandin E2 production (PGE2), and expression of the PGE2 receptor subtype EP1 in amnion WISH cell culture. Amnion WISH cell cultures were incubated in increasing concentrations of TNF-alpha (0-50 ng/ml). Changes in cyclooxygenase and EP1 receptor proteins were evaluated by Western blot analysis. Changes in EP1 mRNA were evaluated by Northern blot, and culture fluid concentrations of PGE2 were estimated by enzyme immunoassay (EIA). EP1 protein ($p < 0.01$), EP1 mRNA ($p < 0.05$), cyclooxygenase-2 (COX-2) protein ($p < 0.001$), and PGE2 concentrations ($p < 0.01$) all increased with increasing concentrations of TNF-alpha. Changes in COX-1 protein were not observed following TNF-alpha-incubation. The results suggest that TNF-alpha may play a role in infection-induced preterm labor by its pleiotropic ability to simultaneously stimulate COX-2 activity, PGE2 concentrations, and PGE2 EP1 receptor levels in human amnion.

PMID: 9877447 [PubMed - indexed for MEDLINE]

111: Skin Pharmacol Appl Skin Physiol. 2003 May-Jun;16(3):143-50. Related Articles, Links



Transcriptional activity of potent glucocorticoids: relevance of glucocorticoid receptor isoforms and drug metabolites.

Spika I, Hammer S, Kleuser B, Korting HC, Schafer-Korting M.

Institut für Pharmazie, Abteilung für Pharmakologie und Toxikologie, Freie Universität Berlin, Berlin, Germany.

As compared to standard glucocorticoids (GC), prednicarbate (PC) is favorable in the treatment of eczema due to its high benefit/risk ratio. The remarkable anti-inflammatory effects of PC are in strong contrast to its reported low glucocorticoid receptor (GR) binding affinity. In transfected COS-7 cells we related the transcriptional potencies of PC, its metabolites and conventional GC to their receptor binding properties. Moreover, the expression pattern of the human GR isoform hGRalpha and its mutual dominant negative inhibitor hGRbeta in skin cells have been investigated as well as the influence of hGRbeta on receptor binding and transactivation. hGRalpha mRNA and protein was largely overexpressed in skin cells. hGRbeta showed no influence on hGRalpha binding and transactivation. Concentration response curves indicated the greater transactivation potency of betamethasone 17-valerate followed by dexamethasone and prednisolone 17-ethylcarbonate. Native PC appeared almost as potent as dexamethasone. With both a strong correlation was observed between transactivation and GR binding. Copyright 2003 S. Karger AG, Basel

PMID: 12677094 [PubMed - indexed for MEDLINE]



Specific inhibition of AQP1 water channels in isolated rat intrahepatic bile duct units by small interfering RNAs.

Splinter PL, Masyuk AI, LaRusso NF.

Center for Basic Research in Digestive Diseases, Division of Gastroenterology and Hepatology, Mayo Medical School, Clinic, and Foundation, Rochester, Minnesota 55905, USA.

Cholangiocytes express water channels (i.e. aquaporins (AQPs)), proteins that are increasingly recognized as important in water transport by biliary epithelia. However, direct functional studies demonstrating AQP-mediated water transport in cholangiocytes are limited, in part because of the lack of specific AQP inhibitors. To address this issue, we designed, synthesized, and utilized small interfering RNAs (siRNAs) selective for AQP1 and investigated their effectiveness in altering AQP1-mediated water transport in intrahepatic bile duct units (IBDUs) isolated from rat liver. Twenty-four hours after transfection of IBDUs with siRNAs targeting two different regions of the AQP1 transcript, both AQP1 mRNA and protein expression were inhibited by 76.6-92.0 and 57.9-79.4%, respectively. siRNAs containing the same percent of base pairs as the AQP1-siRNAs but in random sequence (i.e. scrambled siRNAs) had no effect. Suppression of AQP1 expression in cholangiocytes resulted in a decrease in water transport by IBDUs in response to both an inward osmotic gradient (200 mosm) or a secretory agonist (forskolin), the osmotic water permeability coefficient ($P(f)$) decreasing up to 58.8% and net water secretion ($J(v)$) decreasing up to 87%. A strong correlation between AQP1 protein expression and water transport in IBDUs transfected with AQP1-siRNAs was consistent with the decrease in water transport by IBDUs resulting from AQP1 gene silencing by AQP1-siRNAs. This study is the first to demonstrate the feasibility of utilizing siRNAs to specifically reduce the expression of AQPs in epithelial cells and provides direct evidence of the contribution of AQP1 to water transport by biliary epithelia.

PMID: 12468529 [PubMed - indexed for MEDLINE]

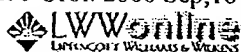
Type IV collagenase (M(r) 72,000) expression in human prostate: benign and malignant tissue.

Stearns ME, Wang M.

Department of Pathology, Medical College of Pennsylvania, Philadelphia 19129.

The expression of type IV collagenase (M(r) 72,000) has been examined in tissues from patients with benign prostatic hyperplasia (6 patients) and varying Gleason grades of malignant prostate cancer (18 patients). Immunoperoxidase labeling indicated that expression of the type IV collagenase was weak or nonexistent in benign tissue but consistently strong in the glandular and ductal epithelial cells of prostate tumors diagnosed at Gleason grades 1-8. In moderate to advanced cancer (i.e., Gleason grades 2 to 8), invasive tumor foci in the stromal tissue produced relatively modest amounts of type IV collagenase. The normal stromal tissue (i.e., fibroblasts) uniformly failed to produce detectable levels of type IV collagenase in the 24 patients examined. Northern and quantitative slot blot hybridization assays demonstrated that collagenase type IV mRNA levels were low in benign tissue and high in malignant tumors. In contrast, the stromal cells did not express significant amounts of type IV collagenase mRNA. Enzyme-linked immunosorbent assays demonstrated that the amounts of type IV collagenase protein correlated directly with the mRNA levels in the tumor tissue. The studies suggest that type IV collagenase may be selectively overexpressed by malignant, preinvasive prostatic epithelial cells.

PMID: 7679051 [PubMed - indexed for MEDLINE]



The decompensated detrusor III: impact of bladder outlet obstruction on sarcoplasmic endoplasmic reticulum protein and gene expression.

Stein R, Gong C, Hutcheson JC, Canning DA, Zderic SA.

Division of Urology, Children's Hospital of Philadelphia, Philadelphia, Pennsylvania, USA.

PURPOSE: Regulation of calcium ion homeostasis has a significant role in smooth muscle contractility. The sarcoplasmic endoplasmic reticulum, calcium, magnesium, adenosine triphosphatase (SERCA) is a regulatory ion pump that may have a role in the functional outcome after outlet obstruction. We investigate what correlation if any existed between SERCA protein and gene expression, and the contractile properties in the same bladder. **MATERIALS AND METHODS:** Standardized partial bladder outlet obstructions were created in adult New Zealand white rabbits, which were divided into control, sham operated and obstructed groups. Muscle strip studies subcategorized the obstructed group into compensated (force greater than 50% of control) and decompensated (force less than 50% of control). Microsomal membrane and total RNA fractions were prepared from the same bladder tissue. Membrane proteins were used for Western blot analysis using a SERCA specific monoclonal antibody, and total RNA was assessed with Northern blot analysis. **RESULTS:** The relative intensities of signals for the Western and Northern blots demonstrated a strong correlation between protein and gene expression. Furthermore there was a strong association between the loss of SERCA messenger RNA and protein expression and loss of bladder function. **CONCLUSIONS:** Bladder contractility after outlet obstruction is influenced in part by smooth muscle cell ability to maintain calcium homeostasis via SERCA. The loss of SERCA protein expression is mediated by down-regulation in gene expression in the same bladder. These data suggest that smooth muscle ion pump gene expression is in part mechanically (pressure work) regulated.

PMID: 10958733 [PubMed - indexed for MEDLINE]

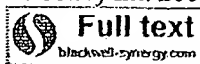
TNF-alpha and IL-8 are upregulated in the epidermis of normal human skin after UVB exposure: correlation with neutrophil accumulation and E-selectin expression.

Strickland I, Rhodes LE, Flanagan BF, Friedmann PS.

Department of Dermatology, University of Liverpool, United Kingdom.

The in vivo response to ultraviolet B (UVB) radiation in skin is characterized by the accumulation of both mononuclear and polymorphonuclear cells within the dermis and an induction of vascular endothelial adhesion molecules. Epidermal production of cytokines (IL-8 and TNF-alpha) has been strongly implicated in the development of UVB-induced inflammation. In the current study, we examined the time course of IL-8 and TNF-alpha mRNA and protein expression in the epidermis over a 24-h period after in vivo UVB irradiation. Also, the induction of adhesion molecule expression and the accumulation of neutrophils within the dermis were followed. We found constitutive expression of both cytokines (mRNA and protein) in the epidermis of unirradiated skin. IL-8 was rapidly upregulated after irradiation and mRNA and protein increased at 4 h, reaching a maximum between 8 and 24 h. TNF-alpha mRNA and protein was minimally increased by 8 h after UVB irradiation and reached a maximum by 24 h. No significant alteration in ICAM-1 or VCAM-1 expression was observed. E-selectin expression, which was absent from control samples, was increased from 4 h onward and also reached a maximum at 24 h, coinciding with peak neutrophil accumulation. A strong correlation ($r = 0.96$) was found between number of E-selectin-positive vessels and numbers of infiltrating neutrophils at this time. Moreover, because E-selectin expression was increased before any apparent increase in TNF-alpha protein (4 h), TNF-alpha does not appear to be involved in the early induction of the adhesion molecule, but cytokines such as TNF-alpha and IL-8 may act subsequently to augment the inflammatory response.

PMID: 9129230 [PubMed - indexed for MEDLINE]



Basic fibroblast growth factor expression is increased in human renal fibrogenesis and may mediate autocrine fibroblast proliferation.

Strutz F, Zeisberg M, Hemmerlein B, Sattler B, Hummel K, Becker V, Muller GA.

Department of Nephrology and Rheumatology, Georg-August-University Gottingen, Germany. fstrutz@gwdg.de

BACKGROUND: Interstitial fibroblasts play a critical role in renal fibrogenesis, and autocrine proliferation of these cells may account for continuous matrix synthesis. Basic fibroblast growth factor (FGF-2) is mitogenic for most cells and exerts intracrine, autocrine, and paracrine effects on epithelial and mesenchymal cells. The aims of the present studies were to localize and quantitate the expression of FGF-2 in normal and pathologic human kidneys and to study the *in vitro* effects of FGF-2 on proliferation, differentiation, and matrix production of isolated cortical kidney fibroblasts. **METHODS:** FGF-2 protein expression was localized by immunofluorescence double labelings in normal and fibrotic human kidneys. Subsequently, interstitial FGF-2 labeling was determined semiquantitatively in 8 normal kidneys and 39 kidneys with variable degrees of interstitial fibrosis and was correlated with the morphometrically determined interstitial cortical volume. In addition, FGF-2 expression was quantitated by immunoblot analysis in three normal and six fibrotic kidneys. FGF-2 mRNA was localized by *in situ* hybridizations. Seven primary cortical fibroblast lines were established, and expression of FGF-2 and FGF receptor-1 (FGFR-1) were examined. The effects of FGF-2 on cell proliferation were determined by bromodeoxyuridine incorporation and cell counts, those on differentiation into myofibroblasts by staining for alpha-smooth muscle actin, and those on matrix synthesis by enzyme-linked immunosorbent assay for collagen type I and fibronectin. Finally, proliferative activity *in vivo* was evaluated by expression of MIB-1 (Ki-67 antigen). **RESULTS:** In normal kidneys, FGF-2 expression was confined to glomerular, vascular, and a few tubular as well as interstitial fibroblast-like cells. The expression of FGF-2 protein was increased in human kidneys, with tubulointerstitial scarring correlating with the degree of interstitial fibrosis ($r = 0.84$, $P < 0.01$). Immunoblot analyses confirmed a significant increase in FGF-2 protein expression in kidneys with interstitial scarring. *In situ* hybridization studies demonstrated low-level detection of FGF-2 mRNA in normal kidneys. However, FGF-2 mRNA expression was robustly up-regulated in interstitial and tubular cells in end-stage kidneys, indicating that these cells are the source of excess FGF-2 protein. Primary cortical fibroblasts express FGF-2 and FGFR-1 *in vitro*. FGF-2 induced a robust growth response in these cells that could be blocked specifically by a neutralizing FGF-2 antibody. Interestingly, the addition of the neutralizing antibody alone did reduce basal proliferation up to 31.5%. In addition, FGF-2 induced expression of alpha-smooth muscle actin up to 1.6-fold, but no significant effect was observed on the synthesis of collagen type I and fibronectin. Finally, staining for MIB-1 revealed a good correlation of interstitial FGF-2 positivity

with interstitial and tubular proliferative activity ($r = 0.71$, $P < 0.01$ for interstitial proliferation, $N = 30$). CONCLUSIONS: Interstitial FGF-2 protein and mRNA expression correlate with interstitial scarring. FGF-2 is a strong mitogen for cortical kidney fibroblasts and may promote autocrine fibroblast growth. Expression of FGF-2 correlates with interstitial and tubular proliferation in vivo.

PMID: 10760088 [PubMed - indexed for MEDLINE]



Adiposity elevates plasma MCP-1 levels leading to the increased CD11b-positive monocytes in mice.

Takahashi K, Mizuarai S, Araki H, Mashiko S, Ishihara A, Kanatani A, Itadani H, Kotani H.

Banyu Tsukuba Research Institute in collaboration with Merck Research Laboratories, Tsukuba, Ibaraki 300-2611, Japan.

Obesity is currently considered as an epidemic in the western world, and it represents a major risk factor for life-threatening diseases such as heart attack, stroke, diabetes, and cancer. Taking advantage of DNA microarray technology, we tried to identify the molecules explaining the relationship between obesity and vascular disorders; comparing mRNA expression of about 12,000 genes in white adipose tissue between normal, high fat diet-induced obesity (DIO) and d-Trp34 neuropeptide Y-induced obesity in mice. Expression of monocyte chemoattractant protein-1 (MCP-1) mRNA displayed a 7.2-fold increase in obese mice as compared with normal mice, leading to substantially elevated MCP-1 protein levels in adipocytes. MCP-1 levels in plasma were also increased in DIO mice, and a strong correlation between plasma MCP-1 levels and body weight was identified. We also showed that elevated MCP-1 protein levels in plasma increased the CD11b-positive monocyte/macrophage population in DIO mice. Furthermore, infusion of MCP-1 into lean mice increased the CD11b-positive monocyte population without inducing changes in body weight. Given the importance of MCP-1 in activation of monocytes and subsequent atherosclerotic development, these results suggest a novel role of adiposity in the development of vascular disorders.

PMID: 13129912 [PubMed - indexed for MEDLINE]

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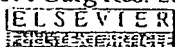
Augmented expression of neuronal nitric oxide synthase in the atria parasympathetically decreases heart rate during acute myocardial infarction in rats.

Takimoto Y, Aoyama T, Tanaka K, Keyamura R, Yui Y, Sasayama S.

Department of Cardiovascular Medicine, Graduate School of Medicine, Kyoto University, Kyoto, Japan.

BACKGROUND: Nitric oxide (NO) synthesized within sinoatrial cells recently has been shown to participate in the autonomic control of heart rate. We hypothesized that NO in the neuronal cells in the heart was increased and parasympathetically regulated heart rate after myocardial infarction (MI). **METHODS AND RESULTS:** We examined heart rate dynamics and neuronal NO synthase (nNOS) expression and activities in the atria of rats with MI 1, 3, 7, and 14 days after MI (n=7 to 22 for each group). Both the mRNA levels of nNOS in the atria determined by competitive reverse transcriptase-polymerase chain reaction and the protein levels determined by Western blotting were significantly increased compared with controls 1, 3, and 7 days after MI. nNOS activity in the atria 1 day after infarction was also increased in MI rats. nNOS immunoreactivity was observed in nerve fibers in the atria. After infusion of a specific inhibitor of nNOS and iNOS, 1-(2-trifluoromethylphenyl) imidazole (TRIM) (50 mg/kg IV), heart rate was significantly ($P<0.01$) increased in MI rats compared with controls 1, 3, and 7 days after MI. The iNOS-specific inhibitor, 1400W (10 mg/kg SC), did not significantly affect the heart rate in rats with MI. The effect of TRIM was abolished by pretreatment with L-arginine (25 mg/kg IV) or by parasympathetic blockade with atropine but not by propranolol. There was a strong correlation ($r=0.837$, $P<0.0001$) between the nNOS protein expression and heart rate change after TRIM infusion. **CONCLUSIONS:** These results indicate that increased nNOS parasympathetically decreased heart rate via the production of NO in rats with acute MI.

PMID: 11815433 [PubMed - indexed for MEDLINE]



Differential upregulation of cellular adhesion molecules at the sites of oxidative stress in experimental acute pancreatitis.

Telek G, Ducroc R, Scoazec JY, Pasquier C, Feldmann G, Roze C.

INSERM U 410, Universite Paris 7 Denis Diderot, 75870 Paris, France.

BACKGROUND: Severe acute pancreatitis (AP)(2) is associated with exaggerated leukocyte adherence and activation. Endothelial cellular adhesion molecules (CAMs) can be induced by cytokines, but also directly by oxygen free radicals (OFRs), mediated by nuclear factor kappa-B (NF-kappa B). We investigated the behavior of inducible CAMs in relation to pancreatic oxidative stress. Our novel modification of cerium capture histochemistry (reaction of OFRs with cerium produces laser reflective Ce perhydroxide precipitates) combined with reflectance confocal laser scanning microscopy (CLSM) allows the histological codemonstration of in vivo OFR production and immunolabeled CAMs, or NF-kappa B. **METHODS:** Taurocholate AP was induced in rats; sham operated and normal animals served as controls. To achieve in situ, in vivo reaction of cerium with OFRs, animals were perfused with CeCl(3) solution at different time points (1, 2, 8, 24 h) and then sacrificed. E-selectin, P-selectin, ICAM-1, VCAM, and NF-kappa B p65 were labeled by immunofluorescence (IF) on frozen sections of cerium perfused pancreata. IF and Ce perhydroxide reflectance were simultaneously detected by CLSM. Pancreatic gene expression of the same CAMs was quantified by competitive RT-PCR (MIMIC internal control). **RESULTS:** Control pancreata showed negligible reflectance and minimal CAM expression. Early (1, 2 h) AP samples were characterized by intense, heterogeneous acinar OFR production, strong P-selectin, and increasing ICAM expression, with nuclear translocation of p65, histologically all colocalizing with the areas of acinar oxidative stress. Adherent polymorphonuclear leukocytes (PMNs) displayed weak OFR formation. Later (8, 24 h), a slowly declining P-selectin, but persisting ICAM-1 expression, was paralleled by widespread adherence of PMNs producing surprisingly large amounts of OFRs. VCAM and E-selectin showed a mild increase at 24 h. CAM gene activation was in good correlation with the protein expression. **CONCLUSIONS:** The early acinar oxidative stress is colocalized with NF-kappa B activation, preferential P-selectin, and ICAM upregulation in this AP model. Subsequently, adherent, activated PMNs become the major source of OFRs, thereby contributing to tissue damage. Copyright 2001 Academic Press.

PMID: 11180997 [PubMed - indexed for MEDLINE]

Myotonic dystrophy: an unstable CTG repeat in a protein kinase gene.

Timchenko L, Monckton DG, Caskey CT.

Department of Molecular and Human Genetics, Baylor College of Medicine, Texas Medical Center, Houston 77030, USA.

Myotonic dystrophy (DM) is caused by the amplification of CTG repeats in the 3' untranslated region of a gene encoding a protein homologous to serine/threonine protein kinases. In DM patients the CTG repeats are extremely unstable, varying in length from patient to patient and generally increasing in length in successive generations. There is a strong correlation between the size of the repeats and the age of onset and severity of the disease. The molecular basis of the effect of the CTG expansion on the development of the DM phenotype continues to be investigated. The first working hypothesis of the molecular mechanism of DM was a reduction in steady-state myotonin-protein kinase (Mt-PK) mRNA and protein levels. However, although the consensus finding is that the Mt PK mRNA and protein levels are decreased in DM patients, it is still not clear if this reduction leads directly to the DM phenotype. In this short review we discuss the molecular aspects of CTG instability and the expression of the myotonin-protein kinase gene in normal and DM populations.

Publication Types:

- [Review](#)
- [Review, Tutorial](#)

PMID: 7620117 [PubMed - indexed for MEDLINE]

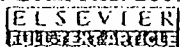
Induction of class 3 aldehyde dehydrogenase in the mouse hepatoma cell line Hepa-1 by various chemicals.

Torronen R, Korkalainen M, Karenlampi SO.

Department of Physiology, University of Kuopio, Finland.

The mouse hepatoma cell line Hepa-1 was shown to express an aldehyde dehydrogenase (ALDH) isozyme which was inducible by TCDD and carcinogenic polycyclic aromatic hydrocarbons. The induced activity could be detected with benzaldehyde as substrate and NADP as cofactor (B/NADP ALDH). As compared with rat liver and hepatoma cell lines, the response was moderate (maximally 5-fold). There was an apparent correlation between this specific form of ALDH and aryl hydrocarbon hydroxylase (AHH) in the Hepa-1 wild-type cell line--in terms of inducibility by several chemicals. However, the magnitude of the response was clearly smaller for ALDH than for AHH. Southern blot analysis showed that a homologous gene (class 3 ALDH) was present in the rat and mouse genome. The gene was also expressed in Hepa-1, and there was a good correlation between the increase of class 3 ALDH-specific mRNA and B/NADP ALDH enzyme activity after exposure of the Hepa-1 cells to TCDD. It is concluded that class 3 ALDH is inducible by certain chemicals in the mouse hepatoma cell line, although the respective enzyme is not inducible in mouse liver in vivo.

PMID: 1505055 [PubMed - indexed for MEDLINE]



Relationship between cyclin D1 and p21(Waf1/Cip1) during differentiation of human myeloid leukemia cell lines.

Ullmannova V, Stockbauer P, Hradcova M, Soucek J, Haskovec C.

Department of Molecular Genetics, Institute of Hematology and Blood Transfusion, U Nemocnice 1, 128 20 Prague 2, Czech Republic. ullman@uhkt.cz

Expression of cell cycle-regulating genes was studied in human myeloid leukemia cell lines ML-1, ML-2 and ML-3 during induction of differentiation in vitro. Myelomonocytic differentiation was induced by phorbol ester (12-o-Tetradecanoyl-phorbol-13-acetate, TPA), tumor necrosis factor alpha (TNFalpha) or interferon gamma (INFgamma), or their combination. Differentiation (with the exception of TNFalpha alone) was accompanied by inhibition of DNA synthesis and cell cycle arrest. Inhibition of proliferation was associated with a decrease in the expression of cdc25A and cdc25B, cdk6 and Ki-67 genes, and with increased p21(Waf1/Cip1) gene expression, as measured by comparative RT-PCR. Expression of the following genes was not changed after induction of differentiation: cyclin A1, cyclin D3, cyclin E1 and p27(Kip1). Surprisingly, cyclin D1 expression was upregulated after induction by TPA, TNFalpha with IFNgamma or BA. Cyclin D2 was upregulated only after induction by BA. The results of the expression of the tested genes obtained by comparative RT-PCR were confirmed by quantitative real-time (RQ) RT-PCR and Western blotting. Quantitative RT-PCR showed as much as a 288-fold increase of cyclin D1 specific mRNA after a 24h induction by TPA. The upregulation of cyclin D1 in differentiating cells seems to be compensated by the upregulation of p21(Waf1/Cip1). These results, besides others, point to a strong correlation between the expression of cyclin D1 and p21(Waf1/Cip1) on the one hand and differentiation on the other hand in human myeloid leukemic cells and reflect a rather complicated network regulating proliferation and differentiation of leukemic cells.

PMID: 12921950 [PubMed - indexed for MEDLINE]



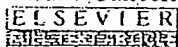
Intestinal carbamoyl phosphate synthase I in human and rat. Expression during development shows species differences and mosaic expression in duodenum of both species.

Van Beers EH, Rings EH, Posthuma G, Dingemans MA, Taminiou JA, Heymans HS, Einerhand AW, Buller HA, Dekker J.

Pediatric Gastroenterology and Nutrition, Department Pediatrics, Emma Children's Hospital, Academic Medical Center, Amsterdam, The Netherlands.

The clinical importance of carbamoyl phosphate synthase I (CPSI) relates to its capacity to metabolize ammonia, because CPSI deficiencies cause lethal serum ammonia levels. Although some metabolic parameters concerning liver and intestinal CPSI have been reported, the extent to which enterocytes contribute to ammonia conversion remains unclear without a detailed description of its developmental and spatial expression patterns. Therefore, we determined the patterns of enterocytic CPSI mRNA and protein expression in human and rat intestine during embryonic and postnatal development, using in situ hybridization and immunohistochemistry. CPSI protein appeared during human embryogenesis in liver at 31-35 e. d. (embryonic days) before intestine (59 e.d.), whereas in rat CPSI detection in intestine (at 16 e.d.) preceded liver (20 e.d.). During all stages of development there was a good correlation between the expression of CPSI protein and mRNA in the intestinal epithelium. Strikingly, duodenal enterocytes in both species exhibited mosaic CPSI protein expression despite uniform CPSI mRNA expression in the epithelium and the presence of functional mitochondria in all epithelial cells. Unlike rat, CPSI in human embryos was expressed in liver before intestine. Although CPSI was primarily regulated at the transcriptional level, CPSI protein appeared mosaic in the duodenum of both species, possibly due to post-transcriptional regulation.

PMID: 9446830 [PubMed - indexed for MEDLINE]



Expression of deoxycytidine kinase in leukaemic cells compared with solid tumour cell lines, liver metastases and normal liver.

van der Wilt CL, Kroep JR, Loves WJ, Rots MG, Van Groenigen CJ, Kaspers GJ, Peters GJ.

Department of Medical Oncology, VU University Medical Center, Amsterdam, The Netherlands.

Deoxycytidine kinase (dCK) is required for the phosphorylation of several deoxyribonucleoside analogues that are widely employed as chemotherapeutic agents. Examples include cytosine arabinoside (Ara-C) and 2-chlorodeoxyadenosine (CdA) in the treatment of acute myeloid leukaemia (AML) and gemcitabine to treat solid tumours. In this study, expression of dCK mRNA was measured by a competitive template reverse transcriptase polymerase chain reaction (CT RT-PCR) in seven cell lines of different histological origin, 16 childhood and adult AML samples, 10 human liver samples and 11 human liver metastases of colorectal cancer origin. The enzyme activity and protein expression levels of dCK in the cell lines were closely related to the mRNA expression levels ($r=0.75$, $P=0.026$ and $r=0.86$, $P=0.007$). In AML samples, dCK mRNA expression ranged from 1.16 to 35.25 ($\times 10^{-3}$) dCK/beta-actin. In the cell line panel, the range was 2.97-56.9 ($\times 10^{-3}$) dCK/beta-actin of dCK mRNA expression. The enzyme activity in liver metastases was correlated to dCK mRNA expression ($r=0.497$, $P=0.05$). In the liver samples, these were not correlated. dCK mRNA expression showed only a 36-fold range in liver while a 150-fold range was observed in the liver metastases. In addition, dCK activity and mean mRNA levels were 2.5-fold higher in the metastases than in the liver samples. Since dCK is associated with the sensitivity to deoxynucleoside analogues and because of the good correlation between the different dCK measurements in malignant cells and tumours, the CT-RT PCR assay will be useful in the selection of patients that can be treated with deoxycytidine analogues.

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Expression of cytokines and growth factors in human glomerulonephritides.

Waldherr R, Noronha IL, Niemir Z, Kruger C, Stein H, Stumm G.

Department of Pathology, University of Heidelberg, Germany.

Numerous experimental studies point to the potential role of cytokines and growth factors in the pathogenesis of renal disease. However, from the various autocrine and paracrine mediators identified in vitro and in animal models, so far only a few have been demonstrated in selected human glomerulopathies. We examined two types of glomerulonephritis (GN): extracapillary GN with anti-neutrophil cytoplasmic autoantibodies (ANCA), an example of an acute form of GN, and mesangial IgA GN, usually a chronic form of GN, with immunocytochemistry, in situ hybridization and the polymerase chain reaction. Normal renal tissue from tumour nephrectomies served as a control. In ANCA-positive GN with active renal lesions (crescents, glomerular and vascular necrosis), infiltrating mononuclear cells in glomeruli and in the interstitium expressed interleukin (IL)-1 beta, tumour necrosis factor (TNF)-alpha, IL-2, interferon (IFN)-gamma, platelet-derived growth factor (PDGF) and transforming growth factor (TGF)-beta. Cytokine expression was also observed in activated resident cells, including endothelial cells, capsular epithelial cells, smooth muscle cells of vessel walls, fibroblasts and some tubular epithelial cells. In addition, we noted an increase in the cytokine and growth factor receptors TNF-R, IL-1R type II, IL-2R, IFN-gamma R and PDGF beta-R. In contrast, in mesangial IgA-GN, IL-1 beta, TNF-alpha, IFN-gamma and IL-2 were usually absent in glomeruli. Mesangial expansion in this disorder was accompanied by an increased expression of PDGF, PDGF beta-R, TGF-beta and IL-6 in mesangial areas. In both conditions a good correlation was observed between cytokine expression at the mRNA (in situ hybridization) and protein level (immunocytochemistry). (ABSTRACT TRUNCATED AT 250 WORDS)

Publication Types:

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Malignant transformation of the human endometrium is associated with overexpression of lactoferrin messenger RNA and protein.

Walmer DK, Padin CJ, Wrona MA, Healy BE, Bentley RC, Tsao MS, Kohler ME, McLachlan JA, Gray KD.

Department of Obstetrics and Gynecology, Duke University Medical Center, Durham, North Carolina 27710.

In the mouse uterus, lactoferrin is a major estrogen-inducible uterine secretory protein, and its expression correlates directly with the period of peak epithelial cell proliferation. In this study, we examine the expression of lactoferrin mRNA and protein in human endometrium, endometrial hyperplasias, and adenocarcinomas using immunohistochemistry, Western immunoblotting, and Northern and in situ RNA hybridization techniques. Our results reveal that lactoferrin is expressed in normal cycling endometrium by a restricted number of glandular epithelial cells located deep in the zona basalis. Two thirds (8 of 12) of the endometrial adenocarcinomas examined overexpress lactoferrin. This tumor-associated increase in lactoferrin expression includes an elevation in the mRNA and protein of individual cells and an increase in the number of cells expressing the protein. In comparison, only 1 of the 10 endometrial hyperplasia specimens examined demonstrates an increase in lactoferrin. We also observe distinct cytoplasmic and nuclear immunostaining patterns under different fixation conditions in both normal and malignant epithelial cells, similar to those previously reported in the mouse reproductive tract. Serial sections of malignant specimens show a good correlation between the localization of lactoferrin mRNA and protein in individual epithelial cells by in situ RNA hybridization and immunohistochemistry. Although the degree of lactoferrin expression in the adenocarcinomas did not correlate with the tumor stage, grade, or depth of invasion in these 12 patients, there was a striking inverse correlation between the presence of progesterone receptors and lactoferrin in all 8 lactoferrin-positive adenocarcinomas. In summary, lactoferrin is expressed in a region of normal endometrium known as the zona basalis which is not shed with menstruation and is frequently overexpressed by progesterone receptor-negative cells in endometrial adenocarcinomas.

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Malignant Transformation of the Human Endometrium Is Associated with Overexpression of Lactoferrin Messenger RNA and Protein

David K. Walmer,¹ Cheryl J. Padin, Mark A. Wrona, Bridget E. Healy, Rex C. Bentley, Ming-Sound Tsao, Matthew F. Kohler, John A. McLachlan, and Karen D. Gray

Department of Obstetrics and Gynecology, Duke University Medical Center, Durham, North Carolina 27710 [D. K. W., B. E. H., M. F. K.]; Department of Obstetrics and Gynecology, University of Michigan, Ann Arbor, Michigan 48109 [C. J. P.]; School of Medicine, University of Maryland, Baltimore, Maryland 21201 [M. A. W.]; Department of Pathology, Duke University Medical Center, Durham, North Carolina 27710 [R. C. B.]; Department of Pathology, Montreal General Hospital, Montreal, Quebec, Canada H3G1A4 [M.-S. T.]; Laboratory of Reproductive and Developmental Toxicology, National Institute of Environmental Health Sciences, Research Triangle Park, North Carolina 27709 [J. A. M.]; and Department of Obstetrics and Gynecology, Uniformed Services University of the Health Sciences, Bethesda, Maryland 20814-7799 [K. D. G.]

ABSTRACT

In the mouse uterus, lactoferrin is a major estrogen-inducible uterine secretory protein, and its expression correlates directly with the period of peak epithelial cell proliferation. In this study, we examine the expression of lactoferrin mRNA and protein in human endometrium, endometrial hyperplasias, and adenocarcinomas using immunohistochemistry, Western immunoblotting, and Northern and *in situ* RNA hybridization techniques. Our results reveal that lactoferrin is expressed in normal cycling endometrium by a restricted number of glandular epithelial cells located deep in the zona basalis. Two thirds (8 of 12) of the endometrial adenocarcinomas examined overexpress lactoferrin. This tumor-associated increase in lactoferrin expression includes an elevation in the mRNA and protein of individual cells and an increase in the number of cells expressing the protein. In comparison, only 1 of the 10 endometrial hyperplasia specimens examined demonstrates an increase in lactoferrin. We also observe distinct cytoplasmic and nuclear immunostaining patterns under different fixation conditions in both normal and malignant epithelial cells, similar to those previously reported in the mouse reproductive tract. Serial sections of malignant specimens show a good correlation between the localization of lactoferrin mRNA and protein in individual epithelial cells by *in situ* RNA hybridization and immunohistochemistry. Although the degree of lactoferrin expression in the adenocarcinomas did not correlate with the tumor stage, grade, or depth of invasion in these 12 patients, there was a striking inverse correlation between the presence of progesterone receptors and lactoferrin in all 8 lactoferrin-positive adenocarcinomas. In summary, lactoferrin is expressed in a region of normal endometrium known as the zona basalis which is not shed with menstruation and is frequently overexpressed by progesterone receptor-negative cells in endometrial adenocarcinomas.

INTRODUCTION

The uterus is a sex steroid-responsive organ that plays a major role in women's health. Hysterectomies were the most frequently performed major surgical procedures in a 20-year study interval (1965-1984; Ref. 1). Fifty-eight to 80% of these 12.5 million procedures were performed for estrogen-related disorders of proliferation. Chronic unopposed estrogen exposure, most commonly associated with type II ovulatory disorders, eventually leads to the development of complex endometrial hyperplasia and adenocarcinoma. Since the sex steroids, estrogen and progesterone, act on their target tissues by regulating the expression of a wide variety of signaling molecules, identifying these regulatory factors will provide critical information towards understanding normal reproduction and reproductive tract pathology. Our current knowledge of estrogen and progesterone action on the reproductive tract is based to a great extent on information collected from rodents (2). Although differences exist

between the reproductive physiology of rodents and humans, the mouse has been a useful model for studying steroid hormone action in the human female reproductive tract (3, 4). One potential regulatory molecule shown to be regulated by estrogen in the mouse reproductive tract is lactoferrin. Lactoferrin is a basic glycoprotein with an extraordinarily high affinity for iron that was originally discovered in milk. This protein is expressed in a wide variety of tissues, most notably in polymorphonuclear leukocytes and most mammalian exocrine glandular secretions. In the mammary gland (5) and the female reproductive tract of the mouse (6-8), lactoferrin is regulated by endocrine hormones. Prolactin stimulates lactoferrin synthesis in the breast; whereas in the uterus and vagina, the ovarian sex steroid, 17 β -estradiol, is the inducer. (6, 7, 9). To date, lactoferrin is one of the few genes that have been identified in the rodent that are directly regulated by estradiol. The lactoferrin gene contains an ERE² that is important for regulating its expression *in vivo* in the mouse reproductive tract. Being linked to estradiol, the expression of lactoferrin by the uterine epithelium parallels the onset of DNA synthesis. Although sequencing information suggests that the human lactoferrin gene also contains a functional imperfect ERE in the 5'-flanking promoter region (10, 11), there is very little data regarding lactoferrin expression in the human female reproductive tract.

The purpose of our study was to examine the expression of lactoferrin in the human endometrium under normal and pathological conditions by immunohistochemistry, immunoblotting, and Northern and *in situ* RNA hybridization techniques. In addition, we looked for correlations between lactoferrin expression and several parameters, such as the stage of the menstrual cycle, the distribution of estrogen and progesterone receptors, HER-2/*neu* expression, markers of cell proliferation, and the histopathological grade and extent of myometrial invasion in the adenocarcinomas. Our data demonstrates that lactoferrin is expressed in a very restricted number of glands in the basal region of normal human endometrium and is markedly overexpressed in a significant number of the uterine adenocarcinomas by PR-negative cells.

MATERIALS AND METHODS

Tissue Preparation and Histological Evaluation. Surgical pathology specimens were obtained from Duke University Medical Center (Durham, NC) and the Department of Pathology at Montreal General Hospital (Quebec, Montreal, Canada). Cycling endometrium was obtained from 22 women (ages 31-49), and atrophic endometrium was obtained from 7 postmenopausal women (ages 64-77). Hysterectomies from cycling women were performed for subserosal leiomyomas ($n = 6$), pelvic relaxation ($n = 8$), pelvic pain ($n = 4$), peritoneal endometriosis ($n = 2$), and cancer of either the exocervix ($n = 1$) or the ovary ($n = 1$). In addition, 12 adenocarcinomas, 3 atypical complex hyperplasias, 5 complex hyperplasias without atypia, and 4 simple hyperplasias were analyzed. Each human uterus was bivalved shortly after

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¹ To whom requests for reprints should be addressed, at Department of Obstetrics and Gynecology, Box 3145, Duke University Medical Center, Durham, NC 27710.

² The abbreviations used are: ERE, estrogen response element; PCNA, proliferating cell nuclear antigen; ER, estrogen receptor; PR, progesterone receptor.

hysterectomy, and endometrium was removed from the fundal region. A full thickness biopsy was placed into either 10% neutral-buffered formalin or Bouin's solution overnight at room temperature before dehydration, paraffin embedding, and sectioning at 4 μ m on silanized slides. Histological evaluations of hematoxylin and eosin-stained slides were performed blindly by one board-certified pathologist. Normal endometrial samples were dated by the criteria of Noyes *et al.* (12). Endometrial hyperplasias and carcinomas were classified according to the current recommendations of the International Society of Gynecological Pathologists under the auspices of WHO (13). Histological grading of tumors was performed according to Federation Internationale des Gynecologues et Obstetristes criteria (14). Each specimen was read a minimum of three times, and only specimens that were read consistently the same way were included in the study. Unstained sections of the same tissues were used for the cytochemical analysis of protein and mRNA expression using specific reagents. A few endometrial samples were frozen for subsequent protein and RNA extraction, which were evaluated by Western and Northern blotting, respectively. All human tissues were handled with the precautions and the guidelines required by Duke University and National Institute of Environmental Health Sciences.

Immunolocalization. Slides chosen for study were deparaffinized and rinsed in 20% glacial acetic acid at 4°C for 15 s to inhibit endogenous alkaline phosphatase. All subsequent incubations and washes were at room temperature. Sections were next equilibrated in PBS for 20 min and incubated for 20 min with 1.5% normal goat serum diluted in PBS to block nonspecific binding. Detection of lactoferrin was performed primarily with a rabbit anti-human lactoferrin polyclonal antiserum generated in our laboratory and affinity purified. Similar results were also seen with a nonaffinity-purified commercial antiserum (Biogenex, San Ramon, Ca). Following incubation at room temperature with primary antiserum for 60 min, the sections were washed in PBS twice for 10 min each, and lactoferrin was localized using an alkaline phosphatase-biotin-spectavidin detection system (Vectastain ABC-AP kit; Vector Laboratories, Burlingame, CA; or the Super Sensitive Detection System; Biogenex, San Ramon, CA). To identify nonspecific staining, preimmune rabbit serum was used in place of the primary antibody. The immunoreaction was quantitated by determining the percentage of glands and the percentage of cells staining for lactoferrin in the zona basalis and the zona functionalis, with a minimum of 300 cells counted in each region. PR antibody was provided by Geoffrey Greene (KD68), and a commercial source was also used (Biogenex, San Ramon, CA). Identical staining patterns were confirmed with both preparations. Other commercially obtained antisera include PCNA (Biogenex, San Ramon, CA), ER (ER1D5; AMAC, Westbrook, Ma), MIB-1 (AMAC), and HER-2/*neu* (Biogenex, San Ramon, CA). The primary antiserum incubations were 2 h for the PR, 1 h for PCNA, MIB-1, and ER, and 30 min for HER-2/*neu*. Antisera dilutions were 1:100 for MIB-1 and 1:20 for HER-2/*neu*. Antigen retrieval (Biogenex, San Ramon, CA) was performed before adding the progesterone primary antiserum.

Western Blot Analysis. Proteins were extracted from endometrial biopsies by homogenization on ice in 1% Triton-X and 20 mM Tris-HCl (pH 7.4) with protease inhibitors (10 μ g/ml leupeptin, 200 KU/ml aprotinin, and 20 μ g/ml phenylmethylsulfonyl fluoride) and clarified by centrifugation at 45,000 rpm for 30 min in a Beckman 70.1 Ti rotor; then the supernatant was analyzed for protein concentration by the BCA protein assay (Pierce, Rockford, IL). Aliquots of 200 μ g were separated by electrophoresis on an 8.5% SDS polyacrylamide gel, blotted onto nitrocellulose membranes, incubated with polyclonal rabbit antihuman lactoferrin antiserum, and localized with an 125 I-labeled donkey anti-rabbit immunoglobulin, as described previously (7).

In Situ Hybridization. All slides were pretreated with 0.2 N HCl for 30 min at room temperature, digested with 1 μ g/ml proteinase-K (Sigma Chemical Co., St. Louis, MO) in 0.1 M Tris-HCl (pH 7.4)-0.05 M EDTA for 15 min at 37°C, and then treated with 0.1 M methanolamine-0.25% acetic anhydride for 5 min at room temperature and 0.1 M Tris-glycine (pH 7.4) for 30 min at room temperature. The sections were subsequently dehydrated with graded ethanol, air dried, and prehybridized at 50°C for 1 h in 2 \times SSC, 10 mM DTT, 5 \times Denhardt's solution, 100 μ g/ml of both salmon sperm DNA and yeast tRNA, and 50% formamide (15). The slides were then hybridized overnight at 50°C in the same medium with 10% dextran sulfate and 2 \times 10⁶ cpm/ μ l of the specific RNA probe. The lactoferrin oligonucleotide probe was amplified by PCR using primers that spanned nucleotides 718-1654 (10) and cloned into pGEM-4Z. 32 S-labeled sense and antisense RNA probes were made with the

Promega Riboprobe kit (Promega, Madison, WI), washed twice in 1 \times SSC for 10 min at room temperature, digested with RNase [2.8 μ g/ml RNase-A, 0.3 μ g/ml RNase-T1, 10 mM Tris-HCl (pH 7.4), and 15 mM NaCl], and washed again with 1 \times SSC twice for 20 min each time at 50°C, twice for 20 in 0.1 \times SSC at 55°C, and once for 20 min at 60°C. The sections were then dehydrated and dipped in Kodak autoradiographic emulsion (NTB-2) for detection of specific mRNA expression. The slides were allowed to develop for 2 weeks. After this period, the slides were developed using Kodak D19 developer and Kodak Rapid Fixer.

Northern RNA Analysis. Total cellular RNA was purified from frozen tissue by the guanidine isothiocyanate-cesium chloride method, and poly(A)⁺ RNA was isolated by oligo(dT)-cellulose chromatography using methods described previously (15). For Northern blot analysis, poly(A)⁺ RNA was resolved by electrophoresis on 1.5% formaldehyde agarose gels, stained with ethidium bromide, and transferred to a nylon membrane. The membrane was probed with a 32 P-labeled lactoferrin cDNA derived from human uterus (nucleotides 718-1654; accession no. S52659) using PCR techniques, followed by cloning into pGEM-4Z (Promega, Madison, WI). In order to insure that the quality and quantity of RNA analyzed by Northern blotting was equivalent between control and treated groups, the blot was probed simultaneously for glyceraldehyde-3-phosphate dehydrogenase.

Statistical Analysis. Values are presented as means \pm SD. Differences between the zona basalis and functionalis were tested by the two-tailed Student's *t* test.

RESULTS

Immunohistochemical Analysis of Lactoferrin Protein

Normal Cycling Endometrium. Immunohistochemical studies of normal cycling human endometrium localize lactoferrin protein predominantly to the glandular epithelium deep in the zona basalis and not to the functionalis (Fig. 1A). The association of lactoferrin protein expression with the zona basalis is statistically significant ($P < 0.001$; Fig. 2). Two to 56% of the glands express lactoferrin at any given time during the menstrual cycle. Within positive glands, lactoferrin protein immunolocalization is heterogeneous in that positively staining epithelial cells are interspersed with cells negative for lactoferrin expression. No apparent differences in morphology, PCNA, ER, or PR expression are seen to account for the heterogeneous pattern of intra- and intergland lactoferrin expression in normal endometrium. Similar to our previous findings in mouse uterine epithelial cells, the positive-staining glandular cells of the human endometrium demonstrate two distinct immunostaining patterns for lactoferrin, cytoplasmic and nuclear (Fig. 1B), seen with both formalin and Bouin's fixation. In evaluating the temporal expression of lactoferrin, there is a trend towards more glands expressing lactoferrin during the secretory phase. Because of the large variance, the trend is not statistically significant. As expected, the polymorphonuclear leukocytes in the endometrium also stain intensely for lactoferrin protein, which is stored in their secondary granules (Fig. 3). These results demonstrate, for the first time, that lactoferrin protein is expressed in the human endometrium predominately by polymorphonuclear leukocytes and epithelial cells of glands located deep in the zona basalis.

Proliferative Endometrial Disorders: Hyperplasias and Adenocarcinomas. Immunohistochemistry reveals that the expression of lactoferrin protein is increased in 66.6% (8 of 12) of the endometrial adenocarcinomas examined. In one-half of these cases (4 of 12), lactoferrin is intensely expressed by malignant epithelial cells throughout the entire tumors (Fig. 4). The other four adenocarcinomas demonstrate increased staining for lactoferrin in concentrated regions of the tumors. In all eight cases where lactoferrin expression is elevated, the cells expressing lactoferrin have one similarity with normal positive glands in that they demonstrate heterogeneous staining of interspersed positive and negative cells. However, the expression of lactoferrin by the malignant cells clearly differs from normal

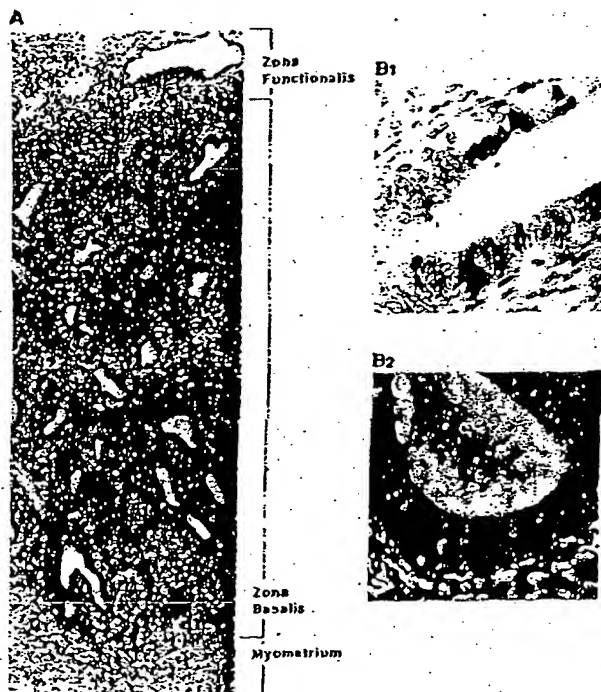


Fig. 1. Localization of lactoferrin protein in normal cycling endometrium by immunohistochemistry using a specific polyclonal antibody. Our analysis reveals that lactoferrin protein is present in a limited number of glands located in the zona basalis of the endometrium (A) $\times 10$. Also, note that lactoferrin is heterogeneous within positive glands, i.e., cells staining for lactoferrin are interspersed with negative-staining epithelial cells throughout the gland. Two immunohistochemical staining patterns are noted for lactoferrin in normal uterine epithelium. In one pattern, lactoferrin protein is immunolocalized primarily over the cytoplasm (B1), and in the other, the staining is seen over the nucleus (B2) $\times 40$.

positive glands in that the lactoferrin is not limited to the basal regions of the tumors, many more cells are positive, and the relative intensity of the staining over individual cells is increased. Although increased lactoferrin expression is associated with malignant transformation, we do not find a correlation between lactoferrin protein presence and the stage, nuclear grade, Fédération Internationale des Gynécologues et Obstétristes grade, or the depth of myometrial invasion in the 12 tumors studied (Table 1). In sharp contrast to the common dysregulation of lactoferrin expression found in the malignant endometrium, only 1 of 10 endometrial hyperplasia specimens evaluated contained an increased number of cells staining for lactoferrin. The hyperplastic specimen overexpressing lactoferrin was read as complex without atypia.

In Situ and Northern Analyses of Lactoferrin mRNA Expression in Normal and Malignant Endometrium

To further our understanding of the location of lactoferrin protein synthesis in the human endometrium, we examined lactoferrin mRNA expression by *in situ* and Northern hybridization using specific 32 S-labeled probes for human lactoferrin. No detectable RNA hybridization is observed in the normal endometrium by *in situ* hybridization, even in the presence of immunodetectable protein, using equivalent hybridization conditions and development times as used for the adenocarcinomas. Consistent with the results obtained by *in situ* hybridization, long exposure times were required to demonstrate lactoferrin mRNA in normal endometrium by Northern blot using poly(A⁺) mRNA. This indicates that lactoferrin mRNA is present in normal

tissue but in very low levels, consistent with the very limited pattern of protein expression in normal endometrium. Equivalent RNA loading and quality for each specimen was demonstrated by ethidium bromide staining of the RNA gels (data not shown) and by probing for the housekeeping gene glyceraldehyde-3-phosphate dehydrogenase that does not fluctuate significantly with the metabolic state of the tissue. A representative Northern blot is shown in Fig. 5. *In situ* hybridization with several adenocarcinomas reveals that there is a direct correlation between the localization of lactoferrin mRNA and the immunostaining of expressed lactoferrin protein (Fig. 4). Lactoferrin mRNA is not associated with polymorphonuclear leukocytes by *in situ* hybridization in either normal or malignant tissue.

Western Blot Analysis

To confirm the specificity of the antisera that we used for immunohistochemistry, we performed Western blot analysis on proteins extracted from both normal and malignant endometrium which were separated by 8.5% SDS-polyacrylamide gel electrophoresis (Fig. 6). Immunoblotting identified a single broad protein band with a molecular weight between 70,000–80,000 in both normal and neoplastic endometrial tissue homogenates, consistent with the reported molecular weight of human lactoferrin. Supporting the immunocytochemical analysis, a representative immunoblot clearly demonstrates that the proportion of protein that is lactoferrin is markedly increased in the adenocarcinomas in comparison to the normal endometrium. The molecular weight of lactoferrin in the adenocarcinomas appears to have a slightly higher molecular weight than the predominant form in normal tissue.

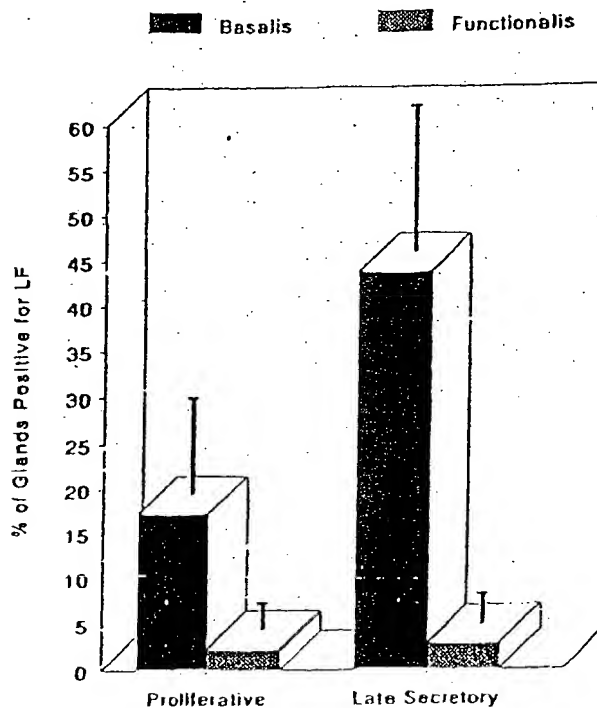


Fig. 2. Percentage of endometrial glands expressing lactoferrin protein by immunohistochemistry. Significantly more glands are positive in the region of the zona basalis than in the zona functionalis of the endometrium ($P < 0.001$). Zona basalis, \blacksquare ; zona functionalis, \square . Although there is a trend towards more of the basalis glands expressing lactoferrin in the secretory phase (right \blacksquare compared with the left \blacksquare), this was not statistically significant.

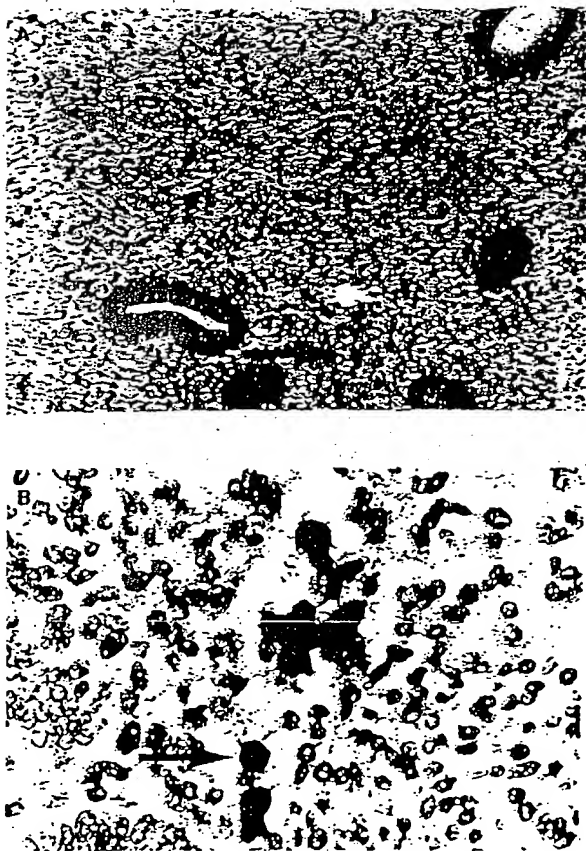


Fig. 3. Polymorphonuclear leukocytes (arrows) are scattered throughout the endometrium and stain intensely for lactoferrin. Lactoferrin is a known component of the secondary granules in polymorphonuclear leukocytes. The presence of a segmented nucleus and lactoferrin protein is an excellent method for identifying this group of inflammatory cells. A, $\times 20$. B, $\times 60$.

Correlation of Lactoferrin Expression with the Expression of PCNA, Ki-67, HER-2/*neu*, ER, and PR

In an attempt to characterize the phenotype of endometrial cells which express lactoferrin, we performed immunohistochemistry on serial sections for the Ki-67 antigen, PCNA, HER-2/*neu*, lactoferrin, ER, and PR. In normal tissue, Ki-67 and PCNA expression are cell cycle-specific markers of cell proliferation (16, 17). Upon analysis of normal cycling endometrium, no relationship between lactoferrin protein expression and ER, PR, or Ki-67 expression was observed. Similarly, in most of the adenocarcinomas evaluated, no relationship was noted between lactoferrin and PCNA protein expression. However, in one adenocarcinoma (Fig. 7), there was a clear inverse relationship seen between lactoferrin and PCNA localization, which was present throughout the entire tumor. Most dramatic, however, was a striking inverse correlation seen between lactoferrin and PR expression in 8 of 8 PR-positive uterine adenocarcinomas (Fig. 7). Two tumors negative for PR also did not express lactoferrin. Although an inverse correlation was also suggested with HER-2/*neu* and PR, the inverse correlation was more precise with lactoferrin in these tumors.

DISCUSSION

In the mouse uterus, lactoferrin is an estrogen-induced uterine secretory protein that is present throughout the epithelium (7), and it

is expressed concomitantly with epithelial cell proliferation. In contrast to lactoferrin's ubiquitous expression in the estrogenized mouse uterine epithelium, lactoferrin protein is limited to glandular epithelial cells in the basal regions of normal human endometrium and usually to glands that were directly adjacent to the myometrium (i.e., the deepest glands of the zona basalis). This regional localization of lactoferrin expression is not surprising in that other biochemical parameters have been reported to show site specificity in primate endometrium. These parameters include the proliferative index and the expression of the secretory component of IgA (18). Similar to our observations in the endometrium, lactoferrin is also expressed regionally in the mammary gland. In bovine breast tissue, lactoferrin is localized primarily to the basal alveolar cells (19); whereas in human breast tissue, the ductal epithelium appears to be the primary source of secreted lactoferrin during lactation (20).

Examining the endometrium on different days of the menstrual cycle demonstrates a trend towards increased lactoferrin expression during the luteal phase. Although this data is not statistically significant, the cyclic variation may be biologically relevant. Kim *et al.* (21) recently reported that the basal endometrial epithelial cells are unique because they proliferate during the postovulatory luteal phase. Interestingly, in the mouse uterus, there is a direct correlation between lactoferrin expression and epithelial cell proliferation (6). Therefore, lactoferrin may have a similar role in the human and mouse endometrium. If lactoferrin expression is cyclic, the ERE in the 5'-flanking promoter region of the human lactoferrin gene may be activated during the luteal phase (10, 11).

Another similarity between human and mouse uterine lactoferrin expression is the observation of two immunohistochemical staining patterns. In one pattern, the antisera binds primarily over the cytoplasm, and in the other, the nucleus is the primary site of localization. Although this could represent a fixation artifact, we have now observed this pattern in two species and under different fixation conditions. It has been demonstrated that signaling peptides, i.e., platelet-derived growth factor (22), Int-2 (23), and probasin (24), can be selectively directed into the nucleus, cytoplasm, or secreted. It is believed that this differential processing may allow proteins to have intracrine, autocrine, and paracrine roles, depending on the physiological state of the cell. The two localization patterns observed suggest that lactoferrin might have signaling sequences that direct the final destination of the mature peptide.

In normal endometrium, lactoferrin mRNA is present but in very low levels. Prolonged exposure times are needed to visualize the mRNA band with Northern analysis. Although with *in situ* hybridization the lactoferrin mRNA signal is easily seen in adenocarcinomas, we failed to localize the lactoferrin mRNA in normal endometrium using an equivalent exposure time. These low levels of mRNA in normal endometrium suggest that the synthesis and degradation of lactoferrin mRNA is more tightly regulated in normal tissue than in the adenocarcinomas. Because of the low levels of message in normal tissue, we are unable at this time to definitively conclude that lactoferrin mRNA is synthesized by the same epithelial cells which express the protein.

In endometrial adenocarcinomas, malignant transformation of the endometrium is associated with the up-regulation of lactoferrin mRNA and protein biosynthesis. The up-regulation at the RNA level is demonstrated by an increase in steady-state RNA levels using both *in situ* hybridization and Northern analysis techniques. In these cancers, we also observe an increase in the number of lactoferrin-positive cells, which express both the protein and mRNA. In this study, 8 of the 12 adenocarcinomas evaluated overexpress lactoferrin, compared with only 1 of 10 hyperplastic specimens. The form of lactoferrin protein extracted from endometrial adenocarcinomas appears to have

LACTOFERRIN DYSREGULATION IN ENDOMETRIAL CANCER

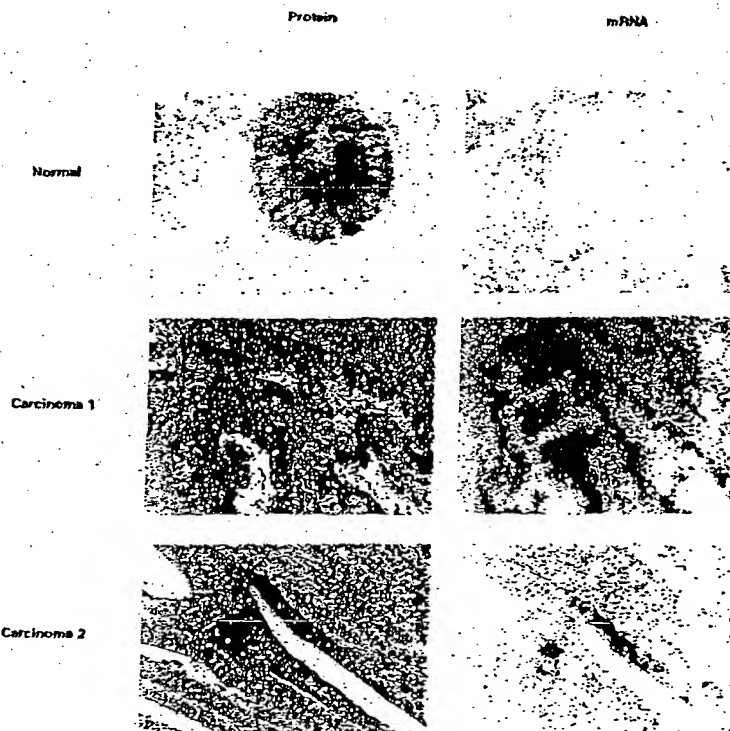


Fig. 4. Colocalization of lactoferrin protein (left panels) and mRNA (right panels) in a normal proliferative endometrium and endometrial adenocarcinomas by performing immunohistochemistry and *in situ* RNA hybridization on serial sections. Dual analysis of protein and mRNA expression reveals that glands in normal endometrium do not have detectable mRNA, as measured by *in situ* hybridization (top panels; X 40), whereas analysis of the adenocarcinomas clearly demonstrates a direct correlation between protein and RNA expression for lactoferrin (middle and bottom panels; X 10). Note that lactoferrin protein and mRNA is distributed in a heterogeneous pattern in the epithelial cells of the adenocarcinomas. As is shown in Fig. 1, a heterogeneous staining pattern for lactoferrin protein is also seen frequently in normal endometrium.

a slightly higher molecular weight than the protein present in normal tissue. This could be due to alterations in the processing of the lactoferrin mRNA, protein, or glycosylation by the malignant cells. Alternatively, there could be minor differences between lactoferrin protein which is present in neutrophils and the form synthesized by uterine epithelial cells. We suggest two hypotheses to explain lactoferrin overexpression in endometrial adenocarcinomas. In the first hypothesis, lactoferrin biosynthesis is deregulated by the same processes that lead to the malignant transformation of endometrium. If this hypothesis is true then lactoferrin may be a useful marker for endometrial adenocarcinoma investigation, and further research is needed to determine whether lactoferrin plays a contributing role in the malignant transformation. A second hypothesis is that lactoferrin-positive human endometrial adenocarcinomas evolve from the clonal expansion of cells residing in the regenerative zone (zona basalis) of normal endometrium. It is interesting to speculate that lactoferrin expression in endometrial cancer may be linked to estrogen action in

some way, since proliferative disorders of human endometrium are linked to chronic estrogen exposure over several years and sequencing data suggests that the promoter for the human lactoferrin gene does contain an ERE.

Although the function of lactoferrin is unknown, a variety of biological roles have been proposed for lactoferrin which could link this protein to a role in cancer, including the regulation of DNA synthesis (25-29), modulation of the immune response (25, 30, 31), and iron transport (32). Some forms of lactoferrin are reported to have RNase activity (33, 34). Secreted RNases are involved in development, reproductive function, neoplasia, angiogenesis, and immune suppression. (35, 36) If angiogenesis and immunosuppression are components of lactoferrin RNase activity, these properties could promote tumor growth.

In the endometrial adenocarcinomas, we observed a heterogeneous expression pattern for lactoferrin, PCNA, Her-2/*neu*, ER, and PR. With regard to prognosis, patient survival is reportedly worse when

Table 1 Correlation of lactoferrin and PR expression in human endometrial adenocarcinomas

An inverse correlation indicates that lactoferrin and progesterone were not expressed in the same regions of the tumor by immunohistochemical analysis. PR was detected with the antisera KD68, and lactoferrin was detected by a specific polyclonal antisera.

| Type of cancer | Age | Stage | Therapy | FIGO ^a grade | Lymph nodes | Myometrial invasion % | Lactoferrin expressed | Inverse correlation with PR |
|--|-----|-------|----------|-------------------------|-------------|-----------------------|-----------------------|-----------------------------|
| Endometrioid | 37 | 1 | None | 1 | 0/12 | 0 | No | No |
| Endometrioid | 61 | 1 | Estrogen | 1-3 | ND | 60 | Yes | Yes |
| Endometrioid | 67 | 1 | Estrogen | 2-5 | ND | 60 | Yes | Yes |
| Endometrioid, squamous differentiation | 35 | 1a | None | 2 | ND | 0 | Yes | Yes |
| Endometrioid | 69 | 1b | None | 1 | ND | 29 | No | No |
| Endometrioid | 57 | 1b | None | 1 | ND | 36 | Yes | Yes |
| Endometrioid | 72 | 1b | None | 2-2 | 0/15 | 5 | Yes | Yes |
| Endometrioid | 67 | 1c | Estrogen | 1 | ND | 50 | Yes | Yes |
| Endometrioid, squamous differentiation | 62 | 2a | None | 2 | ND | 80 | No | No |
| Endometrioid | 76 | 2a | None | 2 | 0/17 | 5 | Yes | Yes |
| Endometrioid, squamous differentiation | 61 | 3c | None | 3 | 1/17 | 50 | No | No |
| Endometrioid | 84 | 4b | None | 2-3 | ND | 75 | Yes | Yes |

^a FIGO, Federation Internationale des Gynecologues et Obstetristes; ND, not done.

1 2 3 4

Fig. 5. Northern analysis of lactoferrin mRNA expression confirms that endometrial adenocarcinomas (Lanes 3 and 4) significantly overexpress the 2.5-kilobase transcript of human lactoferrin in comparison to normal endometrium (Lanes 1 and 2). The Northern hybridization data supports the *in situ* RNA results and confirms that lactoferrin RNA expression is dysregulated in uterine adenocarcinomas. Normal uterine tissues appear to contain low steady-state RNA levels of lactoferrin, reflecting a controlled pattern of protein expression. Equivalent RNA loading and quality for each specimen was demonstrated by ethidium bromide staining of the RNA gels (data not shown) and by probing for a housekeeping gene (glyceraldehyde-3-phosphate dehydrogenase) that does not fluctuate significantly with the metabolic state of the tissue.

endometrial adenocarcinomas lose sex steroid receptors (37, 38), have a higher proliferative index (39), and demonstrate DNA aneuploidy. During the tumor progression of endometrial adenocarcinomas, it appears that the loss of steroid hormone receptors occurs earlier than either the increase in proliferation rate or the development of DNA aneuploidy (40). In our study, we note a striking inverse correlation between the expression of lactoferrin and PR in the endometrial adenocarcinomas. An inverse relationship also has been described for HER-2/*neu* and PR in endometrial adenocarcinomas that correlates with patient prognosis. Furthermore, in cancers of the human endometrium, ovary and breast Her-2/*neu* expression has been associated with advanced disease and poor survival (41-43). HER-2/*neu* is an oncogene that shares sequence homology with the epidermal growth factor receptor and is speculated to contribute to aberrant growth. Of note is that lactoferrin biosynthesis in the mouse uterus is associated with the expression of the epidermal growth factor. Like HER-2/*neu*, the epidermal growth factor receptor is also frequently overexpressed in PR-negative cells of endometrial adenocarcinomas (44). The amplification of growth factor receptor expression in PR-negative endometrial adenocarcinomas may be associated with the acquisition of growth autonomy and hormone independence, which may contribute to the poorer prognosis of PR-negative endometrial carcinomas (45). Some endometrial adenocarcinomas, including recurrent tumors, can be treated successfully with progesterone therapy (46, 47). Although the significance of the inverse relationship between lactoferrin and PR expression is not known, we speculate that the PR-negative cells do not undergo the normal growth inhibition and secretory differentiation normally associated with progesterone action.

A survey of human tissues reveals that lactoferrin is expressed by most normal mammalian exocrine glands and may be a prognostic marker in tumors (20). Lactoferrin is found in normal ductile breast epithelium and in primary breast carcinomas. In breast tumors, there is an inverse correlation between lactoferrin and ERs (20). Notably, lactoferrin expression in breast cancer may fall into the same category

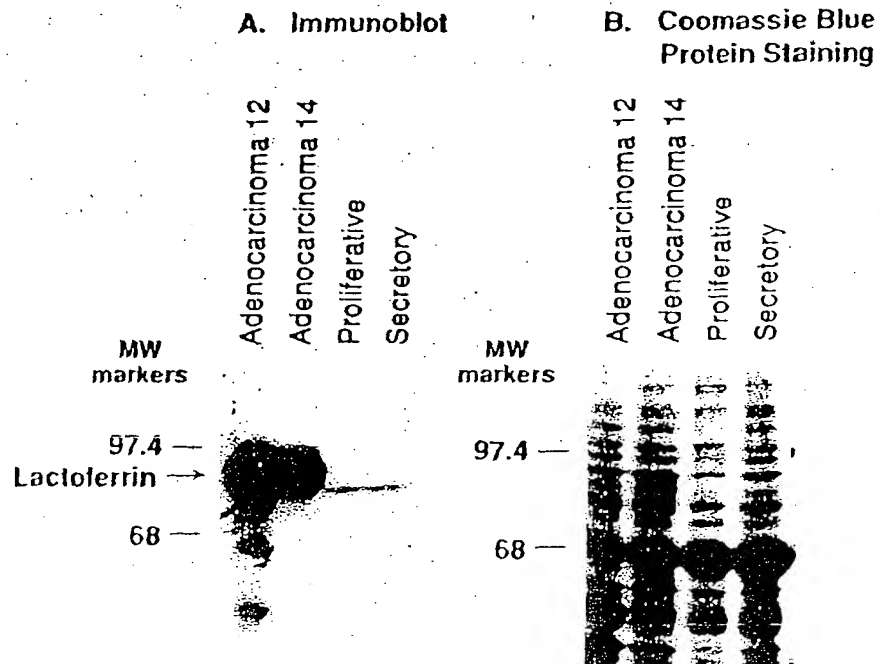


Fig. 6. Western blot analysis using an antiserum specific for human lactoferrin was performed on proteins extracted from normal and malignant endometrium and separated by SDS-polyacrylamide gel electrophoresis. A broad protein band with a molecular weight between 70,000 and 80,000 is detected in both normal and neoplastic endometrial tissue homogenates, consistent with the reported molecular weight of human lactoferrin (A). The most significant observation from the immunoblotting studies is that lactoferrin protein is markedly elevated in the adenocarcinomas, in comparison to normal endometrium, which supports the immunocytochemical analysis that demonstrates a greater number of cells positive for lactoferrin protein in the uterine tumors. B, the relative amount of protein loaded in each lane by Coomassie blue staining.

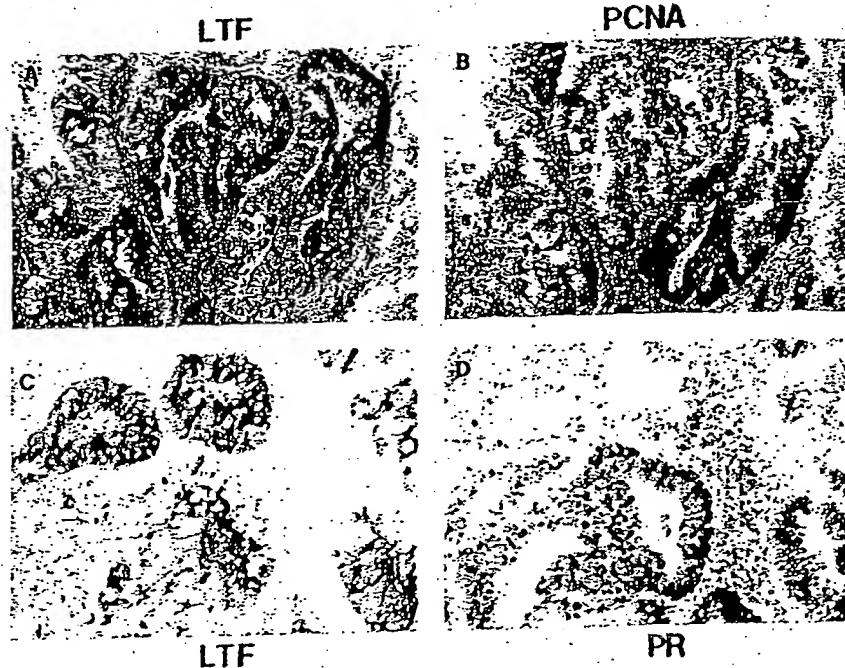


Fig. 7. Correlation of lactoferrin protein expression with the PCNA, a marker of proliferation, and PR as measured by immunohistochemistry performed on serial sections; X 20. In most cases, no relationship between lactoferrin and PCNA expression is found in either normal or malignant endometrium (data not shown). However, in 1 of 12 adenocarcinomas, an inverse correlation is seen between lactoferrin (A) and PCNA (B) localization. This striking pattern was consistent throughout the entire tumor, suggesting the possibility of cell cycle regulation of lactoferrin expression in this adenocarcinoma. The bottom panels exhibit an inverse correlation between lactoferrin (C) and PR expression (D), which was seen in all eight endometrial adenocarcinomas which expressed PR.

as the other markers for ER-negative tumors, such as amplification of EGF receptor, HER-2/*neu*, and transforming growth factor α expression, which are associated with poor prognosis. In gastric carcinomas, lactoferrin expression is associated with transformation of specific cell types including intestinal-type carcinomas, adenomas, and incomplete intestinal metaplasias (28). Although this is complete speculation at this time, perhaps lactoferrin overexpression in the various malignancies may complement the actions of the growth factor pathway molecules and contribute to the autonomous growth of these tumors.

In conclusion, our studies reveal that lactoferrin is associated with a unique population of epithelial cells in the zona basalis and that lactoferrin overexpression may be associated with malignant transformation of the human endometrium. Further studies are needed to elucidate the role of lactoferrin in normal and pathological endometrial physiology.

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REFERENCES

- Pokras, R., and Hulsnaep, V. Hysterectomies in the United States. In: *National Health Survey*, DHHS Publication PHS 88-1753, Series 13, #92, pp. 1-26, 1987.
- Pollard, J. Regulation of polypeptide growth factor synthesis and growth factor-related gene expression in the rat and mouse uterus before and after implantation. *J. Reprod. Fertil.*, 68: 721-731, 1990.
- Marini, L. Estrogens, anti-estrogens and the regulation of cell proliferation in the female reproductive tract in vivo. In: J. McLachlan (ed.), *Estrogens in the Environment*, pp. 105-121. New York: Elsevier North Holland, Inc., 1979.
- Marini, L., and Finn, C. A. Hormonal regulation of cell division in epithelial and connective tissues of the mouse uterus. *J. Endocrinol.*, 41: 565-571, 1968.
- Green, M. R., and Pastuszka, J. V. Lactoferrin is a marker for prolactin response in mouse mammary explants. *Endocrinology*, 103: 1510-1513, 1978.
- Walmer, D. K., Wrons, M. A., Hughes, C. L., and Nelson, K. G. Lactoferrin expression in the mouse reproductive tract during the natural estrous cycle: correlation with circulating estradiol and progesterone. *Endocrinology*, 131: 1458-1466, 1992.
- Teng, C. T., Fentress, B. T., Chen, Y. H., Newbold, R. R., Eddy, E. M., and McLachlan, J. A. Lactoferrin gene expression in the mouse uterus and mammary gland. *Endocrinology*, 124: 992-999, 1989.
- McMaster, M. T., Teng, C. T., Dey, S. K., and Andrews, G. K. Lactoferrin in the mouse uterus: analyses of the preimplantation period and regulation by ovarian steroids. *Mol. Endocrinol.*, 6: 101-111, 1992.
- Teng, C. T., Walker, M. P., Bhattacharya, S. N., Klapper, D. G., DiAugustine, R. P., and McLachlan, J. A. Purification and properties of an estrogen-stimulated mouse uterine glycoprotein (approx. 70 kDa). *Biochem. J.*, 240: 413-422, 1986.
- Rey, M. W., Woloshuk, S. L., deBorja, H. A., and Pierer, F. R. Complete nucleotide sequence of human mammary gland lactoferrin. *Nucleic Acids Res.*, 18: 5286, 1990.
- Teng, C. T., Liu, Y., Yang, N., Walmer, D. K., and Panella, T. Differential molecular mechanism of the estrogen action that regulates lactoferrin gene in human and mouse. *Mol. Endocrinol.*, 6: 1969-1981, 1992.
- Noyes, R. W., Herrig, A. T., and Rock, J. Dating the endometrial biopsy. *Fertil. Steril.*, 3: 25-35, 1950.
- Silverberg, S. G., and Kurman, R. J. Tumor classification. In: S. G. Silverberg and R. T. Kurman (eds.), *Tumors of the Uterine Corpus and Gestational Trophoblastic Disease*, pp. 13. Washington, DC: Armed Forces Institute of Pathology, 1992.
- Announcement, FIGO Stages: 1988 Revision. *Gynecol. Oncol.*, 35: 125-126, 1989.
- Sambrook, J., Fritsch, E. F., and Maniatis, T. *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory, 1989.
- Galand, P., and Degraef, C. Cyclin/PCNA immunostaining as an alternative to unlabeled thymidine pulse labelling for marking S phase cells in paraffin sections from animal and human tissues. *Cell Tissue Kinet.*, 22: 383-392, 1989.
- Kokuguchi, S., Hayase, R., and Sekiba, K. Proliferative activity in normal endometrial and endometrial carcinoma measured by immunohistochemistry using Ki-67 and anti-DNA polymerase α antibody, and by flow cytometry. *Acta Medica Okayama*, 46: 115-121, 1992.
- Suzuki, M., Ogawa, M., Tamada, T., Nagura, H., and Watanabe, K. Immunohistochemical localization of secretory component and IgA in the human endometrium in relation to menstrual cycle. *Acta Histochem. Cytochem.*, 17: 223-229, 1968.
- Husley, W. L., and Rejman, J. J. Bovine lactoferrin in involuting mammary tissue. *Cell Biology Int.*, 17: 283-289, 1993.
- Campbell, T., Skilton, R. A., Coombes, R. C., Shousha, S., Graham, M. D., and Lugmani, Y. A. Isolation of a lactoferrin cDNA clone and its expression in human breast cancer. *Br. J. Cancer*, 65: 19-26, 1992.
- Kim, S., Goodman, A. L., Williams, R. F., Hodgen, G. D., Hsu, J. G., and Chwalisz, K. Progesterone induces cyclic renewal in the "basalis" layer of primate endometrium: a possible site of action in anti-progesterone-induced endometrial regression. The American Fertility Society Annual Meeting Program, O-065, S32-S35, 1994. (Abstract)
- Lee, B. A., Maher, D. W., Hannink, M., and Donoghue, D. J. Identification of a signal for nuclear targeting in platelet-derived growth factor-related molecules. *Mol. Cell Biol.*, 7: 3527-3537, 1987.

25. Acland, P., Dixon, M., Peters, G., and Dickson, C. Subcellular fate of the int-2 oncoprotein is determined by choice of initiation codon. *Nature (Lond.)*, 343: 662-665, 1990.
26. Spence, A., Sheppard, P., Davie, J., Matsuo, Y., Nishi, N., McKeehan, W., Dodd, J., and Natusik, R. Regulation of a bifunctional mRNA results in synthesis of secreted and nuclear proteins. *Proc. Natl. Acad. Sci. USA*, 86: 7843-7847, 1989.
27. Gentile, P., and Broxmeyer, H. E. Interleukin-6 ablates the accessory cell-mediated suppressive effects of lactoferrin on human hematopoietic progenitor cell proliferation *in vitro*. *Ann. NY Acad. Sci.*, 628: 74-83, 1991.
28. Rejman, J. J., Oliver, S. P., Muenchen, R. A., and Turner, J. D. Proliferation of the MAC-T bovine mammary epithelial cell line in the presence of mammary secretion whey proteins. *Cell Biol. Int. Rep.*, 16: 993-1001, 1992.
29. Broxmeyer, H. E. Suppressors cytokines and regulation of myelopoiesis. Biology and possible clinical uses. *Ann. J. Pediatr. Hematol.-Oncol.*, 14: 22-30, 1992.
30. Tuccari, G., Barresi, G., Arena, F., and Inferrera, C. Immunocytochemical detection of lactoferrin in human gastric carcinomas and adenomas. *Arch. Pathol. Lab. Med.*, 113: 912-915, 1989.
31. Lugmani, Y. A., Campbell, T. A., Bennett, C., Coombes, R. C., and Paterson, I. M. Expression of lactoferrin in human stomach. *Int. J. Cancer*, 49: 684-687, 1991.
32. Djeha, A., and Brock, J. H. Effect of transferrin, lactoferrin, and chelated iron on human T-lymphocytes. *Br. J. Haematol.*, 80: 235-241, 1992.
33. Broxmeyer, H. E., Mäntel, C., Gentile, P., Srivastava, C., Miyazawa, K., Zucali, J. R., Rado, T. A., Levi, S., and Arosio, P. Actions of H-subunit ferritin and lactoferrin as suppressor molecules of myelopoiesis *in vitro* and *in vivo*. *Curr. Stud. Hematol. Blood Transfus.*, 178-181, 1991.
34. Sanchez, L., Calvo, M., and Brock, J. H. Biological role of lactoferrin. *Arch. Dis. Child.*, 67: 657-661, 1992.
35. Furmanski, P., and Li, Z. P. Multiple forms of lactoferrin in normal and leukemic human granulocytes. *Exp. Hematol.*, 18: 932-935, 1990.
36. Furmanski, P., Li, Z. P., Fortune, M. B., Swamy, C. V., and Das, M. R. Multiple molecular forms of human lactoferrin: identification of a class of lactoferrins that possess ribonuclease activity and lack iron-binding capacity. *J. Exp. Med.*, 170: 415-429, 1989.
37. Feit, J. W., Soydom, D. J., Lobb, R. R., Alderman, E. M., Bethune, J. L., Riordan, J. F., and Valler, B. L. Isolation and characterization of angiogenin, an angiogenic protein from human carcinoma cells. *Biochemistry*, 24: 5480-5486, 1985.
38. D'Alessio, G., Di Donato, A., Parente, A., and Piccoli, R. Seminal RNase: a unique member of the ribonuclease superfamily. *Trends Biochem. Sci.*, 16: 104-106, 1991.
39. Mayer, T. K., and Mooney, R. A. Laboratory analyses for steroid hormone receptors, and their applications to clinical medicine. *Clin. Chim. Acta*, 172: 1-33, 1988.
40. Vihko, R., Alanko, A., Isomaa, V., and Kauppila, A. The predictive value of steroid hormone receptor analysis in breast, endometrial and ovarian cancer. *Med. Oncol. Tumor Pharmacother.*, 3: 197-210, 1986.
41. Rosenberg, P., Wingren, S., Simonsen, E., Stal, O., Risberg, B., and Nordenskjöld, B. Flow cytometric measurements of DNA index and S-phase on paraffin-embedded early stage endometrial cancer: an important prognostic indicator. *Gynecol. Oncol.*, 35: 50-54, 1989.
42. Ponnonen, R., Mätilä, J., Kuoppala, T., and Koivula, T. DNA ploidy, cell proliferation and steroid hormone receptors in endometrial hyperplasia and early adenocarcinoma. *J. Cancer Res. Clin. Oncol.*, 119: 426-429, 1993.
43. Slamon, D. J., Clark, G. M., Wong, S. G., Levin, W. J., Ulrich, A., and McGuire, W. L. Human breast cancer: correlation of relapse and survival with amplification of the HER-2/*neu* oncogene. *Science (Washington DC)*, 235: 177-182, 1987.
44. Slamon, D. J., Godolphin, W., Jones, L. A., et al. Studies of the HER-2/*neu* proto-oncogene in human breast and ovarian cancer. *Science (Washington DC)*, 244: 707-712, 1989.
45. Beichuck, A., Rodriguez, G., Kinney, R. B., Soper, J. T., Dodge, R. K., Clarke-Pearson, D. L., and Basl, R. C., Jr. Overexpression of HER-2/*neu* in endometrial cancer is associated with advanced stage disease. *Am. J. Obstet. Gynecol.*, 164: 15-21, 1991.
46. Bigby, R. M., Li, A. X., Bomalaski, J., Siehman, F. B., Look, K. Y., and Seltin, G. P. Immunohistochemical study of HER-2/*neu*, epidermal growth factor receptor, and steroid receptor expression in normal and malignant endometrium. *Obstet. Gynecol.*, 79: 95-100, 1992.
47. Lindahl, B., Ferno, M., Gullberg, B., Norgren, A., and Wilén, R. 5-year survival rate in endometrial carcinoma stage I-II related to steroid receptor concentration, degree of differentiation, age and myometrial invasion. *Anticancer Res.*, 12: 409-412, 1992.
48. Koborn, E. I. Gestagens and endometrial carcinoma. *Gynecol. Oncol.*, 4: 398-411, 1976.
49. Creasman, W. T. Clinical correlates of estrogen and progesterone binding protein in human endometrial carcinoma. *Obstet. Gynecol.*, 55: 363-376, 1980.

Cell proliferation in human soft tissue tumors correlates with platelet-derived growth factor B chain expression: an immunohistochemical and in situ hybridization study.

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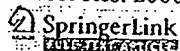
Department of Pathology, University of Washington, Seattle 98195.

The authors tested the hypothesis that the B chain of the platelet-derived growth factor (PDGF), a known connective tissue mitogen and growth factor, could be expressed by human soft tissue tumors, and that its expression could play a role in the control of cell proliferation in these tumors. Using a set of 56 soft tissue tumors, including benign tumors and all three grades of sarcomas, PDGF-B chain protein was localized using immunohistochemistry and PDGF-B mRNA was localized using in situ hybridization. The hypothesis that PDGF-B expression was related to cell proliferation was tested by simultaneously demonstrating the expression of the proliferating cell nuclear antigen in sequential tissue sections of the same tumors. Sixty and 82% of tumors had demonstrable PDGF-B mRNA and protein, respectively, with a strong correlation between their degrees of expression ($P = 0.0001$). Among the sarcomas, a strong correlation between PDGF-B expression and increasing malignant tumor grade ($P = 0.006$), and between PDGF-B expression and increasing proliferating cell nuclear antigen index ($P = 0.01$) was found. All tumors were also demonstrated to express the beta receptor of PDGF via immunohistochemistry. These studies suggest that PDGF-B expression may be an important mediator of cell proliferation control, via an autocrine mechanism, in human soft tissue tumors and may correlate with clinical outcome in the sarcomas.

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Expression of cadherins and catenins in paired tumor and non-neoplastic primary prostate cultures and corresponding prostatectomy specimens.

Wang J, Krill D, Torbenson M, Wang Q, Bisceglia M, Stoner J, Thomas A, DeFlavia P, Dhir R, Becich MJ.

Department of Pathology, University of Pittsburgh Medical Center, PA, USA.

Cadherins are a family of transmembrane proteins that play a crucial role in cell differentiation, cell migration, and intercellular adhesion. Cadherins are associated with catenins through their highly conserved cytoplasmic domain. Down-regulation of E-cadherin protein has been shown in various human cancers. This study examined the expression of cadherins and associated catenins at the mRNA level. Paired tumor and nonneoplastic primary prostate cultures were obtained from surgical specimens. Quantitative multiplex fluorescence reverse transcriptase-polymerase chain reaction (QMF RT-PCR) and quantitative analysis were performed and correlated with immunostain results. Six of seven cases of neoplastic cultures showed moderately-to-markedly decreased levels of E-cadherin and P-cadherin mRNA. Similar losses of alpha-catenin and beta-catenin mRNA were also observed. The results of QMF RT-PCR showed good correlation with the results of immunohistochemical studies based on corresponding formalin-fixed sections. In conclusion, this paper presents a coordinated down-regulation in the expression of E-cadherin and associated catenins at the mRNA and protein level in most of the cases studied. This down-regulation may play an important role in the pathogenesis of prostate cancer.

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ORIGINAL PAPER

Jianzhou Wang · Diane Krill · Michael Torbenson
 Qi Wang · Michelle Bisceglia · Judy Stoner
 Angela Thomas · Petrina DeFlavia · Rajiv Dhir
 Michael J. Becich

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Abstract Cadherins are a family of transmembrane proteins that play a crucial role in cell differentiation, cell migration, and intercellular adhesion. Cadherins are associated with catenins through their highly conserved cytoplasmic domain. Down-regulation of E-cadherin protein has been shown in various human cancers. This study examined the expression of cadherins and associated catenins at the mRNA level. Paired tumor and non-neoplastic primary prostate cultures were obtained from surgical specimens. Quantitative multiplex fluorescence reverse transcriptase-polymerase chain reaction (QMF RT-PCR) and quantitative analysis were performed and correlated with immunostain results. Six of seven cases of neoplastic cultures showed moderately-to-markedly decreased levels of E-cadherin and P-cadherin mRNA. Similar losses of α -catenin and β -catenin mRNA were also observed. The results of QMF RT-PCR showed good correlation with the results of immunohistochemical studies based on corresponding formalin-fixed sections. In conclusion, this paper presents a coordinated down-regulation in the expression of E-cadherin and associated catenins at the mRNA and protein level in most of the cases studied. This down-regulation may play an important role in the pathogenesis of prostate cancer.

Key words Prostate adenocarcinoma · Cadherin · Catenin · Adhesion molecules

J. Wang · D. Krill · M. Torbenson
 Q. Wang · M. Bisceglia · J. Stoner · A. Thomas
 P. DeFlavia · R. Dhir · M. J. Becich
 Department of Pathology,
 University of Pittsburgh Medical Center, Pittsburgh, PA
 M. J. Becich (✉)
 UPMC Shadyside,
 Department of Pathology,
 5230 Centre Avenue, Pittsburgh, PA 15232, USA
 e-mail: becich@pitt.edu
 Tel.: +1-412-6233941; Fax: +1-412-6232814

Introduction

Prostate cancer is the most common malignant tumor and the second leading cause of cancer death in men. The clinical course of patients with prostate cancer varies widely, and different factors contribute to this marked clinical variability, including genetic background, hormonal environment, and the invasive potential of the tumor. Invasion and metastasis are the hallmarks of malignancy and have been closely linked to alterations in cell-to-cell adhesion, cell migration, and interactions with extracellular matrix components [22].

Cadherins are a family of transmembrane glycoproteins responsible for maintaining the integrity of tissue and are involved in cell differentiation, cell migration, and intercellular adhesion through a calcium-dependent mechanism characterized by homotypic adhesion [35–37]. Their highly conserved cytoplasmic domains associate with catenins, a group of intracellular proteins that mediate contact between the cadherins and the microfilaments of the cytoskeleton. Each cadherin subclass shows a unique tissue distribution: E-cadherin is predominantly expressed in epithelial cells and P-cadherin is restricted to decidua tissue and the basal or lower layers of stratified epithelium [30].

The accumulating evidence suggests a decrease or loss of function in E-cadherin and P-cadherin in several human carcinomas [3, 9, 23, 32]. Loss of heterozygosity (LOH) at chromosome 16 in the location of the E-cadherin gene is present in a high percentage of prostate cancers [6, 21, 26, 28]. Decreased expression of E-cadherin is seen in various human malignant tumor cell lines, and the level of decrease correlates with the invasive potential of the tumor cell lines [1, 9, 12, 24, 40, 42]. In addition, many, but not all, immunohistochemical studies using formalin-fixed, paraffin-embedded tissue have shown that the E-cadherin protein is decreased in prostate cancer and the decrease is correlated with tumor grade [4, 8, 13, 27, 38]. Other studies have shown decreased-to-absent P-cadherin levels, but variable E-cadherin levels [33].

α - and β -catenins bind the cytoplasmic domain of E-cadherin and link it to the cytoskeleton [16, 31]. Down-regulation of expression and deletion of α -catenin genes were identified in several human cancer cell lines [19, 24, 42]. Immunohistochemical studies showed decreased α -catenin staining, which correlated well with the loss of E-cadherin staining and patient survival [25, 29, 39]. In addition, decreased β -catenin protein expression appears to be associated with malignant transformation of epithelial tissue [34]. These results suggest that cadherins and catenins may function as tumor invasion-suppressor genes. However, most of these results were obtained in studies using transformed tumor cell lines and formalin-fixed, paraffin-embedded tissue. To our knowledge, no studies to date have examined the co-expression of cadherins and catenins at the mRNA level using material derived from surgical specimens.

Recently, we have developed the methodology to cultivate primary epithelial cells under defined conditions from surgical prostatectomy specimens [20]. Areas of both carcinoma and non-neoplastic tissue are identified grossly, verified histologically, and then cultured separately, resulting in paired primary cultures of both non-neoplastic and neoplastic epithelium from the same patient. The non-neoplastic tissue cultures serve as an important control of any person-to-person variability in the expression of the genes of interest.

In this study, we investigated the co-expression of cadherin and catenin mRNA from multiple paired primary prostatectomy cultures derived from surgical prostatectomy specimens using quantitative multiplex fluorescence reverse transcriptase-polymerase chain reaction (QMF RT-PCR) [41, 44]. Immunohistochemistry studies of cadherins and catenins were performed on the corresponding paraffin-embedded prostate tissue.

Materials and methods

Tissue specimen and primary prostate cultures

We studied patients with adenocarcinoma of the prostate who underwent prostatectomy at the University of Pittsburgh Medical Center during the time interval January 1996–January 1998. No patient had received treatment prior to surgery. Fresh prostatectomy specimens were sectioned and grossly examined, representative samples of neoplastic and non-neoplastic tissue were collected, and the diagnoses were confirmed by examination of hematoxylin and eosin (H&E)-stained sections. Epithelial cells from tumor and

non-neoplastic areas of the specimen were isolated and then cultured with a chemically defined medium (CDM) without addition of growth factors as previously described. Tissue fragments were cut into small pieces and underwent a series of collagenous digestions. Following each digestion, the cells were pelleted by centrifugation at 1,000 rpm for 4 MIN [20]. To selectively promote epithelial cell growth, the pellets were resuspended and maintained in serum-free CDM supplemented with epithelial growth factor (EGF) [20]. The culture's morphology was examined daily, and epithelial cells were allowed to grow until confluence was reached, between days 7 and 10.

Isolation of total RNA and synthesis of cDNA

Total RNA was extracted from the cultured primary prostate epithelial cells at first passage according to the Trizol solution (Gibco BRL, Rockville, MD) modified method of Chomczynski and Sacchi [7]. The RNA was then quantitated spectrophotometrically. Two micrograms of total RNA were used for first strand cDNA synthesis using oligo-dT primers and MMLV reverse transcriptase (Gibco BRL, Rockville, MD).

PCR primers and quantitative multiplex fluorescence PCR

PCR primers for hepatocyte growth factor (HGF) and c-myc were synthesized as previously described [17]. PCR primers for E-cadherin, P-cadherin, α -catenin, β -catenin, and β -actin cDNA were designed according to cDNA sequences provided by the GeneBank (WWW2.ncbi.nlm.gov/genebank/query). The primers were selected to amplify 150–250 bp target genes and the PCR products from each target gene were designed to have a different size (Table 1). The reverse primers were synthesized with fluorescent molecules covalently attached to the 5' end (BRL, Rockville, MD). Twenty-five microliters of PCR reactions for QMF-PCR contained primers (20 μ M each), cDNA corresponding to 50 ng of total RNA, dNTPs, and reaction buffer. The reactions were amplified for 21 cycles at 94 °C for 1 min, 57 °C for 2 min, and 72 °C for 2 min. Five microliters of QMF-PCR reactions were mixed with an equal volume of sequencing gel loading buffer, denatured, and aliquots were electrophoresed on an ABI 373A automated sequencer (Applied Biosystems, Foster City, CA) using a matrix specific for fluorescein (Fig. 1A).

Automated sequencer gels were run for 6 h at 30 W using Genescan software (ABI, Foster City, CA). Lane assignments and areas of the peaks corresponding to fluorescent peaks were assigned and quantitated by the Genescan software using Photomultiplier tube (PMT) voltages (Fig. 1B). All experiments were done in triplicate and the results presented as means and standard deviations (SD).

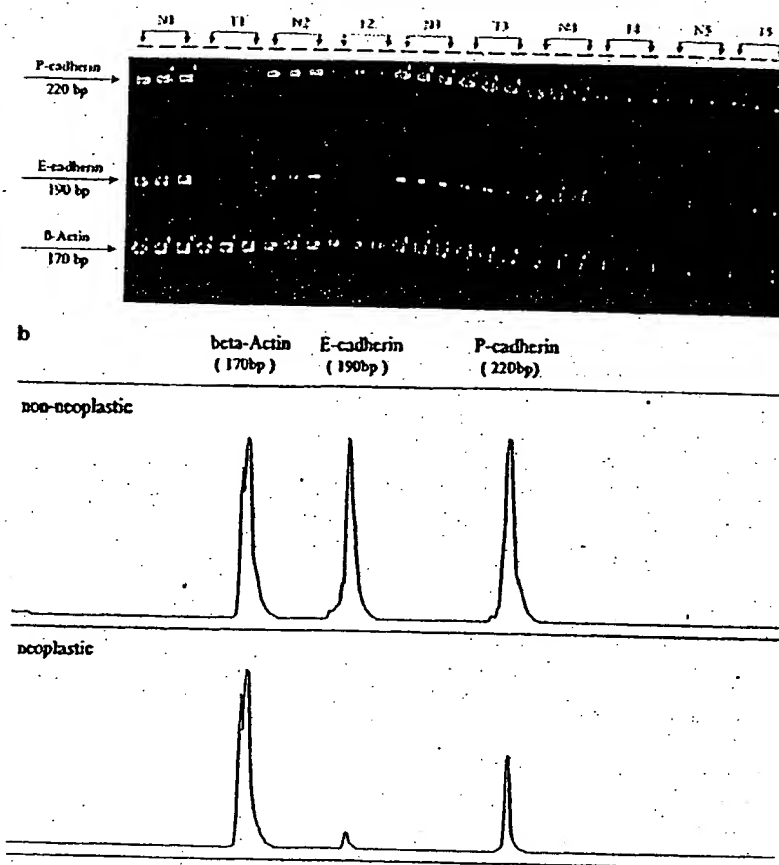
Antibodies and immunohistochemistry

Five-micron sections were obtained from formalin-fixed, paraffin-embedded tissue blocks. They were deparaffinized and hydrated with graduated ethanols. Slides were then microwaved in 1.1 M

Table 1 Summary of PCR primer sequences and PCR products

| Primer | Sequence | Target gene | Size of PCR product (bp) |
|--------|---------------------------------|-------------------|--------------------------|
| 1 | Forward ccacactgtgccatctacg | β -Actin | 170 |
| 2 | Reverse gcttctcttaatgicacg | | |
| 3 | Forward caaagtgggcacagatgggtg | E-cadherin | 190 |
| 4 | Reverse ctgcttggtatccagaaacgg | | |
| 5 | Forward gcaagagccagctctgtttagc | P-cadherin | 220 |
| 6 | Reverse acttgagctgattcagctcttgg | | |
| 7 | Forward gatggacaactatgagccagg | α -Catenin | 182 |
| 8 | Reverse tataccagggggaagcctcg | | |
| 9 | Forward ttctgggtccactaccacagc | β -Catenin | 218 |
| 10 | Reverse tgcattgctctatctaatgic | | |

Fig. 1 A Genescan image of QMF RT-PCR of five paired non-neoplastic and neoplastic primary prostate cultures. cDNA corresponding to 50 ng of total RNA was subjected to 21 cycles of PCR, and all reactions were done in triplicate. RNA of both E-cadherin and P-cadherin showed moderate-to-marked reduction in primary neoplastic cultures in four of five cases (Cases 1, 2, 4, and 5). **B** Representative automated sequencer traces (ABI 373A) from 21 cycles of QMF RT-PCR using cDNA from a paired non-neoplastic (*top panel*) and neoplastic (*bottom panel*) primary prostate cultures (Case 2), and copy number of PCR products (α -actin, E-cadherin, and P-cadherin) was calculated using peak areas. Neoplastic culture showed markedly decreased expression of both E-cadherin and P-cadherin



citrate buffer (pH 6.0) for antigen retrieval. The avidin-biotin complex method for immunohistochemistry was utilized from Vector Laboratories (Burlingame, Calif., USA). The E-cadherin, P-cadherin, α -catenin, and β -catenin antibodies, all mouse monoclonal antibodies, were purchased from Transduction Laboratories (Lexington, Ky., USA).

Evaluation of immunostaining

The percentage of positive cells, intensity of the staining, and cellular localization of the staining were examined by two independent observers using normal prostate epithelium from the same specimen as an internal control. The intensity of the signal was graded as strong, moderate, weak, and negative. The staining pattern of the tumor was compared with that of normal epithelium from the same specimens.

Results

Expression of E-cadherin and P-cadherin

Since many prostate cancers are known to be histologically heterogeneous, adjacent H&E-stained sections of the tissue fragments sent to culture were reviewed

to assess tissue homogeneity and to rule out the presence of other diseases. The paired primary cultures for this study were selected based on the following histologic features: (1) non-neoplastic tissue showed no cancerous foci or high-grade prostatic intraepithelial neoplasia (PIN); (2) neoplastic sections contained less than 5% non-cancerous epithelium. Seven out of 38 pairs of cultures met the selection criteria and were included in this study. Histologically, all seven cases were moderately differentiated adenocarcinomas, with Gleason scores ranging from 5 to 7 (median = 6). In six of these cases, adjacent sections of tumor contained no benign prostate epithelium, and in one case (specimen 4), there was less than 5% non-cancerous epithelium. Prostate stromal cells express HGF, whereas the epithelial cells express c-met, the receptor for HGF [17, 20]. The cases included in this study showed no detectable HGF expression after 21 cycles of QMF RT-PCR (data not shown). This indicates there was no significant stromal cell contamination in the current epithelial cultures.

QMF-PCR is an accurate method of measuring the relative levels of mRNA in small tissue samples [41]. In

this study, we used this method to quantitate the mRNA levels of E-cadherin and P-cadherin, relative to β -actin. All the RNA samples contained no DNA contamination, as shown by the absence of automated signals when RT was omitted from the RT-PCR reactions. We observed the expected linear increases in β -actin, E-cadherin, and P-cadherin signal intensities between cycle numbers 18 and 24, with a cDNA input corresponding to 50 ng of total RNA. The ratios of β -actin to E-cadherin and P-cadherin were constant, as was the E-cadherin/P-cadherin ratio. These results indicated that the target genes were consistently amplified in the reactions. Twenty-one cycles of PCR were chosen for all subsequent experiments. In addition, the raw sequencer quantitation of peak areas for β -actin using the same cDNA input was similar among the paired primary prostate cultures (data not shown).

When compared with the paired non-neoplastic primary cultures and normalized with the β -actin internal controls, four of the neoplastic cultures showed marked (>85%) reduction of E-cadherin mRNA levels, with one case showing no detectable E-cadherin mRNA. The other three neoplastic cultures showed mild-to-moderate reductions (Table 2).

Six of seven neoplastic cultures showed moderately to markedly decreased P-cadherin mRNA levels when compared with non-neoplastic cultures. Interestingly,

the four cases showing marked reduction in levels of E-cadherin mRNA, and also demonstrated significant losses of P-cadherin mRNA. The case with no detectable E-cadherin mRNA also demonstrated near total loss of P-cadherin mRNA (Case 1). Another case (Case 3), with only mild reduction in E-cadherin mRNA, showed no significant change in P-cadherin mRNA. Genetic variations among the patients were evident in the marked differences in the baseline levels of E-cadherin and P-cadherin expression seen in the non-neoplastic primary prostate cultures.

Expression of α -catenin and β -catenin

The highly conserved intracytoplasmic domain of the cadherins interacts with α - and β -catenins, with the catenins serving as a link between the cadherins and the microfilaments of the cytoskeleton. Six cases of neoplastic cultures showed mild-to-moderate reductions in α -catenin mRNA levels, ranging from 26 to 62%. These cases also demonstrated more severe reductions of β -catenin mRNA levels and generally correlated with the changes of cadherins in the same specimen (Table 3). Interestingly, the case with no significant change of P-cadherin mRNA and only a mild loss of E-cadherin mRNA also showed no change in mRNA levels of both

Table 2 E-cadherin and P-cadherin expression in paired non-neoplastic and neoplastic primary prostate cultures derived from prostatectomy specimens

| Case | E-cadherin ^a | | Percentage loss in tumor ^b | P-cadherin ^a | | Percentage loss in tumor ^b |
|------|-------------------------|-----------------|---------------------------------------|-------------------------|----------------|---------------------------------------|
| | Non-neoplastic | Tumor | | Non-neoplastic | Tumor | |
| 1 | 71.0 \pm 2.2 | ND ^c | 100 | 65.7 \pm 2.0 | 0.8 \pm 0.1 | 99 |
| 2 | 27.4 \pm 6.4 | 3.7 \pm 0.3 | 86 | 53.4 \pm 6.5 | 17.8 \pm 1.3 | 67 |
| 3 | 18.0 \pm 0.2 | 12.6 \pm 1.9 | 30 | 88.8 \pm 6.5 | 86.4 \pm 9.5 | 3 |
| 4 | 93.5 \pm 12.0 | 5.4 \pm 0.9 | 94 | 87.8 \pm 7.0 | 25.5 \pm 2.5 | 71 |
| 5 | 41.5 \pm 3.0 | 24.4 \pm 4.0 | 41 | 30.5 \pm 1.4 | 18.1 \pm 2.4 | 41 |
| 6 | 56.7 \pm 1.1 | 8.1 \pm 1.0 | 86 | 71.8 \pm 2.9 | 17.1 \pm 2.2 | 76 |
| 7 | 53.8 \pm 6.1 | 22.2 \pm 0.7 | 59 | 62.7 \pm 3.5 | 19.4 \pm 2.2 | 69 |

^a Level of E-cadherin and P-cadherin are normalized with β -actin from the same sample: (E-cadherin or P-cadherin)/actin \times 100

^b Percentage loss in tumor primary culture: [(N - T)/N] \times 100%

^c ND, not detectable

Table 3 α -Catenin and β -catenin expression in paired non-neoplastic and neoplastic primary prostate cultures derived from prostatectomy specimens

| Case | α -catenin ^a | | Percentage loss in tumor ^b | β -catenin ^a | | Percentage loss in tumor ^b |
|------|--------------------------------|----------------|---------------------------------------|-------------------------------|-----------------|---------------------------------------|
| | Non-neoplastic | Tumor | | Non-neoplastic | Tumor | |
| 1 | 45.1 \pm 7.1 | 17.3 \pm 0.5 | 62 | 19.7 \pm 3.6 | ND ^c | 100 |
| 2 | 49.2 \pm 1.6 | 36.2 \pm 4.0 | 26 | 21.7 \pm 1.5 | 3.9 \pm 0.2 | 82 |
| 3 | 48.0 \pm 5.9 | 47.8 \pm 9.0 | <1 | 40.1 \pm 5.3 | 38.9 \pm 6.2 | 3 |
| 4 | 61.1 \pm 2.5 | 40.8 \pm 2.6 | 33 | 36.8 \pm 1.8 | 22.1 \pm 1.9 | 40 |
| 5 | 58.2 \pm 1.8 | 27.8 \pm 1.6 | 52 | 29.8 \pm 2.0 | 15.2 \pm 1.1 | 49 |
| 6 | 46.7 \pm 4.7 | 33.6 \pm 4.7 | 28 | 31.5 \pm 4.2 | 13.5 \pm 1.6 | 57 |
| 7 | 43.2 \pm 2.6 | 26.4 \pm 1.7 | 39 | 22.3 \pm 1.5 | 11.6 \pm 1.4 | 48 |

^a Level of α -catenin and β -catenin are normalized with β -actin from the same sample: (catenin/actin) \times 100

^b Percentage loss in tumor primary culture: [(N - T)/N] \times 100%

^c ND, not detectable

α - and β -catenins. Moderate interspecimen variation was observed in the baseline expression of α -catenin and β -catenin mRNA levels in the non-neoplastic cultures.

Immunohistochemical studies of cadherins and catenins

In benign prostate tissue, E-cadherin was, in all cases, uniformly localized to the membranes of luminal glandular epithelial cells, predominantly at cell-cell junctions (Fig. 3A). One case of prostate cancer showed complete negative staining for E-cadherin (Fig. 3B), and the remaining six cases demonstrated reduced immunostaining for E-cadherin, with 25–75% of cancer cells positive (Fig. 3C). The cancerous glands generally showed reduced signal intensity and an altered heterogeneous staining pattern, which included focal cytoplasmic staining and reduced membranous staining (Fig. 3C, Table 4).

Benign prostate tissue showed uniform basal cell immunoreactivity for P-cadherin, with principally cytoplasmic and focal membranous pattern staining. The

benign, glandular non-basal epithelial cells and stromal cells were negative for P-cadherin (Fig. 3D). P-cadherin immunoreactivity was completely absent in two cases of prostate cancer (Fig. 3E), and the remaining five cases showed variable focal positivity, which was predominantly cytoplasmic (Fig. 3F). This focal P-cadherin immunostaining positivity was confirmed by staining multiple sections and by using different monoclonal antibodies. In some cases, serial sections also appeared to show immunostaining for both E- and P-cadherins with the same neoplastic cells.

In all cases of benign prostate tissue, α - and β -catenin protein expression showed strong homogeneous staining of the luminal glandular epithelium and the basal cells. In a pattern similar to that of normal E-cadherin, the α - and β -catenins were localized predominately at luminal epithelial cell-cell borders (Fig. 2A, C). In all cases of prostate cancer, there was a mild-to-moderate reduction in staining for both catenins with 50–75% of cells positive, and the staining tended to be heterogeneous (Fig. 2B, D; Table 4).

Table 4 Immunohistochemical expression of cadherins and catenins in prostatectomy specimens corresponding to primary prostate culture

| Case | E-cadherin | P-cadherin | α -catenin | β -catenin |
|------|------------|------------|-------------------|------------------|
| 1 | – | – | ++ | ++ |
| 2 | ++ | + | ++ | ++ |
| 3 | +++ | ++ | +++ | +++ |
| 4 | + | – | ++ | +++ |
| 5 | +++ | + | +++ | +++ |
| 6 | ++ | + | +++ | +++ |
| 7 | ++ | + | ++ | +++ |

–, Negative; +, <25% positive; ++, 25–50% positive; +++, 50–75% positive; + + + +, >75% positive

Discussion

In this study, we observed a coordinated down-regulation of the expression in the genes involved in the cadherin and catenin mediated cell-cell pathway at the mRNA level. The protein levels, as demonstrated by the immunohistochemical studies on the corresponding tissue sections, were generally consistent with the mRNA data as well as with that reported in the literature [18, 33, 34, 38, 39].

E-cadherin showed the most consistent loss of expression at both the mRNA level and the protein level.

Fig. 2A–D Immunohistochemical staining of α - and β -catenin in non-neoplastic prostate and in prostate adenocarcinoma. Original magnification $\times 115$. A, C Normal membranous expression of α -catenin and β -catenin in non-neoplastic prostatic epithelium (Case 1). B Decreased immunostaining for α -catenin in prostate adenocarcinoma (Case 1). D Decreased and heterogenous immunostaining for β -catenin in prostate adenocarcinoma (Case 1)

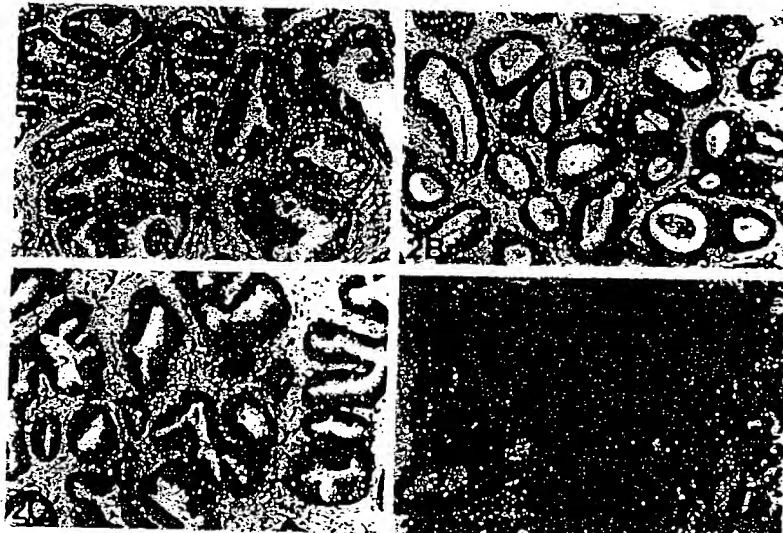
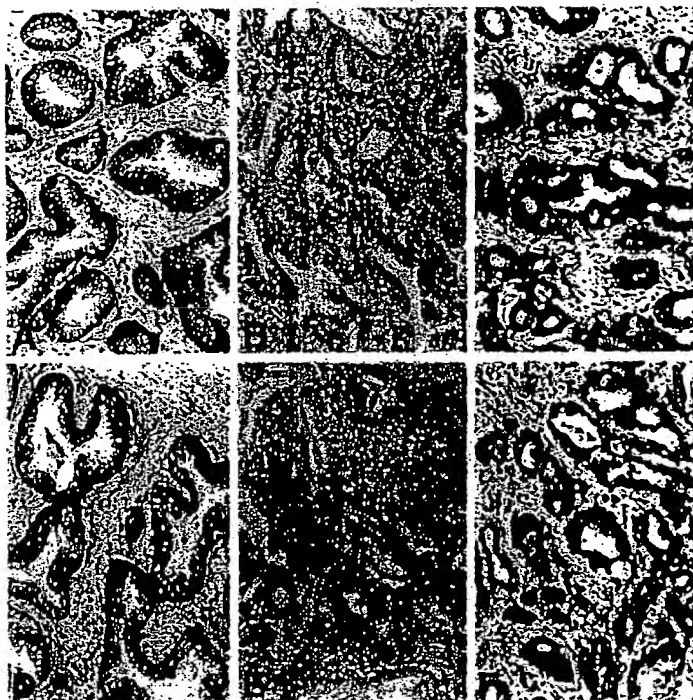


Fig. 3 A Immunohistochemical staining of E- and P-cadherin in non-neoplastic prostate and in prostate adenocarcinoma. Original magnification $\times 115$. A Normal membranous expression of E-cadherin in non-neoplastic prostatic epithelium (Case 1). B, C Complete negative (Case 1) and focal heterogeneous (Case 3) immunostaining for E-cadherin in prostate adenocarcinoma. D Normal continuous basal layer expression of P-cadherin in a portion of non-neoplastic prostatic epithelium (Case 1). E, F Complete negative (Case 1) and mild focal heterogeneous (Case 3) immunostaining for P-cadherin in prostate adenocarcinoma



LOH at chromosome 16q, where both E-cadherin and P-cadherin are located, occurs in up to 30% of prostate cancers. Four of our cases showed a greater than 50% reduction in mRNA, ranging from 87 to 100%, suggesting that mechanisms in addition to LOH may play a role in the reduction of E-cadherin mRNA levels below that of the 50% predicted by LOH alone.

Previous studies have suggested that P-cadherin could serve as a specific marker for basal cell differentiation and was not expressed in prostate cancer, although a recent study has shown focal P-cadherin expression in some prostate tumors [18]. In this study, we demonstrated P-cadherin mRNA in six of seven neoplastic cultures, although it was significantly reduced in all six cases. It is impossible to rule out the possibility that some of the P-cadherin mRNA expression may have resulted from potential contamination by small numbers of non-neoplastic basal cells admixed with the neoplastic cells within the culture material. However, we favor the interpretation that low levels of P-cadherins are expressed in cultured tumor cells, as well as weakly in some tumors *in vivo* owing to disruption of normal gene regulation. This interpretation is favored by the presence of focal immunostaining for P-cadherin protein in histologically neoplastic cells in five of seven cases. The histologic selection criteria (requiring minimal to no benign prostate glands) should also have minimized major contamination. Despite the presence of mRNA in primary tumor cultures and focal positive immuno-

staining, P-cadherin immunostaining may still serve as a useful basal cell marker because the staining pattern was distinctly abnormal in the cases where it was focally present.

In this study, the levels of α - and β -catenin mRNA were also lower than normal in six of seven cases, though the reductions were relatively less than those of cadherins, especially for the α -catenins. Catenin protein expression was generally moderate to weak by immunohistochemistry and showed a heterogeneous cytoplasmic and weak membranous staining pattern. The immunopositivity was generally similar in most cases, with 25–75% cells showing positivity. The results of the immunohistochemical studies were generally consistent with the mRNA data; although not in all cases (e.g., Case 1); perhaps due to tumor heterogeneity.

In this study, we also observed a coordinated down-regulation of E-cadherin and the catenins in most cases; this was most observable at the mRNA level. The cadherins are tightly regulated during embryogenesis and appear to serve the need for precise spatio-temporal regulation. The promoter sequences of both E-cadherin and P-cadherin have been cloned and functionally analyzed [2, 5, 10, 15]. Both promoters have similar regulatory elements, such as GC-rich regions and CCAAT boxes. Although the two promoters share similar sequences, *in vitro* binding studies suggest that the two promoters are regulated by different transcriptional factors [11]. The tissue specificity of these promoters

appears to be derived by different combinations of a relatively few factors common to many types of tissues, and does not appear to be derived from transcription factors specific for each type of tissue. In our study, the degree of loss of expression of α - and β -catenins at the mRNA level was correlated with reductions in the levels of E-cadherin expression. Although the exact mechanism of this coordinated down-regulation is not known, the coordinated pattern supports the hypothesis that loss or alteration of some regulatory factors occurs during prostate tumor progression. Possible mechanisms include transcriptional factor alterations or hypermethylation of the promoter region [14, 43]. Limited information is available on the transcriptional regulation of the catenins, but it is possible that similar mechanisms may play a role.

In summary, this is the first comparative study of the expression of the genes involved in the cadherin-mediated cell-cell adhesion pathway at the mRNA level using paired neoplastic and non-neoplastic primary cultures derived from prostatectomy specimens. Our results indicate that (1) there is a marked patient-to-patient variation in the normal levels of the cadherins and catenins; (2) mRNA levels of E-cadherin as well as catenins are significantly reduced in some prostate cancer primary cultures, and the reduction tends to be to the same degree in each tumor, suggesting a defect in a regulating mechanism common to all of these genes; (3) P-cadherin appears to be present at both the mRNA level and the protein level in some prostate cancers. This coordinated down-regulation of E-cadherin and catenin-mediated adhesion pathways may play a crucial role in tumor pathogenesis and metastasis.

References

- Behrens J, Marcel M, Fiers W, van Roy F, Birchmeier W (1989) Dissecting tumor cell invasion: epithelial cells acquire invasive properties after the loss of uvomorulin-mediated cell-cell adhesion. *J Cell Biol* 108: 2435
- Behrens J, Lowrick O, Klein-Hitpass L, Birchmeier W (1991) The E-cadherin promoter: functional analysis of the GC-rich region and an epithelial cell-specific palindromic regulatory element. *Proc Natl Acad Sci USA* 88: 11,495
- Birchmeier W, Behrens J (1994) Cadherin expression in carcinomas: role in the formation of cell junctions and prevention of invasiveness. *Biochem Biophys Acta* 1198: 11
- Bussemakers MJ, van Moorselaar RJ, Girolodi LA, Ichikawa T, Isaacs JT, Takeichi M, Debruyne FM, Schalken JA (1992) Decreased expression of E-cadherin in the progression of rat prostate cancer. *Cancer Res* 52: 2916
- Bussemakers MJ, Girolodi LA, van Bokhoven A, Schalken JA (1994) Transcriptional regulation of the human E-cadherin gene in human prostate cell lines: characterization of the human E-cadherin promoter. *Biochem Biophys Res Commun* 203: 1284
- Carter BS, Ewing CM, Ward WS, Teiger BF, Alders TW, Schalken JA, Epstein JI, Isaacs WB (1990) Allelic loss of chromosomes 16q and 10q in human prostate cancer. *Proc Natl Acad Sci USA* 87: 8751
- Chomczynski P, Sacchi N (1987) Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Ann Biochem* 162: 156
- Cohen M, Griebing T, Ahaghotu C, Rokhlin O, Ross J (1997) Cellular adhesion molecules in urologic malignancies. *Am J Clin Pathol* 107: 56
- Doki Y, Shiozaki H, Tahara H, Inoue M, Oka H, Iihara K, Kadowaki T, Takeichi M, Mori T (1993) Correlation between E-cadherin expression and invasiveness in vitro in a human esophageal cancer cell line. *Cancer Res* 53: 3421
- Faraldo ML, Cano A (1993) The 5' flanking sequences of the mouse P-cadherin gene: homologies to 5' sequences of the E-cadherin gene and identification of a first 215 base-pair intron. *J Mol Biol* 231: 935
- Faraldo ML, Rodrigo J, Behrens J, Birchmeier W, Cano A (1997) Analysis of the E-cadherin and P-cadherin promoters in murine keratinocyte cell lines from different stages of mouse skin carcinogenesis. *Mol Carcinog* 20: 33
- Frixen UH, Behrens J, Sachs M, Eberle G, Voss B, Warda A, Lochner D, Birchmeier W (1991) E-cadherin-mediated cell-cell adhesion prevents invasiveness of human carcinoma cells. *J Cell Biol* 113: 173
- Girolodi LA, Schalken JA (1993) Decreased expression of the intracellular adhesion molecule E-cadherin in prostate cancer: biologic significance and clinical implications. *Cancer Metastasis Rev* 12: 29
- Graff JR, Herman JG, Lapidus RG, Chopra H, Xu R, Jarrard DF, Isaacs WB, Pitha PM, Davidson NE, Baylin SB (1995) E-cadherin expression is silenced by DNA hypermethylation in human breast and prostate carcinomas. *Cancer Res* 55: 5195
- Hatta M, Takeichi M (1994) Complex cell type-specific transcriptional regulation by the promoter and an intron of the mouse P-cadherin gene. *Dev Growth Differ* 36: 509
- Hinck L, Nathke IS, Papkoff J, Nelson WJ (1994) Dynamics of cadherin/catenin complex formation: novel protein interactions and pathways of complex assembly. *J Cell Biol* 125: 1327
- Humphrey PA, Zhu X, Zarnegar R, Swanson PD, Ratliff TL, Vollmer RT, Day ML (1995) Hepatocyte growth factor and its receptor (c-MET) in prostatic carcinoma. *Am J Pathol* 147: 386
- Jarrard DF, Paul R, Vanbokhoven A, Nguyen SH, Bova GS, Wheelock MJ, Johnson KR, Schalken J, Bussemakers M, Isaacs WB (1997) P-cadherin is a basal cell-specific epithelial marker that is not expressed in prostate cancer. *Clin Cancer Res* 3: 2121
- Kadowaki T, Shiozaki H, Inoue M, Tamura S, Oka H, Doki Y, Iihara K, Matsui S, Iwazawa T, Nagafuchi A (1994) E-cadherin and α -catenin expression in human esophageal cancer. *Cancer Res* 54: 291
- Krill D, Shuman M, Thompson MT, Becich MJ, Strom SC (1997) A simple method for the isolation and culture of epithelial and stromal cells from benign and neoplastic prostates. *Urology* 49: 981
- Kunimi K, Bergerheim US, Larsson IL, Ekman P, Collins VP (1991) Allelotyping of prostate carcinoma. *Genomics* 11: 530
- Liotta LA (1986) Tumor invasion and metastasis: role of extracellular matrix. *Cancer Res* 46: 1
- Mayer B, Johnson JP, Leill F, Jauch KW, Heiss MM, Schildberg FW, Birchmeier W, Funke I (1993) E-cadherin expression in primary and metastatic gastric cancer: down-regulation correlates with cellular dedifferentiation and glandular disintegration. *Cancer Res* 53: 1690
- Morton R, Ewing C, Nagafuchi A, Tsukita S, Isaacs W (1993) Reduction of E-cadherin levels and deletion of the α -catenin gene in human prostate cancer cells. *Cancer Res* 53: 3585
- Murant SJ, Handley J, Stower M, Reid N, Cussenot O, Maitland NJ (1997) Coordinated changes in expression of cell-cell adhesion molecules in prostate cancer. *Eur J Cancer* 33: 263
- Natt E, Magenis RE, Zimmer J, Mansouri A, Scherer G (1989) Regional assignment of the human foci for uvomorulin and chymotrypsinogen B with the help of two overlapping deletions on the long arm of chromosome 16. *Cytogenet Cell Genet* 50: 145
- Paul R, Ewing C, Jarrard D, Isaacs W (1997) The cadherin cell-cell adhesion pathway in prostate cancer progression. *Br J Urol* 79: 37

28. Philips SM, Kinsella AR, Lee SJ, Morton DG (1994) Chromosome 16q and E-cadherin loss in prostate cancer. *Br J Cancer* (69 Suppl) 21: 19
29. Richmond PJ, Karayiannakis AJ, Nagafuchi A, Kaisary AV, Pignatelli M (1997) Aberrant E-cadherin and α -catenin expression in prostate cancer: correlation with patient survival. *Cancer Res* 57: 3189
30. Sakaki T, Wato M, Kaji R, Mushimoto K, Shirasu R, Tanaka A (1994) Correlation of E- and P-cadherin expression with differentiation grade and mode of invasion in gingival carcinoma. *Pathol Int* 44: 280
31. Shiozaki H, Oka H, Inoue M, Tamura S, Monden M (1996) E-cadherin mediated adhesion system in cancer cells. *Cancer Suppl* 77: 1605
32. Siitonen S, Kononen J, Helin H, Rantala I, Holli K, Isola J (1996) Reduced E-cadherin expression is associated with invasiveness and unfavorable prognosis in breast cancer. *Am J Clin Pathol* 105: 394
33. Soler AP, Harner GD, Knudsen KA, McBrearty FX, Grujic E, Salazar H, Han AC, Keshgegian AA (1997) Expression of P-cadherin identifies prostate-specific-negative cells in epithelial tissues of male sexual accessory organs and in prostate carcinomas. *Am J Pathol* 151: 471
34. Takayama T, Shiozaki H, Shibamoto S, Oka H, Kimura Y, Tamura S, Inoue M, Monden T, Ito F, Monden M (1996) β -catenin expression in human cancers. *Am J Pathol* 148: 39
35. Takeichi M (1988) Cell-cell adhesion molecules controlling animal morphogenesis. *Development* 102: 639
36. Takeichi M (1990) Cadherins: a molecular family important in selective cell-cell adhesion. *Annu Rev Biochem* 61: 147
37. Takeichi M (1991) Cadherin cell adhesion receptor as a morphogenetic regulator. *Science* 251: 1451
38. Umbas R, Isaacs WB, Bringuier PP, Schaafama HE, Karthaus HF, Oosterhof GO, Debruyne FM, Schalken JA (1994) Decreased E-cadherin expression is associated with poor prognosis in patients with prostate cancer. *Cancer Res* 49: 2128
39. Umbas R, Isaacs W, Bringuier P, Xue Y, Debruyne F, Schalken J (1997) Relation between aberrant α -catenin expression and loss of E-cadherin function in prostate cancer. *Int J Cancer* 74: 374
40. Vlemmckx K, Vakaet JL, Marcel M, Fiers W, van Roy F (1991) Genetic manipulation of E-cadherin expression by epithelial cell tumor cells reveals an invasion suppressor role. *Cell* 66: 107
41. Wang J, Pegoraro E, Menegazzo E, Gennarelli M, Hoop RC, Angelini C, Hoffman EP (1995) Myotonic dystrophy: evidence for a possible dominant-negative RNA mutation. *Hum Mol Genet* 4: 599
42. Watabe M, Nagafuchi A, Tsukita S, Takeichi M (1994) Induction of polarized cell-cell association and retardation of growth by activation of the E-cadherin-catenin adhesion system in a dispersed carcinoma line. *J Cell Biol* 127: 247
43. Yoshiura K, Kanai Y, Ochiai A, Shimoyama Y, Sugimura T, Hirohashi S (1995) Silencing of the E-cadherin invasion-suppressor gene by CpG methylation in human carcinomas. *Proc Natl Acad Sci USA* 92: 7416
44. Zhou J, Spier SJ, Beech J, Hoffman EP (1994) Pathophysiology of sodium channelopathies: correlation of normal/mutant mRNA ratios with clinical phenotype in dominantly inherited periodic paralysis. *Hum Mol Genet* 3: 1599

Down-regulation of prostate-specific antigen expression by finasteride through inhibition of complex formation between androgen receptor and steroid receptor-binding consensus in the promoter of the PSA gene in LNCaP cells.

Wang LG, Liu XM, Kreis W, Budman DR.

Department of Medicine, New York University, Manhasset 11030, USA.

As a specific competitive inhibitor of 5alpha-reductase, an intracellular enzyme that converts testosterone to dihydrotestosterone, finasteride is being extensively used for the treatment of benign prostatic hyperplasia and in experimental settings for prostate cancer. In this study, we showed that finasteride markedly inhibited prostate-specific antigen (PSA) secretion and expression. The promoter of the PSA gene contains several well-known cis-regulatory elements. Among them, steroid receptor-binding consensus (SRBC) has been identified as a functional androgen-responsive element. Our previous study showed that PSA was not only present in conditioned medium of the PSA-positive LNCaP cells but was also detectable in small amounts in PSA-negative cell lines, PC-3 and DU-145 (L. G. Wang et al., *Oncol. Rep.*, 3: 911-917, 1996). A strong correlation between binding of nuclear factors to SRBC and the level of PSA present in the conditioned medium and cell extracts was found in these three cell lines, whereas no such correlation with binding was obtained using Sp1 oligonucleotide as a probe. Binding of LNCaP cell nuclear proteins to SRBC was diminished when the cells were exposed to 25 micromM finasteride, at which concentration 50% of both PSA mRNA and protein were inhibited. As a major component of DNA-protein complexes, the level of androgen receptor was dramatically decreased in the cells treated with finasteride. Our data indicate that inhibition of complex formation between SRBC and nuclear proteins due to the remarkable decrease in the level of androgen receptor plays a key role in the down-regulation of PSA gene expression by finasteride in LNCaP cells.

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Expression of calcyclin in human melanocytic lesions.

Wettermann MA, van Muijen GN, Bloemers HP, Ruiter DJ.

Department of Biochemistry, University of Nijmegen, The Netherlands.

When comparing two subsequent stages of melanocytic tumor progression we identified calcyclin as a new potential progression marker, the expression of which was correlated with metastatic behavior of various human melanoma cell lines in nude mice. In this study, we describe a good correlation between RNA and protein levels in the xenografts of these cell lines and extended these experiments to a panel of 120 routinely processed human melanocytic cutaneous lesions. Northern blot analysis demonstrated that calcyclin RNA expression was elevated in melanoma metastases as compared to several types of nevocellular nevi. Calcyclin staining using a specific polyclonal antiserum showed a more complex pattern. A stronger staining in a higher percentage of positive cells was observed in thick primary melanoma (≥ 1.5 mm) as compared to thin primary melanoma (< 1.5 mm). Calcyclin expression was also present in a higher percentage of cells showing a stronger staining in melanomas with higher Clark levels ($> II$) corresponding to the vertical growth phase of primary melanomas. Protein expression in nevocellular nevi was confined to the dermal part and was highest in the lower parts of the dermis. Remarkably, dysplastic nevi (atypical moles), potential precursors of melanoma, did not show any expression at all, either in junctional or dermal parts. Confinement of the expression to the dermal part of nondysplastic nevi and primary melanomas may reflect interactions with the microenvironment of the reticular dermis that occurs with vertical growth.

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Estrogen regulation of the cytochrome P450 3A subfamily in humans.

Williams ET, Leyk M, Wrighton SA, Davies PJ, Loose DS, Shipley GL, Strobel HW.

Department of Biochemistry, Medical School, University of Texas Health Science Center at Houston, 6431 Fannin, MSB 6.200, Houston, TX 77030, USA.

This study examines the possible role of estrogen in regulating the expression of the human CYP3A subfamily: CYP3A4, CYP3A5, CYP3A7, and CYP3A43. To accomplish this goal, mRNA was quantified from human livers and endometrial samples, and total CYP3A protein levels were evaluated by Western immunoblot analysis of the liver samples. The human endometrial samples were from premenopausal and postmenopausal women. The premenopausal endometrium was either in the proliferative or secretory phase, whereas for the postmenopausal endometrium samples, the women had been treated with either a placebo or estropipate, an estrogen substitute. After analyses, CYP3A4 mRNA was shown to have lower hepatic expression in females than in males. In the endometrium, CYP3A4 and CYP3A43 are down-regulated by estrogen, whereas CYP3A5 is expressed at higher levels during the secretory phase. CYP3A7 was not detected in the endometrium. In addition, the CYP3A subfamily showed increased mRNA expression in the liver as age increased. The expression levels of total CYP3A protein and total CYP3A mRNA showed good correlation. Despite apparent regulation of CYP3A4 mRNA expression by estrogen, the effects of estrogen may be overshadowed by additional regulators of gene expression.

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Severely decreased MARCKS expression correlates with ras reversion but not with mitogenic responsiveness.

Wojtaszek PA, Stumpo DJ, Blackshear PJ, Macara IG.

Department of Pathology, University of Vermont College of Medicine, Burlington 05405.

Phorbol ester-inducible phosphorylation of MARCKS, the '80-kDa' substrate of protein kinase C, was undetectable in several phenotypically dominant, non-transformed revertants independently derived from the ras-transformed cell line NIH3T3 DT-ras. Extremely low expression of MARCKS protein accounted for this apparent lack of phosphorylation. MARCKS-encoding mRNA levels were correspondingly decreased relative to normal and ras-transformed cells in all four ras revertant cell lines studied: C-11 and F-2, derived by 5-azacytidine treatment and selection with ouabain; CHP 9CJ, derived by ethylmethane sulfonate mutagenesis and selection with cis-hydroxy-L-proline; and 12-V3, derived by transfection with the human Krev-1 gene. However, re-expression of MARCKS after transfection of a cloned MARCKS cDNA into the C-11 ras revertant cells was not sufficient to induce retransformation. In fact, no significant difference in sensitivity to mitogenic stimulation by phorbol esters was observed among several cell lines expressing widely varying levels of MARCKS. This evidence argues against a direct role for MARCKS in mitogenic signaling. However, the strong correlation between attenuation of MARCKS expression and phenotypically dominant ras reversion suggests that a common negative regulatory mechanism might be responsible for both effects, presenting a potentially useful strategy for identifying factors involved in transducing the ras signal.

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Research

Prostate stem cell antigen (PSCA) expression in human prostate cancer tissues and its potential role in prostate carcinogenesis and progression of prostate cancer

Zhao Zhigang¹ ✉ and Shen Wenlv² ✉

¹Department of Urology, Shantou University Medical College, Shantou, Guangdong, China

²Department of Urology, No 2. Affiliated Hospital of Shantou University Medical College, Shantou, Guangdong, China

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Keywords: Prostate, Neoplasm, Prostate stem cell antigen (PSCA)

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Abstract

Background

Prostate stem cell antigen (PSCA) is a recently defined homologue of the Thy-1/Ly-6 family of glycosylphosphatidylinositol (GPI)-anchored cell surface antigens. The purpose of the present study was to examine the expression status of PSCA protein and mRNA in clinical specimens of human prostate cancer (Pca) and to validate it as a potential molecular target for diagnosis and treatment of Pca.

Materials and Methods

Immunohistochemical (IHC) and *in situ* hybridization (ISH) analyses of PSCA expression were simultaneously performed on paraffin-embedded sections from 20 benign prostatic hyperplasia (BPH), 20 prostatic intraepithelial neoplasm (PIN) and 48 prostate cancer (Pca) tissues, including 9 androgen-independent prostate cancers. The level of PSCA expression was semiquantitatively scored by assessing both the percentage and intensity of PSCA-positive staining cells in the specimens. Then compared PSCA expression between BPH, PIN and Pca tissues and analysed the correlations

of PSCA expression level with pathological grade, clinical stage and progression to androgen-independence in Pca.

Results

In BPH and low grade PIN, PSCA protein and mRNA staining were weak or negative and less intense and uniform than that seen in HGPIN and Pca. There were moderate to strong PSCA protein and mRNA expression in 8 of 11 (72.7%) HGPIN and in 40 of 48 (83.4%) Pca specimens examined by IHC and ISH analyses, with statistical significance compared with BPH (20%) and low grade PIN (22.2%) samples ($p < 0.05$, respectively). The expression level of PSCA increased with high Gleason grade, advanced stage and progression to androgen-independence ($p < 0.05$, respectively). In addition, IHC and ISH staining showed a high degree of correlation between PSCA protein and mRNA overexpression.

Conclusions

Our data demonstrate that PSCA as a new cell surface marker is overexpressed by a majority of human Pca. PSCA expression correlates positively with adverse tumor characteristics, such as increasing pathological grade (poor cell differentiation), worsening clinical stage and androgen-independence, and speculatively with prostate carcinogenesis. PSCA protein overexpression results from upregulated transcription of PSCA mRNA. PSCA may have prognostic utility and may be a promising molecular target for diagnosis and treatment of Pca.

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Prostate cancer (Pca) is the second leading cause of cancer-related death in American men and is becoming a common cancer increasing in China. Despite recently great progress in the diagnosis and management of localized disease, there continues to be a need for new diagnostic markers that can accurately discriminate between indolent and aggressive variants of Pca. There also continues to be a need for the identification and characterization of potential new therapeutic targets on Pca cells. Current diagnostic and therapeutic modalities for recurrent and metastatic Pca have been limited by a lack of specific target antigens of Pca.

Although a number of prostate-specific genes have been identified (i.e. prostate specific antigen, prostatic acid phosphatase, glandular kallikrein 2), the majority of these are secreted proteins not ideally suited for many immunological strategies. So, the identification of new cell surface antigens is critical to the development of new diagnostic and therapeutic approaches to the management of Pca.

Reiter RE et al [1] reported the identification of prostate stem cell antigen (PSCA), a cell surface antigen that is predominantly prostate specific. The PSCA gene encodes a 123 amino acid glycoprotein, with 30% homology to stem cell antigen 2 (Sca 2). Like Sca-2, PSCA also belongs to a member of the Thy-1/Ly-6 family and is anchored by a glycosylphosphatidylinositol (GPI) linkage. mRNA *in situ* hybridization (ISH) localized PSCA expression in

normal prostate to the basal cell epithelium, the putative stem cell compartment of prostatic epithelium, suggesting that PSCA may be a marker of prostate stem/progenitor cells.

In order to examine the status of PSCA protein and mRNA expression in human Pca and validate it as a potential diagnostic and therapeutic target for Pca, we used immunohistochemistry (IHC) and *in situ* hybridization (ISH) simultaneously, and conducted PSCA protein and mRNA expression analyses in paraffin-embedded tissue specimens of benign prostatic hyperplasia (BPH, $n = 20$), prostate intraepithelial neoplasm (PIN, $n = 20$) and prostate cancer (Pca, $n = 48$). Furthermore, we evaluated the possible correlation of PSCA expression level with Pca tumorigenesis, grade, stage and progression to androgen-independence.

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Table 1
Correlation of PSCA expression with Gleason score

Table 2
Correlation of PSCA expression with clinical stage

Tissue samples

All of the clinical tissue specimens studied herein were obtained from 80 patients of 57–84 years old by prostatectomy, transurethral resection of prostate (TURP) or biopsies. The patients were classified as 20 cases of BPH, 20 cases of PIN, 40 cases of primary Pca, including 9 patients with recurrent Pca and a history of androgen ablation therapy (orchiectomy and/or hormonal therapy), who were referred to as androgen-independent prostate cancers. Eight specimens were harvested from these androgen-independent Pca patients prior to androgen ablation treatment. Each tissue sample was cut into two parts, one was fixed in 10% formalin for IHC and the other treated with 4% paraformaldehyde/0.1 M PBS PH 7.4 in 0.1% DEPC for 1 h for ISH analysis, and then embedded in paraffin. All paraffin blocks examined were then cut into 5 μ m sections and mounted on the glass slides specific for IHC and ISH respectively in the usual fashion. H&E-stained section of each Pca was evaluated and assigned a Gleason score by the experienced urological pathologist at our institution based on the criteria of Gleason score [2]. The Gleason sums are summarized in Table 1. Clinical staging was performed according to Jewett-whitmore-prout staging system, as shown in Table 2. In the category of PIN, we graded the specimens into two groups, i.e. low grade PIN (grade I – II) and high grade PIN (HGPIN, grade III) on the basis of literatures [3,4].

Immunohistochemical (IHC) analysis

Briefly, tissue sections were deparaffinized, dehydrated, and subjected to microwaving in 10 mmol/L citrate buffer, PH 6.0 (Boshide, Wuhan, China) in a 900 W oven for 5 min to induce epitope retrieval. Slides were allowed to cool at room temperature for 30 min. A primary mouse antibody specific to human PSCA (Boshide, Wuhan, China) with a 1:100 dilution was applied to incubate with the slides at room temperature for 2 h. Labeling was detected by sequentially adding biotinylated secondary antibodies and streptavidin-peroxidase, and localized using 3,3'-diaminobenzidine reaction. Sections were then counterstained with hematoxylin. Substitution of the primary antibody with phosphate-buffered-saline (PBS) served as a negative-staining control.

mRNA *in situ* hybridization (ISH)

Five- μ m-thick tissue sections were deparaffinized and dehydrated, then digested in pepsin solution (4 mg/ml in 3% citric acid) for 20 min at 37.5°C, and further processed for ISH. Digoxigenin-labeled sense and antisense human PSCA RNA probes (obtained from Boshide, Wuhan, China) were hybridized to the sections at 48°C overnight. The posthybridization wash with a high stringency was performed sequentially at 37°C in 2 × standard saline citrate (SSC) for 10 min, in 0.5 × SSC for 15 min and in 0.2 × SSC for 30 min. The slides were then incubated to biotinylated mouse anti-digoxigenin antibody at 37.5°C for 1 h followed by washing in 1 × PBS for 20 min at room temperature, and then to streptavidin-peroxidase at 37.5°C for 20 min followed by washing in 1 × PBS for 15 min at room temperature. Subsequently, the slides were developed with diaminobenzidine and then counterstained with hematoxylin to localize the hybridization signals. Sections hybridized with the sense control probes routinely did not show any specific hybridization signal above background. All slides were hybridized with PBS to substitute for the probes as a negative control.

Scoring methods

To determine the correlation between the results of PSCA immunostaining and mRNA *in situ* hybridization, the same scoring manners are taken in the present study for PSCA protein staining by IHC and PSCA mRNA staining by ISH. Each slide was read and scored by two independently experienced urological pathologists using Olympus BX-41 light microscopes. The evaluation was done in a blinded fashion. For each section, five areas of similar grade were analyzed semiquantitatively for the fraction of cells staining. Fifty percent of specimens were randomly chosen and rescored to determine the degree of interobserver and intraobserver concordance. There was greater than 95% intra- and interobserver agreement.

The intensity of PSCA expression evaluated microscopically was graded on a scale of 0 to 3+ with 3 being the highest expression observed (0, no staining; 1+, mildly intense; 2+, moderately intense; 3+, severely intense). The staining density was quantified as the percentage of cells staining positive for PSCA with the primary antibody or hybridization probe, as follows: 0 = no staining; 1 = positive staining in <25% of the sample; 2 = positive staining in 25%-50% of the sample; 3 = positive staining in >50% of the sample. Intensity score (0 to 3+) was multiplied by the density score (0-3) to give an overall score of 0-9 [1,5]. In this way, we were able to differentiate specimens that may have had focal areas of increased staining from those that had diffuse areas of increased staining [6]. The overall score for each specimen was then categorically assigned to one of the following groups: 0 score, negative expression; 1-2 scores, weak expression; 3-6 scores, moderate expression; 9 score, strong expression.

Statistical analysis

Intensity and density of PSCA protein and mRNA expression in BPH, PIN and Pca tissues were compared using the Chi-square and Student's *t*-test. Univariate associations between PSCA expression and Gleason score, clinical stage and progression to androgen-independence were calculated using Fisher's Exact Test. For all analyses, $p < 0.05$ was considered statistically significant.

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Figure 1
Representatives of PSCA
IHC and ISH staining in Pca
(A. IHC staining, B. ISH
staining, x200
magnification)

PSCA expression in BPH

In general, PSCA protein and mRNA were expressed weakly in individual samples of BPH. Some areas of prostate expressed weak levels (composite score 1–2), whereas other areas were completely negative (composite score 0). Four cases (20%) of BPH had moderate expression of PSCA protein and mRNA (composite score 4–6) by IHC and ISH. In 2/20 (10%) BPH specimens, PSCA mRNA expression was moderate (composite score 3–6), but PSCA protein expression was weak (composite score 2) in one and negative (composite score 0) in the other. PSCA expression was localized to the basal and secretory epithelial cells, and prostatic stroma was almost negative staining for PSCA protein and mRNA in all cases examined.

PSCA expression in PIN

In this study, we detected weak or negative expression of PSCA protein and mRNA (≤ 2 scores) in 7 of 9 (77.8%) low grade PIN and in 2 of 11 (18.2%) HGPIN, and moderate expression (3–6 scores) in the rest 2 low grade PIN and 5 of 11 (45.5%) HGPIN. One HGPIN with moderate PSCA mRNA expression (6 score) was found weak staining for PSCA protein (2 score) by IHC. Strong PSCA protein and mRNA expression (9 score) were detected in the remaining 3 of 11 (27.3%) HGPIN. There was a statistically significant difference of PSCA protein and mRNA expression levels observed between HGPIN and BPH ($p < 0.05$), but no statistical difference reached between low grade PIN and BPH ($p > 0.05$).

PSCA expression in Pca

In order to determine if PSCA protein and mRNA can be detected in prostate cancers and if PSCA expression levels are increased in malignant compared with benign glands, Forty-eight paraffin-embedded Pca specimens were analysed by IHC and ISH. It was shown that 19 of 48 (39.6%) Pca samples stained very strongly for PSCA protein and mRNA with a score of 9 and another 21 (43.8%) specimens displayed moderate staining with scores of 4–6 (Figure 1). In addition, 4 specimens with moderate to strong PSCA mRNA expression (scores of 4–9) had weak protein staining (a score of 2) by IHC analyses. Overall, Pca expressed a significantly higher level of PSCA protein and mRNA than any other specimen category in this study ($p < 0.05$, compared with BPH and PIN respectively). The result demonstrates that PSCA protein and mRNA are overexpressed by a majority of human Pca.

Correlation of PSCA expression with Gleason score in Pca

Using the semi-quantitative scoring method as described in Materials and Methods, we compared the expression level of PSCA protein and mRNA with Gleason grade of Pca, as shown in Table 1. Prostate adenocarcinomas were graded by Gleason score as 2–4 scores = well-differentiation, 5–7 scores = moderate-differentiation and 8–10 scores = poor-differentiation [7]. Seventy-two percent of Gleason scores 8–10 prostate cancers had very

strong staining of PSCA compared to 21% with Gleason scores 5-7 and 17% with 2-4 respectively, demonstrating that poorly differentiated Pca had significantly stronger expression of PSCA protein and mRNA than moderately and well differentiated tumors ($p < 0.05$). As depicted in Figure 1, IHC and ISH analyses showed that PSCA protein and mRNA expression in several cases of poorly differentiated Pca were particularly prominent, with more intense and uniform staining. The results indicate that PSCA expression increases significantly with higher tumor grade in human Pca.

Correlation of PSCA expression with clinical stage in Pca

With regards to PSCA expression in every stage of Pca, we showed the results in Table 2. Seventy-five percent of locally advanced and node positive cancers (i.e. C-D stages) expressed statistically high levels of PSCA versus 32.5% that were organ confined (i.e. A-B stages) ($p < 0.05$). The data demonstrate that PSCA expression increases significantly with advanced tumor stage in human Pca.

Correlation of PSCA expression with androgen-independent progression of Pca

All 9 specimens of androgen-independent prostate cancers stained positive for PSCA protein and mRNA. Eight specimens were obtained from patients managed prior to androgen ablation therapy. Seven of eight (87.5%) of these androgen-independent prostate cancers were in the strongest staining category (score = 9), compared with three out of eight (37.5%) of patients with androgen-dependent cancers ($p < 0.05$). The results demonstrate that PSCA expression increases significantly with progression to androgen-independence of human Pca.

It is evident from the results above that within a majority of human prostate cancers the level of PSCA protein and mRNA expression correlates significantly with increasing grade, worsening stage and progression to androgen-independence.

Correlation of PSCA immunostaining and mRNA *in situ* hybridization

In all 88 specimens surveyed herein, we compared the results of PSCA IHC staining with mRNA ISH analysis. Positive staining areas and its intensity and density scores evaluated by IHC were identical to those seen by ISH in 79 of 88 (89.8%) specimens (18/20 BPH, 19/20 PIN and 42/48 Pca respectively). Importantly, 27/27 samples with PSCA mRNA composite scores of 0-2, 32/36 samples with scores of 3-6 and 22/24 samples with a score of 9 also had PSCA protein expression scores of 0-2, 3-6 and 9 respectively. However, in 5 samples with PSCA mRNA overall scores of 3-6 and in 2 with scores of 9 there were less or negative PSCA protein expression (i.e. scores of 0-4), suggesting that this may reflect posttranscriptional modification of PSCA or that the epitopes recognized by PSCA mAb may be obscured in some cancers. The data demonstrate that the results of PSCA immunostaining were consistent with those of mRNA ISH analysis, showing a high degree of correlation between PSCA protein and mRNA expression.

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PSCA is homologous to a group of cell surface proteins that mark the earliest phase of hematopoietic development. PSCA mRNA expression is prostate-specific in normal male tissues and is highly up-regulated in both androgen-dependent and-independent Pca xenografts (LAPC-4 tumors). We hypothesize that PSCA may play a role in Pca tumorigenesis and progression, and may serve as a target for Pca diagnosis and treatment. In this study, IHC and ISH showed that in general there were weak or absent PSCA protein and mRNA expression in BPH and low grade PIN tissues. However, PSCA protein and mRNA are widely expressed in HGPIN, the putative precursor of invasive Pca, suggesting that up-regulation of PSCA is an early event in prostate carcinogenesis. Recently, Reiter RE et al [1], using ISH analysis, reported that 97 of 118 (82%) HGPIN specimens stained strongly positive for PSCA mRNA. A very similar finding was seen on mouse PSCA (mPSCA) expression in mouse HGPIN tissues by Tran C. P et al [8]. These data suggest that PSCA may be a new marker associated with transformation of prostate cells and tumorigenesis.

We observed that PSCA protein and mRNA are highly expressed in a large percentage of human prostate cancers, including advanced, poorly differentiated, androgen-independent and metastatic cases. Fluorescence-activated cell sorting and confocal/ immunofluorescent studies demonstrated cell surface expression of PSCA protein in Pca cells [9]. Our IHC expression analysis of PSCA shows not only cell surface but also apparent cytoplasmic staining of PSCA protein in Pca specimens (Figure 1). One possible explanation for this is that anti-PSCA antibody can recognize PSCA peptide precursors that reside in the cytoplasm. Also, it is possible that the positive staining that appears in the cytoplasm is actually from the overlying cell membrane [5]. These data seem to indicate that PSCA is a novel cell surface marker for human Pca.

Our results show that elevated level of PSCA expression correlates with high grade (i.e. poor differentiation), increased tumor stage and progression to androgen-independence of Pca. These findings support the original IHC analyses by Gu Z et al [9], who reported that PSCA protein expressed in 94% of primary Pca and the intensity of PSCA protein expression increased with tumor grade, stage and progression to androgen-independence. Our results also collaborate the recent work of Han KR et al [10], in which the significant association between high PSCA expression and adverse prognostic features such as high Gleason score, seminal vesicle invasion and capsular involvement in Pca was found. It is suggested that PSCA overexpression may be an adverse predictor for recurrence, clinical progression or survival of Pca. Hara H et al [11] used RT-PCR detection of PSA, PSMA and PSCA in 1 ml of peripheral blood to evaluate Pca patients with poor prognosis. The results showed that among 58 Pca patients, each PCR indicated the prognostic value in the hierarchy of PSCA>PSA>PSMA RT-PCR, and extraprostatic cases with positive PSCA PCR indicated lower disease-progression-free survival than those with negative PSCA PCR, demonstrating that PSCA can be used as a prognostic factor. Dubey P et al [12] reported that elevated numbers of PSCA + cells correlate positively with the onset and development of prostate carcinoma over a long time span in the prostates of the TRAMP and PTEN +/- models compared with its

normal prostates. Taken together with our present findings, in which PSCA is overexpressed from HGPIN to almost frank carcinoma, it is reasonable and possible to use increased PSCA expression level or increased numbers of PSCA-positive cells in the prostate samples as a prognostic marker to predict the potential onset of this cancer. These data raise the possibility that PSCA may have diagnostic utility or clinical prognostic value in human Pca.

The cause of PSCA overexpression in Pca is not known. One possible mechanism is that it may result from PSCA gene amplification. In humans, PSCA is located on chromosome 8q24.2 [1], which is often amplified in metastatic and recurrent Pca and considered to indicate a poor prognosis [13-15]. Interestingly, PSCA is in close proximity to the c-myc oncogene, which is amplified in >20% of recurrent and metastatic prostate cancers [16,17]. Reiter RE et al [18] reported that PSCA and MYC gene copy numbers were co-amplified in 25% of tumors (five out of twenty), demonstrating that PSCA overexpression is associated with PSCA and MYC coamplification in Pca. Gu Z et al [9] recently reported that in 102 specimens available to compare the results of PSCA immunostaining with their previous mRNA ISH analysis, 92 (90.2%) had identically positive areas of PSCA protein and mRNA expression. Taken together with our findings, in which we detected moderate to strong expression of PSCA protein and mRNA in 34 of 40 (85%) Pca specimens examined simultaneously by IHC and ISH analyses, it is demonstrated that PSCA protein and mRNA overexpressed in human Pca, and that the increased protein level of PSCA was resulted from the upregulated transcription of its mRNA.

At present, the regulation mechanisms of human PSCA expression and its biological function are yet to be elucidated. PSCA expression may be regulated by multiple factors [13]. Watabe T et al [19] reported that transcriptional control is a major component regulating PSCA expression levels. In addition, induction of PSCA expression may be regulated or mediated through cell-cell contact and protein kinase C (PKC) [20]. Homologues of PSCA have diverse activities, and have themselves been involved in carcinogenesis. Signalling through SCA-2 has been demonstrated to prevent apoptosis in immature thymocytes [21]. Thy-1 is involved in T cell activation and transduces signals through src-like tyrosine kinases [22]. Ly-6 genes have been implicated both in tumorigenesis and in cell-cell adhesion [23-25]. Cell-cell or cell-matrix interaction is critical for local tumor growth and spread to distal sites. From its restricted expression in basal cells of normal prostate and its homology to SCA-2, PSCA may play a role in stem/progenitor cell function, such as self-renewal (i.e. anti-apoptosis) and/or proliferation [1]. Taken together with the results in the present study, we speculate that PSCA may play a role in tumorigenesis and clinical progression of Pca through affecting cell transformation and proliferation. From our results, it is also suggested that PSCA as a new cell surface antigen may have a number of potential uses in the diagnosis, therapy and clinical prognosis of human Pca. PSCA overexpression in prostate biopsies could be used to identify patients at high risk to develop recurrent or metastatic disease, and to discriminate cancers from normal glands in prostatectomy samples. Similarly, the detection of PSCA-overexpressing cells in bone marrow or peripheral blood may identify and predict metastatic progression better than current assays, which identify only PSA-positive or PSMA-positive prostate cells.

In summary, we have shown in this study that PSCA protein and mRNA are maintained in expression from HGPIN through all stages of Pca in a majority of cases, which may be associated with prostate carcinogenesis and correlate positively with high tumor grade (poor cell differentiation), advanced stage and androgen-independent progression. PSCA protein overexpression is due to the upregulation of its mRNA transcription. The results suggest that PSCA may be a promising molecular marker for the clinical prognosis of human Pca and a valuable target for diagnosis and therapy of this tumor.

Competing interests

None declared.

Outline

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References

1. Reiter RE, Gu Z, Watabe T, Thomas G, Szigeti K, David E, Wähl M, Nisitani S, Yamashiro J, Le Beau MM, Loda M, Witte ON: **Prostate stem cell antigen: a cell surface marker overexpressed in prostate cancer.** *Proc Natl Acad Sci USA* 1998, **95**:1735-1740. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1] [2] [3] [4] [5]
2. Gleason DF: **Histologic grading and clinical staging of prostatic carcinoma.** *In: Urologic Pathology: The Prostate (Edited by: Tannebaum M).* Philadelphia, Lea & Febiger 1977, 171-197.
Return to citation in text: [1]
3. Brawer MK: **Prostatic intraepithelial neoplasia: a premalignant lesion.** *Hum Pathol* 1992, **23**:242-248. [PubMed Abstract]
Return to citation in text: [1]
4. Amin MB, Ro JY, Ayala AC: **Prostatic intraepithelial neoplasia: relationship to adenocarcinoma of prostate.** *Pathol Annu* 1994, **29**:1-30. [PubMed Abstract]
Return to citation in text: [1]
5. Amara N, Palapattu GS, Schrage M, Gu Z, Thomas GV, Dorey F, Said J, Reiter RE: **Prostate stem cell antigen is overexpressed in human transitional cell carcinoma.** *Cancer Res* 2001, **61**:4660-4665. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1] [2]
6. Hanas JS, Lerner MR, Lightfoot SA, Raczowski C, Kastens DJ, Brackett DJ, Postier RG: **Expression of the cyclin-dependent kinase inhibitor p21 (WAF1/CIP1) and p53 tumor suppressor in dysplastic progression and adenocarcinoma in Barrett esophagus.** *Cancer (Phila)* 1999, **86**:756-763. [Publisher Full Text]
Return to citation in text: [1]
7. Egevad L, Gramfors T, Karlberg L: **Prognostic value of the Gleason score in prostate cancer.**


BJU Int 2002, **89**:538-542. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1]

8. Tran CP, Lin C, Yamashiro J, Reiter RE: **Prostate stem cell antigen is a marker of late intermediate prostate epithelial cells.**
Mol Cancer Res 2002, **1**:113-121. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1]
9. Gu Z, Thomas G, Yamashiro J, Shintaku IP, Dorey F, Raitano A, Witte ON, Said JW, Loda M, Reiter RE: **Prostate stem cell antigen (PSCA) expression increases with high Gleason score, advanced stage and bone metastasis in prostate cancer.**
Oncogene 2000, **19**:1288-1296. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1] [2] [3]
10. Han KR, Seligson DB, Liu X, Horvath S, Shintaku PI, Thomas GV, Said JW, Reiter RE: **Prostate stem cell antigen expression is associated with gleason score, seminal vesicle invasion and capsular invasion in prostate cancer.**
J Urol 2004, **171**:1117-1121. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1]
11. Hara H, Kasahara T, Kawasaki T, Bilim V, Obara K, Takahashi K, Tomita Y: **Reverse Transcription-Polymerase Chain Reaction Detection of Prostate-specific Antigen, Prostate-specific Membrane Antigen, and Prostate Stem Cell Antigen in One Milliliter of Peripheral Blood.**
Clin Cancer Res 2002, **8**:1794-1799. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1]
12. Dubey P, Wu H, Reiter RE, Witte ON: **Alternative pathways to prostate carcinoma activate prostate stem cell antigen expression.**
Cancer Res 2001, **61**:3256-3261. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1]
13. Visa korpi T, Kallioniemi AH, Syvanen AC, Hyytinen ER, Karhu R, Tammela T, Isola JJ, Kallioniemi OP: **Genetic changes in primary and recurrent prostate cancer by comparative genomic hybridization.**
Cancer Res 1995, **55**:342-347. [PubMed Abstract]
Return to citation in text: [1]
14. Sato K, Qian J, Slezak JM, Lieber MM, Bostwick DG, Bergstralh EJ, Jenkins RB: **Clinical significance of alterations of chromosome 8 in high-grade, advanced, nonmetastatic prostate carcinoma.**
J Natl Cancer Inst 1999, **91**:1574-1580. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1]
15. Van Den Berg C, Guan XY, Von Hoff D, Jenkins R, Bittner J, Griffin C, Kallioniemi O, Visakorpi T, McGill J, Herath J, Epstein J, Sarosdy M, Meltzer P, Trent J: **DNA sequence amplification in human prostate cancer identified by chromosome microdissection: potential prognostic implications.**
Clin Cancer Res 1995, **1**:11-18. [PubMed Abstract]
Return to citation in text: [1]
16. Jenkins RB, Qian J, Lieber MM, Bostwick DG: **Detection of c-myc oncogene amplification and chromosomal anomalies in metastatic prostatic carcinoma by fluorescence in situ hybridization.**
Cancer Res 1997, **57**:524-531. [PubMed Abstract]
Return to citation in text: [1]

17. Nupponen NN, Kakkola L, Koivisto P, Visakorpi T: **Genetic alterations in hormone-refractory recurrent prostate carcinomas.**
Am J Pathol 1998, **153**:141-148. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1]
18. Reiter RE, Sato I, Thomas G, Qian J, Gu Z, Watabe T, Loda M, Jenkins RB: **Coamplification of prostate stem cell antigen (PSCA) and MYC in locally advanced prostate cancer.**
Genes Chromosomes Cancer 2000, **27**:95-103. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1] [2]
19. Watabe T, Lin M, Donjacour AA, Cunha GR, Witte ON, Reiter RE: **Growth, regeneration, and tumorigenesis of the prostate activates the PSCA promoter.**
Proc Natl Acad Sci USA 2002, **99**:401-406. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1]
20. Bahrenberg G, Brauers A, Joost HG, Jakse G: **PSCA expression is regulated by phorbol ester and cell adhesion in the bladder carcinoma cell line RT112.**
Cancer Lett 2001, **168**:37-43. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1]
21. Noda S, Kosugi A, Saitoh S, Narumiya S, Hamaoka T: **Protection from anti-TCR/CD3-induced apoptosis in immature thymocytes by a signal through thymic shared antigen-1/stem cell antigen-2.**
J Exp Med 1996, **183**:2355-2360. [PubMed Abstract]
Return to citation in text: [1]
22. Thomas PM, Samelson LE: **The glycoposphatidylinositol-anchored Thy-1 molecule interacts with the p60fyn protein tyrosine kinase in T cells.**
J Biol Chem 1992, **267**:12317-12322. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1]
23. Bamezai A, Rock KL: **Overexpressed Ly-6A.2 mediated cell-cell adhesion by binding a ligand expressed on lymphoid cells.**
Proc Natl Acad Sci USA 1995, **92**:4294-4298. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1]
24. Katz BZ, Eshel R, Sagi-Assif O, Witz IP: **An association between high Ly-6A/E expression on tumor cells and a highly malignant phenotype.**
Int J Cancer 1994, **59**:684-691. [PubMed Abstract]
Return to citation in text: [1]
25. Brakenhoff RH, Gerretsen M, Knippels EM, van Dijk M, van Essen H, Weghuis DO, Sinke RJ, Snow GB, van Dongen GA: **The human E48 antigen, highly homologous to the murine Ly-6 antigen ThB, is a GPI-anchored molecule apparently involved in keratinocyte cell-cell adhesion.**
J Cell Biol 1995, **129**:1677-1689. [PubMed Abstract] [Publisher Full Text]
Return to citation in text: [1]

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Expression of superoxide dismutases, catalase, and glutathione peroxidase in glioma cells.

Zhong W, Yan T, Lim R, Oberley LW.

Radiation Research Laboratory, Department of Radiology, The University of Iowa, Iowa City 52242, USA.

Four primary antioxidant enzymes were measured in both human and rat glioma cells. Both manganese-containing superoxide dismutase (MnSOD) and copper-zinc-containing superoxide dismutase (CuZnSOD) activities varied greatly among the different glioma cell lines. MnSOD was generally higher in human glioma cells than in rat glioma cells and relatively higher than in other tumor types. High levels of MnSOD in human glioma cells were due to the high levels of expression of MnSOD mRNA and protein. Heterogeneous expression of MnSOD was present in individual glioma cell lines and may be due to subpopulations or cells at different differentiation stages. Less difference in CuZnSOD, catalase, or glutathione peroxide was found between human and rat glioma cells. The human glioma cell lines showed large differences in sensitivity to the glutathione modulating drugs 1,3-bis (2-chloroethyl)-1-nitrosourea (BCNU) and buthionine sulfoximine (BSO). A good correlation was found between sensitivity to BCNU and the activities of catalase in these cell lines. Only one cell line was sensitive to BSO and this line had low CuZnSOD activity.

PMID: 10641728 [PubMed - indexed for MEDLINE]

[Expression of human telomerase reverse transcriptase in cervix cancer and its significance]

[Article in Chinese]

Xi L, Zhu T, Wu P, Xu Q, Huang L, Li KZ, Lu YP, Ma D.

Department of Obstetrics and Gynecology, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan 430030, China.

OBJECTIVE: To investigate the expression of human telomerase reverse transcriptase (hTERT) mRNA and protein in cervix cancer, cervical intraepithelial neoplasia (CIN) and normal cervix. **METHODS:** Expression of hTERT mRNA and the other two subunits of telomerase, human telomerase RNA component (hTR), human telomerase-associated protein (hTP1) was determined by RT-PCR in 3 cervix cancer cell lines, 2 diploid cell lines, 38 cases of cervix cancer, 16 cases of CIN and 20 cases of normal cervix. Telomerase activity was also examined by telomeric repeat amplification protocol enzyme-linked immunosorbent assay (TRAP-ELISA). Expression of hTERT protein was detected in all the cell lines and 101 cases of paraffinized cervix tissue sections. **RESULTS:** hTERT mRNA expression was detected in all of the three cervix cancer cell lines, 81.6% of cervix cancer, 37.5% of CIN, 5.0% of normal cervix, while in neither of the two diploid cell lines. The other two subunits of telomerase were prevalently expressed in all of the cell lines and most cervix tissues. There was a strong correlation between hTERT mRNA expression and telomerase activity. Immunostaining also revealed that hTERT protein was expressed in all three cervix cancer cell lines, 65.5% of cervix cancer, 28.0% of CIN and 4.8% of normal cervix. **CONCLUSION:** Up-regulation of hTERT may play an important role in the development of CIN and cervix cancer, hTERT could be used as an early diagnostic biomarker for cervix cancer.

PMID: 16008894 [PubMed - in process]

ORIGINAL ARTICLE

Gene expression signatures and biomarkers of noninvasive and invasive breast cancer cells: comprehensive profiles by representational difference analysis, microarrays and proteomics

GM Nagaraja¹, M Othman², BP Fox¹, R Alsaber¹, CM Pellegrino³, Y Zeng², R Khanna², P Tamburini³, A Swaroop² and RP Kandpal¹

¹Department of Biological Sciences, Fordham University, Bronx, NY, USA; ²Department of Ophthalmology and Visual Sciences, University of Michigan, Ann Arbor, MI, USA and ³Bayer Corporation, West Haven, CT, USA

We have characterized comprehensive transcript and proteomic profiles of cell lines corresponding to normal breast (MCF10A), noninvasive breast cancer (MCF7) and invasive breast cancer (MDA-MB-231). The transcript profiles were first analysed by a modified protocol for representational difference analysis (RDA) of cDNAs between MCF7 and MDA-MB-231 cells. The majority of genes identified by RDA showed nearly complete concordance with microarray results, and also led to the identification of some differentially expressed genes such as lysyl oxidase, copper transporter ATP7A, EphB6, RUNX2 and a variant of RUNX2. The altered transcripts identified by microarray analysis were involved in cell–cell or cell–matrix interaction, Rho signaling, calcium homeostasis and copper-binding/sensitive activities. A set of nine genes that included GPCR11, cadherin 11, annexin A1, vimentin, lactate dehydrogenase B (upregulated in MDA-MB-231) and GREB1, S100A8, amyloid β precursor protein, claudin 3 and cadherin 1 (downregulated in MDA-MB-231) were sufficient to distinguish MDA-MB-231 from MCF7 cells. The downregulation of a set of transcripts for proteins involved in cell–cell interaction indicated these transcripts as potential markers for invasiveness that can be detected by methylation-specific PCR. The proteomic profiles indicated altered abundance of fewer proteins as compared to transcript profiles. Antisense knockdown of selected transcripts led to inhibition of cell proliferation that was accompanied by altered proteomic profiles. The proteomic profiles of antisense transfectants suggest the involvement of peptidyl-prolyl isomerase, Raf kinase inhibitor and 80 kDa protein kinase C substrate in mediating the inhibition of cell proliferation.

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Keywords: representational difference analysis; microarrays; proteomics; breast carcinoma; biomarkers; copper homeostasis

Introduction

The transformation of a normal cell into a cancer cell has been correlated to altered expression of a variety of genes (Perou *et al.*, 2000; Becker *et al.*, 2005). The expression of some of these genes is a direct result of sequence mutation, whereas other changes occur due to alterations in gene products that participate in specific pathways. The changes in gene expression have been routinely characterized by classical subtraction hybridization and differential display approaches (Cerosaletti *et al.*, 1995; Alpan *et al.*, 1996). With the availability of the human genome sequence and sequences for a number of other model organisms, traditional methods have largely been replaced by gene microarrays (Khan *et al.*, 2001). These analyses have been used to characterize the molecular basis of a variety of diseases including cancer. A comprehensive analysis of a large number of cancer cell lines allowed clustering of genes into groups based on their expression patterns in phenotypically related cell lines (Khan *et al.*, 2001; Dan *et al.*, 2002; Rosenwald *et al.*, 2002; van't Veer *et al.*, 2002). The results of profiling experiments indicated expression of specific gene clusters in cell lines that have the same origin or have arisen from the same organ (Ross *et al.*, 2000). A complementary approach that has been used in limited ways is proteomics. Proteomics scores for changes in different proteins and peptides in cells with characteristic phenotypic differences. However, a comparative analysis of transcripts and proteins to establish a relationship between transcript changes and protein levels has not yet become routine.

Although expression profiling of tumor tissue and its comparison with normal tissue, in principle, is most appropriate to obtain the genetic signatures of a tumor type, such comparisons have not been free of attendant

Correspondence: Dr RP Kandpal, Department of Biological Sciences, Larkin Hall, Room 250, 441 E Fordham Road, Fordham University, Bronx, NY 10458, USA.

E-mail: kandpal@fordham.edu

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complications. These complications arise due to heterogeneity of tumor specimens wherein any cell type-specific changes are likely to be masked by other cell types that constitute the tumor specimen. For this reason, well-characterized cell lines established from tumor tissue may prove more informative and have been considered useful by cancer researchers. Comparing gene profiles between cell lines has the potential to reveal genes that could be causative for the phenotype and other genes that can serve as tumor biomarkers.

Our investigations are aimed at designating a subset of transcripts that could distinguish a normal breast cell from a breast cancer cell and help to predict tumorigenic or metastatic potential of a transformed cell. We describe here transcript and proteomic profiles of a normal breast cell line, a tumorigenic but noninvasive breast carcinoma cell line and an invasive breast carcinoma cell line, and summarize them as a set of candidate biomarkers or targets for therapeutic intervention. The comparison of transcript profiles with proteomic profiles demonstrated that altered proteins were not always represented in the microarray designated profiles and *vice versa*. Furthermore, we have targeted five transcripts that were upregulated in MCF7 cells for investigating their role in cell proliferation pathways. The proteomic profiles have revealed that inhibition of cell proliferation by antisense knockdown was mediated by a specific set of proteins.

Results

Representational difference analysis

As described in the Materials and methods section, RDA was performed by using cDNAs from MCF7 and MDA-MB-231 as tester/driver or driver/tester combination. The difference product in the first case represents the genes that are either upregulated in or specific to MCF7. On the other hand, the difference product of MDA-MB-231 (tester) and MCF7 (driver) hybridization resulted in the isolation of cDNAs that are either upregulated in or specific to MDA-MB-231. The initial linkers used in this protocol had internal *Bgl*III sites. One strand of the linker was used to amplify both the tester and driver cDNAs after linkers had been ligated to cDNAs. After removal of linkers from amplified cDNAs by digestion with *Bgl*III, a dephosphorylated *Bgl*III adaptor was ligated to tester DNA. The *Bgl*III adaptor had an internal *Eco*RI site. The difference product was digested with *Eco*RI and cloned in pBlue-Script vector. The cloning efficiency of the difference product was very low (5×10^4 c.f.u./ μ g of DNA). The low efficiency of cloning is attributed to a substantial fraction of amplified DNA product that is refractory to restriction digestion. The sequencing of a set of 100 clones each from the difference libraries revealed 50 different kinds of clones. The majority of these sequences were short fragments and represented either 3' regions or internal fragments of transcripts. A summary of these clones is presented in Table 1. The

involvement of the majority of these cDNAs is well characterized either in tumorigenesis or in metastasis. The phenotypic characteristics of MCF-7 and MDA-MB-231 ideally match with the biological significance of these genes. The alterations in transcripts for Rho signaling proteins, Ca^{2+} binding/requiring proteins, tight junctions/anchoring junctions/gap junctions, copper binding or sensitive proteins, and RUNX2 are particularly noteworthy.

The differential expression of a representative number of RDA clones was validated by semiquantitative PCR. As shown in Figure 1, these transcripts were either specific to or upregulated in the cell line that was used as a tester. Such analyses demonstrated that more than 90% of the clones were differentially expressed. The abundance of transcripts and the results of RT-PCR were also confirmed by Northern blotting (Figure 2). The pattern of hybridization clearly indicates that all these transcripts showed differential expression in MCF7 and MDA-MB-231 cells that were used as driver/tester combinations for the RDA.

Gene microarrays

After obtaining preliminary molecular signatures of MCF7 and MDA-MB-231 cells by RDA, we used Affymetrix gene microarrays to expand the above analysis to identify a comprehensive set of transcripts that is deregulated in invasive breast carcinoma cells. The comparisons of cell lines on the basis of transcripts that are either present or absent as shown in Figure 3 revealed that a set of 123 genes distinguishes MDA-MB-231 cells from MCF7 and MCF10A. These genes can be classified by their involvement in functional classes such as transcription, signal transduction, cell adhesion, cell cycle, metabolism, transport, response genes and development (Figure 4). The majority of these genes participated in the process of signal transduction followed by transcription, cell adhesion and metabolism, respectively. A few transcripts in these classes were tested by real-time RT-PCR to confirm their altered abundance. The selected transcripts showed changes ranging between two- and 10-fold, 11- and 20-fold and greater than 20-fold, and were in close agreement with the results of microarray analysis. The qualitative pattern of change observed in microarrays analysis was readily reproduced by real-time or semiquantitative RT-PCR for all transcripts tested.

The number of altered transcripts was over 1000 based on a change of twofold or greater, and a majority of these genes show changes varying between two- and fourfold (Figure 5). Interestingly, with all comparisons combined, there were 21 genes downregulated more than 50-fold and 55 genes that were upregulated more than 18-fold when specific cell line pairs were compared (Figure 5). The transcripts that represent the extremes of upregulated and downregulated scale can allow distinction between MCF7 and MDA-MB-231 cells. These transcripts include GPCR11, cadherin 11, annexin A1, vimentin, lactate dehydrogenase B (upregulated in MDA-MB-231) and GREB1, S100A8, amyloid

Table 1 Differentially expressed transcripts identified by RDA

| Upregulated in MDA-MB-231 | Downregulated in MDA-MB-231 |
|--|---|
| <i>Extracellular matrix/matrix-crosslinking proteins</i> LOX Laminin β 1 Collagen VI- α 1 | <i>Calcium-binding proteins</i> Calgranulin B |
| <i>Calcium-binding proteins</i> Reticulocalbin 1 S100A8 Cullin 5 | <i>Transcription factors/promoter-binding proteins:</i> Chromosome 4 ORF Estrogen receptor 1 RUNX2 variant (exon 8 deleted) |
| <i>Transcription factors/promoter-binding proteins</i> RUNX2 c-Jun Fra-1 | <i>Cell-cell adhesion/cell-surface receptor proteins</i> Claudin 3 Amyloid β precursor protein Triose phosphate isomerase Plakoglobin Cdh 1 Cdh 3 Annexin A9 RAR- α Connexin 31 |
| <i>Cell-cell adhesion/cell-surface receptor proteins:</i> Cdh11 CYK61 MHC class II antigen γ chain Protease-activated receptor-1 Protease-activated receptor-4 | <i>ATPase/GTPase and signal transduction proteins</i> RhoD RhoB TGF- β R1 |
| <i>ATPase/GTPase and signal transduction proteins</i> ATP7a Caveolin 2 AXL receptor tyrosine kinase Rho GEF 3 Rho/Rac GEF 18 P21-Rac2 | <i>Stress-response proteins</i> Protein kinase H11 |
| <i>Metalloproteases and MMP inhibitor proteins</i> TIMP-2 MT1-MMP | <i>Cytoskeletal component and binding proteins</i> Keratin 18 Tubulin δ 1 Microtubule-associated protein τ |
| <i>Stress-response proteins</i> Dual specificity phosphatase (DUSP) | <i>Cell-cycle regulation and growth/differentiation/apoptosis proteins</i> S100A13 S100C Aurora kinase AIK2 Nucleosidediphosphate kinase |
| <i>Cytoskeletal component and binding proteins</i> Moesin Vimentin Filamin B | <i>Secreted proteins and growth factors</i> Treffol factor 3 (TFF3) Treffol factor 1 (TFF1) Four and a half LIM domain1 Solute carrier family 16 SLC16A6 |
| <i>Cell-cycle regulation and growth/differentiation/apoptosis proteins</i> Cyclin B1 Cyclin E Cyclin A2 Bcl2-like 1 protein | <i>DNA replication</i> DNA replication complex GINS-PSF2 |
| <i>Secreted proteins and growth factors</i> Milk fat globule protein TGF- α SMAD-specific E3 ub ligase 2 | <i>Miscellaneous</i> Serine protease inhibitor type 1 (SPINT1) Human homolog of <i>Xenopus</i> protein XAG Hypothetical protein FLJ22222 Hypothetical protein 20171 Hypothetical protein MGC3265 |
| <i>Miscellaneous</i> Prion protein | |

β precursor protein, claudin 3 and cadherin 1 (down-regulated in MDA-MB-231). The distinction between MCF7 and MCF10A may be made based on keratin 19, serine protease, amyloid β precursor, neuropeptide Y receptor Y1 (upregulated in MCF7) and caldesmon, annexin A1, epithelial membrane protein 1, S100A2,

keratin 15 (downregulated in MCF7). Likewise, MDA-MB-231 cells differ from MCF10A in vimentin, epithelial membrane protein 3, cadherin 11, GPCR 116, collagen type XIII α 1, Bcl2-associated athanogene 2 (upregulated in MDA-MB-231) and keratin 15, cystatin A, cadherin 1, CD24, calcium-activated chlor-

ide channel, S100P, GPCR 87 (downregulated in MDA-MB-231). Thus, a small subset of transcripts may serve as an accurate signature of these cell lines. Several of these gene products have been shown to participate in tumorigenesis and invasiveness of breast carcinoma cell lines.

The invasiveness phenotype of MDA-MB-231 cells specifically relates to changes in the following functional classes: (a) cell adhesion molecules, (b) Ca^{2+} requiring, Ca^{2+} binding or Ca^{2+} regulatory genes, (c) copper-sensitive or copper-transporting proteins and (d) specific regulatory proteins of Rho signaling. Among these functional groups, 23 transcripts involved in cell-cell or cell-matrix interactions are underexpressed in MDA-

MB-231 cells and 21 transcripts were overexpressed in this cell line (Table 2). The comparison of Ca^{2+} -requiring/binding genes indicated downregulation of 26 transcripts and upregulation of 26 transcripts in invasive cells as compared to noninvasive cells (Table 3). While Ca^{2+} homeostasis is extensively investigated in human cancers, copper homeostasis is an underexplored area. The alterations in copper homeostasis in breast carcinoma cells were reflected by changes in transcripts corresponding to a variety of copper-binding or copper-sensitive proteins/enzymes (Tables 1 and 4). The deregulation of Rho signaling was evident from changes in various proteins involved in this pathway (Table 4).

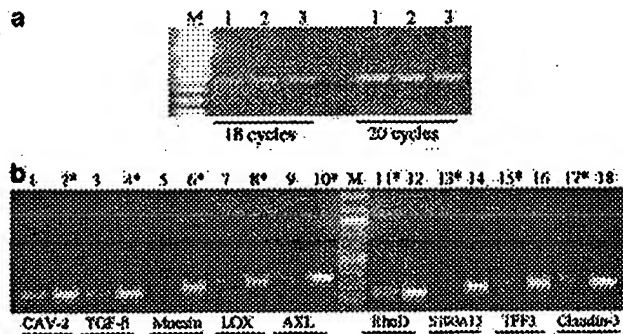


Figure 1 Semiquantitative evaluation of selected transcripts. (a) RNA was isolated from confluent culture dishes containing MCF10A (lanes 1 and 4), MCF7 (lanes 2 and 5) or MDA-MB-231 (lanes 3 and 6) cells. The amount of RNA was first determined spectrophotometrically. Equal amounts of RNA, as determined by absorbance at 260 nm, were amplified with primers specific to actin gene for 18 cycles (lanes 1–3) or 20 cycles (lanes 4–6). The lane containing size markers is labeled as M. (b) A set of primers corresponding to caveolin 2 (lanes 1 and 2), TGF- α (lanes 3 and 4), Moesin (lanes 5 and 6), LOX (lanes 7 and 8), Axl receptor (lanes 9 and 10), RhoD (lanes 11 and 12), S100A13 (lanes 13 and 14), TFF3 (lanes 15 and 16) and Claudin 3 (lanes 17 and 18) were amplified for 32 cycles. Lanes 1, 3, 5, 7, 9, 12, 14, 16 and 18 represent amplified products from MCF7 and lanes 2, 4, 6, 8, 10, 11, 13, 15 and 17 represent MDA-MB-231 cells. The lanes containing PCR products from MDA-MB-231 cells are marked with an asterisk.

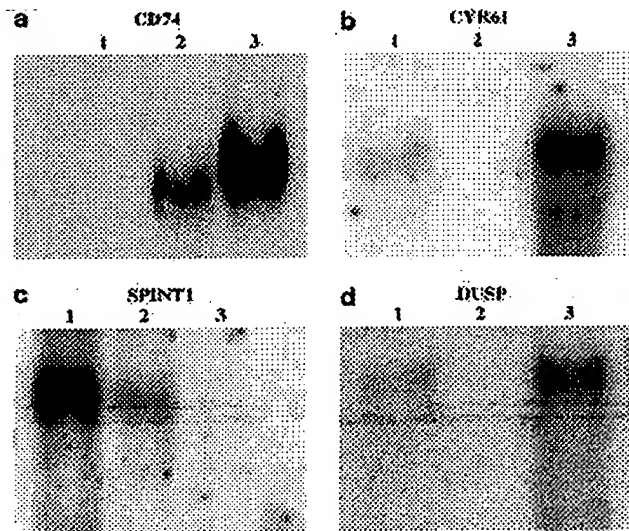


Figure 2 Analysis of selected transcripts by Northern hybridization. The probes specific to CD74 (a), CYR61 (b), SPINT1 (c) and DUSP (d) were labeled with a ^{32}P nucleotide and hybridized to blots containing RNA from MCF10A (lane 1), MCF7 (lane 2) and MDA-MB-231 (lane 3). The blots were washed stringently and developed as described. The amounts of RNA loaded were normalized as in Figure 1.

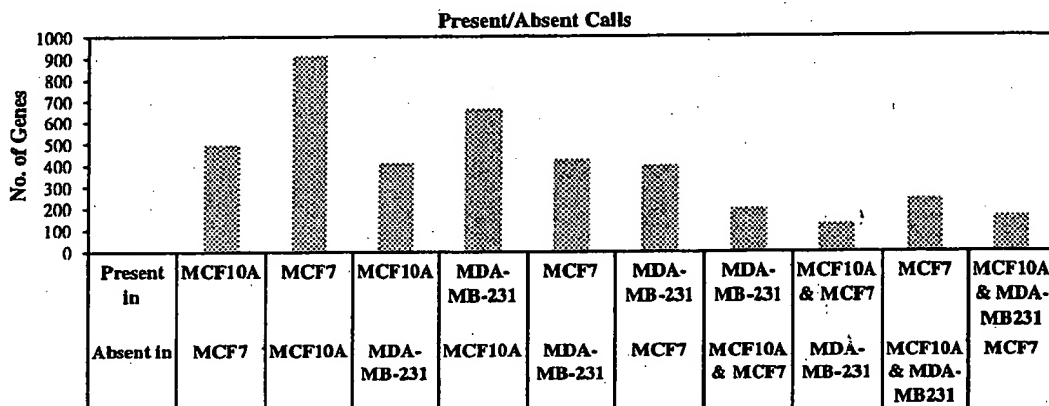


Figure 3 Comparison of cell lines based on the presence or absence of transcripts. The absence or presence of a transcript in the Affymetrix chip was scored by the fluorescence read-out as described in the Materials and methods section.

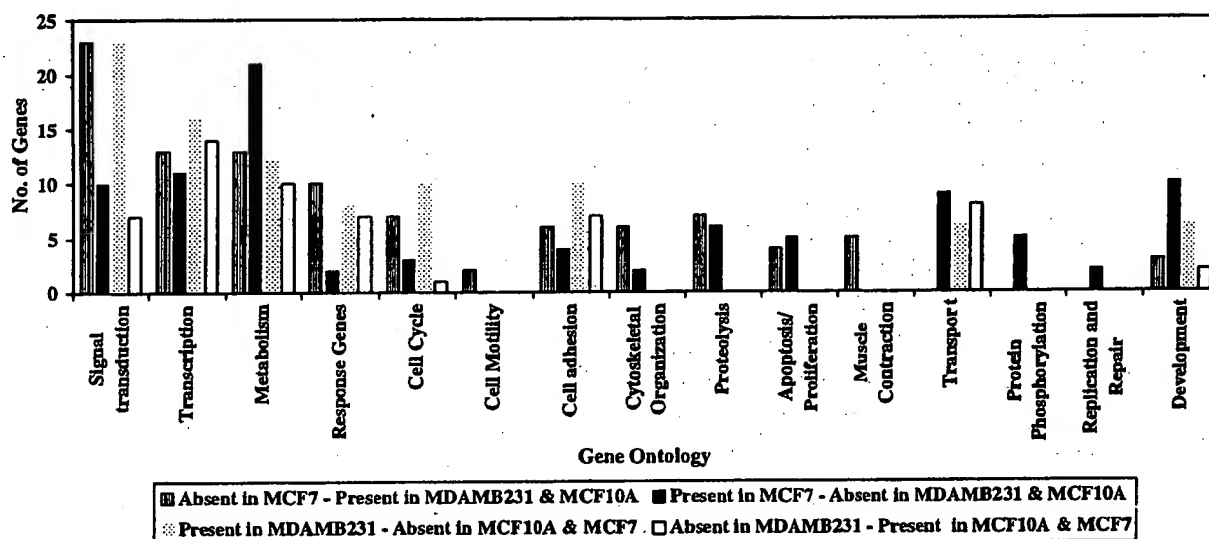


Figure 4 Functional classes of transcripts that differentiate a cell line pair. The transcripts identified as present or absent were classified based on their functional importance.

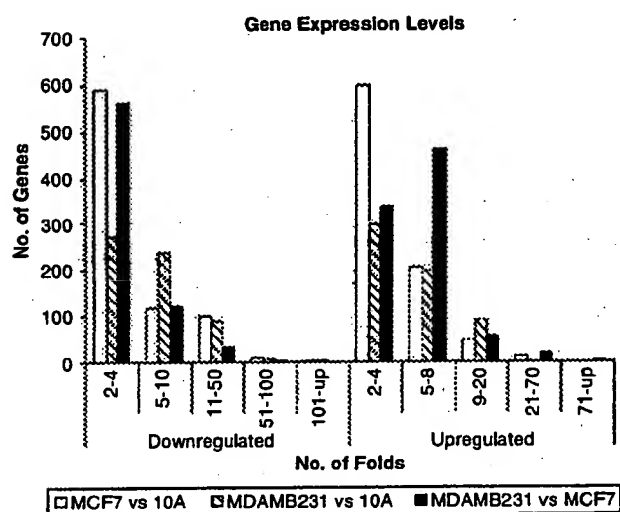


Figure 5 Distribution of altered number of transcripts as a function of fold change. The altered transcripts were categorized in groups based on the magnitude of change in their abundance.

Table 2 Cell adhesion molecules altered in breast carcinoma cells

| Upregulated in MDA-MB-231 | Downregulated in MDA-MB-231 |
|------------------------------|-----------------------------------|
| Cadherin 4 | Claudin3 |
| Cadherin 11 | Cadherin-1 |
| Catenin | Cadherin-3 |
| Integrin α 6 | Cadherin-18 |
| Transmembrane anchor protein | Cadherin, LAG seven pass receptor |
| Eph B2 | Down syndrome cell adhesion |
| Dystonin | Catenin- δ 2 |
| Laminin β 1 | Eph A4 |
| Lamin | Ephrin A4 |
| Filamin B | Annexin A9 |
| Filamin C | Ankyrin 3 |
| Tailin 1 | Sarcoglycan |
| Butyrophilin | Keratin 8 |
| Spectrin- α | MAP-7 |
| Spectrin- β | MAP- τ |
| Thrombospondin | Plakoglobin |
| Plastin 3 | Plakophilin |
| Adducin 3 γ | Discoidin domain receptor |
| Lamin B receptor | Zona occludens 3 |
| Laminin β 2 | Periplakin |
| Lamin A/C | Protocadherin α 9 |
| | Laminin γ 2 |
| | Laminin α 3 |

Proteome analysis

To identify altered abundance of proteins and relate it to transcript profiles, we characterized the protein profiles of MCF-10A, MCF-7 and MDA-MB-231 cells. Typically, > 300 protein spots could be visualized in silver-stained gels, and there were far fewer protein spots in gels that were stained with Coomassie blue. The comparison of MCF7 or MDA-MB-231 proteins with MCF-10A revealed that MCF-7 had 11 unique protein spots, while MDA-MB-231 had 15 spots that were not seen in MCF-10A. These proteins were either specific to or upregulated in these cell lines. The identity of these proteins is shown in Table 5. Out of these 26 protein spots, only 25 yielded amino-acid sequence. As shown in the table, the list includes proteins involved in stress

response, protein-tagging activities, calcium-binding and calcium homeostasis proteins and some regulatory proteins. Prominent among these changes were proteins involved in calcium homeostasis such as calreticulin, calyculin and reticulocalbin. The changes in signaling pathways between the two cell lines were indicated by altered levels of Rho GDP dissociation inhibitor 1, an apoptosis/differentiation regulating protein galectin, Myc expression regulator far upstream binding protein-1 and the microtubule regulator protein stathmin. The translation initiation factors 5A and 4H were also selectively upregulated in MDA-MB-231 cells.

Table 3 Calcium binding or sensitive transcripts

| Upregulated in MDA-MB-231 | Downregulated in MDA-MB-231 |
|--|--|
| Reticulocalbin | S100A8 |
| Dystonin | S100A7 |
| Follistatin like 1 | S100A13 |
| Cullin | Tumor associated Ca ²⁺ signal transducer |
| Annexin A5 | Notch homolog 3 |
| EF hand domain containing-2 | PKD2 |
| Hippocalcin like-2 | Adenylate cyclase |
| LDLR | Phospholipase-C |
| Steroid sulfatase | Chemokine ligand 12 |
| MT-actin crosslinking factor | Ubiquitin-specific protease |
| Inositol-1,4,5-triphosphate receptor 3 | PKC- η |
| Sorcin | Ret-protooncogene |
| Guanine nucleotide-binding protein- γ | Signal peptide-CUB domain |
| GAS6 | Mannosidase α |
| SWP-70 protein | Bradykinin receptor B2 |
| Jagged 1 | Solute carrier family 24 |
| EGF-containing fibrulin-like ECM | Ca ²⁺ channel voltage-dependent β 3 |
| Transglutaminase 2 | Regulator of G protein signaling 17 |
| Thrombin receptor-like 1 | Dystrobin- α |
| Plastin 3 | Synaptogamin 1 |
| FYN oncogene | Matrix gla protein |
| PKC- α | EF hand domain family member D1 |
| Calnegin | PK-cyclic AMP dependent |
| Calpain | ATPase-Ca ²⁺ transporting |
| HEG homolog | Calmodulin 1 |
| Cyr61 | CaM kinase |

Table 4 Altered transcripts involved in copper homeostasis and Rho signaling

| Copper-binding proteins | Rho signaling proteins |
|-------------------------------------|--|
| LOX | Rho3 |
| LOX-1, LOX-2 | Rho/Rac GEF 18 |
| SCO cyt oxidase-deficient homolog 2 | Rho GEF 12 |
| COX 17 homolog | Ras related C3 botulinum toxin substrate 2 |
| Metallothionein 1E, 1F and 2a | Cdc 42 effector protein 3 |
| Ring finger protein7 | Rho GEF 3 |
| Amiloride-binding protein 1 | Rho GDP dissociation inhibitor β |
| Neurotrypsin/motopsin | RhoD |

Changes in proliferation characteristics and protein profiles in response to transfection with antisense constructs of selected transcripts

We had observed significant upregulation of transcripts for DNA replication complex protein GINS PSF2, trefoil factor 3, aurora kinase AIK2, protein kinase H11 and secreted protein XAG in MCF7 cells. We reasoned that antisense knockdown of the above genes in MCF7 cells might indicate pathways involved in tumorigenesis and invasiveness.

MCF7 cells were transfected with empty vector pCDNA3.1 or antisense constructs of the above genes. A semiquantitative amplification of pCDNA marker gene by RT-PCR confirmed the presence of the transfected construct in a significant proportion of the cell population. The transfected cells also showed a decrease in the target transcripts as observed by RT-

Table 5 Proteomic profiles of MCF7 and MDA-MB-231 cells as compared to MCF10A cells

| MCF7 cells | MDA-MB-231 cells |
|--|---|
| <i>Cell-cell adhesion/cell-surface receptor proteins</i> | <i>Calcium-binding proteins</i> |
| Triose-phosphate isomerase | Calcyclin |
| | Calreticulin |
| | Crocalbin |
| | Reticulocalbin |
| <i>Stress-response proteins</i> | <i>Transcription Factors/Promoter-binding proteins</i> |
| Hsp27 | Far upstream element binding protein-1 |
| Superoxide dismutase | Far upstream element binding protein-2 |
| Peroxioredoxin 2 | |
| <i>Cytoskeletal component/binding proteins</i> | <i>Cell-cell adhesion/cell-surface receptor proteins</i> |
| Stathmin | Galectin |
| <i>Cell-cycle regulation and growth/differentiation/apoptosis proteins</i> | <i>ATPase/GTPase/signal transduction/trafficking proteins</i> |
| Nucleoside diphosphate kinase A | Rho GDP dissociation inhibitor 1 |
| S100C | |
| <i>Secreted proteins and growth factors</i> | <i>Stress-response proteins</i> |
| Macrophage migration inhibitory protein | Hsp 70 |
| | Peroxioredoxin 2 |
| <i>Miscellaneous</i> | <i>Cytoskeletal component/binding proteins</i> |
| Cyt c oxidase VIb | Stathmin |
| Peptidyl-prolyl <i>cis-trans</i> isomerase | |
| Ubiquitin | <i>Miscellaneous</i> |
| | Heterogeneous nuclear ribonucleo protein H |
| | eIF4H (translation) |
| | eIF5A (translation) |

PCR. The effects of antisense transfections were scored by growth characteristics of the transfectants. The cell proliferation was reduced between 15 and 40% when antisense transfectants were compared to cells transfected with empty vector.

In order to relate decreased proliferation of transfectants to altered proteins, proteomic profiles of transfectants were compared with vector controls. The comparison of protein profiles of cells transfected with empty vector or antisense construct revealed alterations in several proteins for each transfectant (Table 6). These proteins included stress-response proteins, calcium-regulating proteins, translation factors, ubiquitin, proteins of electron transport chain and oxidative phosphorylation, signaling proteins, cytokeratins, actin and actin regulating proteins and general regulatory factors. The number of altered proteins varied between 5 and 15 for various transfections. Peptidyl prolyl *cis-trans* isomerase, calcium-regulating proteins, SOD, galectin, histidine triad protein and PKC substrate were prominent among altered proteins. We performed database searches to identify interactors for all proteins that were altered in transfected cells and identified nearly 350 proteins (data not shown). A significant number of these interacting proteins are involved in transcriptional regulation.

Table 6 Altered proteins in MCF7 cells after antisense knockdown of specific transcripts

| After transfection with as-trefoil factor 3 (TFF3) | After transfection with as-protein kinase H11 |
|---|---|
| Calcium-binding proteins Calmodulin Cell-cell adhesion/cell-surface receptor proteins Retinoic acid-binding protein II ATPase-GTPase/signal transduction/trafficking proteins Raf kinase inhibitor Stress-response proteins Hsp27 Peroxiredoxin I Cytoskeletal component/binding proteins Cofilin-nonmuscle isoform Cell-cycle regulation and growth/differentiation/apoptosis proteins Chromobox protein homolog 1 Chromobox protein homolog 3 Translationally controlled tumor protein Miscellaneous 40S ribosomal protein S12 ATP synthase D chain Cancer-associated Sm-like protein Cyt c oxidase polypeptide Va eIF5A (translation) Ferritin heavy chain His triad nucleotide-binding protein Histone H2B.n Peptidyl-prolyl isomerase Proteasome subunit β -type 6 RNA-binding protein 8A Thioredoxin Ubiquitin crossreactive protein After transfection with as-aurora kinase AIK2 Calcium-binding proteins Calgranulin B Calgranulin A Stress-response proteins Hsp27 Superoxide dismutase Miscellaneous 60S acidic ribosomal protein P2 Peptidyl-prolyl isomerase Ubiquitin-crossreactive protein | Cell-cell adhesion/cell-surface receptor proteins Galectin Retinoic acid-binding protein II ATPase-GTPase/signal transduction/trafficking proteins PKC substrate Stress-response proteins Hsp27 Cytoskeletal component/binding proteins Actin- α , skeletal muscle Cytokeratin 18 Miscellaneous 3OH-acyl CoA dehydrogenase II ATP synthase A chain Cyt c oxidase peptide Va Enhancer of rudimentary homolog Thioredoxin Ubiquinol-cyt C reductase No match After transfection with as-DNA replication complex GINS PSF2 Calcium Binding proteins: Calreticulin ATPase-GTPase/signal transduction/trafficking proteins 14-3-3 η PKC substrate Cytoskeletal component/binding proteins: Cytokeratin 1 Cytokeratin 18 Cell-cycle regulation and & growth/differentiation/apoptosis proteins Chromobox protein homolog 1 Chromobox protein homolog 3 MAP/MT affinity regulator Miscellaneous ATP synthase D chain Peptidyl-prolyl isomerase Ubiquinol-cyt c reductase After transfection with as-human homolog of XAG Calcium-binding proteins Calreticulin Stress-response proteins Hsp27 Miscellaneous ATP synthase A chain Peptidyl-prolyl isomerase Ubiquitin crossreactive protein No match |

Discussion

The results presented here validate the gene profiles obtained from different expression platforms ranging from subtractive hybridization to gene microarrays and proteomic analysis. The RDA protocol is powerful enough to yield important genes that show significant alterations in their expression between cell lines, and can lead to isolation of full-length cDNAs by using appropriate modifications (Baskaran *et al.*, 1996; Jacob *et al.*, 1997). The detection of RUNX2, variant of RUNX2, EphB6, prion protein, lysyl oxidase and a copper transporter ATP7A transcripts by RDA warrant specific mention. RUNX transcription factors bind specific motifs on target gene promoters and regulate gene expression leading to cell growth, proliferation and

differentiation (Pratap *et al.*, 2003). RUNX2 and its variant have differential repression activity toward the promoter of the cyclin-dependent kinase inhibitor (p21 CIP1) (Westendorf *et al.*, 2002). The loss of EphB6 expression due to methylation of its promoter is related to invasiveness of MDA-MB-231 (Fox and Kandpal, 2004; unpublished observations). Lysyl oxidase, a copper-sensitive enzyme, causes oxidative deamination of lysine and hydroxy lysines of collagen to aldehyde forms to stabilize collagen fibrils (Siegel, 1976) that are found in invasive breast carcinoma cells (Akiri *et al.*, 2003). The activation of LOX is dependent on copper that is internalized and then transported to trans golgi network by copper transporter ATP7A (Pase *et al.*, 2004), a protein mutated in Menkes disease (Moller *et al.*, 2005). Prion protein has also been reported as a

chaperone for copper (Jones *et al.*, 2004). Thus, EphB6 can serve as a marker for invasiveness, and LOX and ATP7A may be exploited as relevant targets for therapeutic intervention.

The downregulation of junctional proteins along with inactivation of TIMPs as shown here is in agreement with other reports describing their relationship with invasiveness of carcinoma cells (Johnson, 1991; Kousidou *et al.*, 2004; Shao *et al.*, 2005) and promoter methylation (Costa *et al.*, 2004). As several transcripts coding for junctional proteins are downregulated in invasive cells, we postulate that methylation-specific PCR can be exploited to use these transcripts as biomarkers of tumor cells in general and invasiveness in particular. The changes in cell-cell interaction correlate to cell phenotypes because such interactions influence Rho/Ras signal transduction pathways and *vice versa* (Malliri and Collard, 2003; Nagaraja and Kandpal, 2004; Ridley, 2004), and lend credence to the significance of altered transcripts for Rho and Rho GEFs as presented here.

Early changes in calcium homeostasis as measured by calcium excretion have been reported in breast cancer (Campbell *et al.*, 1983), and altered calcium signaling has been shown in invasive lung carcinoma cells (Amuthan *et al.*, 2002). Prominent among calcium-binding proteins are S-100 protein, a group of intracellular messengers that respond to transient changes in calcium concentration by binding to specific receptors (Marenholz *et al.*, 2004) and regulate cell growth, differentiation and motility, transcription and cell cycle. The S-100 proteins detected in the present study map to chromosome 1q21, a region of genome that is frequently altered in human breast cancer cells (Bieche *et al.*, 1995). Calcium ions act as a second messenger in specific signaling pathways in a variety of cancers (Missiaen *et al.*, 2000) and are known to alter calcineurin to activate transcription factors such as NFATc (Luo *et al.*, 1996).

As dictated by post-transcriptional regulation, protein profiles showed far fewer changes as compared to transcript profiles, and the knockdown of five selected genes in MCF7 cells produced interesting changes in protein profiles. These genes, namely, XAG (secretory *Xenopus laevis* protein), trefoil factors 3, human aurora2 kinase AIK2, protein kinase H11 and DNA replication complex GINS PSF2, have been shown to be estrogen responsive, oncogenic or involved in tumorigenesis (Yu *et al.*, 2001; Fletcher *et al.*, 2003; Katoh, 2003; Warner *et al.*, 2003; Takayama *et al.*, 2003). The antisense constructs of these genes appeared to work as siRNAs as suggested by the reduction in the transcript detected in RT-PCR of RNA isolated from the transfected cells. The involvement of the above transcripts in invasive potential is apparent from the observed upregulation of calcium-binding proteins in transfected MCF7 cells, which is comparable to their levels in MDA-MB-231 cells. The proteins that appear to mediate inhibition of proliferation in antisense-transfected cells include PKC substrate, Raf kinase inhibitor, histidine triad nucleotide-binding protein and peptidyl-prolyl isomerase (Pin1). We believe histidine triad protein effects are

most likely mediated via its interaction with ATM protein. Raf kinase inhibitor (Keller *et al.*, 2004) and ATM (Hall, 2005) have been conclusively linked to transformation of cells, and the activity of Pin1 has been related to p53-mediated signaling pathways (Mantovani *et al.*, 2004; Berger *et al.*, 2005). In this context, p53 activation has also been hypothesized by Cu-SOD prion-like enzyme (Wiseman, 2005). Thus, alterations in copper homeostasis and p53-mediated signaling may be considered as a significant regulatory mechanism in tumorigenesis.

In summary, we have presented here a set of candidate genes that can serve as biomarkers for tumorigenesis and invasiveness, and some of these markers may be used to develop DNA-based diagnostic tests. The alterations in transcripts for copper homeostasis genes suggest copper chelation or inhibition of copper transporter ATP7A as potential targets for therapeutic application. The modulation of RUNX2 splicing variants by chemicals that affect splicing machinery may also be explored as a therapeutic modality. The changes in EphB6 expression, if confirmed in tumor specimens, may have prognostic significance.

Materials and methods

Breast cancer cell lines

We used MCF-10A, a cell line established from normal breast, and two breast carcinoma cell lines MCF-7 and MDA-MB-231 that vary in their *in vitro* and *in vivo* invasiveness. All cells were cultured at 37°C/7% CO₂. MCF-10A cells were grown in 1:1 DMEM:F12 media (Gibco) with 5% horse serum (Gibco), 20 mM HEPES, 10 ng/ml EGF (Invitrogen), 10 ml/l PenStrep-Glutamine (10 000 U/ml penicillin, 10 000 µg/ml streptomycin and 29.2 mg/ml L-glutamine), 10 µg/ml insulin (Invitrogen), 0.1 µg/ml Cholera Toxin (Sigma) and 500 ng/ml hydrocortisone (Sigma). MCF-7 and MDA-MB-231 cells were grown in DMEM (Gibco) supplemented with 2 mM L-glutamine (Gibco), 1 mM sodium pyruvate (Gibco), 5 ml/l penstrep (5000 U/ml penicillin and 5000 µg/ml streptomycin), and 10% fetal bovine serum (Hyclone).

Total RNA isolation

RNA was isolated from 85 to 95% confluent 10 cm tissue culture dishes using TRI reagent (Molecular Research Center Inc.) with slight modifications to the recommended protocol. Approximately 10 million cells were mixed with 1.0 ml Tri Reagent, the mixture was extracted with chloroform and the aqueous phase containing RNA was separated. The RNA was precipitated with isopropanol, the pellet washed sequentially with 80 and 100% ethanol, then dried and resuspended in DEPC-treated water. RNA was stored in aliquots at -70°C. The quality of RNA was visualized by running on a formaldehyde gel. The appearance of ribosomal RNA bands indicated that RNA was not degraded during the procedure. The amount of RNA was determined by its absorbance at 260 nm.

DNAase treatment of total RNA

To remove DNA contamination, 20 µg of RNA (quantified spectrophotometrically) was treated with 500 ng DNAase I,

80 U RNasin (Promega) and 1 mM MgCl₂ in Tris buffer in a total volume of 50 μ l. The reaction was carried out at 37°C for 1 h and the DNAase inactivated by heating to 65°C for 30 min.

Representational difference analysis (RDA)

RDA of cDNAs is a modification of genomic RDA (Lisitsyn et al., 1993). We performed RDA in the following two ways. In one experiment, MCF-7 cDNA was used as a driver and MDA-MB-231 cDNA as a tester. In the second experiment, MCF-7 cDNA was used as a tester and MDA-MB-231 cDNA as a driver. The protocol has been described previously (Jacob et al., 1997). Briefly, first-strand synthesis was carried out using a commercial cDNA synthesis kit as per the manufacturer's protocol. A linker with a BglIII site was ligated to the tester as well as the driver cDNA. A primer specific to one of the linker strands was used to PCR amplify the linker-ligated cDNAs. The linkers were then removed by digesting the cDNA with BglIII and the digested cDNA was gel purified. A second set of unphosphorylated BglIII adaptors was ligated to the tester cDNA only. The tester and driver DNAs were hybridized in a 5 μ l reaction volume at a ratio of 1:40. After hybridization, the ends of the tester homoduplexes were repaired with Klenow polymerase and 1 μ l of the reaction mixture was diluted to 100 μ l. The difference product was obtained by amplifying 1 μ l of the diluted mixture using the top strand of the ligated adaptor as a primer. The amplified difference product was digested with EcoRI and cloned in a pBlueScript vector. Individual clones were picked up and sequenced by Sanger's dideoxy chain termination method. Representative clones were validated by Northern analysis and semiquantitative RT-PCR.

Microarray analysis

The GeneChips, Human Genome U133A 2.0, (Affymetrix, Santa Clara, CA, USA) used in this study contained approximately 22000 probe sets corresponding to 18400 transcripts and variants, including 14500 well-characterized human genes.

Total RNA was converted into double-stranded cDNA by using SuperScript II (Invitrogen, Carlsbad, CA, USA) and an oligo-dT primer containing a heel of the T7 RNA polymerase promoter sequence. The reaction mixture containing double-stranded cDNA was extracted with phenol-chloroform, precipitated with ethanol, and dissolved in 12 μ l RNase-free water. The cDNA was transcribed *in vitro* by using a RNA transcription labeling kit (Enzo Biochem, Farmingdale, NY, USA) with 6 μ l of double-stranded cDNA in the presence of ATP, CTP, GTP, UTP, bio-11-CTP and bio-16-UTP. The biotinylated RNA was purified by using an affinity column (Qiagen, Valencia, CA, USA) and fragmented randomly, by heating to 95°C in the presence of fragmentation buffer, between sizes of 35 and 200 bases. The GeneChips were hybridized overnight at 45°C in hybridization oven in a solution containing fragmented cRNA, control oligonucleotide B2, 20 \times eucaryotic hybridization controls, herring sperm DNA, acetylated BSA and 2 \times hybridization buffer. The GeneChips were washed and stained with streptavidin-phycoerythrin and the antibody in 2 \times MES stain buffer, acetylated BSA, and optically read at a resolution of 6 μ m with a Affymetrix GeneChip scanner 3000. Affymetrix MICROARRAY SUITE³ was used for initial data preparation (generation of .CHP files). Normalization (quantile method) and calculation of signal intensities was performed with the software package RMA from the R project (<http://www.r-project.org/>). For every cell line, three replicates were performed with Affymetrix Gene Chips. The Gene Chip data were used for further calculations after the raw image and

MAS5 analysis revealed a positive quality report. Ratios of average signal intensity (log 2) were calculated for the probe sets between pairs of cell lines and then converted to an average fold change (AFC). Statistical validation was performed on probe sets as described (Yoshida et al., 2004). The statistical method used to assign *P*-values to the fold changes of gene responses is described by Yoshida et al. (2004) and is a two-step procedure based on the Benjamini and Yekutieli construction of false discovery rate confidence intervals (FDRCI) (Reiner et al., 2003). Functional annotation of proteins was assigned through Gene Ontology (<http://www.geneontology.org>) or Locuslink (<http://www.ncbi.nlm.nih.gov/LocusLink>) classifications obtained through appropriate public databases.

Quantitative RT-PCR

RNA was reverse transcribed with SuperScript II (Invitrogen, Carlsbad, CA, USA) RT by priming with oligodT. The primers specific to validated genes were synthesized from the 3' untranslated region using Primer 3 software. PCR reactions were then performed in triplicates in an I-cycler Thermocycler with optical module (BioRad, Hercules, CA, USA). The amplified products were quantified by reading fluorescence of SybrGreen I (Molecular Probes, Eugene, OR, USA). Average fold changes were calculated by differences in threshold cycles (C_t) between pairs of samples to be compared. *HPRT* gene was used as a control.

Semiquantitative RT-PCR

The spectrophotometrically determined concentration of RNA was confirmed by amplifying actin message at different cycles. The cycling conditions that yielded proportional increment of amplified product was used to normalize the RNA concentration. The normalized RNA was used as template to determine relative abundance of transcripts corresponding to clones identified by RDA experiments. The conditions were standardized in the range of cycles that yielded a PCR product for at least one of the pairs of compared RNAs. Such experiments dictated cycles between 30 and 35 to be appropriate to compare abundance of selected transcripts in MCF7 and MDA-MB-231 cells.

Northern analysis

The expression pattern of selected transcripts in cell lines was also analysed in Northern blots. RNA (20 μ m), as determined by spectrophotometer and confirmed by actin amplification, was electrophoresed on a formaldehyde agarose gel. A RNA ladder was used as a size marker. The RNAs were transferred from the gel to a Hybond nylon membrane by capillary transfer. The RNA was fixed onto the membrane by irradiation in a Stratalinker. The blot was hybridized at 65°C for 12–15 h with a radioactive probe and the blot was subsequently washed with 0.1 \times SSC and 1% SDS at 65°C. The hybridized probe was detected by autoradiography.

Transfection of MCF-7 cells with antisense constructs

The genes selected on the basis of their upregulation were cloned in antisense orientation in pCDNA3.1 vector (Invitrogen). MCF-7 cells were grown to 70–80% confluence. Approximately 4 μ g of DNA was transfected into MCF-7 cells by using Lipofectamine 2000. The transfected cells were grown in the presence of G418 (400 μ g/ml). The transfectants were processed for protein isolation. A control set of cells was transfected with an empty pCDNA3.1. The proteins were analysed by two-dimensional electrophoresis, and altered bands were excised for mass spectrometry.

Protein isolation

The cultured cells were harvested by trypsinization and centrifuged at 220 g for 5 min at 4°C. The cell pellet was washed once with ice-cold 1 × PBS. The proteins were isolated by using a commercial kit (BioRad, Hercules, CA, USA). Briefly, pelleted cells (0.05 ml) were mixed with 0.5 ml ice-cold CPEB solution containing protease inhibitors cocktail (Roche), vortexed and stored on ice for 30 min. The cell suspension was passed through a syringe needle (20 gauge) for 10–20 strokes to ensure complete cell lysis. The cytoplasmic protein fraction was collected by centrifugation at 100 g for 10 min at 4°C. The nuclear pellet was washed once again with 0.5 ml CPEB solution. The nuclear pellet was resuspended in 0.75 ml PSB buffer, vortexed briefly and centrifuged at 1000 g for 10 min at 4°C, and the supernatant containing nuclear protein was collected into a new tube. The samples were quantified using 2D Quant kit (Amersham Biosciences), aliquoted and stored at –80°C to prevent protein degradation. To reduce streaking, background staining and the other gel artefacts associated with substances contaminating 2D/IEF samples, the samples were cleaned with 2D Clean up kit (Bio Rad, Hercules, CA, USA) before running on the gel.

Two-dimensional gel electrophoresis

The protein mixtures were separated based on isoelectric points by using commercial pre-cast pH gradient gel strips according to the manufacturer's instructions. The protein sample (175 µg) in 185 µl of sample buffer (8 M urea, 2% CHAPS, 0.2% biolytes, 3/10 ampholytes, 65 mM DTT and 0.002% bromophenol blue) was loaded in the sample loading trays at the end of 11 cm immobilized rehydrated strips (pH 3–10) (Bio Rad, Hercules, CA, USA). Following isoelectric focusing, proteins were reduced and alkylated by successive 15 min treatments with equilibration buffer (6 M urea, 0.375 M Tris-HCl pH 8.8, 2% SDS, 20% glycerol, 2% DTT) and 2.5% (W/V) iodoacetamide, respectively. Proteins were then resolved in the second dimension on 8–16% gradient SDS-PAGE gel (Bio Rad, Hercules, CA, USA). The protein spots were visualized by staining with either silver stain or

Coomassie blue stain. The gel images were compared and bands showing significant (greater than twofold) alterations in intensity were excised and processed for mass spectrometry. Comparisons were made between protein lysates from MCF-10A, MCF-7 and MDA-MB-231 cell lines or between MCF-7 and MCF-7 cells transfected with specific antisense constructs.

Protein identification by enzymatic digestion followed by mass spectrometry

Prior to performing trypsin digestion, the gel pieces containing protein spots were washed sequentially once with water and twice with acetonitrile. The gel pieces were then allowed to swell in 100 mM ammonium bicarbonate and finally washed with acetonitrile. The washed slices were dried in a Speed Vac concentrator, and subsequently incubated with 20 µl of Promega's autocatalysis-resistant trypsin (12.5 ng/µl in 50 mM ammonium bicarbonate and 5 mM CaCl₂ pH 8.0) overnight at 37°C. The supernatant (5 µl) from tryptic digest was injected for peptide sequence analysis using on-line capillary liquid chromatography-electrospray ionization-tandem mass spectrometry (LC-MS/MS). The front end HPLC utilized a Dionex (San Francisco, CA, USA) Vydac 300 µm inner diameter × 15 mm C18 column. The linear acetonitrile gradient (3%/min, containing 0.02% TFA) was developed using a Hewlett-Packard 1100 pump operating at 0.1 ml/min, and the flow was split before the injector such that the flow rate through the column was 3 µl/min. Peptides were detected at 215 nM. The in-line mass spectrometer was a ThermoElectron LCQ-DECA instrument operated in data-dependent MS/MS mode, and proteins were identified by searching a nonredundant protein database using the Sequest program.

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References

- Akiri G, Sabo E, Dafni H, Vadasz Z, Kartvelishvili Y, Gan N et al. (2003). *Cancer Res* 63: 1657–1666.
- Alpan RS, Sparvero S, Pardee AB. (1996). *Mol Med* 2: 469–478.
- Amuthan G, Biswas G, Ananadatheerthavarada HK, Vijaya-sarathy C, Shephard HM, Avadhani NG. (2002). *Oncogene* 21: 7839–7849.
- Baskaran N, Kandpal RP, Bhargava AK, Glynn MW, Bale A, Weissman SM. (1996). *Genome Res* 6: 633–638.
- Becker M, Sommer A, Kratzschmar JR, Seidel H, Pohlenz HD, Fichtner I. (2005). *Mol Cancer Ther* 4: 151–168.
- Berger M, Stahl N, Del Sal G, Haupt Y. (2005). *Mol Cell Biol* 25: 5380–5388.
- Bieche I, Champeme MH, Lidereau R. (1995). *Clin Cancer Res* 1: 123–127.
- Campbell FC, Blamey RW, Woolfson AM, Elston CW, Hosking DJ. (1983). *Br J Surg* 70: 202–204.
- Cerosaletti KM, Shaperro MH, Fournier RE. (1995). *Genomics* 25: 226–237.
- Costa FF, Verbisck NV, Salim AC, Ierardi DF, Pires LC, Sasahara RM et al. (2004). *Oncogene* 23: 1481–1488.
- Dan S, Tsunoda T, Kitahara O, Yanagawa R, Zembutsu H, Katagiri T et al. (2002). *Cancer Res* 62: 1139–1147.
- Fletcher GC, Patel S, Tyson K, Adam PJ, Schenker M, Loader JA et al. (2003). *Br J Cancer* 88: 579–585.
- Fox BP, Kandpal RP. (2004). *Biochem Biophys Res Commun* 318: 882–892.
- Hall J. (2005). *Cancer Lett* 227: 105–114.
- Jacob AN, Baskaran N, Kandpal G, Narayan D, Bhargava AK, Kandpal RP. (1997). *Somatic Cell Mol Genet* 23: 83–95.
- Johnson JP. (1991). *Metast Rev* 10: 11–22, Review.
- Jones CE, Abdelraheim SR, Brown DR, Viles JH. (2004). *J Biol Chem* 279: 32018–32027.
- Katoh M. (2003). *Int J Mol Med* 12: 3–9.
- Keller ET, Fu Z, Yeung K, Brennan M. (2004). *Cancer Lett* 207: 131–137.
- Khan J, Wei JS, Ringner M, Saal LH, Ladanyi M, Westermann F et al. (2001). *Nat Med* 7: 673–679.
- Kousidou OC, Roussidis AE, Theocharis AD, Karamanos NK. (2004). *Anticancer Res* 24: 4025–4030.
- Lisitsyn N, Lisitsyn N, Wigler M. (1993). *Science* 259: 946–951.
- Luo C, Shaw KT, Raghavan A, Aramburu J, Garcia-Cozar F, Perrino BA et al. (1996). *Proc Natl Acad Sci USA* 93: 8907–8912.

- Malliri A, Collard JG. (2003). *Curr Opin Cell Biol* 15: 583–589.
- Mantovani F, Gostissa M, Collavin L, Del Sal G. (2004). *Cell Cycle* 3: 905–911.
- Marenholz I, Heizmann CW, Fritz G. (2004). *Biochem Biophys Res Commun* 322: 1111–1122.
- Missiaen L, Robberecht W, van den Bosch L, Callewaert G, Parys JB, Wuytack F et al. (2000). *Cell Calcium* 28: 1–21.
- Moller LB, Bukrinsky JT, Molgaard A, Paulsen M, Lund C, Tumer Z et al. (2005). *Hum Mutat* 26: 84–93.
- Nagaraja GM, Kandpal RP. (2004). *Biochem Biophys Res Commun* 313: 654–665.
- Pase L, Voskoboinik I, Greenough M, Camakaris J. (2004). *Biochem J* 378: 1031–1037.
- Perou CM, Sorlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA et al. (2000). *Nature* 406: 747–752.
- Pratap J, Galindo M, Zaidi SK, Vradii D, Bhat BM, Robinson JA et al. (2003). *Cancer Res* 63: 5357–5362.
- Reiner A, Yekutieli D, Benjamini Y. (2003). *Bioinformatics* 19: 368–375.
- Ridley AJ. (2004). *Breast Cancer Res Treat* 84: 13–19.
- Rosenwald A, Wright G, Chan WC, Connors JM, Campo E, Fisher RI et al. (2002). *N Engl J Med* 346: 1937–1947.
- Ross DT, Scherf U, Eisen MB, Perou CM, Rees C, Spellman P et al. (2000). *Nat Genet* 24: 227–235.
- Shao Q, Wang H, McLachlan E, Veitch GI, Laird DW. (2005). *Cancer Res* 65: 2705–2711.
- Siegel RC. (1976). *J Biol Chem* 251: 5786–5792.
- Takayama Y, Kamimura Y, Okawa M, Muramatsu S, Sugino A, Araki H. (2003). *Genes Dev* 17: 1153–1165.
- van't Veer LJ, Dai H, van de Vijver MJ, He YD, Hart AA, Mao M et al. (2002). *Nature* 415: 530–536.
- Warner SL, Bearss DJ, Han H, Von Hoff DD. (2003). *Mol Cancer Ther* 2: 589–595.
- Westendorf JJ, Zaidi SK, Cascino JE, Kahler R, van Wijnen AJ, Lian JB et al. (2002). *Mol Cell Biol* 22: 7982–7992.
- Wiseman A. (2005). *Med Hypotheses* 65: 32–34.
- Yoshida S, Mears AJ, Friedman JS, Carter T, He S, Oh E et al. (2004). *Hum Mol Genet* 13: 1487–1503.
- Yu YX, Heller A, Liehr T, Smith CC, Aurelian L. (2001). *Int J Oncol* 18: 905–911.

Anuradha Waghray¹
Farhana Feroze¹
Megan S. Schober¹
Fayl Yao¹
Chris Wood²
Eric Puravs²
Melissa Krause²
Samir Hanash²
Yong Q. Chen^{1,3}

¹Department of Pathology,
Wayne State University,
Detroit, MI, USA

²Department of Pediatrics,
University of Michigan,
Ann Arbor, MI, USA

³Center for Molecular Medicine
and Genetics, Wayne State
University, Detroit, MI, USA

Identification of androgen-regulated genes in the prostate cancer cell line LNCaP by serial analysis of gene expression and proteomic analysis

A common therapy for nonorgan-confined prostate cancer involves androgen deprivation. To develop a better understanding of the effect of androgen on prostatic cells, we have analyzed gene expression changes induced by dihydrotestosterone (DHT) in the androgen responsive prostate cancer line LNCaP, at both RNA and protein levels. Changes at the RNA level induced by DHT were determined by means of serial analysis of gene expression (SAGE), and protein profiling was done by means of quantitative two-dimensional polyacrylamide gel electrophoresis. Among 123 371 transcripts analyzed, a total of 28844 distinct SAGE tags were identified representing 16 570 genes. Some 351 genes were significantly affected by DHT treatment at the RNA level ($p < 0.05$), of which 147 were induced and 204 repressed by androgen. In two independent experiments, the integrated intensity of 32 protein spots increased and 12 decreased at least two-fold in response to androgen, out of a total of 1031 protein spots analyzed. The change in intensity for most of the affected proteins identified could not be predicted based on the level of their corresponding RNA. Our study provides a global assessment of genes regulated by DHT and suggests a need for profiling at both RNA and protein levels for a comprehensive evaluation of patterns of gene expression.

Keywords: Androgen / Prostate / LNCaP / Gene expression / Serial analysis of gene expression
PRO 0137

1 Introduction

Androgens affect numerous aspects of prostate biology including development, growth, and maintenance. They also play a critical role in tumorigenesis and progression of prostate cancer. Androgen deprivation is an established treatment modality for prostate cancer. However, the disease eventually progresses into a hormone refractory cancer. Several mechanisms have been identified which may contribute to androgen independence: (1) Mutations in the androgen receptor (AR) lead to ligand-independent activation or promiscuity of the receptor [1, 2]. The ability of the receptor to activate or repress downstream genes can also be affected by mutation. AR mutation is associated with advanced phases of prostate cancer [3, 4]; (2) AR can be activated in a ligand-independent manner by specific growth factors and cytokines [5]; (3) AR gene amplification has been found to occur in

28–30% of tumors that recurred post androgen-ablation therapy [6, 7]. Wallen *et al.* [8] have recently shown that one-third of locally recurrent hormone refractory prostate cancer contain AR gene amplification; (4) Coactivator amplification and corepressor down-regulation have been shown to increase receptor transactivation [9, 10]. Regardless of which pathway(s) is taken by the tumor cells, androgen-regulated genes may ultimately be the key players in the development of hormone refractory cancer. As a step towards a better understanding of the effect of androgen on gene expression, we have undertaken a comprehensive assessment of gene expression changes induced by dihydrotestosterone (DHT) in the androgen responsive prostate cancer cell line LNCaP. An important feature of our study is the parallel assessment of expression changes at both RNA and protein levels.

2 Materials and methods

2.1 Prostate cell line and DHT treatment

LNCaP human prostate cancer cell line (American Type Culture Collection, Rockville, MD, USA) was cultured for 3 d in phenol-free RPMI 1640 supplemented with 5% charcoal-stripped fetal bovine serum (FBS) at 37°C. Half of the cultures were then treated with 10^{-9} M DHT for 24 h. Total RNA

Correspondence: Dr. Y. Q. Chen, Department of Pathology, Wayne State University, 540 E. Canfield, Detroit, MI 48201, USA
E-mail: yqchen@med.wayne.edu
Fax: +1-313-5770057

Abbreviations: DHT, dihydrotestosterone; G3PD, glyceraldehyde-3-phosphate dehydrogenase; PSA, prostate specific antigen; RT-PCR, reverse transcription-polymerase chain reaction; SAGE, serial analysis of gene expression

and protein were extracted from untreated and DHT treated cells using TRIzol (Invitrogen, Carlsbad, CA, USA) reagent for RNA and a solubilization cocktail for proteins, consisting of 9 M urea, β -mercaptoethanol and 1% NP-40. For kinetics experiments 1×10^6 cells were plated in phenol-free RPMI medium with 5% charcoal-stripped FBS. On day three, cells were treated with 5 mL of media containing 10^{-9} M DHT and total RNA was extracted at different time points from 0–96 h. To determine the direct/indirect effect of androgen on RNA levels, cells were plated as above and treated with 5 μ g/mL of cyclohexamide in the presence or absence of 10^{-9} M DHT for 24 h and total RNA was extracted.

2.2 Serial analysis of gene expression

Serial analysis of gene expression (SAGE) was performed as described previously [11] with the following modifications: ditags were PCR amplified using biotinylated primers and digested with *Nla*III enzyme [12]. Concatemers were heated for 15 min at 65°C and chilled on ice for 10 min before separation on an 8% polyacrylamide gel [13]. The concatemers were then cloned into the *Sph*I site of the pZero vector (Invitrogen). Concatenated tags were screened by PCR using M13 forward and M13 reverse primers. PCR products with inserts greater than 500 bp were isolated and sequenced with M13 forward primer on an automated 3700 DNA sequencer (Perkin-Elmer, Norwalk, CT, USA). For microSAGE, 1 μ g of total RNA per tube was used for cDNA synthesis in two tubes, with the mRNA Capture Kit (Boehringer Mannheim, Indianapolis, IN, USA). cDNAs were cleaved with *Nla*III, ligated to linkers and then digested with *Bsm*FI enzyme. The released tags were ligated, and processed for the rest of steps as with the standard SAGE protocol.

2.3 SAGE data analysis

SAGE tags were extracted using the SAGE software V 4.12 [11]. Tags were matched to the SAGE reliable map (release 10–26–2000) (<http://www.ncbi.nlm.nih.gov/SAGE/>). Due to the fact that some tags map to multiple genes and some genes have multiple tags, SAGE data were analyzed in two different ways: (1) exclusion method: tags that match to multiple genes were discarded. Only tags that match to a single gene were tabulated and composite counts analyzed for their significance; (2) inclusion method: tags that match to multiple genes were counted at 100% toward each gene. All tags were tabulated and composite counts analyzed for their statistical significance. Lists of differentially expressed genes ($p < 0.05$) obtained from the exclusion and inclusion methods were compared, and finally only genes that have a p value < 0.05 in both lists were considered statis-

tically significant. The total number of genes identified was estimated by $N_m + (N_{um} - 0.1 N_{um})/3.5$, where N_m is the number of genes matched to SAGE tags, N_{um} is the number of SAGE tags that do not match to known genes or ESTs, with 10% representing the estimated sequencing error per SAGE tag and 3.5, the average number of tags per gene in the SAGE reliable map (release 10–26–2000).

2.4 RT-PCR and real-time PCR quantification

For reverse transcription-PCR (RT-PCR), 1 μ g of total RNA was reverse transcribed into cDNA. One fortieth of cDNA was used for PCR reaction. Samples were collected at different cycles and separated on a 2% agarose gel with ethidium bromide. Image was captured and quantified using NucleoVision 760 Image Workstation (Nucleotech, CA, USA). Amplification curves were generated. Subsequently, RT-PCR was done at cycles within the log phase of amplification.

Real-time quantification was performed in the iCycler (Bio-Rad, Hercules, CA, USA). Briefly, one fortieth of cDNA was used in each reaction. Six reactions were carried out for each gene and three independent experiments were performed. PCR mix comprised of 1X PCR buffer, 1.5 mM $MgCl_2$, 0.1 mM dNTP, 200 nM primers (listed below), 0.05 U platinum Taq polymerase (Invitrogen) and 0.1x SYBR green (Molecular Probes, Eugene, OR, USA). PCR was carried out at 94°C for 2 min, and 40 cycles of 94°C for 30 s, 55°C for 30 s, and 72°C for 30 s. Primers used for real-time PCR quantification: PSA (prostate specific antigen) (Hs.171995, 5'-GGAAATGACCAGGCCAAGAC-3', 5'-CAACCCTG GAC C TCACACCTA-3'), SCMH1 (Hs.57475, 5'-GCCTTGACC ACATCACTCCAT-3', 5'-AGGCCTAGGGCTGCAAAAG-3'), and clusterin (Hs.75106, 5'-GCAGGAATACCGCAAAAA GC-3', 5'-GACTCAAGATGCCCCCGTAAG-3'). Standard samples (50, 25, 10, 5, 2.5, 1 and 0.5 μ L cDNA) and experimental samples were used in real-time quantification PCR. Each sample was run in quadruple reactions. Standard curve ($C_t = mX + B$) was obtained, where C_t is threshold cycle number, X is log quantity of target molecules, m is curve slope and B is Y-axis intercept value. Number of fold induction or repression for a given target molecule was calculated by Q_a/Q_b , where Q_a is the target quantity in experimental sample A and Q_b is the target quantity in experimental sample B. $C_{ta} = m \log Q_a + B$ or $Q_a = 10^{(C_{ta}-B)/m}$, therefore, $Q_a/Q_b = 10^{(C_{ta}-C_{tb})/m}$.

2.5 2-D PAGE

The procedure followed was as previously described [14]. Cells were solubilized in 200 μ L of lysis buffer containing 9.5 M urea (Bio-Rad), 2% NP-40, 2% carrier

ampholytes pH 4–8 (Gallard/Schlessinger, Carle Place, NY, USA), 2% β -mercaptoethanol and 10 mM PMSF. Aliquots containing approximately 5×10^6 cells, were applied onto isofocusing gels. IEF was conducted using pH 4 to 8 carrier ampholytes at 700 V for 16 h, followed by 1000 V for an additional 2 h. The 1-D gel was loaded onto the 2-D gel, after equilibration in 125 mM Tris, pH 6.8, 10% glycerol, 2% SDS, 1% DTT and bromophenol blue. For the second dimension separation, a gradient of 11–14% of acrylamide (Serva, Hauppauge, NY, USA) was used. Following 2-D PAGE, proteins were visualized by silver staining of the gels or transferred to an Immobilon-P PVDF membrane (Millipore, Bedford, MA).

2.6 Protein identification by mass spectrometry

The 2-D gels were silver-stained by successive incubations in 0.02% sodium thiosulfate for 2 min, 0.1% silver nitrate for 40 min and 0.014% formaldehyde plus 2% sodium carbonate. The proteins of interest were excised from the 2-D gels and destained for 5 min in 15 mM potassium ferricyanide and 50 mM sodium thiosulfate as described [15]. Following three washes with water, the gel pieces were dehydrated in 100% acetonitrile for 5 min and dried for 30 min in a vacuum centrifuge. Digestion was performed by addition of 100 ng of trypsin (Promega, Madison, WI, USA) in 200 mM ammonium bicarbonate or by addition of 100 ng of the endoprotease Glu-C (Promega) in 100 mM ammonium bicarbonate. The Lys-C digestion was performed with 500 ng of the endoproteinase Lys-C (Roche, Mannheim, Germany) in 100 mM Tris-HCl, pH 9. Following enzymatic digestion overnight at 37°C, the peptides were extracted twice with 50 μ L of 60% acetonitrile/1% TFA. After removal of acetonitrile by centrifugation in a vacuum centrifuge, the peptides were concentrated by using pipette tips C18 (Millipore).

Analyses were performed primarily using a PerSeptive Biosystem MALDI-TOF Voyager-DE mass spectrometer (Framingham, MA, USA), operated in delayed extraction mode. Peptide mixtures were analyzed using a saturated solution of cyano-4-hydroxycinnamic acid (CHA) (Sigma, St. Louis, MO, USA) in acetone containing 1% TPA (Sigma). Peptides were selected in the mass range of 800–4000 Da. Spectra were calibrated using calibration mixture 2 of the Sequazyme peptide mass standards kit (PerSeptive Biosystems). The search program MS-Fit, developed by the University of California at San Francisco (<http://prospector.ucsf.edu>), was used for searches in the NCBI database. Search parameters were as follows: maximum allowed peptide mass error of 400 ppm, consideration of one incomplete cleavage *per* peptide and

pH range between 4 and 8. MALDI-TOF mass spectrometry was also used for molecular weight determination as described [16]. In some cases, the amino acid sequence of some peptides of interest was determined by ESI MS analysis.

3 Results

3.1 SAGE analysis of the effect of androgen on gene expression

SAGE libraries were generated from LNCaP cells cultured in the presence or absence of DHT. A total of 123 371 tags were generated of which 62 878 were from the LNCaP cell line and 60 493 from the LNCaP cells treated with DHT for 24 h (Table 1). Sequence analysis identified a total of 28 844 distinct tags representing 16 570 genes, 11 243 from LNCaP and 12 203 from DHT treated cells. A total of 351 transcripts were differentially expressed at a significant level ($p < 0.05$). Eighty-seven percent of transcripts matched GenBank entries; 79% corresponded to known sequences and 8% to ESTs. RNA levels for 147 genes were increased and 204 genes were decreased after DHT treatment (Table 2). Therefore, at the RNA level more genes were repressed than activated by androgens. Of these androgen-regulated genes, 149 were changed \geq five-fold by DHT treatment (Table 3).

Table 1. Summary of SAGE analysis in LNCaP cells

| Sample | Total transcripts | Distinct tags | Number of genes |
|--------|-------------------|---------------|-----------------|
| – DHT | 62 878 | 17 050 | 11 243 |
| + DHT | 60 493 | 18 510 | 12 203 |
| Total | 123 371 | 28 844 | 16 570 |

– DHT: LNCaP cells without dihydrotestosterone; + DHT: LNCaP cells treated with 10^{-8} M dihydrotestosterone for 24 h

Table 2. DHT regulated genes in LNCaP cells

| Genes | LNCaP vs LNCaP + DHT (gene #) | |
|------------------------------|-------------------------------|------------|
| | $p < 0.05$ | $p < 0.01$ |
| Known genes | 277 | 147 |
| ESTs | 29 | 8 |
| No match | 45 | 13 |
| Total | 351 | 168 |
| Up-regulated by DHT (24 h) | 147 | 65 |
| Down-regulated by DHT (24 h) | 204 | 103 |

Table 3. List of candidate genes that are regulated by DHT (≥ 5 fold)

| UGD or Tag | CAP | DHT | Change | P value | Description |
|-------------|-----|-----|-----------|----------|---|
| 283 305 | 1 | 82 | Induced | 0 | immunoglobulin heavy constant alpha 1 |
| 183 752 | 0 | 42 | Induced | 0 | microseminoprotein, beta- |
| 140 | 1 | 44 | Induced | 0 | immunoglobulin heavy constant gamma 3 (Gm marker) |
| 75 415 | 18 | 95 | Induced | 0 | beta-2-microglobulin |
| 84 298 | 0 | 17 | Induced | 1.00E-05 | CD74 antigen (invariant polypeptide of major histocompatibility complex, class II antigen-associated) |
| 9615 | 0 | 16 | Induced | 1.00E-05 | myosin regulatory light chain 2, smooth muscle isoform |
| 77 443 | 0 | 16 | Induced | 1.00E-05 | actin, gamma 2, smooth muscle, enteric |
| 1119 | 0 | 15 | Induced | 2.33E-05 | nuclear receptor subfamily 4, group A, member 1 |
| 84 753 | 23 | 2 | Repressed | 3.00E-05 | KIAA0246 protein |
| 75 777 | 0 | 14 | Induced | 3.66E-05 | transgelin |
| | 0 | 14 | Induced | 3.66E-05 | immunoglobulin J polypeptide, linker protein for immunoglobulin alpha and mu polypeptides |
| 76 325 | | | | | |
| 1852 | 1 | 17 | Induced | 4.33E-05 | acid phosphatase, prostate |
| 78 465 | 0 | 13 | Induced | 9.00E-05 | v-jun avian sarcoma virus 17 oncogene homolog |
| 78 344 | 1 | 16 | Induced | 1.20E-04 | myosin, heavy polypeptide 11, smooth muscle |
| 83 006 | 19 | 2 | Repressed | 1.50E-04 | CGI-139 protein |
| 263 812 | 15 | 1 | Repressed | 3.19E-04 | nuclear distribution gene C (A. nidulans) homolog |
| TACGGGGATA | 0 | 11 | Induced | 4.20E-04 | Novel |
| GCCTGGGTGG | 11 | 0 | Repressed | 6.63E-04 | Novel |
| GA CTGACACT | 16 | 2 | Repressed | 8.40E-04 | Novel |
| 284 296 | 0 | 10 | Induced | 8.60E-04 | Homo sapiens SURF-4 mRNA, complete cds |
| 75 105 | 13 | 1 | Repressed | 1.19E-03 | emopamil-binding protein (sterol isomerase) |
| 128 075 | 10 | 0 | Repressed | 1.30E-03 | ESTs |
| 154 162 | 15 | 2 | Repressed | 1.43E-03 | ADP-ribosylation factor-like 2 |
| 6895 | 1 | 12 | Induced | 1.49E-03 | actin related protein 2/3 complex, subunit 3 (21 kD) |
| 143 240 | 17 | 3 | Repressed | 1.63E-03 | ESTs |
| 211 582 | 0 | 9 | Induced | 1.66E-03 | myosin, light polypeptide kinase |
| 180 266 | 0 | 9 | Induced | 1.66E-03 | tropomyosin 2 (beta) |
| 93 002 | 14 | 2 | Repressed | 2.53E-03 | ubiquitin carrier protein E2-C |
| 17 883 | 14 | 2 | Repressed | 2.53E-03 | protein phosphatase 1G (formerly 2C), magnesium-dependent, gamma isoform |
| 126 023 | 14 | 2 | Repressed | 2.53E-03 | ESTs, Highly similar to NTC1_HUMAN NEUROGENIC LOCUS NOTCH PROTEIN HOMOLOG 1 PRECURSOR |
| 69 469 | 2 | 13 | Induced | 2.92E-03 | dendritic cell protein |
| 77 899 | 2 | 13 | Induced | 2.92E-03 | tropomyosin 1 (alpha) |
| 285 501 | 2 | 13 | Induced | 3.07E-03 | Human rearranged immunoglobulin lambda light chain mRNA |
| 119 209 | 0 | 8 | Induced | 3.07E-03 | insulin-like growth factor binding protein 7 |
| 173 043 | 0 | 8 | Induced | 3.07E-03 | metastasis-associated 1-like 1 |
| 75 866 | 0 | 8 | Induced | 3.07E-03 | dimethylarginine dimethylaminohydrolase 1 |
| 171 695 | 0 | 8 | Induced | 3.07E-03 | dual specificity phosphatase 1 |
| TACGGGGATT | 0 | 8 | Induced | 3.34E-03 | Novel |
| 172 791 | 13 | 2 | Repressed | 4.19E-03 | ubiquitously-expressed transcript |
| 13 561 | 13 | 2 | Repressed | 4.19E-03 | ESTs, Weakly similar to dJ37E16.5 [H. sapiens] |
| 98 260 | 8 | 0 | Repressed | 4.34E-03 | ESTs |
| CATAAGACTT | 8 | 0 | Repressed | 4.64E-03 | Novel |
| TACGGGGACA | 2 | 12 | Induced | 5.76E-03 | Novel |
| 180 034 | 12 | 2 | Repressed | 6.91E-03 | cleavage stimulation factor, 3' pre-RNA, subunit 3, 77kD |
| 5753 | 12 | 2 | Repressed | 6.91E-03 | inositol(myo)-1(4)-monophosphatase 2 |
| 278 941 | 10 | 1 | Repressed | 6.94E-03 | PRO0628 protein |
| 19 500 | 0 | 7 | Induced | 7.25E-03 | nuclear localization signal deleted in velocardiofacial syndrome |

Table 3. Continued

| UGD or Tag | CAP | DHT | Change | P value | Description |
|------------|-----|-----|-----------|----------|---|
| 9006 | 0 | 7 | Induced | 7.25E-03 | VAMP (vesicle-associated membrane protein)-associated protein A (33kD) |
| 26471 | 0 | 7 | Induced | 7.25E-03 | <i>Homo sapiens</i> clone HQ0692 |
| AACTGCTGGC | 12 | 2 | Repressed | 7.63E-03 | Novel |
| GCTGACCGTC | 7 | 0 | Repressed | 8.59E-03 | Novel |
| CCCCCCTGTC | 7 | 0 | Repressed | 8.59E-03 | Novel |
| 243901 | 2 | 11 | Induced | 8.82E-03 | <i>Homo sapiens</i> cDNA FLJ20738 fis, clone HEP08257 |
| 119000 | 1 | 9 | Induced | 9.12E-03 | actinin, alpha 1 |
| 78596 | 1 | 9 | Induced | 9.12E-03 | proteasome (prosome, macropain) subunit, beta type, 5 |
| 171955 | 7 | 0 | Repressed | 9.61E-03 | trophinin associated protein (tastin) |
| 180545 | 7 | 0 | Repressed | 9.61E-03 | <i>Homo sapiens</i> mRNA for hypothetical protein (TR2/D15 gene) |
| 77719 | 7 | 0 | Repressed | 9.61E-03 | gamma-glutamyl carboxylase |
| 4877 | 7 | 0 | Repressed | 9.61E-03 | CGI-51 protein |
| 22795 | 7 | 0 | Repressed | 9.61E-03 | ESTs |
| 79335 | 7 | 0 | Repressed | 9.61E-03 | <i>Homo sapiens</i> SWI/SNF-related, matrix-associated, actin-dependent regulator of chromatin D1 (SMARCD1) mRNA |
| 119177 | 9 | 1 | Repressed | 1.26E-02 | ADP-ribosylating factor 3 |
| CGGGAGCACC | 9 | 1 | Repressed | 1.29E-02 | Novel |
| 182217 | 0 | 6 | Induced | 1.32E-02 | succinate-CoA ligase, ADP-forming, beta subunit |
| 75106 | 0 | 6 | Induced | 1.32E-02 | clusterin (complement lysis inhibitor, SP-40, 40, sulfated glycoprotein 2, testosterone-repressed prostate message 2) |
| 256311 | 0 | 6 | Induced | 1.32E-02 | granin-like neuroendocrine peptide precursor |
| 103180 | 0 | 6 | Induced | 1.32E-02 | DC2 protein |
| TACGGGGATG | 1 | 8 | Induced | 1.70E-02 | Novel |
| AAACAAATCA | 2 | 10 | Induced | 1.70E-02 | Novel |
| 136644 | 2 | 10 | Induced | 1.71E-02 | CS box-containing WD protein |
| 8036 | 2 | 10 | Induced | 1.71E-02 | glioblastoma overexpressed |
| 74284 | 2 | 10 | Induced | 1.71E-02 | ESTs, Weakly similar to Similar to <i>S. cerevisiae</i> hypothetical protein L3111 |
| ATGGCTGATC | 6 | 0 | Repressed | 1.75E-02 | Novel |
| ATCACTGCCC | 6 | 0 | Repressed | 1.75E-02 | Novel |
| ACATCATCAG | 6 | 0 | Repressed | 1.75E-02 | Novel |
| CCAGTCCAAG | 6 | 0 | Repressed | 1.75E-02 | Novel |
| 54842 | 1 | 8 | Induced | 1.78E-02 | ESTs |
| 1526 | 1 | 8 | Induced | 1.78E-02 | ATPase, Ca++ transporting, cardiac muscle, slow twitch 2 |
| 7911 | 1 | 8 | Induced | 1.78E-02 | KIAA0323 protein |
| 227400 | 1 | 8 | Induced | 1.78E-02 | mitogen-activated protein kinase kinase kinase 3 |
| 77269 | 1 | 8 | Induced | 1.78E-02 | guanine nucleotide binding protein (G protein), alpha inhibiting activity polypeptide 2 |
| 7943 | 6 | 0 | Repressed | 1.81E-02 | RPB5-mediating protein |
| 278544 | 6 | 0 | Repressed | 1.81E-02 | acetyl-Coenzyme A acetyltransferase 2 (acetoacetyl Coenzyme A thiolase) |
| 23111 | 6 | 0 | Repressed | 1.81E-02 | phenylalanine-tRNA synthetase-like |
| 31442 | 6 | 0 | Repressed | 1.81E-02 | RecQ protein-like 4 |
| 38628 | 6 | 0 | Repressed | 1.81E-02 | hypothetical protein |
| 77422 | 6 | 0 | Repressed | 1.81E-02 | proteolipid protein 2 (colonic epithelium-enriched) |
| 211973 | 6 | 0 | Repressed | 1.81E-02 | homolog of Yeast RRP4 (ribosomal RNA processing 4), 3'-5'-exoribonuclease |
| 171075 | 6 | 0 | Repressed | 1.81E-02 | replication factor C (activator 1) 5 (36.5kD) |
| 89781 | 6 | 0 | Repressed | 1.81E-02 | upstream binding transcription factor, RNA polymerase I |
| 154149 | 6 | 0 | Repressed | 1.81E-02 | apurinic/apyrimidinic endonuclease(APEX nuclease)-like 2 protein |

Table 3. Continued

| UGD or Tag | CAP | DHT | Change | P value | Description |
|------------|-----|-----|-----------|----------|---|
| 279772 | 8 | 1 | Repressed | 2.28E-02 | brain specific protein |
| 205091 | 8 | 1 | Repressed | 2.28E-02 | ESTs, Weakly similar to WW domain binding protein 11 (<i>M. musculus</i>) |
| 75658 | 8 | 1 | Repressed | 2.28E-02 | phosphorylase, glycogen; brain |
| GGGCAGCTGT | 8 | 1 | Repressed | 2.37E-02 | Novel |
| 105440 | 0 | 5 | Induced | 2.43E-02 | hepatocyte nuclear factor 3, alpha |
| 118244 | 0 | 5 | Induced | 2.43E-02 | protein phosphatase 2, regulatory subunit B (B56), delta isoform |
| 82389 | 0 | 5 | Induced | 2.43E-02 | CGI-118 protein |
| 31638 | 0 | 5 | Induced | 2.43E-02 | restin (Reed-Steinberg cell-expressed intermediate filament-associated protein) |
| 12013 | 0 | 5 | Induced | 2.43E-02 | ATP-binding cassette, sub-family E (OABP), member 1 |
| 12797 | 0 | 5 | Induced | 2.43E-02 | DEAD/H (Asp-Glu-Ala-Asp/His) box polypeptide 16 |
| 7736 | 0 | 5 | Induced | 2.43E-02 | hypothetical protein |
| 274479 | 0 | 5 | Induced | 2.43E-02 | NME7 |
| 153138 | 0 | 5 | Induced | 2.43E-02 | origin recognition complex, subunit 5 (yeast homolog)-like |
| 16034 | 0 | 5 | Induced | 2.43E-02 | ESTs |
| 270072 | 0 | 5 | Induced | 2.43E-02 | ESTs |
| 187035 | 0 | 5 | Induced | 2.43E-02 | ESTs |
| 140452 | 0 | 5 | Induced | 2.43E-02 | cargo selection protein (mannose 6 phosphate receptor binding protein) |
| 182265 | 0 | 5 | Induced | 2.43E-02 | keratin 19 |
| 19762 | 0 | 5 | Induced | 2.43E-02 | ESTs, Weakly similar to unknown (<i>D. melanogaster</i>) |
| 317 | 0 | 5 | Induced | 2.43E-02 | topoisomerase (DNA) |
| 6236 | 0 | 5 | Induced | 2.43E-02 | ESTs |
| GCTGGAGCCT | 5 | 0 | Repressed | 3.03E-02 | Novel |
| CCAGTGCTCA | 5 | 0 | Repressed | 3.03E-02 | Novel |
| ACCCTACATA | 5 | 0 | Repressed | 3.03E-02 | Novel |
| GGGGAAATCT | 5 | 0 | Repressed | 3.03E-02 | Novel |
| ACTGGTACTG | 5 | 0 | Repressed | 3.03E-02 | Novel |
| GCTCCGGTGT | 5 | 0 | Repressed | 3.03E-02 | Novel |
| ACAGTGGTGA | 5 | 0 | Repressed | 3.03E-02 | Novel |
| 7869 | 1 | 7 | Induced | 3.04E-02 | lysophosphatidic acid acyltransferase-delta |
| 12101 | 1 | 7 | Induced | 3.04E-02 | hypothetical protein |
| 266940 | 1 | 7 | Induced | 3.04E-02 | t-complex-associated-testis-expressed 1-like 1 |
| 6196 | 1 | 7 | Induced | 3.04E-02 | integrin-linked kinase |
| 366 | 1 | 7 | Induced | 3.04E-02 | 6-pyruvoyltetrahydropterin synthase |
| 173611 | 1 | 7 | Induced | 3.04E-02 | NADH dehydrogenase (ubiquinone) Fe-S protein 2 (49kD) (NADH-coenzyme Q reductase) |
| 102469 | 1 | 7 | Induced | 3.04E-02 | putative nuclear protein |
| 78825 | 1 | 7 | Induced | 3.04E-02 | matrin 3 |
| 284465 | 1 | 7 | Induced | 3.04E-02 | ESTs |
| 30738 | 1 | 7 | Induced | 3.04E-02 | hypothetical protein FLJ10407 |
| 78687 | 1 | 7 | Induced | 3.04E-02 | neutral sphingomyelinase (N-SMase) activation associated factor |
| 92381 | 1 | 7 | Induced | 3.04E-02 | nudix (nucleoside diphosphate linked moiety X)-type motif 4 |
| 242039 | 5 | 0 | Repressed | 3.26E-02 | EST |
| 4766 | 5 | 0 | Repressed | 3.26E-02 | DKFZP586O0120 protein |
| 283109 | 5 | 0 | Repressed | 3.26E-02 | hypothetical protein DKFZp762L1710 |
| 192853 | 5 | 0 | Repressed | 3.26E-02 | ubiquitin-conjugating enzyme E2G 2 (homologous to yeast UBC7) |
| 153678 | 5 | 0 | Repressed | 3.26E-02 | reproduction 8 |

Table 3. Continued

| UGD or Tag | CAP | DHT | Change | P value | Description |
|-------------|-----|-----|-----------|----------|--|
| 251317 | 5 | 0 | Repressed | 3.26E-02 | EST |
| 279623 | 5 | 0 | Repressed | 3.26E-02 | selenoprotein X |
| 150319 | 5 | 0 | Repressed | 3.26E-02 | ESTs |
| 102456 | 5 | 0 | Repressed | 3.26E-02 | survival of motor protein interacting protein 1 |
| 284250 | 5 | 0 | Repressed | 3.26E-02 | AD-003 protein |
| 251871 | 5 | 0 | Repressed | 3.26E-02 | CTP synthase |
| 270480 | 7 | 1 | Repressed | 3.92E-02 | ESTs, Weakly similar to ALU5_HUMAN ALU SUBFAMILY SC SEQUENCE |
| 26655 | 7 | 1 | Repressed | 3.92E-02 | glucose-6-phosphatase, transport (glucose-6-phosphate) protein 1 |
| 8118 | 7 | 1 | Repressed | 3.92E-02 | leukotriene A4 hydrolase |
| CTCCGCCGGC | 7 | 1 | Repressed | 4.47E-02 | Novel |
| AGGAAATGCT | 7 | 1 | Repressed | 4.47E-02 | Novel |
| GCTGACCGAGG | 7 | 1 | Repressed | 4.47E-02 | Novel |
| CCCATAGTCC | 7 | 1 | Repressed | 4.47E-02 | Novel |

CAP: LNCaP cells without dihydrotestosterone

DHT: LNCaP cells treated with 10^{-8} dihydrotestosterone for 24 h

UGD: unigene ID

Numbers in CAP and DHT columns are the number of tags observed in respective samples

3.2 Confirmation of SAGE data

To confirm the differential expression pattern at the RNA level, semiquantitative RT-PCR assay was performed on a group of selected genes. One μ g of total RNA was reverse transcribed into cDNA with oligo dT. cDNA was amplified by PCR for various cycles. PCR products were separated on agarose gels and quantified by densitometry. PCR amplification curves were plotted for each gene, and data within the logarithmic phase of amplification were used for quantification (Fig. 1A, B and C). The reproducibility of gene regulation by androgen was confirmed in three independent experiments by monitoring the induction of PSA in the presence or absence of cyclohexamide (Fig. 1B). The kinetics of gene induction or repression was determined for PSA (Hs.171995), clusterin (Hs.75106) and SCMH1 (Hs.57475) genes. Typical examples of kinetics are shown in Figure 1C. PSA was induced at 4–6 h, peaked between 6–20 h, and gradually declined after 20 h post-treatment of DHT. Clusterin was induced within 0.5–1 h and gradually declined after 6–12 h. SCMH1 was repressed 2–4 h post-treatment. Expression of PSA, clusterin, and SCMH1 was quantified by real-time PCR (Fig. 1D).

3.3 Changes in protein expression induced by DHT

Protein lysates of LNCaP cells cultured in the presence of DHT for 72 h and cells cultured in parallel in the absence of DHT were subjected to 2-D PAGE (Fig. 2). Following sil-

ver staining, gels were digitized prior to computer-based matching and quantitative analysis, as previously described [17]. Of a total of 1031 protein spots matched between 2-D patterns of DHT treated and untreated cells, a set of 32 protein spots increased in intensity by at least two-fold in two independent experiments. Likewise, a set of 12 protein spots decreased in intensity by at least two-fold in these two independent experiments. A two-fold change in integrated intensity by silver staining represents on average a three-fold change in amount of protein, based on prior quantitative studies [17].

The 44 protein spots that changed in intensity as a result of DHT treatment were excised from the gels, digested with trypsin, and subsequently analyzed by MALDI-TOF MS. The resulting spectra were used to identify the proteins using the MS-Fit search program. Of the 44 spots excised from the gels, 29 were identified without ambiguity and consisted of 21 up-regulated and eight down-regulated proteins (Table 4). The identified proteins represented a heterogeneous group that included cytoskeletal proteins (e.g. tropomyosin, α tubulin), metabolic enzymes (e.g. adenine phosphoribosyl transferase, β 1,4 galactosyl transferase, galactocerebrosidase), and the products of previously described androgen responsive genes (e.g. SRY, selenium binding protein) [18, 19]. Specific antibodies confirmed the identification based on MS for all proteins analyzed by Western blotting, which included α tubulin, myosin light chain isoforms, nucleoside diphosphate kinase A, glyceraldehyde 3-phosphate dehydrogenase (G3PD), and tropomyosin (data not shown).

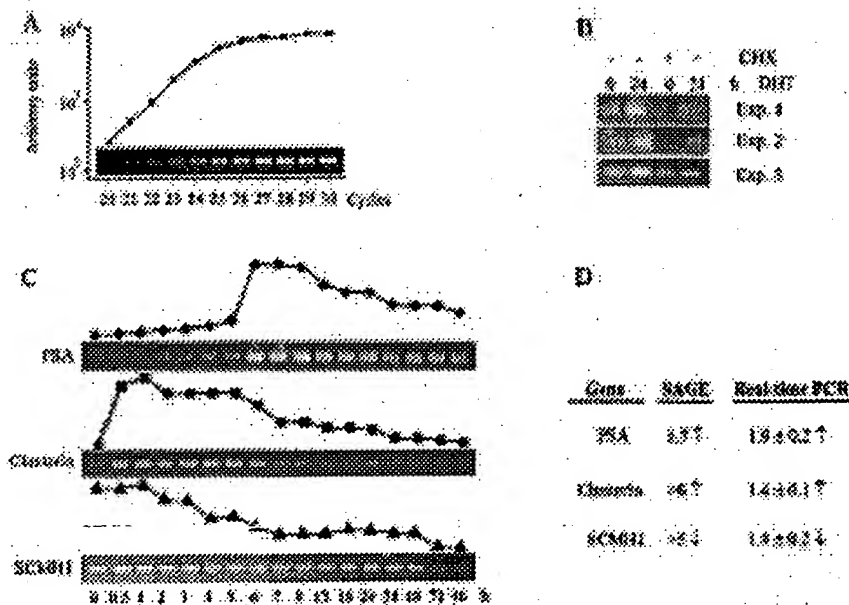


Figure 1. Kinetics and quantification of androgen-regulated genes. Total RNA from untreated and DHT treated LNCaP cells was reverse transcribed. cDNA was PCR amplified for various cycles. PCR amplification curves were plotted for each gene. A cycle number within the logarithmic phase of amplification was selected for semi-quantitative analysis. cDNA was also used for real-time quantitative PCR. (A) An example of RT-PCR amplification of PSA at cycles between 20–30. (B) RT-PCR of PSA in three independently treated LNCaP samples. h: hours; 0: no DHT; 24: treated with 10^{-8} M DHT for 24 h; -:

without cycloheximide; +: with 5 μ g/mL of cycloheximide for 24 h. (C) An example of three independent kinetics experiments for PSA, clusterin and SCM1. (D) Comparison of SAGE results with that of real-time quantitative PCR for PSA, clusterin and SCM1. ↑: induction; ↓: repression.

Corresponding SAGE data were available for most of the proteins affected by DHT treatment that were identified. It was therefore of interest to determine if the changes observed at the protein levels were matched by concordant changes at the mRNA level. Remarkably, for most of the proteins identified, there was no appreciable concordant change at the RNA level (Table 4).

4 Discussion

We have compared the gene expression profile of LNCaP, an androgen responsive prostate cancer cell line, with or without androgen (DHT) treatment. Approximately 350 of 16570 expressed genes detected at the RNA level were affected by dihydrotestosterone at the $p < 0.05$ level. The DHT responsive genes included known genes, ESTs, and novel genes. As expected, we saw an induction of genes that are well known to be regulated by androgens. For instance, we found a 1.7-fold induction in kallikrein 3/PSA, a 7.6-fold induction in prostatic kallikrein 2, and a 15.7-fold induction in prostatic acid phosphatase by DHT in LNCaP cells. We also saw a five-fold induction in NKX3.1/NKX3A and three-fold in fatty acid synthase; two previously identified androgen-regulated genes [20, 21]. Interestingly β -microseminoprotein, reported to be reduced or lost in prostate tumor [22], was up-regulated more than 40-fold in

LNCaP cells by DHT. More significantly, our data indicate that genes involved in a variety of tumor cell functions such as growth, apoptosis, and metastasis, are directly or indirectly regulated by androgens. In addition, it is noteworthy that both β -actin and G3PD, the two most frequently used loading controls, were up-regulated approximately two-fold by DHT ($p < 0.01$). Thus, these two genes may not be appropriate controls in some experiments.

Serial analysis of androgen-regulated gene expression provides us with a list of candidate genes. However, many factors such as the dose of DHT and the time of cell exposure to DHT and other unknown experimental variations will affect the level of gene expression. Therefore, it is important to confirm the SAGE results with alternative methods. We performed semiquantitative RT-PCR (Fig. 1B) on approximately 20 genes and real-time quantitative PCR on 10 genes in at least three independently DHT treated LNCaP samples. We noticed experimental variations in every gene determined. Some genes such as PSA (Fig. 1B) have smaller variations and some have larger variations. In addition, the fold of alteration in expression identified by SAGE is different for some genes compared to that identified by real-time quantitative PCR. This is probably due to the technical limitation of SAGE. Serial analysis of gene expression is highly quantitative for genes whose tags are detected at large numbers.

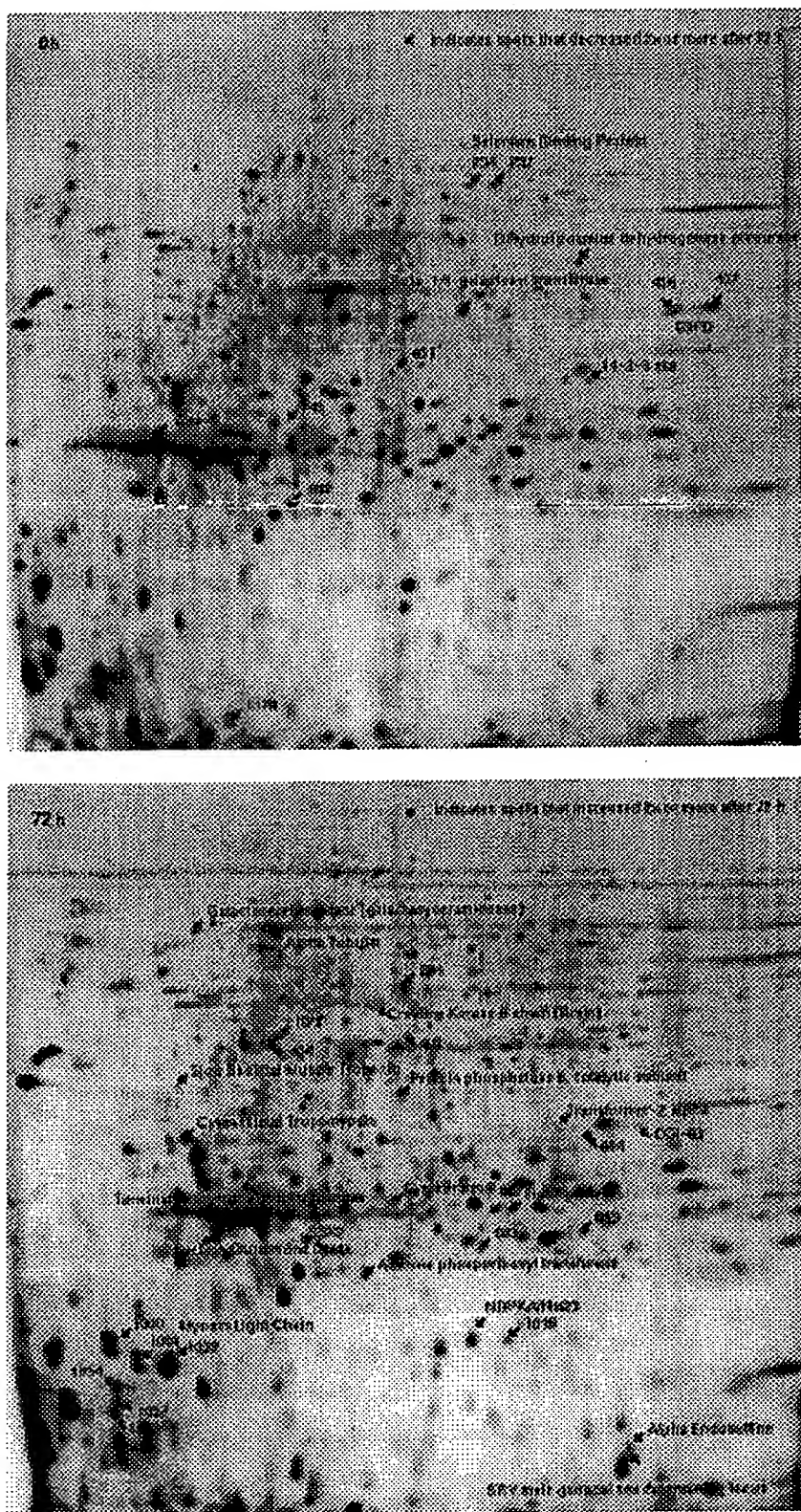


Figure 2. 2-D PAGE patterns of untreated (A) and LNCaP cells treated with DHT for 72 h (B). Identified proteins that were reduced in levels following DHT treatments are shown in (A) and induced proteins are shown in (B). Numbered spots represent either proteins that were changed in level that have not been identified or isoforms of identified proteins.

Table 4. List of proteins that are altered by DHT (≥ 2 -fold)

| Protein name or sport number | Unigene ID | Protein | | Change in protein | mRNA | | Change in mRNA |
|------------------------------------|------------|---------|-------|-------------------|------|------|----------------|
| | | 0 h | 72 h | | 0 h | 24 h | |
| Adenine phosphoribosyl transferase | 28 914 | 0.448 | 1.084 | ↑ | 36 | 18 | ↓ |
| Alpha Endosulfine | 111 680 | 0.066 | 0.679 | ↑ | 2 | 5 | ↑ |
| Alpha Tubulin | 278 242 | 0.924 | 1.932 | ↑ | 48 | 38 | ↓ |
| Nucleophosmin, B23 | 173 205 | 0.256 | 0.522 | ↑ | 16 | 5 | ↓ |
| CGI-83 | 118 554 | 0.063 | 0.291 | ↑ | 2 | 3 | ↑ |
| Creatine Kinase B chain (brain) | 173 724 | 0.508 | 1.142 | ↑ | 59 | 32 | ↓ |
| Cytokeratin 8 | 242 463 | 0.134 | 0.393 | ↑ | 35 | 40 | ↑ |
| Cytoskeletal tropomyosin, NTRK1 | 85 844 | 0.414 | 1.174 | ↑ | 14 | 18 | ↑ |
| Galactocerebrosidase | 273 | 0.095 | 0.405 | ↑ | 3 | 3 | = |
| Lactoyl Glutathione Lyase | 75 207 | 0 | 1.256 | ↑ | 27 | 22 | ↓ |
| Myosin Light Chain (1000) | 77 385 | 0.724 | 6.102 | ↑ | 118 | 88 | ↓ |
| Myosin Light Chain (1001) | 77 385 | 0.6 | 3.212 | ↑ | 118 | 88 | ↓ |
| Myosin Light Chain (1032) | 77 385 | 2.573 | 5.694 | ↑ | 118 | 88 | ↓ |
| Nm23 | 118 638 | 0.583 | 1.978 | ↑ | 35 | 25 | ↓ |
| Protein phosphatase 6, catalytic | 279 563 | 0.287 | 0.737 | ↑ | 2 | 2 | = |
| Protein | 7 016 | 0 | 0.448 | ↑ | 2 | 5 | ↑ |
| Slow Skeletal Muscle Troponin | 84 673 | 0.067 | 0.195 | ↑ | 1 | 0 | ↓ |
| SRY male gonadal sex determining | 1 992 | 0 | 0.735 | ↑ | ND | ND | |
| Synovial sarcoma, X breakpoint 2 | 289 105 | 0.225 | 0.593 | ↑ | ND | ND | |
| Terminal deoxynucleotidyl | 272 537 | 0.392 | 0.995 | ↑ | ND | ND | |
| Transformer-2 alpha | 119 523 | 0.074 | 0.201 | ↑ | 7 | 13 | ↑ |
| 284 | | 0.173 | 0.461 | ↑ | | | |
| 402 | | 0 | 0.33 | ↑ | | | |
| 456 | | 0.154 | 0.314 | ↑ | | | |
| 664 | | 0.297 | 0.629 | ↑ | | | |
| 807 | | 0 | 0.377 | ↑ | | | |
| 863 | | 0.246 | 0.665 | ↑ | | | |
| 903 | | 0.271 | 0.632 | ↑ | | | |
| 1018 | | 0 | 0.725 | ↑ | | | |
| 1054 | | 0.47 | 1.394 | ↑ | | | |
| 1057 | | 0.451 | 0.959 | ↑ | | | |
| 1082 | | 0.609 | 1.738 | ↑ | | | |
| 14-3-3 eta | 75 544 | 0.232 | 0.114 | ↓ | 1 | 5 | ↑ |
| Beta-1,4-galactosyl transferase | 158 540 | 1.306 | 0.707 | ↓ | ND | ND | |
| Cox11 | 241 515 | 1.643 | 1.453 | ↓ | 3 | 0 | ↓ |
| Dihydrolipoamide dehydrogenase | 74 635 | 0.844 | 0.132 | ↓ | 9 | 8 | ↓ |
| G3PD (456) | 169 476 | 1.191 | 0.279 | ↓ | 75 | 137 | ↑ |
| G3PD (457) | 169 479 | 2.051 | 0.336 | ↓ | 75 | 137 | ↑ |
| Selenium Binding Protein (234) | 7 833 | 0.411 | 0.201 | ↓ | 26 | 31 | ↑ |
| Selenium Binding Protein (237) | 7 833 | 0.574 | 0.22 | ↓ | 26 | 31 | ↑ |
| 651 | | 0.31 | 0.115 | ↓ | | | |
| 745 | | 0.21 | 0.067 | ↓ | | | |
| 892 | | 0.674 | 0.454 | ↓ | | | |
| 1178 | | 0.933 | 0.305 | ↓ | | | |

Numbers in protein are the intensity of protein spots

Numbers in mRNA columns are the number of SAGE tags detected

ND: not detected

↑: increase; ↓: decrease; =: equal

When the tag number detected for a given gene is close to zero, the quantitative nature of SAGE is compromised. For instance, in approximately 60 000 transcripts of each sample, the PSA tag was detected 42 times in LNCaP and 70 times in LNCaP treated with DHT. However, the tag for clusterin was detected 0 times and 6 times, and the tag for SCMH1 5 times and 0 times in the corresponding samples. Therefore, the SAGE data of PSA is more reliable than that of clusterin and SCMH1. Indeed, SAGE data of PSA is virtually identical to that of real-time quantitative PCR, whereas the SAGE data of clusterin and SCMH1 differ from that of quantitative PCR (Fig. 1D).

PSA is probably the best-known androgen-regulated gene. We were surprised to see only 1.7-fold induction by DHT. However, kinetics experiments indicate that PSA is induced at 4–6 h, peaked between 6–20 h, and gradually declined 20 h post-treatment of DHT. Since the SAGE experiment was done in samples treated for 24 h with DHT, PSA mRNA level was likely to have declined from its peak level. Indeed, we could detect 5–10 fold induction of PSA at 6–8 h post-treatment. Clusterin was reported as an androgen-repressed gene in the rat prostate [23]. Recent evidence indicates that clusterin may not be directly androgen-repressed, but regulated by apoptotic stimuli [24]. Our results suggest that clusterin was induced within 0.5–1 h, gradually declined after 6–12 h, and after 24 h reduced to a lower level than that of untreated cells (Fig. 1C).

Another important question is how many of the androgen-regulated genes identified are directly induced or repressed by androgens. In order to address this question, detailed analyses must be done for each gene. As the first step, we will divide the androgen-regulated genes identified in this study into two groups: the cyclohexamide-sensitive group whose induction or reduction of expression by DHT is blocked by the protein synthesis inhibitor and the cyclohexamide-insensitive group whose induction or reduction is not affected by cyclohexamide. We consider that genes in the latter group are directly regulated by DHT. Preliminary results suggest that approximately 20% of the 20 genes studied by RT-PCR are cyclohexamide-insensitive. Experiments are under way to determine the cyclohexamide sensitivity of all of the 149 genes listed in Table 3. Further characterization of androgen-regulated genes may provide some clues on the transition from hormone sensitive to hormone refractory prostate cancer.

A relatively small set of genes could be analyzed at the protein level, largely due to the limited sensitivity of 2-D PAGE. Nevertheless, a substantial number of detected proteins (44 of 1031 proteins analyzed, 4.3%) were affected by DHT treatment. Some of the proteins in this

subset that were identified included the products of genes that were previously shown to be affected by androgens, namely selenium binding protein, brain specific creatine kinase, SRY protein, B23 and G3PD. Using a subtractive approach, the human selenium-binding protein gene was shown to be differentially expressed in LNCaP and reversibly down-regulated by exogenous androgen in a concentration-dependent manner, in concordance with our findings at the protein level [19]. An increase in levels of the brain specific creatine kinase B chain in response to androgen has also been described [25]. Likewise the SRY gene has been found to be responsive to androgen stimulation in LNCaP cells [18]. Also, a number of glycolytic enzymes including G3PD which were affected at the protein level have been found to be responsive to androgen stimulation [26]. Androgenic regulation of the amount and phosphorylation of the protein B23, included in our list of affected proteins, has been previously described and found to be related to the early changes associated with androgen mediated growth of the prostate [27].

Corresponding SAGE data was available for most of the proteins affected by DHT treatment that were identified. Interestingly, for most of the proteins identified, there was no appreciable concordant change at the RNA level. There are several potential explanations for this lack of concordance. A particular gene may be represented by more than one protein isoform in 2-D gels. For example, in the identified group, three proteins (myosin light chain, G3PD and selenium binding protein) were represented by more than one isoform. Thus, a source of discordance between RNA and protein data may be that the protein change is limited to a particular isoform of a protein and not to overall protein products of a particular gene. Nevertheless, a change in a particular isoform is informative and biologically meaningful and may not be predictable from RNA data. A lack of concordance between RNA and protein data may also reflect either translational control, post-translational modifications, or changes in protein turnover due to DHT treatment. Yet another explanation for a lack of concordance could be a lag time for changes at the RNA level to be reflected in a protein change. It follows from the above considerations that monitoring gene expression at both RNA and protein levels may provide complementary information that could not be ascertained by solely measuring RNA or protein.

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5 References

- [1] Taplin, M. E., Bubley, G. J., Shuster, T. D., Frantz, M. E., et al., *N. Engl. J. Med.* 1995, 332, 1393-1398.
- [2] Elo, J. P., Kvist, L., Lelonen, K., Isomaa, V., et al., *J. Clin. Endocrinol. Metab.* 1995, 80, 3494-3500.
- [3] Taplin, M. E., Bubley, G. J., Ko, Y. J., Small, E. J., et al., *Cancer Res.* 1999, 59, 2511-2515.
- [4] Marcelli, M., Ittmann, M., Mariani, S., Sutherland, R., et al., *Cancer Res.* 2000, 60, 944-949.
- [5] Weigel, N. L., Zhang, Y., *J. Mol. Med.* 1998, 76, 469-479.
- [6] Visakorpi, T., Hyythén, E., Koivisto, P., Tanner, M., et al., *Nat. Genet.* 1995, 9, 401-406.
- [7] Koivisto, P., Kononen, J., Palmberg, C., Tammela, T., et al., *Cancer Res.* 1997, 57, 314-319.
- [8] Wallén, M. J., Linja, M., Kaartinen, K., Schleutker, J., Visakorpi, T., *J. Pathol.* 1999, 189, 559-563.
- [9] Bautista, S., Valles, H., Walker, R. L., Anzick, S., et al., *Clin. Cancer Res.* 1998, 4, 2925-2929.
- [10] Smith, C. L., Nawaz, Z., O'Malley, B. W., *Mol. Endocrinol.* 1997, 6, 657-666.
- [11] Velculescu, V. E., Zhang, L., Vogelstein, B., Kinzler, K. W., *Science* 1995, 270, 484-487.
- [12] Powell, J., *Nucleic Acid Res.* 1998, 26, 3445-3446.
- [13] Kenzelmann, M., Muhlemann, K., *Nucleic Acid Res.* 1999, 27, 917-918.
- [14] Strahler, J. R., Juick, R., Hanash, S. M., In: Creighton, T. E. [Ed.] *Protein Structure: A Practical Approach*, IRL Press, England 1989, pp. 65-92.
- [15] Gharahdaghi, F., Weinberg, C. R., Meagher, D. A., Imal, B. S., Mische, S. M., *Electrophoresis* 1999, 20, 601-605.
- [16] Liang, X., Bai, J., Liu, Y.-H., Lubman, D. M., *Anal. Chem.* 1996, 68, 1012-1018.
- [17] Hanash, S. M., In: Hames, B. D., [Ed.], *Gel Electrophoresis of Proteins*, IRL Press, Oxford, 1998, pp. 189-211.
- [18] Lau, Y. F., Zhang, J., *Mol. Carcinog.* 2000, 27, 308-321.
- [19] Yang, M., Sytkowski, A. J., *Cancer Res.* 1998, 58, 3150-3153.
- [20] Prescott, J. L., Blok, L., Tindall, D. J., *Prostate* 1998, 35, 71-80.
- [21] Swinnen, J. V., Uhlir, W., Heyns, W., Verhoeven, G., *Proc. Natl. Acad. Sci. USA* 1997, 94, 12975-12980.
- [22] Chan, P. S., Chan, L. W., Xuan, J. W., Choi, H. L., Chan, F. L., *Prostate* 1999, 41, 99-109.
- [23] Léger, J. G., Montpetit, M. L., Tenniswood, M., *Biochem. Biophys. Res. Commun.* 1987, 147, 196-203.
- [24] Miyake, H., Nelson, C., Rennie, P. S., Gleave, M. E., *Cancer Res.* 2000, 60, 170-176.
- [25] Somjen, D., Lundgren, S., Kaye, A. M., *J. Steroid Biochem. Mol. Biol.* 1997, 62, 89-96.
- [26] Tawfic, S., Goueli, S. A., Olson, M. O., Ahmed, K., *Cell. Mol. Biol. Res.* 1993, 39, 43-51.
- [27] Brooks, D. E., *Biochem. J.* 1976, 156, 527-537.

REGULAR ARTICLE

Web-based data warehouse on gene expression in human colorectal cancer

Emil Sagynaliev¹, Ralf Steinert², Gerd Nestler², Hans Lippert², Manfred Knoch¹ and Marc-André Reymond²

¹ Department of Surgery, Johanniter Krankenhaus, Stendal, Germany

² Department of Surgery, University of Magdeburg, Magdeburg, Germany

Based on biomedical literature databases, we tried a first step for constructing a gene expression "data warehouse" specific to human colorectal cancer (CRC). Results of genome-wide transcriptomic research were available from 12 studies, using various technologies, namely, SAGE, cDNA and oligonucleotide arrays, and adaptor-tagged amplification. Three studies analyzed CRC cell lines and nine studies of human samples. The total number of patients was 144. Out of 982 up- or down-regulated genes, 863 (88%) were found to be differentially expressed in a single study, 88 in two studies, 22 in three studies, 7 in four studies, and only 2 genes in six studies. Eight large-scale proteomics studies were published in CRC, using 2-D-, SDS- or free-flow electrophoresis, involving only 11 patients. Out of 408 differentially expressed proteins, 339 (83%) were found to be differentially expressed only in a single study, 16 in three studies, 10 in four studies, 3 in five, and 1 in eight studies. Confirmation at proteome level of results obtained with large-scale transcriptomics studies was possible in 25%. This proportion was higher (67%) for reproducing proteome results using transcriptomics technologies. Obviously, reproducibility and overlapping between published gene expression results at proteome and transcriptome level are low in human CRC. Thus, the development of standardized processes for collecting samples, storing, retrieving, and querying gene expression data obtained with different technologies is of central importance in translational research.

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1 Introduction

Translational research in human colorectal cancer (CRC) is applying a large spectrum of molecular biology, cellular biology, and advanced validation tools. In particular, genome-wide techniques are now applied to decipher modifications

in gene expression. In recent years, transcriptomics and proteomics tools have been broadly applied in CRC. The hope is that the new data obtained will now allow a classification of disease on molecular basis, deep insights into the pathophysiology of CRC, prognostic statements, and finally a systematic search for diagnostic and therapeutic targets.

Because of the complexity of the biological system under investigation, the most significant contribution of translational research in CRC is expected to derive not from the analysis of single experiments but from libraries of experiments. In other words, the results obtained so far by translational research tools in different clinical and experimental settings need to be compared, contrasted, and if possible synthesized. Thus, based on the biomedical literature databases, we tried a first step for constructing such "data warehouse" specific to human CRC.

Correspondence: Professor Marc-André Reymond, University of Magdeburg, Chairman and Head, Department of Surgery, Evangelic Hospital Bielefeld, Burgsteig 13, D-33617 Bielefeld, Germany

E-mail: marc.reymond@evkb.de

Fax: +49-521-144-4551

Abbreviations: cDNA, complementary DNA; CRC, colorectal cancer

The specific aims of the present study were to screen transcriptomics and proteomics studies published in CRC, and to elaborate a kind of "meta-analysis of gene expression" in CRC. Our hypothesis was that we would be able to determine a common set of genes and gene products that are up- or down-regulated in human CRC by overlapping results obtained by different authors.

2 Materials and methods

This is a literature survey of gene expression data published in human CRC. A Medline search of reports published in English from 1990 to 2004 using the terms "colon cancer" and "gene expression" with the limit "human" was performed and yielded 1979 articles. All abstracts were reviewed and a related article search was performed on appropriate abstracts. Articles were selected by a consensus of two reviewers (E.S., M.R.) that satisfied these predetermined criteria: sample origin (human and/or cell lines) and preparation detailed, technology for gene expression studies defined (transcriptomics and/or proteomics studies), and quantitative results on overexpression or underexpression available. Studies concerning single genes or arbitrarily selected genes were discarded. Results obtained in animal models were not considered.

Data were entered into an Excel working sheet (Microsoft, Seattle, USA). Since gene expression data were not always obtained with quantitative, but in most cases semi-quantitative gene expression analysis technologies, no expression values or ratios were entered into the database. Only genes reported as being over- or underexpressed were entered. No threshold was defined so that some genes defined as differentially expressed might have shown only

marginal differences. Only descriptive statistics are provided, which were obtained with the in-built tools of the Excel software.

Various methods were used for large-scale translational research in CRC, including transcriptomics and proteomics technologies. Results of genome-wide transcriptomic research are available from 12 studies, using various technologies, namely, SAGE (five studies), complementary DNA (cDNA) arrays (five studies), and oligonucleotide array (one study), as well as adaptor-tagged amplification (one study). Three studies analyzed CRC cell lines and nine other studies analyzed human samples. The total number of patients profiled in all studies was 144. Out of the nine studies with human samples, only two studies with a total of only 22 patients were performed using purified epithelial cells (one of them using cDNA arrays, the other one using SAGE technology). An overview of these studies is provided in Table 1.

Eight large-scale proteomics studies were published in CRC using various technologies, namely, 2-D PAGE (six studies), SDS-PAGE (one study), and free-flow electrophoresis (one study). In total, all proteome studies in CRC involved only 11 patients. An overview of these studies is provided in Table 2.

3 Results

One thousand two-hundred and forty genes have been reported to be dysregulated (up- and/or down-regulated) in human CRC, representing about 5% of the 20 000–25 000 human genes.

* The complete dataset can be consulted:
<http://www.chirurgiebethel.de>

Table 1. Large-scale or genome-wide transcriptomics studies in human CRC, using cell lines or human tissues. All studies together involved only 144 patients

| Author | Year | Sample | Preparation | Method | Number of patients |
|------------------------------------|------|---|---------------------------|-----------------------|--------------------|
| Zhang <i>et al.</i> [18] | 1997 | Normal mucosa and primary tumors | Whole tissue | SAGE | 2 |
| SAGE-NET [19] | 1997 | Primary tumors and cell lines | Cell lines | SAGE | – |
| Zhang <i>et al.</i> [18] | 1997 | Cell lines | Cell lines | SAGE | – |
| Parle-McDermott <i>et al.</i> [20] | 2000 | Cell lines | Cell lines | SAGE | – |
| Yanagawa <i>et al.</i> [21] | 2001 | Paired normal and cancer | Whole tissue | cDNA | 20 |
| Takemasa <i>et al.</i> [22] | 2001 | Paired normal and primary tumors | Whole tissue | cDNA | 20 |
| Nottermann <i>et al.</i> [23] | 2001 | Paired normal mucosa and primary tumors, or adenoma | Whole tissue | Oligonucleotide array | 22 |
| Buckhaults <i>et al.</i> [24] | 2001 | Normal mucosa, adenoma and primary tumors | Purified epithelial cells | SAGE | 6 |
| Birkenkamp <i>et al.</i> [25] | 2002 | Paired normal and primary tumors | Whole tissue | cDNA | 27 |
| Lin <i>et al.</i> [26] | 2002 | Paired normal mucosa and primary tumors, or adenoma | Purified epithelial cells | cDNA | 16 |
| Muro <i>et al.</i> [27] | 2003 | Normal and cancer | Whole tissue | Adaptor-tagged | 11 |
| Williams <i>et al.</i> [28] | 2003 | Paired normal mucosa and primary tumors, normal and adenoma | Whole tissue | cDNA | 20 |

Table 2. Large-scale or proteomics studies in human CRC, using cell lines or human tissues. All studies together involved only nine patients

| Author | Publication year | Sample | Method | Number of patients |
|-----------------------------|------------------|---------------------------|---------------------------|--------------------|
| Reymond <i>et al.</i> [9] | 1997 | Purified epithelial cells | 2-D PAGE | 1 |
| Simpson <i>et al.</i> [29] | 2000 | Cell lines | SDS-PAGE | – |
| Lawrie <i>et al.</i> [30] | 2001 | Purified epithelial cells | 2-D PAGE | 4 |
| Simpson <i>et al.</i> [31] | 2001 | Cell lines | Free-flow electrophoresis | – |
| Medjahed <i>et al.</i> [32] | 2003 | <i>In silico</i> | 2-D PAGE | – |
| Demalte <i>et al.</i> [33] | 2003 | Cell lines | 2-D PAGE | – |
| Stierum <i>et al.</i> [34] | 2003 | Cell lines | 2-D PAGE | – |
| Friedman <i>et al.</i> [35] | 2004 | Human whole tissue | 2-D PAGE | 6 |

The vast majority of dysregulated genes in human CRC was found using transcriptomics tools: a total of 982 genes was found to be differentially expressed in at least one of these 12 transcriptomics studies. Out of these 982 genes, 863 (88%) were found to be differentially expressed only in a single study, in other words these results have not been reproduced so far. The other findings could be reproduced in two or more transcriptomics studies: 88 genes were found to be differentially expressed in two studies, 22 in three studies, 7 in four studies, no gene in five studies, and 2 genes in six studies. The most cited genes are listed in Tables 3 (up-regulation) and 4 (down-regulation).

A total of 408 proteins were found to be differentially expressed in human CRC in at least one study. Out of these 408 molecules, 339 (83%) were mentioned in a single study, in other words these results could not be reproduced. Differential expression of the remaining 70 proteins were mentioned with following frequency: 40 were mentioned in two proteomics studies, 16 in three studies, 10 in four studies, 3 in five studies, and a single protein in eight studies. The most cited proteins are listed in Table 5. It has to be noted that only a single study [1] provided differential display protein expression data obtained in the human patient, using whole tissue biopsy.

It is also difficult to reproduce transcriptomics results with proteomics tools. Out of 982 genes found to be differentially expressed in human CRC by genome-wide transcriptomics technologies (Table 6a), only 177 (18%) have been confirmed using proteome technologies. When the genes reported to be differentially expressed in three and more transcriptomics studies ($n = 31$) are compared with those reported to be differentially expressed in three and more proteomics studies ($n = 30$), only two genes (actin: ACTB_HUMAN and creatin-kinase: KCRB_HUMAN) can be matched. Thus, the probability of reproducing a gene expression result obtained at transcriptome level is low when using proteomics technologies.

In contrast, there is a better reproducibility when proteomics results are verified using transcriptomics tools (Table 6b). In fact, when the subset of 30 proteins that are consistently (in three studies or more) reported to be dysregulated in CRC is compared with transcriptomics results, 20

genes can be matched, representing about two-thirds. Thus, the probability of being able to reproduce a proteomics result using transcriptomics tools is more than three times higher than the opposite way (67 vs. 18%). However, it has to be noted that in 16% of cases, results remain unclear or even contradictory.

Both in the transcriptomics and proteomics studies, many genes and factors were found to be differentially regulated that obviously do not play a causal role in CRC carcinogenesis.

4 Discussion

This study was aimed at screening transcriptomics and proteomics studies published in CRC, and to elaborate a kind of meta-analysis of gene expression in CRC in order to generate a data warehouse that would be useful in translational research. A significant number of translational research studies have been published in CRC, starting in 1997 with SAGE technology, followed by an increasing number of array-based transcriptomics studies since 2001, and more recently by several proteomics studies. This has created a significant amount of data which we were then able to compile.

Our starting hypothesis was that we would be able to determine a common set of genes and gene products that are up- or down-regulated in human CRC by comparing results obtained by different authors. This endeavor was not very successful; obviously, overlapping between published gene expression results both at proteome and transcriptome level is low in human CRC. In fact, more than 80% of results could not be reproduced. Several explanations can be provided to explain this lack of reproducibility.

First, the number of patients included in the studies is low (altogether 144 patients in transcriptomics, 11 in proteomics studies); some high impact publications having been conducted on samples of only two patients [18]. This is a problem because interindividual genetic variability is high in human CRC [2]. Thus, it is allowed to hypothesize that some results attributed to gene dysregulation might be caused by genetic diversity rather than by cancer-specific traits.

Table 3. Results of transcriptomics research in human CRC. Most cited up-regulated genes out of 12 genome-wide or large-scale studies

| | | | | TRANSCRIPTOMICS | | | | | | | | | | | |
|------------------|------------|------------|---|-----------------|----------------|------|------|------|------|------|------|------|------|------|-----------------------|
| Accession Number | Swiss-Prot | Entry Name | Name | cDNA | Adaptor-tagged | SAGE | cDNA | cDNA | cDNA | SAGE | SAGE | SAGE | SAGE | cDNA | Oligonucleotide Array |
| AF041260 | O75363 | IFM2_HUMAN | Interferon-inducible protein 1-Bd | 1 | | 1 | | 1 | | | 1 | | | | |
| AA232508 | Q9Y397 | SAHH_HUMAN | S-Adenosylhomocysteine hydrolase AHcy) | | | 1 | 1 | | | | | 1 | 1 | | |
| AA007218 | Q96QY8 | BGH3_HUMAN | Transforming growth factor, beta-induced, 66 kDa | 1 | | 1 | 1 | | | | | 1 | | | |
| G01119 | | TCPD_HUMAN | Chaperonin containing TCP-1, subunit 4 (delta) | | | | | | 1 | | | | 1 | 1 | |
| X75821 | Q04984 | CA11_HUMAN | COL1A1 | | | | | 1 | 1 | | 1 | | | | |
| U22055 | Q96AG0 | TF3A_HUMAN | General transcription factor IIIA | | | | 1 | | 1 | | | | | 1 | |
| U14631 | P80365 | GRO_HUMAN | GRO1 oncogene (melanoma growth stimulating activity, alpha) | | | | 1 | | | | | | 1 | 1 | |
| M92439 | P42704 | MDP1_HUMAN | MPDP4, MDP7 microsomal dipeptidase | | | 1 | 1 | | | | 1 | | | | |
| AF029082 | P31947 | MPI2_HUMAN | M-phase inducer phosphatase 2 | 1 | 1 | 1 | | | | | | | | | |
| M86400 | P29312 | PROC_HUMAN | Pyrroline 5-carboxylate reductase mRNA | 1 | 1 | 1 | | | | | | | | | |
| L76465 | P15428 | SPRC_HUMAN | Secreted protein, acidic, cysteine-rich (osteonectin) | 1 | 1 | 1 | | | | | | | | | |
| X57352 | Q01628 | Q96AG0 | 100 kDa coactivator mRNA | 1 | 1 | | | | | | | | | | |
| X03205 | P62888 | RL30_HUMAN | 50S Ribosomal protein L30 | 1 | 1 | | | | | | | | | | |
| M95787 | Q01995 | RL23_HUMAN | 60s ribosomal protein L23 | 1 | 1 | | | | | | | | | | |
| Z25821 | P42126 | HS9B_HUMAN | 90-kDa HSP | 1 | 1 | | | | | | | | | | |
| U96132 | Q99714 | RLA2_HUMAN | Acidic ribosomal phosphoprotein P2 mRNA | 1 | 1 | | | | | | | | | | |
| AB028893 | P04643 | CA14_HUMAN | Alpha-1 chain of collagen IV | 1 | 1 | | | | | | | | | | |
| X79239 | Q02546 | APA1_HUMAN | Apolipoprotein A-I | 1 | 1 | | | | | | | | | | |
| M13934 | P06366 | BMP4_HUMAN | Bone morphogenetic protein-2B (BMP-2B) | 1 | 1 | | | | | | | | | | |
| M60854 | P17008 | CSE1_HUMAN | CAS chromosome segregation gene homologue | 1 | 1 | | | | | | | | | | |
| M13932 | P08708 | CD81_HUMAN | CD81 antigen | 1 | 1 | | | | | | | | | | |
| X69150 | P25232 | TCPZ_HUMAN | Chaperonin containing TCP1, subunit 6A(zeta1) | 1 | 1 | | | | | | | | | | |
| U06498 | P17075 | CA21_HUMAN | Collagen alpha-2 type 1 | 1 | 1 | | | | | | | | | | |
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Table 3. Continued

| | | | | TRANSCRIPTOMICS | | | | | | | | | | |
|------------------|------------|------------|---|--------------------------|----------------------|----------------------------|----------------------------|--------------------------|---------------------|--------------------------|-----------------------|--------------------------|-----------------------|--------------------------|
| | | | | cDNA | Adaptor-tagged | SAGE | cDNA | cDNA | cDNA | SAGE | SAGE | SAGE | SAGE | cDNA |
| Accession Number | Swiss-Prot | Entry Name | Name | Yanagawa et al 2001 [21] | Muro et al 2003 [27] | Buckhaults et al 2001 [24] | Birkenkamp et al 2002 [25] | Takemasa et al 2001 [22] | Lin et al 2002 [26] | Paré-McDermott 2000 [20] | Zhang et al 1997 [18] | SAGE-NET et al 1997 [19] | Zhang et al 1997 [18] | Williams et al 2003 [28] |
| L04483 | P35265 | CO2_HUMAN | Complement component C2 | 1 | 1 | | | | | | | | | |
| M64716 | P25111 | R10A_HUMAN | Csa-19mRNA | 1 | 1 | | | | | | | | | |
| X77770 | P02383 | ROA1_HUMAN | DNA binding protein UPI, liver mRNA fragment | 1 | 1 | | | | | | | | | |
| W52460 | P14798 | TP2A_HUMAN | DNA topoisomerase II (top2) | 1 | 1 | | | | | | | | | |
| X55715 | P23396 | IF39_HUMAN | Eukaryotic translation initiation factor(EIF3)mRNA | 1 | 1 | | | | | | | | | |
| M77234 | P49241 | IF2B_HUMAN | Eukaryotic translation initiation factor 2, subunit 2 | 1 | 1 | | | | | | | | | |
| M58458 | P12750 | GGH_HUMAN | Gamma-glutamyl hydrolase | 1 | 1 | | | | | | | | | |
| D16992 | P10660 | G3P2_HUMAN | Glyceraldehyde 3 phosphate dehydrogenase | 1 | 1 | | | | | | | | | |
| M77233 | P23821 | SYG_HUMAN | Glycyl-tRNA synthetase | 1 | 1 | | | | | | | | | |
| F16294 | P09058 | RS8_HUMAN | H19 RNA | 1 | 1 | | | | | | | | | |
| X61156 | P08865 | HS9A_HUMAN | Heat shock Protein HSP 90-Alpha | 1 | 1 | | | | | | | | | |
| L36055 | Q13541 | HMG1_HUMAN | Hmg1 mRNA for high mobility group protein I | 1 | 1 | | | | | | | | | |
| AA316619 | P04645 | RS23_HUMAN | Human homolog of yeast ribosomal protein S23 (D14530) | 1 | 1 | | | | | | | | | |
| L14599 | Q15233 | IMD2_HUMAN | IMP (inosine monophosphate) dehydrogenase 2 | 1 | 1 | | | | | | | | | |
| X79234 | P39026 | IFM1_HUMAN | Interferon induced transmembrane protein 1(9–27) | 1 | 1 | | | | | | | | | |
| L06505 | P30050 | LDHB_HUMAN | Lactate dehydrogenase B(LDH-B) | 1 | 1 | | | | | | | | | |
| X64707 | P26373 | RS2_HUMAN | LLRep 3 | 1 | 1 | | | | | | | | | |
| X56932 | P40429 | 4F2_HUMAN | Lymphocyte activation antigen 4F2 large subunit | 1 | 1 | | | | | | | | | |
| L25899 | P39030 | NPL1_HUMAN | Nucleosome assembly protein 1-like 1 | 1 | 1 | | | | | | | | | |
| X63527 | P14118 | GSHH_HUMAN | Phospholipid hydroperoxide glutathione peroxidase | 1 | 1 | | | | | | | | | |
| X89401 | P46778 | GDFF_HUMAN | Prostate differentiation factor | 1 | 1 | | | | | | | | | |
| H59771 | P23131 | RL6_HUMAN | Ribosomal protein L 6 | 1 | 1 | | | | | | | | | |
| M94314 | P38663 | RL18_HUMAN | Ribosomal protein L18(RPL18) mRNA | 1 | 1 | | | | | | | | | |
| L19527 | P08526 | RL1X_HUMAN | Ribosomal protein L18a mRNA, complete cds | 1 | 1 | | | | | | | | | |
| U14968 | P46776 | RL7_HUMAN | Ribosomal protein L7 | 1 | 1 | | | | | | | | | |
| U14969 | P46779 | RL8_HUMAN | Ribosomal protein L8 | 1 | 1 | | | | | | | | | |
| L38941 | P49207 | RS5_HUMAN | Ribosomal protein S5 | 1 | 1 | | | | | | | | | |
| U12465 | P42766 | RS19_HUMAN | S 19 ribosomal protein | 1 | 1 | | | | | | | | | |
| F19234 | P18077 | MYC_HUMAN | V-myc myelocytomatosis viral oncogene homolog (avain) | 1 | 1 | | | | | | | | | |
| X66699 | P12751 | Q99497 | EST(PARK7; DJ1) | 1 | 1 | | | | | | | | | |

Table 4. Results of transcriptomics research in human CRC. Most cited down-regulated genes out of 12 genome-wide or large-scale studies

| | | TRANSCRIPTOMICS | | | | | | | | | | | | | |
|------------|--|-----------------|--------------------------|----------------|-------------------|--------------------------|-------------------------|-------------------------|------------------------------------|------------|---------------------|-------------------------------|-----------|---------------------------------------|------------------------------------|
| Entry Name | Name | cDNA | Paired normal and cancer | Adaptor-tagged | Normal and cancer | N.muc,adenoma and pr.tum | Paired n. and pr.tumors | Paired n. and pr.tumors | Paired n.muc and pr.tum or adenoma | Cell lines | Norm.muc and Pr.tum | Primary tumors and cell lines | Cell line | Paired n.muc and pr.tum,n and adenoma | Paired n.muc and pr.tum or adenoma |
| CAH2_HUMAN | Carbonic anhydrase II | 1 | | | | 1 | | 1 | | | | 1 | | 1 | 1 |
| CEA1_HUMAN | EST (BILIARY GLYCOPROTEIN) | | | | | 1 | 1 | 1 | | | | 1 | | 1 | 1 |
| BENE_HUMAN | BENE | 1 | | | | | | 1 | | | 1 | | | | 1 |
| CAH1_HUMAN | Carbonic anhydrase I | 1 | | | 1 | | | | | | | 1 | | | 1 |
| GUAU_HUMAN | GCAP-II guanylate cyclase activator 2B; Uroguanylin; UGN | 1 | | | | | 1 | | | | | 1 | | | 1 |
| FABL_HUMAN | L-FABP liver fatty acid-binding protein 1 | 1 | | | | | 1 | 1 | | | | 1 | | | |
| ACDS_HUMAN | Acyl-CoenzymeA dehydrogenase C-2 to C-3 short chain | | | | | | 1 | | | | | | | 1 | 1 |
| ADHG_HUMAN | ADH gama 2 subunit (aa-1-375) | | | | | | 1 | 1 | | | | | | | 1 |
| T4S3_HUMAN | Co-029; transmembrane 4 superfamily member 3 | | | | | 1 | 1 | | | | | 1 | | | |
| DRA_HUMAN | Colon mucosa-associated (DRA) | | | | | | 1 | | | | | 1 | | | 1 |
| KCRB_HUMAN | Creatine kinase | 1 | | | | | 1 | | | | | 1 | | | |
| GUAN_HUMAN | GUCA1B guanylate cyclase activator 1B/guanylin | | | | | | 1 | | | | | 1 | | | 1 |
| O95784 | IgG Fc-binding protein | 1 | | | | | 1 | 1 | | | | | | | |
| ITMC_HUMAN | Integral membrane protein 2C | | | | | | | 1 | 1 | | | 1 | | | |
| MK03_HUMAN | MAPK3 | | | | | | 1 | | | | | 1 | | 1 | |
| FXY3_HUMAN | MAT8 protein | | | | | | 1 | 1 | | | | 1 | | | |
| ACTB_HUMAN | mRNA fragment encoding cytoplasmic actin | 1 | | | | | | | | 1 | 1 | | | | |
| SELP_HUMAN | Selenoprotein P | | | | | | 1 | 1 | | | | | 1 | | |
| EST1_HUMAN | Liver carboxylesterase 1 [Precursor] | | | | | | | | | | | 1 | 1 | | 1 |
| CFAD_HUMAN | Adipsin, complement factor DCDA (EST) | | | | | | 1 | | | | | | | | 1 |
| ATPB_HUMAN | ATP5B ATP synthase | | | | | | 1 | 1 | | | | | | | |
| CAHC_HUMAN | CA 12 | | | | | | 1 | 1 | | | | | | | |
| CANS_HUMAN | Calcium dependent protease (small subunit) | | | | | | 1 | | | | 1 | | | | |
| CALM_HUMAN | Calmodulin-I (CALM1) mRNA, 3' UTR, partial sequence | | | | | | | | | | 1 | | | 1 | |
| CAH4_HUMAN | Carbonic anhydrase IV | | | | | | 1 | | 1 | | | | | | 1 |

Table 4. Continued

| | | TRANSCRIPTOMICS | | | | | | | | | | |
|------------|--|-----------------|----------------|------|------|------|------|------|------|------|------|------|
| Entry Name | Name | cDNA | Adaptor-tagged | SAGE | cDNA | cDNA | cDNA | SAGE | SAGE | SAGE | SAGE | cDNA |
| MT1E_HUMAN | cDNA similar to gb: M10942_cds1 human | | | | | | | | | | | |
| O00748 | CES 2 carboxylesterase 2 | | | | 1 | 1 | | | | 1 | | 1 |
| CMGA_HUMAN | CgA | | | | 1 | | | | | | | 1 |
| CLUS_HUMAN | clusterin | | | | | | | | | | | 1 |
| K1CT_HUMAN | Cytokeratin 20 | | | | | | | | | 1 | | 1 |
| KDGA_HUMAN | Diacylglycerol kinase | | | | 1 | | | | | | | 1 |
| DTD_HUMAN | DTD sulfate transporter | | | | 1 | 1 | | | | | | |
| PLA8_HUMAN | EST 122594 5_ | | | | | | | | | 1 | | 1 |
| ATPA_HUMAN | F1-ATPase alpha subunit | | | | 1 | 1 | | | | | | |
| LEG3_HUMAN | Galectin-3 (Galactose-specific lectin 3) (MAC-2 antigen) | | | | 1 | 1 | | | | | | |
| ABP_HUMAN | HP-DAO1 (diamine oxidase) | | | | | | | | | | 1 | 1 |
| K1CS_HUMAN | Keratin 19 | | | | | | 1 | | | | 1 | |
| K2C8_HUMAN | Keratin, type II cytoskeletal 8 (cytokeratin 8) (K 8) (CK 8) | | | | | | | 1 | | | 1 | |
| DHB2_HUMAN | L11708 Estradiol-17 beta- dehydrogenase 2 | | | | | | 1 | | | | | 1 |
| MT1H_HUMAN | Metallothionein 1H | | | | | | | | | | | 1 |
| MUC2_HUMAN | Mucin 2, intestinal/tracheal | | | | | | | 1 | 1 | | | |
| MEPA_HUMAN | PPH alpha gene | | | | | | | | | | 1 | 1 |
| PA2A_HUMAN | RASF-A PLA2 gene | | | | | 1 | | | | | | 1 |
| MT1F_HUMAN | RNA helicase-related protein | | | | | | | | | | | 1 |
| SBP1_HUMAN | SBP selenium-binding protein | | | | | | 1 | 1 | | | | |
| MT1L_HUMAN | Serine threonine kinase 39 (STE20/SPS1 homolog,yeast) | | | | | | 1 | | | | | 1 |
| MYL6_HUMAN | Myosin light polypeptide 6 | | | | | | | | | 1 | | 1 |
| TETN_HUMAN | TNA tetranectin | | | | | | 1 | | | | | 1 |
| VIPR_HUMAN | VIPR1 vasoactive intestinal polypeptide receptor 1 | | | | | | 1 | | | | | 1 |
| PLA8_HUMAN | Placenta-specific gene 8 protein | | | | | | | | | | 1 | 1 |
| Q9NXM9 | Hypothetical protein FLJ20151 | | | | | | 1 | | | | | 1 |
| T4S1_HUMAN | Transmembrane 4 superfamily member 1 | | | | 1 | | | | | | 1 | |
| ALU2_HUMAN | Alu subfamily SB sequence contamination warning entry | | | 1 | | 1 | | | | | | |
| S116_HUMAN | S100 calcium-binding protein A16 | | | | | | | | | 1 | 1 | |
| C14A_HUMAN | Dual specificity protein phosphatase CDC14A | | | | | | | | | | 1 | 1 |

Second, results have been obtained using heterogeneous samples in particular cell lines, whole tissue biopsies, and epithelial cells purified from surgical specimens. Results obtained in cell lines do not allow accurate comparison between normal and cancer cells, and the presence/absence of proteins of interest has to be confirmed in biopsies. For example, when 2-D PAGE protein patterns of normal human

colonic crypts were compared with the CRC cell line LIM 1863, the proteins spots from normal crypts matched only 75–80% of the cell line spots and the relative expression levels of a large number of proteins differed [3]. When clinical biopsies are examined, results of phenotypic comparisons depend on the type of samples examined (e.g., heterogeneous whole tissue biopsies with inflammatory cells,

Table 5. Results of proteomics research in human CRC. Most cited up-regulated (orange), down-regulated (green), or mentioned (yellow) proteins out of 12 large-scale proteomics studies

| Accession Number | Swiss-Prot | Entry Name | Name | SDS-PAGE | Free-flow electrophoresis | 2-D PAGE | 2-D PAGE | 2-D PAGE | 2-D PAGE | 2-D PAGE | 2-D PAGE | 2-D PAGE |
|------------------|------------|-------------|--|-------------------------|---------------------------|-------------------------|------------------------|--------------------------|------------------------|-------------------------|-----------------------|----------|
| | | | | Simpson et al 2000 [29] | Simpson et al 2001 [31] | Dematte et al 2003 [33] | Reymond et al 1999 [9] | Medjahed et al 2003 [32] | Lawrie et al 2001 [30] | Friedman et al 2004[35] | Sterum et al 2003[34] | |
| AF041260 | D75363 | ACTB_HUMAN | mRNA fragment encoding cytoplasmic actin | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| AA232508 | Q9Y397 | ACTG_HUMAN | Actin, Cytoplasmic 2 (Gamma-Actin) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| AA007218 | Q96QY8 | GTP_HUMAN | Glutathione S-transferase M3 (brain) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| X77584 | G01119 | THIO_HUMAN | Thioredoxin (ATL-derived factor) (ADF) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| X75821 | Q04984 | ANX3_HUMAN | Annexin III (Lipocortin III) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| U22055 | Q96AG0 | ANX4_HUMAN | Annexin IV (Lipocortin IV) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| U14631 | P80365 | K1CR_HUMAN | Keratin, type I cytoskeletal 18 (cytokeratin 18) (K18) (CK18) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| M92439 | P42704 | ATP5B_HUMAN | ATP5B ATP synthase | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| AF029082 | P31947 | PPIA_HUMAN | Peptidyl-Prolyl Cis-Trans Isomerase A (Cyclophilin A) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| M86400 | P29312 | POA3_HUMAN | Probable protein disulfide isomerase ER-60 precursor | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| L76465 | P15428 | TPIS_HUMAN | Triosephosphate isomerase (TIM) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| X57352 | Q01628 | TCTP_HUMAN | Translationally controlled tumor protein (TCTP)(p23) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| X03205 | P10809 | CH60_HUMAN | Heat shock protein 60 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| M95787 | Q01995 | VINC_HUMAN | Vinculin | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Z25821 | P42126 | EF1G_HUMAN | Elongation factor 1-gamma mRNA | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| U96132 | Q99714 | TBA1_HUMAN | Tubulin alpha-1 chain, brain specific | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| AB028893 | P04643 | CATD_HUMAN | Cathepsin D (EC 3.4.23.5).(GENE:CTSD)Homo sapiens | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| X79239 | Q02546 | EF1I_HUMAN | Elongation factor 1-alpha, mRNA | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| M13934 | P06366 | G3P2_HUMAN | Glyceraldehyde 3 phosphate dehydrogenase | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| M60854 | P17008 | GR78_HUMAN | 78 kDa glucose-regulated protein precursor (GRP 78) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| M13932 | P08708 | ENOA_HUMAN | Alpha enolase (2-phospho-D-glycerate hydrolyase) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| X69150 | P25232 | CRTC_HUMAN | Calreticulum precursor (CR55) (calregulin) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| L06498 | P17075 | COF1_HUMAN | Cofilin, non-muscle isoform | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| L04483 | P35265 | HS7C_HUMAN | Heat shock Cognate 71 kDa protein | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| M64716 | P25111 | HBB_HUMAN | Hemoglobin beta chain.(GENE: HBB) h.s | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| X77770 | P02383 | K2C8_HUMAN | Keratin, type II cytoskeletal 8 (cytokeratin 8) (K 8) (CK 8) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| W52460 | P14798 | RM12_HUMAN | Mitochondrial 60s ribosomal protein L7/L12 precursor | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| X55715 | P23396 | GR75_HUMAN | Mitochondrial stress-70 protein precursor (GRP 75) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| M77234 | P49241 | SODC_HUMAN | Superoxide Dismutase (Cu-Zn) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| M58458 | P12750 | KCRB_HUMAN | Creatine kinase | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| D16992 | P10660 | I43Z_HUMAN | 14-3-3 protein zeta/delta (protein kinase C inhibitor protein-1) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| M77233 | P23821 | RL4_HUMAN | 60s ribosomal protein L4 (L1) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| F16294 | P09058 | ANX1_HUMAN | Annexin I (lipocortin I) (calpactinII) (chromobindin 9) (p35) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| X61156 | P08865 | ROA1_HUMAN | DNA binding protein UPI, liver mRNA fragment | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| L36055 | Q13541 | FABL_HUMAN | L-FABP liver fatty acid-binding protein 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |

Table 5. Continued

| Accession Number | Swiss-Prot | Entry Name | Name | SDS-PAGE | Free-flow electrophoresis | 2-D PAGE | 2-D PAGE | 2-D PAGE | 2-D PAGE | 2-D PAGE | 2-D PAGE | 2-D PAGE |
|------------------|------------|------------|---|----------|---------------------------|----------|----------|----------|----------|----------|----------|----------|
| | | | | | | | | | | | | |
| AA316619 | P04645 | PAB1_HUMAN | Poly(A) binding protein, mRNA | | | | | | | | | |
| L14599 | Q15233 | RL3_HUMAN | Ribosomal protein L 3 | | | | | | | | | |
| X79234 | P39026 | RS19_HUMAN | S 19 ribosomal protein | | | | | | | | | |
| L06505 | P30050 | SAHH_HUMAN | S-Adenosylhomocysteine hydrolase AHCY | | | | | | | | | |
| X64707 | P26373 | TERA_HUMAN | Transitional endoplasmic reticulum ATPase (TER ATPase) | | | | | | | | | |
| X56932 | P40429 | GDIR_HUMAN | Rho GDP-dissociation inhibitor 1 | | | | | | | | | |
| L25899 | P39030 | ACON_HUMAN | Aconitate hydratase, mitochondrial precursor (aconitase) | | | | | | | | | |
| X63527 | P14118 | AKA1_HUMAN | Alcohol dehydrogenase (NADP+) (aldehyde reductase) | | | | | | | | | |
| X89401 | P46778 | AAC4_HUMAN | Alpha-Actinin 4 | | | | | | | | | |
| H59771 | P23131 | NPM_HUMAN | B23 nucleophosmin | | | | | | | | | |
| M94314 | P38663 | CATA_HUMAN | Catalase | | | | | | | | | |
| L19527 | P08526 | CD44_HUMAN | CD44 antigen (Phagocytic glycoprotein I) | | | | | | | | | |
| U14968 | P46776 | TCP2_HUMAN | Chaperonin containing TCP1, subunit 6A(zeta1) | | | | | | | | | |
| U14969 | P46779 | COXA_HUMAN | Cytochrome c oxidase polypeptide Va, mitochondrial (EC 1.9.3.1) (GENE:COX5A)H.s | | | | | | | | | |
| L38941 | P49207 | COXB_HUMAN | Cytochrome C oxidase polypeptide VB precursor.COX5B | | | | | | | | | |
| U12465 | P42766 | EF1B_HUMAN | Elongation factor 1-beta (EF-1-beta) | | | | | | | | | |
| F19234 | P18077 | ER29_HUMAN | Endoplasmic reticulum protein ERp29 (ERp31) (ERp28). | | | | | | | | | |
| X66699 | P12751 | EZRI_HUMAN | Ezrin (p81) (cytovillin)(villin-2) | | | | | | | | | |
| D23660 | P36578 | ALFA_HUMAN | Fructose-Biphosphate Aldolase A (Muscle-Type Aldolase) | | | | | | | | | |
| U14966 | P46777 | DHE3_HUMAN | Glutamate dehydrogenase 1 precursor (GDH) | | | | | | | | | |
| X06705 | P11518 | HS9A_HUMAN | Heat shock Protein HSP 90-Alpha | | | | | | | | | |
| D14531 | P32969 | ROL_HUMAN | Heterogeneous nuclear ribonucleoprotein L (HNRPN L) | | | | | | | | | |
| U30255 | P52209 | K2C1_HUMAN | Keratin, type II cytoskeletal 1 (cytokeratin 1) (K1) (CK1) | | | | | | | | | |
| M81182 | P28288 | PDX1_HUMAN | Peroxiredoxin 1 (EC 1.11.1.-) (Thioredoxin peroxidase 2) | | | | | | | | | |
| U04627 | P40939 | O60506 | PRM RNA binding protein Gry-rbp (GRY-RBP)mRNA | | | | | | | | | |
| X87949 | P11021 | PHB_HUMAN | Prohibitin | | | | | | | | | |
| M16660 | P08238 | PDI_HUMAN | Protein disulfide isomerase precursor (PDI) | | | | | | | | | |
| U79725 | Q99795 | SPCB_HUMAN | Spectrin beta chain, erythrocyte (Beta-I spectrin) | | | | | | | | | |
| D10511 | P24752 | PDX3_HUMAN | Thioredoxin-dependent peroxide reductase, mitochondrial | | | | | | | | | |
| M55040 | P22303 | TPM3_HUMAN | Tropomyosin alpha 3 chain (Tropomyosin 3) | | | | | | | | | |
| AF102542 | O95395 | NDKA_HUMAN | Non-metastatic cells 1, protein (NM23A)expressed in | | | | | | | | | |
| W45148 | P24666 | 143S_HUMAN | 14-3-3 PROTEIN SIGMA | | | | | | | | | |
| M17885 | P05388 | RL24_HUMAN | 60s ribosomal protein L24(L30) | | | | | | | | | |
| M17886 | P05386 | APA1_HUMAN | Apolipoprotein A-I | | | | | | | | | |
| M17887 | P05387 | DHSA_HUMAN | SDH2 succinate dehydrogenase flavoprotein subunit | | | | | | | | | |

Table 6. Reproducibility of transcriptome results using proteomics tools (a) and *vice versa* (b). Proteomics results have been reproduced in 67% of cases in transcriptomics studies. Results from transcriptomics studies are more difficult to reproduce using proteomics tools (25%). Only two gene products can be retrieved in three and more proteomics and transcriptomics studies (actin and creatin-kinase)

a. Transcriptomics > Proteomics

| a. Transcriptomics > Proteomics | | | | TRANSCRIPTOMICS | | | | | | | | | | PROTEOMICS | | | | | | | | | | | |
|---------------------------------|----------|------------|------------|--|--------------------------|--------------------------|-------------------|-------------------|-------------------|--------------------------|--------------------------|-------------------------------------|------------|----------------------|-------------------------------|-----------|-------------------------------------|------------------------------------|------------|------------|---------------------------|-----------|---------------------------|--------------------|------------|
| | | | | cDNA | Adaptor-tagged | SAGE | cDNA | cDNA | cDNA | SAGE | SAGE | SAGE | SAGE | cDNA | Oligonucleotide Array | SDS-PAGE | Free-flow electrophoresis | 2-D PAGE | 2-D PAGE | 2-D PAGE | 2-D PAGE | 2-D PAGE | 2-D PAGE | | |
| Accession | Number | Swiss-Prot | Entry Name | Name | Yanagawa et al 2001 [21] | Paired normal and cancer | Normal and cancer | Normal and cancer | Normal and cancer | Paired n. and pr. tumors | Paired n. and pr. tumors | Paired n. and pr. tumors or adenoma | Cell lines | Normal and Pr. tumor | Primary tumors and cell lines | Cell line | Paired n. and pr. tumor and adenoma | Paired n. and pr. tumor or adenoma | Cell lines | Cell lines | Purified epithelial cells | In silico | Purified epithelial cells | Human whole tissue | Cell lines |
| | AF041260 | D75363 | CAH2_HUMAN | Carbonic anhydrase II | | | | | | | | | | | | | | | | | | | | | |
| | AA232508 | Q9Y397 | CEA1_HUMAN | EST (BILIARY GLYCOPROTEIN) | | | | | | | | | | | | | | | | | | | | | |
| | AA007218 | Q96QY8 | BENE_HUMAN | BENE | | | | | | | | | | | | | | | | | | | | | |
| | X05014 | G01119 | CAH1_HUMAN | Carbonic anhydrase I | | | | | | | | | | | | | | | | | | | | | |
| | X75821 | Q04984 | GUAU_HUMAN | GCAP-II guanylate cyclase activator 2B; Uroguanylin; UGN | | | | | | | | | | | | | | | | | | | | | |
| | U22055 | Q96AG0 | IFM2_HUMAN | Interferon-inducible protein 1-Bd | | | | | | | | | | | | | | | | | | | | | |
| | U14631 | P80365 | FAB1_HUMAN | L-FABP liver fatty acid-binding protein 1 | | | | | | | | | | | | | | | | | | | | | |
| | M92439 | P42704 | SAHH_HUMAN | S-Adenosylhomocysteine hydrolase (AHCY) | | | | | | | | | | | | | | | | | | | | | |
| | AF029082 | P31947 | BGH3_HUMAN | Transforming growth factor, beta-induced, 66 kDa | | | | | | | | | | | | | | | | | | | | | |
| | M86400 | P29312 | ACDS_HUMAN | Acyl-CoenzymeA dehydrogenase C-2 to C-3 short chain | | | | | | | | | | | | | | | | | | | | | |
| | L76465 | P15428 | ADHG_HUMAN | ADH gamma 2 subunit (aa-1-375) | | | | | | | | | | | | | | | | | | | | | |
| | X57352 | Q01628 | CC16_HUMAN | APC5 or CDC16Hs cell division cycle 16 | | | | | | | | | | | | | | | | | | | | | |
| | X03205 | P50991 | TCPD_HUMAN | Chaperonin containing TCP-1, subunit 4 (delta) | | | | | | | | | | | | | | | | | | | | | |
| | M95787 | Q01995 | T4S3_HUMAN | Co-029;transmembrane 4 superfamily member 3 | | | | | | | | | | | | | | | | | | | | | |
| | Z25821 | P42126 | CA11_HUMAN | COL1A1 | | | | | | | | | | | | | | | | | | | | | |
| | U96132 | Q99714 | DRA_HUMAN | Colon mucosa- associated (DRA) | | | | | | | | | | | | | | | | | | | | | |
| | AB028893 | P04643 | KCRB_HUMAN | Creatine kinase | | | | | | | | | | | | | | | | | | | | | |
| | X79239 | Q02546 | TF3A_HUMAN | General transcription factor IIIA | | | | | | | | | | | | | | | | | | | | | |
| | M13934 | P06366 | GRO_HUMAN | GRO1 oncogene(melanoma growth stimulating activity, alpha) | | | | | | | | | | | | | | | | | | | | | |
| | M60854 | P17008 | GUAU_HUMAN | GUCA1B guanylate cyclase activator 1B/guanylin | | | | | | | | | | | | | | | | | | | | | |
| | M13932 | P08708 | O95784 | IgG Fc-binding protein | | | | | | | | | | | | | | | | | | | | | |
| | X69150 | P25232 | ITMC_HUMAN | Integral membrane protein 2C | | | | | | | | | | | | | | | | | | | | | |
| | L06498 | P17075 | MK03_HUMAN | MAPK3 | | | | | | | | | | | | | | | | | | | | | |
| | L04483 | P35265 | FXY3_HUMAN | MAT8 protein | | | | | | | | | | | | | | | | | | | | | |
| | M64716 | P25111 | MDP1_HUMAN | MPDP4, MDP7 microsomal dipeptidase | | | | | | | | | | | | | | | | | | | | | |
| | X77770 | P02383 | MPI2_HUMAN | M-phase inducer phosphatase 2 | | | | | | | | | | | | | | | | | | | | | |
| | W52460 | P14738 | ACTB_HUMAN | mRNA fragment encoding cytoplasmic actin | | | | | | | | | | | | | | | | | | | | | |
| | X55715 | P23396 | PRQC_HUMAN | Pyrroline 5-carboxylate reductase mRNA | | | | | | | | | | | | | | | | | | | | | |
| | M77234 | P49241 | SPRC_HUMAN | Secreted protein,acidic, cysteine-rich (osteonectin) | | | | | | | | | | | | | | | | | | | | | |
| | M58458 | P12750 | SELP_HUMAN | Selenoprotein P | | | | | | | | | | | | | | | | | | | | | |
| | O16992 | P10660 | EST1_HUMAN | EST | | | | | | | | | | | | | | | | | | | | | |
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Table 6. Continued
b. Proteomics > Transcriptomics

| | | | | TRANSCRIPTOMICS | | | | | | | | | | PROTEOMICS | | | | | | | | | |
|------------------|------------|------------|---|-----------------|----------------|------|------|------|------|------|------|------|------|------------|-----------------------|----------|---------------------------|---------|---------|---------|---------|---------|---------|
| | | | | cDNA | adaptor-tagged | SAGE | cDNA | cDNA | cDNA | SAGE | SAGE | SAGE | SAGE | cDNA | Oligonucleotide Array | SDS-PAGE | Free-flow electrophoresis | 2D-PAGE | 2D-PAGE | 2D-PAGE | 2D-PAGE | 2D-PAGE | 2D-PAGE |
| Accession Number | Swiss-Prot | Entry Name | Name | | | | | | | | | | | | | | | | | | | | |
| W52460 | P14798 | ACTB_HUMAN | mRNA fragment encoding cytoplasmic actin | | | | | | | | | | | | | | | | | | | | |
| M17887 | P05387 | GTP_HUMAN | Glutathione S-transferase M3 (brain) | | | | | | | | | | | | | | | | | | | | |
| AA496676 | P20749 | ACTG_HUMAN | Actin, Cytoplasmic 2 (Gamma-Actin) | | | | | | | | | | | | | | | | | | | | |
| U53445 | Q8IUM3 | THIO_HUMAN | Thioredoxin (ATL-derived factor) (ADF) | | | | | | | | | | | | | | | | | | | | |
| X56932 | P40429 | ATPB_HUMAN | ATP5B ATP synthase | | | | | | | | | | | | | | | | | | | | |
| U14631 | P80365 | K1CR_HUMAN | Keratin, type I cytoskeletal 18 (cytokeratin 18) (K18) (CK18) | | | | | | | | | | | | | | | | | | | | |
| M33987 | P00915 | ANX4_HUMAN | Annexin IV (Lipocortin IV) | | | | | | | | | | | | | | | | | | | | |
| X51346 | P17535 | CH60_HUMAN | Heat shock protein 60 | | | | | | | | | | | | | | | | | | | | |
| X17206 | P15860 | PPIA_HUMAN | Peptidyl-Prolyl Cis-Trans Isomerase A (Cyclophilin A) | | | | | | | | | | | | | | | | | | | | |
| M36035 | P30536 | TPIS_HUMAN | Triosephosphate isomerase (TIM) | | | | | | | | | | | | | | | | | | | | |
| X75821 | Q04984 | ANX3_HUMAN | Annexin III (Lipocortin III) | | | | | | | | | | | | | | | | | | | | |
| W52942 | P29312 | PDA3_HUMAN | Probable protein disulfide isomerase ER-60 precursor | | | | | | | | | | | | | | | | | | | | |
| W89072 | Q01628 | TCTP_HUMAN | Translationally controlled tumor protein (TCTP)(p23) | | | | | | | | | | | | | | | | | | | | |
| W85876 | Q01995 | VINC_HUMAN | Vinculin | | | | | | | | | | | | | | | | | | | | |
| AB028893 | P04643 | KCRB_HUMAN | Creatine kinase | | | | | | | | | | | | | | | | | | | | |
| U80040 | Q99798 | G3P2_HUMAN | Glyceraldehyde 3 phosphate dehydrogenase | | | | | | | | | | | | | | | | | | | | |
| AA461325 | Q9UEY8 | K2C8_HUMAN | Keratin, type II cytoskeletal 8 (cytokeratin 8) (K 8) (CK 8) | | | | | | | | | | | | | | | | | | | | |
| D31840 | P54259 | GR78_HUMAN | 78 kDa glucose-regulated protein precursor (GRP 78) | | | | | | | | | | | | | | | | | | | | |
| J04469 | P12532 | CATD_HUMAN | Cathepsin D (EC 3.4.23.5)(GENE:CTSD)Homo sapiens | | | | | | | | | | | | | | | | | | | | |
| X73502 | P35900 | COF1_HUMAN | Cofilin, non-muscle isoform | | | | | | | | | | | | | | | | | | | | |
| Z11692 | P13639 | EF11_HUMAN | Elongation factor 1-alpha, mRNA | | | | | | | | | | | | | | | | | | | | |
| X84694 | P49411 | EF1G_HUMAN | Elongation factor 1-gamma mRNA | | | | | | | | | | | | | | | | | | | | |
| U20272 | Q92664 | HSC7_HUMAN | Heat shock Cognate 71 kDa protein | | | | | | | | | | | | | | | | | | | | |
| D16111 | P30086 | TBA1_HUMAN | Tubulin alpha-1 chain, brain specific | | | | | | | | | | | | | | | | | | | | |
| M13332 | P08708 | ENO4_HUMAN | Alpha enolase (2-phospho-D-glycerate hydro-lyase) | | | | | | | | | | | | | | | | | | | | |
| Z70701 | P25232 | CRTC_HUMAN | Calreticulum precursor (CR55) (calregulin) | | | | | | | | | | | | | | | | | | | | |
| F15744 | P25111 | HBB_HUMAN | Hemoglobin beta chain (GENE: HBB) h.s | | | | | | | | | | | | | | | | | | | | |
| T74426 | Q9UNH5 | RM12_HUMAN | Mitochondrial 60s ribosomal protein L7/L12 precursor | | | | | | | | | | | | | | | | | | | | |
| D60944 | Q9Y365 | GR75_HUMAN | Mitochondrial stress-70 protein precursor (GRP 75) | | | | | | | | | | | | | | | | | | | | |
| Hs.7369 | P49241 | SODC_HUMAN | Superoxide Dismutase (Cu-Zn) | | | | | | | | | | | | | | | | | | | | |

necrosis, blood, stool, etc.), so that standardized sample preparation procedures are critical for obtaining reproducible results. Several sample preparation methods have been described, in particular fluorescence-activated cell-sorting (FACS) [4], laser capture microdissection (LCM) [5–8], immunomagnetic beads separation [9], and cellular fractionation [10]. Unfortunately, these sample preparation procedures were barely applied in CRC. In our experience, the beads-based method is characterized by several advantages when compared with other cell purification procedures [11].

Third, gene expression patterns depend on the arrays technology platform. In transcriptomics studies any factors may affect the outcome of a microarray experiment, in particular technical, instrumental, computational, and interpretative factors. In fact, lack of reproducibility and accuracy is a major concern in microarray studies [12]. When cross-platform comparison was performed, reproducibility was insufficient: only four genes from a set of 185 common genes selected behaved consistently on three array platforms, and agreement of about 30% was found between two brands [13].

Fourth, in proteomics studies, 2-D PAGE or 2-D DIGE have well-known technological limitations. In CRC, even after epithelial cell enrichment using magnetic beads, the mean CV of repeated 2-D PAGE analysis with silver staining was found to lie between 20 and 28%. Only 47% (interrun) to 76% (intrarun) of spots could be matched within a triplicate experiment. Interindividual phenotypic variability was high. Thus, even under well-defined experimental conditions, 2-D PAGE parallel analysis of paired CRC samples is hampered by a significant variability [2].

Fifth, the methods applied for generating, formatting, storing, retrieving, and querying data are of outmost importance to assess methodological and biological variation in gene expression analysis. Unfortunately, due to the small sample size (number of patients), large number of variables examined at once, and absence of double or triple experiments (arrays and gels are expensive and samples are rare) statistical analysis is often not valid. In particular, assessing the reproducibility of a variable is necessary (e.g., using the intraclass correlation coefficient) for comparing multiple samples at once. The use of median values instead of mean values has been shown to improve data correction [14]. It has also been proposed to use housekeeping genes as endogenous controls [15]. A dedicated society, The Microarray Gene Expression Data (MGED) Society, has been formed to facilitate the sharing of gene expression data generated by functional genomics and proteomics experiments [16].

Finally, correlation between results of transcriptomics versus proteomics results is low. For CRC, there is no publication comparing mRNA and protein expression for a cohort of genes. However, extrapolation is reasonable from another epithelial cancer (lung adenocarcinoma), where such comparison has been performed. Only a subset of the proteins (17%) exhibited a significant correlation with mRNA abundance [17].

Obviously, many genes and factors found to be differentially regulated (both in transcriptomics and proteomics studies) do not play a causal role in CRC carcinogenesis. For example in the studies under investigation, at least 17 mRNAs encoding ribosomal proteins were identified to be dysregulated using cDNA arrays and 39 ESTs using SAGE technology. This broad dysfunction of protein synthesis, in particular of small molecules synthesis, has been reported not only in cancer but also in several other human diseases, where etiologies have been linked to mutations in genes of the translational control machinery [36].

However, some findings might be of particular interest in human CRC. For example, a small molecule group found to be dysregulated in human CRC is the 14–3–3 proteins family. 14–3–3 proteins are ubiquitous within all eukaryotic cells and participate in protein kinase signaling pathways. In particular, they are involved in phosphorylation-dependent protein–protein interactions that control progression through the cell cycle, initiation and maintenance of DNA damage checkpoints, activation of MAP kinases, prevention of apoptosis, and coordination of integrin signaling and cytoskeletal dynamics [37]. Given the prevalence of specific 14–3–3 isoforms expression in several human epithelial cancers [38, 39], these proteins may be involved in cancer tumorigenesis and particular isoforms may be useful as therapeutic targets in human CRC.

In summary, we propose a gene expression data warehouse in human CRC that is intended to help researchers active in the field to get an overview of the data available. However, reproducibility of results obtained in different studies was disappointing.

As a matter of fact, the development of some types of unified processes for generating information, formatting, storing, retrieving, and querying data, regardless of the technology used to generate it, is of central importance for building gene expression databases. Such unified processes have been proposed (see above) but have found only limited recognition so far. This is unfortunate because these methodological issues must be solved before trying to integrate experimental data from different sources into functional proteomics studies at the bench or *in silico*. The present meta-analysis of gene expression highlights the need for including a sufficient number of patients, analyzing only purified epithelial cells, using clinical standards (such as the international classification of disease, histopathology, and staging), assessing the variability of the technology applied, considering common standards for microarray data annotation and exchange, and finally for developing software implementing these standards and promoting the sharing of these high quality, well-annotated data within the life sciences community.

In the absence of such unified processes, reproducibility of results within a laboratory will continue to be low, confirmation of results between different groups will stay difficult, and identification of diagnostic and therapeutic targets will remain a lottery. These problems are challenging the whole drug dis-

covery process from the very beginning, namely target identification, so that target validation, assay development, and prioritization of compounds remains a high risk endeavor.

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5 References

- [1] Friedman, D. B., Hill, S., Keller, J. W. et al., *Proteomics* 2004, 4, 793–811.
- [2] Ott, V., Guenther, K., Steinert, R. et al., *Pharmacogenomics J.* 2001, 1, 142–151.
- [3] Ji, H., Whitehead, R. H., Reid, G. E. et al., *Electrophoresis*, 1994, 15, 391–405.
- [4] Reymond, M. A., Sánchez, J. C., Schneider, C. et al., *Electrophoresis* 1997, 18, 622–624.
- [5] Kondo, T., Seike, M., Mori, Y. et al., *Proteomics* 2003, 3, 1758–1766.
- [6] Banks, R. E., Dunn, M. J., Forbes, M. A. et al., *Electrophoresis* 1999, 20, 689–700.
- [7] Craven, R. A., Banks, R. E., *Proteomics* 2001, 1, 1200–1204.
- [8] Craven, R. A., Totty, N., Harnden, P. et al., *Am. J. Pathol.* 2002, 160, 815–822.
- [9] Reymond, M. A., Sanchez, J. C., Hughes, G. J. et al., *Electrophoresis* 1997, 18, 2842–2848.
- [10] Szymczyk, P., Krajewska, W. M., Jakubik, J. et al., *Tumori* 1996, 82, 376–381.
- [11] Kellner, U., Steinert, R., Seibert, V. et al., *Pathol. Res. Pract.* 2004, 200, 155–163.
- [12] Shi, L., Tong, W., Goodsaid, F. et al., *Expert Rev. Mol. Diagn.* 2004, 4, 761–777.
- [13] Tan, P. K., Downey, T. J., Spitznagel, E. L. Jr. et al., *Nucleic Acids Res.* 2003, 31, 5676–5684.
- [14] Pellis, L., Franssen-van Hal, N. L., Burema, J., Keijer, J., *Physiol. Genomics* 2003, 16, 99–106.
- [15] Janssens, N., Janicot, M., Perera, T., Bakker, A., *Mol. Diagn.* 2004, 8, 107–113.
- [16] www.mged.org/, consulted on December 25, 2004.
- [17] Chen, G., Gharib, T. G., Huang, C. C. et al., *Mol. Cell. Proteomics* 2002, 1, 304–313.
- [18] Zhang, L., Zhou, W., Velculescu, V. E. et al., *Science* 1997, 276, 1268–1272.
- [19] www.sagenet.org/cancer/table1.htm, consulted on December 1, 2003.
- [20] Parle-McDermott, A., McWilliam, P., Tighe, O., Dunican, D., Croke, D. T., *Br. J. Cancer* 2000, 83, 725–728.
- [21] Yanagawa, R., Furukawa, Y., Tsunoda, T. et al., *Neoplasia* 2001, 3, 395–401.
- [22] Takemasa, I., Higuchi, H., Yamamoto, H. et al., *Biochem. Biophys. Res. Commun.* 2001, 285, 1244–1249.
- [23] Notterman, D. A., Alon, U., Sierk, A. J., Levine, A. J., *Cancer Res.* 2001, 61, 3124–3130.
- [24] Buckhaults, P., Rago, C., St. Croix, B. et al., *Cancer Res.* 2001, 61, 6996–7001.
- [25] Birkenkamp-Demtroder, K., Christensen, L. L., Olesen, S. H. et al., *Cancer Res.* 2002, 61, 4352–4363.
- [26] Lin, Y. M., Furukawa, Y., Tsunoda, T. et al., *Oncogene* 2002, 21, 4120–4128.
- [27] Miuro, S., Takemasa, I., Oba, S. et al., *Genome Biol.* 2003, 4, R21.
- [28] Williams, N. S., Gaynor, R. B., Scoggin, S. et al., *Clin. Cancer Res.* 2003, 9, 931–946.
- [29] Simpson, R. J., Connolly, L. M., Eddes, J. S. et al., *Electrophoresis* 2000, 21, 1707–1732.
- [30] Lawrie, L. C., Curran, S., McLeod, H. L., Fothergill, J. E., Murray, G. I., *Mol. Pathol.* 2001, 54, 253–258.
- [31] Simpson, R. J., *Proteomics* 2001, 1, 807–818.
- [32] Medjahed, D., Luke, B. T., Tontesh, T. S. et al., *Proteomics* 2003, 3, 1445–1453.
- [33] Demakts, I., Swiss-2-D PAGE database: Search for DLD1_Human. <http://www.expasy.org>, Release 16 and updates up to 17-Nov-2003.
- [34] Stierum, R., Gaspari, M., Dommels, Y., et al., *Biochim. Biophys. Acta* 2003, 1650, 73–91.
- [35] Friedman, D. B., Hill, S., Keller, J. W. et al., *Proteomics* 2004, 4, 793–811.
- [36] Calkhoven, C. F., Muller, C., Leutz, A., *Trends Mol. Med.* 2002, 8, 577–583.
- [37] Wilker, E., Yaffe, M. B., *J. Mol. Cell Cardiol.* 2004, 37, 633–642 (review).
- [38] Qi, W., Liu, X., Qiao, D. et al., *Int. J. Cancer* 2005, 113, 359–363.
- [39] Moreira, J. M., Gromov, P., Celis, J. E., *Mol. Cell. Proteomics* 2004, 3, 410–419.

17

Proteomics

Kathryn Lilley, Azam Razzaq and Michael J. Deery

17.1 Introduction

There are now numerous organisms whose genome sequences are known, for example *Arabidopsis* (*Arabidopsis* Genome Initiative, 2000), *Drosophila* (Adams *et al.*, 2000), human (International Human Genome Sequencing Consortium, 2001) and rice (Yu *et al.*, 2002). The prediction of open-reading frames from these genomic sequences has enabled the comprehensive identification of many putative protein sequences. These proteins can be arranged into three categories, namely those of known function, those with recognisable motifs and hence a vague idea of function, and those with no sequence similarities to any protein (Gabor Miklos and Malenzska, 2001). Many proteins reside in this latter 'functional vacuum' which could represent as much as 30% of the predicted proteins. Determining protein function is key to understanding cellular mechanism. Studying how protein expression is modulated in response to a given set of circumstances, such as infection, disease, developmental stage, senescence or response to drugs, will facilitate the elucidation of disease pathways and thus provide a mechanism for diagnosis and therapy.

DNA chips (mRNA profiling studies) can contribute to the study of gene expression in response to a particular biological perturbation. However, the extrapolation that changes in transcript level will also result in corresponding changes in protein amount or activity cannot always be made. To understand fully, we need integrated data sets from a variety of protein expression studies, providing information on relative abundances, sub-cellular locations, protein complex formation and the profiling of isoforms generated by either alternate mRNA splicing or post-translational modifications. Proteomics is the word now

commonly used to describe the discipline associated with the acquisition of these data sets.

17.2 Definitions and applications

Before embarking on an overview of the techniques used in the field of proteomics, it is necessary to define a few terminologies; proteomics, functional genomics, structural genomics and post-genomics are terms that have crept into the scientific vocabulary with alarming stealth and are used freely, and in some cases interchangeably, especially in cases where the use of such 'buzz words' is likely to increase the attendance at departmental seminars.

- **Genome:** A genome represents the entire DNA content in a particular cell, whether or not it is coding, non-coding, or is located either chromosomally or extra-chromosomally.
- **Genomics:** Genomics is the study of the genome, interrogating the complete genome sequence using both DNA and RNA methodologies.
- **Proteome:** The proteome represents the complete set of proteins encoded by the genome.
- **Proteomics:** Proteomics is the study of the proteome and investigates the cellular levels of all the isoforms and post-translational modifications of proteins that are encoded by the genome of that cell under a given set of circumstances.
- **Functional genomics:** This is the study of the functions of genes and their inter-relationships.

Whilst a genome is more or less static, the protein levels in a particular cell can change dramatically as genes get turned on and off during the cell's response to its environment. The proteome originally was defined over seven years ago as 'all the proteins coded by the genome of an organism' (Wasinger *et al.*, 1995). Nowadays the term 'proteomics' is used more widely and implies an effort to link structure to function by whatever means are appropriate. We can expect the definition to change again with time as the field and the investigator's view of it evolves.

17.3 Stages in proteome analysis

Proteomic analysis can ascertain function either by looking for changes in the expression of either all or a subset of proteins, or by identifying binding partners for particular proteins and seeing how their interaction is affected by biological perturbation. Whatever the rationale of the investigation, or the number of proteins involved, the study of the proteome can be broken down into the following stages of analysis.

ORIGINAL ARTICLE

Identification of putative oncogenes in lung adenocarcinoma by a comprehensive functional genomic approach

R Li¹, H Wang¹, BN Bekele², Z Yin³, NP Caraway¹, RL Katz¹, SA Stass⁴ and F Jiang¹¹Department of Pathology, The University of Texas MD Anderson Cancer Center, Houston, TX, USA; ²Department of Biostatistics & Applied Mathematics, The University of Texas MD Anderson Cancer Center, Houston, TX, USA; ³Department of Internal Medicine, The University of Texas Houston Medical School, Houston, TX, USA and ⁴Department of Pathology, The University of Maryland School of Medicine, Baltimore, MD, USA

Amplification and overexpression of putative oncogenes confer growth advantages for tumor development. We used a functional genomic approach that integrated simultaneous genomic and transcript microarray, proteomics, and tissue microarray analyses to directly identify putative oncogenes in lung adenocarcinoma. We first identified 183 genes with increases in both genomic copy number and transcript in six lung adenocarcinoma cell lines. Next, we used two-dimensional polyacrylamide gel electrophoresis and mass spectrometry to identify 42 proteins that were overexpressed in the cancer cells relative to normal cells. Comparing the 183 genes with the 42 proteins, we identified four genes – *PRDX1*, *EEF1A2*, *CALR*, and *KCIP-1* – in which elevated protein expression correlated with both increased DNA copy number and increased transcript levels (all $r > 0.84$, two-sided $P < 0.05$). These findings were validated by Southern, Northern, and Western blotting. Specific inhibition of *EEF1A2* and *KCIP-1* expression with siRNA in the four cell lines tested suppressed proliferation and induced apoptosis. Parallel fluorescence *in situ* hybridization and immunohistochemical analyses of *EEF1A2* and *KCIP-1* in tissue microarrays from patients with lung adenocarcinoma showed that gene amplification was associated with high protein expression for both genes and that protein overexpression was related to tumor grade, disease stage, Ki-67 expression, and a shorter survival of patients. The amplification of *EEF1A2* and *KCIP-1* and the presence of overexpressed protein in tumor samples strongly suggest that these genes could be oncogenes and hence potential targets for diagnosis and therapy in lung adenocarcinoma. *Oncogene* (2006) 25, 2628–2635. doi:10.1038/sj.onc.1209289; published online 12 December 2005

Keywords: lung cancer; microarrays; proteomics; tissue microarray

Introduction

In lung adenocarcinoma, as in other types of cancer, gene amplification and the consequent overexpression of the amplified oncogene play an important role in the development of tumors, because their overexpression confers a growth advantage. The ability to identify putative oncogenes that are activated during tumorigenesis could facilitate the choice of molecular genetic targets for diagnosis and therapy of the disease. This concept has been exemplified by *HER-2*, which was first found to be amplified in neuroblastomas and subsequently shown to be associated with poor prognosis in breast cancer (Ross and Fletcher, 1999). Now, *HER-2* aberrations are used as a predictor of response to therapy, and treatment of *HER-2*-positive breast cancer with the monoclonal anti-*HER-2* antibody trastuzumab has been shown to improve prognosis (Ross and Fletcher, 1999). Emerging evidence of common amplicons in lung adenocarcinomas (Luk *et al.*, 2001; Jiang *et al.*, 2004; Tonon *et al.*, 2005) suggests that additional oncogenes remain to be identified; however, conventional techniques are ineffective in pinpointing such oncogenes. Parallel measurement of DNA copy number and mRNA levels in cDNA microarrays permits changes in copy number to be compared with transcription levels on a gene-by-gene basis to generate lists of candidate genes within the defining amplicons (Hyman *et al.*, 2002; Pollack *et al.*, 2002). However, use of transcript patterns does not allow assessment of the expression of protein products or identification of proto-oncogenes. Another approach, identifying differentially expressed proteins by proteomic analysis and then comparing the proteins present with mRNA expression in cDNA microarrays from the same specimens, can clarify the extent to which changes in transcript patterns reflect changes in their cognate proteins and post-transcriptional mechanisms (Chen *et al.*, 2002), but this approach cannot be used to identify oncogenes driven by extensive increases of their gene copy number. Moreover, using individual microarrays or proteomic approaches alone cannot distinguish the cancer-driving oncogenes that directly propel tumor progression from the larger number of passenger genes that may be concurrently over-represented but are not biologically relevant in tumor development.

Correspondence: Assistant Professor F Jiang, Department of Pathology, The University of Maryland School of Medicine, 10 South Pine Street, MSTF 7th floor, Baltimore, MD 21201-1192, USA.
E-mail: fjiang@som.umaryland.edu

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In this study, we used a comprehensive approach that integrated simultaneous comparative genomic hybridization (CGH) and transcript microarray with proteomic analyses of six lung adenocarcinoma cell lines. We directly and specifically identified four putative oncogenes that could have been activated through amplification and consequent elevation of transcript expression. We used small interfering RNA (siRNA) to inhibit the expression of two of these four genes in the lung cancer cell lines, which further implicated them in oncogenesis. We then explored the clinical significance of these findings by assessing the expression of these two genes in tissue microarrays of human lung cancer specimens. Our findings underscore the power of integrated functional genomic analyses for identifying putative oncogenes in tumorigenesis; such activated genes could be useful as targets for diagnosis or therapy in lung cancer.

Results

Simultaneous global genomic and transcript analyses identify 183 genes with increases in genomic copy numbers and transcript expression levels

To identify genes in which increased DNA copy number might contribute to increased transcript in lung adenocarcinomas, first we used CGH with microarrays of six lung adenocarcinoma cell lines. We identified 587 genes showing increases in DNA copy number across all six cell lines (Supplementary Table 1S), which were distributed as 90 amplicons on all chromosomes except for chromosomes 13 and Y (Supplementary Table 2S). A subsequent transcript test with the identical arrays of the same cell lines revealed 275 genes that showed increased mRNA levels (Supplementary Table 3S). Using random permutation tests across all cancer cell lines, we identified 183 genes (31%) that showed elevated transcript levels from the 587 genes that were over-represented in the genome (Table 1), suggesting that elevated transcript levels of the 183 genes may reflect their genomic over-representation in the cancer cells. These findings are consistent with previous reports linking genomic changes with altered transcript patterns in breast cancer (Hyman *et al.*, 2002; Pollack *et al.*, 2002). However, our finding that only 31% of the genes showing increased DNA copy numbers had cognate increases in transcript expression in lung adenocarcinomas is different from the overall rates of 40–60% reported for breast cancer (Hyman *et al.*, 2002; Pollack *et al.*, 2002). This discordance may reflect methodologic differences between studies or biological differences between breast cancer and lung adenocarcinoma.

Proteomic analyses identify four genes for which protein abundance was associated with increases in the cognate gene and transcript levels

Analysis of transcript patterns is insufficient for understanding the expression of protein products and the effect of genomic over-representation on the expression

of their cognate proteins. To extend these findings beyond genomic over-representation to expression of the protein products of those genes, we next assessed protein expression in the same cell lines by two-dimensional polyacrylamide gel electrophoresis (PAGE) and found that 42 different proteins, representing 42 individual genes, were significantly increased in the cancer cell lines (Table 2; Supplementary Figures 1S and 2S). Some of these proteins were identified as having multiple isoforms, and all individual isoforms exhibited increases in expression ranging from 4.6 to 12.8 times their expression in normal lung tissue cells. In comparing protein level of the 42 genes with changes in their cognate genomic and mRNA expression from the global microarray analyses, we found that four (9.5%) of those 42 genes – *PRDX1*, *EEF1A2*, *CALR*, and *KCIP-1* – showed statistically significant correlations between elevated protein expression and increases in both copy number and mRNA expression (all $r > 0.84$; $P < 0.05$) (Table 2) in the cancer cell lines. These findings imply that the abundance of these four proteins is attributable to the amplification and consequent elevated transcription of their cognate genes.

Validation of copy number, transcript, and protein expression of PRDX1, EEF1A2, CALR, and KCIP-1 in lung cancer cell lines

To confirm our findings from the high-throughput analyses, we next used Southern, Northern, and Western blotting to assess DNA, RNA, and protein levels for the four genes identified in the six cell lines. For comparison, we arbitrarily chose one gene, *NFKB1*, in which an increase in protein level did not correlate with genetic changes. Overall, we found excellent concordance between the CGH microarray and Southern blotting analyses, transcript array and Northern blotting analyses, and proteomic and Western blotting analyses for all five genes (Figure 1). For example, *KCIP-1* showed fivefold amplification in five of the six cancer cell lines, whereas *NFKB1* showed no such increase in any of the cell lines. As for transcript expression, Northern blotting of *EEF1A2* showed high expression in five of the six cancer cell lines; again, levels of *NFKB1* transcript were not increased in any cancer cell line as compared with normal bronchial epithelial cells. The results of Western blotting were also consistent with the results of the proteomic experiments; for example, five of the cancer cell lines exhibited strong protein bands for *PRDX1* as compared with normal cells. These findings provide strong support for the validity of the results derived from the high-throughput techniques in this study.

These parallel analyses also revealed close correlations in the extent of changes in gene copies, transcript, and protein of each of the four genes in the cancer cell lines. For example, in the five cancer cell lines that showed at least fourfold increases in *EEF1A2* copy number, expression of transcript and protein was also increased by at least a factor of four as well (relative to their expression in normal cells) (Supplementary Figure 3S). The protein abundance of the four genes showing

Table 1 List of 183 genes with statistically significant correlation (0.05) between genomic copy number and transcript level

| Gene symbol | Chro. | Distance from p arm of each chromosome (Mb) | α |
|-------------|-------|---|----------|
| ENO1 | 1 | 8.5 | 0.0085 |
| DDOST | 1 | 20.1 | 0.0111 |
| SFN | 1 | 26.4 | 0.0113 |
| MLP | 1 | 32.2 | 0.0114 |
| AKR1A1 | 1 | 45.4 | 0.0128 |
| PRDX1 | 1 | 45.4 | 0.0122 |
| UQCRH | 1 | 46.2 | 0.0125 |
| RPL7 | 1 | 96.4 | 0.0127 |
| COL11A1 | 1 | 102.6 | 0.0129 |
| MCL1 | 1 | 147.3 | 0.0222 |
| PSMB4 | 1 | 148.1 | 0.0131 |
| JTB | 1 | 150.7 | 0.0134 |
| RPS27 | 1 | 150.7 | 0.0135 |
| HAX1 | 1 | 151 | 0.0266 |
| MUC1 | 1 | 151.9 | 0.0143 |
| CCT3 | 1 | 153.1 | 0.0167 |
| CRABP2 | 1 | 153.4 | 0.0148 |
| TKT | 1 | 159.3 | 0.0152 |
| ATPIB1 | 1 | 165.8 | 0.0234 |
| CHIT1 | 1 | 199.7 | 0.0154 |
| SNRPE | 1 | 200.2 | 0.0165 |
| YWHAQ | 2 | 9.6 | 0.0159 |
| ODC1 | 2 | 10.60 | 0.0119 |
| RPL31 | 2 | 101.20 | 0.0161 |
| BENE | 2 | 110.40 | 0.0169 |
| STAT1 | 2 | 191.80 | 0.0175 |
| HSPD1 | 2 | 198.30 | 0.0277 |
| HSPE1 | 2 | 198.30 | 0.0185 |
| RPL37A | 2 | 217.30 | 0.0388 |
| IGFBP2 | 2 | 217.50 | 0.0189 |
| RPS7 | 2 | 3.30 | 0.0193 |
| RAB1A | 2 | 65.30 | 0.0204 |
| IGKC | 2 | 89.00 | 0.0285 |
| LTF | 3 | 46.3 | 0.0455 |
| PFN2 | 3 | 151 | 0.0207 |
| KPNA4 | 3 | 161.5 | 0.0211 |
| SI00P | 4 | 6.7 | 0.1122 |
| UGDH | 4 | 39.3 | 0.0215 |
| UCHL1 | 4 | 41.1 | 0.0222 |
| SPPI | 4 | 89.3 | 0.0227 |
| TRIM2 | 4 | 154.7 | 0.0231 |
| FGB | 4 | 156 | 0.0235 |
| EGG | 4 | 156 | 0.0441 |
| SDHA | 5 | 0.251 | 0.0243 |
| PDCD6 | 5 | 0.305 | 0.0245 |
| CCT5 | 5 | 10.3 | 0.0446 |
| PTPRF | 5 | 14.2 | 0.0248 |
| RPL37 | 5 | 40.8 | 0.0251 |
| ENCI | 5 | 74 | 0.0336 |
| QP-C | 5 | 132.2 | 0.0466 |
| SPINK1 | 5 | 147.2 | 0.0256 |
| CANX | 5 | 179.2 | 0.0263 |
| SOX4 | 6 | 21.7 | 0.0321 |
| HDGF | 6 | 22.6 | 0.0362 |
| RPS10 | 6 | 34.6 | 0.0177 |
| RPL10A | 6 | 35.4 | 0.0369 |
| VEGF | 6 | 43.7 | 0.0372 |
| OSF-2 | 6 | 45.4 | 0.0173 |
| FSCN1 | 7 | 5.3 | 0.0378 |
| CYCS | 7 | 24.9 | 0.0381 |
| CBX3 | 7 | 25.9 | 0.0289 |
| IGFBP3 | 7 | 45.7 | 0.0389 |
| CLDN4 | 7 | 72.7 | 0.0403 |
| HSPB1 | 7 | 75.5 | 0.0433 |
| CALR | 7 | 92.7 | 0.0425 |
| COL1A2 | 7 | 93.6 | 0.0457 |
| ATPSJ2 | 7 | 98.7 | 0.0475 |
| AKR1B10 | 7 | 133.6 | 0.0481 |

Table 1 (continued)

| Gene symbol | Chro. | Distance from p arm of each chromosome (Mb) | α |
|-------------|-------|---|----------|
| RPS20 | 8 | 56.7 | 0.0482 |
| TCEB1 | 8 | 74.6 | 0.0486 |
| LAPTM4B | 8 | 98.5 | 0.0497 |
| RPL30 | 8 | 98.7 | 0.0054 |
| KCIP-1 | 8 | 101.6 | 0.0093 |
| PABPC1 | 8 | 101.78 | 0.0119 |
| EEF1D | 8 | 144.4 | 0.0121 |
| TSTA3 | 8 | 144.5 | 0.0122 |
| RPL8 | 8 | 145.6 | 0.0128 |
| TRA1 | 9 | 117.1 | 0.0136 |
| RPL35 | 9 | 121.1 | 0.0133 |
| HSPA5 | 9 | 121.5 | 0.0135 |
| LCN2 | 9 | 124.4 | 0.0137 |
| DPP7 | 9 | 133.4 | 0.0139 |
| PFKP | 10 | 3.2 | 0.0223 |
| AKRIC1 | 10 | 5.1 | 0.0146 |
| PLAU | 10 | 75.6 | 0.0356 |
| DSP | 10 | 76.7 | 0.0289 |
| TALDO1 | 11 | 0.434 | 0.0143 |
| SLC22A1L | 11 | 2.9 | 0.0151 |
| TSSC3 | 11 | 2.9 | 0.0611 |
| RPL27A | 11 | 8.7 | 0.0156 |
| ST5 | 11 | 8.8 | 0.0162 |
| LDHA | 11 | 18.5 | 0.0168 |
| MDK | 11 | 46.4 | 0.0162 |
| DOC-IR | 11 | 67.5 | 0.0167 |
| MMPI2 | 11 | 102.8 | 0.0177 |
| HYOU1 | 11 | 118.9 | 0.0183 |
| SCNN1A | 12 | 6.3 | 0.0185 |
| LDHB | 12 | 21.7 | 0.0193 |
| KRT7 | 12 | 52.3 | 0.0196 |
| KRT5 | 12 | 52.6 | 0.0197 |
| KRT6E | 12 | 52.6 | 0.0201 |
| ERBB3 | 12 | 56.2 | 0.0212 |
| NACA | 12 | 56.8 | 0.0218 |
| TM4SF3 | 12 | 71.2 | 0.0401 |
| NTS | 12 | 86.2 | 0.0215 |
| ASCL1 | 12 | 103.3 | 0.0219 |
| TXNRD1 | 12 | 104.6 | 0.0223 |
| CKAP4 | 12 | 106.6 | 0.0124 |
| COX6A1 | 12 | 120.7 | 0.0435 |
| BGN | 12 | 122.5 | 0.0235 |
| RAN | 12 | 129.88 | 0.0238 |
| RPL36A | 14 | 48.1 | 0.0243 |
| PGD | 14 | 50.7 | 0.0248 |
| THBS2 | 15 | 37.5 | 0.0251 |
| TRAF4 | 15 | 38.3 | 0.0253 |
| SPINT1 | 15 | 38.7 | 0.0254 |
| RPL17 | 15 | 45.26 | 0.0411 |
| PKM2 | 15 | 70.1 | 0.0258 |
| IDH2 | 15 | 88.2 | 0.0211 |
| RPL23A | 16 | 0.377 | 0.0264 |
| MSLN | 16 | 0.753 | 0.0366 |
| UBE2I | 16 | 1.3 | 0.0271 |
| RPS2 | 16 | 1.95 | 0.0281 |
| CLDN9 | 16 | 3.1 | 0.0329 |
| ARL6IP | 16 | 18.7 | 0.0412 |
| EIF3S8 | 16 | 28.3 | 0.0336 |
| TUFM | 16 | 28.9 | 0.0377 |
| ALDOA | 16 | 30.1 | 0.038 |
| NME4 | 16 | 53.6 | 0.0381 |
| GPR56 | 16 | 57.4 | 0.0386 |
| CDH1 | 16 | 68.5 | 0.0289 |
| NQO1 | 16 | 69.5 | 0.0396 |
| SLC7A5 | 16 | 87.6 | 0.0397 |
| APRT | 16 | 88.6 | 0.0411 |
| GALNS | 16 | 88.6 | 0.0255 |
| RPL13 | 16 | 89.3 | 0.0431 |
| MCP | 17 | 32.4 | 0.0465 |

Table 1 (continued)

| Gene symbol | Chro. | Distance from p arm of each chromosome (Mb) | α |
|-------------|-------|---|----------|
| ERBB2 | 17 | 35.11 | 0.0483 |
| JUP | 17 | 39.8 | 0.0495 |
| CRF | 17 | 40.39 | 0.0505 |
| RPL27 | 17 | 41.1 | 0.0046 |
| NME1 | 17 | 46.59 | 0.0082 |
| COL1A1 | 17 | 48.6 | 0.0108 |
| ABCC3 | 17 | 49.1 | 0.0326 |
| NME2 | 17 | 49.6 | 0.0111 |
| RPL38 | 17 | 72.7 | 0.0117 |
| SMT3H2 | 17 | 73.6 | 0.0119 |
| SYNGR2 | 17 | 76.6 | 0.0122 |
| LGALS3BP | 17 | 77.4 | 0.0127 |
| P4HB | 17 | 80.3 | 0.0126 |
| PPAP2C | 19 | 0.221 | 0.0228 |
| GPI | 19 | 39.55 | 0.0145 |
| HPN | 19 | 40.2 | 0.0129 |
| ZNF146 | 19 | 41.4 | 0.0131 |
| SPINT2 | 19 | 43.4 | 0.0238 |
| PSMD8 | 19 | 43.5 | 0.0132 |
| YIF1P | 19 | 43.5 | 0.0135 |
| RPS16 | 19 | 44.6 | 0.0144 |
| CEACAM5 | 19 | 46.9 | 0.0145 |
| CEACAM6 | 19 | 46.9 | 0.0143 |
| GIPR | 19 | 50.8 | 0.0259 |
| SNRPD2 | 19 | 50.9 | 0.0413 |
| KDELRL1 | 19 | 53.6 | 0.0152 |
| RPL28 | 19 | 60.6 | 0.0156 |
| RPS5 | 19 | 63.6 | 0.0267 |
| TRIM28 | 19 | 63.7 | 0.0158 |
| DAP | 20 | 35.6 | 0.0166 |
| TOP1 | 20 | 40.3 | 0.0172 |
| UBE2C | 20 | 45.1 | 0.0174 |
| RPS21 | 20 | 61.6 | 0.0268 |
| EEF1A2 | 20 | 62.8 | 0.0185 |
| TFF3 | 21 | 42.6 | 0.0186 |
| TFF1 | 21 | 42.7 | 0.0192 |
| CSTB | 21 | 44.1 | 0.0201 |
| MIF | 22 | 22.6 | 0.0202 |
| XBPI | 22 | 27.5 | 0.0204 |
| PRDX4 | X | 22.9 | 0.0198 |
| SYN1 | X | 46.3 | 0.0204 |
| TIMP1 | X | 46.3 | 0.0209 |
| PLP2 | X | 47.8 | 0.0212 |
| MAGED1 | X | 50.3 | 0.0331 |
| RPS4X | X | 71 | 0.0124 |
| SSR4 | X | 152.6 | 0.0232 |

corresponding increases in both DNA copy number and mRNA provides further evidence that these could be oncogenes, the activation of which is reflected by genomic amplification and consequent increases in transcript level in lung adenocarcinoma cell lines.

Specific inhibition of EEF1A2 and KCIP-1 expression by siRNAs led to decreased cell proliferation and induction of apoptosis

To further prove the oncogenic function of the identified genes in lung tumorigenesis, we used siRNAs to inhibit the endogenous expression of EEF1A2 and KCIP-1 protein in four lung cancer cell lines (H1563, H229, H522, and SK-LU). Transfection of the cancer cells with specific siRNAs reduced the level of EEF1A2 and KCIP-1 protein by 70–90% 48 h after transfection

(Supplementary Figure 4S). In contrast, EEF1A2 and KCIP-1 protein levels remained unchanged in mock-treated control cells and in cells transfected with a scrambled siRNA sequence. At 48 h after siRNA transfection, the percentage of proliferation of the transfected cancer cells was reduced to 15–30% as compared with 91–100% of cell proliferation of the same cell lines treated with PBS or scrambled siRNA (Supplementary Figure 5S). Apoptosis of siRNA-transfected cells was 27–34%, whereas only 4% of the same cell lines treated with PBS or scrambled siRNA showed apoptosis. These results strongly support an oncogenic role for the identified genes in lung cancer and confirm their potential usefulness as therapeutic targets for the disease.

Amplification and protein expression of KCIP-1 and EEF1A2 in lung tissue

To further validate these findings and to assess the possible clinical significance of the four potential putative oncogenes identified from the cell lines, we first applied fluorescence *in situ* hybridization and immunohistochemical analysis, in parallel, to commercially available human lung tissue microarrays (Ambion, Austin, TX, USA) to evaluate the status of two of these four genes in lung cancer tissue specimens. (Commercially available antibodies to PRDX1 or CALR were not suitable for use in immunohistochemical analysis when this report was written.) Overexpression of KCIP-1 and EEF1A2 protein in the tumors was concordant with amplification of the corresponding genes ($P=0.0003$ for KCIP-1 and $P=0.0011$ for EEF1A2). For example, 16 (35%) of the 46 lung adenocarcinomas in the microarray showed amplification of KCIP-1, and strong cytoplasmic staining for KCIP-1 protein was seen in 18 tumors (39%) (Figure 2). We next examined whether overexpression of these genes was associated with increased cell proliferation by analysing Ki-67 expression in contiguous sections of the tissue microarrays. Positive Ki-67 expression was found to correlate with positive expression of both KCIP-1 ($P=0.02$) and EEF1A2 ($P=0.01$). To extend these findings, we then studied 11 tissue microarray blocks comprising normal and tumor tissue specimens from 113 patients with pathologic stage I non-small-cell lung cancer who had undergone curative surgery (Wang *et al.*, 2005). Immunohistochemical analysis showed that EEF1A2 was expressed in 32 cases (28%) and KCIP-1 in 29 cases (26%). Univariate and multivariate Cox proportional hazards models were used to detect possible associations between EEF1A2 and KCIP-1 expression and clinicopathologic variables. Expression of EEF1A2 or KCIP-1 was associated with short overall survival time ($P=0.0012$ for EEF1A2 and $P=0.0026$ for KCIP-1) (Supplementary Figure 6S). Age at diagnosis, histologic type of cancer, degree of tumor differentiation, and smoking history were not associated with survival time.

Although only two genes were validated in the lung tissue microarrays (because available antibodies to the other two genes were not suitable for use in

Table 2 Proteins showing significant overexpression in cancer cell lines relative to those in normal bronchial epithelial cell lines and their correlation coefficients with increased DNA copy number or mRNA values^a

| Acc. no. | Gene ID | Gene | Mw/pI | Description | r with genomic copy changes ^b | r with mRNA changes ^b |
|----------|---------|---------|-----------|--|--|----------------------------------|
| Q06830 | 5052 | PRDX1 | 48.4/5.4 | Peroxisiredoxin 1 | 0.92364 | 0.91892 |
| Q05639 | 1917 | EEF1A2 | 50.5/5.7 | Eukaryotic translation elongation factor 1 alpha 2 | 0.90218 | 0.89456 |
| P27797 | 811 | CALR | 61/5.5 | Calreticulin | 0.84128 | 0.86434 |
| P63104 | 7534 | KCIP-1 | 27/6.5 | Tyrosine 3-monooxygenase activation protein, zeta | 0.84467 | 0.85499 |
| P07237 | 5034 | P4HB | 54/6.2 | Procollagen-proline, 2-oxoglutarate 4-dioxygenase | 0.91884 | 0.76786 |
| Q04695 | 3872 | KRT17 | 48.0/4.9 | Keratin 17 | 0.00236 | 0.86892 |
| P09211 | 2950 | GSTP1 | 23.2/4.7 | Glutathione S-transferase pi | 0.84218 | 0.69456 |
| P17936 | 3486 | IGFBP-3 | 31.6/5.8 | Insulin-like growth-factor binding protein 3 | 0.06412 | 0.16434 |
| P26641 | 1937 | EEF1G | 50/6.4 | Eukaryotic translation elongation factor 1 gamma | 0.00446 | 0.85549 |
| P08727 | 3880 | KRT19 | 44.1/5.2 | Keratin 19 | -0.04884 | 0.86786 |
| P04792 | 3315 | HSPB1 | 22/6.5 | Heat shock 27 kDa protein 1 | 0.00364 | 0.31892 |
| P00558 | 5230 | PGK1 | 44.5/4.2 | Phosphoglycerate kinase 1 | 0.50402 | 0.79456 |
| Q01995 | 5876 | TAGLN | 22.5/4.3 | Transgelin | -0.34128 | -0.26434 |
| P08631 | 3055 | JTK9 | 59.5/6.8 | Hemopoietic cell kinase | -0.01446 | 0.02549 |
| P09382 | 3956 | LGALS1 | 16/5.5 | Galectin-1, galactoside-binding, soluble, 1 | 0.026623 | 0.01123 |
| Q92784 | 8110 | DPF3 | 25.8/4.8 | D4, zinc and double PHD fingers, family 3 | 0.094884 | -0.03214 |
| P54257 | 9001 | HAP1 | 75.5/6.5 | Huntington-associated protein 1 | 0.12364 | -0.08108 |
| P05783 | 3875 | KRT18 | 48/5.3 | Keratin 18 | 0.010218 | 0.60544 |
| P05787 | 3856 | KRT8 | 9.2/4.4 | Keratin 8 | 0.041280 | 0.84566 |
| P00738 | 3240 | HP | 55.2/6.2 | Haptoglobin | 0.044679 | -0.14501 |
| P09769 | 2268 | FGR | 59.5/5.2 | Gardner-Rasheed feline sarcoma viral oncogene homolog | 0.031264 | -0.13789 |
| P19838 | 4790 | NFKB1 | 50.4/6.3 | Nuclear factor of kappa light gene enhancer in B-cells 1 | 0.04467 | -0.14501 |
| P29034 | 6273 | S100A2 | 10.9/4.6 | S100 calcium-binding protein A2 | 0.87964 | 0.243214 |
| Q13105 | 7709 | ZBTB17 | 87.9/5.3 | Zinc-finger and BTB domain containing 17 | -0.17636 | 0.048108 |
| Q00987 | 4193 | MDM2 | 75.2/4.8 | Transformed 3T3 cell double minute 2 | -0.19782 | -0.50544 |
| P27816 | 4134 | MAP4 | 111/5.4 | Microtubule-associated protein 4 | 0.25872 | -0.05356 |
| P52732 | 3832 | KIF11 | 119.2/6.2 | Kinesin family member 11 | -0.25778 | -0.53444 |
| P25205 | 4172 | MCM3 | 90.9/5.5 | Minichromosome maintenance deficient 3 | 0.25644 | 0.053666 |
| P08631 | 3055 | HCK | 59.5/5.7 | Hemopoietic cell kinase | 0.65533 | 0.054501 |
| P09237 | 4316 | MMP7 | 22.6/5.8 | Matrix metalloproteinase 7 | 0.234987 | 0.876820 |
| P30305 | 994 | CDC25B | 64.9/4.5 | Cell division cycle 25B | 0.045116 | 0.283214 |
| P50290 | 998 | CDC42 | 21.3/6.1 | Cell division cycle 42 (GTP-binding protein, 25 kDa) | -0.47636 | 0.088108 |
| P61586 | 387 | RHOA | 19.8/6.9 | Ras homolog gene family, member A | -0.49782 | -0.00544 |
| P63000 | 5879 | RAC1 | 21.5/6.8 | Ras-related C3 botulinum toxin substrate 1 | -0.05583 | -0.03566 |
| P07437 | 203068 | TUBB | 49.6/6.5 | Tubulin, beta polypeptide | 0.255533 | 0.145010 |
| P24864 | 898 | CCNE1 | 47.1/4.3 | Cyclin E1 | -0.65116 | 0.232149 |
| P04141 | 1437 | CSF2 | 16.9/6.3 | Colony stimulating factor 2 (granulocyte-macrophage) | -0.64636 | -1.28108 |
| P28072 | 5694 | PSMB6 | 25.3/5.2 | Proteasome (prosome, macropain) subunit, beta type, 6 | -0.69782 | -1.30544 |
| P00352 | 216 | ALD- | 54.7/4.3 | Aldehyde dehydrogenase 1 family, member A1 | -0.75872 | 0.03356 |
| | | H1A1 | | | | |
| Q03013 | 2948 | GTM4 | 25.3/5.0 | Glutathione S-transferase M4 | -0.78533 | 0.134501 |
| P63241 | 1984 | EIF5A | 10/4.4 | Eukaryotic translation initiation factor 5A | -0.97893 | -1.44321 |
| Q01469 | 2171 | EFABP | 18.0/4.2 | Fatty acid-binding protein 5 | 0.25684 | -0.36432 |

^aOnly the gene showing statistically significant increased protein expression with increases in both genomic copy number and transcript simultaneously will be considered as potential putative oncogene in lung adenocarcinoma cells. ^br, Spearman correlation coefficients between proteins and genomic or mRNA values are based on all six cancer cell lines; bold indicates $P < 0.05$, if $r > 0.84000$. Mw, molecular weight; pI, isoelectric point.

immunohistochemical analysis), these findings are consistent with those from our cell lines, demonstrating again that genomic amplification and consequent increases in amounts of transcript may be, at least in part, driving the abundance of proteins in these lung tumors. The association between expression of these genes and that of Ki-67, a known indicator of poor prognosis in lung cancer (Martin *et al.*, 2004), suggests that activation of these genes may be an indicator of tumor aggressiveness. These results also suggest that expression of EEF1A2 and KCIP-1 proteins in stage I non-small-cell lung cancer may be useful as a marker for distinguishing patients with relatively poor prognosis from those who might benefit from adjuvant treatment.

Discussion

Our current study illustrates the power of integrated functional genomic analyses for identifying putative oncogenes and for evaluating their potential clinical significance. Among the four identified oncogenes, three genes (*PRDX1*, *CALR*, and *KCIP-1*) have been implicated in lung tumorigenesis. *PRDX1* is an antioxidant protein involved in regulating cell proliferation, differentiation, and apoptosis. Kim *et al.* (2003) found *PRDX1* expression to be elevated in both lung cancer and adjacent normal lung tissue, suggesting that activation of *PRDX1* may enhance proliferation in lung cancer. *CALR* has a major role in Ca^{2+} binding and the

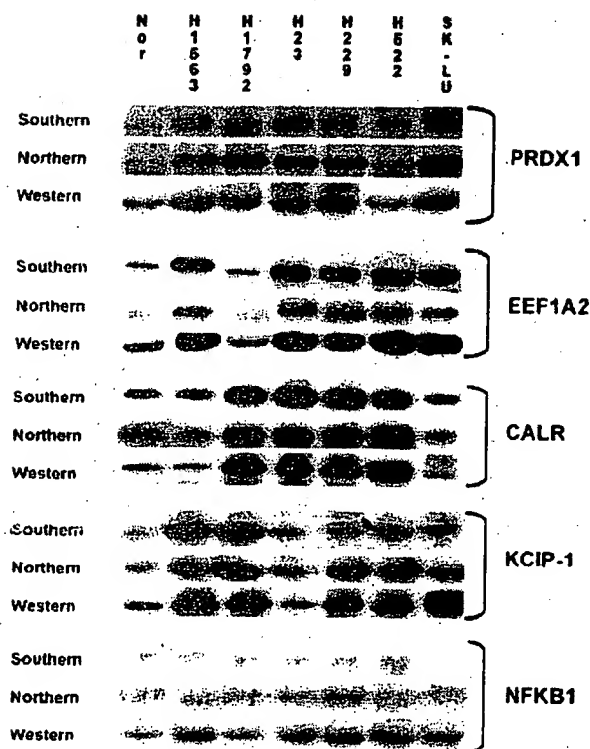


Figure 1 Confirmation by Southern, Northern, and Western blot analyses of increased DNA copies, transcript levels, and protein levels in the four genes identified in high-throughput analyses. For comparison, we arbitrarily chose one gene, *NFKB1*, in which an increased protein level did not correlate with genetic changes. The blotting results are consistent with the results from the CGH array, transcript array, and proteomic analyses. Nor, indicates normal bronchial epithelial cell line. All the experiments were repeated at least three times with each cell line. Means of normalized to β -actin signal intensities on Southern, Northern, and Western blots, along with 95% confidence intervals, were calculated (β -actin signals are not shown in the figure; two different normal bronchial epithelial cell lines were used in the confirmation and only one normal cell line is shown in the figure).

transcriptional regulation of other genes and was recently found to be overexpressed in 73% of 40 lung adenocarcinomas (Oates and Edwards, 2000). *KCIP-1* belongs to the 14-3-3 family, which participates via the MAPK and Wnt signaling pathways in the regulation of many cellular processes including cell proliferation and differentiation as well as tumorigenesis (Thomas *et al.*, 2005). *KCIP-1* was recently found to be expressed in all 12 lung tumors tested in a single-institution study (Qi *et al.*, 2005). Interestingly, *EEF1A2* was originally considered a putative oncogene in ovarian cancer on the basis of its being amplified in 25% and overexpressed in 30% of the same set of ovarian tumors (Anand *et al.*, 2002); functional analyses have established its oncogenic role in cellular transformation (Lee, 2003). Our discovery that *EEF1A2* may be a putative oncogene in lung adenocarcinoma demonstrates the power of our functional genomic strategy for rapidly identifying potential oncogenes.

Although the main focus of this study was to specifically identify putative oncogenes, it should be

noted that 90.7% of the genes showing high protein expression did not show corresponding increases in both DNA copy number and transcript, a finding consistent with that of others that transcriptional, translational, and post-translational regulatory mechanisms can greatly influence the abundance of protein in lung tumorigenesis (Chen *et al.*, 2002). For example, *NFKB1* is a critical arbiter of immune responses, cell survival, and transformation and is often activated in several types of tumors (Chen *et al.*, 2002). Deregulation of *NFKB1* is thought to be modulated through phosphorylation of Ser337 by protein kinase A (Chen *et al.*, 2002). In our study, 68.8% of the genes showing over-representation in the genome did not show elevated transcript levels, implying that at least some of these genes are 'passenger' genes that are concurrently amplified because of their location with respect to amplicons but lack biological relevance in terms of the development of lung adenocarcinoma.

Although the potential oncogenes we identified here are likely to be important, certainly other oncogenes could be involved in the development of lung adenocarcinoma. The oligo microarray we used consists of 22 000 probes, which represent only about 60% of the human genome. Moreover, each probe was designed for the 3' region of expressed sequence tags of the selected genes. Also, our results were initially derived from cancer cell lines, although the findings were later confirmed in human tissue samples. Our ongoing study using microarrays with information on more genes and the development of high-resolution proteomic analyses for use with larger numbers of specimens will allow more comprehensive analyses of the molecular consequences of gene amplifications. Such expanded analyses will very likely lead to the identification of additional oncogenes.

Some of the results of our current study were comparable to those of other studies of lung cancer. For example, genomic copy number and protein levels of *KCIP-1* were previously found to be amplified and overexpressed in primary lung cancers by cDNA clone-based CGH array analysis (Jiang *et al.*, 2004) and proteomic analysis (Chen *et al.*, 2002), respectively. Our functional genomic approach, which integrates simultaneous CGH, transcript microarrays, proteomic analyses, and siRNA, allows us not only to quickly identify potential oncogenes but also to explore their significance as diagnostic and therapeutic targets in tumor progression – more than could be achieved by any technique alone.

Genes identified in this way may serve as promising targets for diagnosis and therapy in lung adenocarcinoma. Further research on the clinical implications of such genes is needed; experiments now underway in our laboratory include overexpression of the genes in normal cells, disruption of the function of these genes in cancer cells, and investigation of how interactions among these genes (or interactions with other known oncogenes) may mediate the expression of the transformed phenotype.

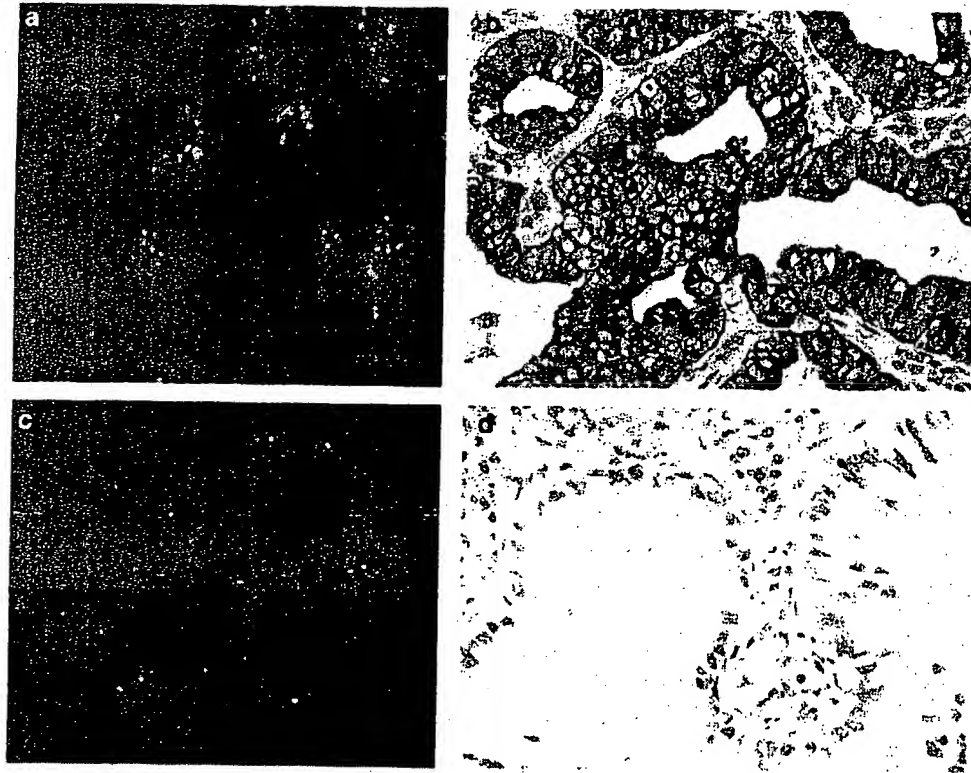


Figure 2 *EEF1A2* amplification is associated with high *EEF1A2* protein expression in lung adenocarcinomas. (a) Cells from a lung adenocarcinoma sample in which *EEF1A2* is amplified show more green signals (*EEF1A2*) than red signals (chromosome 20 centromeric probe) (original magnification, $\times 400$). (b) Immunohistochemical staining of cells from the same tissue sample as in panel a shows strong *EEF1A2* staining in the cytoplasm. (c) A lung adenocarcinoma sample with two copies of *EEF1A2* and chromosome 20 centromeric probe, indicating no *EEF1A2* amplification (original magnification, $\times 400$). (d) Immunohistochemical staining of cells from the same tissue sample as in panel c shows negative staining for *EEF1A2*.

Materials and methods

Cell lines

Six human lung adenocarcinoma cell lines (H23, H229, H1792, SK-LU-1, H522, and H1563) were obtained from the American Type Culture Collection (Manassas, VA, USA). Two normal bronchial epithelial cell lines were obtained from Clontech (Palo Alto, CA, USA). Genomic DNA, mRNA, and protein were derived from a single harvest of these cells.

DNA and RNA profiles by microarray analysis

Genomic DNA labeling and hybridization were performed as described previously (Barrett *et al.*, 2004) with Agilent's Human 1A Oligo Microarray (V2) (Agilent Technologies, Palo Alto, CA, USA), which contains 22 000 unique 60-mer oligos. Details of the protocol for analysing transcripts are available at <http://www.chem.agilent.com>. Map positions for arrayed genes were assigned by identifying the DNA sequence represented in the UniGene cluster and matching it with the Golden Path genome assembly (<http://genome.ucsc.edu/>; Mat 7, 2004 Freeze). Microarray images of DNA copy number and expression were analysed by using AgilentCGH Analytics and Feature Extraction software. DNA copy number profiles that deviated significantly from background signal ratios (measured from normal control cell hybridization, as described elsewhere; Barrett *et al.*, 2004) were interpreted as evidence of true differences in DNA copy number. The criteria for defining genomic over-representation and amplicons are described elsewhere (Hyman *et al.*, 2002); details are given in the

Supplementary Information. An increase in mRNA level was defined as a twofold increase in signal ratio relative to that of the control ($\log_2 > 1$).

Quantitative two-dimensional PAGE and mass spectrometry

Analysis of proteins by two-dimensional PAGE and their identification by mass spectrometry were performed as previously described (Shen *et al.*, 2004). Briefly, protein pellets were solubilized in rehydration buffer, after which the first-dimension isoelectric focusing was carried out with a Protean IEF Cell (Bio-Rad Laboratories) and the second-dimension separation was carried out with Bio-Rad's Ready Gel Precast Gels and the Bio-Rad Criterion Cell apparatus. Protein spots were visualized by silver-based staining, and all gels were assessed with Bio-Rad's PDQuest 2D gel image analysis software. Selected spots were subjected to in-gel tryptic digestion and analysed on a Voyager-DE PRO matrix-assisted laser desorption ionization/time-of-flight mass spectrometer (Applied Biosystems, Foster City, CA, USA). The mass list of the 20 most intense monoisotopic peaks for each sample was entered in the MS-Fit search program (v3.2.1) (<http://prospector.ucsf.edu/ucsfhtml4.0/msfit.htm>) and searched in the National Center for Biotechnology Information protein database.

Southern, Northern, and Western blot analyses

Southern, Northern, and Western blot hybridizations were performed according to standard protocols. cDNA clones for the tested genes were purchased from Invitrogen (Carlsbad,

CA, USA) and prepared as probes for the blot hybridizations. Antibodies used were obtained as follows: PRDX1, CALR, NFKB1, KCIP-1, and β -actin from Santa Cruz Biotechnology (Santa Cruz, CA, USA); and EEFA2 from Upstate Biotechnology (Waltham, MA, USA).

Fluorescence in situ hybridization and immunohistochemical analyses of lung tissue microarrays

Fluorescence *in situ* hybridizations and immunohistochemical analyses of KCIP-1 and EEFA2 were carried out as described elsewhere (Jiang et al., 2002; Wang et al., 2005) with Lung Tissue Microarrays (Ambion, Austin, TX, USA) and 11 homemade microarray blocks containing tissue samples from 113 patients with pathologic stage I non-small-cell lung cancer (Wang et al., 2005). DNA probes specific for KCIP-1 and EEFA2 were obtained by screening a Human BAC Clone library (Invitrogen) by polymerase chain reaction as described previously (Jiang et al., 2002). The antibodies used for the immunohistochemical analyses were the same as those used for the Western blotting. Cell proliferation of the lung tissues was assessed with a Ki-67 monoclonal antibody from Santa Cruz Biotechnology. Definitions of the cutoff value for a positive result of each antibody are shown in Supplementary Information.

siRNA transfection, cellular proliferation assay, and apoptosis analysis

Transfections were carried out by using siPORT Lipid Transfection Agent (Ambion) with siRNAs targeting KCIP-1 or EEFA2 or with a scrambled siRNA duplex (siControl) (Dharmacon Inc., Lafayette, CO, USA), with PBS used as a negative control (Jiang et al., 2002). Cells were fixed 24, 48, or 96 h later and subjected to further tests. All siRNAs were prepared by using a transcription-based method with Silencer siRNA according to the manufacturer's instructions (Ambion). Sequences of the individual siRNAs are listed in Supplementary Table 4S. Inhibition of cell growth by the

siRNAs was determined by MTT staining, and cell growth rate was plotted against the percentage of viable cells in the saline-treated controls (a value arbitrarily set at 100%) (Jiang et al., 2002). Apoptosis was analysed by fluorescence cell cycle analysis of terminal deoxynucleotidyl transferase-mediated dUTP nick-end labeling with FITC-labeled dUTP (Boehringer Mannheim Biochemicals, Mannheim, Germany) (Jiang et al., 2005).

Statistical analyses

Relationships between gene copy number and mRNA level were examined as described elsewhere (Hyman et al., 2002, Supplementary Information). Correlations between protein abundance and DNA copy number and mRNA expression of the corresponding genes were evaluated with the Spearman correlation coefficient. Fisher's exact test and χ^2 -tests were used to analyse associations between amplification and expression of the candidate genes with various histopathologic variables of the samples in the tissue microarrays. Univariate and multivariate analyses were carried out with Cox's proportional hazards model to determine which independent factors might have a joint significant influence on survival. A P -value ≤ 0.05 was considered statistically significant; all statistical tests were based on a two-sided significance level.

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References

- Anand N, Murthy S, Amann G, Wernick M, Porter LA, Cukier IH et al. (2002). *Nat Genet* 31: 301–305.
- Barrett MT, Scheffer A, Ben-Dor A, Sampas N, Lipson D, Kincaid R et al. (2004). *Proc Natl Acad Sci USA* 101: 17765–17770.
- Chen G, Gharib TG, Huang CC, Taylor JM, Misk DE, Kardia SL et al. (2002). *Mol Cell Proteomics* 1: 304–313.
- Hyman E, Kauraniemi P, Hautaniemi S, Wolf M, Mousses S, Rozenblum E et al. (2002). *Cancer Res* 62: 6240–6245.
- Jiang F, Caraway NP, Li RY, Katz RL. (2005). *Oncogene* 24: 3409–3418.
- Jiang F, Lin F, Price R, Gu J, Medeiros LJ, Zhang HZ et al. (2002). *J Mol Diagn* 4: 144–149.
- Jiang F, Yin Z, Caraway NP, Li R, Katz RL. (2004). *Neoplasia* (New York) 6: 623–635.
- Kim HJ, Chae HZ, Kim YJ, Kim YH, Hwang TS, Park EM et al. (2003). *Cell Biol Toxicol* 19: 285–298.
- Lee JM. (2003). *Reprod Biol Endocrinol* 1: 69–73.
- Luk C, Tsao MS, Bayani J, Shepherd F, Squire JA. (2001). *Cancer Genet Cytogenet* 125: 87–99.
- Martin B, Paesmans M, Mascaux C, Berghmans T, Lohthaire P, Meert AP et al. (2004). *Br J Cancer* 91: 2018–2225.
- Oates J, Edwards C. (2000). *Histopathology* 36: 341–347.
- Pollack JR, Sorlie T, Perou CM, Rees CA, Jeffrey SS, Lonning PE et al. (2002). *Proc Natl Acad Sci USA* 99: 12963–12968.
- Qi W, Liu X, Qiao D, Martinez JD. (2005). *Int J Cancer* 113: 359–363.
- Ross JS, Fletcher JA. (1999). *Semin Cancer Biol* 9: 125–138.
- Shen J, Person MD, Zhu J, Abbruzzese JL, Li D. (2004). *Cancer Res* 64: 9018–9026.
- Thomas D, Guthridge M, Woodcock J, Lopez A. (2005). *Curr Top Dev Biol* 67: 285–303.
- Tonon G, Wong KK, Maulik G, Brennan C, Feng B, Zhang Y et al. (2005). *Proc Natl Acad Sci USA* 102: 9625–9630.
- Wang H, Zhang Z, Li R, Ang KK, Zhang H, Caraway NP et al. (2005). *Int J Cancer* 116: 285–290.

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